



#### University of Groningen

# Exploring natural products: from herbal resources, microbial synthesis to animal models

Guan, Zheng

#### DOI: 10.33612/diss.846916968

IMPORTANT NOTE: You are advised to consult the publisher's version (publisher's PDF) if you wish to cite from it. Please check the document version below.

Document Version Publisher's PDF, also known as Version of record

Publication date: 2024

Link to publication in University of Groningen/UMCG research database

Citation for published version (APA): Guan, Z. (2024). Exploring natural products: from herbal resources, microbial synthesis to animal models. [Thesis fully internal (DIV), University of Groningen]. University of Groningen. https://doi.org/10.33612/diss.846916968

#### Copyright

Other than for strictly personal use, it is not permitted to download or to forward/distribute the text or part of it without the consent of the author(s) and/or copyright holder(s), unless the work is under an open content license (like Creative Commons).

The publication may also be distributed here under the terms of Article 25fa of the Dutch Copyright Act, indicated by the "Taverne" license. More information can be found on the University of Groningen website: https://www.rug.nl/library/open-access/self-archiving-pure/taverneamendment.

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

Downloaded from the University of Groningen/UMCG research database (Pure): http://www.rug.nl/research/portal. For technical reasons the number of authors shown on this cover page is limited to 10 maximum.

Introduction and scope of this thesis

## Abstract

The impact of nature on human health is significant, particularly in the field of medicine. In this chapter, a selective overview is provided on the contributions of plant-derived sources, microbial biosynthesis sources, and animal models to the field of medicine, which have been made by advancements in science and technology. Furthermore, this chapter describes the relevant research conducted within this thesis and its potential significance, approached from these three perspectives.

#### Introduction

Nature is a treasure for mankind. We utilize natural resources to eat, to cure, to wear, to exchange, to study, to work, to play, and to live. As a medical scientists, almost everything from nature in our labs can be used as a cure or tool to serve human health. Whereas the resources on the earth are limited, and their attributes usually are not as perfect as we might hope, scientists dedicate their lives to further exploring nature, to reveal its mystery. This helps to promote the sustainable development and deployment of natural resources for making products that promote our health and well-being.

In this thesis, novel tools for natural product discovery and development are being explored and exemplified with some handy species: a herb, *Perilla frutescens*; a bacterium, *Bacillus subtilis*; and an animal, *Meriones unguiculatus*.

#### Herbal resources

For the past millenniums in human history, natural products (NPs) have dominantly occupied the medical area as the "traditional medicine<sup>1</sup>" until the recent centuries. Among all traditional medicines, the herb is the main source of the NPs, for instance, in "The Grand Compendium of Materia Medica (Běn Căo Gāng Mù, 本草纲目)" about 60% of traditional Chinese medicines are herbs<sup>2</sup>.

It is not difficult to find out the landmarks of the drugs derived from herbs in human history (see Table 1).

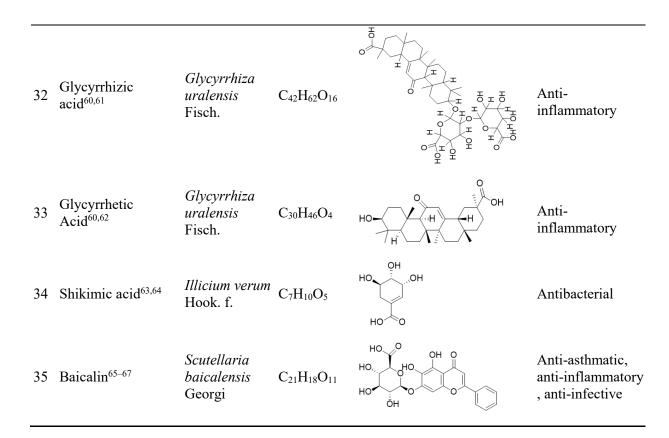
No.	Drug	Source	Formula	Structure	Function
1	Berberine <sup>3–5</sup>	<i>Coptis</i> <i>chinensis</i> Franch.	$C_{20}H_{18}NO_4^+$		Anti-bacterial diarrhea
2	Artemisinin <sup>6,7</sup>	Artemisia annua	C <sub>15</sub> H <sub>22</sub> O <sub>5</sub>		Anti-malarial, anti-infective
3	Camptothecin <sup>8,9</sup>	Camptotheca acuminata	$C_{20}H_{16}N_2O_4$		Antineoplastic, topoisomerase-I inhibitor

### Table 1 Herb-derived drugs.

4	Anisodamine <sup>10,11</sup>	Anisodus tanguticus (Maxim.) Pascher	C <sub>17</sub> H <sub>23</sub> NO <sub>4</sub>	HO H O OH	Spasmolysis, anticholine
5	Paclitaxel <sup>12,13</sup>	Taxus chinensis	C47H51NO14	HO HO HO HO HO HO HO HO HO HO HO HO HO H	Antineoplastic
6	Salvianolic acid A <sup>14,15</sup>	Salvia miltiorrhiza Bge.	C <sub>26</sub> H <sub>22</sub> O <sub>10</sub>		Anti-myocardial infarction, protects cerebrovascular
7	Ephedrine <sup>16,17</sup>	<i>Ephedra sinica</i> Stapf	<sup>4</sup> C <sub>10</sub> H <sub>15</sub> NO	OH H	Relieving asthma, and spasmolysis
8	Puerarin <sup>18,19</sup>	<i>Puerarialobata</i> (Willd.) Ohwi	$^{a}C_{21}H_{20}O_{9}$		Protecting cardiovascular
9	Breviscapine <sup>20,21</sup>	<i>Erigeron</i> <i>breviscapus</i> (Vant.) Hand. -Mazz.	$C_{21}H_{18}O_{12}$		Treating ischemic cerebrovascular disease
10	Gastrodin <sup>22,23</sup>	Gastrodia elata Bl.	$C_{13}H_{18}O_7$		Tranquilizing, and hypnotic
11	Decanoyl acetaldehyde <sup>24,25</sup>	<i>Houttuynia cordata</i> Thunb.	C <sub>12</sub> H <sub>22</sub> O <sub>2</sub>	o H O H O H O H O H O H O H O H O H O H	Antibacterial
12	Matrine <sup>26,27</sup>	Sophora flavescens Alt.	C <sub>15</sub> H <sub>24</sub> N <sub>2</sub> O		Anti-hepatitis B
13	Curcumin <sup>28,29</sup>	Curcuma Longa L.	$C_{21}H_{20}O_{6}$		Anti-hepatitis B

