Check for updates

OPEN ACCESS

EDITED BY Rozeta Sokou, Nikaia General Hospital "Aghios Panteleimon", Greece

REVIEWED BY

Christoph Härtel, University Hospital Würzburg, Germany Maria Oana Sasaran, "George Emil Palade" University of Medicine, Pharmacy, Sciences and Technology of Târgu Mureş, Romania

*CORRESPONDENCE Brynne A. Sullivan Sosa4m@uvahealth.org

RECEIVED 13 November 2023 ACCEPTED 08 January 2024 PUBLISHED 19 January 2024

CITATION

Kumar R, Kausch SL, Gummadi AKS, Fairchild KD, Abhyankar MM, Petri WA Jr and Sullivan BA (2024) Inflammatory biomarkers and physiomarkers of late-onset sepsis and necrotizing enterocolitis in premature infants. Front. Pediatr. 12:1337849. doi: 10.3389/fped.2024.1337849

COPYRIGHT

© 2024 Kumar, Kausch, Gummadi, Fairchild, Abhyankar, Petri and Sullivan. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Inflammatory biomarkers and physiomarkers of late-onset sepsis and necrotizing enterocolitis in premature infants

Rupin Kumar¹, Sherry L. Kausch², Angela K. S. Gummadi², Karen D. Fairchild², Mayuresh M. Abhyankar³, William A. Petri Jr³ and Brynne A. Sullivan²*

¹Department of Pediatrics, Division of Neonatology, University of Kentucky College of Medicine, Lexington, KY, United States, ²Department of Pediatrics, Division of Neonatology, University of Virginia School of Medicine, Charlottesville, VA, United States, ³Department of Internal Medicine, Division of Infectious Diseases, University of Virginia School of Medicine, Charlottesville, VA, United States

Background: Early diagnosis of late-onset sepsis (LOS) and necrotizing enterocolitis (NEC) in very low birth weight (VLBW, <1,500 g) infants is challenging due to non-specific clinical signs. Inflammatory biomarkers increase in response to infection, but non-infectious conditions also cause inflammation. Cardiorespiratory data contain physiological biomarkers, or physiomarkers, of sepsis that may be useful in combination with inflammatory hematologic biomarkers for sepsis diagnosis.

Objectives: To determine whether inflammatory biomarkers measured at the time of LOS or NEC diagnosis differ from times without infection and whether biomarkers correlate with cardiorespiratory sepsis physiomarkers in VLBW infants. **Methods:** Remnant plasma sample collection from VLBW infants occurred with blood draws for routine laboratory testing and suspected sepsis. We analyzed 11 inflammatory biomarkers and a pulse oximetry sepsis warning score (POWS). We compared biomarker levels obtained at the time of gram-negative (GN) bacteremia or NEC, gram-positive (GP) bacteremia, negative blood cultures, and no suspected infection.

Results: We analyzed 188 samples in 54 VLBW infants. Several biomarkers were increased at the time of GN LOS or NEC diagnosis compared with all other samples. POWS was higher in patients with LOS and correlated with five biomarkers. IL-6 had 78% specificity at 100% sensitivity to detect GN LOS or NEC and added information to POWS.

Conclusion(s): Inflammatory plasma biomarkers discriminate sepsis due to GN bacteremia or NEC and correlate with cardiorespiratory physiomarkers.

KEYWORDS

prematurity, neonatal sepsis, biomarkers, inflammation, heart rate, oxygenation, cardiorespiratory

1 Introduction

Sepsis and necrotizing enterocolitis (NEC) lead to significant morbidity and mortality in premature infants (1). Inflammation due to sepsis and NEC can cause end-organ damage (2, 3), including brain injury, which leads to long-term impairment (4–6). Early recognition and treatment may improve outcomes, but signs and symptoms often overlap with non-infectious conditions (7). Therefore, a diagnosis is often made once the

infection is advanced, and on the other hand, empiric antibiotics are given when there is no infection (8). Clinicians have limited tests and data to guide decisions on the likelihood of infection.

Inflammatory molecules, upregulated as part of the immune response to infection, hold promise as biomarkers of neonatal sepsis. A biomarker with sufficient diagnostic accuracy could be a useful screening tool to aid in deciding whether to start antibiotics or continue monitoring for further signs of sepsis. Many studies have evaluated biomarkers for neonatal sepsis in premature infants (9–12). But, few have been translated into clinical care (13). Decisions to start or stop antibiotics in the NICU largely rely on the clinician's assessment of clinical signs and non-specific laboratory tests, such as C-reactive protein (CRP) (14) and complete blood count (CBC) components (15).

