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Howarth, ERI, Szott, ID, Witham, CL, Wilding, CS and Bethell, EJ (2023) Genetic polymorphisms in the serotonin, dopamine and opioid pathways influence social attention in rhesus macaques (Macaca mulatta). PLoS One, 18 (8). pp. 1-29. ISSN 1932-6203

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Citation: Howarth ERI, Szott ID, Witham CL, Wilding CS, Bethell EJ (2023) Genetic polymorphisms in the serotonin, dopamine and opioid pathways influence social attention in rhesus macaques (*Macaca mulatta*). PLoS ONE 18(8): e0288108. https://doi.org/10.1371/journal.pone.0288108

Editor: Asem Surindro Singh, University of Cincinnati, INDIA

Received: March 9, 2023
Accepted: June 20, 2023
Published: August 2, 2023

Peer Review History: PLOS recognizes the benefits of transparency in the peer review process; therefore, we enable the publication of all of the content of peer review and author responses alongside final, published articles. The editorial history of this article is available here: https://doi.org/10.1371/journal.pone.0288108

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Data Availability Statement: All relevant data are within the paper and its Supporting information files.

RESEARCH ARTICLE

Genetic polymorphisms in the serotonin, dopamine and opioid pathways influence social attention in rhesus macaques (*Macaca mulatta*)

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Abstract

Behaviour has a significant heritable component; however, unpicking the variants of interest in the neural circuits and molecular pathways that underpin these has proven difficult. Here, we present a comprehensive analysis of the relationship between known and new candidate genes from identified pathways and key behaviours for survival in 109 adult rhesus macaques (Macaca mulatta). Eight genes involved in emotion were analysed for variation at a total of nine loci. Genetic data were then correlated with cognitive and observational measures of behaviour associated with wellbeing and survival using MCMC-based Bayesian GLMM in R, to account for relatedness within the macaque population. For four loci the variants genotyped were length polymorphisms (SLC6A45-hydroxytryptamine transporter length-polymorphic repeat (5-HTTLPR), SLC6A4 STin polymorphism, Tryptophan 5hydroxylase 2 (TPH2) and Monoamine oxidase A (MAOA)) whilst for the other five (5hydroxytryptamine receptor 2A (HTR2A), Dopamine Receptor D4 (DRD4), Oxytocin receptor (OXTR), Arginine vasopressin receptor 1A (AVPR1a), Opioid receptor mu(μ) 1 (OPRM1)) SNPs were analysed. STin genotype, DRD4 haplotype and OXTR haplotype were significantly associated with the cognitive and observational measures of behaviour associated with wellbeing and survival. Genotype for 5-HTTLPR, STin and AVPR1a, and haplotype for HTR2A, DRD4 and OXTR were significantly associated with the duration of behaviours including fear and anxiety. Understanding the biological underpinnings of individual variation in negative emotion (e.g., fear and anxiety), together with their impact on social behaviour (e.g., social attention including vigilance for threat) has application for managing primate populations in the wild and captivity, as well as potential translational application for understanding of the genetic basis of emotions in humans.

Funding: This research was supported by a National Centre for the Replacement, Refinement and Reduction of Animals in Research (NC3Rs) grant NC/L000539/1to EJB. ERIH was supported by a Liverpool John Moores PhD studentship. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing interests: The authors have declared that no competing interests exist.

Introduction

In humans (*Homo sapiens*) and many other animal species, a significant heritable component of personality [1, 2] and behaviour [3, 4] has been demonstrated. Whilst identification of genetic variants that are consistently associated with such traits would allow a better understanding of the specific neural circuits and molecular pathways that underpin emotional behaviours [5, 6], due to the complexity and number of these pathways involved in behavioural responses, unpicking the variants of interest has often proven difficult.

Phenotypic traits, such as personality, are both quantitative (non-binary) and usually polygenic [7]. Identifying the loci underpinning behavioural traits has sometimes involved their study in mapping populations created by experimental crosses [8]. Such quantitative trait locus (QTL) analysis has led to the identification of specific QTL relating to, for example. avoidance behaviours [9], anxiety [10] and aggressive behaviour [11]. However, QTL are typically broad, and require substantive follow-up work to identify causal genes within the identified QTL. More recently, genome wide association studies (GWAS) in which associations between genotype and phenotype are examined in populations of unrelated individuals have been utilised to study a range of behavioural conditions (e.g., risk-taking [12], antisocial behaviour [13], anxiety [14]).

Whilst there are numerous genomic resources for rhesus macaques (*Macaca mulatta*) [15] due to the genomic nature of the data needed, QTL and particularly GWAS can, however, be expensive. Candidate gene studies involving genotyping variants at a smaller number of preselected candidate loci have long been a more affordable and accessible approach for genetic analysis [16].

With this cost in mind, this study aimed to provide the most comprehensive analysis to date of the relationship between individual candidate genes from identified pathways and associated behaviours and vigilance for threat in adult rhesus macaques. We first review the primate literature for each pathway (with relevant human studies also included, especially where primate data are lacking) to justify our choices of candidate loci. Table 1 provides a summary of the pathways, genes, and variants of interest in this study and an overview of whether published primate and human data are available.

Serotonin pathway

A key candidate gene in the study of anxiety and stress-related disorders is the serotonin transporter (5-hydroxytryptamine transporter or 5-HTT) gene (also known as *SLC6A4* (Solute Carrier Family 6 Member 4)). Serotonin (5-hydroxytryptamine or 5-HT) is a neurotransmitter and 5-HTT regulates the signaling and concentration of synaptic 5-HT [18]. The neural circuitry controlling temperament and mood relies on 5-HT synapses and disturbances in this system result in many psychiatric disorders in humans [18]. *SLC6A4* displays three well-studied polymorphisms in humans: a guanine/thymine (G/T) SNP in a non-coding 3'-untranslated region (3'-UTR), an insertion / deletion polymorphism in the promoter region (the 5-HTT length-polymorphic repeat or 5-HTTLPR) and a 16–17 bp variable number of tandem repeats (VNTR) polymorphism located in intron 2 (*STin*) [19, 20].

The most widely studied polymorphism is the *5-HTTLPR* for which, in humans, the predominant alleles are a 14-repeat short allele (s-allele) and a 16-repeat long allele (l-allele) [18, 21]. The s- and l-alleles differ in their rate of serotonin transporter transcription, with the s-allele having a lower rate of transcriptional efficiency than the l-allele, such that individuals homozygous for the s-allele or who are heterozygous have around a 65% lower 5-HTT mRNA expression level than homozygous l-allele individuals [21, 22]. A similar length variable allelic system is found in rhesus macaques, though here, the alleles are a 23-repeat s-allele and a

Table 1. Candidate SNP or length variants for behavioural phenotypes in primates. Further details are provided in the text. Variant ID refers to the location in the *Macaca mulatta* genome (Mmul_10 assembly [17]).

Pathway	Gene	Variant ID (SNP only) / location (LP only)	Type of variant	Previous published studies	
Serotonin	<i>SLC6A4</i> 5-hydroxytryptamine transporter length-polymorphic repeat (<i>5-HTTLPR</i>)	16:25897958-25898456	Insertion / deletion LP	Primates: ✓ Humans: ✓	
	SLC6A4 STin polymorphism	16:25,882,595–25,882,709	16–17 bp VNTR LP	Primates: X Humans: ✓	
	Tryptophan 5-hydroxylase 2 (TPH2)	11:71,601,619	159 bp insertion LP (TPH2IP) in the 3'-UTR	Primates: ✓ Humans: ✓	
	5-hydroxytryptamine receptor 2A (HTR2A)	rs80365915	Missense SNP	Primates: X Humans: X	
		rs80363349	5' UTR SNP		
		rs196407124	5' UTR SNP		
Dopamine / serotonin	Monoamine oxidase A (MAOA)	X:43,576,386-43,576,511	Length	Primates: ✓ Humans: ✓	
Dopamine	Dopamine Receptor D4 (DRD4)	rs300413141	Upstream SNPs	Primates: X Humans: X	
		rs1079355788			
		rs290724315			
		rs301203363			
Oxytocin	Oxytocin receptor (OXTR)	rs196783445	Intronic SNPs	Primates: X Humans: X	
		rs292502465			
		rs300857875			
		rs308701533			
		rs292035217			
		rs302789768			
		rs283226059			
	Arginine vasopressin receptor 1A (AVPR1a)	11:62,913,090	8bp repeat	Primates: X Humans: X	
Opioid	Opioid receptor mu(μ) 1 (OPRM1)	rs195917455		Primates: ✓ Humans: ✓	

Abbreviations: SNP-single-nucleotide polymorphism; LP- length polymorphism; UTR-untranslated region; VNTR-variable number of tandem repeats.

https://doi.org/10.1371/journal.pone.0288108.t001

24-repeat l-allele. Whilst the effect of these alleles on transcriptional efficiency *in vitro* has not been studied, since 5-HTT genotype plays a role in serotonin regulation, which affects hypothalamic-pituitary-adrenal (HPA) axis function [23] phenotypic effects of genotype at this locus are expected and suggested in some studies. S-allele homozygous infant and juvenile monkeys (n = 128) displayed more anxious and threatened behaviour than l-allele carriers during novel fruit, human intruder, remote-controlled car, and free play tests [24]. Also, interactions between genotype and thalamic levels of 5-HTT, and dysregulation of both the HPA axis and serotonergic system, have been shown to be associated with negative mood states [25], the control of arousal, depression, and anxiety [26, 27]. In a study testing the effect of chronic stress caused by social conflict on 29 female rhesus macaques, 5-HTTLPR genotype was found to influence cortisol response [28]. Individuals homozygous for the s-allele had higher hair cortisol compared to heterozygous or l-allele homozygous individuals. As chronic stress and heightened cortisol levels are associated with the onset of depression in adult female macaques [29, 30], they may be more susceptible to depression if they have low serotonin transporter efficiency (s-allele) and a history of stress.

By contrast to *5-HTTLPR*, the function of the VNTR polymorphism *STin* is less well studied [31]. In humans and primates, including macaques, there are several known alleles for *STin* [32, 33]. In humans, the three most common are *STin*2.9, *STin*2.10 and *STin*2.12, containing

9, 10 and 12 copies of the VNTR element respectively [31, 33]. *In vitro* studies with human embryonic stem cells have shown genotype-dependent reporter gene expression in embryonic stem cells with *STin*2.12 increasing gene expression 29-fold compared to *STin*2.10 [34]. Phenotypic studies have explored the association of these polymorphisms with e.g., tobacco use [20] and migraine susceptibility [33]. The *STin*2.12 allele has also been associated with obsessive-compulsive disorder (OCD) [35]. However, there have been discrepancies and conflicting data published on the effect of the *STin* alleles on psychological disorders [36, 37]. For example, Kaiser et al [38] reported a six- fold increase in the risk of developing a subtype of schizophrenia in human patients that carry the *STin*2.9 allele, with the allele being significantly associated with an increased risk of unipolar disorder and depression [39], while the *STin*2.10 allele has been reported as a predictor of suicide attempts in female members of the Dubla tribe of Daman [40].

The *tryptophan 5-hydroxylase 2* gene (*TPH2*) is also within the serotonin pathway [41]. *TPH2* encodes the protein catalyst, tryptophan hydroxylase, for the first and rate-limiting step in the biosynthesis of serotonin [42]. In macaques there are several known *TPH2* polymorphisms: two mononucleotide repeats, one dinucleotide repeat, 17 SNPs and, a 159 bp insertion polymorphism (*TPH2IP*) in the 3'-UTR [43]. In luciferase reporter gene assays the *TPH2IP* had a profound effect on gene expression, with the l-allele driving substantially higher gene expression than the s-allele. In humans, similar 3'-UTR length polymorphisms have been associated with ADHD [44], bipolar disorder [45] and suicide [46] and, in rhesus macaques, with altered HPA axis functioning and aggressive behaviour [47].

The 5-hydroxytryptamine receptor 2A gene (HTR2A) encodes the 5-HT $_{2A}$ receptor for serotonin, which is expressed mainly in the brain, including in the hippocampus, olfactory tubercle, nucleus accumbens, caudate nucleus and neocortex [48]. Polymorphisms in HTR2A have been associated with neuropsychiatric disorders including impulsive behaviour and schizophrenia [49, 50]. For example, in humans, carriers of the minor A-allele of a SNP \approx 2kb upstream of HTR2A experienced greater depressive symptoms than individuals who are homozygous for the G-allele [51].

No specific candidate polymorphisms have been previously identified in macaques. Here, sequencing of the 5'UTR and first coding exon was undertaken to genotype three SNPs, one exonic (rs80365915) and two SNPs rs80363349 (A/G) and rs196407124 (A/C) in the 5'-UTR. They have not been previously evaluated for association with anxiety or stress-related disorders.

