



Breaking the amyotrophic lateral sclerosis early diagnostic barrier: the promise of general markers

Yizhou Lu^{1†} , Lu He^{2†} , Huanyu Meng¹ , Sheng Chen^{1,3*} , Qinming Zhou^{1*} 

¹Department of Neurology, Ruijin Hospital Luwan Branch, Shanghai Jiao Tong University School of Medicine, Shanghai 200020, China

²Department of Neurology, Ruijin Hospital, Shanghai Jiao Tong University School of Medicine, Shanghai 200025, China

³Co-Innovation Center of Neuroregeneration, Nantong University, Nantong 226001, Jiangsu, China

[†]These authors contributed equally to this work.

***Correspondence:** Sheng Chen, mztcs@163.com; Qinming Zhou, zqmm2005@163.com. Department of Neurology, Ruijin Hospital, Shanghai Jiao Tong University School of Medicine, Shanghai 200025, China

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Abstract

Amyotrophic lateral sclerosis (ALS) is a severe neurodegenerative disease that is associated with selective and progressive loss of motor neurons. As a consequence, the symptoms of ALS are muscle cramps and weakness, and it eventually leads to death. The general markers for early diagnosis can assist ALS patients in receiving early intervention and prolonging their survival. Recently, some novel approaches or previously suggested methods have validated the potential for early diagnosis of ALS. The purpose of this review is to summarize the status of current general markers discovery and development for early diagnosis of ALS, including genes, proteins neuroimaging, neurophysiology, neuroulttrasound, and machine learning models. The main genetic markers evaluated are superoxide dismutase 1 (*SOD1*), chromosome 9 open reading frame 72 (*C9orf72*), transactivation-responsive DNA binding protein 43 (*TARDBP*), and fused in sarcoma (*FUS*) genes. Among proteins, neurofilament light chain is still the most established disease-specific adaptive change in ALS. The expression of chitinases, glial fibrillary acidic protein (*GFAP*), and inflammatory factors are changed in the early stage of ALS. Besides, more patient-friendly and accessible feature assays are explored by the development of neuroimaging, neurophysiology, and neuroulttrasound techniques. The novel disease-specific changes exhibited the promising potential for early diagnosis of ALS. All of these general markers still have limitations in the early diagnosis, therefore there is an urgent need for the validation and development of new disease-specific features for ALS.

Keywords

Amyotrophic lateral sclerosis, marker, genes, neurofilament, magnetic resonance imaging, positron emission tomography, neurophysiology, neuroulttrasound



Introduction

Amyotrophic lateral sclerosis (ALS) is a highly lethal disease characterized by selective and progressive loss of cortical, spinal, and bulbar motor neurons [1]. Initially presenting from focal muscle weakness involving upper or lower limbs, bulbar, or respiratory regions, ALS gradually spreads to global muscles, ultimately resulting in respiratory dysfunction [2]. Epidemiological studies in Europe have shown the global incidence of ALS is approximately 1–2.6 cases per 100,000 individuals annually, with a prevalence of around 6 cases per 100,000 [3, 4]. ALS most commonly occurs around the age of 60, affecting more males than females [4]. The median time from symptom onset to diagnosis is typically 14 months due to the complex heterogeneity. Fatality of ALS patients usually occurs within 3–5 years after diagnosis due to respiratory paralysis [3]. The etiology of ALS remains an unsolved mystery. However, it is currently believed that genetic, environmental, and pathological factors are all implicated.

Interestingly, there is an increasing body of epidemiological evidence linking environmental toxins with neurodegenerative diseases, including ALS, Alzheimer's disease (AD), and Parkinson's disease (PD) [5, 6]. Numerous environmental factors play roles in the pathogenesis of ALS, such as pesticides, electromagnetic fields, smoking, physical activity, body mass index, microbiota structure, cyanobacteria, and cyanotoxins. Recent studies have revealed the presence of the neurotoxin β -N-methylamino-L-alanine (L-BMAA) in the brain and cerebrospinal fluid (CSF) samples of AD and ALS patients [7]. L-BMAA was produced by cyanobacteria and algal species, indicating that exposure to cyanotoxins may contribute to the development of human neurodegenerative diseases [8]. Spencer et al. [9] first proposed a connection between the pathogenesis of ALS/parkinsonism-dementia complex (PDC) and the neurotoxin L-BMAA, which was produced by cyanobacteria of the genus *Nostoc* [9]. However, Montine et al. [10] assayed free L-BMAA in the brains of five control subjects and five patients with AD, who were from the US Pacific Northwest as well as Chamorros with and without PDC, but they detected no free L-BMAA in any of these samples. Cruz-Aguado et al. [11] also supported neither the causal role of L-BMAA in neurodegeneration nor the specific involvement of this amino acid in ALS/PDC. Hence, the involvement of L-BMAA in neurotoxicity and neurodegeneration remains controversial. Transactivation-responsive DNA binding protein 43 (TDP-43), a protein encoded by the TDP-43 gene (*TARDBP*), has been found to overexpress and aggregate both *in vitro* and *vivo* models, such as SH-SY5Y cell lines [12] and primary neurons in rats [13], mice [14], and zebrafish [15], when exposed to L-BMAA. These aberrant forms of TDP-43 are also present in patients with neurodegenerative diseases such as ALS and frontotemporal dementia (FTD) [16], suggesting their potential involvement in the pathogenesis of these neurodegenerative diseases. Recently, a proteomic study of murine neuroblastoma Neuro-2A (N2A) cells following low-dose exposure to saxitoxin revealed alterations in various proteins, which play key roles in the regulation of skeleton maintenance for cells, membrane potentials, mitochondrial functions, and cell apoptotic pathways [17]. It is also noteworthy that low doses of saxitoxins induce a decrease in voltage-dependent anion-selective channel 1 (VDAC1), a multifunctional protein expressed in the mitochondria and other cell compartments. VDAC1 regulates the main metabolic and energetic functions of the cell, such as Ca^{2+} homeostasis, oxidative stress, and mitochondria-mediated apoptosis [8]. VDAC1 serves as the main mitochondrial docking site of many misfolded proteins, such as amyloid β and Tau in AD, as well as several superoxide dismutase 1 (*SOD1*) mutants in ALS [18].

Although there is an absence of effective therapies for neurodegenerative diseases, early diagnosis, and intervention can improve the life quality of ALS patients [19]. However, most clinical trials for ALS treatment have strict inclusion criteria that require a short disease duration. This means that a diagnosis of ALS at an early stage would allow more patients to be enrolled in these trials. Unfortunately, the median time from symptom onset to diagnosis is typically 14 months, as neurologists often struggle with the uncertainty of the diagnosis and repeat investigations for confirmation. ALS is a highly complicated and heterogeneous disease, causing difficulty in early diagnosis. Currently, ALS diagnosis depends on the regional extent of clinical and electrophysiological signs, and many other diseases can mimic ALS in the early stages. No clear evidence indicates the involvement of upper motor neuron (UMN) and lower motor neuron (LMN) across multi-levels at an early stage. In particular, there are no sensitive UMN injury

identification techniques with atypical clinical signs. Thus, it is very challenging to diagnose ALS at an early stage.

A series of criteria have been designed over the years in consensus meetings, regarding the patterns of UMN and LMN signs required for diagnostic certainty, together with the absence of features suggestive of other neurological disorders. These clinical criteria were revised to define more precisely the neurophysiological criteria for LMN signs [20–22]. At all these consensus meetings it was recognized that early features of UMN disorder in ALS were difficult to elicit. To facilitate the early diagnosis of ALS, an international consensus group proposed the Gold Coast criteria, which established an increased diagnostic sensitivity when compared with the previous criteria [23]. However, effective markers can further facilitate the diagnosis, monitor the disease progression, and evaluate the therapeutic effects. In particular, the disease-specific features for early diagnosis assist ALS patients in receiving early intervention and prolonging their survival.

