Nuisance Insects of Rearing mass Asian Citrus Psyllid (ACP) Diaphorina citri Under Controlled Conditions

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> Abstract. Rearing insects for pest experiment it's very critical. The condition of rearing such insects impacted the smooth running of research. This condition also happens in the rearing of Asian Citrus Psyllid (ACP), Diaphorina citri. ACP is one of the most dangerous pests of citrus since responsible for the huanglongbing disease vector in citrus. Many researchers, either laboratory or field-based use a D.citri for object experiments. A clear and recent description of the types of insects or arthropods that disturb the rearing ACP is needed, especially in Indonesia. The research was conducted in the greenhouse of Faculty of Agriculture, Gadjah Mada University, from January 2021 to September 2022. The rearing of ACP uses orange jasmine (Muraya paniculata) and citrus (Citrus nobilis). The collected insect and spider preserved in alcohol or pined in insect tray and identify them based on related literature. This study proposed to assess the nuisance insect that infested on rearing of D.citri based on greenhouse conditions. Our results showing that the many nuisance insects that disturb the growth of rearing in D.citri. A total of nine insect and spider attacking D citri, they are black ants (Dolichoderus sp), aphid (Aphis sp), leaf roller caterpillar (Phyllocnistis citrella), cockroach (Blattella sp), ladybug (Exochomus nigromaculatus), mite (Tetranychus sp), Mealy bug (Pseudoccocus sp), Scale insect (Aonidella aurantia) and Spider. Those nuisance insects and spiders attack D.citri colonies in different ways and stages. The attacking nuisance insect affected to affected the of the ACP size culture.

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1 Introduction

Rearing of any insects is important for providing insects in any level of generation for specific research interests. Rearing main purpose is to provide reliable and affordable high-quality insects (Leppla, 2009), for sure it's dependent on researcher needs. Rearing of insect including ACP, either mass or laboratory scale is key roles for successful of any insect treatment. Researchers or users need to pay attention to insect rearing since some effects on insects from laboratory adaptation, inbreeding depression, inadvertent selection, or direct rearing effects (Sørensen et al., 2012). Since rearing insects is critical to any scope of insect research, the rearing condition becomes very important.

Diaphorina citri (DC) or known as Asian Citrus Psyllids (ACP) are one of the most responsible vectors, transmitting Huanglongbing disease in almost every country around the world (Berk, 2016; Urbaneja et al., 2020). Rearing D.citri has been recently very important for studies of the transmission of bacteria causing hualongbing, Candidatus Liberibacter asiaticus (Las) (Li et al., 2006; Paris et al., 2013). Providing good and healthy D.citri during the experiment are challenging since any nuisance insect or other organisms bothering D.citri growth. The nuisance insect recorded in disturbing D.citri growth and colony such as Tamarixia radiata (Irvin et al., 2021; Kalile et al., 2022), coccinellids (Coleoptera: Coccinellidae), lacewings (Neuroptera: Chrysopidae), syrphids (Diptera: Syrphidae) and spiders (Arachnida: Araneae), and Diaphorencyrtus aligarhensis Shafee, Alam & Argarwal (Corallo et al., 2021; Michaud, 2004). The extensive method of rearing ACP described by (Skelley & Hoy, 2004) includes prerequisites of their parasitoids.

2 Materials and method

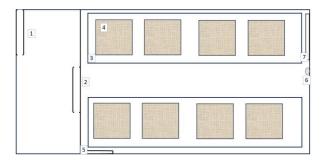
2.1 Rearing set

The rearing of ACP uses two common plants, orange jasmine (Murava paniculata) and Sweet orange (Citrus nobilis). The rearing is set in a 1x1x1 meter square cage covered with net cloth, the cage maintained under a greenhouse with 27±1 °C and 60% Humidity. Every cage contains six to eight individual plants depending on their canopy. The plant was pruned two weeks before use as a feed for ACP. Every month plants are fertilized with common NPK fertilizer through diluted two spoons (equal to 2 grams) of fertilizer in 10 liter fresh water and poured to plant media as needed. The designs of the greenhouse and rearing set are described in Fig 1. Greenhouses typically consist of two separate doors (no.1 and 2) to prevent insects and other unwanted organisms from entering the rearing area. Inside of green house consisted two benches separately (no. 3) and four cages (1X1X1m) (no. 4) on the bench. The greenhouse is also provided with a fan (no.7) and humidifier apparatus (no. 5) to maintain temperature and humidity under optimal conditions for ACP

rearing. Meanwhile, for watering, a water tap equipped with a hose is provided (no. 6)