Despite extensive research, no single blood, stool, or urine biomarker has been adopted in clinical practice for its predictive accuracy to diagnose neonatal sepsis and start antibiotic therapy (16). Physiologic sepsis biomarkers, such as abnormal vital sign patterns, can be detected using analysis of continuous vital sign data from standard bedside monitors. We previously used predictive analytics and a multicenter cohort of very low birth weight (VLBW, <1,500 g) preterm infants to develop and externally validate a cardiorespiratory sepsis risk model (17). The model, called Pulse Oximetry Warning Score (POWS), detects physiologic biomarkers of abnormal heart rate and SpO2 patterns that occur near the clinical diagnosis of late-onset sepsis and calculates the relative risk of blood culture-positive sepsis in the subsequent 24 h (17). POWS has yet to be implemented in clinical practice, so we sought to understand how inflammatory biomarkers will correlate and add to cardiorespiratory markers of sepsis. Combining information from hematologic and physiologic inflammatory biomarkers could improve the accuracy of sepsis diagnosis over using either alone.

While many prior studies have analyzed inflammatory biomarkers in premature infants at the time of suspected sepsis (10, 18–20), few have analyzed baseline levels or changes over time in patients who develop sepsis (21, 22). Inflammatory biomarkers may be less useful in premature infants with noninfectious conditions that lead to inflammation, including respiratory distress syndrome and chronic lung disease. To understand the potential utility of biomarkers in VLBW infants, more data are needed. In this study, we aim to identify inflammatory biomarkers that discriminate sepsis from baseline levels and non-sepsis conditions in VLBW infants. To do this, we measured multiple biomarkers in samples collected from remnant plasma, weekly when available and at the time of evaluation for late-onset sepsis or NEC.

2 Methods

2.1 Study design

This was a prospective, observational cohort study conducted at a single center Level IV academic NICU. The Institutional Review Board approved this study with consent from a parent (IRB #HSR20902). We prospectively enrolled VLBW infants within seven days of birth and collected clinical data and remnant plasma from blood drawn for suspected sepsis and for routine laboratory testing. Clinical data and culture results were recorded from the electronic health record. We classified samples according to the indication for the laboratory tests ordered and the final diagnosis of a workup for LOS or NEC (see "clinical definitions" section below).

2.2 Sample collection

The study protocol did not require any extra blood draws and did not influence the clinical team's decisions on ordering blood tests. Nurses collected blood samples in tubes according to the clinical laboratory guidelines for the laboratory test ordered. Blood for a complete blood count was collected in EDTA tubes, and blood for a basic metabolic panel or CRP was collected in sodium heparin tubes. These were the only clinical laboratory tests used to request remnant plasma for the study. The clinical lab stores samples at 4 degrees Celsius immediately after receipt, and the leftover blood from a clinical sample was requested by the study team. The clinical lab separated plasma from whole blood for samples collected in sodium heparin tubes, while the study team separated plasma from whole blood for samples collected in EDTA tubes by centrifugation (1,800 g × 5 min). We stored all remnant plasma samples with at least 250 microliters at -80° C until analysis.

2.3 Biomarker analysis

We selected subjects with at least two collected samples of adequate plasma volume ($\geq 250 \,\mu$ l) for biomarker analysis. The volume of blood obtained from the patient was not altered for the purpose of this study. Therefore, samples with insufficient remnant plasma were excluded from the study. The multiplex assay measured a panel of inflammatory biomarkers: interleukin (IL)-6, IL-8, IL-10, IL-18, interferon gamma inducible protein (IP)-10, tumor necrosis factor alpha (TNFα), procalcitonin (PCT), human growth factor (HGF), endothelin growth factor (EGF), soluble suppression of tumorigenicity 2 (sST-2), and IL-1 receptor antigen (IL1-ra). We selected these biomarkers based on their role in the inflammatory response to bacterial infections and the existing literature (12, 20, 23, 24). Analyte concentrations were quantified using a customized multiplex Luminex® magnetic bead-based antibody assay (R&D Systems, Minneapolis, USA). Fluorescence signals for each biomarker bead region were analyzed on a Luminex[®]200, a dual-laser flow-based detection instrument. Concentrations below the lowest standard were recorded as the value of the lower limit of detection for statistical analyses. We used a 10-fold dilution to analyze PCT, sST-2, and IL-1ra, and the remaining biomarkers did not require dilution. Results of CRP measurement obtained for clinical use were obtained from the electronic health record. Clinical guidelines at the time of the study recommended measuring CRP at the time of suspected infection and when determining duration of treatment, but the decision to order a CRP was left to the discretion of the clinical team.

