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Serum plant sterols as surrogate markers of dietary compliance in familial dyslipidemias

Rocío Mateo-Gallego, Lucía Baila-Rueda, Theodora Mouratidou, Ana M. Bea, Sofía Perez-Calahorra, Ana Cenarro, Luis A. Moreno, Fernando Civeira



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1 **Title:** Serum plant sterols as surrogate markers of dietary compliance in familial
2 dyslipidemias

3 **Authors:** Rocío Mateo-Gallego¹, Lucía Baila-Rueda¹, Theodora Mouratidou², Ana M. Bea¹,
4 Sofía Perez-Calahorra¹, Ana Cenarro¹, Luis A. Moreno², Fernando Civeira¹.

5 **Centers:** ¹Unidad de Lípidos and Laboratorio de Investigación Molecular, Hospital
6 Universitario Miguel Servet, Instituto Aragonés de Ciencias de la Salud (I+CS), Zaragoza,
7 Spain. ²GENUD (Growth, Exercise, Nutrition and Development) Research Group, Faculty of
8 Health Sciences, University of Zaragoza, C/ Domingo Miral s/n, 50009 Zaragoza, Spain.

9 **Address for correspondence:** Rocío Mateo-Gallego. Unidad de Lípidos. Hospital
10 Universitario Miguel Servet. C/ Padre Arrupe, s/n, 50009, Zaragoza, Spain.

11 Phone: +34 976765500. Fax: +34 976369985. E-mail: rmateo.iacs@aragon.es

12 **Abbreviations**

13 CVD; Cardiovascular disease

14 ADH; Autosomal dominant hypercholesterolemia

15 FCHL; Familial combined hyperlipidemia

16 TC; Total cholesterol

17 BMI; Body mass index

18 CRP; C-reactive protein

19 FFQ; Food frequency questionnaire

20 PCA; Principal component analysis

21 GGT; γ -glutamyl transpeptidase

22 ABSTRACT

23 **Background and Aims:** A well-balanced diet is the first line treatment in hyperlipidemia.

24 The objective was to study the association between serum phytosterols and dietary patterns to
25 use them as surrogate markers of dietary compliance in primary dyslipidemias.

26 **Methods:** 288 patients with primary hyperlipidemias (192 autosomal dominant
27 hypercholesterolemia (ADH) and 96 familial combined hyperlipidemia (FCHL)) were
28 included. Principal factor analysis identified 2 major dietary patterns using a 137-item food
29 frequency questionnaire. “Vegetable & Fruits pattern” was characterized by higher intake of
30 of fruits, green beans, nuts, tomatoes, roasted or boiled potatoes, lettuce and chard and lower
31 of processed baked goods, pizza and beer. “Western pattern” was positively characterized by
32 hamburgers, pasta, sunflower oil, rice, chickpeas, whole milk, veal, red beans and negatively
33 with white fish. Serum non-cholesterol sterols were determined by HPLC-MS/MS.

34 **Results:** Plant sterols to-total cholesterol (TC) levels were lower with a higher adherence to a
35 “Vegetable & Fruits pattern” ($P = 0.009$), mainly in ADH subjects ($R^2 = 0.019$). Their
36 concentration was greater with higher compliance to “Western pattern” especially in FCHL (P
37 $= 0.014$). Higher levels of synthesis markers to TC with a greater adherence to “Vegetable &
38 Fruits pattern” was found ($P = 0.001$) ($R^2 = 0.033$ and $R^2 = 0.109$ in ADH and FCHL
39 respectively).

40 **Conclusion:** In subjects with primary dislipidemia, dietary patterns associate with serum
41 absorption and synthesis markers, but no with lipid concentrations. The influence of diet on
42 non-cholesterol sterols levels is not powerful enough to use them as subrogate markers.

43

44 KEYWORDS

45 Plant sterols, phytosterols, familial hyperlipidemias, dietary patterns, dietary compliance.

46

47 INTRODUCTION

48 The strong relationship between plasma cholesterol and cardiovascular disease (CVD), the
49 leading cause of mortality in the world, is well accepted [1,2]. Lifestyle behavioural changes,
50 including dietary modifications, increased physical activity and weight loss in overweight or
51 obese patients, are the first-line option treatment in hypercholesterolemias. This is also
52 reflected in the National Cholesterol Education Program (NCEP) dietary guidelines for the
53 prevention of CVD recommending a healthy dietary pattern rich in monounsaturated fatty
54 acids provided by plant sources, fruits, vegetables, whole grains, and low-fat dairy products
55 and low in saturated fatty acids and cholesterol [3].

56 Dietary assessment is important in the following-up of the patients so as to identify
57 “unhealthy” dietary habits to promote adherence to healthier dietary patterns. Dietary
58 questionnaires such as food frequency questionnaires, 24 hour recalls or food records are
59 commonly used to assess dietary intake, however, such methods come with limitations as
60 reliance on memory, misreporting and should be considered when evaluating nutrition
61 behaviours [4,5]. Subrogate parameters which could objectively evaluate dietary compliance
62 would be very useful both in epidemiological studies and in clinical practice.

63 Serum levels of phytosterols, commonly known as plant sterols, and cholestanol are
64 positively correlated with cholesterol absorption and their ratios to cholesterol (relative
65 concentrations) are considered to be reliable markers of intestinal sterols absorption efficiency
66 [6,7]. Serum phytosterols are only partially dependent to their amount in the diet although this
67 association has not been studied in subjects with primary dyslipidemias who show an
68 abnormal cholesterol homeostasis [8,9].

69 Dietary pattern analysis has emerged as an alternative approach to examine the
70 relationship between diet and the risk of chronic diseases; conceptually, dietary patterns
71 provide a broader picture of food and nutrient consumption, and may thus be more predictive

72 of disease risk than individual foods or nutrients [10,11]. The aim of the study was to examine
73 the association between serum phytosterols levels and dietary patterns in primary
74 dyslipidemias by proposing them as surrogate markers of dietary compliance in patients with
75 primary dyslipidemias.

76

77 **MATERIALS AND METHODS**

78 **Study population**

79 Patients attending to the Lipid Unit of the Hospital Universitario Miguel Servet (Zaragoza,
80 Spain) from January 2011 to September 2012 were recruited. 288 subjects with familial
81 hypercholesterolemias were recruited as part of a genetic and metabolic wider study whose
82 study details has been already published elsewhere [12]. Inclusion criteria were being over 18
83 years of age and the presence of familial hyperlipidemia by including autosomal dominant
84 hypercholesterolemia (ADH) and familial combined hyperlipidemia (FCHL). ADH was
85 diagnosed in subjects with off-treatment LDL cholesterol levels above the age- and sex-
86 specific 95th percentile of a Spanish reference population, triglyceride below 200 mg/dL and
87 familial vertical transmission with at least one first-degree relative with LDL cholesterol
88 above age- and sex-specific 95th percentiles. The diagnosis of FCHL was based on the
89 presence of primary combined hyperlipidaemia in untreated patients whose serum cholesterol
90 and triglyceride concentrations were above the sex- and age-specific 90th percentiles for the
91 Spanish population, serum total apolipoprotein B levels ≥ 120 mg/dL and there was at least
92 one first-degree relative with hyperlipidemia (total cholesterol (TC) and/or triglycerides >90th
93 percentile) [13]. Secondary causes of hyperlipidaemia (e.g. body mass index (BMI) ≥ 30
94 kg/m², alcohol intake over 30 gr. and 20 gr. in men and women respectively) and subjects
95 with plant sterols supplements intake were excluded. Written informed consent was obtained

96 by all study participants. The study protocol was approved by the Ethical Committee of our
97 Institution (Comité Ético de Investigación Clínica de Aragón).