Nevertheless, there are no widely accepted characteristic changes in ALS diagnosis. With the advancement of diagnostic techniques, novel approaches demonstrate the potential for early ALS diagnosis. The purpose of this review is to summarize the current status of features discovery and development for the early diagnosis of ALS, including genes, proteins, neuroimaging, neurophysiology, neuroultrasound, and machine learning models (Figure 1). The main genetic markers evaluated for ALS diagnosis are *SOD1*, chromosome 9 open reading frame 72 (*C9orf72*), *TARDBP*, and fused in sarcoma (*FUS*) genes. Although neurofilament is the most established disease-specific protein, other proteins, such as chitinases, glial fibrillary acidic protein (GFAP), and inflammation cytokines, can play crucial roles in early diagnosis. Besides, the development of more patient-friendly and accessible feature assays has been facilitated by the advancement in neuroimaging techniques, such as magnetic resonance imaging (MRI), positron emission tomography (PET), and ultrasound. Furthermore, the use of machine learning models as an emerging tool to identify new characteristic changes in ALS will be discussed.

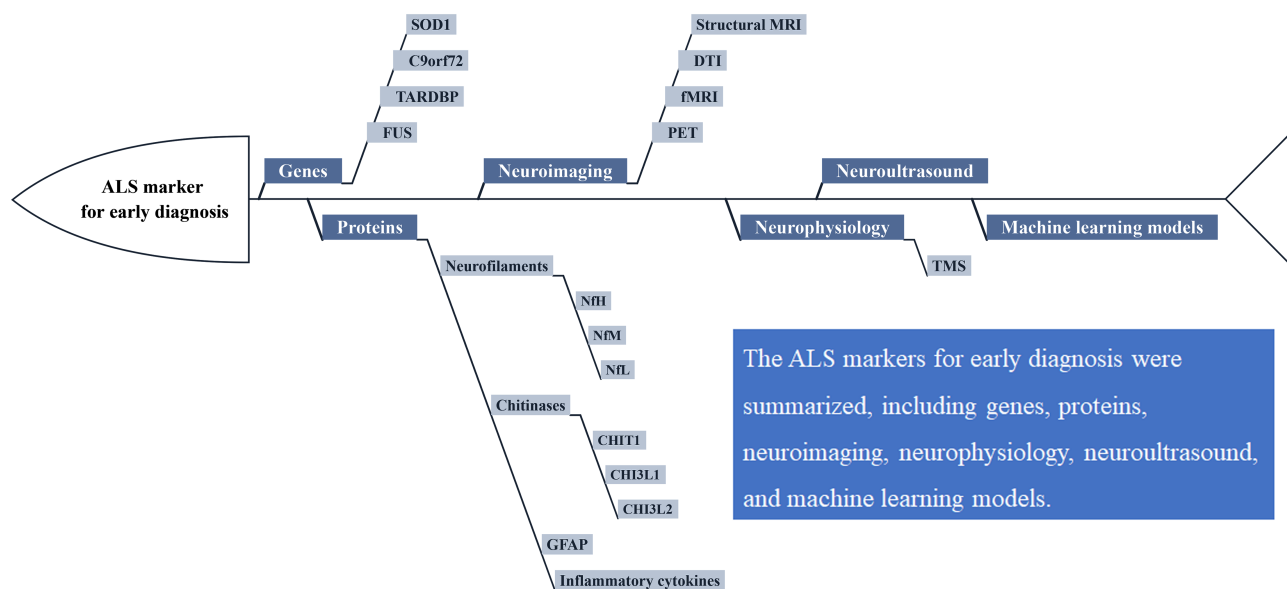


Figure 1. The promising markers for early diagnosis of ALS. DTI: diffusion tensor imaging; fMRI: functional MRI; TMS: transcranial magnetic stimulation; NfH: neurofilament heavy chain; NfM: neurofilament medium chain; NfL: neurofilament light chain; CHIT1: chitinase-3-like protein 1

Genes

About 10% of ALS cases are familial ALS (fALS), with frequent dominant traits and high penetrance [24]. The genetic cause has been elucidated in approximately 60–70% of fALS patients [25]. About 90% of individuals with ALS were classified as sporadic ALS (sALS) without a family history, but genetic factors have also been considered necessary. The estimated heritability of sALS is up to 50% [26]. The first ALS

gene, cytosolic *SOD1* was reported in 1993 [27, 28]. Since then, more than 50 potential ALS genes have been reported. However, validating the pathogenicity of a specific variant remains challenging. Among European populations, four genes account for nearly 70% of fALS, namely, *SOD1*, *C9orf72*, *TARDBP*, and *FUS* [29]. Typical mutants can become diagnostic features and therapeutic targets for ALS. Some loci are also associated with prognosis and can even reflect treatment efficacy. Early genetic screening will assist rapid diagnosis of asymptomatic and pre-symptomatic family members of fALS patients. Moreover, early genetic screening will provide crucial diagnostic evidence in suspected sALS patients. Since some genetic target therapies have been developed, they could control the disease early in patients with gene mutations.

SOD1

SOD1, the first ALS gene identified in 1993, causes autosomal dominant fALS [27, 28]. *SOD1* is a potent antioxidative enzyme that protects cells from oxidative stress damage. More than 170 mutations in *SOD1* have been found to cause ALS, with most occurring in fALS. Approximately 3% of mutations are in sALS [28]. The frequency of the *SOD1* gene mutations is about 12–23% in fALS and 0–7% in sALS [30]. The most common mutations of *SOD1* in ALS patients include D90A, A4V, H46R, and G93A. While G93A is relatively rare, it has been extensively studied [30]. *SOD1* mutations can cause ALS through toxic gain of function generated by aggregating misfolded *SOD1* proteins. High expression levels of human ALS-associated *SOD1* mutants (e.g., SOD1G93A, SOD1G37R, or SOD1G85R) could exhibit overt adult-onset motor neuron disease phenotypes mimicking the clinical ALS symptoms [28, 31, 32]. Thus, *SOD1* could be the features for early diagnosis and a potential therapeutic target. Therapeutic *SOD1* silencing using antisense oligonucleotides or microRNAs has been tested in the SOD1G93A mouse model and non-human primates [33, 34].

The first clinical study of intrathecal delivery of an antisense oligonucleotide, tofersen, was conducted in 2013 to assess its safety and tolerability. Tofersen is an antisense oligonucleotide that mediates the degradation of *SOD1* mRNA to reduce *SOD1* protein synthesis. Seven of eight patients in the placebo group had adverse events, compared to 20 of 24 in the ISIS 333611 (tofersen) group. No serious adverse events occurred in patients given ISIS 333611. Re-enrolment and re-treatment were also well tolerated, showing promising therapeutic potential and no significant safety concerns [35]. A phase 1–2 ascending-dose trial was conducted to evaluate tofersen in adults with ALS due to *SOD1* mutations [36]. In each dose cohort, participants were randomly assigned to receive five doses of tofersen or placebo, administered intrathecally for 12 weeks. Lumbar puncture—related adverse events were most common in participants. Some participants receiving tofersen experienced elevations in CSF white-cell count and protein levels. The change from the baseline CSF *SOD1* concentration to that at day 85 was also recorded. The difference between the tofersen groups and the placebo group was 2 percentage points [95% confidence interval (CI), –18 to 27] for the 20 mg dose, –25 percentage points (95% CI, –40 to –5) for the 40 mg dose, –19 percentage points (95% CI, –35 to 2) for the 60 mg dose, and –33 percentage points (95% CI, –47 to –16) for the 100 mg dose [36]. Recently, the result of trial III of tofersen has been revealed. Neurologic serious adverse events occurred in 7% of tofersen recipients. In persons with *SOD1* ALS, tofersen reduced concentrations of *SOD1* in CSF and of NfLs in plasma over 28 weeks [37].