2.4 Clinical definitions

Samples were classified as "routine" if they were obtained from blood drawn for standard laboratory monitoring, usually a basic metabolic panel, and not within 48 h of suspected infection. We classified samples obtained at the time of a blood culture using the diagnosis of the event according to the following definitions:

- A. Late-onset sepsis (LOS) (25): a positive blood culture obtained after 72 h of age and treated with at least five days of intravenous antibiotics. These were further categorized as sepsis due to Gram-positive (GP) or Gram-negative (GN) bacteremia according to the organism identified by blood culture.
- B. Necrotizing enterocolitis (NEC) (26): radiographic evidence of necrotizing enterocolitis and clinical illness with or without a positive blood culture.
- C. Clinical sepsis (CS): negative blood and urine cultures treated with ≥5 days of antibiotics for presumed infection due to clinical illness.
- D. Sepsis Ruled Out (SRO): a negative blood culture treated with <5 days of empiric antibiotics while awaiting culture results and monitoring symptoms, typically for 48 h.

For the analysis, results of samples classified as SRO and routine were grouped as "no sepsis".

2.5 Cardiorespiratory sepsis risk prediction

We previously developed a multivariable model to predict sepsis using continuous heart rate (HR) and oxygen saturation (SpO₂) data called the Pulse Oximetry Warning Score, or POWS (17). The POWS model calculates the mean, standard deviation, skewness, kurtosis, and cross-correlation of HR and SpO₂ every 10 min and uses logistic regression at each window to predict the relative risk of LOS with a positive blood culture in the next 24 h. We calculated hourly POWS values from continuous bedside monitoring data during the 12 h preceding blood cultures. POWS scores were calculated after discharge and thus did not influence decisions about clinical care. A heart rate characteristics (27, 28) monitoring system for sepsis risk prediction was in use during the study period, but was not routinely used in the decision to order a blood culture.

2.6 Statistical analyses

We compared distributions of inflammatory biomarkers and POWS data using Kruskal–Wallis analysis across the four diagnosis groups, followed by a pairwise Wilcoxon test with corrections for multiple testing. Hierarchical cluster analysis was performed on seven biomarkers in samples associated with a positive blood culture or NEC with no organism identified. Cytokine values were log transformed prior to clustering. The relationships between POWS and individual biomarker levels were assessed using univariate linear regression. The statistical significance of each model's coefficient was adjusted for repeated measures using the Huber-White method (29). A p-value <0.05 was considered statistically significant. The predictive performance of biomarkers and POWS were assessed individually or in combination using sensitivity, specificity at specific thresholds, and area under the receiver operator characteristics curve (AUC) using logistic regression to predict LOS or NEC. All statistical analysis was performed using RStudio (R Version 4.2.3, Vienna, Austria).

3 Results

3.1 Patients and samples

We enrolled 118 VLBW infants with parental consent. Of those enrolled, we had sufficient samples from 54 infants for analysis. The 54 infants included were 48% female, had a median gestational age of 26 weeks (IQR 25-27), and a median birth weight of 790 g (IQR 630-930). Compared with infants analyzed, infants excluded for insufficient samples or data were 43% female, had higher median gestational age (28 weeks, IQR 26-29) and birthweight (980 g, IQR 760-1,240). Table 1 shows clinical characteristics of infants included for sample analysis and compares those with and without LOS or NEC. In total, 188 plasma samples were analyzed, including 68 obtained near the time of blood cultures for suspected LOS or NEC and 120 at the time of routine blood sampling. The median number of samples analyzed per patient was 3 (range 2-6). Seven biomarkers (IP-10, IL-6, IL-10, IL-18, TNFa, IL-8, PCT) were analyzed at all times, while four (HGF, EGF, sST-2, IL-1ra) were successfully measured 80% of the time (152 samples in 45 patients).

Of the 67 blood cultures with study samples analyzed, 28 were diagnosed as LOS or NEC with bacteremia. Of these, there were 22 cases of Gram-positive (GP) bacteremia, 5 Gram-negative (GN) bacteremia, and 1 with NEC and GN bacteremia. Organisms in the positive blood cultures were coagulase-negative Staphylococcus (CONS) species (n = 19), *Escherichia coli* (n = 2), *Enterobacter* species (n = 1), Group B *Streptococcus* (n = 1), *Klebsiella* species (n = 5), and methicillin-susceptible *Staphylococcus aureus* (n = 2). There were 2 cases of NEC with negative blood cultures and 17 negative blood cultures diagnosed as CS. The remaining 141 samples came from blood drawn at times with NS, including routine samples and blood cultures diagnosed as SRO.

TABLE 1 Cohort characteristics overall and grouped by infants with or with late-onset sepsis (LOS) or necrotizing enterocolitis (NEC) diagnosis.

