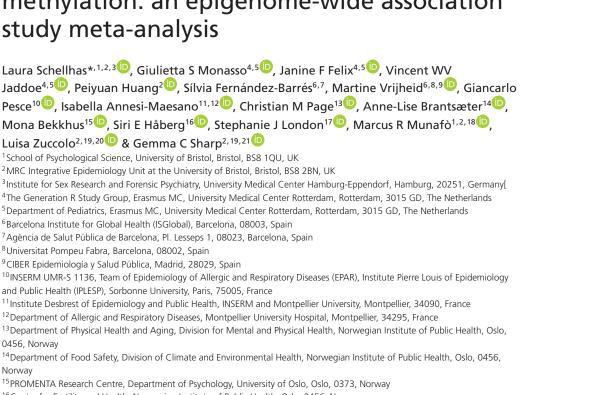
Maternal caffeine consumption during pregnancy and offspring cord blood DNA methylation: an epigenome-wide association study meta-analysis



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Background: Prenatal caffeine exposure may influence offspring health via DNA methylation, but no large studies have tested this. Materials & methods: Epigenome-wide association studies and differentially methylated regions in cord blood (450k or EPIC Illumina arrays) were meta-analyzed across six European cohorts (n = 3725). Differential methylation related to self-reported caffeine intake (mg/day) from coffee, tea and cola was compared with assess whether caffeine is driving effects. Results: One CpG site (cg19370043, PRRX1) was associated with caffeine and another (cg14591243, STAG1) with cola intake. A total of 12–22 differentially methylated regions were detected with limited overlap across caffeinated beverages. Conclusion: We found little evidence to support an intrauterine effect of caffeine on offspring DNA methylation. Statistical power limitations may have impacted our findings.

Plain language summary: Current guidelines recommend pregnant women to limit caffeine intake to less than 200 mg daily, even though there is no clear proof of its effects on human development. A biological explanation for how exposure to caffeine during pregnancy influences development would help clarify if recommended limits are justified. An epigenetic mechanism, called DNA methylation (DNAm), has been

Medicine

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suggested as a potential biological explanation for how caffeine intake during pregnancy influences health development. DNAm can switch genes 'on' or 'off' in response to environmental influences and therefore act as a bridge between genes and the environment. Studies have found that smoking during pregnancy is connected to over 6000 changes in DNAm at birth, with lasting effects into adulthood. To explore the link between caffeine intake during pregnancy and DNAm at birth, we analyzed data from 3725 mother-child pairs living in different European countries. We looked at effects from coffee, tea and cola intake during pregnancy on children's DNAm at birth. We found one change in DNAm to be connected to total caffeine and another to cola consumption during pregnancy. These few connections do not provide convincing evidence that caffeine intake during pregnancy impacts children's DNAm at birth. However, because mothers in our study consumed little caffeine, it is possible that results would be different in studies with participants consuming high amounts of caffeine during pregnancy. Potentially, our study did not include enough people to find very small changes in DNAm that are connected to caffeine consumption during pregnancy.

Tweetable abstract: EWAS meta-analysis of six European cohorts finds no support for an intrauterine effect of caffeine on DNA methylation at birth. Associations are likely driven by diverse confounding structures of caffeinated drinks, not caffeine itself.

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There is growing public and research interest about the effect of caffeine consumption during pregnancy on offspring health. Throughout pregnancy, the metabolic rate of caffeine gradually decreases and in the second and third trimester the half-life of caffeine can be up to four-times longer than outside of pregnancy [1]. Due to pregnancyrelated changes to the caffeine metabolism and the potential of caffeine readily crossing the placenta barrier, the European Food Safety Authority states "... *unborn children to be the most vulnerable group for adverse effects of caffeine among the general population*" [2]. Current pregnancy guidelines for caffeine intake during pregnancy [3]. These guidelines are based on evidence from observational study designs that have found associations with an increased risk for low birth weight [4–8], small for gestational age [4,9] and childhood overweight [10]. However, observational studies are likely affected by selection, measurement and confounding biases [11]. Caffeine consumption is a complex phenotype that is culturally bound, with different caffeinated drinks showing varying confounding structures, even across countries that are perceived to be culturally similar (e.g., UK and The Netherlands) [12].

DNA methylation (DNAm) is an epigenetic mechanism that has been proposed to link prenatal caffeine exposure to later health outcomes in offspring [13]. In this epigenetic modification, a methyl group is added to a CpG site in the genome [14]. DNAm undergoes profound changes during embryonic development, making it a valuable proxy for assessing the quality of the intrauterine environment [15]. Animal studies suggest that prenatal caffeine-induced DNAm may have an effect on offspring growth restriction [16], metabolic, cardiac [17,18], neuroendocrine and hormonal [19–21] development. To our knowledge, only one published study today has investigated prenatal caffeine exposure and offspring DNAm in human cord blood. This small study (n = 378) found only one CpG (cg09460369, nearest gene *RAB2A*) to be differentially methylated in association with prenatal exposure to the caffeine metabolite theobromine [22]. A recent epigenome-wide association study (EWAS) meta-analysis of adult populations found coffee consumption to be associated with own peripheral blood DNAm at 11 CpG sites [23]. Discovering associations between maternal caffeine intake during pregnancy and offspring DNAm could provide initial indication for whether DNAm may pose as a potential biological mechanism explaining the associations found in observational studies.

In this international meta-analysis using data from the Pregnancy and Childhood Epigenetics consortium [15], we explored the association between offspring cord blood DNAm and maternal caffeine consumption during pregnancy across different European cultures (UK, The Netherlands, Spain, France and Norway) using different sources of caffeine (from tea, coffee and cola).

Materials & methods Meta-analysis of EWAS

Participating cohorts

The EWAS meta-analysis included six independent prospective pregnancy and birth cohorts from the Pregnancy and Childhood Epigenetics consortium [15] that had data available on cord blood DNAm and maternal caffeine consumption during pregnancy. The total sample (n = 3731) included two UK-based cohorts (ALSPAC [24,25] and BiB [26,27]), one Dutch (Generation R) [28,29], one Norwegian (MoBa) [30], one Spanish (INMA) [31] and one French (EDEN) [32] cohort. Recruitment periods varied by cohort and took place between the beginning of the 1990s (ALSPAC) and 2010 (BiB). Ethical approval was obtained by local ethics committees and informed consent for the use of data was obtained from all participants. More details about the individual cohorts can be found in the Supplementary Information.