98 **Clinical and laboratory determination**

99 Clinical parameters obtained included anthropometric measures (weight, height and waist
100 circumference) and blood pressure. BMI was calculated (weight in kg. divided by the square
101 of height in meters) and all subjects were assessed for personal and/or family history of early-
102 onset coronary heart disease, clinical history, tobacco consumption and demographic
103 characteristics by a personal interview.

104 Fasting blood was drawn following at least 4 weeks without lipid-lowering drugs
105 treatment. Cholesterol, triglycerides, HDL cholesterol and γ -glutamyl transpeptidase (GGT)
106 were measured by spectrophotometry with standard enzymatic methods. LDL cholesterol was
107 estimated with the Friedewald formula when serum triglycerides were <400 mg/dL. Non-
108 HDL cholesterol was calculated as TC minus HDL cholesterol. Apolipoprotein B,
109 lipoprotein(a), and C-reactive protein (CRP) were determined by nephelometry using
110 IMMAGE-Immunochemistry System (Beckman Coulter).

111 **Dietary assessment**

112 Dietary intakes were determined using an interviewer-administered 137-item food frequency
113 questionnaire (FFQ). One registered dietician (RM-G) performed the interviews. More details
114 of the FFQ validity, which has been previously used to study other diet-disease association
115 including plant sterols, could be found elsewhere [14-16]. Food and nutrient intakes were
116 calculated as frequency x nutrient composition of specified portion sizes, where frequencies
117 were measured in 9 categories (never, 1-3 times a month, 1 time a week, 2-4 times a week, 5-
118 6 times a week, 1 time a day, 2-3 times a day, 4-6 times a day and > 6 times a day) for each
119 food item. The total energy and nutrients intakes were calculated based on previously
120 validated Spanish food composition tables [17]. When possible and applicable, the 137 foods

121 were grouped into categories based on similar nutritional values. Food items (N = 18) with
122 low prevalence of consumption (less than 15%) were not considered in the final analysis (N =
123 119) to avoid possible bias in the dietary patterns calculation.

124 **Serum non-cholesterol sterols determination**

125 Serum non-cholesterol sterol concentrations were analysed by high performance liquid
126 chromatography tandem mass spectrometry (HPLC-MS/MS) [18]. Briefly, ($[^2\text{H}_6]$ cholesterol-
127 26,26,26,27,27,27 D6) (4 $\mu\text{g/g}$) was added to serum (0.1 ml) as the internal standard. After
128 alkaline hydrolysis, extraction and solid phase extraction, the sterols were separated using
129 reverse-phase C18 HPLC. A 40 μl aliquot of the extract (100% 2-propanol) was loaded onto a
130 RP-HPLC column (Zorbax Eclipse Plus C₁₈ 2.1 x 150 mm, 3,5 μm particle; Agilent, Spain)
131 equipped with a guard column (C₁₈, 4 x 2,5 mm). The HPLC (Agilent 1200RRLC) was
132 coupled to a 4000 QTrap triple quadrupole ion trap mass spectrometer (Applied Biosystems,
133 Foster City, CA) through an APCI by Heated Nebulizer (Turbo VTM Source). In each run,
134 cholestanol, campesterol, sitosterol, sitostanol and stigmasterol were quantified.

135 **Statistical analysis**

136 Statistical analysis was performed using SPSS software version 15.0 (Chicago, Illinois, USA)
137 using a significance level of $P < 0.05$.

138 Data are expressed as means \pm standard deviation (SD) for continuous variables with
139 normal distribution and medians (percentile 25 – percentile 75) for variables with a skewed
140 distribution. Student-t or Mann-Whitney tests were used accordingly. Categorical variables
141 were compared using a chi-square test. ANOVA and Kruskal-Wallis tests were performed to
142 multiple independent variables comparison. Non-cholesterol sterols levels were adjusted by
143 those variables which have been shown more influential in its concentration: age, gender,
144 BMI and APOE genotype [19]. PCA with varimax rotation was used to derive dietary patterns
145 based on the 61 foods or food groups [10,20,21]. The factors were rotated by an orthogonal

146 transformation (resulting in uncorrelated factors) to achieve a simpler structure with greater
147 interpretability. In determining the number of factors to retain, we considered components
148 with an eigenvalue >1 , the Scree test and the interpretability of the factors. The factor score
149 for each pattern was constructed by summing observed intakes of the component food items
150 weighted by factor loadings so each subject had a score for each dietary pattern with a higher
151 score indicating higher adherence to the respective pattern [22]. Factor loadings quintiles
152 were calculated for each different dietary pattern to study the association with non-cholesterol
153 sterols levels.

154

155 RESULTS

156 The study group was composed of 288 subjects (48% men), of whom 192 were diagnosed
157 with ADH and 96 subjects with FCHL. **Table 1** presents the main clinical and biochemical
158 characteristics by dyslipidemia type. The FCHL group had a higher percentage of men (65.6%
159 vs. 39.8%, $P = < 0.001$), smokers (35.4% vs. 22.5% in FCHL and ADH respectively, $P =$
160 0.048), had higher BMI (26.3 ± 2.12 vs. 24.3 ± 2.79 in FCHL and ADH respectively, $P < 0.001$)
161 and waist circumference (93.2 ± 8.14 vs. 84.0 ± 10.5 cm. in FCHL and ADH respectively, $P <$
162 0.001) than AHD group. Regarding biochemical parameters, patients diagnosed with ADH, as
163 expected, showed higher levels of LDL cholesterol (219 (198-250) vs. 204 (181-228) mg/dL
164 in ADH and FCHL respectively, $P < 0.001$), HDL cholesterol (56.0 (45.3-68.0) vs. 41.5
165 (36.3-52.8) mg/dL in ADH and FCHL respectively, $P < 0.001$) and apolipoprotein A1
166 (158 ± 35.1 vs. 143 ± 28.1 mg/dL in ADH and FCHL respectively, $P < 0.001$) and lower values
167 of triglycerides (95.5 (77.0-131) vs. 246 (194-382) mg/dL in ADH and FCHL respectively, P
168 < 0.001). Adjusted non-cholesterol sterols concentrations and their ratios to TC by type of
169 dyslipidemia are shown in **Table Supplementary 1**. In general, the ADH group had higher
170 levels of non-cholesterol sterols which have been established as absorption markers such as

171 cholestanol, desmosterol and campesterol especially with their ratios to TC. FCHL group
172 showed higher concentrations of cholesterol synthesis markers particularly of lanosterol
173 concentration.