	LOS/NEC (<i>N</i> = 30)	No LOS/NEC (<i>N</i> = 24)	Overall (N = 54)
Gestational age (weeks), mean (SD)	25.8 (2.1)	26.2 (1.6)	25.9 (1.9)
Birth weight (grams), mean (SD)	780 (203)	831 (245)	803 (222)
Male, <i>n</i> (%)	19 (63%)	9 (38%)	28 (52%)
Race/ethnicity, n (%)			
Black	3 (10%)	4 (17%)	7 (13%)
Hispanic	3 (10%)	1 (4%)	4 (7%)
White	23 (77%)	18 (75%)	41 (76%)
Unknown/other	1 (3%)	1 (4%)	2 (4%)
Died, <i>n</i> (%)	5 (17%)	0 (0%)	5 (9%)

Values are presented as mean (standard deviation) or number (%).



as clinical sepsis (CS, n = 17), Gram-positive bacteremia (GP, n = 22), Gram-negative bacteremia or Necrotizing Enterocolitis (NEC) (GN, n = 8), or no sepsis (NS, n = 141). IL-6 (A), TNF-a (B), IL-8 (C), IL-10 (D), and sST-2 (E) were significantly higher in patients with GN sepsis as compared with NS, CS, and GP sepsis, as indicated by asterisks. Biomarker levels were not different when comparing NS, CS, and GP sepsis samples.

3.2 Biomarkers

Five of the eleven biomarkers (IL-6, TNF- α , IL-8, IL-10, and sST-2) were significantly higher in patients with GN sepsis or NEC than those with NS, CS, and GP sepsis (p < 0.05). Overall, biomarkers at CS or GP diagnosis were not significantly different from NS samples (Figure 1).

CRP was ordered by the clinical team at the time of 92 (49%) of the analyzed remnant plasma samples. CRP was significantly higher in patients with GN sepsis or NEC (median CRP 4.4 g/dl, IQR 2.1–11.0) compared to patients with no sepsis [median CRP 0.1 g/dl, IQR 0.1–0.4 (p < 0.05)] or clinical sepsis [median CRP 0.12 g/dl, IQR 0.1–1.2 (p < 0.05)]. There was no statistically significant difference in CRP for GN sepsis vs. GP sepsis cases [median CRP 2.7 g/dl, IQR 0.1–3.3 (p = 0.16)] or clinical sepsis [median CRP 0.12 g/dl, IQR 0.1–1.2 (p < 0.08)].

Hierarchical cluster analysis of cytokines from 30 LOS or NEC blood samples showed two distinct clusters of biomarker profiles (Figure 2). GN bacteremia or NEC was more prevalent in the cluster of samples with the highest cytokine levels. IL-6 above 20 pg/ml had the best overall test accuracy for diagnosing gramnegative sepsis or NEC with a specificity of 78%, negative predictive value of 100%, and sensitivity of 100%.

3.3 Association with POWS, a cardiorespiratory sepsis risk prediction model

Of the 188 samples, 187 had continuous heart rate and oxygen saturation data available around the time of sample collection to calculate POWS, a pulse oximetry warning score designed as a physiomarker for impending sepsis. POWS was significantly associated with the levels of 5 biomarkers (IL-8, PCT, HGF, sST-2, and IL1-ra) irrespective of the associated diagnosis (Figure 3, p < 0.05). The maximum POWS within the 12 h preceding the sample had an AUC to predict LOS (GN or GP) or NEC (with or without bacteremia) of 0.610. The AUC increased to 0.680 when the IL-6 level of the sample was added to the model.

4 Discussion

We assayed inflammatory biomarkers at the time of blood culture for suspected sepsis and routine laboratory testing in VLBW infants and found differences that distinguish late-onset Gram-negative sepsis and NEC from other diagnoses, but similar biomarker distributions obtained from samples collected at



baseline and at the time of suspected sepsis or Gram-positive sepsis. We also found correlations between inflammatory biomarkers and POWS, an algorithm that detects abnormal patterns of heart rate and oxygen saturation that we previously developed as a physiomarker of sepsis (17).

Our results confirm the findings in a previous study of VLBW infants that multiple biomarkers can discriminate between GN sepsis and GP sepsis at the time of positive blood culture (12). Since Gram-negative infections and NEC are known to carry higher morbidity and mortality, this finding might be a useful adjunct clinical decision support tool in helping pick the appropriate empiric antibiotic therapy and institute treatment early (30–32). The current study adds to our prior work (12) because we analyzed samples remote from suspected sepsis. We found no significant difference in biomarker levels in these samples compared to those obtained at the time of clinical sepsis or Gram-positive sepsis. Some VLBW preterm infants may have a chronic or subacute systemic

inflammatory response associated with lung disease or intracranial hemorrhage, which could confound sepsis prediction using hematologic biomarkers or cardiorespiratory physiomarkers (33–35).