14	Selagine <sup>30,31</sup>	Selaginella tamariscina (Beauv.) Spring	C <sub>15</sub> H <sub>18</sub> N <sub>2</sub> O	O H HNH	Brain protection
15	Bifendate <sup>32</sup>	Schisandra chinensis (Turcz.) Baill.	$C_{20}H_{18}O_{10}$		Liver protection
16	Bicyclol <sup>33,34</sup>	Schisandra chinensis (Turcz.) Baill.	C <sub>19</sub> H <sub>18</sub> O <sub>9</sub>		Liver protection & anti-hepatitis
17	Tanshinone <sup>35,36</sup>	Salvia miltiorrhiza Bge.	$C_{18}H_{12}O_3$		Cardio-cerebrovas cular protection
18	Higenamine <sup>37,38</sup>	Aconitum carmichaelii Debeaux.	C <sub>16</sub> H <sub>17</sub> NO <sub>3</sub>	HO OH	Cardiac function test drug
19	Vinblastine <sup>39</sup>	<i>Catharanthus</i> <i>roseus</i> (L.) G. Don	C <sub>46</sub> H <sub>58</sub> N <sub>4</sub> O <sub>9</sub>	HO, HO, HOHOOO	Antineoplastic
20	Rutin <sup>40,41</sup>	Ruta graveolens L.	C <sub>27</sub> H <sub>30</sub> O <sub>16</sub>		Vascular protection
21	Deoxyschizandrin <sup>42,</sup>	Schisandra chinensis (Turcz.) Baill.	C <sub>24</sub> H <sub>32</sub> O <sub>6</sub>		Liver protection & anti-hepatitis
22	Tetrahydropalmatine 44,45	Corydalis yanhusuo W.T.Wang	C <sub>21</sub> H <sub>25</sub> NO <sub>4</sub>		Non-narcotic analgesic, adrenergic agent

23	Tetramethylpyrazine	<i>Ligusticum</i> <i>chuanxiong</i> Hort.	C <sub>8</sub> H <sub>12</sub> N <sub>2</sub>	N	Vasodilator
24	Sinomenine <sup>47</sup>	Sinomenium acutum	C <sub>19</sub> H <sub>23</sub> NO <sub>4</sub>		Antirheumatic
25	Butylphthalide <sup>48,49</sup>	Apium graveolens	C <sub>12</sub> H <sub>14</sub> O <sub>2</sub>	°	Neuroprotective, platelet aggregation inhibitor
26	Piperine <sup>50,51</sup>	Piper nigrum L.	C <sub>17</sub> H <sub>19</sub> NO <sub>3</sub>		Cytochrome P-450 enzyme inhibitor
27	Shanzhiside methyl ester <sup>52,53</sup>	<i>Lamiophlomis</i> <i>rotata</i> (Benth.) Kudo	C <sub>17</sub> H <sub>26</sub> O <sub>11</sub>		Anti-rheumatic diseases
28	8-O-Acetyl shanzhiside methyl ester <sup>52,54</sup>	<i>Lamiophlomis</i> <i>rotata</i> (Benth.) Kudo	C <sub>19</sub> H <sub>28</sub> O <sub>12</sub>		Analgesic
29	Triptolide <sup>55,56</sup>	<i>Tripterygium wilfordii</i> Hook f	. C <sub>20</sub> H <sub>24</sub> O <sub>6</sub>		Antineoplastic
30	Diosgenin <sup>57,58</sup>	Dioscorea polystachya Turczaninow	C <sub>27</sub> H <sub>42</sub> O <sub>3</sub>		Anti-cholestasis, anti-lipemic
31	Notoginsenoside Fe <sup>59</sup>	Panax notoginseng (Burk.) F. H. Chen ex C. Y. Wu et K.m. Feng	C47H80O17	HO + O + HO + HO + HO + HO + HO + HO +	Hemostasis



As far as we know, all the mentioned herbal drugs in Table 1 are still playing an important role in the clinic. Unfortunately, many of the valuable herbs, are also rare leading to shortages and destruction of natural habitats. Therefore, scientists endeavored to find replacements for medical manufacturing, e.g. same family substitute, recycled herb residues, replacement by using different plant parts, and, finally, biosynthesis. Below are some examples of the alternative resources developed for high-value medicinal plants (see Table 2).

Herb	Medicinal part	Efficacy <sup>68</sup>	Substitute
Panax ginseng C. A. Meyer	Root, rhizome	To tonify the original qi greatly, resume pulse, secure collapse, tonify spleen, replenish kidney, engender fluid, nourish the blood, tranquilize the mind, and replenish wisdom.	Stem and leaf of <i>Panax</i> ginseng; root and rhizome of <i>Codonopsis pilosula</i> (Franch.) Nannf. <sup>69–71</sup>
<i>Cephalotaxus fortune</i> Hook. f.	Seed, root, stem, leaf	To expel parasite, moisten lung for arresting cough, and antineoplastic.	Endophytic fungus of <i>Cephalotaxus fortunei</i> <sup>72</sup>
<i>Dendrobium</i> nobile Lindl	Stem	To boost the stomach, engender fluids, nourish yin, and clear heat.	The whole plant of <i>Pholidota yunnanensis</i> Rolfe <sup>73</sup>
Cistanche	Stem	To tonify the kidney yang, replenish essence and blood, moisten the intestines,	Stem of Cistanche salsa

Table 2	Alternative	resources	of	herbs.
---------	-------------	-----------	----	--------

<i>deserticola</i> Ma		and open the bowels.	(C.A.Mey.) Beck <sup>74</sup>
Daemonorops draco Bl.	Resin	To activate blood, relieve pain, resolve stasis, stanch bleeding, promote tissue regeneration, and promote wound healing.	Artificially induced Resina Draconis <sup>75</sup>
Taxus brevifolia	Bark	To relieve pain and inflammation, anti-hypertensive and antineoplastic.	Taxus cell culture or semi-synthetic (baccatin III) from leafs <sup>76</sup>
<i>Cordyceps</i> sinensis (Berk.) Sacc.	Stroma	To tonify the kidney, replenish lung, stanch bleeding, and resolve phlegm.	Stroma of <i>Cordyceps</i> <i>militaris</i> (L.ex Fr.) Link. <sup>77</sup>

### **Microbial synthesis**

Whereas some scholars have devoted themselves to searching the herbal families and testing the medicinal parts, some others go the other way: biosynthesis. People have engineered bacteria to synthesize herb-derived compounds for the pharmaceutical industry, e.g. Taxol<sup>78</sup> and artemisinin<sup>79</sup>. Among engineered bacteria, *E. coli* and *B. subtilis* are the most frequently used species. Interestingly, *B. subtilis* is more suitable to synthesize medical compounds, since it has the GRAS status (generally regarded as safe).

Actually, *Bacillus subtilis* has many advantages for being used in biosynthesis. It can not only form protective endospores, which permit it to withstand some extreme environment<sup>80,81</sup>, but also possesses a fast-growth rate in simple media, high protein- secretion capacity, and excellent fermentation properties<sup>82,83</sup>. In industries, people can find *B. subtilis* in various fields, e.g. food enzymes, feed additives in agriculture, aquaculture, food and beverage processing, and pharmaceuticals<sup>84,85</sup>.

Although the main use of *B. subtilis* is in producing secreted enzymes, such as proteases and  $\alpha$ -amylases, it is also being used for industrial production of natural products such as riboflavin<sup>86</sup>.