Multiple pathways (serotonin & dopamine)

Monoamine oxidase A is involved in the degradation of circulating serotonin [52]. This mitochondrial enzyme catalyses the oxidative deamination of amines, including serotonin, dopamine, noradrenaline, and adrenaline and is encoded by the *monoamine oxidase A* gene (*MAOA*) [53]. A key polymorphism associated with aggressive behaviour in rhesus macaques are 5-, 6- and 7–18 bp repeat alleles located within the transcriptional control region of the *MAOA* gene [54]. *MAOA* is an X-linked gene and females can be heterozygous or homozygous while males can only be hemizygous (only one member of the chromosome pair or segment is present rather than two) [53]. The 5- and 6- repeat heterozygous, homozygous, or hemizygous variants have a significantly higher activity than the 7-repeat, resulting in a lower rate of degradation of circulating serotonin in 7-homozygotes [54, 55]. There is little *in vitro* or *in vivo* evidence of whether 5/7 and 6/7 repeat heterozygous variants have high or low activity, therefore, it is not known whether they result in a higher or lower rate of serotonin degradation [54, 55]. In the present study, the 5-, 6- and 7–18 bp repeat alleles have been investigated for their link to anxiety and attention bias (AB).

Dopamine pathway

Dopamine (3, 4-Dihydroxytyramine) is associated with reward-motivated behaviour and memory [56]. Dopamine is a catecholamine that can act as a hormone which is released from the hypothalamus, or a neurotransmitter that activates the dopamine receptors [41, 57, 58]. There are five known dopamine receptors (Dr1-5) subdivided into two categories: D1-like receptors (Dr1 and Dr5) and D2-like receptors (Dr2-4). The *Dopamine Receptor D4* gene (*DRD4*) encodes the G-protein coupled D4 subtype of the dopamine receptor [59]. The receptor activates pertussis toxin-sensitive G-proteins [60], inhibits adenylyl cyclase activity and mediates dopamine activity in the central nervous system (CNS) [61]. In humans, *DRD4* contains several polymorphisms including a 48 bp VNTR in the third exon of the gene with the 7-repeat allele associated with psychiatric disorders including ADHD, bulimia, and alcoholism [62, 63]. This allele is also associated with anger, aggression, and delinquency in humans [64, 65] and avoidance of mothers and conspecifics in juvenile rhesus macaques [66]. Here, we sequenced a region of the *DRD4* promoter region to look for associations between alleles at four SNPs (rs30041314, rs1079355788, rs290724315, rs301203363) and AB.

Oxytocin pathway

Oxytocin is a neuromodulatory posterior pituitary hormone that is associated with increased parasympathetic functioning and plays a counter-regulatory role in stress and fear responses [67, 68]. The *oxytocin receptor* (*OXTR*) is a potential key candidate gene in the study of oxytocin levels and the downstream effects of these. *OXTR* encodes G-protein coupled receptor proteins for oxytocin. Polymorphisms in human *OXTR*, including the most studied *OXTR* SNPs rs53576 and rs2254298, are associated with social behaviour and emotional responsiveness [69], reduced positivity [70], and poor social recognition skills [71]. Both rs53576 and rs2254298 are intron variants. For rhesus macaques, there are no previously characterised SNPs for *OXTR*.

OXTR encodes an oxytocin receptor, a member of a subclass of peptide receptors that also includes the arginine vasopressin receptors (1A, 1B, 2). Arginine vasopressin is a neuropeptide and the arginine vasopressin receptor 1A gene (AVPR1a) encodes the vasopressin V1a receptor. There are three known length variant polymorphisms in humans: RS1 a (GATA)₁₄ tetranucleotide repeat, RS3 a complex (CT)₄-TT-(CT)₈-(GT)₂₄ repeat and STR1 a (GT)₂₅ dinucleotide repeat [72]. RS1 and RS3 are located within two blocks, known as DupA and DupB respectively, of a ~350 bp tandem duplicate region [73]. Only the DupB region, and therefore RS3, is found in rhesus macaques [74]. Variation within RS3 is associated with altruism [75], pairbonding behaviour [76], social behaviour including sibling conflict [77] and reproductive behaviour [78]. Walum et al [79] reported 11 RS3 alleles in humans (320, 330, 332, 334, 336, 338, 340, 342, 344, 346, 348 bp). The 334bp allele was the most common allele in the study (40%) and was associated with marital problems with homozygous 334 individuals having double the risk of a marital crisis compared to 334 non-carriers. In this study, AVPR1a RS3 alleles were investigated for their association with anxiety and stress-related behaviours.

Opioid pathway

The endogenous opioid system, including the opioid receptors and opioid peptides, is implicated in the process for reinforcement and reward in the brain and nervous system by interaction with specific receptors [80]. The *opioid receptor mu(\mu) 1 (OPRM1)* gene encodes the μ -opioid receptor which is the principal target of endogenous opioid peptides and opioid analgesic agents such as beta-endorphin and enkephalins, which are associated with neurotransmission and pain modulation [81]. Positive reinforcement is mediated by both indirect (nicotine,

cannabinoids, alcohol) and direct (morphine) activation of the μ -opioid receptor [82]. Mechling et al [83] demonstrated that the μ -opioid receptor shapes the reward/aversion circuitry in the brain of rats. In humans, G-allele carriers at an A/G SNP in *OPRM1* have a dopamine-mediated lowered response to reward during positive reinforcement learning [84]. As a result of the link between reward and reinforcement, the *OPRM1* gene is associated with addiction in humans [85] whilst a different polymorphism (C77G) is linked to increased levels of alcohol consumption in male macaques [86]. This polymorphism is also associated with increased aggressive-threat behaviours in rhesus macaques [87] with G-allele carriers also showing higher baseline attachment behaviours than CC homozygous individuals [88]. Here, the relationship between the C77G polymorphism (rs195917455) and anxiety in rhesus macaques was investigated.

Attention bias

AB describes a tendency to preferentially attend to emotional compared to neutral cues and is influenced by underlying affect [89, 90]. AB is a new approach to assessing animal cognition, affect and anxiety that has shown significant reproducibility between measures in rhesus macaques [91, 92]. Previous studies have shown that genotype can affect social attention in humans [93, 94]. However, there is no published evidence for an association between AB and genotype in rhesus macaques and the extent to which genetic variation influences attention to potentially threatening stimuli is not yet fully understood.

Behaviour

Stressful events can have a significant effect on behaviour [95]. When faced with a potential threat, survival may depend on an appropriate behavioural change associated with either the active fight-flight or passive freeze-hide responses [96]. This behavioural change is often the most overt, easily observable, and biologically economical response to stress as a threat may be avoided by removing oneself from the stimulation, for example, an animal pursued by a predator will avoid the danger by escaping and a heat-stressed animal may move to find water or shade [97]. However, actively avoiding a stressor may not be possible for captive animals as this environment is associated with a range of uncontrollable factors from which the animal cannot escape, for example, inappropriate artificial lighting schedules, temperature variation away from their preferred ranges, space limitations and, sounds and smells of predator species [98]. Captive animals have limited or no opportunities for changing their spatial proximity to conspecifics and caretakers, environmental conditions, and nutrition.

Captive animals will have a different behavioural repertoire to their free-ranging counterparts [99]. However, free-ranging behaviours should be considered as it provides a reference of species-typical behaviour [100]. Free-ranging macaques spend most of their time allogrooming, resting, foraging, and exploring their environment [101–103]. These behaviours should apply to captive group or pair-housed macaques and a change in the magnitude or frequency of these key species-typical behaviours can be indicative of an underlying issue, such as ill health, pain, or stress [104]. In macaques, stress and anxiety can manifest as displacement behaviours and aggression [105]. Camus et al [105] reported five distinct behavioural profiles when recording spontaneous atypical behaviour in 40 captive cynomolgus macaques (*M. fascicularis*). One behavioural profile (E) was associated with a high occurrence of aggressive and displacement behaviours. Displacement behaviours are acts performed by the animal that are irrelevant to the behavioural context [106]. It is thought that these behaviours may occur as a form of energy dissipation or as a conflict between two competing motivations [107]. In primates, displacement behaviours include yawning, shaking, auto-grooming, and scratching

[108]. Facial expressions form a complex suite of behaviour associated with stress and communication. A lip smack is a characteristic affiliative or appeasement display, while a fear grimace, indicated by bared teeth, is a sign of fear or social submission [109]. Macaque facial movements have been described using a macaque Facial Action Coding System (MaqFACS) revealing similarity in facial expression across primate species [110]. However, several speciesspecific specialisations were also described with macaques having more independent control over their ear movements compared to humans or chimpanzees (Pan troglodytes) [110]. Ear movements are thought to play an important role in macaque social communication as grimace and lip-smack displays include prominent ear movements [111]. Enhanced vigilance is a further indicator of anxiety [108] with hypervigilance an indicator of extreme stress in many species [112]. The function of this behaviour is to detect social and predatory threats and, in response to stress, vigilance behaviour increases [113]. Socially stressed, subordinate macaques engage in vigilant scanning more frequently than dominant animals [114]. Vigilance is commonly accompanied by appearement (lip smack) or fear (grimace) behaviours [115, 116]. Here, we examine variation at the nine previously described candidate loci and examine the role of variation at these loci in describing behaviour and AB in a group of captive rhesus macaques.

Materials & methods

Ethics

Protocols were developed following discussion with the facility Home Office Inspector (Nov 2011) and carried out in accordance with ethical guidelines for work with non-human primates [117]. Approval was granted by the Medical Research Council (MRC) Animal Welfare and Ethical Review Body (AWERB) in 2014, Roehampton University Ethics Committee (approval #LSC 14/113) and the Liverpool John Moores University (LJMU) ethics panel (approval #EB/2014-1). Further approval was granted by the MRC AWERB in November 2017, and the LJMU ethics panel (approval #EB_EH/2017-5). Animal health was monitored daily by the care staff, and annually with a full veterinary examination. No regulated procedures were carried out for this study and at no point were animals removed from their home colony at any time. Participation was voluntary insofar as only animals who approached the apparatus for rewards took part. Blood samples for DNA analysis were collected by a trained veterinarian with the primary purpose of colony management. Methods and results are reported according to the ARRIVE guidelines [118].

Animals

Data were collected from 109 adult rhesus macaques (*Macaca mulatta*; 94 female) housed at the Centre for Macaques, MRC Harwell Institute (MRC-CFM), UK (mean age on first day of testing = 8.83 years, range = 2.5–18.3 years). The centre is licenced by the Home Office to breed macaques for provision to UK facilities. Rhesus macaques are a female philopatric species that have a linear hierarchy based on female relatedness [119]. To reflect their typical social system, monkeys at MRC-CFM are housed in social breeding groups comprising one adult male and breeding females. The monkeys that took part in the current research were housed in 29 breeding groups comprising one adult male and between three and 11 related females, plus infants and juveniles. Social rank for each female was categorised on a scale of high-mid-low, determined through consultation with facility staff who care for the animals on a daily basis, and from our published data [120]. All monkeys had previously been trained to station next to a target in order to take part in AB testing [120] and were familiar with researchers at the time testing was conducted. Each group had access to a free-roaming room

 $(3.35 \,\mathrm{m} \times 8.04 \,\mathrm{m} \times 2.8 \,\mathrm{m})$ and an adjacent cage area $(1.5 \,\mathrm{m} \times 6.12 \,\mathrm{m} \times 2.8 \,\mathrm{m})$, accessible through hatches, with a minimum total space of $3.5 \,\mathrm{m}^3$ /breeding animal in the largest groups. Each free-roaming room had a large bay window at one end facing outdoors and allowing a natural day-night cycle. At the other end of each room was an internal window into the hallway used by staff. Internal windows were fitted with movable mirrors so that monkeys could manipulate the mirrors to view activities along the corridor. Rooms were furnished with wooden platforms and poles (horizontal, vertical, diagonal), fire hose, ladders, plastic horse jumps and saddle racks, PVC piping, plastic barrels and balls, and small plastic blocks attached to structures or walls. The floor was covered with a deep layer of straw and shavings. All rooms were temperature controlled $(20\,^{\circ}\mathrm{C} \pm 5)$ with humidity at $55\% \pm 10\%$.

Animals were free to move between the room and cage area at all times during this research, and at no point were the hatches used to retain animals. Adjacent groups were able to see and hear each other from the cage area, but there was no possibility for physical contact. All testing took place in the cage area, which monkeys entered voluntarily, with open access to the free-roaming room at all times. The cage area had three tiers with open access points between them. Individuals tended to have preferred levels they would spend time in and so staff and researchers would work with an individual at their preferred location, a key factor when working with group housed animals.

