

## Article

# Effects of Pre-Germinative Treatments and Temperatures on Tassel Hyacinth [*Muscari comosum* (L.) Mill.] Seeds

Donato Castronuovo <sup>1</sup>, Lorian Cardone <sup>2,\*</sup> and Vincenzo Candido <sup>2</sup>

<sup>1</sup> School of Agricultural, Forest, Food and Environmental Sciences (SAFE), University of Basilicata, Via dell'Ateneo Lucano, 10, 85100 Potenza, Italy; donato.castronuovo@unibas.it

<sup>2</sup> Department of European and Mediterranean Cultures (DiCEM), University of Basilicata, Via Lanera, 20, 75100 Matera, Italy; vincenzo.candido@unibas.it

\* Correspondence: loriana.cardone@unibas.it; Tel.: +39-0971205371

**Abstract:** *Muscari comosum* (L.) Mill. is a spontaneous plant that grows in the whole Mediterranean area, including the Basilicata and Puglia regions (southern Italy), where it has received inclusion in the Italian National List for Traditional Agri-Food Product (TAP). The food and medicinal uses of bulb are ancient due to its antioxidant properties and high variety of nutrients, such as starch, sugars, and minerals. *Muscari* seed is characterized by morpho-physiological dormancy, and in order to achieve uniform germination, some pre-germinative treatments are needed. In this research, the effects of hydro-priming and osmo-priming, i.e., PEG 8000 and KNO<sub>3</sub>, as well as three germination temperatures (4, 10, and 20 °C), have been evaluated. In general, the average results pointed out that the pre-treatments increased the germination index (GI) by 5% and the germination percentage (GP) by 3% compared to the no-primed control. The germination temperature of 10 °C significantly reduced the median germination time (T50) by 5.4 days and the mean germination time (MGT) by 5 days compared to temperature at 4 °C. In particular, the best results were obtained by “hydro-priming treatment × 10 °C” interaction, in terms of T50 (34.9 days) and MGT (36.3 days). This combination decreased the T50 by 10.5 days and the MGT by 9.6 days compared to the “control × 4 °C” interaction. Pearson’s correlation matrix results highlighted a significant positive link between T50 and MGT (r = 0.993). In conclusion, these techniques enhanced the germination potential so that the use of pre-treated seeds could be included in a cultivation protocol to improve the germination phase and satisfy the growing demand for Italian bulbs.

**Keywords:** seed dormancy; hydro-priming; osmo-priming; polyethylene glycol; potassium nitrate; biodiversity; lampascione; wild edible species



**Citation:** Castronuovo, D.; Cardone, L.; Candido, V. Effects of Pre-Germinative Treatments and Temperatures on Tassel Hyacinth [*Muscari comosum* (L.) Mill.] Seeds. *Agronomy* **2024**, *14*, 225. <https://doi.org/10.3390/agronomy14010225>

Academic Editor: Monica Boscaiu

Received: 28 December 2023

Revised: 11 January 2024

Accepted: 19 January 2024

Published: 21 January 2024



**Copyright:** © 2024 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

## 1. Introduction

*Muscari comosum* (L.) Mill. [*Tassel hyacinth*; syn. *Leopoldia comosa* (L.) Parl.] is a widespread herbaceous geophyte belonging to the *Liliaceae* family and distributed over south-western and central Europe, the Mediterranean area, and eastwards to Iran and Arabia [1].

The food and medicinal use of the bulb is ancient, and its excellent properties were known by the Egyptians, the Greeks, and all the other inhabitants of the Mediterranean area and Asia Minor [2,3]. In particular, in Turkey [4] and in Italian regions as Sardinia [5], Sicily [6], Basilicata [3,7], Puglia [8], Campania, Molise, and Calabria [9], the bulb is used in traditional gastronomy due to its characteristic strong bitter taste [2,10] and interesting biological activities such as anti-inflammatory, anti-cancer, anti-hyperglycemic, diuretic, aphrodisiac, and antioxidant effects [9–11]. Recently, some authors have reported the anti-obesity effects of raw bulbs due to their high total polyphenol and flavonoid content [9].

From a botanical point of view, *M. comosum* is characterized by globose-ovate bulbs wrapped in reddish-pinkish tunics, particular racemes with violet-blue sterile flowers above brownish-green fertile flowers, obovoid capsules, blackish and globose seeds, and

linear leaves [1,2,12]. Propagation can occur from seeds, bulbs, and bulblets, i.e., small bulbs that form laterally from the mother bulb during the growing season [12]. In Italy, it is commonly named “lampascione”, and according to dialect, takes other local names such as “lambagione”, “lampagione”, “muscaro”, “cipuddine”, and “pampasciuli” [8]. It received inclusion in the Italian National List for Traditional Agri-Food Products (TAP) of the Italian Ministry of Agricultural, Food, and Forestry Policies for the Puglia and Basilicata regions, and is generally employed in preserves in oil and in various local recipes [13]. The processing methods of these products are practiced on the territory according to traditional and antique rules, characteristics which attract the majority of consumers [14,15]. In the case of “lampascione”, its harvesting occurs in uncultivated fields before the flowering phase, using a tool similar to a spade with a very narrow blade capable of going deep into the soil. At the beginning of the vegetation, a cut is made to leave a small green spike [1]. Unfortunately, today, the availability of Italian bulbs has been reduced due to the spread of agriculture and agronomic operations such as deep ploughing, and due to the uncontrolled harvesting [16]. Therefore, in order to satisfy the growing demand for bulbs and remedy the limited disposability of imported bulbs from northern Africa (particularly Tunisia and Morocco), seed sowing at high density in shallow containers was recommended for the first two years. Two-year-old bulbs, with a mean diameter of 1.5 cm, can be hand-transplanted and will reach the commercial dimensions (2.5–5 cm) in two years [17].