In the past decades, scientists have discovered that *B. subtilis* is a promising host for terpenoid production<sup>87</sup>. Terpenoids, which are composed of isoprene units, are also known as isoprenoids. They form a large group of natural chemicals, comprising around 25% of small natural compounds<sup>88</sup>. Structurally, according to the number of isoprene units, they can be divided into hemiterpenoids (C5), monoterpenoids (C10), sesquiterpenoids (C15), diterpenoids (C20), sesterterpenoids (C25), triterpenoids (C30), tetraterpenoids (C40) and

polyterpenoids<sup>89</sup>. They are everywhere in our daily lives and have been broadly used in numerous fields, including energy, cosmetics, food, and pharmaceuticals.

In the medical area, terpenoids have been used to treat virus and bacterial infection, inflammation, and carcinoma, and many of them have been reported to act as antioxidant<sup>90</sup>. Some examples are artemisinin (antimalarial), paclitaxel (anticancer), ginsenosides (nutrient and anticancer), and carotenoids (anti-oxidant).<sup>91–95</sup> These functional terpenoids can be extracted from plants, however, the concentrations usually are very low. Extracting them, commonly, is not cost-efficient. Moreover, harvesting from the wild will endanger the natural spread of the plants in the environment. Although chemical synthesis can solve part of the issue, normally those compounds have a complex structure and show stereoselectivity. Thus, biosynthesis becomes essential to durably produce those promising natural products.

As a natural isoprene producer, B. subtilis was reported as a propitious potential platform for terpenoid biosynthesis. It has an endogenous 2-C-methyl-D-erythritol 4-phosphate (MEP) pathway for isoprene precursor biosynthesis<sup>96</sup>. The pathway starts with the enzyme 1-deoxy-D-xylulose-5-phosphate synthase (Dxs), which catalyzes the condensation of pyruvate and glyceraldehyde-3-phosphate to form 1-deoxy-D-xylulose-5-phosphate (DXP)<sup>97</sup>. Then the 1-deoxy-D-xylulose-5-phosphate reductoiso-merase IspC reduces and isomerizes the MEP<sup>98</sup>. After these steps<sup>99</sup>, DXP to produce two recognized rate-limiting 4-diphosphocytidyl-2-C-methylerythritol synthase introduces cytidine (IspD) the monophosphate (CMP) moiety to MEP and brings out the 4-diphosphocytidyl-2-Cmethyl-D-erythritol (CDP-ME). Subsequently, IspE adds the phosphate groups to the 4-diphosphocytidyl-2-C-methyl-d-erythritol-2-phosphate CDP-ME, which generates (CDP-ME2P). In the following step, the IspF catalyzed the removal of the cytidyl moiety of CDP-MEP2P and its cyclization, which results in methyl erythritol cyclic diphosphate (MEcDP)<sup>100</sup>. The last reductive steps rely on by IspG and IspH. These two enzymes convert the MEcDP 4-hydroxy-3-methylbut-2-enyl-diphosphate to (HMBDP) and dihydroxylation/isomerization of HMBDP to either isopentenyl pyrophosphate (IPP) or dimethylallyl Diphosphate (DMAPP), respectively<sup>101</sup>. Therefore, numerous terpenoids including those for medical purposes can be produced in *B. subtilis* by building a downstream biosynthesis pathway.

As indicated above, in recent years scientists have made remarkable progress in terpenoid synthesis in *B. subtilis*, such as isoprene, amorphadiene, taxadiene, squalene, menaquinone-7

and carotenoids<sup>87,88,102–105</sup>. However, only a few studies have addressed the biosynthesis of polycyclic triterpenoids, and none of them discusses methods to improve their final production levels, which leaves us with an interesting challenge.

#### **Animal models**

Whereas herbs and bacteria have compounds with interesting pharmaceutical properties, before they can be used in practice they should be tested in disease models. Animals are most often used to establish disease models in medical areas. For example, in the current novel coronavirus disease (COVID-19) outbreak, as early as in 2020, the World Health Organization (WHO) has already assembled an international panel to develop animal models for COVID-19 and has developed models in more than 11 animal species to accelerate the testing of vaccines and therapeutic agents<sup>106</sup>. The importance of these animal models is apparent. In contrast to this speed and abundance in establishing models for acute diseases, for chronic diseases the choice is much more limited. Therefore, much effort has to be dedicated to developing chronic disease animal models.

A classic example is obesity and diabetes mellitus. As it is well known, more than one-third of the global population is overweight and hence at risk of developing type 2 diabetes mellitus (T2DM). This situation has caused the need to use and improve animal models to discover, optimize and validate novel therapeutics for more effective and safe use in humans. According to Matthias's research<sup>107</sup>, people have already developed obesity and diabetes animal models in more than 13 species (including more than 50 models), which has greatly benefited the discovery and development of safer and more potent therapeutics for obesity and T2DM pandemics. However, for some other chronic medical complications, the development of predictive animal models has turned out more difficult. Among these complications, non-alcoholic fatty liver disease (NAFLD) caused liver fibrosis and cirrhosis is a typical case.

Fibrosis and cirrhosis are severe stages of liver dysfunction. They are the third and fourth stages of the natural history of NAFLD, and usually take decades to form clinical symptoms. The long disease progress is difficult to imitate in animal models. This issue seriously hindered drug development and research on disease prevention. To better mimic the pathological process of NAFLD-caused cirrhosis, researchers have developed two types of animal models, one is the genetic model, and the other one is a drug and/or diet-induced model. The genetic models normally relate to lipid metabolism genes, such as db/db mouse, ob/ob mouse, PPAR $\alpha$ -/- mouse, PNPLA3 transgenic mouse, and (fa/fa) Zucker rats. On the

other hand, the drug and/or diet-induced models are more often used including methionine choline-deficient (MCD) diet<sup>108,109</sup>, high-carbohydrate diet (HCD), and high-fat diet (HFD, including high-fat and high-fat, high-cholesterol diets)<sup>110</sup>. However, each of these animal models has deficiencies that cannot be ignored. Firstly, monogenic models cannot represent the real pathological changes of human diseases. Secondly, the drug and/or diet-induced models can rarely reach the "stable" fibrosis stage, not to mention cirrhosis<sup>111,112</sup>.

Therefore, the Mongolian gerbil (*Meriones unguiculatus*) appears to attract researcher's attention, since it is not only a fat-sensitive animal<sup>113–118</sup>, but also a species that is more suitable for carotenoid conversion and metabolism, metabolic disease, and cardiovascular disease studies<sup>119</sup>. Compared to humans, it was indicated that the gerbil's lipemic responses to dietary fat and cholesterol were more sensitive, and the responses can be detected in gerbils even without feeding dietary cholesterol<sup>113–118,120,121</sup>. Moreover, the gerbil also tends to acquire diabetes spontaneously<sup>122</sup>, and has clearly reached the fibrosis stage of the nonalcoholic steatohepatitis (NASH) after diet-induction<sup>123,124</sup>, which makes it into an ideal potential model for NAFLD-caused cirrhosis.