174 Factor analysis (principal components) to derive dietary patterns based on the 61 foods
175 or food groups were performed and 1 major and 3 minor patterns (accounting for 18.8% of the
176 variance) were identified. Thus, we extracted 2 factors in the final model whose factor-
177 loading matrixes are presented in **Table 2**. The larger the loading of a given food item to the
178 factor, the greater the contribution of that food item to a specific factor. The first factor was
179 positively correlated with consumption of oranges, bananas, apples, green beans, nuts,
180 tomatoes, roasted or boiled potatoes, lettuce and chard whilst negatively with processed baked
181 goods, pizza and beer. The second factor was loaded positively with hamburgers, pasta,
182 sunflower oil, rice, chickpeas, whole milk, veal, red beans and negatively with white fish.
183 Components were named as “Vegetable & Fruits dietary pattern” and the second as “Western
184 dietary pattern”, which explained 6.85% and 4.12% of the total diet variance respectively. The
185 two other minor patterns that were identified did not appear to represent a clear dietary pattern
186 and analyses did not suggest a significant association between these patterns and non-
187 cholesterol sterols (data not shown).

188 Subjects characteristics across quintiles of dietary patterns (“Vegetable & Fruits
189 dietary pattern” and “Western dietary pattern”) scores are described in **Table Supplementary**
190 **2** and **Table Supplementary 3** respectively. Those subjects with a higher score for the
191 “Vegetable & Fruits dietary pattern” were more likely to be older, have a higher intake of
192 energy, total fat, phytosterols, fiber and lower carbohydrates consumption. The percentage of
193 men was higher in the lowest quintile although a tendency across quintiles was not observed.
194 Regarding “Western dietary pattern”, those subjects with a higher adherence were younger
195 and presenting an upper intake of energy, carbohydrates, cholesterol and lower of

196 monounsaturated fat consumption. Significant differences were found across quintiles in
197 protein and phytosterols intake although a clear tendency was not found. A higher percentage
198 of smokers was found in the highest quintile of “Western dietary pattern” compliance.

199 The association of non-cholesterol sterols with dietary patterns is described in **Table 3**
200 and **4** (“Vegetable & Fruits dietary pattern” and “Western dietary pattern” respectively).
201 Those subjects with a higher adherence to the “Vegetable & Fruits dietary pattern” showed
202 lower levels of adjusted absorption (phytosterols-to TC) markers compared to those with
203 lower adherence ($2.59 (2.44-2.88) \times 10^{-2}$ mg/dL in Q1 versus $2.48 (2.30-2.61) \times 10^{-2}$ mg/dL in
204 Q5, $P = 0.009$ in all subjects). This was mainly noted in stigmasterol-to-TC ($0.42 (0.36-0.43)$
205 $\times 10^{-2}$ mg/dL in Q1 versus $0.40 (0.34-0.42) \times 10^{-2}$ mg/dL in Q5, $P = 0.002$ in all subjects) and
206 sitosterol-to-TC ($1.65 (1.57-1.87) \times 10^{-2}$ mg/dL in Q1 versus $1.60 (1.51-1.68) \times 10^{-2}$ mg/dL in
207 Q5, $P = 0.044$ in all subjects). This tendency was mainly observed in subjects diagnosed with
208 ADH. Cholestanol-to-TC levels did not show a clear tendency across quintiles, which was not
209 statistically significant in any of both dyslipidemias. The concentration of adjusted synthesis-
210 to-TC (lanosterol and desmosterol) increased across “Vegetable & Fruits dietary pattern”
211 score quintiles which was observed both in ADH and in FCHL ($0.70 (0.67-0.75) \times 10^{-2}$ mg/dL
212 in Q1 versus $0.75 (0.71-0.80) \times 10^{-2}$ mg/dL in Q5, $P = 0.001$ in all subjects). Regarding
213 “Western dietary pattern”, those subjects with a higher adherence were likely to have higher
214 levels of adjusted absorption (phytosterols-to-TC) markers compared to those in the lowest
215 ($2.39 (2.27-2.55) \times 10^{-2}$ mg/dL in Q1 versus $2.54 (2.40-2.71) \times 10^{-2}$ mg/dL in Q5, $P = 0.012$ in
216 all subjects) although the trend was not fully clear. As in the first dietary pattern, this effect
217 was mainly present in campesterol-to-TC ($0.39 (0.36-0.41) \times 10^{-2}$ mg/dL in Q1 versus 0.40
218 ($0.38-0.46) \times 10^{-2}$ mg/dL in Q5, $P = 0.042$ in all subjects), stigmasterol-to-TC ($0.36 (0.35-$
219 $0.41) \times 10^{-2}$ mg/dL in Q1 versus $0.41 (0.35-0.42) \times 10^{-2}$ mg/dL in Q5, $P = 0.022$ in all
220 subjects) and sitosterol-to-TC levels ($1.56 (1.49-1.66) \times 10^{-2}$ mg/dL in Q1 versus $1.62 (1.51-$

221 $1.81) \times 10^{-2}$ mg/dL in Q5, $P = 0.037$ in all subjects). FCHL group had more significant
222 differences among quintiles, mainly comparing the highest quintile vs. the lowest one, but the
223 trend was unclear. The adjusted levels of the synthesis markers levels were likely to be lower
224 in those subjects with higher scores for “Western dietary pattern” but no significant
225 differences were found.

226 Regression analysis showed that “Vegetable & Fruits dietary pattern” adherence was
227 significantly associated to absorption markers, independently of age, BMI, gender and *APOE*
228 genotype, only in ADH subjects (standardized $B = -0.156$; $P = 0.030$) by determining a 1.9% of
229 the variance. The compliance to this pattern was independently associated to synthesis
230 subrogate markers in ADH and FCHL (standardized $B = 0.195$; $P = 0.007$ and standardized $B =$
231 0.345 ; $P = 0.001$ respectively) by determining a 3.3% and 10.9% of levels variance in each
232 case. No significant influence of “Western dietary pattern” compliance in non-cholesterol
233 sterols was founded.

234

235 DISCUSSION

236 The results derived from the present study reveal that there is an influence of diet on non-
237 cholesterol sterols levels although the association is not strong enough to consider them as
238 potential subrogate markers of healthy diet compliance. These data agree with previous
239 studies which have stated the diet influence on serum plant sterols levels concentration, not
240 only with phytosterols intake but with other nutrients too, such as the ratio of
241 polyunsaturated/saturated fatty acids [6]. However, the influence of regular dietary
242 phytosterols intake on circulating plant sterols and in plant sterol-to-TC ratios has been
243 estimated in 4.03% and 3.59% respectively in general population but no data were available
244 for subjects with familial dyslipidemias [19]. Otherwise, the high intake of phytosterols, as
245 supplements, is associated with a marked increase in serum plant sterols concentration

246 [19,23]. According to our data, healthy diet (“Vegetable & Fruits dietary pattern” with a
247 moderate-high intake of plant sterols) compliance independently influence non-cholesterol
248 sterols, mainly in ADH. These levels variance explained by the dietary adherence is around
249 1.9% in absorption subrogate markers and 3.3% and 10.9% (ADH and FCHL respectively) in
250 synthesis subrogate markers. Despite having analysed dietary patterns beyond of isolated
251 nutrients, the influence is not strong enough and their usefulness as subrogate markers of
252 dietary adherence does not seem to be useful.