C9orf72

C9orf72 gene mutation is the most common cause of ALS accounting for 20–50% of fALS cases and 5–20% of sALS cases [38]. Meanwhile, this mutation is also the leading genetic cause of FTD, causing approximately 10–30% of FTD cases [38]. The disease mechanism of mutant *C9orf72* has been proposed to include gain-of-function of protein, loss-of-function of protein [39], gain-of-function of toxic RNA [40, 41], and generating repeat-associated non-AUG initiation (RAN) translation products [42, 43]. RNA fluorescence *in-situ* hybridization (FISH) techniques have been used to detect *C9orf72* mutant foci by examining intranuclear GGGGCC repeat RNA. These foci can be detected in brain and peripheral cells, such as fibroblasts derived from a skin biopsy [40, 44] and blood leukocytes [45]. Thus, these foci can serve as a potential molecular change for diagnosis and evaluating treatment effects in clinical trials, eliminating the repeat expansion through therapeutic intervention.

A pathological mechanism of *C9orf72* gene expansion entails the translation of the expansion into dipeptide repeat proteins [46, 47]. This occurs through RAN translation of the hexanucleotide repeat, resulting in the formation of five dipeptide repeat proteins: glycine-alanine (GA), glycine-arginine (GR), proline-alanine (PA), proline-arginine (PR), and glycine-proline (GP) [47]. Production of poly-PR and poly-GR leads to neurotoxicity via impaired protein translation [48]. Poly-GP has attracted attention as a potential marker in *C9orf72* gene expansion carriers in ALS [49]. While asymptomatic mutation carriers have elevated levels of poly-GP in CSF and peripheral blood mononuclear cells, these levels are further increased in individuals with the disease [50].

TARDBP

TDP-43 encoded by *TARDBP* is a DNA/RNA-binding protein primarily located in the nucleus [51]. TDP-43 plays a crucial regulatory role in RNA metabolism, including mRNA splicing, translation, RNA transportation, and microRNA biogenesis [52–54]. The pathological findings of cytoplasmic aggregation and nucleus depletion of TDP-43 are present in over 97% of ALS and 45% of FTD patients [55]. *TARDBP* mutation occurs in approximately 3% of fALS cases and 1.2% of sALS cases [56]. The c.1144GA (p.A382T) is a founder mutation, highly prevalent in ALS cases from Sardinia, a genetically isolated island [57]. There are more frequent *TARDBP* mutations in European populations in sALS [56]. The causal mutations in *TARDBP* are relatively rare. However, the ubiquitous TDP-43 cytoplasmic inclusions and nucleus depletion described the causal role of the TDP-43 pathological axis in ALS.

FUS

Like *TARDBP*, DNA/RNA binding protein *FUS* mutations can cause ALS and a rare FTD. More than 50 *FUS* mutations have been reported in ALS. The *FUS* mutation frequency is about 3% in fALS and 0.4% in sALS [56], typically showing an autosomal dominant inheritance pattern. Similar to TDP-43 pathology, the *FUS* pathogenesis in ALS arises from the toxic *FUS* aggregation in the cytoplasm, and *FUS* deletion in the nucleus induces nuclear function loss. Expressing the mutations associated with rapidly progressive and juvenile-onset ALS *in vivo* at physiological levels can cause progressive and age-dependent motor neuron degeneration in mice [58].

Recently, Eleanor and Lou Gehrig ALS centers at Columbia University's Irving Medical Center initiated the first clinical trial targeting *FUS*, which is called Jacifusen. It is a multicenter phase 1–3 study, double-blinded, randomized, placebo-controlled of antisense oligonucleotides targeting the *FUS* mRNA and supported by ALS association and Project ALS. The primary outcome is to measure the change in ALS Functional Rating Scale (ALSFERS)-Revised (ALSFERS-R) and Ventilation Assistance-free survival (VAFS) at 505 days while the outcome is unknown yet [59].

Besides, other studies using genome editing with clustered regularly interspaced short palindromic repeats-associated protein 9 (CRISPR/Cas9) have been reported. Researchers use induced pluripotent stem cells (iPSCs) derived from ALS patients with *FUS* mutations to study *FUS* pathogenesis. Wang et al. [60] first revealed the CRISPR/Cas-9-mediated *FUS* (G1566A) correction. After that, Bhinge et al. [61] reported the CRISPR/Cas-9-mediated correction of *FUS* (H517Q) mutation, which showed that the abnormal activation of mitogen-activated protein kinase (MAPK) signaling is related to the pathogenesis process in ALS patients [61]. Furthermore, another study with CRISPR/Cas9-mediated *FUS* (R521H) correction proved that pathological axonal transport defects in motor neurons with *FUS* mutation could be rescued by gene correction [62]. CRISPR/Cas9 mediated gene editing may facilitate the development of new therapies in ALS even though more clinical studies are required.

Other genes

Other ALS-related genes include RNA binding proteins, such as TATA-box binding protein associated factor 15 (*TAF15*), Ewing sarcoma breakpoint region 1 (EWS) RNA binding protein 1 (*EWSR1*), ataxin-2 (*ATXN2*), and heterogeneous nuclear ribonucleoproteins (*hnRNPs*). Moreover, other genes also include mitochondria proteins, such as coiled-coil-helix-coiled-coil-helix domain containing 10 (*CHCHD10*), cytoskeleton-related

proteins, including tubulin alpha-4A (*TUBA4A*), and kinesin family member 5A (*KIF5A*), and several other newly-defined genes. However, mutation prevalence in these genes is relatively rare in ALS cases. Therefore, the most common genes (*SOD1*, *C9orf72*, *TARDBP*, and *FUS*) should be screened at the beginning.

Proteins

Neurofilaments

Neurofilaments are neuronal cytoskeletal proteins that are highly expressed in large caliber myelinated axons, functioning in axonal growth and maintenance. These consist of NfH, NfM, and NfL and α -internecine. A few studies have shown elevated levels of neurofilaments in CSF, plasma, and serum of ALS patients.

Currently, NfL is the most established disease-specific feature in ALS. The CSF NfL levels in ALS patients are significantly higher than in healthy controls and patients with FTD [63]. The CSF and blood NfL correlated with disease progression and survival time, indicating it could be a prognostic disease-specific marker [64, 65]. Additionally, CSF and blood NfL are altered in the early stage of the disease, transitioning from pre-symptomatic to symptomatic [66]. Benatar et al. [67] detected the serum and CSF NfL levels in ALS patients through the ALS-associated gene mutations [*SOD1*, *C9orf72*, *TARDBP*, *FUS*, valosin-containing protein (*VCP*), etc.]. The pre-symptomatic ALS patients were named the individual carriers of the ALS-associated gene mutations without clinical symptoms at the time of enrollment. In the longitudinal study, some pre-symptomatic patients converted to ALS patients with clinical symptoms. Both serum and CSF NfL levels were significantly higher in ALS patients than in healthy controls and pre-symptomatic ALS patients. In clinically converted ALS patients, NfL levels were found to be elevated 12 months before the onset of the earliest clinical symptoms. Moreover, different subtypes of ALS showed varying NfL levels, with bulbar-onset ALS patients having significantly higher plasma NfL levels than the spinal-onset ones [68].

In addition to NfL, NfH also showed diagnostic potential in ALS. Most studies have focused on phosphorylated NfH (pNfH) due to its phosphorylation. Previous studies have revealed elevated pNfH levels in serum, plasma, and CSF in ALS patients, increasing along with the disease progression [69]. Simultaneously, there were significantly increased NfH levels prior to symptom onset. However, serum and CSF pNfH were found to be less sensitive to pre-symptomatic ALS compared to NfL [70].

The increasing levels of neurofilament are observed in various neurological disorders due to axonal damage. This includes inflammatory, neurodegenerative, traumatic, and cerebrovascular diseases, which limits the diagnostic specificity of neurofilaments in ALS. However, neurofilaments remain the most promising and characteristic changes in ALS. They can be measured by easily acquiring serum/plasma, and the detecting techniques have favorable maturity, sensitivity, and reproducibility in assessing neurofilament levels.