#### Scope of this thesis

The aim of this thesis contains three main aspects. The first part involves the comprehensive utilization of the pomace of an herb, perilla seed, for extracting its phenolic compounds. The second part focuses on the biosynthesis of terpenoids in the GRAS (generally regarded as safe) bacterium, *B. subtilis*. The third part employed the gerbil and established a nonalcoholic fatty liver disease cirrhosis model to be used for drug discovery and testing of natural products.

In **Chapter 1**, we describe the sketch and the main object of this thesis. Specifically, the role of natural products in drug discovery is discussed with an emphasis on functional phenolic compounds and terpenoids from herbs. Subsequently, we discuss the sustainable utilization of herbal resources and the improvements in nature products' biosynthesis, e.g. the terpenoid biosynthesis methods, and the pathways in *B. subtilis*. Subsequently, this thesis explores the utilization of animal models as a valuable tool for drug evaluation, with specific emphasis placed on the nonalcoholic fatty liver disease (NAFLD) cirrhosis model. The inclusion of the NAFLD model is justified due to perilla seeds' lipid-lowering and anti-inflammatory effects, as well as the critical role played by squalene as an essential step within the cholesterol biosynthesis pathway.

In **Chapter 2**, we identify and quantify the phenolic compounds from the seed and pomace of *Perilla frutescens* using HPLC/PDA and HPLC-ESI/QTOF/MS/MS. The purpose of the study is to investigate major phenolic compounds in perilla seeds and pomaces, to check if the pomace could be an alternative resource to the seed for nutritional and medical purposes. In this investigation, herb markers selected by principal components analysis (PCA) are then quantified in both seeds and pomaces. Moreover, fingerprinting approach and multiple discriminant analysis are applied to screen the phenolic markers in 22 batches of samples. Ten phenols are tentatively identified, among which four (rosmarinic acid, luteolin, apigenin, and rosmarinic acid-3-*O*-glucoside) are selected as herb markers. Perilla seeds and pomaces have shown similar phenol profiles, however, the pomaces contained almost two times more of the four herb markers compared to the seeds, which indicates perilla pomace is a promising alternative source of phenolic compounds.

In Chapter 3, we discuss the metabolic engineering of *B. subtilis* for terpenoid production and the encountered challenges. Firstly, the inherent terpenoid biosynthetic pathways of *B. subtilis* are summarised, including the inherent terpenoid biosynthesis enzymes of *B. subtilis*. Secondly, we describe the research progress of genetic engineering of MEP pathway enzymes in *B. subtilis*. Thirdly, the detection and metabolomics methods for engineering terpenoid pathways are also outlined, as these are important for setting up assays to evaluate biosynthetic efficiency. We focus especially on reported metabolites and the inherent terpenoid pathway intermediates in *B. subtilis*.

In Chapter 4, we employ *B. subtilis* to produce squalene, the common precursor of triterpenoids, by introducing multiples squalene synthases (SQSs) from bacteria, fungi, and plants into *B. subtilis*. Furthermore, the expression vector, cultivation temperature, and rate-limiting enzymes within the MEP pathway were systematically studied to enhance squalene production. Finally, a 29-fold increase in squalene titer is achieved by overexpressing SQS from *Bacillus megaterium* (BmSQS) and MEP pathway enzymes compared with the original strain. This represents the first trial of squalene synthesis and improvement in *B. subtilis*.

In **Chapter 5**, based on an HPLC-Q-Orbitrap-MS/MS study, we investigated the expression of dehydrosqualene synthase (CrtM) in *Bacillus subtilis* 168. The results have shown that *crtM* from *Staphylococcus aureus* can not only produce dehydrosqualene but also squalene and phytoene. More interestingly, compared to squalene synthase from *Bacillus megaterium* 

expressed in *B. subtilis*, the CrtM from *S. aureus* can produce up to 2.4 times the amount of squalene. Besides, by adjusting the medium to a nutrition-rich medium or inserting *dxs* to upregulate the upstream donors, the CrtM was able to greatly increase the yield of squalene, which provides evidence for its potential in squalene biosynthesis. Moreover, these three products were also observed in *B. subtilis* strains that only contain SQS, indicating the promiscuity of the squalene synthase-like (SSL) enzymes. Looking into the sterostructures of presqualene diphosphate in CrtM and SQS enzymes highlights the resilience of their active sites and lends additional support to our hypothesis that the squalene synthase-like enzymes are promiscuous and the CrtM could be further remolded to produce more squalene.

In Chapter 6, we focus on developing a NAFLD-caused cirrhosis model in gerbil, to mimic the chronic progress characteristics of NAFLD. The dynamic relationship between hepatic lipid metabolism and cirrhosis was examined. The model's pathological process, lipid metabolism, oxidative stress, liver collagen deposition, and presence of relevant cytokines were tested and evaluated during the full-time frame of disease onset. The gerbil model can start non-alcoholic steatohepatitis within 9 weeks and can develop cirrhosis after 21 weeks of induction. The model's lipids metabolism disorder is accompanied by liver damage development. During the NAFLD progression, triglycerides (TG) and free fatty acids (FFA) have shown a distinct rise and fall tendency, and the turning points are at the fibrosis stage. Besides that, the ratios of total cholesterol (CHO) to high-density lipoprotein cholesterol (HDL-C) exhibited constant growth tendency, and have a good linear relationship with hepatic stellate cells (HSC) ( $R^2 = 0.802, P < 0.001$ ). The model possesses a positive correlation between lipids metabolism and cirrhosis. The compelling rise and fall tendency of TG and FFA indicated that the fibrosis progression could lead to impairment in lipoprotein synthesis and engender decreased TG levels. CHO/HDL-C ratios can indicate fibrosis progress and be used as a blood indicator for disease prediction and prevention.

Chapter 7, summarizes the investigations in this thesis, and describes some future perspectives.

#### References

- 1. Butler MS. The Role of Natural Product Chemistry in Drug Discovery. J Nat Prod. 2004;67(12):2141-2153. doi:10.1021/np040106y
- 2. Shizhen L. Compendium of Materia Medica (Bencao Gangmu) 6 Vols. Foreign Language Press; 2006.