253 A clear association of lipid parameters and a healthy dietary adherence was not found
254 in any of both dyslipidemias. Thus, in non obese subjects with primary dyslipidemias, the
255 effect of diet on lipid metabolism seems to be low, in contrast with the substantial
256 improvement of lipid profile with weigh loss and dietary habits enhancement in obese
257 subjects with FCHL that has been previously proved [3,24]. Given that non-cholesterol
258 sterols, both absorption and synthesis markers, are not markedly influenced by diet in non
259 obese subjects with primary dyslipidemias, they could be more useful for the study of the
260 pathogenic mechanism of the lipoprotein disorder, as previously proposed [25].

261 The inverse association of cholesterol absorption with synthesis has been well
262 established in general population and our data confirm this relationship also in subjects with
263 primary dyslipidemias. An increase in cholesterol synthesis by the liver occurs when
264 intestinal cholesterol absorption diminishes due to an increase in phytosterols intake [7,26].
265 An unexpected finding in our study was that those subjects with a high adherence to a healthy
266 diet, rich in vegetables, fruits and phytosterols, had lower plasma plant sterols levels than
267 those subjects with a lower adherence. At the same time, an increase in cholesterol synthesis
268 markers was also observed with a higher adherence which was especially evident in ADH
269 subjects. We hypothesize that a compensatory mechanism could be the responsible for the
270 reduction of serum plant sterols levels when an increase of a healthy diet (“Vegetable & Fruits

271 dietary pattern”) adherence. It maybe possible that in those patients following a healthy diet
272 the excretion of phytosterols to the bile would be increased by compensating the
273 augmentation of those in the serum whereas the cholesterol neosynthesis in the liver would
274 raise, as previously described by Krawczyk M. et al. in gallstone disease [27]. Because of this
275 compensatory mechanism, no differences in the cholesterol levels across quintiles of dietary
276 adherence are observed despite of a better diet and higher intake of phytosterols. These data
277 reinforces the primary cause of the hypercholesterolemia in the studied subjects and explains
278 the poor response to diet modification in some genetic hypercholesterolemias as previously
279 reported [28,29]. Most of our ADH subjects had familial hypercholesterolemia with a
280 pathogenic mutation in the LDLR, a group of subjects with a very limited response to a lipid
281 lowering diet due to the mechanism of their disease. In contrast, the lipid profile of subjects
282 with FCHL is highly dependent of environmental factors, especially to weight gain. However,
283 cholesterol synthesis, the main pathogenic factor in FCHL, is poorly modified by diet in
284 absence of weight loss, in contrast with triglycerides synthesis [30].

285 The study has several limitations such as the assessment of food intake based on
286 subjective self-reports although we have not studied nutrients or foods by separate but dietary
287 patterns which represent better and more realistic the overall diet and represent a broader
288 picture of dietary habits which constitute strength of the present study. Subjective decisions
289 on the number of patterns to be extracted were made based on empirical guidelines rather than
290 on an exact quantitative solution and this should be considered as another study limitation.

291 In conclusion, the influence of diet on non-cholesterol sterols concentration is neither
292 completely clear nor sufficiently powerful and a clear tendency was not observed in patients
293 with primary dyslipidemias. Thus, the usefulness of serum plant sterols levels as subrogate
294 makers of dietary adherence have to be reconsidered.

295

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299 questionnaires.

300

301 STATEMENT OF AUTHORSHIP

302 The author's responsibilities were as follows – RMG: conducted research, collected and
303 analyzed data and performed statistical analysis, wrote the paper and had primary
304 responsibility for its final content; LBR: conducted research, collected and analyzed data;
305 TM: performed statistical analysis and provided significant advice; AMB, SPC and AC:
306 conducted research and provided significant consultation; LAM: provided significant advice;
307 FC: conducted research, analyzed data, performed statistical analysis, wrote the paper,
308 provided significant advice and consultation and had primary responsibility for its final
309 content.

310

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314

315 CONFLICT OF INTEREST STATEMENT

316 None.

317

318 REFERENCES

319 [1] Stamler J, Wentworth D, Neaton JD; for the MRFIT Research Group. Is relationship
320 between serum cholesterol and risk of premature death from coronary heart disease

- 321 continuous and graded? Findings in 356 222 primary screenees of the Multiple Risk
322 Factor Intervention Trial (MRFIT). *JAMA* 1986; 256: 2823-8.
- 323 [2] Global strategy on diet, physical activity and health. Cardiovascular disease: prevention
324 and control. Geneva, Switzerland: World Health Organization, 2007.
- 325 [3] National Cholesterol Education Program (NCEP) Expert Panel on Detection, Evaluation
326 and Treatment of High Blood Cholesterol in Adults (Adult Treatment Panel III) Third
327 Report of the National Cholesterol Education Program (NCEP) Expert Panel on
328 Detection, Evaluation, and Treatment of High Blood Cholesterol in Adults (Adult
329 Treatment Panel III) final report. *Circulation* 2002; 106:3143-421.
- 330 [4] Serra-Majem L, Frost Andersen L, Henríque-Sánchez P, Doreste-Alonso J, Sánchez-
331 Villegas A, Ortiz-Andrelluchi A et al. Evaluating the quality of dietary intake validation
332 studies. *Br J Nutr* 2009; 102 Suppl 1:S3-9.
- 333 [5] Ngo J, Engelen A, Molag M, Roesle J, García-Segovia, P, Serra-Majem L. A review of
334 the use of information and communication technologies for dietary assessment. *Br J Nutr*
335 2009; 101 Suppl. 2:S102-12.
- 336 [6] Miettinen TA, Tilvis RS, Kesäniemi YA. Serum cholestanol and plant sterol levels in
337 relation to cholesterol metabolism in middle-aged men. *Metabolism* 1989; 38:136-40.
- 338 [7] Miettinen TA, Gylling H, Nissinen MJ. The role of serum non-cholesterol sterols as
339 surrogate markers of absolute cholesterol synthesis and absorption. *Nutr Metab*
340 *Cardiovasc Dis* 2011; 21:765-9.
- 341 [8] Escurriol V, Cofán M, Serra M, Bulló M, Basora J, Salas-Salvadó J et al. Serum sterol
342 responses to increasing plant sterol intake from natural foods in the Mediterranean diet.
343 *Eur J Nutr* 2009; 48:373-82.

