

# Mean difference in live-weight per incremental difference in body condition score estimated in multiple sheep breeds and crossbreds

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Body condition score (BCS) is a subjective assessment of the proportion of body fat an animal possesses and is independent of frame size. There is a growing awareness of the importance of mature animal live-weight given its contribution to the overall costs of production of a sector. Because of the known relationship between BCS and live-weight, strategies to reduce live-weight could contribute to the favouring of animals with lesser body condition. The objective of the present study was to estimate the average difference in live-weight per incremental change in BCS, measured subjectively on a scale of 1 to 5. The data used consisted of 19 033 BCS and live-weight observations recorded on the same day from 7556 ewes on commercial and research flocks; the breeds represented included purebred Belclare (540 ewes), Charollais (1484 ewes), Suffolk (885 ewes), Texel (1695 ewes), Vendeen (140 ewes), as well as, crossbreds (2812 ewes). All associations were quantified using linear mixed models with the dependent variable of live-weight; ewe parity was included as a random effect. The independent variables were BCS, breed (n = 6), stage of the interlambing interval (n = 6; pregnancy, lambing, pre-weaning, at weaning, post-weaning and mating) and parity (1, 2, 3, 4 and 5 + ). In addition, two-way interactions were used to investigate whether the association between BCS and live-weight differed by parity, a period of the inter-lambing interval or breed. The association between BCS and live-weight differed by parity, by a period of the inter-lambing interval and by breed. Across all data, a one-unit difference in BCS was associated with 4.82 (SE = 0.08) kg liveweight, but this differed by parity from 4.23 kg in parity 1 ewes to 5.82 kg in parity 5 + ewes. The correlation between BCS and live-weight across all data was 0.48 (0.47 when adjusted for nuisance factors in the statistical model), but this varied from 0.48 to 0.53 by parity, from 0.36 to 0.63 by stage of the inter-lambing interval and from 0.41 to 0.62 by breed. Results demonstrate that consideration should be taken of differences in BCS when comparing ewes on live-weight as differences in BCS contribute quite substantially to differences in live-weight; moreover, adjustments for differences in BCS should consider the population stratum, especially breed.

Keywords: regression, parity, ovine, correlation, lambing interval

# Implications

Results from the present study highlight the importance of measuring body condition score (BCS) concurrently with ewe live-weight when attempting to characterise ewes, thereby enabling differences in BCS to be accounted for in any evaluation of ewe or flock credentials. Moreover, results from the present study highlight that live-weight adjustment for BCS should be undertaken by breed, parity and period of the inter-lambing interval. The impact is more informative metrics for benchmarking ewes or flocks but also more pertinent genetic evaluations.

## Introduction

The necessity for more efficient and environmentally benign ruminant production systems has intensified interest in the monitoring and inter-flock benchmarking of efficiencyrelated key performance indicators. Because of the known large contribution of mature animals to the overall feed expenditure of the respective animal sector (Ferrell and Jenkins, 1985; Montaño-Bermudez *et al.*, 1990), the credentials of the mature flock are coming under ever-increasing scrutiny. Moreover, the often cited association between animal (metabolic) live-weight and maintenance requirements (National Research Council, 2001) has resulted in mature animal live-weight becoming a trait that is receiving special attention as a key performance indicator. The outcome is

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that mature ewe live-weight is being used as a proxy to compare not only individual animals but also flocks on expected feed requirements.

The contribution of BCS to differences in live-weight has already been documented in both dairy (Berry et al., 2006 and 2011) and beef cows (Drennan and Berry, 2006), with BCS being positively correlated with live-weight and explaining between 6% and 41% of its variability. Studies in mature ewes suggest stronger correlations than in cattle between BCS and live-weight (0.77 to 0.89; Sanson et al., 1993; Treacher and Filo, 1995; Sezenler et al., 2011; Morel et al., 2016) with BCS explaining between 60% and 79% of its variability. Thus, when comparing animals or flocks on live-weight, some adjustment should be made for differences in BCS; by not doing so, thinner ewes may, on average, be perceived to be more efficient and thus favoured. The implications on health and fertility of poor BCS in mature females is well established in sheep (Corner-Thomas et al., 2015) and cattle (Berry et al., 2007a and 2007b; Roche et al., 2007).