- 3. Yue SJ, Liu J, Wang WX, et al. Berberine treatment-emergent mild diarrhea associated with gut microbiota dysbiosis. *Biomedicine & Pharmacotherapy*. 2019;116:109002.
- 4. Rabbani G, Butler T, Knight J, Sanyal S, Alam K. Randomized controlled trial of berberine sulfate therapy for diarrhea due to enterotoxigenic Escherichia coli and Vibrio cholerae. *Journal of Infectious Diseases*. 1987;155(5):979-984.
- 5. Chen C, Tao C, Liu Z, et al. A randomized clinical trial of berberine hydrochloride in patients with diarrhea-predominant irritable bowel syndrome. *Phytotherapy Research*. 2015;29(11):1822-1827.
- 6. Cheong DH, Tan DW, Wong FW, Tran T. Anti-malarial drug, artemisinin and its derivatives for the treatment of respiratory diseases. *Pharmacological research*. 2020;158:104901.
- 7. Chaturvedi D, Goswami A, Saikia PP, Barua NC, Rao PG. Artemisinin and its derivatives: a novel class of anti-malarial and anti-cancer agents. *Chemical Society Reviews*. 2010;39(2):435-454.
- 8. Kang J, Kumar V, Yang D, Chowdhury PR, Hohl RJ. Cyclodextrin complexation: influence on the solubility, stability, and cytotoxicity of camptothecin, an antineoplastic agent. *European journal of pharmaceutical sciences*. 2002;15(2):163-170.
- 9. Hertzberg RP, Caranfa MJ, Holden KG, et al. Modification of the hydroxylactone ring of camptothecin: inhibition of mammalian topoisomerase I and biological activity. *Journal of medicinal chemistry*. 1989;32(3):715-720.
- 10. Poupko JM, Baskin SI, Moore E. The pharmacological properties of anisodamine. *Journal of Applied Toxicology: An International Journal*. 2007;27(2):116-121.
- 11. Zhang WW, Song MK, Cui YY, et al. Differential neuropsychopharmacological influences of naturally occurring tropane alkaloids anisodamine versus scopolamine. *Neuroscience letters*. 2008;443(3):241-245.
- 12. Rowinsky E, Donehower R. The clinical pharmacology of paclitaxel (Taxol). In: Seminars in Oncology. Vol 20.; 1993:16-25.
- 13. Alqahtani FY, Aleanizy FS, El Tahir E, Alkahtani HM, AlQuadeib BT. Paclitaxel. In: *Profiles of Drug Substances, Excipients and Related Methodology*. Vol 44. Elsevier; 2019:205-238.
- 14. Wang S bao, Tian S, Yang F, Yang H guang, Yang X ying, Du G hua. Cardioprotective effect of salvianolic acid A on isoproterenol-induced myocardial infarction in rats. *European journal of pharmacology*. 2009;615(1-3):125-132.
- 15. Liu C di, Liu N nan, Zhang S, et al. Salvianolic acid A prevented cerebrovascular endothelial injury caused by acute ischemic stroke through inhibiting the Src signaling pathway. *Acta Pharmacologica Sinica*. 2021;42(3):370-381.
- 16. Drew C, Knight G, Hughes D, Bush M. Comparison of the effects of D-(-)-ephedrine and L-(+)-pseudoephedrine on the cardiovascular and respiratory systems in man. *British journal of clinical pharmacology*. 1978;6(3):221-225.
- 17. Gad MZ, Azab SS, Khattab AR, Farag MA. Over a century since ephedrine discovery: an updated revisit to its pharmacological aspects, functionality and toxicity in comparison to its herbal extracts. *Food & Function*. 2021;12(20):9563-9582.

- 18. Zhou YX, Zhang H, Peng C. Puerarin: a review of pharmacological effects. *Phytotherapy Research*. 2014;28(7):961-975.
- 19. Hou N, Huang Y, Cai S ai, et al. Puerarin ameliorated pressure overload-induced cardiac hypertrophy in ovariectomized rats through activation of the PPARα/PGC-1 pathway. *Acta Pharmacologica Sinica*. 2021;42(1):55-67.
- 20. Lin LL, Liu AJ, Liu JG, Yu XH, Qin LP, Su DF. Protective effects of scutellarin and breviscapine on brain and heart ischemia in rats. *Journal of Cardiovascular Pharmacology*. 2007;50(3):327-332.
- 21. Wen L, He T, Yu Ax, et al. Breviscapine: A Review on its Phytochemistry, Pharmacokinetics and Therapeutic Effects. *The American journal of Chinese medicine*. 2021;49(06):1369-1397.
- 22. Liu Y, Gao J, Peng M, et al. A review on central nervous system effects of gastrodin. *Frontiers in pharmacology*. 2018;9:24.
- 23. Qin B, Luo N, Li Y, et al. Protective effect of gastrodin on peripheral neuropathy induced by anti-tumor treatment with vincristine in rat models. *Drug and Chemical Toxicology*. 2021;44(1):84-91.
- 24. Verma RS, Joshi N, Padalia RC, et al. Chemical Composition and Allelopathic, Antibacterial, Antifungal, and Antiacetylcholinesterase Activity of Fish-mint (Houttuynia cordataThunb.) from India. *Chemistry & biodiversity*. 2017;14(10):e1700189.
- 25. Liu X, Zhong L, Sui Y, et al. Sodium houttuyfonate: A review of its antimicrobial, anti-inflammatory and cardiovascular protective effects. *European Journal of Pharmacology*. 2021;902:174110.
- 26. Huang J, Xu H. Matrine: Bioactivities and structural modifications. *Current Topics in Medicinal Chemistry*. 2016;16(28):3365-3378.
- 27. Li X, Tang Z, Wen L, Jiang C, Feng Q. Matrine: a review of its pharmacology, pharmacokinetics, toxicity, clinical application and preparation researches. *Journal of Ethnopharmacology*. 2021;269:113682.
- 28. Nabavi SF, Daglia M, Moghaddam AH, Habtemariam S, Nabavi SM. Curcumin and liver disease: from chemistry to medicine. *Comprehensive Reviews in Food Science and Food Safety*. 2014;13(1):62-77.
- 29. Musarra-Pizzo M, Pennisi R, Ben-Amor I, Mandalari G, Sciortino MT. Antiviral activity exerted by natural products against human viruses. *Viruses*. 2021;13(5):828.
- 30. Bai D, Tang X, He X. Huperzine A, a potential therapeutic agent for treatment of Alzheimer's disease. *Current Medicinal Chemistry*. 2000;7(3):355-374.
- 31. Wang B, Guan C, Fu Q. The traditional uses, secondary metabolites, and pharmacology of Lycopodium species. *Phytochemistry Reviews*. Published online 2021:1-79.
- 32. Pan SY, Yang R, Dong H, Yu Z ling, Ko KM. Bifendate treatment attenuates hepatic steatosis in cholesterol/bile salt- and high-fat diet-induced hypercholesterolemia in mice. *European Journal of Pharmacology*. 2006;552(1):170-175. doi:https://doi.org/10.1016/j.ejphar.2006.09.011
- 33. Zhao J, Chen H, Li Y. Protective effect of bicyclol on acute alcohol-induced liver injury in mice. *European journal of pharmacology*. 2008;586(1-3):322-331.