- 344 [9] Sanclemente T, Marques-Lopes I, Fajó-Pascual M, Cofán M, Jarauta E, Ros E et al.
345 Naturally-occurring phytosterols in the usual diet influence cholesterol metabolism in
346 healthy subjects. *Nutr Metab Cardiovasc Dis* 2012; 22:849-55.
- 347 [10] Hu FB. Dietary pattern analysis: a new direction in nutritional epidemiology. *Curr*
348 *Opin Lipidol* 2002;13:3-9.
- 349 [11] Duffey KJ, Steffen LM, Van Horn L, Jacobs DR Jr, Popkin BM. Dietary patterns
350 matter: diet beverages and cardiometabolic risks in the longitudinal Coronary Artery Risk
351 Development in Young Adults (CARDIA) Study. *Am J Clin Nutr* 2012; 95:909-15.
- 352 [12] Jarauta E, Mateo-Gallego R, Gilabert R, Plana N, Junyent M, de Groot E et al. Carotid
353 atherosclerosis and lipoprotein particle subclasses in familial hypercholesterolaemia and
354 familial combined hyperlipidaemia. *Nutr Metab Cardiovasc Dis* 2012; 22:591-7.
- 355 [13] Gómez-Gerique JA, Gutiérrez-Fuentes JA, Montoya MT, Porres A, Rueda A,
356 Avellaneda A et al. Lipid profile of the Spanish population: the DRECE (diet and risk of
357 cardiovascular disease in Spain) study. DRECE study group. *Med Clin (Barc)* 1999;
358 113:730–735.
- 359 [14] de la Fuente-Arrillaga C, Vázquez Ruiz Z, Bes-Rastrollo M, Sampson L, Martínez-
360 González MA. Reproducibility of an FFQ validated in Spain. *Public Health Nutr* 2010;
361 13:1364-72.
- 362 [15] Estruch R, Ros E, Salas-Salvadó J, Covas MI, Corella D, Arós F et al.; PREDIMED
363 Study Investigators. Primary prevention of cardiovascular disease with a Mediterranean
364 diet. *N Engl J Med* 2013; 368:1279-90.
- 365 [16] Sanclemente T, Marques-Lopes I, Fajó-Pascual M, Cofán M, Jarauta E, Ros E et al. A
366 moderate intake of phytosterols from habitual diet affects cholesterol metabolism. *J*
367 *Physiol Biochem.* 2009; 65:397-404.

- 368 [17] Mataix Verdú J, Mañas Almendros M. *Tabla de composición de alimentos españoles*
369 *(Spanish Food Composition Tables), 4th edn.*, Universidad de Granada: Granada, Spain,
370 2003.
- 371 [18] Baila-Rueda L, Cenarro A, Cofán M, Orera I, Barcelo-Batllori S, Pocoví M et al.
372 Simultaneous determination of oxysterols, phytosterols and cholesterol precursors by high
373 performance liquid chromatography tandem mass spectrometry in human serum. *Anal.*
374 *Methods* 2013; 5:2249-2257.
- 375 [19] Chan YM, Varady KA, Lin Y, Trautwein E, Mensink RP, Plat J, Jones PJ. Plasma
376 concentrations of plant sterols: physiology and relationship with coronary heart disease.
377 *Nutr Rev* 2006;64:385-402.
- 378 [20] Hu FB, Rimm E, Smith-Warner SA, Feskanich D, Stampfer MJ, Ascherio A et al.
379 Reproducibility and validity of dietary patterns assessed with a food-frequency
380 questionnaire. *Am J Clin Nutr* 1999; 69:243-9.
- 381 [21] Martínez ME, Marshall JR, Sechrest L. Invited commentary: Factor analysis and the
382 search for objectivity. *Am J Epidemiol* 1998; 1148:17-9.
- 383 [22] Kim J-O, Mueller CW. *Factor analysis: statistical methods and practical issues.*
384 Thousand Oaks, CA: Sage Publications, Inc, 1978.
- 385 [23] Racette SB, Lin X, Lefevre M, Spearie CA, Most MM, Ma L et al. Dose effects of
386 dietary phytosterols on cholesterol metabolism: a controlled feeding study. *Am J Clin*
387 *Nutr* 2010; 91:32-8.
- 388 [24] Carmena R, Ascaso JF, Real JT. Impact of obesity in primary hyperlipidemias. *Nutr*
389 *Metab Cardiovasc Dis* 2001; 11:354-9.
- 390 [25] García-Otín AL, Cofán M, Junyent M, Recalde D, Cenarro A, Pocoví M, Ros E,

- 391 Civeira F. Increased intestinal cholesterol absorption in autosomal dominant
392 hypercholesterolemia and no mutations in the low-density lipoprotein receptor or
393 apolipoprotein B genes. *J Clin Endocrinol Metab* 2007; 92:3667-73.
- 394 [26] Miettinen TA, Gylling H. Cholesterol absorption efficiency and sterol metabolism in
395 obesity. *Atherosclerosis* 2000; 153:241-8.
- 396 [27] Krawczyk M, Lütjohann D, Schirin-Sokhan R, Villarroel L, Nervi F, Pimentel F et al.
397 Phytosterol and cholesterol precursor levels indicate increased cholesterol excretion and
398 biosynthesis in gallstone disease. *Hepatology* 2012; 55:1507-17.
- 399 [28] Broekhuizen K, van Poppel MN, Koppes LL, Kindt I, Brug J, van Mechelen W. Can
400 multiple lifestyle behaviours be improved in people with familial hypercholesterolemia?
401 Results of a parallel randomised controlled trial. *PLoS One* 2012; 7:e50032.
- 402 [29] Carmena-Ramón R, Real JT, Ascaso JF, Ordovás JM, Carmena R. Effect of
403 apolipoprotein E genotype on lipid levels and response to diet in familial
404 hypercholesterolemia. *Nutr Metab Cardiovasc Dis* 2000; 10:7-13.
- 405 [30] Mateo-Gallego R, Jarauta E, Calmarza P, Bea AM, Burillo E, Civeira F. Predictors of
406 lipid lowering response to weight loss in familial hyperlipidemias. *Circulation* 2009;
407 120:S529-S530.

Table 1. Factor loading matrix for the two major dietary patterns identified.^a

	“Vegetable & fruit dietary pattern”	“Western dietary pattern”
Oranges	0.755	
Bananas	0.683	
Apples	0.651	
Green beans	0.428	
Nuts	0.426	
Tomatoes	0.416	
Roasted or boiled potatoes	0.367	
Lettuce	0.330	
Chard	0.305	
Processed baked goods	- 0.301	
Pizza	- 0.395	
Hamburguers		0.526
Pasta		0.518
Sunflower oil		0.515
Rice		0.488
Chickpeas		0.460
Whole milk		0.366
Veal		0.319
White fish		- 0.334
Beer	- 0.327	
Red beans		0.329

^aAbsolute values < 0.30 were not listed in the table for simplicity. Food items with factor loadings < 0.30 for any factor were excluded.

Table 2. Clinical and biochemical characteristics of subjects according to clinical diagnosis. ^a

	ADH N= 192	FCH N = 96	<i>p</i>
Males, n (%)	76 (39.8)	63 (65.6)	<0.001
Age, years	44.2±12.3	46.0±11.3	0.208
Tobacco consumption, n (%)			
Smoker	43 (22.5)	34 (35.4)	0.048
Former smoker	53 (27.7)	26 (27.1)	
Non smoker	95 (49.7)	36 (37.5)	
Diabetes, n (%)	0 (0)	1 (1)	0.333
Hypertension, n (%)	27 (14.1)	12 (12.5)	0.433
Systolic blood pressure, mm Hg	127 (119-135)	131 (123-141)	<0.001
Diastolic blood pressure, mm Hg	80.0 (70.0-85.0)	82.5 (77.3-90.0)	<0.001
Body mass index, kg/m ²	24.3±2.79	26.3±2.12	<0.001
Waist circumference, cm	84.0±10.5	93.2±8.14	<0.001
Metabolic syndrome, n (%)	22 (11.5)	50 (52.1)	< 0.001
Total cholesterol, mg/dL	309±48.3	299±44.1	0.106
HDL cholesterol, mg/dL	56.0 (45.3-68.0)	41.5 (36.3-52.8)	<0.001
Non HDL cholesterol, mg/dL	239 (218-271)	246 (222-276)	0.252
LDL cholesterol, mg/dL	219 (198-250)	204 (181-228)	<0.001
Triglycerides, mg/dL	95.5 (77.0-131)	246 (194-382)	<0.001
Apolipoprotein A1, mg/dL	158±35.1	143±28.1	<0.001
Apolipoprotein B, mg/dL	161±33.7	168±31.6	0.104
Lipoprotein(a), mg/dL	27.9 (13.6-59.2)	24.3 (7.60-59.0)	0.193
C reactive Protein, mg/L	1.50 (0.60-3.40)	2.35 (1.10-3.63)	0.009
GGT, IU/L	19.5 (15.0-29.0)	30.0 (19.0-46.0)	<0.001
Glucose, mg/dL	87.8±10.2	92.9±11.1	<0.001
HbA1c, %	5.20 (5.00-5.40)	5.30 (5.10-5.50)	0.033