Plasma cytokines and chemokines are early markers of immune activation in response to infection, and have been shown to rise in septic premature infants (22, 36, 37). Several studies have evaluated the utility of biomarkers for diagnosing sepsis by comparing biomarker levels in cases vs. controls (11). Others have compared cytokine profiles obtained at the time of suspected infection in VLBW infants when the blood culture returns positive vs. negative (38, 39). Instead, we assessed differences in inflammatory biomarkers measured at times when there was clinical suspicion for infection or sepsis and at times remote from infection. Kuster and colleagues also took the approach of measuring cytokines at baseline and at the time of suspected sepsis in VLBW infants and found IL-1ra and IL-6 to have high sensitivity and specificity in 21 cases of late-onset sepsis, even measured from samples taken the



day prior to blood culture (37). A longitudinal study of extremely low birth weight (<1,000 g) measured cytokines at days 1, 3, 14, and 21 after birth and found that IFN- γ , IL-10, IL-18, TGF- β , and TNF- α levels differed among infants who developed fungal or bacterial LOS compared with those who never developed sepsis (22). This study enrolled a large, multicenter cohort but did not evaluate differences relative to the timing of blood cultures. They found that infants with higher levels of immune regulatory cytokines relative to pro-inflammatory cytokines were associated with an increased risk of LOS at any time during the NICU course (21).

Of the biomarkers assayed in our study, IL-6 has been most consistently identified as a promising sepsis biomarker in previous studies, though with variable sensitivity and specificity (39). We found that IL-6 had a high specificity at a threshold set to detect 100% of Gram-negative sepsis and NEC cases but low predictive accuracy for Gram-positive cases, which were mostly due to CONS bacteremia. IL-6 has been shown to predict fatal or severe sepsis in premature infants (40), which occurs more often in Gram-negative than Gram-positive infections due to pathogen virulence factors (41). However, we also found elevated levels of IL-6 measured from samples remote from sepsis with blood draws for routine laboratory tests.

The inclusion of sST-2 as a sepsis biomarker was novel for this population as its utility has mainly been studied in adult populations. It is an IL-1 family receptor that binds IL-33 and has been implicated in diseases involving intestinal inflammation (42). Recent studies in adult patients demonstrate an association between elevated sST-2 and the severity of illness in *Clostridium difficile* colitis (43, 44). Our results indicate that this may also be a useful biomarker for diagnosing NEC and Gram-negative sepsis, where gastrointestinal dysfunction and bacterial translocation cause systemic inflammation.

Cytokines and chemokines such as IL-6, IL-1ra, and IL-8 have been demonstrated to have diagnostic utility as early sepsis markers (37, 45), while acute phase reactants such as CRP and PCT rise during the later phases of systemic inflammation (46). CRP and PCT are also examples of the few inflammatory biomarkers available in U.S. clinical laboratories, which may drive their clinical use despite evidence of low clinical utility (14). A recent meta-analysis showed overall low sensitivity and specificity of CRP for LOS diagnosis (47). We note several limitations of the study. First, the analysis was limited by the small sample size, where most late-onset sepsis events were due to CONS bacteremia, some of which may have represented contamination and not a true infection. Second, events diagnosed as clinical sepsis are heterogeneous in severity with subjective diagnostic criteria. We collected data on clinical characteristics, but not to the level of detail to account for concurrent inflammatory processes, such as lung disease, invasive mechanical ventilation, and minor procedures. Finally, the use of remnant plasma allowed us to enroll patients and collect samples without additional blood draws, but the volume of plasma available was small and therefore did not allow assays to be run in duplicate.

Analysis of plasma biomarkers and physiomarkers of LOS using POWS, a cardiorespiratory sepsis risk score, resulted in promising correlations that warrant further confirmation in larger studies. Continuous cardiorespiratory predictive monitoring used in conjunction with biomarker testing could prove useful, both for early initiation of antibiotics when the likelihood of sepsis is high and for sparing antibiotics when biomarkers and physiomarkers indicate low sepsis risk.

5 Conclusion

In conclusion, in a prospective cohort of VLBW infants, inflammatory biomarkers discriminated between late-onset sepsis due to gram-negative bacteremia or NEC and all other samples, including those collected at times with or without suspected sepsis. Biomarkers measured at the time of sepsis due to grampositive bacteremia did not differ from those measured at times without sepsis. Several inflammatory biomarkers, measured at baseline and at the time of suspected or confirmed sepsis, correlated with cardiorespiratory physiomarkers of sepsis.

Data availability statement

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation.