- 34. Zhao T, Mao L, Yu Z, et al. Therapeutic potential of bicyclol in liver diseases: Lessons from a synthetic drug based on herbal derivative in traditional Chinese medicine. *International Immunopharmacology*. 2021;91:107308. doi:https://doi.org/10.1016/j.intimp.2020.107308
- 35. Park EJ, Zhao YZ, Kim YC, Sohn DH. Preventive effects of a purified extract isolated from Salvia miltiorrhiza enriched with tanshinone I, tanshinone IIA and cryptotanshinone on hepatocyte injury in vitro and in vivo. *Food and Chemical Toxicology*. 2009;47(11):2742-2748. doi:https://doi.org/10.1016/j.fct.2009.08.007
- 36. Lai Z, He J, Zhou C, Zhao H, Cui S. Tanshinones: An update in the medicinal chemistry in recent 5 years. *Current medicinal chemistry*. 2021;28(14):2807-2827.
- 37. Zhang N, Lian Z, Peng X, Li Z, Zhu H. Applications of Higenamine in pharmacology and medicine. *Journal of ethnopharmacology*. 2017;196:242-252.
- 38. Zhu J xing, Ling W, Xue C, et al. Higenamine attenuates cardiac fibroblast abstract and fibrosis via inhibition of TGF-β1/Smad signaling. *European journal of pharmacology*. 2021;900:174013.
- 39. Gigant B, Wang C, Ravelli RB, et al. Structural basis for the regulation of tubulin by vinblastine. *Nature*. 2005;435(7041):519-522.
- 40. Al-Dhabi NA, Arasu MV, Park CH, Park SU. An up-to-date review of rutin and its biological and pharmacological activities. *EXCLI journal*. 2015;14:59.
- 41. Negahdari R, Bohlouli S, Sharifi S, et al. Therapeutic benefits of rutin and its nanoformulations. *Phytotherapy Research*. 2021;35(4):1719-1738.
- 42. Yan F, Zhang QY, Jiao L, et al. Synergistic hepatoprotective effect of Schisandrae lignans with Astragalus polysaccharides on chronic liver injury in rats. *Phytomedicine*. 2009;16(9):805-813.
- 43. Kopustinskiene DM, Bernatoniene J. Antioxidant effects of Schisandra chinensis fruits and their active constituents. *Antioxidants*. 2021;10(4):620.
- 44. Cao FL, Shang GW, Wang Y, Yang F, Li CL, Chen J. Antinociceptive effects of intragastric DL-tetrahydropalmatine on visceral and somatic persistent nociception and pain hypersensitivity in rats. *Pharmacology Biochemistry and Behavior*. 2011;100(1):199-204.
- 45. Liu J, Dai R, Damiescu R, Efferth T, Lee DY. Role of Levo-tetrahydropalmatine and its metabolites for management of chronic pain and opioid use disorders. *Phytomedicine*. 2021;90:153594.
- 46. Kwan C, Daniel E, Chen M. Inhibition of vasoconstriction by tetramethylpyrazine: does it act by blocking the voltage-dependent Ca channel? *Journal of Cardiovascular Pharmacology*. 1990;15(1):157-162.
- 47. Liu W, Zhang Y, Zhu W, et al. Sinomenine Inhibits the Progression of Rheumatoid Arthritis by Regulating the Secretion of Inflammatory Cytokines and Monocyte/Macrophage Subsets. *Frontiers in Immunology*. Published online 2018.
- 48. Abdoulaye IA, Guo YJ. A review of recent advances in neuroprotective potential of 3-N-butylphthalide and its derivatives. *BioMed research international*. 2016;2016.
- 49. Wang B ni, Wu C biao, Chen Z miao, et al. DL-3-n-butylphthalide ameliorates diabetes-associated cognitive decline by enhancing PI3K/Akt signaling and suppressing oxidative stress. *Acta Pharmacologica Sinica*. 2021;42(3):347-360.

- 50. Atal C, Dubey RK, Singh J. Biochemical basis of enhanced drug bioavailability by piperine: evidence that piperine is a potent inhibitor of drug metabolism. *Journal of Pharmacology and Experimental Therapeutics*. 1985;232(1):258-262.
- 51. Zhang W, Zheng Q, Song M, et al. A review on the bioavailability, bio-efficacies and novel delivery systems for piperine. *Food & Function*. 2021;12(19):8867-8881.
- 52. Ghule B, Kotagale N, Patil K. Inhibition of the pro-inflammatory mediators in rat neutrophils by shanzhiside methyl ester and its acetyl derivative isolated from Barleria prionitis. *Journal of Ethnopharmacology*. 2020;249:112374.
- 53. Zhao X, Jiang S, Dong Q, et al. Anti-rheumatoid arthritis effects of iridoid glucosides from Lamiophlomis rotata (Benth.) kudo on adjuvant-induced arthritis in rats by OPG/RANKL/NF-κB signaling pathways. *Journal of Ethnopharmacology*. 2021;266:113402.
- 54. Zhang W, Bai Y, Qiao Y, et al. 8-O-acetyl shanzhiside methylester from Lamiophlomis rotata reduces neuropathic pain by inhibiting the ERK/TNF-α pathway in spinal astrocytes. *Frontiers in Cellular Neuroscience*. 2018;12:54.
- 55. Shi X, Jin Y, Cheng C, et al. Triptolide inhibits Bcr-Abl transcription and induces apoptosis in STI571-resistant chronic myelogenous leukemia cells harboring T315I mutation. *Clinical Cancer Research: an Official Journal of the American Association for Cancer Research*. 2009;15(5):1686-1697.
- 56. Jiang L, Gu Y, Du Y, Tang X, Wu X, Liu J. Engineering exosomes endowed with targeted delivery of triptolide for malignant melanoma therapy. *ACS Applied Materials & Interfaces*. 2021;13(36):42411-42428.
- 57. Son IS, Kim JH, Sohn HY, Son KH, Kim JS, Kwon CS. Antioxidative and hypolipidemic effects of diosgenin, a steroidal saponin of yam (Dioscorea spp.), on high-cholesterol fed rats. *Bioscience, biotechnology, and biochemistry*. 2007;71(12):3063-3071.
- 58. Yu L, Lu H, Yang X, et al. Diosgenin alleviates hypercholesterolemia via SRB1/CES-1/CYP7A1/FXR pathway in high-fat diet-fed rats. *Toxicology and Applied Pharmacology*. 2021;412:115388.
- 59. Gao B, Huang L, Liu H, et al. Platelet P2Y12 receptors are involved in the haemostatic effect of notoginsenoside Ft1, a saponin isolated from P anax notoginseng. *British journal of pharmacology*. 2014;171(1):214-223.
- 60. Richard SA. Exploring the pivotal immunomodulatory and anti-inflammatory potentials of glycyrrhizic and glycyrrhetinic acids. *Mediators of Inflammation*. 2021;2021.
- 61. Yu JY, Ha JY, Kim KM, Jung YS, Jung JC, Oh S. Anti-inflammatory activities of licorice extract and its active compounds, glycyrrhizic acid, liquiritin and liquiritigenin, in BV2 cells and mice liver. *Molecules*. 2015;20(7):13041-13054.
- 62. Finney R, Somers G. The anti-inflammatory activity of glycyrrhetinic acid and derivatives. *Journal of Pharmacy and Pharmacology*. 1958;10(1):613-620.
- 63. Bai J, Wu Y, Liu X, Zhong K, Huang Y, Gao H. Antibacterial activity of shikimic acid from pine needles of Cedrus deodara against Staphylococcus aureus through damage to cell membrane. *International Journal of Molecular Sciences*. 2015;16(11):27145-27155.