Measurement of maternal caffeine intake during pregnancy

Assessment of maternal caffeine consumption varied by cohort and is described in more detail in Supplementary Information. Generally, mothers self-reported the number of cups they consumed of caffeinated coffee, tea and cola in questionnaires between week 12 and 22 of pregnancy. All cohorts used Food Frequency Questionnaires [33] except for Generation R, which also did not have information on caffeinated cola consumption available. Cups per day were transformed to milligrams of caffeine per day (mg/day), based on the assumption that one standard-sized cup of coffee contains 57 mg, one cup of tea contains 27 mg, and one cup of cola contains 20 mg of caffeine [34]. A continuous total caffeine score was calculated by summing the caffeine content from each caffeinated drink in mg/day (Supplementary Information). Current literature claims that even amounts below the 200 mg/day caffeine limit may have an effect on offspring health and recommends to abstain from caffeine intake during pregnancy [35]. Thus, we investigated, in addition to the continuous score, whether any caffeine exposure (regardless of the amount of caffeine) during pregnancy might have an effect on offspring DNAm. For this analysis, we dichotomized total caffeine into 0 mg/day = none, and >0 mg/day = any.

Measurement of DNAm

Cohorts assessed cord blood DNAm data individually, using their own laboratory methods, quality control and normalisation. DNAm data was sampled using the Illumina Infinium[®] HumanMethylation450 (486,425 probes), except for BiB, which used the Illumina EPIC BeadChip array (Illumina, CA, USA; 866,553 probes). Probes on SNPs, crosshybridizing probes [36] and probes on the sex chromosomes were excluded. In the final meta-analysis, only probes that were available in both arrays (maximum 364,678) were included. Methylation was measured using normalized beta values ranging from 0 to 1, representing 0–100% methylation.

Covariates

To adjust for variation in DNAm driven by cell composition, models were adjusted for cell proportions estimated using the Houseman method with a cord blood reference panel [37,38]. Offspring sex was used to conduct sexstratified sensitivity analyses because sex-specific DNAm differences can still be observed even when restricting analyses to autosomes and removing probes that are crossreactive with sex chromosomes [39]. Another sensitivity analysis was conducted where the unstratified models were additionally adjusted for gestational age at birth. Gestational age could be a mediating factor as it is robustly associated with DNAm [40,41] and there is some evidence that it can be associated with prenatal caffeine exposures [42,43]. To avoid introducing collider bias, we adjusted for gestational age in separate models instead of the main models [44]. To adjust for possible technical variation, all cohorts generated 20 surrogate variables and included them in models – as is standard practice in the field [45].

Each model contained the following covariates [45,46]: an ordinal measure representing maternal education as a proxy for socioeconomic position, maternal age in years, maternal BMI (kg/m²), a binary measure of maternal smoking during pregnancy (e.g., in ALSPAC: 0 = no smoking or giving up smoking during the first trimester; 1 = smoking after the first trimester) and a binary assessment of parity (1 = one or more previous children; 0 = no previous children). The Supplementary Information describes the classifications of covariates in each cohort.

Statistical analyses

Cohort-specific statistical analyses

Probe-level analysis

The analysis plan and R script is available on GitHub (https://github.com/ammegandchips/Prenatal_Caffeine). Cohorts were asked to exclude multiple pregnancies (e.g., twins) and siblings so that each mother was only included once in the dataset. If cohorts included more than one major ethnic group, they were asked to run the EWAS analysis separately for each group. The EWAS R script included the following: a function to remove probes classified as outliers according to the Tukey method of outlier removal (values <25th percentile - $3 \times$ interquartile range and values >75th percentile + $3 \times$ interquartile range) [47], a function to generate surrogate variables using the R package SVA [48] and a function to run an EWAS of each model using the R package Limma [49]. In a second sensitivity analysis, the binary (any vs none) and total continuous unstratified models were additionally adjusted for gestational age at birth. In a separate sensitivity analysis without gestational age adjustment, the binary (any vs none) and total continuous caffeine models were stratified by offspring's sex. For quality assurance, an independent shadow meta-analysis was conducted by a coauthor of the University of Bristol.

Prior to meta-analyzing summary results from each cohort, quality checks were conducted to ensure that the EWASs were properly conducted and there were no problems with the data, in line with standard practice in the field [45,50] (Supplementary Information).

Differentially methylated regions

The probe level approach was complemented using a regional analysis, which considers DNAm at clusters of neighboring CpG sites throughout the epigenome. This approach is more statistically powerful and arguably more biologically plausible; neighboring CpG sites are assumed to exert similar biological functions. We used the dmrff method [51] to identify differentially methylated regions (DMRs). In this analysis, first candidate DMRs are identified based on the meta-analyzed summary statistic of each CpG site. Candidate DMRs were defined as regions with a minimum of two CpG sites within 500-base pair proximity, which show the same direction of effect and a p-value < 0.05. Second, summary statistics for these candidate DMRs are calculated within each cohort. Last, a meta-analysis is performed based on the cohort DMR summary statistics. The dmrff meta-analysis applies an inverse-variance weighted fixed effects approach that accounts for dependencies between CpG sites [51,52]. Cohorts were supplied with an R script to conduct the DMR analysis using their own data (https://github.com/ammeg andchips/Prenatal_Caffeine/blob/master/dmrff.mat.caff.EWAS.cohorts.r). Probes were annotated to the human reference genome version 19, build 37h using the annotation data available from the R package *meffil* [53].

Meta-analysis

Probe level meta-analysis

Results were meta-analyzed with fixed effect estimates weighted by the inverse of the variance using the software METAL [54]. Multiple testing was accounted for using a 5% false discovery rate [55]. Meta-analyzed results were scrutinized in a similar manner as the individual cohort results and leave-one-cohort-out analysis using the R package *metafor* [56] was performed on the CpGs that showed evidence to be associated with maternal caffeine consumption. Results were deemed to be driven by a single cohort (and therefore to 'fail' the leave-one-out test), if the meta-analysis effect estimate changed direction, moved toward the null by more than 20% or had a CI that included 0 after removal of a single cohort.

DMR meta-analysis

DMR cohort results were meta-analyzed using an inverse-variance weighted fixed effects approach using the *dmrff.meta* function in the dmrff R package [51]. We defined a DMR as a region with at least two CpG sites with the same direction of effect and a Bonferroni adjusted p-value ($P_{Bonferroni}$) < 0.05.

Causal inference & sensitivity analyses

Beverage-specific effects of the meta-analyzed probe-level and DMR results were investigated by comparing the congruence between associations found using different sources of caffeine (that could have different confounding structures). Whereas high congruence between results of the different caffeine models (in terms of CpG site hits and/or genes annotated to CpG sites found in each model) would provide evidence for caffeine being the causal agent driving effects, beverage-specific effects would indicate that factors other than caffeine are driving associations.