The macaques were fed twice daily by scatter feed, morning and afternoon, with sufficient food to last for a 24 h period. The diet varied daily and included a dried forage mix (cereal, peas, beans, lentils etc.), a range of fruit and vegetables, bread and boiled eggs. Water was available *ad libitum* in both the room and cage area at all times. Further details, images and video of the facility can be found at: https://www.mrc.ac.uk/research/facilities-and-resources-for-researchers/mrc-centre-for-macaques/ and www.nc3rs.org.uk/macaques.

DNA collection

Blood samples were collected by the Named Veterinary Surgeon at MRC-CFM during the macaques' annual health screening. Macaques were sedated with KHCl prior to this procedure. Blood samples, which were primarily used for colony management purposes, were collected into EDTA K2 (anticoagulant) tubes, centrifuged, and wrapped in cotton wool prior to transport to B&K Universal Laboratories (Marshall BioResources), Grimston, East Yorkshire, UK. DNA extraction occurred upon arrival at the laboratory using the DNeasy Blood & Tissue Kit from Qiagen (Catalogue no. 69506) and concentrations and purity measured using a Thermo Fisher Scientific Inc. NanoDrop™ Lite Spectrophotometer. The DNA samples were stored at -20°C at Liverpool John Moores University, James Parsons Building, Liverpool, UK until required for analysis. Samples were stored at 4°C during use.

Genotyping

Eight genes were analysed for variation at a total of nine loci. For four loci the variants genotyped were length polymorphisms (5-HTTLPR, STin, TPH2 and MAOA) whilst for the other five (HTR2A, DRD4, OXTR, AVPR1a and OPRM1) SNPs were analysed. Samples were genotyped for the four length variants using PCR amplification and gel-analysis. SNPs for HTR2A, DRD4, and OXTR were determined by amplification and sequencing, with haplotypes manually reconstructed from multi-locus genotype data, while SNPs for AVPR1a were determined through amplification followed by restriction enzyme digestion and gel-analysis [121]. Seven intron variants for OXTR (rs196783445, rs292502465, rs300857875, rs308701533, rs292035217, rs302789768, rs283226059) were identified using Ensembl [122] all within a short intron tractable for PCR and sequencing. A TaqMan genotyping assay was carried out

for the *OPRM1* C77G polymorphism [123]. PCR cycle times and temperatures were adapted from established protocols and optimised for the current study (<u>Table 2</u>). All PCR products were visualised using GelRed stain during electrophoresis; however, GelRed caused inconsistencies in the accurate sizing of the *5-HTTLPR* length polymorphism. Instead, ethidium bromide was used as the visualization dye for *5-HTTLPR*.

Genetic data preparation

The final dataset consisted of genotypes from nine loci for each monkey (S1 Table). Variant information for each genotype and associated behaviours from the literature are shown in Table 2. For, *HTR2A*, *DRD4* and *OXTR*, the SNPs (3, 4 and 7 respectively) were manually phased into haplotypes prior to analysis. For *AVPR1a*, there were two upstream polymorphisms: a SNP (rs194455405) and an 8bp insertion (11:62913090) which were combined to create three haplotypes. For the genes for which sufficient published data were available (5-HTTLPR, STin, TPH2, MAOA, OPRM1) genotypes were reclassified prior to statical analysis as high or low expressing based on their known biological effects (high or low risk / expression; Table 3).

Attention bias

A full AB protocol is presented in Howarth et al [92]. In brief, unfamiliar threat-neutral male conspecific face pair stimuli were shown to macaques for three seconds and their looking time to each stimulus pair recorded. Looking time towards face pair stimuli was blind coded from video [92]. Cognitive data were then averaged for baseline (assumed non-stressed state) and stress (following veterinary intervention) to create an AB profile for each animal.

Behaviour

An ethogram of behavioural indices (Table 4) was constructed to include key behaviours related to aggressive, anxiety, distraction, fear, foraging, prosocial and inactive behaviour. These categories were chosen as they have previously been identified as key behavioural indices for wellbeing in rhesus macaques [105, 108, 109].

Continuous focal animal behavioural observations were completed using Behavioral Observation Research Interactive Software (BORIS) [129]. Observations were conducted following each AB trial with data collected on four consecutive weekdays (Tuesday—Friday) plus the following Monday. Following completion of the AB trials, groups were given 10 minutes to settle without observation before each monkey was observed for five minutes. Observations were conducted within 60 minutes of the cognitive trials.

Cognitive data preparation

Cognitive data for social attention comprised two continuous response variables: time spent looking at the threat face per three second trial (THR; range = 0 < 3000ms) was included to assess interest in threat faces specifically, and total time spent looking at the threat-neutral face pair overall per trial (TL; range = 0 < 3000ms) was included to assess interest in social images regardless of emotion content.

To reduce the potential influence of outliers, following Howarth et al. [92], we removed cognitive trials for which a disruption was deemed to have occurred. Disruptions may be associated with increased stress, which our previous work has shown influences AB to threat [91]. Disruptions were identified as: receiving treatment for illness, a change to group membership

Table 2. Primer pairs, location, and PCR conditions for amplification of 5-HTTLPR, STin, TPH2, HTR2A, MAOA, DRD4, OPRM1, OXTR, and AVPR1a. All genomic positions are from Mmul_1 [122, 124].

Gene Variant type		Primer and genomic location	PCR cycles & temperatures	Source		
5-HTTLPR	Length	5HTT-F (5'-GCCGCTCTGAATACCAGCAC-3') HTTLPR_intl (5'-CAGGGGAGATCCTGGGAGGG-3') Chromosome 16: 25,898,078-25,898,491	95°C for 5 mins 40 cycles: 95°C for 30 s, 61°C for 30s, 72°C for 1 min Final extension: 72°C for 7 mins	[125, 126]		
STin	Length	STin-F (5'-TGTTCCCAGACTTACACCAGTG-3') STin-R (5'-GTCAGTATCACAGGCTGCGAG-3') Chromosome 16: 25,882,527-25,882,740	95°C for 3 mins 35 cycles: 95°C for 30s, 55°C for 30s, 72°C for 1 min Final extension: 72°C for 5 mins	Primers based on human primers from Ogilvie et al [39] redesigned based on <i>M. mulatta</i> sequence		
TPH2	Length	TPH2-U3F5 (5'-TGTAGGAAACTTCTCATCACAA-3') TPH2-U3R5 (5'-CAGCATAAAAATTCATAGTCCCAAG-3') Chromosome 11: 71,601,416-71,601,688 forward strand	95°C for 3 mins 35 cycles: 94°C for 30s, 55°C for 30s, 72°C for 30s Final extension: 72°C for 5 mins	[43]		
HTR2A	SNP	HTR2A-F (5'-GGCATGACAAGGAAACCCAG-3') HTR2A-R (5'-CCAGGACATTTATCTCCCCGA-3') Chromosome 17: 25,340,395-25,341,115	95°C for 3 mins 35 cycles: 95°C for 30s, 55°C for 30s, 72°C for 1 min Final extension: 72°C for 5 mins	Novel primers, no published PCR protocols		
MAOA	Length	MAOA-F(2) (5'-CAGAAACATGAGCACAAACG-3') MAOA-R(2) (5'-TACGAGGTGTCGTCCAAGTT-3') Chromosome X: 43,576,337-43,576,618	95°C for 5 mins, 40 cycles: 94°C for 30s, 55°C for 30s, 72°C for 30s Final extension: 72°C for 10 mins	[127]		
DRD4 haplotype	SNP	DRD4-PROM-SNP-F (5'-CGGGGGCTGAGCACCAGAGGCTGCT-3') DRD4-PROM-SNP-R (5'-GCATCGACGCCAGAGCCATCCTGCC-3') Chromosome 14: 694,533-694,799	95° for 1min 35 cycles: 95° for 20s, 72°C for 30s final extension: 72°C for 7 mins	[128]		
OXTR	SNP	OXTR- F (5'-CTGGACGCCTTTCTTCTTCG-3') OXTR-R (5'-AACTACTAGGGGCTTGGCTG-3') Chromosome 2: 140,100,939-140,101,567	95°C for 3 mins 30 cycles: 95°C for 30s, 62°C for 15s, 72°C for 30s Final extension: 72°C for 5 mins	Novel primers, no published PCR protocols		
AVPR1a	SNP	AVPR-F (5'-AAGTCGGGAAGGTGAGCTC-3') AVPR-R (5'-CTTCCCGTAGCAAACACAGG-3') Chromosome 11: 62,912,936-62,913,546	95°C for 3 mins 35 cycles: 95°C for 30s, 55°C for 30s, 72°C for 1 min Final extension: 72°C for 5 mins	Novel primers, no published PCR protocols		
OPRM1	SNP	OPRM1_C77G_F (5' - TGGCGCACTCAAGTTGCT-3') OPRM1_C77G_R (5' - GGGACAAGTTGACCCAGGAA-3') Probes: OPRM1_C77G_VIC (5' -CAGCACGCAGCACC-3') labelled with VIC for detecting G allele OPRM1_C77G_FAM (5' -CAGCACCCAGCCC-3') labelled with 6-FAM for detecting C allele Chromosome 4: 60,308,969-60,309,025	95°C for 15mins, 40 cycles: 92°C for 15s, 60°C for 1min	Newly designed Taqman assay		

https://doi.org/10.1371/journal.pone.0288108.t002

Table 3. Overview of variants for 5-HTTLPR, STin, TPH2, HTR2A, MAOA, DRD4, OPRM1, OXTR, and AVPR with frequencies and risk / expression if known.

Gene	Allele_anc / allele_alt	Risk / expression	Associated behaviours from literature		
SLC6A4 5-hydroxytryptamine transporter length-polymorphic	L	High	Anxiety (active), Anxiety (low activity), Distraction, Fear		
repeat (5-HTTLPR)	S	Low	Prosocial		
SLC6A4 STin polymorphism	L	High	Anxiety (active), Anxiety (low activity), Distraction.		
	S	Low			
Tryptophan 5-hydroxylase 2 (TPH2)	L	Low	Aggression, Anxiety (active), Anxiety (low activity).		
	S	High			
5-hydroxytryptamine receptor 2A (HTR2A)	C/G	No polygenic	Anxiety (active), Distraction		
	A/G	data			
	A/C				
Monoamine oxidase A (MAOA)	55	High	Aggression, Anxiety (active), Distraction.		
	56	High			
	57				
	66	High			
	67				
	77	Low			
Dopamine Receptor D4 (DRD4)	A/T	No polygenic	Aggression, Fear		
	C/G	data			
	C/G				
	A/G				
Oxytocin receptor (OXTR)	G/T	No polygenic	Anxiety (active), Anxiety (low activity), Fear, Prosocia		
	C/T	data			
	C/G				
	A/G				
	C/T				
	C/T				
	A/G				
Arginine vasopressin receptor 1A (AVPR1a)	T/C ± insertion	No polygenic data	Anxiety (active), Anxiety (low activity), Fear, Prosocial.		
Opioid receptor mu(μ) 1 (OPRM1)	C/G	No polygenic data	Aggression, Anxiety (active), Anxiety (low activity), Distraction, Fear, Prosocial		

Abbreviations: allele_anc—ancestral allele; allele_alt—alternative allele

https://doi.org/10.1371/journal.pone.0288108.t003

in the preceding week, injury in the preceding 48 hours, cleaning of housing in the last 24 hours and giving birth in the previous 24 hours.

Statistical analysis

Statistical analyses were conducted in R v. 3.4.3v [130]. A maximal model was built per response variable separately. Response variables for social attention were THR, TL; Behaviour: aggression, anxiety, fear, distraction, prosocial. Each genetic variant was entered as a key predictor variable in each model. In addition, we controlled for factors known to influence socioe-motional behaviour: sex, age, and condition [92]. All predictor variables were initially assessed to ensure none were correlated above 0.4 (which could result in collinearity [131]). Response variables were visually inspected for their distribution and transformed to obtain more normal distributions when necessary (this was the case for THR and TL). Appropriate transformations were identified using Tukey's Ladder of Transformation [132] to extract an appropriate

Table 4. Ethogram of behaviours and groups for behavioural observation of captive rhesus macaques (*Macaca mulatta*).