2.2 *D.citri* infestation and nuisance insect observation

Ten to fifteen of matured adults ACP laid on new shoots of M.paniculata and C.nobilis using an aspirator. The colony was covered with a 10 cm diameter of a plastic tube covered with net cloth. The colony was left inside the tube for six days and opened immediately after the egg was laid. Eggs are allowed to hatch naturally and are monitored regularly. During the development of egg D.citri, observations are conducted at least every week to see if nuisance insects are coming and disturbing. The individuals of nuisance insects are collected and noted if available. The collected insect and arthropod are preserved in alcohol or pinned depend on their size and type of body. All the collected insect and and arthropod are identify based on relevant literature and online catalogue such as Encyclopedia of Life (https://eol.org/).



Note: 1) First door; 2) Second door; 3) Bench; 4) cages (1x1x1m); 5) Humidifier; 6) Water tap; 7) Fan

Fig. 1. Design of Greenhouse and rearing set during observations of nuisance insect.

3 Results and discussion

The results during observation rearing facilities of ACP are provided in Table 1. Every insect and arthropod have different type of disturbing even predator, parasitoid, or competitor. Those roles are describing as possible their type of disturbing and stadia of ACP.

 Table 1. Kind of nuisance insect and arthropod in rearing facilities of ACP.

N o	Organism	Possible role	Symptom appearanc e	Stadia disturbe d
1	Black ants (<i>Dolichoderus</i> sp)	Predator	Unknown	Egg and nymph
2	Aphid (<i>Aphis</i> sp)	Competito r	Leaf curling, especially	Egg and nymph

			in new flush	
3	Leaf roller caterpillar (Phyllocnistis citrella)	Competito r	The leaves roll up and stick to each other	Egg and nymph
4	Cockroach (<i>Blattella</i> sp)	Predator	Unknown	Egg, Nymph, and imago
5	Lady bug (Exochomus nigromaculatu s)	Competito r, Predator	pale spots on leaves	Nymph and imago
6	Mite (<i>Tetranychus</i> sp),	Competito r, Predator	solid white spots on the leaf surface	Egg, Nymph, and imago
7	Mealy bug (<i>Pseudoccocus</i> sp),	Competito r	yellow spot and adherent white colonies on leave and stem	Egg, Nymph, and imago
8	Scale insect (Aonidella aurantia)	Competito r	The leaves turn white and the stems dry white	Egg, Nymph, and Imago
9	Spider	Predator	marked by the emergence of the web	Egg, Nymph, and imago

Some insects are commonly pests in the rearing facilities of ACP in every country. Skelley & Hoy (2004) described them very well of Scale insects and Aphid as quarantine pests who mostly attack ACP rearing mass. In our greenhouse and rearing facility's Scale insects, Aonidella aurantia, and Aphid, Aphis sp are two common pests who disturb the rearing colony in almost every season. The possibilities of Scale insects and aphids coming from the unclear plant, Murava, and Citrus, when they were used as prey for rearing. These problems are common in ACP rearing as (Skelley & Hoy, 2004) mentions in their notes. Murava and Citrus are favoured host plants for ACP (Hall, 2014; Setyaningrum et al., 2023) which also naturally favor other insects such as Thrips, Frankliniella spp ((Hall, 2014), scale insects (Poudel et al., 2022), and any other insect from various orders (Zuhran et al., 2021).