Although the mean live-weight associated with each BCS unit, for subsequent use as adjustment factors, is known in dairy cattle (Berry et al., 2006 and 2011), such adjustment factors are not well known in sheep. The BCS adjustment factors that do exist in sheep have originated either from small experimental studies (14 ewes - Sanson et al., 1993; 84 ewes - Treacher and Filo, 1995; 35 ewes - Frutos et al., 1997; 156 ewes - Sezenler et al., 2011; 28 ewes - Morel et al., 2016) or from breeds less common in temperate regions (Treacher and Filo, 1995; Frutos et al., 1997; Sezenler et al., 2011). Furthermore, differences exist among sheep studies in the BCS scale used; Sanson et al. (1993) used a scale of 1 to 9, Sezenler et al. (2011) used a scale of 0 to 5, whereas both Treacher and Filo (1995) and Morel et al. (2016) used a scale of 1 to 5. The objective of the present study was to quantify the mean live-weight associated with each incremental change in BCS (scale 1 to 5) and determine if the extent of this association differed by parity, by a period of the inter-lambing interval and by breed. The breeds included in the present study were: purebred Belclare, Charollais, Suffolk, Texel and Vendeen as well as crossbreds.

## **Material and methods**

## Data

A total of 36 424 live-weight and BCS records from 25 246 ewes collected from 18 flocks between the years 2013 and 2017 (inclusive) were available. The data originated from two main sources: 16 commercial flocks that participate in the national sheep breeding programme operated by Sheep Ireland (http://www.sheep.ie) and two research flocks. Ewe BCS was evaluated by trained assessors on a 1 (emaciated)to 5 (over fat)-point scale (Jefferies, 1961) in increments of one. Ewe live-weight was recorded using electronic scales. Ewe live-weight was defined as the weight of a female who had at least one recorded lambing event. Only recorded ewe live-weights between 45 and 120 kg were retained. Only records where ewe live-weight and BCS were recorded on the same day were retained for analysis.

Data were also available on ewe parity and breed composition. The breed was categorised as: purebred Belclare, Charollais, Suffolk, Texel and Vendeen; all other ewes were crossbred ewes. Ewe parity was categorised as 1, 2, 3, 4 or  $\geq$  5. Inter-lambing interval was defined based on six key production stages: pregnancy (i.e. 100 to 30 days prelambing), lambing (i.e. 20 days pre-lambing to 20 days postlambing), pre-weaning (i.e. 201 to 80 days post-lambing), weaning (i.e. 81 to 130 days post-lambing), post-weaning (i.e. 131 to 180 days post-lambing) and mating (i.e. 181 to 280 days post-lambing). Ewe live-weight during pregnancy was adjusted for the weight of the foetus using the formulas of Wheeler et al. (1971); gestation length was assumed to be 147 days. The adjusted live-weight was used in all subsequent analyses. Contemporary group was defined as flockby-date of weighing, and only contemporary groups with at least 10 records were considered further. The final data set used in the analysis consisted of 19033 records for BCS and live-weight from 7556 ewes. The number of records per parity, per inter-lambing period and per breed is in Tables 1–3, respectively.

## Statistical analyses

The Pearson correlation between BCS and live-weight was estimated across all data and also within each stratum of parity, inter-lambing interval and breed. The correlation between BCS and live-weight across all data was also estimated following adjustment for the class effects of contemporary group, parity, inter-lambing interval and breed. Correlations within each stratum of parity, inter-lambing interval and breed were also estimated following adjustment for the previously mentioned terms except the term representing the stratum under investigation. Whether the estimated correlations differed from each other were based on Fisher's *r*-to-*z* transformation. All regression analyses were undertaken using a mixed model in ASreml (Gilmour *et al.*, 2009) with

**Table 1** Number of records (n), mean and standard deviation for liveweight and body condition score (BCS) in sheep as well as both the regression (b) of live-weight on BCS for each period of the interlambing interval and the raw correlation ( $r_{raw}$ ) between BCS and liveweight and the correlation following adjustment for fixed effects in the model ( $r_{adj}$ )