- 64. Salem MA, El-Shiekh RA, Hashem RA, Hassan M. In vivo antibacterial activity of star anise (Illicium verum Hook.) Extract Using Murine MRSA skin infection model in relation to its metabolite profile. *Infection and drug resistance*. 2021;14:33.
- 65. Liu J, Wei Y, Luo Q, et al. Baicalin attenuates inflammation in mice with OVA-induced asthma by inhibiting NF-κB and suppressing CCR7/CCL19/CCL21. *International Journal of Molecular Medicine*. 2016;38(5):1541-1548.
- 66. Shen J, Cheng J, Zhu S, et al. Regulating effect of baicalin on IKK/IKB/NF-kB signaling pathway and apoptosis-related proteins in rats with ulcerative colitis. *International Immunopharmacology*. 2019;73:193-200.
- 67. Shen J, Li P, Liu S, et al. Traditional uses, ten-years research progress on phytochemistry and pharmacology, and clinical studies of the genus Scutellaria. *Journal of Ethnopharmacology*. 2021;265:113198.
- 68. Pharmacopoeia of the People's Republic of China. China Medical Science Press; 2020.
- 69. Fu YP, Li LX, Zhang BZ, et al. Characterization and prebiotic activity in vitro of inulin-type fructan from Codonopsis pilosula roots. *Carbohydrate Polymers*. 2018;193:212-220.
- 70. Lee SE, Lee SW, Bang JK, Yu YJ, Seong NS. Antioxidant activities of leaf, stem and root of Panax ginseng CA Meyer. *Korean Journal of Medicinal Crop Science*. 2004;12(3):237-242.
- 71. Guo M, Shao S, Wang D, Zhao D, Wang M. Recent progress in polysaccharides from Panax ginseng CA Meyer. *Food & Function*. 2021;12(2):494-518.
- 72. LIU G. Isolation, identification and antimicrobial activities of endophytic fungi from Cephalotaxus fortunei. *Chinese Pharmaceutical Journal*. Published online 2013:165-170.
- 73. Xuemei M, Ping Z, Suping Y, Manfei L. Analysis of the total alkaloids and polysaccharides in Yunnashixiantao Pholidota yunanensis and Shihu Dendrobium nobile. *Zhong cao yao= Chinese Traditional and Herbal Drugs*. 1997;28(9):561-563.
- 74. Wang Y, Zhang S, Luo G, et al. Analysis of phenylethanoid glycosides in the extract of herba Cistanchis by LC/ESI-MS/MS. *Yao xue xue bao= Acta Pharmaceutica Sinica*. 2000;35(11):839-842.
- 75. Chen Q, He L, Mo C, et al. Rapid Evaluation of Chemical Consistency of Artificially Induced and Natural Resina Draconis Using Ultra-Performance Liquid Chromatography Quadrupole-Time-of-Flight Mass Spectrometry-Based Chemical Profiling. *Molecules*. 2018;23(8):1850.
- 76. Croteau R, Ketchum REB, Long RM, Kaspera R, Wildung MR. Taxol biosynthesis and molecular genetics. *Phytochem Rev.* 2006;5(1):75-97. doi:10.1007/s11101-005-3748-2
- 77. Liu Y, Xiao K, Wang Z, Wang S, Xu F. Comparison of metabolism substances in Cordyceps sinensis and Cordyceps militaris cultivated with tussah pupa based on LC-MS. *Journal of Food Biochemistry*. 2021;45(6):e13735.
- 78. Huang Q, Roessner CA, Croteau R, Scott AI. Engineering Escherichia coli for the synthesis of taxadiene, a key intermediate in the biosynthesis of taxol. *Bioorganic & medicinal chemistry*. 2001;9(9):2237-2242.

- 79. Chang YJ, Song SH, Park SH, Kim SU. Amorpha-4, 11-diene synthase of Artemisia annua: cDNA isolation and bacterial expression of a terpene synthase involved in artemisinin biosynthesis. *Archives of Biochemistry and Biophysics*. 2000;383(2):178-184.
- 80. Earl AM, Losick R, Kolter R. Ecology and genomics of Bacillus subtilis. *Trends in microbiology*. 2008;16(6):269-275.
- 81. McKenney PT, Driks A, Eichenberger P. The Bacillus subtilis endospore: assembly and functions of the multilayered coat. *Nature Reviews Microbiology*. 2013;11(1):33-44.
- 82. Liu Y, Liu L, Li J, Du G, Chen J. Synthetic biology toolbox and chassis development in Bacillus subtilis. *Trends in biotechnology*. 2019;37(5):548-562.
- 83. Gu Y, Xu X, Wu Y, et al. Advances and prospects of Bacillus subtilis cellular factories: from rational design to industrial applications. *Metabolic engineering*. 2018;50:109-121.
- 84. Su Y, Liu C, Fang H, Zhang D. Bacillus subtilis: a universal cell factory for industry, agriculture, biomaterials and medicine. *Microbial cell factories*. 2020;19(1):1-12.
- 85. Schallmey M, Singh A, Ward OP. Developments in the use of Bacillus species for industrial production. *Canadian journal of microbiology*. 2004;50(1):1-17.
- 86. Chu R, Li R, Wang C, Ban R. Production of vitamin B2 (riboflavin) by Bacillus subtilis. *Journal* of Chemical Technology & Biotechnology. 2022;97(8):1941-1949. doi:10.1002/jctb.7017
- 87. Pramastya H, Song Y, Elfahmi EY, Sukrasno S, Quax WJ. Positioning Bacillus subtilis as terpenoid cell factory. *Journal of Applied Microbiology*. 2021;130(6):1839-1856.
- 88. Guan Z, Xue D, Abdallah II, et al. Metabolic engineering of Bacillus subtilis for terpenoid production. *Applied Microbiology and Biotechnology*. 2015;99(22):9395-9406.
- 89. Bohlmann J, Keeling CI. Terpenoid biomaterials. The Plant Journal. 2008;54(4):656-669.
- 90. Tholl D. Biosynthesis and biological functions of terpenoids in plants. *Biotechnology of isoprenoids*. Published online 2015:63-106.
- 91. Guo Y hang, Kuruganti R, Gao Y. Recent advances in ginsenosides as potential therapeutics against breast cancer. *Current Topics in Medicinal Chemistry*. 2019;19(25):2334-2347.
- 92. Bai L, Gao J, Wei F, Zhao J, Wang D, Wei J. Therapeutic potential of ginsenosides as an adjuvant treatment for diabetes. *Frontiers in pharmacology*. 2018;9:423.
- 93. Moran NE, Mohn ES, Hason N, Erdman Jr JW, Johnson EJ. Intrinsic and extrinsic factors impacting absorption, metabolism, and health effects of dietary carotenoids. *Advances in Nutrition*. 2018;9(4):465-492.
- 94. Weaver BA. How Taxol/paclitaxel kills cancer cells. *Molecular biology of the cell*. 2014;25(18):2677-2681.
- 95. Tsuruta H, Paddon CJ, Eng D, et al. High-level production of amorpha-4, 11-diene, a precursor of the antimalarial agent artemisinin, in Escherichia coli. *PLoS One*. 2009;4(2):e4489.
- 96. Kunst F, Rapoport G. Salt stress is an environmental signal affecting degradative enzyme synthesis in Bacillus subtilis. *Journal of bacteriology*. 1995;177(9):2403-2407.