^a Values are mean ± standard deviation or median (percentile 25-percentile 75) as applicable.

Table 3. Adjusted non-cholesterol sterols-to-total cholesterol levels among quintiles of “Vegetable & Fruits dietary pattern” scores.^a

		Cholestanol-to-TC	Stigmasterol-to-TC	Campesterol-to-TC	Sitosterol-to-TC	Synthesis markers to-TC ^b	Phytosterols-to-TC ^c
ADH N = 192	Q1	0.60 (0.56-0.63)	0.42 (0.40-0.43)	0.41 (0.38-0.46)	1.68 (1.61-1.90)	0.71 (0.66-0.74)	2.62 (2.50-2.90)
	Q2	0.61 (0.58-0.64)	0.36 (0.35-0.41)	0.40 (0.38-0.45)	1.58 (1.48-1.68)	0.70 (0.65-0.76)	2.45 (2.29-2.63)
	Q3	0.62 (0.58-0.65)	0.36 (0.35-0.42)	0.40 (0.38-0.45)	1.57 (1.50-1.74)	0.70 (0.66-0.75)	2.44 (2.29-2.70)
	Q4	0.62 (0.58-0.67)	0.40 (0.35-0.42)	0.40 (0.36-0.45)	1.60 (1.53-1.79)	0.72 (0.66-0.78)	2.48 (2.34-2.73)
	Q5	0.60 (0.87-0.65)	0.40 (0.34-0.42)	0.39 (0.36-0.41)	1.60 (1.54-1.68)	0.75 (0.70-0.79)	2.48 (2.31-2.61)
	<i>p</i>	0.561	0.005	0.087	0.009	0.031	0.009
FCHL N = 96	Q1	0.57 (0.53-0.60)	0.42 (0.35-0.52)	0.38 (0.35-0.41)	1.60 (1.47-1.87)	0.70 (0.68-0.75)	2.52 (2.26-2.87)
	Q2	0.58 (0.56-0.61)	0.41 (0.35-0.42)	0.36 (0.35-0.40)	1.60 (1.49-1.65)	0.74 (0.69-0.77)	2.51 (2.30-2.54)
	Q3	0.59 (0.57-0.61)	0.36 (0.35-0.41)	0.39 (0.35-0.40)	1.53 (1.41-1.64)	0.73 (0.69-0.77)	2.29 (2.19-2.56)
	Q4	0.58 (0.55-0.63)	0.41 (0.38-0.47)	0.37 (0.34-0.41)	1.62 (1.56-1.82)	0.77 (0.72-0.78)	2.56 (2.38-2.81)
	Q5	0.58 (0.55-0.59)	0.40 (0.35-0.43)	0.36 (0.34-0.38)	1.61 (1.44-1.69)	0.77 (0.72-0.79)	2.45 (2.23-2.66)
	<i>p</i>	0.303	0.220	0.301	0.441	0.018	0.252
All subjects N = 288	Q1	0.58 (0.54-0.61)	0.42 (0.36-0.43)	0.40 (0.37-0.45)	1.65 (1.57-1.87)	0.70 (0.67-0.75)	2.59 (2.44-2.88)
	Q2	0.61 (0.58-0.63)	0.40 (0.35-0.41)	0.39 (0.36-0.43)	1.59 (1.49-1.66)	0.71 (0.66-0.77)	2.47 (2.30-2.57)
	Q3	0.61 (0.58-0.64)	0.36 (0.35-0.42)	0.40 (0.38-0.44)	1.57 (1.48-1.65)	0.71 (0.67-0.75)	2.44 (2.27-2.58)
	Q4	0.62 (0.57-0.65)	0.40 (0.35-0.42)	0.40 (0.36-0.44)	1.61 (1.53-1.80)	0.73 (0.67-0.78)	2.49 (2.35-2.77)
	Q5	0.59 (0.57-0.62)	0.40 (0.34-0.42)	0.38 (0.35-0.41)	1.60 (1.51-1.68)	0.75 (0.71-0.80)	2.48 (2.30-2.61)
	<i>p</i>	0.044	0.003	0.024	0.012	0.001	0.009

^aData (10⁻²) are expressed as median (percentile 25-percentil 75). Non-cholesterol sterols-to-total cholesterol levels are adjusted by gender, age, BMI and APOE genotype. *P* refers to statistical differences among quintiles.

^bSynthesis markers are the sum of lanosterol and desmosterol levels.

^cPhytosterols are the sum of campesterol, sitosterol, sitostanol and stigmasterol levels.

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Table 4. Non-cholesterol sterols-to-total cholesterol levels among quintiles of “Western dietary pattern” scores.^a