Cohort (n)	Weeks of gestation of dietary assessment	Mean total daily caffeine intake (SD)	n users (%) >200 mg of daily caffeine intake [†]	M daily intake of coffee (SD)	M daily intake of tea (SD)	M daily intake of cola (SD)
ALSPAC(n = 729)	18	135 (94)	197 (27)	52 (69)	72 (58)	3 (6)
BiB(Asian; n = 353)	26–28	49 (47)	5 (2)	11 (33)	41 (34)	12 (20)
BiB(White European; n = 306)	26–28	112 (105)	50 (19)	46 (65)	66 (63)	15 (22)
Generation R(n = 798)	18–25	115 (96)	132 (20)	118 (78)	57 (62)	Not available
INMA(n = 378)	12	111 (129)	26 (8)	79 (120)	25 (46)	7 (12)
EDEN(n = 162)	24–28	37 (43)	1 (<1)	25 (42)	6 (15)	6 (9)
MoBa1(n = 999)	22	105 (106)	108 (11)	70 (106)	18 (27)	14 (27)
Total and % or M and SD [‡] (n = 3725)	-	85 (82)	519 (14)	46 (65)	26 (35)	5 (11)

[†]Mothers were grouped as users of caffeine if they indicated to consume more than zero cups of coffee, tea or cola. Caffeine content in milligrams per day. [‡]In the Total row, average caffeine content was calculated by weighting by the inverse variance for each cohort. M: Mean; SD: Standard deviation.

To find out which gene pathways are linked to the CpG sites of the caffeine-associated DMRs, a gene ontology (GO) analysis was run using the R package *missMethyl* [57]. We tested enrichment of GO categories and Kyoto Encyclopaedia of Genes and Genomes (KEGG) pathways.

Results

Sample characteristics

Maternal caffeine consumption during pregnancy

In all cohorts, most mothers (80–94%) consumed at least some caffeine during weeks 1–28 of gestation, with a weighted mean of 85 mg/day over all cohorts, but with large variation within and between cohorts (weighted average standard deviation = 82 mg/day) (Table 1). Approximately 14% of mothers in the total sample consumed more caffeine than the commonly recommended caffeine limit of 200 mg/day. Across all cohorts, coffee and tea were the most common sources of caffeine, with coffee being the most common source in all cohorts except for the UK-based cohorts ALSPAC and BiB, where the most common source was caffeinated tea (Table 1).

Demographics

A general overview over the demographics of the individual cohorts can be found in Table 2. Except for mothers from BiB, cohorts included slightly more mothers with higher (high school diploma or above) than lower educational attainment. Around 15% of mothers smoked after the second trimester of pregnancy. Mothers who consumed caffeine during pregnancy were about twice as likely to have smoked during pregnancy (20%), compared with mothers who did not consume caffeine during pregnancy (11%) (Supplementary Table 1). Furthermore, mothers who consumed caffeine were more likely to already have children (49%) compared with mothers who did not consume caffeine during pregnancy (33%) (Supplementary Table 1).

Association between maternal caffeine consumption & offspring cord blood DNAm

Results of the quality control checks for the individual cohort and meta-analyzed results can be found in Supplementary Figures 1–18. The overall EWAS meta-analysis results of the caffeine models can be found in Table 3. After adjusting for multiple testing, one CpG site (cg19370043, nearest gene *PRRX1*) was negatively associated with total maternal caffeine consumption (estimate = -2.18×10^{-05} ; 95% CI: -2.98×10^{-05} to -1.37×10^{-05} ; p = 1.32×10^{-07}) and one CpG site (cg14591243, nearest gene *STAG1*) with caffeine consumed from cola in mg/day (estimate = 2.78×10^{-05} ; 95% CI: 2.779×10^{-05} to 2.781×10^{-05} ; p = 5.59×10^{-09}) (Table 3). For the caffeinated cola results, drinking one extra cup of cola per day would be associated with a 0.06% increase in DNAm at cg14591243. Only the total maternal caffeine-associated CpG site survived the leave-one-out analysis. For the cola-associated CpG site, the leave-one-out analysis indicated that MoBa was driving the effect (Supplementary Figures 16 & 17).

Cohort	Country and ancestry	DNA methylation array	n high maternal socioeconomic position [†] (%)	M maternal age (SD)	n maternal smoking (%) [‡]	n parity > 0 (%)	M BMI	M gestational age (weeks)
ALSPAC (n = 729)	UK; northern European	450k	375 (51)	29.79 (4.39)	77 (11)	381 (52)	22.79 (3.63)	39.53 (1.52)
BiB(Asian; n = 353)	UK; Pakistani	EPIC	146 (41)	28.21 (5.37)	9 (3)	249 (71)	25.75 (5.23)	39.17 (1.52)
BiB(White European; n = 306)	UK; northern European	EPIC	125 (41)	26.98 (6.15)	93 (30)	159 (52)	27.10 (6.48)	39.29 (1.88)
Generation R (n = 798)	The Netherlands; northern European	450k	458 (57)	30.15 (4.95)	109 (14)	95 (59)	23.06 (3.64)	40.20 (1.48)
INMA (n = 378)	Spain; southern European	450k	277 (73)	31.55 (4.07)	53 (14)	161 (43)	23.79 (4.44)	41.06 (1.34)
EDEN (n = 162)	France; southern and northern European	450k	113 (70)	31.94 (4.10)	26 (16)	324 (41)	23.52 (4.64)	39.51 (1.33)
MoBa1 (n = 999)	Norway; northern European	450k	761 (76)	29.93 (4.35)	287 (29)	580 (58)	24.02 (4.18)	39.95 (1.56)
Total or M (n = 3725)	_	_	2255 (61)	30.01(4.60)	654 (18)	1949 (52)	23.61 (4.12)	39.94 (1.51)

[†]High maternal socioeconomic position: maternal education \geq high school diploma.

[‡]Continued smoking during pregnancy. Parity = one or more previous pregnancies.

M: Mean; SD: Standard deviation.

Model†	CpGs with false discovery rate-corrected p-value < 0.05	CpGs surviving leave-one-out analysis	Meta-analysis sample size	Genomic inflation factor $(\lambda)^{\ddagger}$
Any vs no caffeine				
All offspring (minimally adjusted) †	0	NA	3731	0.97
All offspring (adjusted for covariates)	0	NA	3731	0.97
Female offspring (adjusted for covariates)	0	NA	1797	0.99
Male offspring (adjusted for covariates)	0	NA	1934	1.00
All offspring (adjusted for covariates and gestational age)	0		3731	0.97
Caffeine in mg/day				
All offspring (minimally adjusted) [†]	33	NA	3731	1.03
All offspring (adjusted for covariates)	1	1 (100%)	3731	1.00
Female offspring (adjusted for covariates)	0	NA	1797	1.00
Male offspring (adjusted for covariates)	0	NA	1934	1.04
All offspring (adjusted for covariates and gestational age)	0		3731	0.99
Caffeine from coffee				
All offspring (adjusted for covariates)	0	NA	2779	1.02
Caffeine from tea				
All offspring (adjusted for covariates)	0	NA	3477	1.00
Caffeine from cola				
All offspring (adjusted for covariates)	1	0	2610	1.00