Group	Behaviour	Description			
Aggressive	Aggressive	The animal chases, attacks, threatens, stares at, displaces or lunges towards a conspecific.			
Anxiety (active)	Body shake	Like a dog shake—the animal rapidly moves whole body, usually starting with shaking of the head followed by rest of body.			
	Stereotypic	The behaviour has no obvious function. Includes pacing, bar biting or head tossing.			
	Vigilance	The animal has alert posture scanning their environment or looking at a particular thing (may be out of view).			
Anxiety (low activity)	Self-directed	The animal uses their hands or mouth to clean, scratch or manipulate the skin or fur.			
	Sit hunched	The animal is sitting with their back and head curved round so that the head is below slumped shoulder.			
	Yawn	Animal opens its mouth wide (not directed at conspecific)			
Distraction	Object	The animal uses hands or mouth to investigate and move an inanimate, moveable object in the environment and / or pull or grab parts of the enclosure such as padlocks, sliding adjustable panels and cage dividers.			
	Locomotion	Any behaviour (except those otherwise defined) that involves the animal moving from one location to another, for example, quadrupedal and bipedal walking and running, climbing, descending and jumping. The animal must not be engaged in any other activity.			
Fear	Grimace	The animal's lips are pulled back to expose the teeth.			
	Submissive	Animal moves away, flees, is displaced by or ducks away from a conspecific. They may give out high-pitched screams. Animal may also present its hindquarters in a non-sexual context (not followed by mating).			
Foraging	Foraging	The animal is searching for and / or consuming food or water.			
Inactive	Resting	The animal is lying horizontally with the stomach, back or side touching the floor or the animal us sitting upright			
	Stationary	Weight baring on two or four legs.			
Prosocial	Affiliative	Friendly interaction between the animal and a conspecific includes huddling, being in physical contact with a conspecific and hugging but not grooming behaviour.			
	Allogrooming	Reciprocal grooming. The animal's skin or fur is cleaned, scratched or manipulated by the hands or mouth of a conspecific. The animal uses their hands or mouth to clean, scratch or manipulate the skin or fur of a conspecific.			
	Interaction with baby	The animal interacts with a baby e.g., grooming, playing, carrying or feeding.			
	Lip smack	Animal opens and closes its lips repeatedly without showing its teeth, occasionally making a smacking sound.			
	Sexual behaviour	The animal presents or is presented the hindquarters. The animal is mounted or mounts.			
Other	Other	Any behaviour not otherwise defined.			
	Out of sight	The animal is not visible to the observer.			

https://doi.org/10.1371/journal.pone.0288108.t004

lambda for transformation. Covariates were scaled using a z-transformation to a mean of zero and a standard deviation \pm 1. Scaling continuous variables provides more comparable estimates for interpretation of model output [133]. Participant monkey identity was entered as a random factor in all models.

To account for relatedness between individuals, analyses were conducted using the animal model, in which pedigree was included as a random factor, using the MCMC-based Bayesian GLMM in R [134]. Ignoring family structure can increase type I error (false positive) rate

[135]. Bayesian analysis uses observed or "prior" distribution to estimate parameters of an underlying distribution [136]. For each model, an uninformative prior was specified following de Villemereuil [137]. An uninformative prior was used as the probability distribution was unknown and this type of prior allows for all distributions to be equally likely. Priors are determined based on observed data. As this is a new area of study, there is little prior knowledge that can inform the prior selection.

Each model was plotted and visually inspected to check for convergence. Parameters were set to obtain 1000 iterations, with a burn-in of 1000 and thinning interval of 500 to attain adequate plots, as assessed by eye. Independence of the data points was assessed by using a thinning interval producing autocorrelation of <0.1 assessed using the function autocorr [138]. With these parameters, absence of autocorrelation was assessed as <0.1 for animals at 10000 lag in all cases and the chain was said to have mixed well.

Meaningful effects were evaluated based on whether the 95% credible interval of the estimate overlapped zero [139]. We also provide P values in tables to aid readers in interpretation and for future meta-analyses. The use of Bayesian statistics with uninformative priors automatically controlled for multiplicity, therefore, it was not necessary to use a correction for multiple tests (e.g., Bonferroni correction as used in frequentist approaches [140]).

Results

Genetic and cognitive data were collected from 109 monkeys (94 female). For 95 individuals (81 female), cognitive data were collected at both baseline and following a veterinary stressor. For 14 monkeys (13 female), cognitive data were collected at baseline only. Behavioural data were collected through observation of individuals in the home enclosure for 101 monkeys (93 female). For 77 individuals (69 female) behaviour data were collected at both baseline and following a veterinary stressor. For 12 females, data were collected at baseline only and for 12 females, following the stressor only.

At baseline, 796 trials were collected, and 154 trials were removed from the data set due to disruption, for example, unplanned veterinary visits, cleaning, or births. The cognitive data was then averaged for these trials (2–11 trials; mean = 6) leaving a total of 109 rows of baseline data. A total of 392 trials were collected following the stressor and no trials were removed from the data set. These were averaged for each monkey (1–5 trials; mean = 4) leaving a total of 95 rows of stress data.

Novel haplotypes determined prior to analysis

SNPs for *HTR2A*, *DRD4*, *OXTR* and *AVPR1a* were determined by amplification and sequencing, with haplotypes manually reconstructed from multi-locus genotype data prior to analysis. These novel haplotypes, SNPs and dbSNP numbers are shown in <u>Table 5</u>. This is the first presentation of haplotypes for these polymorphisms in *HTR2A*, *DRD4*, *OXTR* and *AVPR1a*.

Replication of previous evidence for an association of 5-HTTLPR and STin with social attention and behaviour

Of the genes that were analysed with knowledge of which were potentially high and low expressing alleles, *5-HTTLPR* and *STin* were found to be significantly associated with measures of social attention and behaviour. For both *5-HTTLPR* and *STin*, LL was associated with high expression and the s-allele associated with low expression.

The duration of TL and the genotype for *STin* were significantly associated. Macaques with the low expressing alleles had a significantly greater duration of TL (SL, SS: 1.633 ± 0.605 s)

Gene	Haplotype Sequence					
HTR2A	1	CAA				
	2	GGA				
	3	GGC				
	4	GAC				
	dbSNP rs labels: rs8036	dbSNP rs labels: rs80365915, rs80363349, rs196407124				
DRD4	1	AGGG				
	2	TGGG				
	3	ACGG				
	4	TGCA				
	dbSNP rs labels: rs300413141, rs1079355788, rs290724315, rs301203363					
OXTR	1	TTGGCCG				
	2	GCCATTA				
	3	GTGGCCG				
	dbSNP rs labels: rs1967 rs292035217, rs3027892	83445, rs292502465, rs300857875, rs308701533, r68, rs283226059				
AVPR1a	1	T; no insertion (-)				
	2	C; no insertion (-)				
	3	C; with insertion (+)				
	dnSNP rs label: rs19445	55405 plus an 8bp insertion (11:62913090) which we				

Table 5. Sequences for each haplotype for AVPR1a, HTR2A, DRD4 and OXTR.

https://doi.org/10.1371/journal.pone.0288108.t005

than macaques with the high expressing allele (LL; 1.401 ± 0.555 s, 95% CI [-19.049, -1.677]; Table 6).

combined to create three haplotypes

The duration of distraction behaviour was associated with *5-HTTLPR* and, possibly, *STin*. Macaques with the low expressing alleles (SL, SS) for either of these polymorphisms performed more distraction behaviour (*5-HTTLPR*: 23.391 \pm 27.239 s, *STin*: 25.749 \pm 26.370 s) than macaques with the high expressing allele (LL) for *5-HTTLPR* (20.071 \pm 18.296 s, 95% CI [1.466, 14.878]) or *STin* (14.918 \pm 13.574 s, 95% CI [-14.260, 0.306]; Table 6).

Macaques with the high expressing allele for 5-HTTLPR (LL) exhibited more fear behaviour (0.898 \pm 1.250 s) than macaques with the low expressing alleles (SL, SS; 0.308 \pm 0.769, 95% CI [1.603, 9.803]; Table 6). TPH2, MAOA and OPRM1 were not associated with social attention or behaviour.

Table 6. Relationship between high and low expressing alleles for 5-HTTLPR and STin with the mean duration of social attention (n = 109; total duration looking at the threat and neutral faces (TL) and behaviour (n = 90) in rhesus macaques (Macaca mulatta)). Credible intervals not crossing 0 are highlighted in bold.

		HE mean (s)	LE mean (s)	HE df	LE df	Lower CI	Upper CI	P
TL	STin	1.401 ± 0.555	1.633 ± 0.605	45	62	-19.049	-1.677	0.030
Distraction	HTTLPR	20.071 ± 18.296	23.391 ± 27.239	46	42	1.466	14.878	0.022
	STin	14.918 ± 13.574	25.749 ± 26.370	33	55	-14.260	0.306	0.034
Fear	HTTLPR	0.898 ± 1.250	0.308 ± 0.769	46	42	1.603	9.803	0.002

 $Abbreviations: \textbf{HE}-high\ expressing; \textbf{LE}-low\ expressing; \textbf{CI}-credible\ interval$

https://doi.org/10.1371/journal.pone.0288108.t006

New evidence for an association of DRD4 and OXTR with social attention

The durations of the measures of social attention were significantly associated with DRD4 haplotype with individuals homozygous for haplotype 1 (1–1) associated with a short duration of THR and TL (THR: 0.922 ± 0.496 s, TL: 1.598 ± 0.788 s) compared to individuals heterozygous for haplotypes 2–3 (THR: 1.217 ± 0.470 s, 95% CI [0.323, 8.44]; TL: 1.930 ± 0.390 s, 95% CI [0.718, 41.402]) and haplotypes 2–4 (THR: 1.362 ± 0.746 s, 95% CI [2.001, 11.565]; TL: 2.068 ± 0.645 s, 95% CI [4.690, 54.698]; Table 7). The duration of TL was also associated with OXTR haplotype. Individuals heterozygous for haplotypes 1–3 had a significantly longer duration of TL (1.015 ± 0.836) compared to 2–3 individuals (0.766 ± 0.515 s, 95% CI [0.569, 39.556]). None of the polymorphisms for HTR2A or AVPR1a were associated with differences in the duration of THR or TL.

Table 7. Relationship between the new polymorphisms with no previous published data expression or behaviour data (*HTR2A*, *DRD4*, *OXTR*, *AVPR1a*) and the mean duration of social attention (n = 109; duration looking at the threat face (THR), and total duration looking at the threat and neutral faces (TL)) and behaviour (n = 90) in rhesus macaques (*Macaca mulatta*). Credible intervals not crossing 0 are highlighted in bold. Reference variant with most negative test value and a sample size greater than 8 were selected to aid interpretation for each gene.

	Gene (ref variant)	Haplotype	Post mean	n	Lower CI	Upper CI	P
THR	DRD4 (1-1; n = 27)	1-2	0.631	42	-1.198	2.638	0.498
		1-3	2.020	3	-2.5090	6.985	0.398
		1-4	0.738	3	-4.041	5.573	0.746
		2-2	0.344	24	-2.141	2.434	0.748
		2-3	4.400	5	0.323	8.444	0.030
		2-4	6.882	3	2.001	11.565	0.008
TL	DRD4 (1-1; n = 27)	1-2	5.590	42	-4.222	14.952	0.240
		1-3	14.356	3	-8.756	39.908	0.230
		1-4	-3.479	3	-27.101	19.424	0.778
		2–2	2.890	24	-9.164	14.334	0.604
		2-3	22.173	5	0.718	41.402	0.046
		2-4	29.504	3	4.690	54.698	0.022
	OXTR (2-3; n = 10)	1-1	6.573	14	-10.371	26.025	0.492
		1-2	0.905	51	-15.824	15.794	0.892
		1-3	21.298	9	0.569	39.556	0.020
		2-2	3.356	25	-13.657	20.432	0.688
Anxiety (active)	AVPR1a (BC; n = 14)	AB	39.857	21	-80.306	163.781	0.514
		AC	278.582	2	24.414	514.356	0.018
		BB	46.596	52	-52.252	149.498	0.388
Fear	HTR2A (1.1; n = 25)	1.2	-0.864	33	-6.34	3.452	0.730
		1.4	-3.062	6	-11.812	5.503	0.482
		2.2	6.154	14	0.345	12.539	0.048
		2.4	6.600	8	-1.317	14.853	0.110
		3.4	13.606	2	0.495	26.851	0.042
	OXTR (2.3; n = 8)	1.1	9.455	10	1.058	19.083	0.036
		1.2	6.566	43	-0.672	13.884	0.074
		1.3	9.7805	7	0.8113	19.7451	0.052
		2.2	8.4165	22	-8.0152	0.8785	0.832

https://doi.org/10.1371/journal.pone.0288108.t007

New evidence for an association of HTR2A, OXTR and AVPR1a with fearful and anxious behaviour

The duration of fear behaviour was significantly associated with HTR2A haplotype. 1–1 individuals performed fear behaviour for a significantly shorter duration (0.394 \pm 1.015 s) compared to individuals homozygous for haplotype 2 (0.751 \pm 1.069 s, 95% CI [0.345, 12.539]) and heterozygous for haplotypes 3–4 (1.968 \pm 2.178 s, 95% CI [0.495, 26.851]).