The epigeal germination of *M. comosum* occurs between November and December in southern Europe, and the range of temperature is 5–15 °C [1,12]. The germination is an important process which influences the yield and quality of crop vegetables [12,18]. Doussi and Thanos (2002) [12] investigated the ecophysiological aspects of seed germination of *M. comosum* and found that white light, simulating daylight, decreases the germination rate, and the optimal temperature for germination is 10 °C. Seeds of *M. comosum* are characterized by a morpho-physiological dormancy due to a hard coating with low water permeability. The long period of dormancy (7–8 months) and low germination rate represent a problem in *M. comosum* cultivation. Many studies on the positive effects of the pre-germinative treatments, such as decreasing the mean germination time of seeds, increasing germination uniformity, and increasing the tolerance of seeds to stress environmental (such as salt stress and thermal stress), have been investigated in different species [16,18,19].

In the case of *Muscari* spp., Labbaf et al. (2023) [20] studied how sulfuric acid and stratification treatments influence the germination of seeds. These authors indicated that only the combination of scarification treatment with sulfuric acid at 15 min and a 45-day stratification period allowed for the germination of *Muscari armeniacum*, and sulfuric acid treatment at 5 min with a 45-day stratification period seeds resulted in a rapid and uniform germination of *Muscari neglectum*. Kirmizi (2023) [21] studied the effects of moist chilling (3 to 12 months), the application of gibberellic acid ( $GA_3$ ) (250 to 1000 mg L<sup>-1</sup>) and four temperature regimes (20, 20–10, 25–15, and 15–10 °C) on the seed dormancy and germination of *Muscari bourgaei* Baker, an ornamental and endemic plant of Turkey. This author reported that the highest germination (90%) was obtained after 9 months of moist chilling treatment and with 1000 mg L<sup>-1</sup>  $GA_3$  application at 15–10 °C. This type of study provides important information for ex situ conservation protocols of rare plants and allows for the safeguarding of biodiversity as an attraction for winter sports and tourism.

As of today, the literature lacks scientific evidence on the effects of pre-germinative treatment, such as priming and its interaction with temperature germination, of *M. comosum* seeds. Priming is a technique which involves hydration and drying of the seeds with water (hydro-priming), with moistened solid carriers such as vermiculite or calcium silicate (matric-priming) or with osmo-priming agents [19]. The osmo-priming is the most common technique involving the hydration of seeds in aerated solutions with low osmolarity, such as potassium nitrate (KNO<sub>3</sub>) and polyethylene glycol (PEG). This latter is the most used, and it does not damage the seed embryos [19,22].

The aim of the present study is to evaluate the effects of three pre-germinative treatments, hydro-priming and osmo-priming with  $\text{KNO}_3$  or PEG 8000, three germination temperatures (20, 10, and 4 °C), and their interactions.

## 2. Materials and Methods

### 2.1. Experimental Description and Research Design

On *Muscari comosum* (L.) Mill. seeds, collected in June 2018 from spontaneous plants in the countryside of Metaponto in southern Italy (40°24' N, 16°48' E; 30 m a.s.l.), some germination tests were carried out from September to November 2019 at the Vegetable Crops and Floriculture laboratory of the University of Basilicata (Potenza, Italy). In particular, pre-germinative treatments, i.e., hydro-priming and osmo-priming, were applied. The hydro-priming was carried out only with distilled water, whereas potassium nitrate ( $\text{KNO}_3$ ) (Fisher Scientific Italia s.r.l., Segrate, Milano, Italy) and polyethylene glycol 8000 (PEG 8000) (Fisher Scientific Italia s.r.l., Segrate, Milano, Italy) were used for the osmo-priming. Moreover, seeds not subjected to any treatment were considered as control.

On 23 September 2019, for each pre-germinative treatment plus the control, 1000 seeds were selected and weighed using a precision balance (TE214S, Sartorius, Goettingen, Germany). Afterwards, they were disinfected using a 10% solution of hydrogen peroxide for 10 min, and distilled water was used to remove the disinfectant solution [23]. Finally, seeds were subjected to different pre-germination treatments, which were carried out for 24 h at a temperature of  $4 \pm 0.5$  °C and in dark conditions [24].

For each pre-germinative treatment, two glass jars, which had previously been sterilized in an autoclave, were filled with 20 mL of distilled water for the hydro-priming, 3%  $\text{KNO}_3$  solution ( $\Psi = -1.35$  MPa) for the potassium nitrate osmo-priming, and 32% PEG8000 solution ( $\Psi = -1.51$  MPa) for the polyethylene glycol osmo-priming. Then, 500 seeds were placed inside each jar. The distilled water used for the  $\text{KNO}_3$  and PEG 8000 osmo-priming treatments had previously been heated in an oven at a temperature of 40 °C in order to achieve a better dissolution of the used compounds. After the pre-germinative treatments, seeds were filtered in a previously sterilized sieve and then washed in distilled water to eliminate any impurities. Then, seeds were dried for 24 h at room temperature (20 °C).

On 25 September 2019, three replicates of 100 seeds from each pre-germinative treatment were placed on two discs of filter paper (90 mm diameter, Whatman filter No.1, GE Healthcare Life Sciences, Buckinghamshire, UK); afterwards, the discs were placed into acrylic plastic Petri dishes 8.5 cm in diameter. Seeds were moistened with 5 mL of distilled water, and the dishes were closed with parafilm. Then, Petri dishes were placed into three germination chambers (GDH-H 500, Falc Instruments s.r.l., Treviglio, Bergamo, Italy) set at 20 °C, 10 °C (*M. comosum* optimal germination temperature), and 4 °C, respectively, in the dark. The germinated seeds were checked daily starting from the 5th day of the germination period, which lasted 53 days, until at least the 90% of the seeds were germinated.