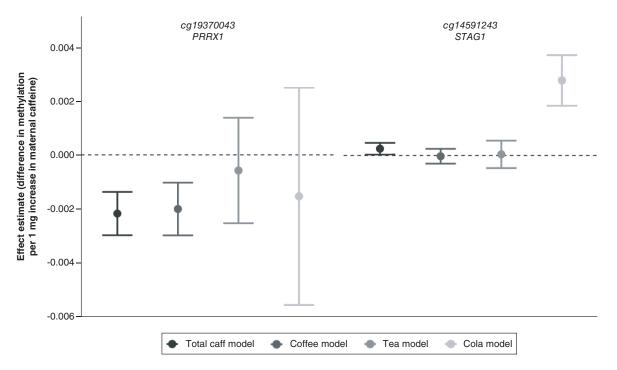
[†]Only adjusted for estimated cell counts and 20 surrogate variables. Covariates: maternal age, maternal smoking, maternal parity, maternal education, maternal BMI, estimated cell counts and 20 surrogate variables.

^{\ddagger}The genomic inflation factor (λ) estimates the extent of bulk inflation of epigenome-wide association study p-values and the excess false positive rate. 1 = no inflation; >1 some evidence of inflation.

NA: Not applicable.

There is a slight deviation in sample sizes between the results and descriptive information (n = 6 participants) because of removal of data in MoBa between 2018 and 2020 due to withdrawal of consent and/or non-Nordic/non-European ancestry.







Beverage-specific effects

Figure 1 displays DNAm at the two CpG sites discovered in the probe-level analysis across the different caffeine models. If these associations were truly driven by caffeine exposure, we would expect to see similar associations of these CpG sites across the different beverage models. The association between DNAm at cg19370043 and maternal total caffeine consumption appears to be mostly driven by coffee and not by caffeine from tea or cola (Figure 1). For coffee intake specifically, the p-value was < 0.05, but the association with coffee did not survive adjustment for multiple testing in the EWAS meta-analysis, probably because of the lower statistical power to detect small effects in the coffee compared with total caffeine analysis (n total caffeine = 3731 vs n coffee = 2779; because the total caffeine model included mothers with missing beverage-specific data). The association between DNAm at cg14591243 appears to be specific to cola consumption rather than general caffeine consumption.

DMR meta-analysis

The regional meta-analysis implemented using dmrff detected 22 DMRs for total maternal caffeine consumption with at least two and a maximum of 15 consecutive CpG sites ($P_{Bonferroni} < 0.05$) (Supplementary Table 2 & Supplementary Figure 19). The strongest evidence was found at a region on chromosome 17, with seven consecutive CpG sites (chr17: 58499679-58499911; estimate = -3.77×10^{-05} ; standard error: 5.02×10^{-06} ; $P_{Bonferroni} = 1.42 \times 10^{-10}$; nearest gene *C17orf64*) (Supplementary Table 2). In the any versus no maternal caffeine consumption model, there was evidence for 11 DMRs (Supplementary Table 3). The strongest evidence was found at a region on chromosome 6, with ten consecutive CpG sites (Chr6:31734147-31734554; estimate = -9.44^{-03} ; standard error: 1.37^{-03} ; $P_{Bonferroni} = 1.93^{-06}$; nearest gene *C6orf27*). DMR analyses from the individual sources of caffeine revealed 12 DMRs for caffeine consumed from coffee (Supplementary Table 4), 18 DMRs for caffeine from tea (Supplementary Table 5) and 14 DMRs for caffeine consumption from cola (Supplementary Table 6) during pregnancy. The analyses from the sex-stratified models showed evidence for total maternal caffeine being associated with 12 DMRs in cord blood in female sex offspring (Supplementary Table 7) and 18 DMRs in male sex offspring (Supplementary Table 8).

For each pairwise combination of models, we calculated the percentage overlap of CpG sites (or closest gene) by dividing the overlap of CpG sites (or genes) between two models by the sum of the models' unique CpG sites/genes (e.g., percentage crossover any caffeine and total caffeine models: $7/[167 + 63 - 7] = 0.03 \times 100 = 3\%$). There was very little overlap in CpG sites (range percentage overlap: 0–12%) or annotated genes (percentage overlapping genes: 0–11%) between the different models (Supplementary Table 9).

Functional analysis of DMRs

A GO analysis was conducted using the R package *missMethyl* [57]. We tested enrichment of GO categories and KEGG pathways. Neither the functional categories defined by GO terms nor any of the KEGG pathways showed evidence for enrichment in genes annotated to CpG sites in the caffeine-associated DMRs (all false discovery rate adjusted p-values > 0.05). Due to the limited number of DMRs available for the functional analysis, insufficient statistical power may have compromised the identification of meaningful enrichment. The top five KEGG pathways and GO terms with the strongest evidence according to the smallest p-values for each list of CpGs in the caffeine-DMRs are available in Supplementary Table 10.

Discussion

Summary & interpretation of findings

We investigated the association of self-reported maternal caffeine consumption during pregnancy with offspring cord blood DNAm using data from six international birth cohorts. For the EWAS meta-analysis, probe-level and regional DMR analyses were applied as hypothesis-free approaches to detect associations between maternal caffeine phenotypes and differential methylation levels in cord blood. We compared caffeine consumption during pregnancy across countries and caffeinated drinks, which reduced the potential for cultural confounding, and analyzed the CpG sites of the maternal caffeine-associated DMRs for their biological function. Results of these analyses show little converging evidence between the associations of the different sources of caffeine, indicating that the associations that we observed are most likely explained by other factors than caffeine exposure.

The probe-level analysis indicated that differences in DNAm at two CpG sites were associated with maternal caffeine intake (one with maternal total caffeine intake, and one with caffeine intake from cola); both showed small effect estimates. According to the genecards database [58], the gene PRRX1, which is annotated to the total caffeine-associated CpG site cg19370043, has been found to be associated with determining mesodermal muscle types by regulating muscle creatine kinase. The gene STAGI, which has been annotated to the cola-associated CpG site cg14591243, has been found to be associated with sister chromatid cohesion during cell division and diseases, including intellectual developmental disorder and Cornelia de Lange syndrome [58]. The coefficients from the regression analyses represent the change in offspring cord blood DNAm at a given CpG site per 1 mg/day increase in maternal caffeine consumption. Putting these results into real-life context, and assuming causality and linearity of effects, if the recommended limit of caffeine consumption during pregnancy were doubled from 200 mg/day to 400 mg/day, this would only be associated with 0.4% reduction in DNAm at cg19370043. This effect size is in line with the small effect sizes found in the EWAS meta-analysis of adult personal caffeine consumption on DNAm by Karabegović and colleagues [23], where an additional cup of coffee (= 57 mg) was associated with a 0.2% decrease in peripheral blood DNAm at a CpG site near AHRR, which would be equivalent to a 0.7% decrease in DNAm per 200 mg/day of caffeine $(0.2\%/57 \text{ mg of caffeine per cup of coffee} \times 200)$. These estimated effect sizes appear to be much smaller than the estimated effect of smoking, which is the lifestyle exposure with the strongest effect on DNAm discovered to date. Sustained smoking during pregnancy was associated with changes of up to \sim 7% decrease in offspring cord blood DNAm at the AHRR gene [46]. As acknowledged by Karabegović et al., because their smoking adjustment did not include the amount of smoking or duration of smoking, the coffee-associated DNAm differences might be explained by residual confounding by smoking [23].