The duration of fear behaviour was associated with the *OXTR* haplotype. Individuals homozygous for haplotype 1 performed a significantly greater duration of fear behaviour $(0.920 \pm 1.127 \text{ s})$ than individuals heterozygous for haplotypes 2–3 $(0.168 \pm 0.315 \text{ s}, 95\% \text{ CI} [1.058, 19.083])$. 1–3 individuals also performed a greater duration of fear behaviour $(1.335 \pm 1.514 \text{ s})$ than 2–3 individuals and this difference approached significance $(0.168 \pm 0.315 \text{ s}, 95\% \text{ CI} [0.811, 19.745])$.

The duration of active anxiety behaviour was associated with AVPR1a genotype. AC individuals spent significantly longer performing active anxiety behaviour (165.091 \pm 77.624 s) compared to BC individuals (68.003 \pm 71.557 s, 95% CI [24.414, 514.356]). None of the haplotypes were significantly associated with aggressive, low activity anxiety, distraction, prosocial behaviour, or inactive behaviour.

Discussion

We assessed the relationship between eight genes (nine loci) involved in emotion with two measures of social attention (the total time spent looking at the threat-neutral face pair overall per trial (TL) and the time spent looking at the threat face per three second trial (THR)), and eight behavioural categories in a relatively large sample of 109 rhesus macaques. Our results support previous findings that the 5-HTT length-polymorphic repeat in *SLC6A4* (5-HTTLPR) and intron 2 of the *serotonin transporter* (*STin*) are associated with distraction and fear behaviors. Additionally, we demonstrate for the first time that *STin* is also associated with social attention. We found an association between four novel polymorphisms in 5-hydroxytryptamine receptor 2A (HTR2A), Dopamine Receptor D4 (DRD4), Oxytocin receptor (OXTR) and Arginine vasopressin receptor 1A (AVPR1a) with measures of social attention and behaviour which we recommend for further investigation.

Social attention

Three polymorphisms in candidate genes were found to be associated with measures of social attention. TL was associated with the genotype for the 16–17 bp VNTR polymorphism in *STin* and the haplotypes for both *DRD4* and *OXTR*. THR was also associated with the haplotype for *DRD4*. The influence of these variants on social attention and behaviour has not previously been studied in primates and, only the *STin* polymorphism has been explored in humans. No studies have explicitly looked at the link between attention bias (AB) and *STin*; however, the results of the present study and the available human literature suggest that the high expressing *STin* allele may be associated with increased anxiety and attention to social stimuli in both macaques and humans.

In macaques, the high expressing allele for *STin* (LL) was associated with a significantly shorter duration of TL than macaques with the low expressing allele (SL), i.e., those with the high expressing allele spent less time looking at social stimuli. This may suggest that individuals with the high expressing allele could be more susceptible to stress than individuals with the low expressing allele as previous cognitive studies have demonstrated that macaques become more avoidant of negative stimuli following a stressor [91].

Macaques and humans have different patterns of response to threat in cognitive trials using face stimuli: macaques showi greater avoidance of threat while anxious or stressed, compared to humans [141]. In humans, the high expressing *STin* allele (*STin*2.12) is associated with emotional disorders including anxiety, OCD, and depression [34, 35, 142]. OCD shares the underlying anxiety psychopathology with other disorders such as post-traumatic stress disorder and depression [143]. Cognitive factors implicated in the maintenance and development of OCD include the overestimation of threat and AB towards threatening information [144, 145]. Human participants with OCD struggle to disengage with disgust faces during cognitive trails of social attention [146, 147]. When shown threatening social stimuli, OCD patients spend significantly longer looking at the faces compared to healthy controls [147]. OCD is also associated with lower AB variability [148] and individuals with OCD showed lower within-individual variability of cognitive measure of social attention [149].

The polymorphisms for *DRD4* and *OXTR* presented here have not previously been studied in macaques or humans. Here, we found an influence of haplotype for both polymorphisms on social attention I macaques. *DRD4* individuals homozygous for haplotype 1 (1–1), were more avoidant of social stimuli compared to individuals heterozygous for haplotypes 2–3 or 2–4. For *OXTR*, individuals heterozygous for haplotypes 2–3 were more avoidant of social stimuli than those heterozygous for haplotypes 1–3. This suggests that macaques with *DRD4* haplotype 1 or *OXTR* haplotypes 2–3 may be more vulnerable to stress than macaques with other haplotypes for these genes. Other polymorphisms within these genes have been associated with a range of conditions (e.g., *DRD4*: anger, aggression, and delinquency [64, 65]; *OXTR*: reduced positivity [70], and poor social recognition skills [71]) and the results demonstrated here suggest that these novel polymorphisms could be key for future candidate gene studies for cognitive measures of social attention in rhesus macaques.

Further research to increase the sample size, and therefore the statistical power [150, 151] would be beneficial, especially for confirming the effects of the rarer haplotypes. However, the data presented here were from 109 macaques and the sample size was an improvement on other published rhesus macaque studies (e.g., n = 20 [152], n = 20 [153], n = 9 [154], n = 7 [155], n = 29 [28]).

Behaviour

Six polymorphisms in candidate genes were found to be associated with differences in behaviour. Genotypes for both polymorphisms within the serotonin transporter (5-HTTLPR and STin) were associated with distraction behaviour. The duration of fear behaviour was associated with 5-HTTLPR genotype and haplotype for HTR2A and OXTR. AVPR1a haplotype was associated with the duration of active anxiety behaviour. Apart from 5-HTTLPR and STin, these variants have not previously been studied in macaques or humans. Our data suggest they are worth further investigation.

The low expressing alleles for both 5-HTTLPR and STin were associated with a longer duration of distraction behaviour. S-allele carriers spent longer engaged in distraction behaviour compared to l-allele homozygotes for both 5-HTTLPR and STin. In both humans and macaques, the s-allele for 5-HTTLPR has a lower rate of serotonin transporter transcriptional efficiency [21, 22, 24]. Juvenile rhesus macaques homozygous for the s-allele displayed more anxious and threatened behaviour than l-allele carriers during novel fruit, human intruder, remote-controlled car, and free play tests [24].

Here, distraction behaviour included locomotion and manipulation of objects. Both behaviours have previously been identified as self-soothing responses to stress in primates. For example, juvenile macaques show an increase in locomotion following maternal separation

[156, 157], agitated locomotion increases following agonistic interactions with macaque conspecifics [158] and, orangutans (*Pongo pygmaeus*), frustrated during delayed computer trials, displayed more object manipulation behaviour than those not subject to the delayed trials [159]. However, it is important to note that both behaviours could be categorised as "exploratory" and classed as positive or neutral responses. For example, reduced locomotor activity could indicate withdrawal [160] and object manipulation is often considered a natural behaviour associated with tool use [161]. The difference in interpretation of locomotor and manipulation behaviours means that there is disagreement between the Bethea et al [24] study and the data presented here on the impact of 5-HTTLPR genotype on affective state.

Bethea et al [24] reported that genotype for 5-HTTLPR had a significant association with activity during a free play test. S-allele homozygotes were significantly less active than l-allele carriers. This reduced activity was measured using percentage of time engaged in locomotor behaviour and percentage of time spent playing with toys, the opposite of the results presented in the present study. Previous studies have also revealed that macaques with the lower expressing alleles have a heightened hormonal response to stress. In response to maternal separation, juvenile macaques heterozygous for the l-allele had lower adrenocorticotropic hormone (ACTH) levels than s/l heterozygotes [23, 153]. ACTH stimulates the adrenal cortices of the adrenal glands to release mineralocorticoid and glucocorticoid hormones, including cortisol [162] leading to higher cortisol responses to stress in s-allele homozygotes [28]. Fear behaviour is also influenced by 5-HTTLPR genotype. In the present study, the high expressing allele (LL) for 5-HTTLPR was associated with a longer duration of fear behaviour while Bethea et al [24] revealed s-allele homozygotes to engage in fear behaviour more frequently than l-allele carriers.

Much of the previous literature focuses on the interaction between 5-HTTLPR genotype and response to a specific stressor, for example, separation [23, 153] or consecutive fear and anxiety eliciting tests [24]. Studies that applied no specific acute stressor showed no behavioural differences associated with genotype for 5-HTTLPR [28]. In the present study, no specific fear or anxiety inducing tests were conducted. This project piggybacked onto the macaques' routine annual veterinary health check to ensure further stress was not caused as a result of this research. Data were collected before and after the health check and the procedures involved (e.g., ketamine sedation) have been shown to acutely compromise macaque welfare [91, 163–165]. Here we found no effect of the veterinary intervention on behaviour or measures of social attention. Refinements in veterinary and scientific practice and techniques involved in the veterinary health check [166] may mean that they are no longer sufficiently stressful to compromise welfare. In addition, all macaques were familiar with the veterinary intervention; novelty is known to be stressful for primates [167], therefore, this reduced novelty is likely to also have decreased the negative impact on their welfare and dampened the behavioural response.

The age and rearing environment difference between the studies may also be a contributing factor. Much of the previously published macaque literature focused on infant and juvenile macaques (\leq 6 months [23, 153]; \leq 1 year [24]) that experienced early life stress (i.e., early weaning). The typical age at which juvenile rhesus macaques stop suckling and are nutritionally weaned is between 10 and 14 months old [168–170]. Early weaning of captive macaques is defined as removal from the mother before 12 months and can be a source of considerable early life stress [171]. Peer-reared juvenile s/l heterozygote macaques, separated from their mothers at birth, had significantly heightened hormonal responses to separation stress compared to mother reared macaques of any 5-HTTLPR genotype [23]. Peer-reared juvenile s/l heterozygote macaques, weaned at 30 days, also had significantly lower levels of 5-hydroxyindoleacetic acid in the cerebrospinal fluid (CSF 5-HIAA) compared to peer reared l-

homozygotes [124]. In these studies, the mother reared infants were also weaned before the recommended age (six months [23], seven months [124]).

Mother abused infant macaques with the 5-HTTLPR s/l genotype had heighted anxiety and hormonal response to stress compared to infants that did not experience abuse, while s-homozygotes showed more behavioural reactivity (resistance to handling, tantrums) than l-homozygotes [153]. In this study, l/l homozygotes appeared to be buffered from the physiological effects of mother abuse and had lower levels of cortisol than non-abused infants of the other genotypes. The authors suggested that l-homozygotes are better able to adapt to initially heightened cortisol levels caused by their mothers' abuse.

The macaques involved in the present study were housed at the MRC-CFM. MRC-CFM has been involved in several research projects focusing on weaning including Prescott et al [171] and as such macaques are not weaned before 10 to 14 months [172]. As a result, only a small number of the older macaques had been weaned early and only two were homozygous for the 5-HTTLPR s-allele. Early weaning was not included in the analysis here due to low sample size resulting from improved management practices and, therefore, the impact of the interaction between early life stress and the low expressing genotype cannot be explored in this data set.

Fear behaviour was associated with haplotype for HTR2A and OXTR. HTR2A encodes the serotonin 2A receptor which plays an important role in the modulation of mood and anxiety [48, 173, 174]. Polymorphisms in HTR2A have been associated with neuropsychiatric disorders including impulsive behaviour and schizophrenia in humans [49, 50]. In mice ($Mus\ sp$.), HTR2A-expressing cells in the central amygdala control innate and learned freezing [175]. Freezing is a characteristic fear response in mice [176]. Inactivation of the HTR2A-expressing cells downregulated learned-freezing response and upregulated innate freezing response. In marmosets ($Callithrix\ jacchus$), mRNA expression levels of HTR2A were shown to have a selective-relationship between trait-like anxiety suggesting that the serotonin receptor may only have a limited role in regulating serotonin signaling in the downstream pathway [177]. The authors pointed to potential sample size issues (n = 13); highlighting why further studies on polymorphisms within HTR2A are important for determining the role of this gene within anxiety and fear behaviours.

OXTR encodes G-protein coupled receptor proteins for oxytocin. Oxytocin is a neuromodulatory hormone that plays a counter-regulatory role in stress and fear responses [67, 68]. Other polymorphisms within OXTR have been shown to be associated with differences in oxytocin receptor density in the brain and Depression Anxiety and Stress Scale (DASS) scores in humans [178]. No OXTR polymorphisms have previously been studied in macaques and therefore, the polymorphisms presented here represent important variants for future study.

AVPR1a haplotype was associated with the duration of active anxiety behaviour. The polymorphisms investigated here were RS3 alleles which, in humans, are associated with altruism [75], pair-bonding behaviour [76], social behaviour including sibling conflict [77] and reproductive behaviour [78]. In humans, the high-risk genotype is 327 bp and the low risk in non-327 [179] with the 327 bp allele being associated with lower altruism and non-optimal mothering [180]. Other alleles, for example, the 334 bp allele was associated with relationship and marital problems [79]. These human variants do not exactly map onto the macaque polymorphisms studied here but point to AVPR1a haplotype influencing general and social anxiety in primates.