Chitinases

The chitinases belong to the family 18 of glycosyl hydrolases (GH18), which can cleave chitin. The GH18 family is widely expressed across various organisms. Mammalian chitinases include the enzymatically active chitinases: CHIT1, acidic mammalian chitinase (AMCase), CHI3L1, CHI3L2 [71].

Several studies have revealed that ALS patients have significantly elevated CSF levels of CHIT1, CHI3L1, and CHI3L2 than healthy controls, mimics, and asymptomatic mutation carriers [72–74]. Among human chitinases, CHIT1 is the most abundant in monocytes and macrophages. Increased CHIT1 is a disease-specific change of microglia activation in the CSF of ALS patients. Some studies have indicated that CHIT1 levels in the CSF were elevated, distinguishing ALS from healthy controls, other neurodegenerative disease patients, and diseases mimicking ALS [73–76]. CHIT1 levels have been associated with disease progression and survival time in ALS patients, suggesting its potential as an early marker for the disease [74]. Based on these findings, Steinacker et al. [77] conducted a study investigating the diagnostic and prognostic potential of CHIT1 in 275 early symptomatic ALS patients from 8 European neurological centers. However, they observed that CHIT1 levels were elevated in both early and symptomatic ALS and did not correlate with the

rate of disease progression. CHIT1 could only predict disease progression in ALS patients with a short history and classified within low clinical certainty diagnostic categories. However, Steinacker et al. [77] revealed that the levels of three chitinases (CHIT1, CHI3L1, and CHI3L2) were also increased in other neurodegenerative conditions, with their discriminatory power outperformed by NfH and NfL [77]. Therefore, CSF chitinase proteins may have limited value as independent diagnostic and stratification index in ALS. However, they offer a window into non-autonomous mechanisms of motor neuron loss in ALS, particularly during the evaluation of responses to therapies targeting neuroinflammatory pathways [74].

GFAP

GFAP is released by astrocytes and acts as an intermediate filament protein during astrogliosis [78]. GFAP has been considered to be a potential feature for FTD based on the elevated GFAP levels in the CSF [79] and plasma in FTD patients [80]. In ALS patients and controls, serum GFAP similarly moderately correlated with age. Nevertheless, ALS patients were found to carry higher serum GFAP levels compared to controls [81]. GFAP is also elevated in CSF samples in the previous report [79]. However, as a measure of astrogliosis, GFAP is common in other neurodegenerative diseases, such as AD and dementia with Lewy bodies, making it impossible to be used as a specific index for ALS.

Inflammatory cytokines

The role of neuroinflammation in the pathogenesis of ALS patients has already been identified. A systematic review was conducted to study the status of proteins and anti-inflammatory cytokines in neurodegeneration diseases, including ALS [82]. Interleukin-2 (IL-2), IL-4, IL-5, IL-10, IL-17, and interferon-gamma (IFN- γ) remained unchanged, but tumor necrosis factor-alpha (TNF- α), TNF receptor 1, IL-6, IL-1 β , IL-8, and vascular endothelial growth factor (VEGF) levels were significantly elevated in the peripheral blood of ALS patients [83]. Granulocyte colony-stimulating factor, IL-2, IL-15, IL-17, monocyte chemoattractant protein-1, macrophage inflammatory protein (MIP)-1 α , TNF- α , and VEGF levels were significantly increased in the CSF of ALS patients, but granulocyte-macrophage colony-stimulating factor, IFN- γ , IL-4, IL-5, IL-6, IL-7, IL-8, IL-10, IL-12p70, IL-13, MIP-1 β , and regulated upon activation, normal T cell expressed, and presumably secreted (RANTES) levels were not significantly different in the CSF samples of ALS patients [84]. These results confirm the presence of inflammatory response in ALS, but these inflammatory cytokines currently cannot act as disease-specific changes due to the lack of specificity.

Neuroimaging

Structural MRI

Focal brain atrophy is a crucial feature in ALS patients and can be analyzed using structural MRI. An extended follow-up study was conducted in asymptomatic *C9orf72* carriers, which showed that cortical changes could be detected up to 20 years before the estimated symptom onset [85]. A large multi-center study in FTD patients carrying *C9orf72* mutation revealed that the insula and temporal lobe volume decreased 10 years before expected symptom onset [85]. In addition, another study demonstrated that pre-symptomatic *C9orf72* carriers had experienced a loss of white matter integrity in tracts connecting the frontal lobe, thalamic radiation, and tracts related to motor functioning compared to healthy controls [86]. However, previous studies did not have consistent results regarding brain atrophy in ALS patients, disputing the early diagnostic value of structural MRI in ALS.

DTI

An essential feature of ALS is the degenerating white matter fibers, particularly the corticospinal tracts and corpus callosum. DTI, a mathematical model depending on diffusion MRI, is used to evaluate the degeneration of the white matter bundle. Several studies have reported a decrease in fractional anisotropy and an increase in mean diffusivity in the corticospinal tracts [87]. Additionally, some studies have observed changes in white matter tracts using DTI in pre-symptomatic individuals carrying the *C9orf72* mutation, especially in those under the age of 40 [88, 89].

fMRI

Task-based and resting state fMRI is performed to understand the functional brain network. The pre-symptomatic *C9orf72* carriers had prominent intrinsic connectivity deficits within salience and medial pulvinar thalamus-seeded networks [90]. Another study revealed that pre-symptomatic *C9orf72* carriers possessed abnormal longitudinal trajectories of intra-network homogeneity in the somatomotor, dorsal attention, and default mode networks compared to healthy controls and symptomatic carriers [91].

PET

¹⁸F-fluorodeoxyglucose (FDG)-PET for assessing brain glucose metabolism indicated the diagnostic potential in early-stage ALS. Popuri et al. [92] observed that pre-symptomatic *C9orf72* carriers demonstrated changes in brain glucose metabolism up to 10 years before symptom onset and significant gray matter volume changes. Some studies generated diagnostic algorithms that discriminated ALS from controls [93–95]. The algorithms had favorable sensitivity, but low specificity in the multicenter validation. PET can analyze disease mechanisms using specific molecular tracers sensitive to early pathological changes, and detect UMN degeneration in suspected ALS patients. However, consistent diagnostic criteria for PET have not been reached. Large longitudinal studies are required to determine the application value of PET in distinguishing UMN dysfunction before clinical signs emerge.

Neurophysiology

Currently, cortical hyperexcitability could be an early and specific feature of ALS related to excitotoxicity from excessive glutamate receptor activity at the synaptic cleft [96]. Cortical hyperexcitability is potentially useful as an objective diagnostic marker in ALS, which can be captured using TMS. The involvement of UMN dysfunction is difficult to measure traditionally. The pathological spread could be measured using hyperexcitability as a surrogate by recording the response at the abductor pollicis brevis. Thus, TMS offered some diagnostic utility at an early stage. TMS detection revealed that the pre-symptomatic ALS patients had reduced short-interval intracortical inhibition and increased intracortical facilitation [97]. TMS showed a high sensitivity (73.21%) and specificity (80.88%) at early disease stages [98]. TMS could differentiate ALS from neuromuscular mimicking disorders [99, 100] and advance ALS diagnosis by eight months [98].

Neuroulttrasound

Fasciculations are acute denervation of motor neurons equivalent to fibrillations and positive sharp waves, essential for ALS diagnosis. Misawa et al. [101] observed that fasciculations were detected using ultrasound in the tongue (60%), biceps brachii (88%), and tibialis anterior muscles (83%). They were more frequently detected than by electromyogram (EMG) [101]. Combining ultrasound and EMG could be a quicker, more sensitive, and less invasive approach for diagnosing ALS. However, no accepted algorithm for combining ultrasound and EMG exists to investigate patients with suspected ALS.