Ethics statement

The studies involving humans were approved by University of Virginia institutional review board. The studies were conducted in accordance with the local legislation and institutional

References

1. Shane AL, Sánchez PJ, Stoll BJ. Neonatal sepsis. Lancet. (2017) 390 (10104):1770-80. doi: 10.1016/S0140-6736(17)31002-4

2. Garg PM, Paschal JL, Ansari MAY, Block D, Inagaki K, Weitkamp J-H. Clinical impact of NEC-associated sepsis on outcomes in preterm infants. *Pediatr Res.* (2022) 92(6):1705–15. doi: 10.1038/s41390-022-02034-7

3. Wynn JL, Kelly MS, Benjamin DK, Clark RH, Greenberg R, Benjamin DK, et al. Timing of multiorgan dysfunction among hospitalized infants with fatal

requirements. Written informed consent for participation in this study was provided by the participants' legal guardians/next of kin.

Author contributions

RK: Conceptualization, Data curation, Formal Analysis, Writing – original draft, Writing – review & editing. SK: Data curation, Formal Analysis, Investigation, Methodology, Visualization, Writing – original draft, Writing – review & editing. AG: Data curation, Project administration, Investigation, Writing – review & editing. KF: Investigation, Methodology, Supervision, Writing – review & editing, Conceptualization. MA: Methodology, Validation, Writing – review & editing, Data curation, Formal Analysis, Investigation, Resources. WP: Supervision, Validation, Writing – review & editing, Resources. BS: Conceptualization, Data curation, Investigation, Methodology, Supervision, Validation, Visualization, Writing – original draft, Funding acquisition, Writing – review & editing.

Funding

The author(s) declare financial support was received for the research, authorship, and/or publication of this article.

We acknowledge the following grant for funding the work presented in this manuscript: K23 HD097254 (PI: B Sullivan); R01 AI124214, R01 AI152477 (PI: W. Petri)

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

fulminant sepsis. Am J Perinatol. (2017) 34(7):633-9. doi: 10.1055/s-0036-1597130

4. Adams-Chapman I, Stoll BJ. Neonatal infection and long-term neurodevelopmental outcome in the preterm infant. *Curr Opin Infect Dis.* (2006) 19 (3):290–7. doi: 10.1097/01.qco.0000224825.57976.87

5. Mukhopadhyay S, Puopolo KM, Hansen NI, Lorch SA, DeMauro SB, Greenberg RG, et al. Neurodevelopmental outcomes following neonatal late-onset sepsis and

blood culture-negative conditions. Arch Dis Child Fetal Neonatal Ed. (2021) 106 (5):467-73. doi: 10.1136/archdischild-2020-320664

6. Cai S, Thompson DK, Anderson PJ, Yang JY-M. Short- and long-term neurodevelopmental outcomes of very preterm infants with neonatal sepsis: a systematic review and meta-analysis. *Children (Basel)*. (2019) 6(12):19–21. doi: 10. 3390/children6120131

7. Shane AL, Stoll BJ. Neonatal sepsis: progress towards improved outcomes. J Infect. (2014) 68(Suppl 1):S24–32. doi: 10.1016/j.jinf.2013.09.011

8. Cantey JB. The spartacus problem: diagnostic inefficiency of neonatal sepsis. *Pediatrics.* (2019) 144(5):1. doi: 10.1542/peds.2019-2576

9. Chauhan N, Tiwari S, Jain U. Potential biomarkers for effective screening of neonatal sepsis infections: an overview. *Microb Pathog.* (2017) 107:234–42. doi: 10. 1016/j.micpath.2017.03.042

10. Ng S, Strunk T, Jiang P, Muk T, Sangild PT, Currie A. Precision medicine for neonatal sepsis. *Front Mol Biosci.* (2018) 5:70. doi: 10.3389/fmolb.2018.00070

11. Ye Q, Du L-Z, Shao W-X, Shang S-Q. Utility of cytokines to predict neonatal sepsis. *Pediatr Res.* (2017) 81(4):616–21. doi: 10.1038/pr.2016.267

12. Raynor LL, Saucerman JJ, Akinola MO, Lake DE, Moorman JR, Fairchild KD. Cytokine screening identifies NICU patients with gram-negative bacteremia. *Pediatr Res.* (2012) 71(3):261–6. doi: 10.1038/pr.2011.45

13. Cantey JB, Lee JH. Biomarkers for the diagnosis of neonatal sepsis. Clin Perinatol. (2021) 48(2):215–27. doi: 10.1016/j.clp.2021.03.012

14. Cantey JB, Bultmann CR. C-reactive protein testing in late-onset neonatal sepsis: hazardous waste. *JAMA Pediatr.* (2020) 174(3):235–6. doi: 10.1001/jamapediatrics. 2019.5684

15. Mussap M. Laboratory medicine in neonatal sepsis and inflammation. J Matern Fetal Neonatal Med. (2012) 25(Suppl 4):32–4. doi: 10.3109/14767058.2012.715000