Therefore, a randomized block design with 12 treatments (3 types of pre-germination treatments  $\times$  3 germination temperatures plus a control, i.e., no pre-germination treatment  $\times$  3 germination temperatures), with three replicates consisting each one by a single Petri dish, was carried out for a total of 36 dishes.

As reported in Table 1, the experimental treatments are indicated as follows: S-A (hydro-priming pre-germinative treatment and germination temperature of 20 °C), S-O (hydro-priming pre-germinative treatment and germination temperature of 10 °C), S-F (hydro-priming pre-germinative treatment and germination temperature of 4 °C), K-A ( $\text{KNO}_3$  osmo-priming pre-germinative treatment and germination temperature of 20 °C), K-O ( $\text{KNO}_3$  osmo-priming pre-germinative treatment and germination temperature of 10 °C), K-F ( $\text{KNO}_3$  osmo-priming pre-germinative treatment and germination temperature of 4 °C), P-A (PEG 8000 osmo-priming pre-germinative treatment and germination temperature of 20 °C), P-O (PEG 8000 osmo-priming pre-germinative treatment and germination temperature of 10 °C), P-F (PEG 8000 osmo-priming pre-germinative treatment and germination temperature of 4 °C), C-AA (no pre-germinative treatment and germination

temperature of 20 °C), C-O (no pre-germinative treatment and germination temperature of 10 °C), and P-F (no pre-germinative treatment and germination temperature of 4 °C).

**Table 1.** Trial experimental theses.

Experimental Theses	Pre-Germinative Treatments	Seeds Germination Temperature (°C)
S-A	Hydro-priming	20
S-O	Hydro-priming	10
S-F	Hydro-priming	4
K-A	KNO <sub>3</sub> osmo-priming	20
K-O	KNO <sub>3</sub> osmo-priming	10
K-F	KNO <sub>3</sub> osmo-priming	4
P-A	PEG 8000 osmo-priming	20
P-O	PEG 8000 osmo-priming	10
P-F	PEG 8000 osmo-priming	4
C-A	Control	20
C-O	Control	10
C-F	Control	4

## 2.2. Germination Indexes

In order to evaluate the effects of the different applied treatments on the seeds, the median germination time (T50), mean germination time (MGT), germination index (GI), synchrony of germination (Z), and percentage of germination (GP) were calculated.

In particular, the T50, which indicates the time required to achieve 50% of the final maximum germination [25], was calculated using the Coolbear et al. (1984) [26] formula, modified by Farooq et al. (2005) [27], as follows:

$$T50 = T_i + \frac{\left(\frac{N}{2} - N_i\right) * (T_i - T_j)}{N_i - N_j} \quad (1)$$

where N is the final number of germinated seeds and N<sub>i</sub> and N<sub>j</sub> are the numbers of germinated seeds at times T<sub>i</sub> and T<sub>j</sub>, with N<sub>i</sub> <  $\frac{N}{2}$  < N<sub>j</sub>.

The MGT, representing the average length of time until the maximum germination of the seeds [28], was calculated using the Ellis and Roberts formula (1981) [29]:

$$MGT = \frac{\sum(n_i * d_i)}{N} \quad (2)$$

where n<sub>i</sub> is the number of seeds germinated on each day, d<sub>i</sub> is the number of days from the beginning of the test, and N is the total number of seeds germinated at the termination of the experiment.

The GI, defined as a weighted sum of the daily numbers of germinated seeds and representing an index that describes the germination percentage/speed relationship [30], was calculated using the formula of the Association of Official Seed Analysts [31]:

$$GI = \frac{G_1}{D_1} + \frac{G_2}{D_2} + \dots + \frac{G_n}{D_n} \quad (3)$$

where G<sub>1</sub>, G<sub>2</sub>, and G<sub>n</sub> represent the number of seeds germinated each day, and D<sub>1</sub>, D<sub>2</sub>, and D<sub>n</sub> indicate the day on which the seeds germinated, counting the days from sowing until the last counting day.

The synchrony of germination was calculated using the following formula:

$$Z = \frac{\sum C_{n_i^2}}{N} \quad (4)$$

where  $C_{n_i^2} = \frac{n_i(n_i-1)}{2}$ ,  $N = \frac{\sum n_i(\sum n_i-1)}{2}$ , and n<sub>i</sub> is the number of seeds germinated at time i. Z is a value that varies between 0 and 1, with Z = 1 if all seeds germinate at the same time and Z = 0 if at least two seeds germinate on the same day [26,28].

The germination percentage, representing the seed viability index, was calculated using the formula below:

$$GP = \frac{n}{N} \times 100 \quad (5)$$

where  $n$  is the number of germinated seeds, and  $N$  is the number of total seeds sown.

### 2.3. Statistical Analysis

The Shapiro–Wilk ( $p \leq 0.05$ ) and Bartlett ( $p \leq 0.05$ ) tests were applied to test the normality and homogeneity of variances, respectively. Then, the data were subjected to an analysis of variance (two-way ANOVA) according to the randomized block experimental design, with the typology of the pre-germinative treatment and the germination temperatures as sources of variation. Mean values were separated with the Student–Newman–Keuls (SNK) test at a significance level of  $p \leq 0.05$ .

Moreover, a correlation matrix, implemented using the Pearson correlation coefficient method, was utilized to evaluate the correlations between the pre-germinative treatments and the seeds' germination temperatures using the investigated germination parameters.

All statistical procedures were computed using the software RStudio: Integrated Development for R, version 2023.09.1 Build 494 [32].

## 3. Results

### 3.1. Effects of the Pre-Germinative Treatments on Seeds Germination

As shown in Table 2, the comparison of the studied pre-germinative treatments on the analysed seeds' germination parameters reported significant differences for all the investigated indices. In particular, the pre-germinative treatments yielded similar data, whilst the control always achieved the worst results (Table 2).