Machine learning models

ALS is a complex syndrome with significant heterogeneity and diverse clinical presentations. One feature or technique cannot depict the complicated disease status. Recently some researchers fed a large amount of disease data into the artificial intelligence system to establish a machine-learning model for ALS diagnosis. Geevasinga et al. [102] developed a novel diagnostic score called the ALS diagnostic index (ALSDI). ALSDI exhibited more than 80% sensitivity, specificity, and diagnostic accuracy involving clinical, conventional neurophysiologic, and TMS measures. It reliably differentiated ALS from mimicking disorders early in the disease process. Tang et al. [103] employed model-based and model-free machine-learning methods to predict the ALS disease progression (changes within the ALSFRS) using an extensive ALS data archive of 8,000 patients with 3 million records and 200 clinical features tracked over 12 months. Their models showed 70% accuracy in predicting univariate clinical outcomes. Fukushima et al. [104] observed 15 muscles and recorded fasciculations using muscle ultrasonography in 100 ALS patients, with 50 early-stage

ALS patients within nine months. Then a machine learning-based model was established with eight muscles in the four body regions. The model showed a high sensitivity, specificity, and positive predictive value for early-stage ALS patients [104]. Therefore, machine learning models describe the usefulness of evaluating a combination of multiple pathways rather than having a single target.

Conclusions

It is still difficult to diagnose and manage ALS early due to the heterogeneous presentation, and the overlap of symptoms and signs with other illnesses. Several characteristic changes and techniques have been developed recently to aid early ALS diagnosis. The updated diagnostic criteria shortened the time from disease onset to diagnosis. Genetic markers provide a relatively definitive diagnosis of ALS but are limited to a small subgroup. NfL is still the most promising fluid disease-specific change but cannot discriminate ALS from other mimic diseases due to limited specificity. Similarly, chitinases had limited value in early diagnosis. Some novel neuroimaging, neurophysiological, and ultrasonic techniques have also been applied to early ALS diagnosis. In particular, TMS capturing cortical hyperexcitability can be complementary to traditional electromyography. Thus, the algorithm or models including multiple diagnostic factors may be a favorable choice due to the high heterogeneity of ALS. To expedite the early diagnosis of ALS, the simpler diagnostic criteria, genetic testing, and NfL measurement could be applied in the future. In addition, the novel techniques should be validated in a larger ALS population. The research efforts will improve the clinical outcomes of ALS patients.

Abbreviations

AD: Alzheimer's disease

ALS: amyotrophic lateral sclerosis

C9orf72: chromosome 9 open reading frame 72

CHI3L1: chitinase-3-like protein 1

CHIT1: chitotriosidase

CI: confidence interval

CRISPR/Cas9: clustered regularly interspaced short palindromic repeats-associated protein 9

CSF: cerebrospinal fluid

DTI: diffusion tensor imaging

EMG: electromyogram

fALS: familial amyotrophic lateral sclerosis

fMRI: functional magnetic resonance imaging

FTD: frontotemporal dementia

FUS: fused in sarcoma

GFAP: glial fibrillary acidic protein

GP: glycine-proline

IL-2: interleukin-2

L-BMAA: β -*N*-methylamino-*L*-alanine

LMN: lower motor neuron

MRI: magnetic resonance imaging

NfH: neurofilament heavy chain

NfL: neurofilament light chain
PDC: parkinsonism-dementia complex
PET: positron emission tomography
pNfH: phosphorylated neurofilament heavy chain
sALS: sporadic amyotrophic lateral sclerosis
SOD1: superoxide dismutase 1
TARDBP: transactivation-responsive DNA binding protein 43 gene
TDP-43: transactivation-responsive DNA binding protein 43
TMS: transcranial magnetic stimulation
TNF: tumor necrosis factor
UMN: upper motor neuron
VDAC1: voltage-dependent anion-selective channel 1

Declarations

Author contributions

YL, LH, and HM: Writing—original draft. SC: Conceptualization. QZ: Conceptualization, Writing—original draft. All authors read and approved the submitted version.

Conflicts of interest

The authors declare that they have no conflicts of interest.

Ethical approval

Not applicable.

Consent to participate

Not applicable.

Consent to publication

Not applicable.

Availability of data and materials

Not applicable.

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References

1. Zhou Q, He L, Hu J, Gao Y, Shen D, Ni Y, et al. Increased expression of coronin-1a in amyotrophic lateral sclerosis: a potential diagnostic biomarker and therapeutic target. *Front Med.* 2022;16:723–35.

2. Zhou QM, Zhang JJ, Li S, Chen S, Le WD. *n*-butylidenephthalide treatment prolongs life span and attenuates motor neuron loss in SOD1^{G93A} mouse model of amyotrophic lateral sclerosis. *CNS Neurosci Ther.* 2017;23:375–85.
3. Talbott EO, Malek AM, Lacomis D. The epidemiology of amyotrophic lateral sclerosis. In: Aminoff MJ, Boller F, Swaab DF, editors. *Handbook of Clinical Neurology.* Amsterdam: Elsevier; 2016. pp. 225–38.
4. van Es MA, Hardiman O, Chio A, Al-Chalabi A, Pasterkamp RJ, Veldink JH, et al. Amyotrophic lateral sclerosis. *Lancet.* 2017;390:2084–98.
5. Spencer PS, Palmer VS, Kisby GE. Cycad β -*N*-methylamino-L-alanine (BMAA), methylazoxymethanol, genotoxicity, and neurodegeneration. *Toxicol.* 2018;155:49–50.
6. Cox PA, Kostrzewa RM, Guillemin GJ. BMAA and neurodegenerative illness. *Neurotox Res.* 2018;33:178–83.
7. Pablo J, Banack SA, Cox PA, Johnson TE, Papapetropoulos S, Bradley WG, et al. Cyanobacterial neurotoxin BMAA in ALS and Alzheimer's disease. *Acta Neurol Scand.* 2009;120:216–25.
8. Ra D, Sa B, Si B, Js M, Sj M, DA D, et al. Is exposure to BMAA a risk factor for neurodegenerative diseases? A response to a critical review of the BMAA hypothesis. *Neurotox Res.* 2021;39:81–106.
9. Spencer PS, Nunn PB, Hugon J, Ludolph AC, Ross SM, Roy DN, et al. Guam amyotrophic lateral sclerosis-parkinsonism-dementia linked to a plant excitant neurotoxin. *Science.* 1987;237:517–22.
10. Montine TJ, Li K, Perl DP, Galasko D. Lack of β -methylamino-l-alanine in brain from controls, AD, or Chamorro with PDC. *Neurology.* 2005;65:768–9.
11. Cruz-Aguado R, Winkler D, Shaw CA. Lack of behavioral and neuropathological effects of dietary β -methylamino-L-alanine (BMAA) in mice. *Pharmacol Biochem Behav.* 2006;84:294–9.
12. de Munck E, Muñoz-Sáez E, Miguel BG, Solas MT, Ojeda I, Martínez A, et al. β -*N*-methylamino-L-alanine causes neurological and pathological phenotypes mimicking Amyotrophic Lateral Sclerosis (ALS): the first step towards an experimental model for sporadic ALS. *Environ Toxicol Pharmacol.* 2013;36:243–55.
13. Scott L, Downing T. Dose-dependent adult neurodegeneration in a rat model after neonatal exposure to β -*N*-methylamino-L-alanine. *Neurotox Res.* 2019;35:711–23.
14. Yin HZ, Yu S, Hsu CI, Liu J, Acab A, Wu R, et al. Intrathecal infusion of BMAA induces selective motor neuron damage and astrogliosis in the ventral horn of the spinal cord. *Exp Neurol.* 2014;261:1–9.
15. Martin RM, Bereman MS, Marsden KC. BMAA and MCLR interact to modulate behavior and exacerbate molecular changes related to neurodegeneration in larval zebrafish. *Toxicol Sci.* 2021;179:251–61.
16. Sini P, Dang TBC, Fais M, Galioto M, Padedda BM, Lugliè A, et al. Cyanobacteria, cyanotoxins, and neurodegenerative diseases: *dangerous liaisons.* *Int J Mol Sci.* 2021;22:8726.
17. Berntzon L, Ronnevi LO, Bergman B, Eriksson J. Detection of BMAA in the human central nervous system. *Neuroscience.* 2015;292:137–47.
18. Brooks BW, Lazorchak JM, Howard MD, Johnson MV, Morton SL, Perkins DA, et al. Are harmful algal blooms becoming the greatest inland water quality threat to public health and aquatic ecosystems? *Environ Toxicol Chem.* 2016;35:6–13.
19. Xu X, Shen D, Gao Y, Zhou Q, Ni Y, Meng H, et al. A perspective on therapies for amyotrophic lateral sclerosis: Can disease progression be curbed? *Transl Neurodegener.* 2021;10:29.
20. Brooks BR, Miller RG, Swash M, Munsat TL; World Federation of Neurology Research Group on Motor Neuron Diseases. El Escorial revisited: revised criteria for the diagnosis of amyotrophic lateral sclerosis. *Amyotroph Lateral Scler Other Motor Neuron Disord.* 2000;1:293–9.
21. Feldman EL, Goutman SA, Petri S, Mazzini L, Savelieff MG, Shaw PJ, et al. Amyotrophic lateral sclerosis. *Lancet.* 2022;400:1363–80.
22. Costa J, Swash M, de Carvalho M. Awaji criteria for the diagnosis of amyotrophic lateral sclerosis: a systematic review. *Arch Neurol.* 2012;69:1410–6.