		Live-weight		BCS				
Parity	n	Mean	SD	Mean	SD	<i>b</i> (SE)	<b>r</b> <sub>raw</sub>	<i>r</i> <sub>adj</sub>
1 2 3 4	5155 3336 2287	77.85 79.60 80.75	11.53 11.98 12.21	3.69 3.66 3.59	0.70 0.74 0.76	4.23 (0.13) <sup>a</sup> 4.65 (0.13) <sup>b</sup> 5.02 (0.15) <sup>c</sup> 5.29 (0.18) <sup>cd</sup>	0.48 <sup>b</sup> 0.53 <sup>a</sup> 0.50 <sup>ab</sup>	0.47 <sup>b</sup> 0.50 <sup>cd</sup> 0.50 <sup>bd</sup>
<del>-</del> 5 +	1774	79.47	12.71	3.45	0.81	5.82 (0.20) <sup>d</sup>	0.49 <sup>ab</sup>	0.49 <sup>bo</sup>

<sup>a,b,c,d</sup>Different superscripts within column signify differences at P < 0.05.

**Table 2** Number of records (n), mean and standard deviation for live-weight and body condition score (BCS) in sheep as well as both the regression (b) of live-weight on BCS for each period of the inter-lambing interval and the raw correlation ( $r_{raw}$ ) between BCS and live-weight and the correlation following adjustment for fixed effects in the model ( $r_{adj}$ )

Periods	n	Live-weight		BCS				
		Mean	SD	Mean	SD	<i>b</i> (SE)	<i>r</i> <sub>raw</sub>	r <sub>adj</sub>
Pregnancy	2743	78.77	12.54	3.66	0.58	4.90 (0.20) <sup>a</sup>	0.36 <sup>a</sup>	0.38 <sup>a</sup>
Lambing	396	79.43	14.44	3.71	0.76	6.26 (0.50) <sup>b</sup>	0.41 <sup>abe</sup>	0.43 <sup>ab</sup>
Pre-weaning	5576	75.26	12.65	3.50	0.71	4.82 (0.14) <sup>a</sup>	0.43 <sup>b</sup>	0.46 <sup>b</sup>
Weaning	3345	74.35	12.88	3.60	0.82	4.65 (0.16) <sup>a</sup>	0.54 <sup>c</sup>	0.48 <sup>b</sup>
Post-weaning	2348	76.11	13.39	3.69	0.81	6.87 (0.22) <sup>c</sup>	0.63 <sup>d</sup>	0.57 <sup>c</sup>
Mating	4625	76.26	10.50	3.64	0.67	4.07 (0.14) <sup>d</sup>	0.47 <sup>e</sup>	0.48 <sup>b</sup>

 $\overline{a,b,c,d,e}$  Different superscripts within column signify differences at P < 0.05.

**Table 3** Number of records (n), mean and standard deviation for live-weight and body condition score (BCS) in sheep as well as both the regression (b) of live-weight on BCS for each period of the inter-lambing interval and the raw correlation ( $r_{raw}$ ) between BCS and live-weight and the correlation following adjustment for fixed effects in the model ( $r_{adj}$ )

Breeds	n	Live-weight		BCS				
		Mean	SD	Mean	SD	<i>b</i> (SE)	<i>r</i> <sub>raw</sub>	<i>r</i> <sub>adj</sub>
Belclare	552	70.73	11.88	4.00	0.69	6.36 (0.57) <sup>c</sup>	0.42 <sup>a</sup>	0.40 <sup>a</sup>
Charollais	1484	82.84	13.51	4.12	0.73	8.67 (0.32) <sup>b</sup>	0.54 <sup>b</sup>	0.54 <sup>b</sup>
Suffolk	1660	85.18	14.31	3.75	0.71	6.94 (0.28) <sup>c</sup>	0.57 <sup>b</sup>	0.57 <sup>b</sup>
Texel	2625	79.65	12.06	3.97	0.71	5.15 (0.23) <sup>a</sup>	0.41ª	0.43 <sup>a</sup>
Vendeen	140	78.96	14.63	4.11	0.79	9.79 (0.97) <sup>b</sup>	0.62 <sup>b</sup>	0.62 <sup>b</sup>
Crossbreds	12 572	73.48	10.88	3.42	0.65	4.16 (0.09) <sup>d</sup>	0.43 <sup>a</sup>	0.43 <sup>a</sup>

<sup>a,b,c,d</sup>Different superscripts within column signify differences at P < 0.05.

live-weight included as the dependent variable and ewe parity considered as random. Initially, the association between BCS (treated as a continuous variable) and live-weight was estimated across all data at once; fixed effects included in this model were the contemporary group of flock-date of recording, parity, inter-lambing interval and breed. In subsequent analyses, a two-way interaction between BCS with parity, interlambing interval or breed was included to determine if the association between BCS and live-weight differed by stratum. Higher order regressions on BCS were also investigated to determine if non-linear associations between live-weight and BCS existed.