In the regional analyses, we identified 12–22 DMRs for each of the caffeine models. Yet, lack of congruence of associations across models, which was evident in the probe-level and regional analysis, provided evidence for beverage-specific effects instead of the effects being driven by caffeine (which is common to all included beverages).

Strengths & limitations

This was the first large international EWAS meta-analysis investigating associations between offspring DNAm and caffeine from coffee, tea and cola during pregnancy. A major strength of this study is the consideration of the effects of other common sources of caffeine besides coffee. Consumption of the different sources of caffeine might

be differentially socially patterned, allowing capturing a larger spectrum of the caffeine-consuming population. For instance, British and non-European ethnicities consume more caffeinated tea than coffee [12,59]. Further, it is indicated that the main source of caffeine might change during pregnancy, with even habitual coffee drinkers preferring caffeinated tea to coffee during pregnancy [60,61]. Last, in contrast to previous research, this study assessed maternal caffeine consumption through mg/day instead of cups per day, which is a useful approach to isolate the effect of caffeine, and allow comparison between different caffeinated beverages and a more fine-tuned assessment of the effects of different caffeine dosages. We also adjusted for maternal smoking, an important potential confounder in analyses of caffeine intake, along with other potential confounding variables.

The findings should be considered in the light of the following limitations. Caffeine assessment in the metaanalysis relied on self-report, which might be underestimated [62] or underreported during pregnancy because of social stigma around maternal health behaviors [63,64]. However, this would be more obvious for more recent cohorts and less and less likely for older cohorts, such as ALSPAC, due to awareness of the potential toxicity of caffeine in pregnancy emerging only recently. Most of the cohorts of this study were assessed in the beginning of 2000s except for ALSPAC, which was assessed in the beginning of the 1990s. Caffeine consumption across the included cohorts does not show a clear pattern of change over time. There is a lack of nationally representative studies for caffeine consumption during pregnancy but a systematic review of caffeine consumption in the general population found that consumption remained stable between 1997 and 2015 [65]. The questions used to assess caffeine consumption in the cohorts only allowed for rough estimations of maternal caffeine consumption [66], which could reduce power. Generalizability of the results to other populations might be limited by examining only second trimester consumption, the relatively low caffeine intake and the tendency for birth cohorts to enroll more advantaged families [25]. Effects of caffeine exposure during pregnancy on offspring cord blood DNAm were only assessed at regions available on the 450k array, which only covers around 2% of CpG sites of the entire epigenome [67]. Thus, differentially methylated CpGs or regions not covered by the array may have been missed in this study. We only assessed offspring DNAm in blood. There is some evidence suggesting that DNAm levels in blood might be able to proxy for DNAm levels in other tissues, yet we cannot rule out that maternal caffeine during pregnancy might be influencing DNAm differentially in other tissue types [68]. Finally, this study indicates that, if maternal caffeine consumption influences cord blood DNAm, the effect is likely to be small. Although our meta-analysis maximized sample sizes, even larger sample sizes with more variable levels of caffeine consumption may be required to detect small effects of prenatal caffeine exposure on offspring DNAm. Due to the dynamic nature of DNAm, results of EWAS are prone to capture associations of confounding variables [69]. Though attempts were made to reduce confounding, we cannot rule out that confounding influenced the results.

Future research

Future research should aim to use a more accurate assessment of caffeine consumption during pregnancy by considering differing types of coffee, brewing times and cups sizes, and/or assessing biomarkers of caffeine such as plasma concentrations of the caffeine metabolite paraxanthine [70]. Also, future research should investigate the effects of high caffeine consumption on offspring DNAm (e.g., comparison of above vs below the commonly recommended limit of 200 mg/caffeine). Triangulation strategies may be applied to disentangle confounded from causal effects of caffeine exposure during pregnancy on offspring DNAm. These might include further exploring the different confounding structures of various caffeinated beverages and considering individual differences in the maternal metabolism of caffeine. Maternal caffeine entabolism might influence intensity of exposure during pregnancy and might change the effect of caffeine on offspring DNAm. For instance, studies could conduct analyses using genetic variants that account for differences in caffeine metabolism [71] and/or consider prepregnancy caffeine consumption to account for differences in the tolerance to effects of caffeine during pregnancy [66]. Also, more assessments of prenatal paternal caffeine consumption would enable the conduction of negative control analyses to investigate intrauterine effects [72,73], as well as investigating the effects of paternal caffeine consumption prior to pregnancy and its effect on offspring DNAm in its own right [73].

Conclusion

In conclusion, results of this large scale EWAS meta-analysis indicate little evidence for a strong association between maternal caffeine consumption during the second trimester of pregnancy and offspring cord blood DNAm.

Summary points

- This large-scale meta-analysis of epigenome-wide association studies across six European cohorts does not support an intrauterine effect of caffeine on offspring cord blood DNA methylation.
- Lack of overlap between associations with different caffeinated drinks suggest that any (weak) associations were driven by diverse confounding structures of different caffeinated drinks, rather than caffeine *per se*.
- More research is needed to understand the biological mechanisms driving potential effects of caffeine on offspring health.

Supplementary data

To view the supplementary data that accompany this paper please visit the journal website at: www.futuremedicine.com/doi/ suppl/10.2217/epi-2023-0263

Author contributions

L Schellhas and GC Sharp contributed to the study conceptualization and design of the study. L Schellhas conducted the cohort analyses in ALSPAC and BiB, conducted the meta-analysis and quality control analysis under supervision of GC Sharp, and wrote the manuscript. GC Sharp, MR Munafò and L Zuccolo were substantially involved in the interpretation of data for the study and contributed to critically revise the work for important intellectual content. GS Monasso conducted the cohort analysis using Generation R data under supervision of JF Felix. VWV Jaddoe contributed with data acquisition for Generation R. P Huang conducted the shadow meta-analysis. S Fernández-Barrés conducted the analysis in INMA under the supervision of M Vrijheid. G Pesce conducted the analysis in EDEN under the supervision of I Annesi-Maesano. C Page conducted the cohort analysis using data of MoBa. SE Håberg contributed with data acquisition for MoBa. AL Brantsæter generated the caffeine variables in MoBa. M Bekkhus contributed to the study design and generated the outcome measures (psychometric variables) using MoBa data. SJ London contributed with data generation for MoBa and also initiated the Pregnancy and Childhood Epigenetics consortium, of which this study is part. All coauthors reviewed/edited the manuscript and approved the final version.