23. Hannaford A, Pavey N, van den Bos M, Geevasinga N, Menon P, Shefner JM, et al. Diagnostic utility of Gold Coast criteria in amyotrophic lateral sclerosis. *Ann Neurol*. 2021;89:979–86.
24. Taylor JP, Brown RH Jr, Cleveland DW. Decoding ALS: from genes to mechanism. *Nature*. 2016;539:197–206.
25. Mead RJ, Shan N, Reiser HJ, Marshall F, Shaw PJ. Amyotrophic lateral sclerosis: a neurodegenerative disorder poised for successful therapeutic translation. *Nat Rev Drug Discov*. 2023;22:185–212.
26. van Rheenen W, van der Spek RAA, Bakker MK, van Vugt JJFA, Hop PJ, Zwamborn RAJ, et al. Common and rare variant association analyses in amyotrophic lateral sclerosis identify 15 risk loci with distinct genetic architectures and neuron-specific biology. *Nat Genet*. 2021;53:1636–48.
27. Deng HX, Hentati A, Tainer JA, Iqbal Z, Cayabyab A, Hung WY, et al. Amyotrophic lateral sclerosis and structural defects in Cu,Zn superoxide dismutase. *Science*. 1993;261:1047–51.
28. Rosen DR, Siddique T, Patterson D, Figlewicz DA, Sapp P, Hentati A, et al. Mutations in Cu/Zn superoxide dismutase gene are associated with familial amyotrophic lateral sclerosis. *Nature*. 1993;362:59–62.
29. Hardiman O, Al-Chalabi A, Chio A, Corr EM, Logroscino G, Robberecht W, et al. Amyotrophic lateral sclerosis. *Nat Rev Dis Primers*. 2017;3:17071.
30. Kaur SJ, McKeown SR, Rashid S. Mutant SOD1 mediated pathogenesis of Amyotrophic Lateral Sclerosis. *Gene*. 2016;577:109–18.
31. Gurney ME, Pu H, Chiu AY, Dal Canto MC, Polchow CY, Alexander DD, et al. Motor neuron degeneration in mice that express a human Cu,Zn superoxide dismutase mutation. *Science*. 1994;264:1772–5.
32. Wong PC, Pardo CA, Borchelt DR, Lee MK, Copeland NG, Jenkins NA, et al. An adverse property of a familial ALS-linked SOD1 mutation causes motor neuron disease characterized by vacuolar degeneration of mitochondria. *Neuron*. 1995;14:1105–16.
33. Smith RA, Miller TM, Yamanaka K, Monia BP, Condon TP, Hung G, et al. Antisense oligonucleotide therapy for neurodegenerative disease. *J Clin Invest*. 2006;116:2290–6.
34. Borel F, Gernoux G, Sun H, Stock R, Blackwood M, Brown RH Jr, et al. Safe and effective superoxide dismutase 1 silencing using artificial microRNA in macaques. *Sci Transl Med*. 2018;10:eaau6414.
35. Miller TM, Pestronk A, David W, Rothstein J, Simpson E, Appel SH, et al. An antisense oligonucleotide against *SOD1* delivered intrathecally for patients with *SOD1* familial amyotrophic lateral sclerosis: a phase 1, randomised, first-in-man study. *Lancet Neurol*. 2013;12:435–42.
36. Miller T, Cudkowicz M, Shaw PJ, Andersen PM, Atassi N, Bucelli RC, et al. Phase 1-2 trial of antisense oligonucleotide tofersen for SOD1 ALS. *N Engl J Med*. 2020;383:109–19.
37. Miller TM, Cudkowicz ME, Genge A, Shaw PJ, Sobue G, Bucelli RC, et al.; VALOR and OLE Working Group. Trial of antisense oligonucleotide tofersen for SOD1 ALS. *N Engl J Med*. 2022;387:1099–110.
38. Mendez EF, Sattler R. Biomarker development for C9orf72 repeat expansion in ALS. *Brain Res*. 2015;1607:26–35.
39. Ciura S, Lattante S, Le Ber I, Latouche M, Tostivint H, Brice A, et al. Loss of function of C9orf72 causes motor deficits in a zebrafish model of amyotrophic lateral sclerosis. *Ann Neurol*. 2013;74:180–7.
40. Almeida S, Gascon E, Tran H, Chou HJ, Gendron TF, Degroot S, et al. Modeling key pathological features of frontotemporal dementia with *C9ORF72* repeat expansion in iPSC-derived human neurons. *Acta Neuropathol*. 2013;126:385–99.
41. Donnelly CJ, Zhang PW, Pham JT, Haeusler AR, Mistry NA, Vidensky S, et al. RNA toxicity from the ALS/FTD *C9ORF72* expansion is mitigated by antisense intervention. *Neuron*. 2013;80:415–28.
42. Gendron TF, Belzil VV, Zhang YJ, Petrucelli L. Mechanisms of toxicity in C9FTLD/ALS. *Acta Neuropathol*. 2014;127:359–76.