# **Results and discussion**

Mean (standard deviation in parenthesis) BCS and liveweight of all ewes in the data set was 3.6 (0.72) units and 76.0 (12.40) kg, respectively. Parity, inter-lambing interval and breed were all associated (P < 0.001) with live-weight. The correlation between BCS and live-weight in the entire data set was 0.48, implying that 23% of the variability in live-weight was explained by differences in BCS; when adjusted for contemporary group, parity, inter-lambing interval and breed, the correlation was 0.47. Treacher and Filo (1995) reported a stronger correlation of 0.77 between live-weight and BCS in 84 Awassi ewes, whereas Sanson *et al.* (1993) also reported a stronger correlation of 0.89 in 14 mature western-range ewes at slaughter. Across all ages and breeds, Sezenler et al. (2011) reported strong correlations of 0.73 to 0.82 between BCS and live-weight in 156 sheep of three indigenous sheep breeds in Turkey. In a population of 28 mixed-aged Romney-cross ewes, Morel et al. (2016) reported a correlation of 0.81 between BCS and live-weight. Hence, the raw correlation of 0.48 between BCS and live-weight observed in the present study across all data is considerably weaker than observed in other sheep populations, although it is similar to the correlation of 0.49 and 0.55 between BCS and live-weight reported in dairy cows (Berry et al., 2006 and 2011). One possible reason for the weaker correlation between BCS and live-weight in the present study may have been due to several (trained) personnel being used to assess BCS in the present study; although details on the number of assessors used in other studies are not always provided, their relatively small population size over a short period of time suggests one or just a few assessors.

The regression coefficient of live-weight on BCS for the entire data set in the present study was 4.82 (SE = 0.081), implying that a one-unit difference in BCS was associated with, on average, 4.82 kg live-weight. Although a quadratic regression coefficient of live-weight on BCS was statistically significant, the trajectory of the mean live-weight per unit change in BCS was almost identical when BCS was included as a linear or a quadratic term; hence only the solutions from

a model with BCS included as a linear term are discussed further. Treacher and Filo (1995) using a population of 84 Awassi ewes reported a regression coefficient of 11.8 kg liveweight per unit BCS on a scale of 1 to 5, which is more than twice that observed in the present study. However, if the covariance structure among records within ewe was not accounted for in the present study, and instead only a fixed effects model was used, the regression coefficient of liveweight on BCS was 7.25 (SE = 0.10).

# Parity

The association between BCS and live-weight differed (P < 0.001) by parity; no such investigation appears to exist in sheep, although Sezenler et al. (2011) did investigate if the association differed by age of the ewe. However, the association between BCS and live-weight has been reported to differ by parity in dairy cattle (Berry et al., 2006 and 2011). The raw mean live-weight and BCS for each parity as well as the correlation between BCS and live-weight by parity and the regression coefficient of live-weight on BCS by parity are provided in Table 1. Mean BCS and live-weight both increased with parity up to parity 3 or 4, respectively, after which they declined. The raw correlation between BCS and live-weight varied little among parities ranging from 0.48 (parity 2) to 0.53 (parity 1 and 3); although some of these correlations differed (P < 0.05) from each other, the biological significance of this difference is small. The correlation between BCS and live-weight, following adjustment for the contemporary group, inter-lambing interval and breed, varied from 0.42 (parity 1) to 0.50 (parity 3 and 4). The regression coefficient of live-weight on BCS for each parity varied from 4.23 (parity 1) to 5.82 (parity 5). The correlations reported between live-weight and BCS at breeding in three indigenous breeds of sheep in Turkey varied from 0.66 to 0.84 with age of ewe (Sezenler et al., 2011); the correlations between live-weight and BCS at breeding from ages 2 to 4 years varied from just 0.83 to 0.84 implying little difference by age of ewe (Sezenler *et al.*, 2011) similar to that observed in the present study. Moreover, while Sezenler et al. (2011) did not undertake any formal statistical test of whether these correlations actually differed from each other, our calculations reveal that in fact their correlations did not differ (P > 0.05) from each other, although (lack of) statistical power may have contributed to type II errors. The regression of live-weight on BCS at breeding in the same three breeds of indigenous Turkish sheep varied from 5.4 to 6.8 (BCS scale of 0 to 5; Sezenler et al., 2011) with no consistent trend of regression coefficients by ewe age in years; no standard errors of the regression coefficients were provided to facilitate calculation as to whether the coefficients truly differed from each other. The correlation between live-weight and BCS in Irish and New Zealand dairy cows by parity varied from 0.51 to 0.59 (Berry et al., 2011) and from 0.49 to 0.63 (Berry et al., 2006), respectively, with many of them being different (P < 0.05) from each other.