16. Boscarino G, Migliorino R, Carbone G, Davino G, Dell'Orto VG, Perrone S, et al. Biomarkers of neonatal sepsis: where we are and where we are going. *Antibiotics (Basel.* (2023) 12(8):1. doi: 10.3390/antibiotics12081233

17. Kausch SL, Brandberg JG, Qiu J, Panda A, Binai A, Isler J, et al. Cardiorespiratory signature of neonatal sepsis: development and validation of prediction models in 3 NICUs. *Pediatr Res.* (2023) 93(7):1913–21. doi: 10.1038/ s41390-022-02444-7

18. Hibbert J, Strunk T, Simmer K, Richmond P, Burgner D, Currie A. Plasma cytokine profiles in very preterm infants with late-onset sepsis. *PLoS One*. (2020) 15 (5):e0232933. doi: 10.1371/journal.pone.0232933

19. Ruoss JL, Wynn JL. Biomarkers in the diagnosis of neonatal sepsis. In: Benitz WE, Smith PB, editors. *Infectious disease and pharmacology: Neonatology questions and controversies*. Orlando: Elsevier (2019). p. 103–12. Available at: https://www.sciencedirect.com/science/article/pii/B9780323543910000096 (Cited March 14, 2019).

20. Lam HS, Ng PC. Biochemical markers of neonatal sepsis. *Pathology.* (2008) 40 (2):141-8. doi: 10.1080/00313020701813735

21. Schelonka RL, Maheshwari A, Carlo WA, Taylor S, Hansen NI, Schendel DE, et al. T cell cytokines and the risk of blood stream infection in extremely low birth weight infants. *Cytokine*. (2011) 53(2):249–55. doi: 10.1016/j.cyto.2010.11.003

22. Sood BG, Shankaran S, Schelonka RL, Saha S, Benjamin DK, Sánchez PJ, et al. Cytokine profiles of preterm neonates with fungal and bacterial sepsis. *Pediatr Res.* (2012) 72(2):212–20. doi: 10.1038/pr.2012.56

23. Escobar GJ. Effect of the systemic inflammatory response on biochemical markers of neonatal bacterial infection: a fresh look at old confounders. *ClinChem*. (2003) 49(1):21–2. doi: 10.1373/49.1.21

24. Angurana SK, Bansal A, Muralidharan J, Aggarwal R, Singhi S. Cytokine levels in critically ill children with severe sepsis and their relation with the severity of illness and mortality. *J Intensive Care Med.* (2020) 36(5):885066620912989. doi: 10.1177/0885066620912989

25. Stoll BJ, Hansen N, Fanaroff AA, Wright LL, Carlo WA, Ehrenkranz RA, et al. Late-onset sepsis in very low birth weight neonates: the experience of the NICHD neonatal research network. *Pediatrics*. (2002) 110(2 Pt 1):285–91. doi: 10.1542/peds. 110.2.285

26. Walsh MC, Kliegman RM. Necrotizing enterocolitis: treatment based on staging criteria. *Pediatr Clin North Am.* (1986) 33(1):179–201. doi: 10.1016/S0031-3955(16) 34975-6

27. Sullivan BA, Fairchild KD. Predictive monitoring for sepsis and necrotizing enterocolitis to prevent shock. *Semin Fetal Neonatal Med.* (2015) 20(4):255-61. doi: 10.1016/j.siny.2015.03.006

28. Griffin MP, O'Shea TM, Bissonette EA, Harrell FE, Lake DE, Moorman JR. Abnormal heart rate characteristics preceding neonatal sepsis and sepsis-like illness. *Pediatr Res.* (2003) 53(6):920–6. doi: 10.1203/01.PDR.0000064904.05313.D2

29. Huber PJ. The behavior of maximum likelihood estimation under nonstandard conditions. In: Le Cam LM, Neyman J, editors. *Proceedings of the Fifth Berkeley Symposium on Mathematical Statistics and Probability*, Vol 1. Durham: Regents of the University of California (1967). p. 221–33.