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BiB: BiB is only possible because of the enthusiasm and commitment of the children and parents in BiB. We are grateful to all the participants, practitioners and researchers who have made BiB happen. 450K DNAm and genotype array data was generated in the Bristol Bioresource Laboratory Illumina Facility, University of Bristol. Ethical approval for the data collection was granted by Bradford Research Ethics Committee (ref. 07/H1302/112). On registration with the study, pregnant mothers gave written informed consent for themselves and on behalf of their child.

Generation R: The Generation R Study is conducted by Erasmus Medical Center, University Medical Center Rotterdam, in close collaboration with the School of Law and Faculty of Social Sciences of the Erasmus University Rotterdam, the Municipal Health Service Rotterdam area, Rotterdam, the Rotterdam Homecare Foundation, Rotterdam and the Stichting Trombosedienst and Artsenlaboratorium Rijnmond (STAR-MDC), Rotterdam. We gratefully acknowledge the contribution of children and parents, general practitioners, hospitals, midwives and pharmacies in Rotterdam. The study protocol was approved by the Medical Ethical Committee of the Erasmus Medical Centre, Rotterdam. Written informed consent was obtained for all participants. The generation and management of the Illumina 450K methylation array data (epigenome-wide association study [EWAS] data) for the Generation R Study was executed by the Human Genotyping Facility of the Genetic Laboratory of the Department of Internal Medicine, Erasmus MC, The Netherlands. We thank M Verbiest, M Jhamai, S Higgins, M Verkerk and L Stolk for their help in creating the EWAS database. We thank A Teumer for his work on the quality control and normalization scripts.

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Ethical conduct of research

The authors state that they have obtained appropriate institutional review board approval or have followed the principles outlined in the Declaration of Helsinki for all human or animal experimental investigations.

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References

Papers of special note have been highlighted as: • of interest; •• of considerable interest

- Grosso LM, Bracken MB. Caffeine metabolism, genetics, and perinatal outcomes: a review of exposure assessment considerations during pregnancy. Ann. Epidemiol. 15(6), 460–466 (2005).
- 2. EFSA Panel on Dietetic Products, Nutrition, Allergies (NDA). Scientific opinion on the safety of caffeine. EFSA J. 13(5), 4102 (2015).
- 3. Reyes CM, Cornelis M. Caffeine in the diet: country-level consumption and guidelines. Nutrients 10(11), 1772 (2018).
- Greenwood DC, Thatcher NJ, Ye J *et al.* Caffeine intake during pregnancy and adverse birth outcomes: a systematic review and dose–response meta-analysis. *Eur. J. Epidemiol.* 29(10), 725–734 (2014).
- This systematic review and meta-analysis included 53 cohort and case-control studies. Findings indicate that caffeine intake was associated with a higher risk of spontaneous abortion, stillbirth, preterm delivery, low birth weight (LBW) and children being small for gestational. However, there was substantial variation in the results across studies and indication for small study bias. The review found no evidence for a threshold below which associations diminished, but effect sizes were modest. The study suggests maintaining existing recommendations for caffeine intake during pregnancy as a precaution.
- 5. Rhee J, Kim R, Kim Y *et al.* Maternal caffeine consumption during pregnancy and risk of low birth weight: a dose-response meta-analysis of observational studies. *PLOS ONE* 10(7), e0132334 (2015).
- 6. Chen L-W, Wu Y, Neelakantan N, Chong MF-F, Pan A, van Dam RM. Maternal caffeine intake during pregnancy is associated with risk of low birth weight: a systematic review and dose-response meta-analysis. *BMC Med.* 12(1), 174 (2014).
- 7. Soltani S, Salari-Moghaddam A, Saneei P *et al.* Maternal caffeine consumption during pregnancy and risk of low birth weight: a dose–response meta-analysis of cohort studies. *Crit. Rev. Food Sci. Nutr.* 63(2), 224–233 (2023).
- This systematic review and meta-analysis included seven cohort studies that investigated maternal caffeine intake during pregnancy and risk for LBW. In five of those seven studies, smoking was adjusted for. The study found evidence for an association between maternal caffeine intake during pregnancy and an elevated risk for LBW, with an increment of 100 mg/day caffeine being associated with an 1.12 increased risk of LBW (95% CI: 1.03–1.22).