Conclusion & future applications

We analysed eight genes for variation at a total of nine loci with cognitive and behavioural correlates in rhesus macaques. Of these, five had not previously been studied in primates and four

had not previously been studied in humans. Our results revealed a significant association between *STin* genotype, *DRD4* haplotype and *OXTR* haplotype and cognitive measures of social attention in rhesus macaques. Genotype for *5-HTTLPR*, *STin* and *AVPR1a* and haplotype for *HTR2A* and *OXTR* were significantly associated with the duration of behaviours including fear and anxiety. Due to the novelty of these findings, we recommend future work focus particularly on the association between *HTR2A*, *DRD4*, *OXTR* and *AVPR1a* with cognitive and observational measures of behaviour associated with wellbeing and survival in both macaques other species including humans. Understanding the biological underpinnings of individual variation in negative emotion (e.g., fear and anxiety), together with their impact on social behaviour (e.g., social attention including vigilance for threat) has application for managing primate populations in the wild and captivity, and has translational value for understanding the evolution and function of negative emotion in our own species.

Supporting information

S1 Table. Final dataset of cognitive and behaviour data with genotypes from nine loci for each rhesus macaque (*Macaca mulatta*). (XLSX)

Acknowledgments

We thank the staff at MRC-CFM, particularly Faye Peters and Sebastian Merritt who assisted with data collection.

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References

- Verhulst EV, Mateman AC, Zwier MV, Caro SP, Verhoeven KJF, van Oers K. Evidence from pyrosequencing indicates that natural variation in animal personality is associated with *DRD4* DNA methylation. Mol Ecol. 2016; 25(8): 1801–1811.
- 2. Long T, Yuan M, Yuan H, Lu W. Heritability of animal individuality in fish: Distribution, behavior, metabolism and stress response. Aquaculture. 2021; 536: 736415.
- Persson ME, Roth LSV, Johnsson M, Wright D, Jensen P. Human-directed social behaviour in dogs shows significant heritability. Genes Brain Behav. 2015; 14(4): 337–344. https://doi.org/10.1111/gbb.12194 PMID: 25703740
- Wolc A, Settar P, Fulton JE, Arango J, Rowland K, Lubritz D, et al. Heritability of perching behavior and its genetic relationship with incidence of floor eggs in Rhode Island Red chickens. Genet Sel Evol. 2021; 53: 1–9.

- Insel T, Cuthbert, Garvey M, Heinssen R, Pine DS, Quinn K, et al. Research domain criteria (RDoC): Toward a new classification framework for research on mental disorders. Am J Psychiatry. 2010; 167: 748–751. https://doi.org/10.1176/appi.ajp.2010.09091379 PMID: 20595427
- Clinton SM, Shupe EA, Glover ME, Unroe KA, McCoy CR, Cohen JL, et al. Modeling heritability of temperamental differences, stress reactivity, and risk for anxiety and depression: Relevance to research domain criteria (RDoC). Eur J Neurosci. 2021; 55(9–10): 2076–2107. https://doi.org/10.1111/ejn.15158 PMID: 33629390
- Rosenberg NA, Edge MD, Pritchard JK, Feldman MW. Interpreting polygenic scores, polygenic adaptation, and human phenotypic differences. Evo Med Public Health, 2019; 2019(1): 26–34. https://doi.org/10.1093/emph/eoy036 PMID: 30838127
- Flint J. Analysis of quantitative trait loci that influence animal behaviour. J Neurobiol. 2003; 54(1): 46– 77
- Turri MG, Datta SR, DeFries J, Henderson ND, Flint J. QTL analysis identifies multiple behavioural dimensions in ethological tests of anxiety in laboratory mice. Curr Biol. 2001; 11(10): 725–734.
- Fernandez-Teruel A, Escorihuela RM, Gray JA, Aguilar R, Gil L, Gimenez-Llort L, et al. A quantitative trait locus influencing anxiety in the laboratory rat. Genome Res. 2002; 12(4): 618–626. https://doi.org/ 10.1101/gr.203402 PMID: 11932246
- Edwards AC, Mackay TFC. Quantitative trait loci for aggressive behaviour in *Drosophila melanoga-ster*. Genetics. 2009; 182(3): 889–897.
- Strawbridge RJ, Ward J, Cullen B, Tunbridge EM, Hartz S, Bierut L, et al. Genome-wide analysis of self-reported risk-taking behaviour and cross-disorder genetic correlations in the UK Biobank cohort. Transl Psychiatry. 2018; 8: 39. https://doi.org/10.1038/s41398-017-0079-1 PMID: 29391395
- Tielbeek JJ, Johansson A, Polderman TJC, Rautianinen M-R, Jansen P, Taylor M, et al. Genomewide association studies of a broad spectrum of antisocial behaviour. JAMA Psychiatry. 2018; 74(12): 1242–1250.
- Zvrskovec J, Breen G, Eley T, Iniesta R, ter Kuile A. Tu38. Joint latent factor GWA study using highdefinition LD scores outlines the specific genetic architecture of anxiety and mood disorders. Eur Neuropsychopharmacol. 2021; 51: e115.
- Rogers J. Genomic resources for rhesus macaques (Macaca mulatta). Mamm Genome. 2022; 33: 91–99
- Flint J, Munafo MR. Candidate and non-candidate genes in behavior genetics. Curr Opin Neurobiol. 2013; 23(1): 57–61. https://doi.org/10.1016/j.conb.2012.07.005 PMID: 22878161
- Warren WC, Harris RA, Haukness M, Fiddes IT, Murali SC, Fernandes J, et al. Sequence diversity analyses of an improved rhesus macaque genome enhance its biomedical utility. Science. 2020; 370 (6523): 6617. https://doi.org/10.1126/science.abc6617 PMID: 33335035
- Houwing DJ, Buwalda B, van der Zee EA, de Boer SF, Oliver JDA. The serotonin transporter and early life stress: translational perspectives. Frontiers in Cellular Neuroscience. 2017; 11: 117. https://doi.org/10.3389/fncel.2017.00117 PMID: 28491024
- de Lima KW, Guembarovski RL, Oda JM, Ramos G, Oliveira BV, Cavalli IJ, et al. Association between the STin2 VNTR polymorphism and smoking behavior in oral cancer patients and healthy individuals. Int J of Clin Exp Med. 2012; 12(1): 13–19. https://doi.org/10.1007/s10238-011-0140-y PMID: 21626393
- 20. de Castro MRP, Maes M, Guembarovski RL, Ariza CB, Reiche EDV, Vargas HO, et al. SLC6A4 STin2 VNTR genetic polymorphism is associated with tobacco use disorder, but not with successful smoking cessation or smoking characteristics: a case control study. BMC Genet. 2014; 15: 78.
- Peterson IT, Bates JE, Goodnight JA, Dodge KA, Lansford JE, Pettit GS, et al. Interaction between serotonin transporter polymorphism (5-HTTLPR) and stressful life events in adolescents' trajectories of anxious/depressed symptoms. Dev Psychol. 2012; 48(5): 1463–1475. https://doi.org/10.1037/ a0027471 PMID: 22390669
- Wankerl M, Miller R, Kirschbaum C, Hennig J, Stalder T, Alexander N. Effects of genetic and early environmental risk factors for depression on serotonin transporter expression and methylation profiles. Transl Psychiatry. 2014; 4: e402. https://doi.org/10.1038/tp.2014.37 PMID: 24937096
- Barr CS, Newman TK, Shannon C, Parker C, Dvoskin RL, Becker ML, et al. Rearing condition and rh5-HTTLPR interact to influence limbic-hypothalamic-pituitary-adrenal axis response to stress in infant macaques. Biol Psychiatry. 2004; 55: 733–738. https://doi.org/10.1016/j.biopsych.2003.12.008 PMID: 15039002
- Bethea CL, Stricher JM, Coleman K, Pau FKY, Moessner R, Cameron JL. Anxious behaviour and fenfluramine-induced prolactin secretion in young rhesus macaques with different alleles of the serotonin reuptake transporter polymorphism (5-HTTLPR). Behav Genet. 2004; 34(3): 295–307.

- Caspi A, Sugden K, Moffitt TE, Tayor A, Craig IW, Harrington H, et al. Influence of life stress on depression: moderation by a polymorphism in the 5-HTT gene. Science. 2003; 301(5631): 386–389. https://doi.org/10.1126/science.1083968 PMID: 12869766
- Lesch KP, Bengel D, Heils A, Sabol SZ, Greenberg BD, Petri S, et al. Association of anxiety-related traits with a polymorphism in the serotonin transporter gene regulatory region. Science. 1996; 274 (5292): 1527–1531. https://doi.org/10.1126/science.274.5292.1527 PMID: 8929413
- Goenjian AK, Bailey JN, Walling DP, Steinberg AM, Schmidt D, Dandekar U, et al. Association of TPH1, TPH2, and 5HTTLPR with PTSD and depressive symptoms. J Affect Disord. 2012; 140(3): 244–252.
- Qin D, Rizak J, Feng X, Yang S, Yang L, Fan X, et al. Cortisol responses to chronic stress in adult macaques: Moderation by a polymorphism in the serotonin transporter gene. Behav Brain Res. 2015; 278: 280–285. https://doi.org/10.1016/j.bbr.2014.10.001 PMID: 25311283
- Shively CA, Register TC, Friedman DP, Morgan TM, Thompson J, Lanier T. Social stress-associated depression in adult female cynomolgus macaques (*Macaca fascicularis*). Biol Psychol. 2005; 69(1): 67–84
- Qin DD, Rizak J, Feng XL, Yang SC, Lü LB, Pan L, et al. Prolonged secretion of cortisol as a possible mechanism underlying stress and depressive behaviour. Sci Rep. 2016; 6: 30187. https://doi.org/10. 1038/srep30187 PMID: 27443987
- Yilmaz M, Erdal ME, Herken H, Cataloluk O, Barlas O, Bayazit YA. Significance of serotonin transporter gene polymorphism in migraine. J of Neurol Sci. 2001; 186(1–2): 27–30. https://doi.org/10.1016/s0022-510x(01)00491-9 PMID: 11412868
- Inoue-Murayama M, Hibino E, Iwatsuki H, Inoue E, Hong KW, Nishida T, et al. Interspecies and intraspecies variations in the serotonin transporter gene intron 3 VNTR in nonhuman primates. *Primates*. 2008; 49: 139–142. https://doi.org/10.1007/s10329-007-0077-7 PMID: 18204817
- Joshi G, Pradhan S, Mittal B. No direct association of serotonin transporter (STin2 VNTR) and receptor (HT 102T>C) gene variants in genetic susceptibility to migraine. Dis Markers. 2010; 29(5): 223–229.
- 34. Fiskerstrand CE, Lovejoy EA, Quinn JP. An intronic polymorphic domain often associated with susceptibility to affective disorders has allele dependent differential enhancer activity in embryonic stem cells. FEBS Lett. 1999; 458: 171–174. https://doi.org/10.1016/s0014-5793(99)01150-3 PMID: 10481059
- 35. Baca-Garcia E, Vaquero-Lorenzo C, Diaz-Hernandez M, Rodrigues-Salgado B, Doolengevich-Segal H, Arrojo-Romero M, et al. Association between obsessive-compulsive disorder and a variable number of tandem repeats polymorphism in intron 2 of the serotonin transporter gene. Prog Neuropsychopharmacol Biol Psychiatry. 2007; 31(2): 416–420. https://doi.org/10.1016/j.pnpbp.2006.10.016 PMID: 17174018
- 36. Lovejoy EA, Scott AC, Fiskerstrand CE, Bubb VJ, Quinn JP. The serotonin transporter intronic VNTR enhancer correlated with a predisposition to affective disorders has distinct regulatory elements within the domain based on the primary DNA sequence of the repeat unit. Eur J Neurosci. 2003; 17: 417–420. https://doi.org/10.1046/j.1460-9568.2003.02446.x PMID: 12542679
- Haddley K, Bubb VJ, Breen G, Parades-Esquivel UM, Quinn JP. Behavioural genetics of the serotonin transporter. Curr Top Behav Neurosci. 2012; 12: 503–535. https://doi.org/10.1007/7854_2011_186
 PMID: 22261701
- Kaiser R, Tremblay PB, Schmider J, Henneken M, Dettling M, Muller-Oerlinghausen B, et al. Serotonin transporter polymorphisms: no association with response to antipsychotic treatment, but associations with the schizoparanoid and residual subtypes of schizophrenia. Mol Psychiatry. 2001; 6(2): 179–185. https://doi.org/10.1038/sj.mp.4000821 PMID: 11317220
- **39.** Ogilvie AD, Battersby S, Budd VJ, Fink G, Harmer AJ, Goodwin GM, et al. Polymorphism in serotonin transporter gene associated with susceptibility to major depression. Lancet. 1996; 347: 731–733. https://doi.org/10.1016/s0140-6736(96)90079-3 PMID: 8602004
- **40.** Saha S, Newmei MK, Pasi S, Singh PK, Rao V. Stress and 5-HTT (SLC6A4) as an indicator of suicidal behavior: a population study among the Dubla tribe of Daman. Mol Cytogenet. 2014; 7(S1): 44.
- Pavlov KA, Chistiakov DA, Chekhonin VP. Genetic determinants of aggression and impulsivity in humans. J Appl Genet, 2012; 53(1): 61–82. https://doi.org/10.1007/s13353-011-0069-6 PMID: 21994088
- Walther DJ, Peter JU, Bashammakh S, Hörtnagl H, Voits M, Fink H, et al. Synthesis of serotonin by a second tryptophan hydroxylase isoform. Science. 2003; 299(5603): 76. https://doi.org/10.1126/ science.1078197 PMID: 12511643
- **43.** Chen GL, Novak MA, Hakim S, Xie Z, Miller GM. Tryptophan hydroxylase-2 gene polymorphisms in rhesus monkeys: association with hypothalamic–pituitary–adrenal axis function and *in vitro* gene expression. Mol Psychiatry. 2006; 11(10): 914–928.