		Cholestanol-to-TC	Stigmasterol-to-TC	Campesterol-to-TC	Sitosterol-to-TC	Synthesis markers-to-TC ^b	Phytosterols-to-TC ^c
ADH N = 192	Q1	0.62 (0.60-0.66)	0.36 (0.35-0.41)	0.40 (0.38-0.43)	1.57 (1.50-1.74)	0.71 (0.67-0.79)	2.37 (2.28-2.67)
	Q2	0.60 (0.57-0.63)	0.41 (0.34-0.42)	0.39 (0.35-0.44)	1.60 (1.50-1.76)	0.73 (0.69-0.76)	2.49 (2.28-2.75)
	Q3	0.62 (0.58-0.66)	0.40 (0.35-0.42)	0.40 (0.36-0.45)	1.61 (1.54-1.77)	0.73 (0.65-0.76)	2.49 (2.38-2.73)
	Q4	0.61 (0.58-0.64)	0.40 (0.34-0.42)	0.41 (0.37-0.45)	1.63 (1.53-1.76)	0.69 (0.67-0.76)	2.50 (2.32-2.71)
	Q5	0.61 (0.57-0.65)	0.41 (0.35-0.42)	0.40 (0.38-0.46)	1.62 (1.52-1.81)	0.71 (0.68-0.73)	2.53 (2.41-2.78)
	<i>p</i>	0.163	0.317	0.521	0.647	0.554	0.350
FCH N = 96	Q1	0.58 (0.57-0.60)	0.40 (0.34-0.41)	0.36 (0.35-0.40)	1.56 (1.46-1.61)	0.73 (0.70-0.78)	2.45 (2.25-2.51)
	Q2	0.58 (0.56-0.61)	0.40 (0.34-0.41)	0.36 (0.35-0.40)	1.58 (1.47-1.66)	0.76 (0.70-0.79)	2.44 (2.25-2.53)
	Q3	0.51 (0.50-0.60)	0.51 (0.42-0.52)	0.36 (0.34-0.38)	1.85 (1.57-1.90)	0.76 (0.72-0.81)	2.89 (2.56-2.95)
	Q4	0.58 (0.55-0.59)	0.41 (0.35-0.42)	0.38 (0.34-0.39)	1.57 (1.46-1.66)	0.73 (0.70-0.78)	2.44 (2.24-2.62)
	Q5	0.58 (0.55-0.61)	0.41 (0.35-0.43)	0.41 (0.37-0.48)	1.63 (1.49-1.85)	0.71 (0.67-0.76)	2.54 (2.29-2.61)
	<i>p</i>	0.068	0.016	0.019	0.077	0.284	0.014
All subjects N = 288	Q1	0.61 (0.58-0.64)	0.36 (0.35-0.41)	0.39 (0.36-0.41)	1.56 (1.49-1.66)	0.73 (0.69-0.78)	2.39 (2.27-2.55)
	Q2	0.59 (0.57-0.62)	0.40 (0.34-0.42)	0.38 (0.35-0.41)	1.58 (1.49-1.69)	0.74 (0.70-0.77)	2.47 (2.27-2.64)
	Q3	0.60 (0.57-0.66)	0.41 (0.35-0.43)	0.39 (0.36-0.45)	1.61 (1.55-1.83)	0.74 (0.66-0.78)	2.51 (2.38-2.82)
	Q4	0.60 (0.57-0.63)	0.41 (0.35-0.42)	0.39 (0.36-0.42)	1.60 (1.51-1.70)	0.71 (0.67-0.77)	2.48 (2.32-2.64)
	Q5	0.60 (0.56-0.63)	0.41 (0.35-0.42)	0.40 (0.38-0.46)	1.62 (1.51-1.81)	0.71 (0.68-0.75)	2.54 (2.40-2.71)
	<i>p</i>	0.400	0.110	0.061	0.098	0.242	0.042

^aData (10⁻²) are expressed as median (percentile 25-percentile 75). Non-cholesterol sterols-to-total cholesterol levels are adjusted by gender, age, BMI and APOE genotype. *P* refers to statistical differences among quintiles.

^bSynthesis markers are the sum of lanosterol and desmosterol levels.

^cPhytosterols are the sum of campesterol, sitosterol, sitostanol and stigmasterol levels.

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Table Supplementary 1. Clinical, biochemical and dietary characteristics of subjects among quintiles of “Vegetable & Fruits dietary pattern” scores.^a

	Q1 N = 56	Q2 N = 57	Q3 N = 57	Q4 N = 57	Q5 N = 57	<i>p</i>
ADH/ FCH, %	17.4 / 24.5	20.0 / 20.2	21.6 / 17.0	22.1 / 16.0	18.9 / 22.3	0.433
Males, %	38 (67.9)	27 (47.4)	18 (31.6)	23 (40.4)	32 (56.1)	0.001
Age, years	37.7±9.48	44.8±9.85	45.1±7.83	48.0±9.82	48.4±8.87	< 0.001
Tobacco consumption, %						
Smoker	22 (39.3)	19 (33.9)	13 (22.8)	16 (28.1)	7 (12.3)	0.125
Former smoker	13 (23.2)	15 (26.8)	16 (28.1)	15 (26.3)	18 (31.6)	
Non smoker	21 (37.5)	22 (39.3)	28 (49.1)	26 (45.6)	32 (56.1)	
Packets/day x Years	16.5 (7.75-30.0)	23.0 (10.5-33.0)	20.0 (10.8-30.0)	13.0 (8.00-23.0)	20.0 (10.5-31.0)	0.489
Systolic blood pressure, mm Hg	127 (113-138)	130 (120-141)	126 (121-132)	134 (120-140)	130 (123-139)	0.219
Diastolic blood pressure, mm Hg	78.0 (70.0-87.0)	82.5 (73.0-88.0)	80.0 (77.0-87.0)	82.0 (76.0-88.0)	80.0 (79.5-84.0)	0.417
Body mass index, kg/m ²	25.8±3.41	25.1±2.11	24.9±1.88	25.2±2.74	25.3±2.54	0.768
Waist circumference, cm	90.7±12.7	88.5±8.18	87.0±8.65	88.9±10.5	90.0±11.1	0.676
Total cholesterol, mg/dL	306±55.4	296±42.1	307±46.5	296±33.8	315±55.6	0.534
HDL cholesterol, mg/dL	46.0 (39.0-58.0)	46.0 (36.8-54.0)	52.0 (44.0-64.5)	46.0 (38.0-63.0)	41.0 (37.0-56.5)	0.159
Non HDL cholesterol, mg/dL	243 (222-287)	245 (220-279)	241 (223-272)	234 (225-261)	270 (226-293)	0.741
LDL cholesterol, mg/dL	218 (192-250)	216 (189-247)	214 (190-242)	208 (196-227)	229 (184-269)	0.845
Triglycerides, mg/dL	145 (94.0-221)	130 (91.3-168)	133 (103-206)	179 (85.0-277)	270 (85.5-272)	0.892
Apolipoprotein A1,mg/dL	146±34.0	144±27.8	156±31.2	151±32.3	144±31.9	0.538
Apolipoprotein B, mg/dL	168±29.4	165±31.5	167±32.2	164±28.9	167±34.0	0.989
Lipoprotein(a), mg/dL	24.1 (12.0-55.0)	45.7 (15.5-69.2)	31.8 (13.0-75.2)	14.0 (5.27-37.1)	21.4 (9.54-48.7)	0.057

C reactive Protein, mg/L	1.55 (0.58-3.43)	2.75 (0.78-4.40)	1.30 (0.55-2.80)	2.60 (1.33-5.38)	2.00 (1.20-3.25)	0.108
GGT, IU/L	27.0 (18.0-38.0)	24.5 (17.8-32.0)	30.0 (15.5-50.5)	26.0 (18.0-65.0)	22.0 (15.5-38.0)	0.698
Glucose, mg/dL	88.7±10.2	90.2±10.3	87.5±9.73	93.6±11.0	92.8±15.0	0.181
HbA1c, %	5.10 (5.00-5.30)	5.40 (5.05-5.50)	5.20 (4.93-5.38)	5.30 (5.08-5.60)	5.30 (5.13-5.40)	0.053
<i>Dietary intake</i>						
Energy, kcal/day	2111 (1777-2466)	2104 (1695-2584)	2104 (1691-2545)	2093 (1803-2416)	2384 (2073-2879)	0.006
Carbohydrates, %	43.3±7.93	43.3±7.18	44.8±6.86	46.3±6.62	48.7±5.88	< 0.001
Protein, %	16.4±2.50	16.6±2.46	16.6±2.89	16.3±2.39	16.2±2.41	0.869
Fat, %	36.1±7.25	36.7±6.42	35.7±6.30	34.8±6.64	32.7±5.04	0.010
Monounsaturated fat, %	17.6±4.65	17.9±4.13	17.2±4.01	16.6±3.90	15.4±3.27	0.007
Polyunsaturated fat, %	4.30 (3.63-5.06)	4.69 (4.06-5.44)	4.66 (3.86-5.70)	4.53 (3.76-6.09)	5.01 (4.11-7.02)	0.056
Saturated fat, %	10.2±2.08	9.96±2.59	9.81±2.40	9.65±2.30	8.72±1.96	0.010
Cholesterol, mg/day ^b	347±129	328±109	349±101	314±89.7	358±125	0.219
Phytosterols, mg/day ^b	293 (230-340)	320 (257-391)	335 (274-410)	363 (314-417)	440 (374-520)	< 0.001
Fiber, g/day ^b	14.0 (12.3-16.9)	19.3 (16.6-22.1)	22.5 (20.0-26.5)	23.9 (21.8-29.1)	32.1 (28.2-37.0)	< 0.001
Alcohol, g/day ^b	6.74 (2.15-15.9)	8.23 (1.42-17.3)	5.08 (1.20-12.2)	4.38 (0.68-11.2)	4.38 (0.69-11.9)	0.296