# Period of inter-lambing interval

The association between live-weight and BCS differed (P < 0.001) by a period of the inter-lambing interval and both the correlations and regression coefficients are summarised in Table 2. The correlation between BCS and live-weight varied from 0.36 (during pregnancy) to 0.63 (post-weaning); the correlation between BCS and live-weight, following adjustment for the contemporary group, parity and breed, varied from 0.38 (during pregnancy) to 0.57 (post-weaning). The regression of live-weight on BCS by period of the interlambing interval varied from 4.07 at mating to 6.87 postweaning (Table 2). Across all data, Sezenler et al. (2011) reported correlations between BCS and live-weight in sheep at breeding, at lambing and weaning of 0.82 (n = 156), 0.73 (n=152) and 0.75 (n=135), respectively; analysis of the provided data revealed that none were actually different (P > 0.05) from each other. The regression coefficients of live-weight on BCS (scale 0 to 5) by stage of the interlambing interval evaluated by Sezenler et al. (2011) varied from 6.77 to 7.07, but no standard errors were provided.

In their analysis of 925 records from 299 beef cows, Drennan and Berry (2006) reported a range in correlations between BCS and live-weight of between 0.24 to 0.51 by period of the year; given the seasonal calving system operated in Irish beef herds (Berry and Evans, 2014), period of the year is synonymous with inter-calving interval. In their analysis of Irish Holstein-Friesian dairy cows, Berry *et al.* (2011) reported a range of correlations between BCS and liveweight of between 0.50 and 0.59 by stage of inter-calving interval; a range of 0.36 to 0.48 was reported by Berry *et al.* (2006) in New Zealand Holstein-Friesian dairy cows across stages of the inter-calving interval.

# Breed

Breed in the present study was used as a proxy for frame size; the Suffolk is known to be a relatively large framed animal, whereas the Belclare and Vendeen would be regarded as smaller-framed breeds. Because BCS is a measure of subcutaneous fat, each unit increase in BCS is therefore expected to represent a greater mass of fat in larger-framed animals. This hypothesis was somewhat substantiated with the regression of live-weight on BCS varying from 5.15 and 6.36 in the smaller-framed Texel and Belclare, respectively, up to 6.94 and 8.67 in the larger-framed Suffolks and Charollais, respectively. Furthermore, the correlation between BCS and live-weight per breed varied from 0.41 (Texel) to 0.62 (Vendeen); the correlation between BCS and live-weight, following adjustment for the contemporary group, parity and inter-lambing interval, varied from 0.40 (Belclare) to 0.62 (Vendeen). Sezenler et al. (2011) also estimated the correlations between live-weight and BCS in three breeds of sheep, but calculations based on the information provided in that study revealed no significant difference in the correlation among breeds. Nonetheless, considerable inter-breed differences in the regression coefficients of live-weight on BCS were evident in the present study and should be considered especially in multi-breed genetic evaluations where one of the goal traits may be mature ewe live-weight.

# Conclusions

The expected difference in live-weight between a BCS of 2 units and a BCS of 4 units (scale 1 to 5) was 9.64 kg in the present study when estimated across the entire data set; this represents 13% of the mean live-weight of the entire population. Moreover, the standard deviation in BCS across the entire data set was 0.72 units. Hence, large variability exists in BCS, and the impact on live-weight is substantial. Therefore any strategy, genetic or otherwise, that advocates lighter ewes should consider differences in BCS to avoid simply promoting thinner animals. The results from the present study indicate that BCS should ideally be measured concurrent with any weighing of animals and account should be taken of differences in BCS in any evaluation of ewe or flock characteristics. Moreover, the adjustment factors, either in the statistical model of analysis or through preadjustment factors, should be undertaken by breed, by parity and by a period of the inter-lambing interval as appropriate.