- Walitza S, Renner TJ, Dempfle A, Konrad K, Wewetzer CH, Halbach A, et al. Transmission disequilibrium of polymorphic variants in the tryptophan hydroxylase-2 gene in attention-deficit/hyperactivity disorder. Mol Psychiatry. 2005; 10(12): 1126-1132. https://doi.org/10.1038/sj.mp.4001734 PMID: 16116490
- Lopez VA, Detera-Wadleigh S, Cardona I, Kassem L, McMahon FJ. Nested association between genetic variation in tryptophan hydroxylase II, bipolar affective disorder, and suicide attempts. Biol Psychiatry. 2007; 61(2): 181-186. https://doi.org/10.1016/j.biopsych.2006.03.028 PMID: 16806105
- Zill P, Büttner A, Eisenmenger W, Möller HJ, Bondy B, Ackenheil M. Single nucleotide polymorphism and haplotype analysis of a novel tryptophan hydroxylase isoform (TPH2) gene in suicide victims. Biol Psychiatry. 2004; 56(8): 581-586. https://doi.org/10.1016/j.biopsych.2004.07.015 PMID: 15476687
- Chen GL, Novak MA, Meyer JS, Kelly BJ, Vallender EJ, Miller GM. The effect of rearing experience and TPH2 genotype on HPA axis function and aggression in rhesus monkeys: A retrospective analysis. Horm Behav. 2010; 57(2): 184-191.
- D'Souza UM, Craig IW. Genetic organization of the serotonergic system. In: Muller CP, Jacobs BL, editors. Handbook of Behavioural Neuroscience (Vol. 21). London: Elsevier; 2010. pp. 23-50.
- Nomura M. Nomura Y. Psychological, neuroimaging and biochemical studies on functional association between impulsive behaviour and 5-HT2A receptor gene polymorphisms in humans. Ann N Y Acad Sci. 2006; 1086: 132-143.
- D'Souza UM. Craig IW. Functional genetic polymorphisms in serotonin and dopamine gene systems and their significance in behavioural disorders. Prog Brain Res. 2008; 172: 73–98. https://doi.org/10. 1016/S0079-6123(08)00904-7 PMID: 18772028
- Smith RM, Papp AC, Webb A, Ruble CL, Munsie LM, Nisenbaum LK, et al. Multiple regulatory variants modulate expression of 5-hydroxytryptamine 2A receptors in human cortex. Biol Psychiatry. 2013; 73 (6): 546-554. https://doi.org/10.1016/j.biopsych.2012.09.028 PMID: 23158458
- Mendelson SD. Psychiatric medications and metabolic syndrome. In: Metabolic Syndrome and Psychiatric Illness. Burlington: Academic Press; 2008. pp. 73-91.
- Goldman D, Landi P, Ducci F. Addictive disorders. In: Rimoin D, Pyeritz R, Korf B, editors. Emery and Rimoin's Principles and Practice of Medical Genetics. 6th ed. London: Academic Press; 2013.
- 54. Newman TK, Syagailo YV, Barr CS, Wendland JR, Champoux M, Graessle M, et al. Monoamine oxidase A gene promoter variation and rearing experience influences aggressive behavior in rhesus monkeys. Biol Psychiatry. 2005; 57(2): 167-172. https://doi.org/10.1016/j.biopsych.2004.10.012 PMID: 15652876
- Kinnally EL, Karere GM, Lyons LA, Mendoza SP, Mason WA, Capitanio JP. Serotonin pathway genegene and gene-environment interaction influence behavioural stress response in infant rhesus macagues. Dev Psychopathol. 2010; 22(1): 35-44.
- 56 Girault JA, Greengard P. The neurobiology of dopamine signaling. JAMA Neurol. 2004; 61(5): 641-644. https://doi.org/10.1001/archneur.61.5.641 PMID: 15148138
- 57. Knab AM, Lightfoot JT. Does the difference between physically active and couch potato lie in the dopamine system? Int J Biol Sci. 2010; 6(2): 133-150. https://doi.org/10.7150/ijbs.6.133 PMID: 20224735
- 58. Stott SRW, Ang SL. The generation of midbrain dopaminergic neurons. In: Rubenstein JLR, Rakic P, editors. Patterning and Cell Type Specification in the Developing CNS and PNS. London: Academic Press; 2013. pp. 435-453.
- Asghari V, Sanyal S, Buchwaldt S, Paterson A, Jovanovic V, Von Tol HHM. Modulation of intracellular cyclic AMP levels by different human dopamine D4 receptor variants. J Neurochem. 1994; 65(3): 1157-1165.
- Kazmi MA, Snyder LA, Cypess AM, Graber SG, Sakmar TP. Selective reconstitution of human D4 dopamine receptor variants with Gia subtypes. Biochem. 2000; 39(13): 3734-3744.
- Oak JN, Oldenhof J, Van Tol HHM. The dopamine D4 receptor: one decade of research. Eur J of Pharmacol. 2000; 405(1-3): 303-327. https://doi.org/10.1016/s0014-2999(00)00562-8 PMID: 11033337
- Kaplan AS, Levitan RD, Yilmaz Z, David C, Tharmalingham S, Kennedy JL. A DRD4/BDNF gene-62. gene interaction associated with maximum BMI in women with bulimia nervosa. Int J Eat Disord. 2007; 41(1): 22-28.
- Chen D, Liu F, Shang Q, Song X, Miao X, Wang Z. Association between polymorphisms of DRD2 and DRD4 and opioid dependence: Evidence from the current studies. Am J Med Genet B Neuropsychiatr Genet. 2011; 156(6): 661-670.
- Hohmann S, Becker K, Fellinger J, Banaschewski T, Schmidt MH, Esser G, et al. Evidence for epistasis between the 5-HTTLPR and the dopamine D4 receptor polymorphisms in externalizing behavior among 15-year-olds. J Neural Transm. 2009; 116(12): 1621-1629. https://doi.org/10.1007/s00702-009-0290-1 PMID: 19696961

- Dmitrieva J, Chen C, Greenberger E, Ogunseitan O, Ding YC. Gender-specific expression of the DRD4 gene on adolescent delinquency, anger and thrill seeking. Soc Cogn Affect Neurosci. 2011; 6 (1): 82–89. https://doi.org/10.1093/scan/nsq020 PMID: 20203140
- 66. Coyne SP, Lindell SG, Clemente J, Barr CS, Parker KJ, Maestripieri D. Dopamine D4 receptor genotype variation in free-ranging rhesus macaques and its association with juvenile behavior. Behav Brain Res. 2015; 292: 50–55. https://doi.org/10.1016/j.bbr.2015.06.014 PMID: 26073765
- Sawchenko PE, Swanson LW. Immunohistochemical identification of neurons in the paraventricular nucleus of the hypothalamus that project to the medulla or to the spinal cord in the rat. J Comp Neurol. 1982; 205: 260–272. https://doi.org/10.1002/cne.902050306 PMID: 6122696
- Dreifuss JJ, Dubois-Dauphin M, Widmer H, Raggenbass M. Electrophysiology of oxytocin actions on central neurons. Ann N Y Acad Sci. 1992; 652: 46–57. https://doi.org/10.1111/j.1749-6632.1992. tb34345.x PMID: 1320838
- 69. Feldman R, Monakakhov M, Pratt M, Ebstein RP. Oxytocin pathway genes: Evolutionary ancient system impacting on human affiliation, sociality, and psychopathology. Biol Psychiatry. 2016; 79(3): 174–184. https://doi.org/10.1016/j.biopsych.2015.08.008 PMID: 26392129
- **70.** Saphire-Bernstein S, Way BM, Kim HS, Sherman DK, Taylor SE. Oxytocin receptor gene (*OXTR*) is related to psychological resources. Proc Natl Acad Sci U S A. 2011; 108(37): 15118–15122.
- Skuse DH, Lori A, Cubells JF, Lee I, Conneely KN, Puura K, et al. Common polymorphism in the oxytocin receptor gene (OXTR) is associated with human social recognition skills. Proc Natl Acad Sci U S A. 2014; 111(5): 1987–1992. https://doi.org/10.1073/pnas.1302985111 PMID: 24367110
- 72. Thibonnier M, Graves MK, Wagner MS, Chatelain N, Soubrier F, Corvol P, et al. Study of V₁-vascular vasopressin receptor gene microsatellite polymorphisms in human essential hypertension. J Mol Cell Cardiol. 2000; 32(4): 557–564.
- 73. Staes N, Koski SE, Helsen P, Fransen E, Eens M, Stevens JMG. Chimpanzee sociability is associated with vasopressin (*Avpr1a*) but not oxytocin receptor gene (OXTR) variation. Horm Behav. 2015; 75: 84–90.
- Donaldson ZR, Kondrashov FA, Putnam A, Bai Y, Stoinski TL, Hammock EAD, et al. Evolution of a behavior-linked microsatellite-containing element in the 5' flanking region of the primate AVPR1A gene. BMC Evol Biol. 2008; 8: 180.
- Yirmiya N, Rosenberg C, Levi S, Salomon S, Shulman C, Nemanov L, et al. Association between the arginine vasopressin 1a receptor (AVPR1a) gene and autism in a family-based study: mediation by socialization skills. Mol Psychiatry. 2006; 11(5): 488–494.
- Young LJ, Wang Z. The neurobiology of pair bonding. Nat Neurosci. 2004; 7(10): 1048–1054. https://doi.org/10.1038/nn1327 PMID: 15452576
- 77. Bachner-Melman R, Zohar AH, Bacon-Shnoor N, Elizur Y, Nemanov L, Gritsenko I, et al. Link between vasopressin receptor *AVPR1A* promoter region microsatellites and measures of social behavior in humans. J Individ Differ. 2005; 26(1): 2–10.
- **78.** Prichard ZM, Mackinnon AJ, Jorm AF, Easteal S. *AVPR1A* and *OXTR* polymorphisms are associated with sexual and reproductive behavioral phenotypes in humans. Hum Mutat. 2007; 28 (11): 1150.
- 79. Walum H, Westberg L, Henningsson S, Neiderhiser JM, Reiss D, Igl W, et al. Genetic variation in the vasopressin receptor 1a gene (AVPR1A) associates with pair-bonding behavior in humans. Proc Natl Acad Sci U S A. 2008; 105(37): 14153–14156. https://doi.org/10.1073/pnas.0803081105 PMID: 18765804
- 80. Gianoulakis C. Endogenous opioids and addiction to alcohol and other drugs of abuse. Curr Top Med Chem. 2004; 4: 39–50. https://doi.org/10.2174/1568026043451573 PMID: 14754375
- Veening JG, Barendregt HP. The effects of Beta-Endorphin: state change modification. Fluids Barriers CNS, 2015; 12: 3. https://doi.org/10.1186/2045-8118-12-3 PMID: 25879522
- Contet C, Kieffer BL, Befort K. Mu opioid receptor: a gateway to drug addiction. Curr Opin Neurobiol. 2004; 14(3): 370–378. https://doi.org/10.1016/j.conb.2004.05.005 PMID: 15194118
- **83.** Mechling AE, Arefin T, Lee HL, Bienert T, Reisert M, Hamida SB, et al. Deletion of the mu opioid receptor gene in mice reshapes the reward–aversion connectome. Proc Natl Acad Sci U S A. 2016; 113(41): 11603–11608. https://doi.org/10.1073/pnas.1601640113 PMID: 27671662
- **84.** Lee MR, Gallen CL, Zhang X, Hodgkinson CA, Goldman D, Stein EA, et al. Functional polymorphism of the mu-opioid receptor gene (*OPRM1*) influences reinforcement learning in humans. PLoS ONE. 2011; 6(9): e24203.
- 85. Riju R, Ruparel K, Newberg A, Wileyto EP, Loughead JW, Divgi C, et al. Human mu opioid receptor (OPRM1 A118G) polymorphism is associated with brain mu-opioid receptor binding potential in smokers. Proc Natl Acad Sci U S A. 2011; 108(22): 9268–9273. https://doi.org/10.1073/pnas.1018699108 PMID: 21576462