43. Ash PE, Bieniek KF, Gendron TF, Caulfield T, Lin WL, DeJesus-Hernandez M, et al. Unconventional translation of *C9ORF72* GGGGCC expansion generates insoluble polypeptides specific to c9FTD/ALS. *Neuron*. 2013;77:639–46.
44. Lagier-Tourenne C, Baughn M, Rigo F, Sun S, Liu P, Li HR, et al. Targeted degradation of sense and antisense *C9orf72* RNA foci as therapy for ALS and frontotemporal degeneration. *Proc Natl Acad Sci U S A*. 2013;110:E4530–9.
45. Zu T, Liu Y, Bañez-Coronel M, Reid T, Pletnikova O, Lewis J, et al. RAN proteins and RNA foci from antisense transcripts in *C9ORF72* ALS and frontotemporal dementia. *Proc Natl Acad Sci U S A*. 2013;110:E4968–77.
46. Shi Y, Lin S, Staats KA, Li Y, Chang WH, Hung ST, et al. Haploinsufficiency leads to neurodegeneration in *C9ORF72* ALS/FTD human induced motor neurons. *Nat Med*. 2018;24:313–25.
47. Kumar V, Hasan GM, Hassan MI. Unraveling the role of RNA mediated toxicity of *C9orf72* repeats in C9-FTD/ALS. *Front Neurosci*. 2017;11:711.
48. Kanekura K, Yagi T, Cammack AJ, Mahadevan J, Kuroda M, Harms MB, et al. Poly-dipeptides encoded by the *C9ORF72* repeats block global protein translation. *Hum Mol Genet*. 2016;25:1803–13.
49. Gendron TF, Chew J, Stankowski JN, Hayes LR, Zhang YJ, Prudencio M, et al. Poly(GP) proteins are a useful pharmacodynamic marker for *C9ORF72*-associated amyotrophic lateral sclerosis. *Sci Transl Med*. 2017;9:eaai7866.
50. Meeter LHH, Gendron TF, Sias AC, Jiskoot LC, Russo SP, Donker Kaat L, et al. Poly(GP), neurofilament and grey matter deficits in *C9orf72* expansion carriers. *Ann Clin Transl Neurol*. 2018;5:583–97.
51. Tollervey JR, Curk T, Rogelj B, Briese M, Cereda M, Kayikci M, et al. Characterizing the RNA targets and position-dependent splicing regulation by TDP-43. *Nat Neurosci*. 2011;14:452–8.
52. Deshaies JE, Shkreta L, Moszczynski AJ, Sidibé H, Semmler S, Fouillen A, et al. TDP-43 regulates the alternative splicing of hnRNP A1 to yield an aggregation-prone variant in amyotrophic lateral sclerosis. *Brain*. 2018;141:1320–33.
53. Neelagandan N, Gonnella G, Dang S, Janiesch PC, Miller KK, Küchler K, et al. TDP-43 enhances translation of specific mRNAs linked to neurodegenerative disease. *Nucleic Acids Res*. 2019;47:341–61.
54. Kawahara Y, Mieda-Sato A. TDP-43 promotes microRNA biogenesis as a component of the Drosha and Dicer complexes. *Proc Natl Acad Sci U S A*. 2012;109:3347–52.
55. Kim G, Gautier O, Tassoni-Tsuchida E, Ma XR, Gitler AD. ALS genetics: gains, losses, and implications for future therapies. *Neuron*. 2020;108:822–42.
56. Zou ZY, Zhou ZR, Che CH, Liu CY, He RL, Huang HP. Genetic epidemiology of amyotrophic lateral sclerosis: a systematic review and meta-analysis. *J Neurol Neurosurg Psychiatry*. 2017;88:540–9.
57. Chiò A, Borghero G, Pugliatti M, Ticca A, Calvo A, Moglia C, et al.; Italian Amyotrophic Lateral Sclerosis Genetic (ITALSGEN) Consortium. Large proportion of amyotrophic lateral sclerosis cases in Sardinia due to a single founder mutation of the TARDBP gene. *Arch Neurol*. 2011;68:594–8.
58. Korobeynikov VA, Lyashchenko AK, Blanco-Redondo B, Jafar-Nejad P, Shneider NA. Antisense oligonucleotide silencing of FUS expression as a therapeutic approach in amyotrophic lateral sclerosis. *Nat Med*. 2022;28:104–16.
59. Fang T, Je G, Pacut P, Keyhanian K, Gao J, Ghasemi M. Gene therapy in amyotrophic lateral sclerosis. *Cells*. 2022;11:2066.
60. Wang L, Yi F, Fu L, Yang J, Wang S, Wang Z, et al. CRISPR/Cas9-mediated targeted gene correction in amyotrophic lateral sclerosis patient iPSCs. *Protein Cell*. 2017;8:365–78.
61. Bhingre A, Namboori SC, Zhang X, VanDongen AMJ, Stanton LW. Genetic correction of SOD1 mutant iPSCs reveals ERK and JNK activated AP1 as a driver of neurodegeneration in amyotrophic lateral sclerosis. *Stem Cell Reports*. 2017;8:856–69.

62. Guo W, Naujock M, Fumagalli L, Vandoorne T, Baatsen P, Boon R, et al. HDAC6 inhibition reverses axonal transport defects in motor neurons derived from FUS-ALS patients. *Nat Commun.* 2017;8:861.
63. Gaetani L, Blennow K, Calabresi P, Di Filippo M, Parnetti L, Zetterberg H. Neurofilament light chain as a biomarker in neurological disorders. *J Neurol Neurosurg Psychiatry.* 2019;90:870–81.
64. Skillbäck T, Mattsson N, Blennow K, Zetterberg H. Cerebrospinal fluid neurofilament light concentration in motor neuron disease and frontotemporal dementia predicts survival. *Amyotroph Lateral Scler Frontotemporal Degener.* 2017;18:397–403.
65. Brodovitch A, Boucraut J, Delmont E, Parlanti A, Grapperon AM, Attarian S, et al. Combination of serum and CSF neurofilament-light and neuroinflammatory biomarkers to evaluate ALS. *Sci Rep.* 2021;11:703.
66. van der Ende EL, Bron EE, Poos JM, Jiskoot LC, Panman JL, Papma JM, et al.; GENFI consortium. A data-driven disease progression model of fluid biomarkers in genetic frontotemporal dementia. *Brain.* 2022;145:1805–17.
67. Benatar M, Wu J, Andersen PM, Lombardi V, Malaspina A. Neurofilament light: a candidate biomarker of presymptomatic amyotrophic lateral sclerosis and phenoconversion. *Ann Neurol.* 2018;84:130–9.
68. Behzadi A, Pujol-Calderón F, Tjust AE, Wuolikainen A, Höglund K, Forsberg K, et al. Neurofilaments can differentiate ALS subgroups and ALS from common diagnostic mimics. *Sci Rep.* 2021;11:22128.
69. Boylan KB, Glass JD, Crook JE, Yang C, Thomas CS, Desaro P, et al. Phosphorylated neurofilament heavy subunit (pNF-H) in peripheral blood and CSF as a potential prognostic biomarker in amyotrophic lateral sclerosis. *J Neurol Neurosurg Psychiatry.* 2013;84:467–72.
70. Benatar M, Wu J, Lombardi V, Jeromin A, Bowser R, Andersen PM, et al. Neurofilaments in pre-symptomatic ALS and the impact of genotype. *Amyotroph Lateral Scler Frontotemporal Degener.* 2019;20:538–48.
71. Gaur N, Perner C, Witte OW, Grosskreutz J. The chitinases as biomarkers for amyotrophic lateral sclerosis: signals from the CNS and beyond. *Front Neurol.* 2020;11:377.
72. Varghese AM, Sharma A, Mishra P, Vijayalakshmi K, Harsha HC, Sathyaprabha TN, et al. Chitotriosidase - a putative biomarker for sporadic amyotrophic lateral sclerosis. *Clin Proteomics.* 2013;10:19.
73. Oeckl P, Weydt P, Steinacker P, Anderl-Straub S, Nordin F, Volk AE, et al.; German Consortium for Frontotemporal Lobar Degeneration. Different neuroinflammatory profile in amyotrophic lateral sclerosis and frontotemporal dementia is linked to the clinical phase. *J Neurol Neurosurg Psychiatry.* 2019;90:4–10.
74. Thompson AG, Gray E, Bampton A, Raciborska D, Talbot K, Turner MR. CSF chitinase proteins in amyotrophic lateral sclerosis. *J Neurol Neurosurg Psychiatry.* 2019;90:1215–20.
75. Gille B, De Schaepdryver M, Dedeene L, Goossens J, Claeys KG, Van Den Bosch L, et al. Inflammatory markers in cerebrospinal fluid: independent prognostic biomarkers in amyotrophic lateral sclerosis? *J Neurol Neurosurg Psychiatry.* 2019;90:1338–46.
76. Steinacker P, Verde F, Fang L, Feneberg E, Oeckl P, Roeber S, et al.; FTLDC study group. Chitotriosidase (CHIT1) is increased in microglia and macrophages in spinal cord of amyotrophic lateral sclerosis and cerebrospinal fluid levels correlate with disease severity and progression. *J Neurol Neurosurg Psychiatry.* 2018;89:239–47.
77. Steinacker P, Feneberg E, Halbgebauer S, Witzel S, Verde F, Oeckl P, et al. Chitotriosidase as biomarker for early stage amyotrophic lateral sclerosis: a multicenter study. *Amyotroph Lateral Scler Frontotemporal Degener.* 2021;22:276–86.
78. Middeldorp J, Hol EM. GFAP in health and disease. *Prog Neurobiol.* 2011;93:421–43.