**Table 2.** Effects of the pre-germinative treatments on seeds' germination parameters.

Treatments <sup>1</sup>	T50 (Days)	MGT (Days)	GI (-)	Z (-)	GP (%)
Hydro-priming	38.57 b	39.33 b	14.27 a	0.116 a	99.25 a
KNO <sub>3</sub> osmo-priming	37.31 b	38.11 b	14.05 a	0.122 a	99.16 a
PEG 8000 osmo-priming	36.76 b	37.57 b	13.99 a	0.123 a	99.50 a
Control	44.85 a	44.36 a	9.26 b	0.106 b	95.92 b
Significance <sup>2</sup>	*	*	*	*	*

<sup>1</sup> Mean values followed by different letters are significantly different at  $p \leq 0.05$ , according to SNK test. <sup>2</sup> \*, significance at  $p \leq 0.05$ .

### 3.2. Effects of the Germination Temperature on Seed Germination

Since no seed germination occurred at 20 °C, Table 3 reports data only for the germination temperatures of 10 and 4 °C (Table 3).

**Table 3.** Effects of the germination temperature on seeds' germination parameters.

Germination Temperature <sup>1</sup>	T50 (Days)	MGT (Days)	GI (-)	Z (-)	GP (%)
4 °C	42.33 a	42.85 a	12.83	0.130 a	98.25
10 °C	36.92 b	37.84 b	12.96	0.109 b	97.83
Significance <sup>2</sup>	*	*	n.s.	*	n.s.

<sup>1</sup> Mean values followed by different letters are significantly different at  $p \leq 0.05$ , according to SNK test. <sup>2</sup> n.s., no significant difference; \*, significance at  $p \leq 0.05$ .

The germination temperature had effects with significant differences on T50, MGT, and Z; on the contrary, no significant differences were found for GI and GP (Table 3). In all

the parameters with differences, the germination temperature of 4 °C had higher values for the seeds' germination parameters (Table 3).

### 3.3. Interactive Effects of Pre-Germination Treatments and Seeds' Germination Temperatures

The interactive effects of the pre-germinative treatments and the seeds' germination temperature are reported in Figure 1. Statistically, significant differences were observed for T50 (Figure 1a), MGT (Figure 1b), and GI (Figure 1c), whereas Z (Figure 1d) and GP (Figure 1e) showed no significant differences. Concerning to T50, the best results were recorded by S-O, K-O, and P-O, with the shortest median time of germination, while the treatment C-F had the longest T50 time (Figure 1a). The MGT followed the T50 trend, with the S-O, K-O, and P-O treatments achieving the shortest length to maximum seed germination and the C-F resulting the worst one (Figure 1b). Lastly, the germination index (GI) reported higher values for S-O, K-O, and P-O, indicating that those treatments had high seed germination percentages and speeds (Figure 1c).

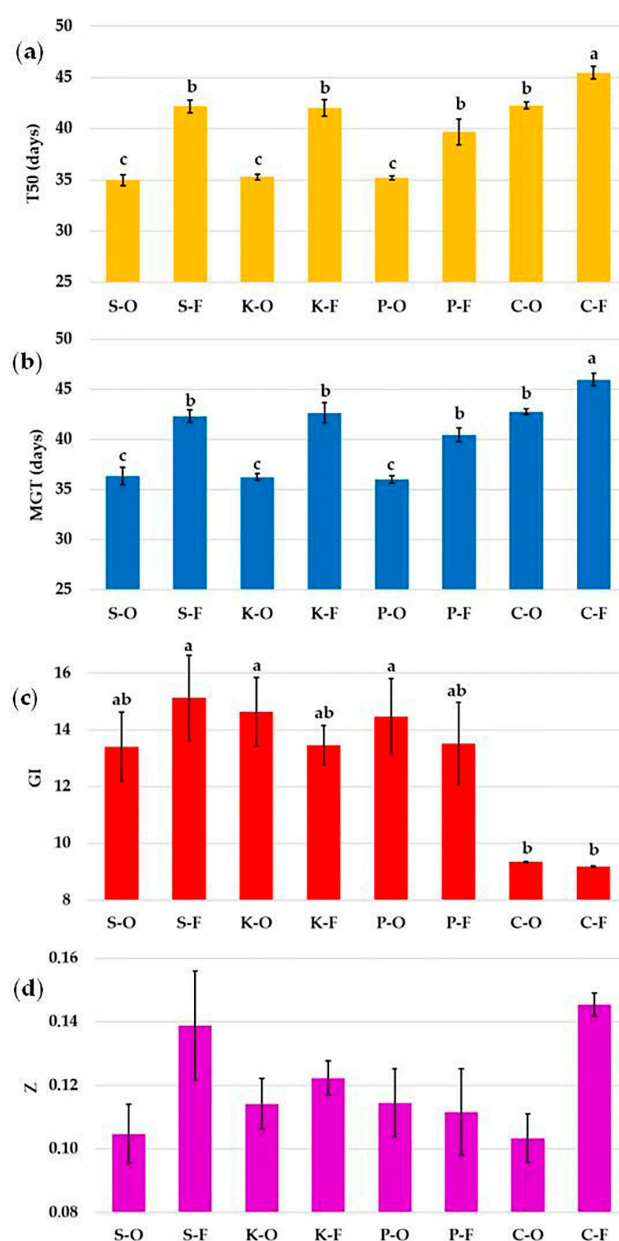
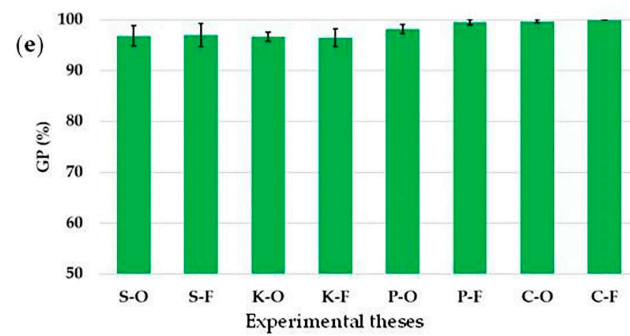


Figure 1. Cont.