Spider, especially from the Aranea order is the mostly diverse group, consisting of over 42,000 species in 109 families, which are all predators and primarily feed on insects (Humenik et al., 2011). This situation makes Aranae very easy to find in every place where insects settle. In tropical rearing ACP facilities such as, it's almost found in every season both wet and dry season. It's also happened in nature or citrus orchards such as West Kalimantan (Zuhran et al., 2021). Mites are citrus pests, that have a symptom bronzing or silvering effect on the leaves where they attacked on(KENNETT et al., 1999). Differences from the spider, mites mostly attacked plants rather than *D.citri* colony. In these situations, mites become competitors or specific predators, especially on eggs of D.citri.

The black ant, Dolichoderus sp is commonly associated with seasonal fruit trees, including Sapodilla and Citrus (Jutsum et al., 1981; Van Mele & Cuc, 2001). Black ants in some cases came to the citrus trees associated with another insect, such as Scale insect (Jutsum et al., 1981; Michael Githae et al., 2020). These situations may happen in the greenhouse, since we found Scale insects Aonidella aurantia and Dolichoderus sp, in citrus and Muraya. Controlling Scale insects will impact directly or indirectly to Dolichoderus infestation inside rearing facilities. Mostly, the infestation of scale insects comes from the previous stock of plant rearing. As we know, scale insects are one of the serious insects that are difficult to control even when applying pesticides. It could happen also in the Leaf roller caterpillar (*Phyllocnistis citrella*) which is a common pest in citrus (Moore & Duncan, 2017), especially in the dry season (Foda et al., 2021). The leaf roller disturbing rearing since young leaves, which D.citri are the suitable places for growth. By infestation leaf roller, D.citri is unable to lay eggs. The possibilities of pathway's nuisance insect and arthropod-infested D.citri rearing are described in Fig. 2.

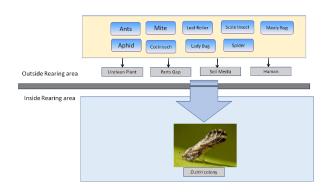


Fig. 2. Nuisance Insect and Arthropod Pathways Disturbance of ACP rearing

4 Conclusion

Nuisance insects of rearing ACP are black ants (*Dolichoderus sp*), aphid (*Aphis sp*), leaf roller caterpillar (Phyllocnistis citrella), cockroach (*Blattella sp*), ladybug (*Exochomus nigromaculatus*), mite

(*Tetranychus sp*), Mealy bug (*Pseudoccocus sp*), Scale insect (*Aonidella aurantia*) and Spider. That nuisance insect played a different role in disturbing the growth of ACP colony. The possible role could be competitor or predator which affected to any stages of ACP growth. The potential pathways that allow nuisance insects and arthropods to pass through unclean plant host sources, gaps in rearing facilities, or through human movement during the rearing process.

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6 Selected references

1. Leppla, N. C. Rearing of Insects. Encycl. Insects 866– 869 (2009) doi:10.1016/B978-0-12-374144-8.00227-7.

2. Sørensen, J. G., Addison, M. F. & Terblanche, J. S. Mass-rearing of insects for pest management: Challenges, synergies and advances from evolutionary physiology. Crop Prot. 38, 87–94 (2012).

3. Urbaneja, A. et al. Citrus pests in a global world. The Genus Citrus 333–348 (2020) doi:10.1016/B978-0-12-812163-4.00016-4.

4. Berk, Z. Diseases and pests. Citrus Fruit Process. 83– 93 (2016) doi:10.1016/B978-0-12-803133-9.00005-9.

5. Li, W., Hartung, J. S. & Levy, L. Quantitative realtime PCR for detection and identification of Candidatus Liberibacter species associated with citrus huanglongbing. J. Microbiol. Methods 66, 104–115 (2006).

6. Paris, T. M., Rohde, B. B., Allan, S. A., Mankin, R. W. & Stansly, P. A. SYNCHRONIZED REARING OF MATED AND UNMATED DIAPHORINA CITRI (HEMIPTERA: LIVIIDAE) OF KNOWN AGE. Florida Entomol. 96, 1631–1634 (2013).

7. Irvin, N. A., Pierce, C. & Hoddle, M. S. Evaluating the potential of flowering plants for enhancing predatory hoverflies (Syrphidae) for biological control of Diaphorina citri (Liviidae) in California. Biol. Control 157, 104574 (2021).