79. Ishiki A, Kamada M, Kawamura Y, Terao C, Shimoda F, Tomita N, et al. Glial fibrillar acidic protein in the cerebrospinal fluid of Alzheimer's disease, dementia with Lewy bodies, and frontotemporal lobar degeneration. *J Neurochem*. 2016;136:258–61.
80. Heller C, Foiani MS, Moore K, Convery R, Bocchetta M, Neason M, et al.; GENFI. Plasma glial fibrillary acidic protein is raised in progranulin-associated frontotemporal dementia. *J Neurol Neurosurg Psychiatry*. 2020;91:263–70.
81. Verde F, Milone I, Maranzano A, Colombo E, Torre S, Solca F, et al. Serum levels of glial fibrillary acidic protein in patients with amyotrophic lateral sclerosis. *Ann Clin Transl Neurol*. 2023;10:118–29.
82. Tanaka M, Toldi J, Vécsei L. Exploring the etiological links behind neurodegenerative diseases: inflammatory cytokines and bioactive kynurenines. *Int J Mol Sci*. 2020;21:2431.
83. Hu Y, Cao C, Qin XY, Yu Y, Yuan J, Zhao Y, et al. Increased peripheral blood inflammatory cytokine levels in amyotrophic lateral sclerosis: a meta-analysis study. *Sci Rep*. 2017;7:9094.
84. Van Everbroeck B, Dewulf E, Pals P, Lübke U, Martin JJ, Cras P. The role of cytokines, astrocytes, microglia and apoptosis in Creutzfeldt-Jakob disease. *Neurobiol Aging*. 2002;23:59–64.
85. Rohrer JD, Nicholas JM, Cash DM, van Swieten J, Dopfer E, Jiskoot L, et al. Presymptomatic cognitive and neuroanatomical changes in genetic frontotemporal dementia in the Genetic Frontotemporal dementia Initiative (GENFI) study: a cross-sectional analysis. *Lancet Neurol*. 2015;14:253–62.
86. Pappa JM, Jiskoot LC, Panman JL, Dopfer EG, den Heijer T, Donker Kaat L, et al. Cognition and gray and white matter characteristics of presymptomatic *C9orf72* repeat expansion. *Neurology*. 2017;89:1256–64.
87. Chiò A, Pagani M, Agosta F, Calvo A, Cistaro A, Filippi M. Neuroimaging in amyotrophic lateral sclerosis: insights into structural and functional changes. *Lancet Neurol*. 2014;13:1228–40.
88. Bertrand A, Wen J, Rinaldi D, Houot M, Sayah S, Camuzat A, et al.; Predict to Prevent Frontotemporal Lobar Degeneration and Amyotrophic Lateral Sclerosis (PREV-DEMALS) Study Group. Early cognitive, structural, and microstructural changes in presymptomatic *C9orf72* carriers younger than 40 years. *JAMA Neurol*. 2018;75:236–45.
89. Wen J, Zhang H, Alexander DC, Durrleman S, Routier A, Rinaldi D, et al.; Predict to Prevent Frontotemporal Lobar Degeneration and Amyotrophic Lateral Sclerosis (PREV-DEMALS) Study Group. Neurite density is reduced in the presymptomatic phase of *C9orf72* disease. *J Neurol Neurosurg Psychiatry*. 2019;90:387–94.
90. Lee SE, Sias AC, Mandelli ML, Brown JA, Brown AB, Khazenzon AM, et al. Network degeneration and dysfunction in presymptomatic *C9ORF72* expansion carriers. *Neuroimage Clin*. 2017;14:286–97.
91. Waugh RE, Danielian LE, Shoukry RFS, Floeter MK. Longitudinal changes in network homogeneity in presymptomatic *C9orf72* mutation carriers. *Neurobiol Aging*. 2021;99:1–10.
92. Popuri K, Beg MF, Lee H, Balachandar R, Wang L, Sossi V, et al. FDG-PET in presymptomatic *C9orf72* mutation carriers. *Neuroimage Clin*. 2021;31:102687.
93. Van Weehaeghe D, Ceccarini J, Delva A, Robberecht W, Van Damme P, Van Laere K. Prospective validation of ¹⁸F-FDG brain PET discriminant analysis methods in the diagnosis of amyotrophic lateral sclerosis. *J Nucl Med*. 2016;57:1238–43.
94. Van Laere K, Vanhee A, Verschueren J, De Coster L, Driesen A, Dupont P, et al. Value of ¹⁸fluorodeoxyglucose-positron-emission tomography in amyotrophic lateral sclerosis: a prospective study. *JAMA Neurol*. 2014;71:553–61.
95. D'hulst L, Van Weehaeghe D, Chiò A, Calvo A, Moglia C, Canosa A, et al. Multicenter validation of [¹⁸F]-FDG PET and support-vector machine discriminant analysis in automatically classifying patients with amyotrophic lateral sclerosis *versus* controls. *Amyotroph Lateral Scler Frontotemporal Degener*. 2018;19:570–7.

96. Saba L, Viscomi MT, Caioli S, Pignataro A, Bisicchia E, Pieri M, et al. Altered functionality, morphology, and vesicular glutamate transporter expression of cortical motor neurons from a presymptomatic mouse model of amyotrophic lateral sclerosis. *Cereb Cortex*. 2016;26:1512–28.
97. Eisen A, Braak H, Del Tredici K, Lemon R, Ludolph AC, Kiernan MC. Cortical influences drive amyotrophic lateral sclerosis. *J Neurol Neurosurg Psychiatry*. 2017;88:917–24.
98. Menon P, Geevasinga N, Yiannikas C, Howells J, Kiernan MC, Vucic S. Sensitivity and specificity of threshold tracking transcranial magnetic stimulation for diagnosis of amyotrophic lateral sclerosis: a prospective study. *Lancet Neurol*. 2015;14:478–84.
99. Menon P, Kiernan MC, Vucic S. Cortical hyperexcitability precedes lower motor neuron dysfunction in ALS. *Clin Neurophysiol*. 2015;126:803–9.
100. Vucic S, Cheah BC, Yiannikas C, Kiernan MC. Cortical excitability distinguishes ALS from mimic disorders. *Clin Neurophysiol*. 2011;122:1860–6.
101. Misawa S, Noto Y, Shibuya K, Iose S, Sekiguchi Y, Nasu S, et al. Ultrasonographic detection of fasciculations markedly increases diagnostic sensitivity of ALS. *Neurology*. 2011;77:1532–7.
102. Geevasinga N, Howells J, Menon P, van den Bos M, Shibuya K, Matamala JM, et al. Amyotrophic lateral sclerosis diagnostic index: toward a personalized diagnosis of ALS. *Neurology*. 2019;92:e536–47.
103. Tang M, Gao C, Goutman SA, Kalinin A, Mukherjee B, Guan Y, et al. Model-based and model-free techniques for amyotrophic lateral sclerosis diagnostic prediction and patient clustering. *Neuroinformatics*. 2019;17:407–21.
104. Fukushima K, Takamatsu N, Yamamoto Y, Yamazaki H, Yoshida T, Osaki Y, et al. Early diagnosis of amyotrophic lateral sclerosis based on fasciculations in muscle ultrasonography: a machine learning approach. *Clin Neurophysiol*. 2022;140:136–44.