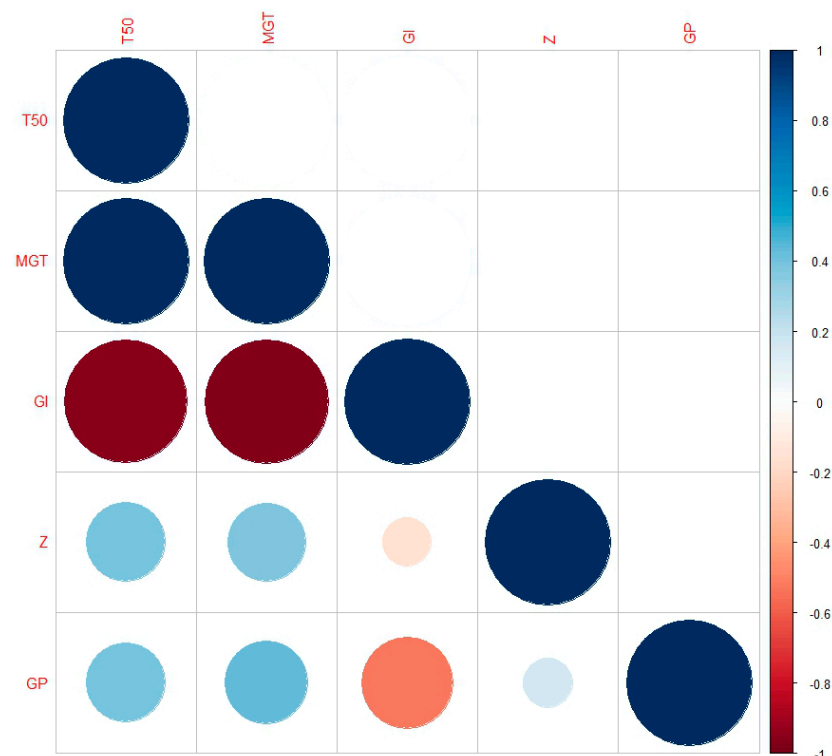


**Figure 1.** Interactive effects of pre-germination treatments and seeds' germination temperatures on (a) T50, (b) MGT, (c) GI, (d) Z, and (e) GP. Mean values followed by different letters are significantly different at  $p \leq 0.05$ , according to SNK test.

### 3.4. Correlation Matrix

As shown in Table 4, the correlation matrix, built using the Pearson correlation coefficient method, highlighted the correlations of the analyzed seeds' germination parameters. A high correlation was found between T50 and MGT, with a Pearson coefficient equal to 0.993 (Table 4).

Moreover, for a visual reading of the correlation matrix (Table 4), a correlogram was built, with positive correlations displayed in blue and negative correlations in red, and with the color intensity and the size of the circles proportional to the correlation coefficients (Figure 2).



**Figure 2.** Correlogram of the studied germination parameters. Color intensity and the size of the circles are proportional to the correlation coefficients.

**Table 4.** Correlation matrix of the studied germination parameters.

	T50	MGT	GI	Z	GP
T50	1.000				
MGT	0.993	1.000			
GI	−0.385	−0.400	1.000		
Z	0.552	0.544	0.326	1.000	
GP	0.403	0.446	−0.002	0.463	1.000

#### 4. Discussion

The process of seed germination is mainly influenced by environmental factors such as temperature and light [33], and can be distinguished into three principal phases: an initial phase (phase I), which involves water imbibitions by the seeds; phase II, which concerns the reactivation of metabolism (hydrolytic enzyme synthesis, respiration, translocation of nutrients and subcellular structures, and cell elongation); and phase III, characterized by the protrusion of radicles. The most critical phase is II, where the pre-germinative treatments act at a biochemical level [34,35].

In general, the species with the most difficult germination, such as the “lampascione”, are subjected to pre-germination treatments, such as scarification, hydro-priming, osmo-priming, matrix-priming, and bio-priming, to obtain rapid and uniform germination [34–36].

In this study, the hydro-priming and osmo-priming treatments, as well as the effect of germination temperature on *M. comosum* seeds, were evaluated. The results reveal that the pre-germinative treatments significantly influenced all of the studied parameters, and the treated seeds presented more interesting values than the untreated ones (Table 2). In particular, these techniques increased the germination index, synchrony of germination, and germination percentage compared to the control (Table 2). On the other hand, these pre-treatments decreased the median germination time and mean germination time (Table 2).

The positive obtained effects of pre-germinative treatments are similar to those of Labbaf et al. (2023) [20], who, by investigating the effects of sulfuric acid and stratification priming on seed germination, found a decrease in germination time to 15 days using a 5 min sulfuric acid treatment plus a 45-day stratification treatment.

The favorable effects of hydro-priming and osmo-priming on *M. comosum* can be attributed to the improvement of the metabolic processes of the seeds and the physiological modifications, such as the efficiency of water use, the accumulation of osmolytes (proline, glycinebetaine, and polyamines), the biosynthesis of some hormones (gibberellins, abscisic acid, and ethylene), and the translocation of reserved food materials [18,24,34]. In addition, the effect of priming can be attributed to the early replication of DNA, an increase in RNA and protein synthesis, greater availability of ATP, and faster growth of the embryos [34].

The beneficial effects of priming have been observed in several field crops, such as tomato [26], wheat [34], maize [37], rice [22,38], sorghum [39], melon [24], alfalfa [35], carrot [40], and faba bean [41], but no studies on *M. comosum* have been reported in the literature. When studying the effect of seed priming on *Liliaceae* species, Selvarani and Umarani (2011) [40] reported an increase in the vigor of the seedlings, greater formation, and mobilization of reserves for the seedling growth of onion plants.