^aData (10⁻²) are expressed as mean±standard deviation or median (percentile 25-percentile 75) as applicable. *P* refers to statistical differences among quintiles.

^bEnergy-adjusted.

Table Supplementary 2. Clinical, biochemical and dietary characteristics of subjects among quintiles of “Western dietary pattern” scores.^a

	Q1 N = 56	Q2 N = 57	Q3 N = 57	Q4 N = 57	Q5 N = 57	<i>p</i>
ADH/ FCH, %	18.4/22.3	17.9/24.5	24.2/11.7	21.1/18.1	18.4/23.4	0.099
Males, %	19 (33.9)	31 (54.4)	25 (43.9)	29 (50.9)	34 (59.6)	0.061
Age, years	48.1±9.95	46.7±9.97	44.4±8.85	43.8±10.1	39.9±9.47	0.010
Tobacco consumption, %						
Smoker	13 (23.2)	9 (15.8)	16 (28.6)	13 (22.8)	26 (45.6)	0.021
Former smoker	19 (33.9)	17 (29.8)	18 (32.1)	15 (26.3)	8 (14.0)	
Non smoker	24 (42.9)	31 (54.4)	22 (39.3)	29 (50.9)	23 (40.4)	
Packets/day x Years	24.4 (8.50-36.0)	20.0 (8.75-34.3)	17.0 (10.0-32.8)	14.0 (9.13-22.9)	18.0 (10.0-31.0)	0.767
Systolic blood pressure, mm Hg	130 (120-134)	132 (122-140)	130 (120-139)	130 (123-139)	128 (117-138)	0.548
Diastolic blood pressure, mm Hg	80.0 (72.3-87.0)	83.5 (80.0-89.0)	80.0 (74.3-88.0)	82.0 (80.0-86.0)	80.0 (71.5-85.5)	0.219
Body mass index, kg/m ²	25.3±2.56	25.8±2.36	25.3±2.87	25.4±2.09	24.7±2.92	0.618
Waist circumference, cm	89.1±11.1	90.7±9.937	88.3±10.8	89.4±9.16	88.2±11.1	0.899
Total cholesterol, mg/dL	309±42.6	299±40.2	292±44.1	318±55.9	301±49.6	0.240
HDL cholesterol, mg/dL	47.0 (40.0-58.8)	45.0 (38.8-60.8)	49.0 (39.0-62.3)	52.0 (36.3-62.8)	43.0 (34.8-52.0)	0.529
Non HDL cholesterol, mg/dL	271 (221-284)	254 (222-271)	230 (213-251)	252 (227-297)	241 (223-283)	0.148
LDL cholesterol, mg/dL	226 (191-261)	226 (181-246)	202 (189-224)	218 (200-268)	222 (196-240)	0.566
Triglycerides, mg/dL	143 (94.0-277)	158 (98.0-210)	124 (89.8-185)	140 (89.8-235)	154 (94.0-300)	0.859
Apolipoprotein A1, mg/dL	153±28.7	156±31.6	151±28.1	147±34.8	136±32.0	0.097
Apolipoprotein B, mg/dL	174±31.7	167±28.9	159±33.3	168±30.2	164±28.6	0.365
Lipoprotein(a), mg/dL	20.5 (6.77-32.7)	26.8 (12.0-73.4)	25.7 (12.7-62.7)	16.0 (7.05-51.3)	43.5 (12.5-64.4)	0.208
C reactive Protein, mg/L	1.80 (0.60-4.10)	1.65 (0.50-2.50)	1.50 (0.70-3.55)	2.60 (0.95-4.03)	2.30 (0.90-3.85)	0.626
GGT, IU/L	25.5 (19.3-50.3)	30.5 (16.0-48.8)	25.5 (16.0-33.8)	21.5 (15.5-51.3)	26.0 (18.0-32.0)	0.818

Glucose, mg/dL	89.3±9.00	91.8±10.3	90.4±13.0	89.6±13.9	91.3±10.1	0.907
HbA1c, %	5.30 (5.10-5.48)	5.25 (4.88-5.50)	5.20 (5.00-5.40)	5.30 (5.10-5.40)	5.20 (5.10-5.48)	0.765
<i>Dietary intake</i>						
Energy, kcal/day	2054 (1684-2341)	2091 (1680-2600)	1940 (1659-2244)	2364 (1972-2701)	2454 (2140-2865)	< 0.001
Carbohydrates, %	42.9±7.87	45.3±6.89	44.1±6.81	47.4±6.38	46.7±7.09	0.004
Protein, %	16.4±2.62	16.7±2.65	17.4±2.44	16.2±2.28	15.5±2.34	0.002
Fat, %	37.1±7.41	34.8±6.12	35.4±6.39	33.9±5.37	34.8±6.71	0.100
Monounsaturated fat, %	18.6±5.02	17.1±3.69	17.2±3.93	15.8±3.48	16.1±3.67	0.002
Polyunsaturated fat, %	4.98 (4.05-5.52)	4.57 (3.89-5.38)	4.43 (3.74-5.85)	5.57 (3.71-5.68)	4.72 (3.80-7.32)	0.706
Saturated fat, %	9.29±2.31	9.42±2.18	9.76±2.45	9.78±2.22	10.0±2.42	0.451
Cholesterol, mg/day ^b	302±101	338±107	311±95.1	360±118	384±119	< 0.001
Phytosterols, mg/day ^b	341 (284-405)	354 (266-430)	311 (269-359)	377 (320-447)	389 (316-470)	0.002
Fiber, g/day ^b	22.5 (17.7-26.6)	21.9 (18.7-29.6)	22.1 (16.7-27.7)	23.5 (19.3-31.9)	21.6 (15.8-28.0)	0.344
Alcohol, g/day ^b	4.42 (0.68-15.5)	5.08 (0.35-16.3)	5.14 (1.42-15.2)	4.38 (1.46-11.8)	5.92 (1.79-15.6)	0.811

^aData (10⁻²) are expressed as mean±standard deviation or median (percentile 25-percentile 75) as applicable. *P* refers to statistical differences among quintiles.

^bEnergy-adjusted.