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#### **Declaration of interest**

The authors declare no conflict of interest.

#### **Ethics statement**

The data used in the present study originated from an already existing database.

#### Software and data repository resources

The data sets and programmes used in the current study are available from the corresponding author on reasonable request.

#### References

Berry DP, Buckley F and Dillon P 2011. Relationship between live weight and body condition score in Irish Holstein-Friesian dairy cows. Irish Journal of Agricultural and Food Research 50, 141–147.

Berry DP and Evans RD 2014. Genetics of reproductive performance in seasonal calving beef cows and its association with performance traits. Journal of Animal Science 92, 1412–1422.

Berry DP, Lee JM, Macdonald KA and Roche JR 2007a. Body condition score and body weight effects on dystocia and stillbirths and consequent effects on postcalving performance. Journal of Dairy Science 90, 4201–4211.

Berry DP, Lee JM, Macdonald KA, Stafford K, Matthews L and Roche JR 2007b. Associations among body condition score, body weight, somatic cell count, and clinical mastitis in seasonally calving dairy cattle. Journal of Dairy Science 90, 637–648.

Berry DP, Macdonald KA, Penno JW and Roche JR 2006. Association between body condition score and liveweight in pasture-based Holstein-Friesian dairy cows. Journal of Dairy Research 73, 487–491.

Corner-Thomas RA, Hickson RE, Morris ST, Back PJ, Ridler AL, Stafford KJ and Kenyon PR 2015. Effects of body condition score and nutrition in lactation on twin-bearing ewe and lamb performance to weaning. New Zealand Journal of Agricultural Research 58, 156–169.

Drennan MJ and Berry DP 2006. Factors affecting body condition score, live weight and reproductive performance in spring-calving suckler cows. Irish Journal of Agricultural and Food Research 45, 25–38.

Ferrell CL and Jenkins TG 1985. Cow type and the nutritional environment: nutritional aspects. Journal of Animal Science 61, 725–741.

Frutos P, Mantecón AR and Giráldez FJ 1997. Relationship of body condition score and live weight with body composition in mature Churra ewes. Animal Science 64, 447–452.

Gilmour AR, Gogel B, Cullis BR and Thompson R 2009. ASReml user guide release 3.0. VSN International Ltd, Hemel Hempstead, UK.

Jefferies BC 1961. Body condition scoring and its use in management. Tasmanian Journal of Agriculture 32, 19–21.

Montaño-Bermudez M, Nielsen MK and Deutscher GH 1990. Energy requirements for maintenance of crossbred beef cattle with different genetic potential for milk. Journal of Animal Science 68, 2279–2288.

Morel PCH, Schreurs NM, Corner-Thomas RA, Greer AW, Jenkinson CMC, Ridler AL and Kenyon PR 2016. Live weight and body composition associated with an increase in body condition score of mature ewes and the relationship to dietary energy requirements. Small Ruminant Research 143, 8–14.

National Research Council (NRC) 2001. Nutrient requirements of dairy cattle, 7th revised edition. National Academy Press, Washington, DC, USA.

Roche JR, MacDonald KA, Burke CR, Lee JM and Berry DP 2007. Associations among body condition score, body weight and reproductive performance in seasonal-calving dairy cattle. Journal of Dairy Science 90, 376–391.

Sanson DW, West TR, Tatman WR, Riley ML, Judkins MB and Moss GE 1993. Relationship of body composition of mature ewes with condition score and body weight. Journal of Animal Science 71, 1112–1116.

Sezenler T, Ozder M, Yildirir M, Ceyhan A and Yuksel MA 2011. The relationship between body weight and body condition score some indigenous sheep breeds in Turkey. Journal of Animal and Plant Science 21, 443–447.

Treacher TT and Filo S 1995. Relationships between fat depots and body condition score or live weight in Awassi ewes. Options Méditerranéennes Série A, Séminaires Méditerranéens 27, 13–17.

Wheeler JL, Reardon TF, Hedges DA and Rocks RL 1971. The contribution of the conceptus to weight change in pregnant Merino ewes at pasture. Journal of Agricultural Science 76, 347–353.