8. Kalile, M. O. et al. UV light attracts Diaphorina citri and its parasitoid. Biol. Control 170, 104928 (2022).

9. Corallo, A. B., Pechi, E., Bettucci, L. & Tiscornia, S. Biological control of the Asian citrus psyllid, Diaphorina citri Kuwayama (Hemiptera: Liviidae) by Entomopathogenic fungi and their side effects on natural enemies. Egypt. J. Biol. Pest Control 31, 1–9 (2021).

10. Michaud, J. P. Natural mortality of Asian citrus psyllid (Homoptera: Psyllidae) in central Florida. Biol. Control 29, 260–269 (2004).

11. Skelley, L. H. & Hoy, M. A. A synchronous rearing method for the Asian citrus psyllid and its parasitoids in quarantine. Biol. Control 29, 14–23 (2004).

12. Hall, D. G. Interference by Western Flower Thrips in rearing Asian citrus psyllid: Damage to host plants and facultative predation. Crop Prot. 60, 66–69 (2014).

13. Setyaningrum, H., Martono, E., Soffan, A. & Mo, J. Best Practices Intercropping Citrus Controlling Asian Citrus Psyllids (Diaphorina citri) in Indonesia. Adv. Biol. Sci. Res. 29, 591–596 (2023).

14. Poudel, A., Sapkota, S., Pandey, N., Oli, D. & Regmi, R. Causes of citrus decline and its management practices adopted in Myagdi district, Nepal. Heliyon 8, e09906 (2022).

15. Zuhran, M., Mudjiono, G. & Dyah Puspitarini, R. Pengaruh pengelolaan agroekosistem terhadap kelimpahan kutu loncat jeruk Diaphorina citri Kuwayama (Hemiptera: Liviidae). J. Entomol. Indones. 18, 102–102 (2021).

16. Humenik, M., Scheibel, T. & Smith, A. Spider silk: Understanding the structure-function relationship of a natural fiber. Prog. Mol. Biol. Transl. Sci. 103, 131–185 (2011).

17. KENNETT, C. E., MCMURTRY, J. A. & BEARDSLEY, J. W. Biological Control in Subtropical and Tropical Crops. Handb. Biol. Control 713–742 (1999) doi:10.1016/B978-012257305-7/50074-6.

18. Van Mele, P. & Cuc, N. T. T. Farmers' Perceptions and Practices in Use of Dolichoderus thoracicus (Smith) (Hymenoptera: Formicidae) for Biological Control of Pests of Sapodilla. Biol. Control 20, 23–29 (2001).

19. Jutsum, A. R., Cherrett, J. M. & Fisher, M. Interactions Between the Fauna of Citrus Trees in Trinidad and the Ants Atta cephalotes and Azteca Sp. J. Appl. Ecol. 18, 187 (1981).

20. Michael Githae, George O. Ong'amo, John Nderitu, Gillian W. Watson & Wanja Kinuthia. Ants (Hymenoptera: Formicidae) associated with scale insects (Hemiptera: Coccomorpha) on citrus trees in Coastal and Lower Eastern Counties, Kenya. journal of Agricultural Science and Practice 245–249

https://www.researchgate.net/publication/348381949_Ants_ Hymenoptera_Formicidae_associated_with_scale_insects_He miptera_Coccomorpha_on_citrus_trees_in_Coastal_and_Lo wer Eastern Counties Kenya (2020).

21. Moore, S. D. & Duncan, L. W. Microbial Control of Insect and Mite Pests of Citrus. Microb. Control Insect Mite Pests From Theory to Pract. 283–298 (2017) doi:10.1016/B978-0-12-803527-6.00019-6.

22. Foda, Y. L., Wibowo, L., Lestari, P. & Hasibuan, R. INVENTARISASI DAN INTENSITAS SERANGAN HAMA TANAMAN JERUK (Citrus sinensis L.) DI KECAMATAN SEKAMPUNG UDIK KABUPATEN LAMPUNG TIMUR. J. Agrotek Trop. 9, 367 (2021).