Interaction effect between pre-germinative treatments and temperature was found significant for all parameters, except for Z and GP (Figure 1). In particular, the best results were obtained using the “hydro-priming treatment × 10 °C” interaction in terms of T50 (34.9 days) and MGT (36.3 days). This combination decreased the T50 by 10.5 days and the MGT by 9.6 days compared to the “control × 4 °C” interaction (Figure 1).

The most important factor in the breaking of dormancy and germination is the temperature [21]. No seed germination was found at a temperature of 20 °C. This result is in agreement with Casoria et al. (1999) [42], who reported that germination does not occur above 15 °C. A germination temperature of 10° significantly reduced the median germination time, mean germination time, and synchrony of germination. In particular, the



temperature at 10 °C reduced the T50 and MGT by about 5 days compared to temperature of 4 °C (Table 3).

To provide a more thorough understanding of the plant's response to different environmental conditions, intermediary ranges of temperatures, such as between 4 and 9 °C and between 10 and 15 °C, will be evaluated in future research. Being that the seed germination responses to temperature are species-specific [43], future research could be carried out to study the effect of pre-treatments on different *Muscari* spp. widely present in Basilicata region, such as *Muscari atlanticum* Boiss. et Reuter and *Muscari botryoides* (L.) Mill., whose boiled or fried bulbs are used in traditional gastronomy [44].

The mean germination time, median germination time, and germination index were the best indices for the assessment of the germination performance compared to the synchrony of germination and final germination percentage, as reported in Figure 2.

Seed germination is a crucial process that influences crop yield and quality [45]. Further studies of the effects of seed pre-treatment on the growth, development, yield, and quality of “lampascione” bulbs will be conducted. It will be interesting to determine how these pre-treatments can help heighten to the profitability of “lampascione” cultivation by increasing the fresh weight and diameter of bulbs. On the other hand, germination pre-treatments could enhance seed quality through the accumulation of antioxidants (total ascorbate, dehydroascobate and catalase activity) responsible for the improvement of cell membrane integrity [46,47].

The study of molecular mechanisms underlying the seed dormancy and germination of *M. comosum* could be important for improvement of crop yield and quality. Further studies are necessary in order to identify these mechanisms of the regulation of the metabolic and signaling aspects of different plant hormones, mainly abscisic acid and gibberellins [45,48].

## 5. Conclusions

In recent years, and mainly during the COVID-19 pandemic, the consumption of local and traditional agri-food products has increased due to their high food safety and nutritional aspects. “Lampascione” bulbs are characterized by a long cultivation cycle, which could be considered sustainable if seeds are sown in containers for the first two years. This study suggests that the performance of *M. comosum* seed germination can be improved by employing seed priming techniques, such as hydro-priming and osmo-priming with PEG and KNO<sub>3</sub>. These techniques—mostly hydro-priming—are cheap, eco-friendly, and easy to implement. Moreover, further studies are also needed because seed priming treatments can improve the plant defense system through increased activity of antioxidant enzymes (superoxide dismutase, catalase, and glutathione reductase), allowing the plant to overcome abiotic stresses (high temperature and drought) and adapt better to climate change. Furthermore, the aforementioned seed treatments may be useful for reducing the vegetative cycle of *M. comosum* in order to obtain marketable bulbs earlier. In conclusion, this study provides practical information for farmers regarding improving the seed germination and the subsequent field performance of “lampascione”.

**Author Contributions:** Conceptualization, D.C., L.C. and V.C.; methodology, D.C., L.C. and V.C.; software, D.C., L.C. and V.C.; validation, D.C., L.C. and V.C.; formal analysis, D.C., L.C. and V.C.; investigation, D.C., L.C. and V.C.; resources, V.C.; data curation, D.C., L.C. and V.C.; writing—original draft preparation, D.C., L.C. and V.C.; writing—review and editing, D.C., L.C. and V.C.; visualization, D.C., L.C. and V.C.; supervision, V.C.; project administration, V.C.; funding acquisition, V.C. All authors have read and agreed to the published version of the manuscript.

**Funding:** This research received no external funding.

**Data Availability Statement:** The data presented in this study are available upon request from the corresponding author.