30. Prieto CL, Colomer BF, Sastre JBL. Prognostic factors of mortality in very lowbirth-weight infants with neonatal sepsis of nosocomial origin. *Am J Perinatol.* (2013) 30(5):353–8. doi: 10.1055/s-0032-1324701

31. Goh GL, Lim CSE, Sultana R, De La Puerta R, Rajadurai VS, Yeo KT. Risk factors for mortality from late-onset sepsis among preterm very-low-birthweight infants: a single-center cohort study from Singapore. *Front Pediatr.* (2021) 9:801955. doi: 10.3389/fped.2021.801955

32. Makhoul IR, Sujov P, Smolkin T, Lusky A, Reichman B. Epidemiological, clinical, and microbiological characteristics of late-onset sepsis among very low birth weight infants in Israel: a national survey. *Pediatrics*. (2002) 109(1):34–9. doi: 10.1542/peds.109.1.34

33. Fairchild KD, Sinkin RA, Davalian F, Blackman AE, Swanson JR, Matsumoto JA, et al. Abnormal heart rate characteristics are associated with abnormal neuroimaging and outcomes in extremely low birth weight infants. *J Perinatol.* (2014) 34(5):375–9. doi: 10.1038/jp.2014.18

34. Collaco JM, McGrath-Morrow SA, Griffiths M, Chavez-Valdez R, Parkinson C, Zhu J, et al. Perinatal inflammatory biomarkers and respiratory disease in preterm infants. *J Pediatr.* (2022) 246:34–39.e3. doi: 10.1016/j.jpeds.2022.04.028

35. Bose CL, Dammann CEL, Laughon MM. Bronchopulmonary dysplasia and inflammatory biomarkers in the premature neonate. *Arch Dis Child Fetal Neonatal Ed.* (2008) 93(6):F455-61. doi: 10.1136/adc.2007.121327

36. Arnon S, Litmanovitz I. Diagnostic tests in neonatal sepsis. *CurrOpinInfect Dis*. (2008) 21(3):223-7. doi: 10.1097/QCO.0b013e3282fa15dd

37. Küster H, Weiss M, Willeitner AE, Detlefsen S, Jeremias I, Zbojan J, et al. Interleukin-1 receptor antagonist and interleukin-6 for early diagnosis of neonatal sepsis 2 days before clinical manifestation. *Lancet.* (1998) 352(9136):1271-7. doi: 10. 1016/S0140-6736(98)08148-3

38. Gonzalez BE, Mercado CK, Johnson L, Brodsky NL, Bhandari V. Early markers of late-onset sepsis in premature neonates: clinical, hematological and cytokine profile. *J Perinat Med.* (2003) 31(1):60–8. doi: 10.1515/JPM.2003.009

39. Ng PC, Li K, Wong RPO, Chui K, Wong E, Li G, et al. Proinflammatory and anti-inflammatory cytokine responses in preterm infants with systemic infections. *Arch Dis Child Fetal Neonatal Ed.* (2003) 88(3):F209–13. doi: 10.1136/fn.88.3.F209

40. Kurul Ş, Simons SHP, Ramakers CRB, De Rijke YB, Kornelisse RF, Reiss IKM, et al. Association of inflammatory biomarkers with subsequent clinical course in suspected late onset sepsis in preterm neonates. *Crit Care.* (2021) 25(1):12. doi: 10. 1186/s13054-020-03423-2

41. Shah J, Jefferies AL, Yoon EW, Lee SK, Shah PS, Network CN. Risk factors and outcomes of late-onset bacterial sepsis in preterm neonates born at <32 weeks' gestation. *Am J Perinatol.* (2015) 32(7):675–82. doi: 10.1055/s-0034-1393936

42. Nunes T, Bernardazzi C, de Souza HS. Interleukin-33 and inflammatory bowel diseases: lessons from human studies. *Mediators Inflamm.* (2014) 2014:423957. doi: 10.1155/2014/423957

43. Madden GR, Rigo I, Boone R, Abhyankar MM, Young MK, Basener W, et al. Novel biomarkers, including tcdB PCR cycle threshold, for predicting recurrent clostridioides difficile infection. *Infect Immun.* (2023) 91(4):e0009223. doi: 10.1128/ iai.00092-23

44. Abhyankar MM, Ma JZ, Scully KW, Nafziger AJ, Frisbee AL, Saleh MM, et al. Immune profiling to predict outcome of clostridioides difficile infection. *MBio.* (2020) 11(3):2–5. doi: 10.1128/mBio.00905-20

45. Ng PC, Ang IL, Chiu RWK, Li K, Lam HS, Wong RPO, et al. Host-response biomarkers for diagnosis of late-onset septicemia and necrotizing enterocolitis in preterm infants. *J Clin Invest.* (2010) 120(8):2989–3000. doi: 10.1172/JCI40196

46. Eschborn S, Weitkamp J-H. Procalcitonin versus C-reactive protein: review of kinetics and performance for diagnosis of neonatal sepsis. J Perinatol. (2019) 39 (7):893–903. doi: 10.1038/s41372-019-0363-4

47. Brown JVE, Meader N, Wright K, Cleminson J, McGuire W. Assessment of Creactive protein diagnostic test accuracy for late-onset infection in newborn infants: a systematic review and meta-analysis. *JAMA Pediatr.* (2020) 174(3):260–8. doi: 10. 1001/jamapediatrics.2019.5669