- 97. Wagner WP, Helmig D, Fall R. Isoprene biosynthesis in Bacillus subtilis via the methylerythritol phosphate pathway. *Journal of natural products*. 2000;63(1):37-40.
- 98. Hoeffler JF, Tritsch D, Grosdemange-Billiard C, Rohmer M. Isoprenoid biosynthesis via the methylerythritol phosphate pathway: Mechanistic investigations of the 1-deoxy-d-xylulose 5-phosphate reductoisomerase. *European Journal of Biochemistry*. 2002;269(18):4446-4457.
- 99. Xue J, Ahring BK. Enhancing isoprene production by genetic modification of the 1-deoxy-d-xylulose-5-phosphate pathway in Bacillus subtilis. *Applied and environmental microbiology*. 2011;77(7):2399-2405.
- 100. Liu Z, Jin Y, Liu W, Tao Y, Wang G. Crystal structure of IspF from Bacillus subtilis and absence of protein complex assembly amongst IspD/IspE/IspF enzymes in the MEP pathway. *Bioscience reports*. 2018;38(1).
- 101.Lee M, Gräwert T, Quitterer F, et al. Biosynthesis of isoprenoids: crystal structure of the [4Fe–4S] cluster protein IspG. *Journal of molecular biology*. 2010;404(4):600-610.
- 102. Mahdinia E, Demirci A, Berenjian A. Enhanced vitamin K (Menaquinone-7) production by Bacillus subtilis natto in biofilm reactors by optimization of glucose-based medium. *Current pharmaceutical biotechnology*. 2018;19(11):917-924.
- 103. Xue D, Abdallah II, de Haan IE, Sibbald MJ, Quax WJ. Enhanced C30 carotenoid production in Bacillus subtilis by systematic overexpression of MEP pathway genes. *Applied microbiology and biotechnology*. 2015;99(14):5907-5915.
- 104. Abdallah II, Pramastya H, Van Merkerk R, Quax WJ. Metabolic engineering of Bacillus subtilis toward taxadiene biosynthesis as the first committed step for taxol production. *Frontiers in microbiology*. 2019;10:218.
- 105. Cui, S., Lv, X., Wu, Y., Li, J., Du, G., Ledesma-Amaro, R. and Liu, L.,. Engineering a bifunctional Phr60-Rap60-Spo0A quorum-sensing molecular switch for dynamic fine-tuning of menaquinone-7 synthesis in Bacillus subtilis. ACS Synthetic Biology. 2019;8(8):1826-1837.
- 106. Muñoz-Fontela C, Dowling WE, Funnell SGP, et al. Animal models for COVID-19. *Nature*. 2020;586(7830):509-515. doi:10.1038/s41586-020-2787-6
- 107. Kleinert M, Clemmensen C, Hofmann SM, et al. Animal models of obesity and diabetes mellitus. *Nat Rev Endocrinol.* 2018;14(3):140-162. doi:10.1038/nrendo.2017.161
- 108. Chae MK, Park SG, Song SO, et al. Pentoxifylline attenuates methionine- and choline-deficient-diet-induced steatohepatitis by suppressing TNF-α expression and endoplasmic reticulum stress. *Exp Diabetes Res.* 2012;2012:762565. doi:10.1155/2012/762565
- 109. Loyer X, Paradis V, Hénique C, et al. Liver microRNA-21 is overexpressed in non-alcoholic steatohepatitis and contributes to the disease in experimental models by inhibiting PPARα expression. *Gut.* 2016;65(11):1882-1894.
- 110. Feldstein AE, Canbay A, Guicciardi ME, Higuchi H, Bronk SF, Gores GJ. Diet associated hepatic steatosis sensitizes to Fas mediated liver injury in mice. *Journal of hepatology*. 2003;39(6):978-983.
- 111. Kucera O, Cervinkova Z. Experimental models of non-alcoholic fatty liver disease in rats. *World journal of gastroenterology: WJG*. 2014;20(26):8364.

- 112. Hebbard L, George J. Animal models of nonalcoholic fatty liver disease. *Nature reviews Gastroenterology & hepatology*. 2011;8(1):35-44.
- 113. Hegsted DM, Gallagher A. Dietary fat and cholesterol and serum cholesterol in the gerbil. *Journal of lipid research*. 1967;8(3):210-214.
- 114. Wijendran V, Pronczuk A, Bertoli C, Hayes KC. Dietary trans-18: 1 raises plasma triglycerides and VLDL cholesterol when replacing either 16: 0 or 18: 0 in gerbils. *The Journal of nutritional biochemistry*. 2003;14(10):584-590.
- 115. Dictenberg JB, Pronczuk A, Hayes KC. Hyperlipidemic effects of trans fatty acids are accentuated by dietary cholesterol in gerbils. *The Journal of nutritional biochemistry*. 1995;6(7):353-361.
- 116. Mercer NJ, Holub BJ. Response of free and esterified plasma cholesterol levels in the Mongolian gerbil to the fatty acid composition of dietary lipid. *Lipids*. 1979;14(12):1009-1014.
- 117. Nicolosi RJ, Marlett JA, Morello AM, Flanagan SA, Hegsted DM. Influence of dietary unsaturated and saturated fat on the plasma lipoproteins of Mongolian gerbils. *Atherosclerosis*. 1981;38(3-4):359-371.
- 118. Andersen DB, Holub BJ. Effects of dietary cholesterol level and type of dietary carbohydrate on hepatic and plasma sterols in the gerbil. *Canadian Journal of Physiology and Pharmacology*. 1982;60(7):885-892.
- 119. Lee CM, Boileau AC, Boileau TWM, et al. Review of Animal Models in Carotenoid Research. *The Journal of Nutrition*. 1999;129(12):2271-2277. doi:10.1093/jn/129.12.2271
- 120. Forsythe III WA. Comparison of dietary casein or soy protein effects on plasma lipids and hormone concentrations in the gerbil (Meriones unguiculatus). *The Journal of nutrition*. 1986;116(7):1165-1171.
- 121. Tasker TE, Potte SM. Effects of dietary protein source on plasma lipids, HMG CoA reductase activity, and hepatic glutathione levels in gerbils. *The Journal of Nutritional Biochemistry*. 1993;4(8):458-462.
- 122. Boquist L. Obesity and pancreatic islet hyperplasia in the Mongolian gerbil. *Diabetologia*. 1972;8(4):274-282.
- 123. Ying HZ, Liu YH, Yu B, Wang ZY, Zang JN, Yu CH. Dietary quercetin ameliorates nonalcoholic steatohepatitis induced by a high-fat diet in gerbils. *Food and Chemical Toxicology*. 2013;52:53-60.
- 124. Li W, Shi QJ, Guo HG, Lou Q, Lu LQ, Sa XY. Dynamic analysis of the pathogenesis and biochemistry of nonalcoholic fatty liver disease in gerbils. *Chinese Journal of Laboratory Animal Science*. 2011;(21):44-48.