**Acknowledgments:** We thank Matteo Pandolfo for his technical support.

**Conflicts of Interest:** The authors declare no conflicts of interest.

## References

- Candido, V.; Castronuovo, D.; Fascetti, S.; Rosati, L.; Potenza, G. Seed-propagated *Muscari comosum* (L.) Mill.: Effects of sowing date and growing conditions. *Plant Biosyst.* **2017**, *151*, 484–492. [CrossRef]
- Borgonovo, G.; Caimi, S.; Morini, G.; Scaglioni, L.; Bassoli, A. Taste-active compounds in a traditional Italian food: ‘Lampascioni’. *Chem. Biodivers.* **2008**, *5*, 1184–1194. [CrossRef] [PubMed]
- Pieroni, A.; Nebel, S.; Santoro, R.F.; Heinrich, M. Food for two seasons: Culinary uses of non-cultivated local vegetables and mushrooms in a south Italian village. *Int. J. Food Sci. Nutr.* **2005**, *56*, 245–272. [CrossRef] [PubMed]
- Kiran, Y.; Pinar, S.M.; Dogan, G.; Eroglu, H. A karyomorphological study on the subgenus *Leopoldia* of the genus *Muscari* growing in Turkey. *Cytologia* **2020**, *85*, 79–83. [CrossRef]
- Satta, C. *Piante Officinali Spontanee di Sardegna*; Zonza Editori: Cagliari, Italy, 2016.
- Lentini, F.; Venza, F. Wild food plants of popular use in Sicily. *J. Ethnobiol. Ethnomed.* **2007**, *3*, 15–21. [CrossRef] [PubMed]
- Guarrera, P.M.; Salerno, G.; Caneva, G. Folk phytotherapeutical plants from Maratea area (Basilicata, Italy). *J. Ethnopharmacol.* **2005**, *99*, 367–378. [CrossRef] [PubMed]
- Bianco, V.; Mariani, M.; Santamaria, P. *Piante Spontanee nella Cucina Tradizionale Molese*; Edizioni Levante: Mola di Bari, Italy, 2009.
- Casacchia, T.; Sofo, A.; Casaburi, I.; Marrelli, M.; Conforti, F.; Statti, G.A. Antioxidant, Enzyme-Inhibitory and Antitumor Activity of the Wild Dietary Plant *Muscari comosum* (L.) Mill. *Int. J. Plant Biol.* **2017**, *8*, 6895. [CrossRef]
- Giglio, F.; Castiglione Morelli, M.A.; Matera, I.; Sinisgalli, C.; Rossano, R.; Ostuni, A. *Muscari comosum* L. Bulb Extracts Modulate Oxidative Stress and Redox Signaling in HepG2 Cells. *Molecules* **2021**, *26*, 416. [CrossRef]
- Larocca, M.; Di Marsico, M.; Riccio, P.; Rossano, R. The in vitro antioxidant properties of *Muscari comosum* bulbs and their inhibitory activity on enzymes involved in inflammation, post-prandial hyperglycemia, and cognitive/neuromuscular functions. *J. Food Biochem.* **2018**, *42*, 12580. [CrossRef]
- Doussi, M.A.; Thanos, C.A. Ecophysiology of seed germination in Mediterranean geophytes. 1. *Muscari* ssp. *Seed Sci. Res.* **2002**, *12*, 193–201. [CrossRef]
- Ministero dell’Agricoltura e della Sovranità Alimentare, Ventitreesima Revisione dei Prodotti Agroalimentari Tradizionali. Available online: <https://www.politicheagricole.it/flex/cm/pages/ServeBLOB.php/L/IT/IDPagina/19693> (accessed on 12 December 2023).
- Didonna, A.; Renna, M.; Santamaria, P. Traditional Italian Agri-Food Products: A Unique Tool with Untapped Potential. *Agriculture* **2023**, *13*, 1313. [CrossRef]
- Renna, M.; Signore, A.; Santamaria, P. I Prodotti Agroalimentari Tradizionali (PAT), Espressione Del Territorio e Del Patrimonio Culturale Italiano. *Italus Hortus* **2018**, *25*, 1–13. [CrossRef]
- Candido, V.; Matteo, D.; Lacarpia, F.; Castronuovo, D.; Miccolis, V. La propagazione per seme di *Muscari comosum* (L.) Mill. *Italus Hortus* **2010**, *17* (Suppl. S2), 30.
- Bonasia, A.; Conversa, G.; Lazzizzera, C.; La Rotonda, P.; Elia, A. Weed control in lampascione-*Muscari comosum* (L.) Mill. *Crop Prot.* **2012**, *36*, 65–72. [CrossRef]
- Adhikari, B.; Dhital, P.R.; Ranabhat, S.; Poudel, H. Effect of seed hydro-priming durations on germination and seedling growth of bitter melon (*Momordica charantia*). *PLoS ONE* **2021**, *16*, e0255258. [CrossRef] [PubMed]
- Lei, C.; Bagavathiannan, M.; Wang, H.; Sharpe, S.M.; Meng, W.; Yu, J. Osmopriming with Polyethylene Glycol (PEG) for Abiotic Stress Tolerance in Germinating Crop Seeds: A Review. *Agronomy* **2021**, *11*, 2194. [CrossRef]
- Labaf, N.; Rohollahi, I.; Naji, A.M. *Muscari* seed germination enhancement by using sulfuric acid, and stratification priming. *Ornam. Hort.* **2023**, *29*, 171–180. [CrossRef]
- Kirmizi, S. Effects of Pre-Treatments on Seed Dormancy and Germination of Endemic *Muscari bourgaei* Baker. *Agronomy* **2023**, *13*, 2438. [CrossRef]
- Ruttanaruangboworn, A.; Chanprasert, W.; Tobunluepop, P.; Onwimol, D. Effect of seed priming with different concentrations of potassium nitrate on the pattern of seed imbibition and germination of rice (*Oryza sativa* L.). *J. Integr. Agric.* **2017**, *16*, 605–613. [CrossRef]
- Di Girolamo, G.; Barbanti, L. Treatment conditions and biochemical processes influencing seed priming effectiveness. *Ital. J. Agron.* **2012**, *7*, 25. [CrossRef]
- Farooq, M.; Basra, S.M.A.; Rehman, H.; Ahmad, N.; Saleem, B.A. Osmopriming improves the germination and early seedling growth of melons (*Cucumis melo* L.). *Pak. J. Agri. Sci.* **2007**, *44*, 529–536.
- Kumar, S.; Basu, S.; Anand, A.; Kumar Lal, S.; Tomar, B.S. Identification of the Best Germination Indices Represents Seed Quality Status in Unaged and Aged Onion Seeds. *Int. J. Curr. Microbiol. App. Sci.* **2021**, *10*, 76–85. [CrossRef]
- Coolbear, P.; Francis, A.; Grierson, D. The effect of low temperature pre-sowing treatment on the germination performance and membrane integrity of artificially aged tomato seeds. *J. Exp. Bot.* **1984**, *35*, 1609–1617. [CrossRef]
- Farooq, M.; Basra, S.M.A.; Hafeez, K.; Ahmad, N. Thermal hardening: A new seed vigour enhancement tool in rice. *J. Integr. Plant Biol.* **2005**, *47*, 187–193. [CrossRef]
- Czabator, F.J. Germination Value: An Index Combining Speed and Completeness of Pine Seed Germination. *For. Sci.* **1962**, *8*, 386–396.
- Ellis, R.A.; Roberts, E.H. The quantification of aging and survival in orthodox seeds. *Seed Sci. Technol.* **1981**, *9*, 373–409.