- Poole R, Kennedy OJ, Roderick P, Fallowfield JA, Hayes PC, Parkes J. Coffee consumption and health: umbrella review of meta-analyses of multiple health outcomes. *BMJ* 359 (2017).
- 9. Askari M, Bazshahi E, Payande N, Mobaderi T, Fahimfar N, Azadbakht L. Relationship between caffeine intake and small for gestational age and preterm birth: a dose–response meta-analysis. *Crit. Rev. Food Sci. Nutr.* 1–11 (2023).
- 10. Jin F, Qiao C. Association of maternal caffeine intake during pregnancy with low birth weight, childhood overweight, and obesity: a meta-analysis of cohort studies. *Int. J. Obes.* 45(2), 279–287 (2021).
- 11. Hammerton G, Munafo MR. Causal inference with observational data: the need for triangulation of evidence. *Psychol. Med.* 51(4), 563–578 (2021).
- 12. Treur JL, Taylor AE, Ware JJ *et al.* Associations between smoking and caffeine consumption in two European cohorts: smoking and caffeine consumption. *Addiction* 111(6), 1059–1068 (2016).
- •• This study found a positive association between smoking and caffeine consumption in population-based samples from The Netherlands and the UK. Smoking initiation and persistence were linked to higher caffeine consumption, with each additional cigarette per day associated with increased daily caffeine consumption. Smoking was positively associated with coffee consumption and to a lesser extent with cola and energy drinks. Tea consumption showed mixed results.
- 13. Ding Q, Xu Y-M, Lau ATY. The epigenetic effects of coffee. Molecules 28(4), 1770 (2023).
- 14. Bird A. Perceptions of epigenetics. Nature 447(7143), 396-398 (2007).
- Felix JF, Joubert BR, Baccarelli AA et al. Cohort profile: Pregnancy And Childhood Epigenetics (PACE) consortium. Int. J. Epidemiol. 47(1), 22–23u (2018).
- •• The Pregnancy And Childhood Epigenetics consortium conducted a meta-analysis across 13 cohorts, revealing over 6000 differentially methylated CpG sites in newborns associated with maternal smoking during pregnancy. The findings suggest epigenetic mechanisms underlying the effects of maternal smoking during pregnancy with implications for child health and development extending into later childhood.
- 16. Huang J, Zhou S, Ping J *et al.* Role of p53-dependent placental apoptosis in the reproductive and developmental toxicities of caffeine in rodents. *Clin. Exp. Pharmacol. Physiol.* 39(4), 357–363 (2012).
- Fang X, Mei W, Barbazuk WB, Rivkees SA, Wendler CC. Caffeine exposure alters cardiac gene expression in embryonic cardiomyocytes. Am. J. Physiol. Regul. Integr. Comp. Physiol. 307(12), R1471–R1487 (2014).
- 18. Buscariollo DL, Fang X, Greenwood V, Xue H, Rivkees SA, Wendler CC. Embryonic caffeine exposure acts via A1 adenosine receptors to alter adult cardiac function and DNA methylation in mice. *PLOS ONE* 9(1), (2014).
- 19. Ping J, Wang J, Liu L *et al.* Prenatal caffeine ingestion induces aberrant DNA methylation and histone acetylation of steroidogenic factor 1 and inhibits fetal adrenal steroidogenesis. *Toxicology* 321, 53–61 (2014).
- Wu D-M, He Z, Ma L-P, Wang L-L, Ping J, Wang H. Increased DNA methylation of scavenger receptor class B type I contributes to inhibitory effects of prenatal caffeine ingestion on cholesterol uptake and steroidogenesis in fetal adrenals. *Toxicol. Appl. Pharmacol.* 285(2), 89–97 (2015).
- 21. Xu D, Zhang B, Liang G *et al.* Caffeine-induced activated glucocorticoid metabolism in the hippocampus causes hypothalamic–pituitary–adrenal axis inhibition in fetal rats. *PLOS ONE* 7(9), (2012).
- 22. Polinski KJ, Purdue-Smithe A, Robinson SL *et al.* Maternal caffeine intake and DNA methylation in newborn cord blood. *Am. J. Clin. Nutr.* 115(2), 482–491 (2022).
- •• In this small study (n = 378) the association between offspring cord blood DNA methylation and maternal caffeine intake preconception and during pregnancy was investigated. Results showed one association between a caffeine metabolite (theobromine) during pregnancy and cord blood DNA methylation (DNAm) near the *RAB2A* gene. Another association was found between self-reported caffeine intake preconception and DNAm near the *GLIS3* gene.
- 23. Karabegović I, Portilla-Fernandez E, Li Y et al. Epigenome-wide association meta-analysis of DNA methylation with coffee and tea consumption. *Nat. Commun.* 12(1), 2830 (2021).
- This large-scale epigenome-wide association study on coffee and tea consumption combined data from 15 cohort studies and found DNAm signals associated with coffee intake but no associations with tea consumption, suggesting differential epigenetic mechanisms underlying the health effects of these beverages.
- Boyd A, Golding J, Macleod J et al. Cohort profile: the 'Children of the 90s' the index offspring of the Avon Longitudinal Study of Parents and Children. Int. J. Epidemiol. 42(1), 111–127 (2013).
- Fraser A, Macdonald-Wallis C, Tilling K et al. Cohort profile: the Avon Longitudinal Study of Parents and Children: ALSPAC mothers cohort. Int. J. Epidemiol. 42(1), 97–110 (2013).
- 26. Wright J, Small N, Raynor P *et al.* Cohort profile: the Born in Bradford multi-ethnic family cohort study. *Int. J. Epidemiol.* 42(4), 978–991 (2013).
- 27. Raynor P. Born in Bradford Collaborative Group. Born in Bradford, a cohort study of babies born in Bradford, and their parents: protocol for the recruitment phase. *BMC Public Health* 8, 327 (2008).

- 28. Kooijman MN, Kruithof CJ, van Duijn CM *et al.* The Generation R Study: design and cohort update 2017. *Eur. J. Epidemiol.* 31(12), 1243–1264 (2016).
- 29. Kruithof CJ, Kooijman MN, van Duijn CM *et al.* The Generation R Study: biobank update 2015. *Eur. J. Epidemiol.* 29(12), 911–927 (2014).
- Magnus P, Birke C, Vejrup K *et al.* Cohort profile update: the Norwegian Mother and Child Cohort Study (MoBa). *Int. J. Epidemiol.* 45(2), 382–388 (2016).
- Guxens M, Ballester F, Espada M et al. Cohort profile: the INMA Infancia y Medio Ambiente (Environment and Childhood) Project. Int. J. Epidemiol. 41(4), 930–940 (2012).
- 32. Heude B, Forhan A, Slama R *et al.* Cohort profile: the EDEN mother–child cohort on the prenatal and early postnatal determinants of child health and development. *Int. J. Epidemiol.* 45(2), 353–363 (2016).
- 33. Thompson FE, Subar AF. Dietary assessment methodology. In: *Nutrition in the Prevention and Treatment of Disease*. Elsevier Inc., 5–48 (2017).
- Farrow A, Shea KM, Little RE. Birthweight of term infants and maternal occupation in a prospective cohort of pregnant women. the ALSPAC study team. Occup. Environ. Med. 55(1), 18–23 (1998).
- 35. James JE. Maternal caffeine consumption and pregnancy outcomes: a narrative review with implications for advice to mothers and mothers-to-be. *BMJ Evid. Based Med.* 26(3), 114–115 (2021).
- 36. Chen YA, Lemire M, Choufani S *et al.* Discovery of cross-reactive probes and polymorphic CpGs in the Illumina Infinium HumanMethylation450 microarray. *Epigenetics* 8(2), 203–209 (2013).
- 37. Gervin K, Page CM, Aass HCD *et al.* Cell type specific DNA methylation in cord blood: a 450K-reference data set and cell count-based validation of estimated cell type composition. *Epigenetics* 11(9), 690–698 (2016).
- 38. Houseman EA, Accomando WP, Koestler DC *et al.* DNA methylation arrays as surrogate measures of cell mixture distribution. *BMC Bioinformatics* 13(1), (2012).
- 39. Yousefi P, Huen K, Davé V, Barcellos L, Eskenazi B, Holland N. Sex differences in DNA methylation assessed by 450 K BeadChip in newborns. *BMC Genomics* 16 (2015).
- 40. Merid SK, Novoloaca A, Sharp GC et al. Epigenome-wide meta-analysis of blood DNA methylation in newborns and children identifies numerous loci related to gestational age. *Genome Med.* 12(1), 25 (2020).
- 41. York TP, Latendresse SJ, Jackson-Cook C *et al.* Replicated umbilical cord blood DNA methylation loci associated with gestational age at birth. *Epigenetics* 15(11), 1243–1258 (2020).
- 42. Bakker R, Steegers EA, Obradov A, Raat H, Hofman A, Jaddoe VW. Maternal caffeine intake from coffee and tea, fetal growth, and the risks of adverse birth outcomes: the Generation R Study. *Am. J. Clin. Nutr.* 91(6), 1691–1698 (2010).
- 43. Hoyt AT, Browne M, Richardson S, Romitti P, Druschel C. Maternal caffeine consumption and small for gestational age births: results from a population-based case–control study. *Matern. Child Health J.* 18(6), 1540–1551 (2014).
- 44. Elwert F, Winship C. Endogenous selection bias: the problem of conditioning on a collider variable. Annu. Rev. Sociol. 40, 31-53 (2014).
- Sharp GC, Alfano R, Ghantous A et al. Paternal body mass index and offspring DNA methylation: findings from the PACE consortium. Int. J. Epidemiol. 50(4), 1297–1315 (2021).
- 46. Joubert BR, Felix JF, Yousefi P *et al.* DNA methylation in newborns and maternal smoking in pregnancy: genome-wide consortium meta-analysis. *Am. J. Hum. Genet.* 98(4), 680–696 (2016).
- 47. Tukey JW. Biometric Journal. Exploratory Data Analysis. Addison-Wesley Publishing Company Reading, Mass 23(4), 413-414 (1981).
- Leek JT, Johnson WE, Parker HS et al. sva: Surrogate Variable Analysis. R package version 3.38.0. (2020). https://bioconductor.org/packages/release/bioc/html/sva.html
- 49. Ritchie ME, Phipson B, Wu D et al. limma powers differential expression analyses for RNA-sequencing and microarray studies. Nucleic Acids Res. 43(7), e47 (2015).
- Van der Most PJ, Küpers LK, Snieder H, Nolte I. QCEWAS: automated quality control of results of epigenome-wide association studies. *Bioinformatics* 33(8), 1243–1245 (2017).
- 51. Suderman M, Staley JR, French R, Arathimos R, Simpkin A, Tilling K. dmrff: identifying differentially methylated regions efficiently with power and control. *bioRxiv* doi: 10.1101/508556 (2018).
- 52. Odintsova VV, Suderman M, Hagenbeek FA et al. DNA methylation in peripheral tissues and left-handedness. Sci. Rep. 12, 5606 (2022).
- 53. Suderman M, Hemani G, Min JL. *meffil: Efficient algorithms for DNA methylation. R package version* 1.1.1 (2021). https://github.com/perishky/meffil
- 54. Willer CJ, Li Y, Abecasis GR. METAL: fast and efficient meta-analysis of genomewide association scans. *Bioinformatics* 26(17), 2190–2191 (2010).
- 55. Benjamini Y, Hochberg Y. Controlling the false discovery rate: a practical and powerful approach to multiple testing. *Journal of the Royal Statistical Society Series B (Methodological)* 57(1), 289–300 (1995).