30. Kader, M.A. A comparison of seed germination calculation formulae and the associated interpretation of resulting data. *J. Proc. R. Soc. New South Wales* **2005**, *138*, 65–75. [[CrossRef](#)]
31. AOSA. *Seed Vigor Testing Handbook*; Association of Official Seed Analysts: Washington, DC, USA, 1983.
32. R Core Team. *R: A Language and Environment for Statistical Computing*; R Foundation for Statistical Computing: Vienna, Austria, 2023.
33. Ali, A.S.; Elozeiri, A.A. Metabolic processes during seed germination. *Adv. Seed Biol.* **2017**, *2017*, 141–166. [[CrossRef](#)]
34. Lemmens, E.; Deleu, L.J.; De Brier, N.; De Man, W.L.; De Proft, M.; Prinsen, E.; Delcour, J.A. The Impact of Hydro-Priming and Osmo-Priming on Seedling Characteristics, Plant Hormone Concentrations, Activity of Selected Hydrolytic Enzymes, and Cell Wall and Phytate Hydrolysis in Sprouted Wheat (*Triticum aestivum* L.). *ACS Omega* **2019**, *4*, 22089–22100. [[CrossRef](#)]
35. Mouradi, M.; Bouzigaren, A.; Farissi, M.; Makoudi, B.; Kabbadj, A.; Very, A.A.; Sentenac, H.; Qaddoury, A.; Ghoulam, C. Osmopriming improves seeds germination, growth, antioxidant responses and membrane stability during early stage of Moroccan alfalfa populations under water deficit. *Chil. J. Agric. Res.* **2016**, *76*, 265–272. [[CrossRef](#)]
36. Mirmazloun, I.; Kiss, A.; Erdélyi, É.; Ladányi, M.; Németh, É.Z.; Radácsi, P. The Effect of osmopriming on seed germination and early seedling characteristics of *Carum carvi* L. *Agriculture* **2020**, *10*, 94. [[CrossRef](#)]
37. Jayesh, V.; Meeta, J. Influence of halopriming and hydropriming on seed germination and growth characteristics of *Zea mays* L. cv. GSF-2 under salt stress. *Res. J. Chem. Environ.* **2015**, *19*, 10.
38. Jisha, K.C.; Puthur, J.T. Seed halopriming outdo hydropriming in enhancing seedling vigor and osmotic stress tolerance potential of rice varieties. *J. Crop Sci. Biotechnol.* **2014**, *17*, 209–219. [[CrossRef](#)]
39. Singh, A.; Dahiru, R.; Musa, M. Osmopriming Duration Influence on Germination, Emergence and Seedling Growth of Sorghum. *Seed Technol.* **2012**, *34*, 111–118.
40. Selvarani, K.; Umarani, R. Evaluation of seed priming methods to improve seed vigour of onion (*Allium cepa* cv. aggregatum) and carrot (*Daucus carota*). *J. Agric. Sci. Technol.* **2011**, *7*, 857–867.
41. Damalas, C.A.; Koutroubas, S.D.; Fotiadis, S. Hydro-priming effects on seed germination and field performance of faba bean in spring sowing. *Agriculture* **2019**, *9*, 201. [[CrossRef](#)]
42. Casoria, P.; Menale, B.; Muoio, R. Muscari comosum, Liliaceae, in the Food Habits of South Italy. *Econ. Bot.* **1999**, *53*, 113–115.
43. Yi, F.; Wang, Z.; Baskin, C.C.; Baskin, J.M.; Ye, R.; Sun, H.; Zhang, Y.; Ye, X.; Liu, G.; Yang, X.; et al. Seed germination responses to seasonal temperature and drought stress are species-specific but not related to seed size in a desert steppe: Implications for effect of climate change on community structure. *Ecol. Evol.* **2019**, *9*, 2149–2159. [[CrossRef](#)]
44. Pieroni, A.; Nebel, S.; Quave, C.; Münz, H.; Heinrich, M. Ethnopharmacology of liakra: Traditional weedy vegetables of the Arbëreshë of the Vulture area in southern Italy. *J. Ethnopharmacol.* **2002**, *81*, 165–185. [[CrossRef](#)]
45. Tuan, P.A.; Sun, M.; Nguyen, T.-N.; Park, S.; Ayele, B.T. Molecular Mechanisms of Seed Germination. In *Sprouted Grains*; Feng, H., Nemzer, B., DeVries, J.W., Eds.; AACC International Press: St. Paul, MN, USA, 2019; pp. 1–24, ISBN 978-0-12-811525-1. [[CrossRef](#)]
46. Siri, B.; Vichitphan, K.; Kaewnaree, P.; Vichitphan, S.; Klanrit, P. Improvement of quality, membrane integrity and antioxidant systems in sweet pepper (*Capsicum annuum* Linn.) seeds affected by osmopriming. *Aust. J. Crop Sci.* **2013**, *7*, 2068–2073.
47. Chen, K.; Arora, R. Dynamics of the antioxidant system during seed osmopriming, post-priming germination, and seedling establishment in Spinach (*Spinacia oleracea*). *Plant Sci.* **2011**, *180*, 212–220. [[CrossRef](#)] [[PubMed](#)]
48. Farooq, M.A.; Ma, W.; Shen, S.; Gu, A. Underlying Biochemical and Molecular Mechanisms for Seed Germination. *Int. J. Mol. Sci.* **2022**, *23*, 8502. [[CrossRef](#)] [[PubMed](#)]

**Disclaimer/Publisher’s Note:** The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.