- 56. Viechtbauer W. Conducting meta-analyses in R with the metafor package. J. Stat. Softw. 36(3), 1-48 (2010).
- 57. Phipson B, Maksimovic J, Oshlack A. missMethyl: an R package for analysing methylation data from Illumina's HumanMethylation450 platform. *Bioinformatics* btv560 doi: 10.1093/bioinformatics/btv560 (2015).
- 58. Stelzer G, Rosen N, Plaschkes I *et al.* The GeneCards suite: from gene data mining to disease genome sequence analyses. *Curr. Protoc. Bioinformatics* 54(1), (2016).
- The Coffee and Caffeine Genetics Consortium, International Parkinson's Disease Genomics Consortium, North American Brain Expression Consortium *et al.* Genome-wide meta-analysis identifies six novel loci associated with habitual coffee consumption. *Mol. Psychiatry* 20(5), 647–656 (2015).
- Chen L, Bell EM, Browne ML, Druschel CM, Romitti PA. Exploring maternal patterns of dietary caffeine consumption before conception and during pregnancy. *Matern. Child Health J.* 18(10), 2446–2455 (2014).
- 61. Lawson CC, LeMasters GK, Wilson KA. Changes in caffeine consumption as a signal of pregnancy. *Reprod. Toxicol.* 18(5), 625–633 (2004).
- Schreiber GB, Maffeo CE, Robins M, Masters MN, Bond AP. Measurement of coffee and caffeine intake: implications for epidemiologic research. *Prev. Med.* 17(3), 280–294 (1988).
- 63. Sharp GC, Lawlor DA, Richardson SS. It's the mother!: how assumptions about the causal primacy of maternal effects influence research on the developmental origins of health and disease. *Soc. Sci. Med.* 213, 20–27 (2018).
- 64. Murphy C, Brown T, Trickey H *et al.* It remains unclear whether caffeine causes adverse pregnancy outcomes; but naive policy recommendations could cause harm [Letter to the editor] (2020). https://ebm.bmj.com/content/26/3/114.responses#it-remains-unclea r-whether-caffeine-causes-adverse-pregnancy-outcomes-but-naive-policy-recommendations-could-cause-harm
- 65. Verster JC, Koenig J. Caffeine intake and its sources: a review of national representative studies. Crit. Rev. Food Sci. Nutr. 58(8), 1250–1259 (2018).
- 66. van Dam RM, Hu FB, Willett WC. Coffee, caffeine, and health. N. Engl. J. Med. 383(4), 369-378 (2020).
- 67. Lövkvist C, Dodd IB, Sneppen K, Haerter JO. DNA methylation in human epigenomes depends on local topology of CpG sites. *Nucleic Acids Res.* 44(11), 5123–5132 (2016).
- 68. Walton E, Relton CL, Caramaschi D. Using openly accessible resources to strengthen causal inference in epigenetic epidemiology of neurodevelopment and mental health. *Genes* 10(3), 193 (2019).
- 69. Rakyan VK, Down TA, Balding DJ, Beck S. Epigenome-wide association studies for common human diseases. *Nat. Rev. Genet.* 12(8), 529–541 (2011).
- 70. Boylan SM, Cade JE, Kirk SFL et al. Assessing caffeine exposure in pregnant women. Br. J. Nutr. 100(4), 875-882 (2008).
- Cornelis M, Kacprowski T, Menni C et al. Genome-wide association study of caffeine metabolises provides new insights to caffeine metabolism and dietary caffeine-consumption behavior. Hum. Mol. Genet. 5472–5482 doi: 10.1093/hmg/ddw334. (2016).
- 72. Davey-Smith G. Assessing intrauterine influences on offspring health outcomes: can epidemiological studies yield robust findings? *Basic Clin. Pharmacol. Toxicol.* 102(2), 245–256 (2008).
- 73. Easey KE, Sharp GC. The impact of paternal alcohol, tobacco, caffeine use and physical activity on offspring mental health: a systematic review and meta-analysis. *Reprod. Health* 18, 214 (2021).