- Barr CS, Schwandt M, Lindell SG, Chen SA, Goldman D, Suomi SJ, et al. Association of a functional polymorphism in the µ-opioid receptor gene with alcohol response and consumption in male rhesus macaques. Arch Gen Psychiatry. 2007; 64: 369-376.
- Miller GM, Bendor J, Tiefenbacher S, Yang H, Novak MA, Madras BK. A mu-opioid receptor single nucleotide polymorphism in rhesus monkey: association with stress response and aggression. Mol Psychiatry. 2004; 9: 99-108. https://doi.org/10.1038/sj.mp.4001378 PMID: 14699447
- Barr CS, Schwandt ML, Lindell SG, Higley JD, Maestripieri D, Goldman D, et al. Variation at the muopioid receptor gene (OPRM1) influences attachment behavior in infant primates. Proc Natl Acad Sci USA. 2008; 105(13): 5277-5281. https://doi.org/10.1073/pnas.0710225105 PMID: 18378897
- 89. Bar-Haim Y, Lamy D, Pergamin L, Bakermans-Kranenburg MJ, van IJzendoorn MH. Threat-related attentional bias in anxious and nonanxious individuals: A meta-analytic study. Psychol Bull. 2007; 133 (1): 1-24. https://doi.org/10.1037/0033-2909.133.1.1 PMID: 17201568
- 90. Crump A, Arnott G, Bethell EJ. Affect-driven attention biases as animal welfare indicators: Review and methods. Animals. 2018; 8: 136. https://doi.org/10.3390/ani8080136 PMID: 30087230
- Bethell EJ, Holmes A, MacLarnon A, Semple S. Evidence that emotion mediates social attention in rhesus macaques. PLoS ONE. 2012; 7: 1-9. https://doi.org/10.1371/journal.pone.0044387 PMID: 22952968
- Howarth ERI, Kemp C, Thatcher HR, Szott ID, Farningham D, Witham CL, et al. Developing and vali-92. dating attention bias tools for assessing trait and state affect in animals: A worked example with Macaca mulatta. Appl Anim Behav Sci. 2021; 234: 105198.
- 93. Beevers CG, Gibb BE, McGeary JE, Miller IW. Serotonin transporter genetic variation and biased attention for emotional word stimuli among psychiatric inpatients. J Abnorm Psychol. 2007; 116: 208-212. https://doi.org/10.1037/0021-843X.116.1.208 PMID: 17324031
- Fox E, Ridgewel A, Ashwin C. Looking on the bright side: biased attention and the human serotonin transporter gene. Proc R Soc B. 2009; 276: 1747-1751. https://doi.org/10.1098/rspb.2008.1788 PMID: 19324793
- Worlein JM. Nonhuman primate models of depression: effects of early experience and stress. ILAR J. 2014; 55(2): 259-273. https://doi.org/10.1093/ilar/ilu030 PMID: 25225305
- Rupia EJ, Binning SA, Roche DG, Lu W. Fight-flight or freeze-hide? Personality and metabolic pheno-96. type mediate physiological defense responses in flatfish. J Anim Ecol. 2016; 85(4): 927-937.
- Moberg GP. Biological response to stress: implications for animal welfare. In: Moberg GP, Mench JA, editors. The Biology of Animal Stress-Basic Principles and Implications for Animal Welfare. Oxon: CABI publishing; 2000. pp. 1-22.
- Morgan KN, Tromborg CT. Sources of stress in captivity. Appl Anim Behav Sci. 2007; 102: 262–302.
- Veasey J, Waran NK, Young RJ. On comparing the behaviour of zoo housed animals with wild conspecifics as a welfare indicator. Anim Welf. 1996; 5(1): 13-24.
- 100. Kagan R, Veasey J. Challenges of zoo animal welfare. In: Kleiman DG, Allen M, Thompson K, editors. Effects of Captivity on the Behavior of Wild Mammals. 2nd ed. Chicago: University of Chicago Press; 2010.
- Hambali K, Ismail A, Md-Zain BM. Daily activity budget of long-tailed macaques (Macaca fascicularis) 101. in Kuala Selangor Nature Park. Int J Sci Basic Appl Res. 2012; 12(4): 47-52.
- 102. Jaman MF. Huffman MA. The effect of urban and rural habitats and resource type on activity budgets of commensal rhesus macaques (Macaca mulatta) in Bangladesh. Primates. 2013; 54: 49-59.
- 103. Majolo B, McFarland R, Young C, Qarro M. The effect of climate factors on the activity budgets of Barbary macagues (Macaca sylvanus). Int J Primatol. 2013; 34: 500-514.
- 104. Weary DM, Niel L, Flower FC, Fraser D. Identifying and preventing pain in animals. Appl Anim Behav Sci. 2006; 100(1-2): 64-76.
- Camus SMJ, Blois-Heulin C, Li Q, Hausberger M, Bezard E. Behavioural profiles in captive-bred cyno-105. molgus macaques: Towards monkey models of mental disorders? PLoS ONE. 2013; 8: e62141. https://doi.org/10.1371/journal.pone.0062141 PMID: 23658620
- 106. Breed MD, Moores J. Homeostasis and time budgets. In: Animal Behaviour. 2nd ed. Colorado: Academic Press; 2016.
- 107. Anselme P. Abnormal patterns of displacement activities: A review and reinterpretation. Behav Processes. 2008; 79(1): 48-58. https://doi.org/10.1016/j.beproc.2008.05.001 PMID: 18554824
- Coleman K, Pierre PJ. Assessing anxiety in nonhuman primates. ILAR J. 2014; 55(2): 333-346. https://doi.org/10.1093/ilar/ilu019 PMID: 25225310
- 109. Maestripieri D, Wallen K. Affiliative and submissive communication in rhesus macaques. Primates. 1997; 38: 127-138.

- 110. Parr LA, Waller BM, Burrows AM, Gothard KM, Vick SJ. MagFACS: A muscle-based facial movement coding system for the rhesus macague. Am J Phys Anthropol. 2010; 143(4): 625-630.
- Partan S. Single and multichannel signal composition: facial expressions and vocalizations of rhesus macaques (Macaca mulatta). Behaviour. 2002; 139: 993-1027.
- Peters A, McEwan BS. Stress habituation: body shape and cardiovascular mortality. Neurosci Biobe-112. hav Rev. 2015; 56: 139-150. https://doi.org/10.1016/j.neubiorev.2015.07.001 PMID: 26148986
- Allan ATL, Hill RA. What have we been looking at? A call for consistency in studies of primate vigi-113. lance. Am J Phys Anthropol. 2018; 165(S65): 4-22.
- Shively CA, Day SM. Social inequalities in health in nonhuman primates. Neurobiol Stress. 2015; 1: 114. 156-163. https://doi.org/10.1016/j.ynstr.2014.11.005 PMID: 27589665
- Shively CA, Laber-Laird K, Anton RF. Behavior and physiology of social stress and depression in female cynomolgus monkeys. Biol Psychiatry. 1997; 41: 871-882. https://doi.org/10.1016/S0006-3223(96)00185-0 PMID: 9099414
- Shively CA. Social subordination stress, behavior, and central monoaminergic function in female cynomolgus monkeys. Biol Psychiatry. 1998; 44: 882-291. https://doi.org/10.1016/s0006-3223(97)00437x PMID: 9807643
- 117. NC3Rs. Welfare assessment. 2015. https://www.nc3rs.org.uk/macaques/welfare-assessment/.
- Kilkenny C, Browne WJ, Cuthill IC, Emerson M, Altman DG. Improving bioscience research reporting: The ARRIVE guidelines for reporting animal research. PLoS Biol. 2010; 8(6): e1000412. https://doi. org/10.1371/journal.pbio.1000412 PMID: 20613859
- de Waal FBM, Luttrell LM. The formal hierarchy of rhesus macaques: an investigation of the bared teeth display. Am J Primatol. 1985; 9(2): 73-85. https://doi.org/10.1002/ajp.1350090202 PMID: 32102494
- 120. Kemp C, Thatcher H, Farningham D, Witham C, MacLarn A, Holmes A, et al. A protocol for training group-housed rhesus macaques (Macaca mulatta) to cooperate with husbandry and research procedures using positive reinforcement. Appl Animl Behav Sci. 2017; 197: 90-100. https://doi.org/10.1016/ j.applanim.2017.08.006 PMID: 29200543
- Roberts RJ. How restriction enzymes became the workhorses of molecular biology. Proc Natl Acad 121. Sci U S A. 2005; 102(17): 5905-5908. https://doi.org/10.1073/pnas.0500923102 PMID: 15840723
- 122. Cunningham F, Achuthan P, Akanni W, Allen J, Amode AR, Armean IM, et al. Ensembl 2019. Nucleic Acids Res. 2019; 47(D1): D745-D751. https://doi.org/10.1093/nar/gky1113 PMID: 30407521
- Szott I. The effect of genotype on attention bias in rhesus macagues, Macaca mulatta, as a welfare 123. indicator. MPhil Thesis, Liverpool John Moores University. 2015. http://researchonline.ljmu.ac.uk/id/ eprint/4508/.
- Gibbs RA, Rogers J, Katze MG, Bumgarner R, Weinstock GM, Mardis ER, et al. Evolutionary and bio-124. medical insights from the rhesus macaque genome. Science, 2007; 316: 222-234. https://doi.org/10. 1126/science.1139247 PMID: 17431167
- Bennett AJ, Lesch KP, Heils A, Long JC, Lorenz JG, Shoaf SE, et al. Early experience and serotonin 125. transporter gene variation interact to influence CNS function. Mol Psychiatry, 2002; 7: 118–122.
- Karere GM, Sullivan E, Kinnally EL, Capitanio JP, Lyons LA. Enhancing genotyping of MAOA-LPR 126. and 5-HTT-LPR in rhesus macaques (Macaca mulatta). J Med Primatol. 2012; 41: 407-411.
- Sullivan EC, Mendoza SP, Capitanio JP. Similarity in temperament between mother and offspring 127. Rhesus monkeys: Sex differences and the role or monoamine oxidase-A and serotonin transporter promoter polymorphism genotypes. Dev Psychobiol. 2011; 53: 549-563.
- Okuyama Y, Ishiguro H, Nankai M, Shibuya H, Watanabe A, Arinami T. Identification of a polymor-128. phism in the promoter region of DRD4 associated with the human novelty seeking personality trait. Mol Psychiatry. 2000; 5: 64-69. https://doi.org/10.1038/sj.mp.4000563 PMID: 10673770
- Friard O, Gamba M. BORIS: a free, versatile open-source event-logging software for video/audio cod-129. ing and live observations. Methods Ecol Evol. 2016; 7(11): 1325-1330.
- R Core Team. R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. 2018. https://www.R-project.org/.
- 131. Crawley MJ. The R Book. Chichester, England: John Wiley & Sons Ltd; 2007.
- 132. Tukey JW. Exploratory Data Analysis. Reading, Massachusetts: Addison-Wesley; 1977.
- Schielzeth H. Simple means to improve the interpretability of regression coefficients. Methods Ecol 133. Evol. 2010: 1: 103-113.
- 134. Hadfield JD. MCMC methods for multi-response generalized linear mixed models: the MCMCqlmm R package. J Stat Softw. 2010; 33(2): 1-22.

- 135. McArdle PF, O'Connell JR, Pollin TI, Baumgarten M, Shuldiner AR, Peyser PA, et al. Accounting for relatedness in family based genetic association studies. Hum Hered. 2007; 64(4): 234–242. https://doi.org/10.1159/000103861 PMID: 17570925
- Gelman A, Carlin J, Stern H, Rubin D. Bayesian Data Analysis. Boca Raton, FL: Chapman & Hall;
 1995.
- 137. de Villemereuil P. Estimation of a biological trait heritability using the animal model: How to use the MCMCglmm R package. 2012. http://devillemereuil.legtux.org/wp-content/uploads/2012/12/tuto_en.pdf.
- 138. Venables WN, Ripley BD. Modern Applied Statistics with S. 4th ed. New York, NY: Springer-Verlag; 2002.
- 139. Hespanhol L, Vallio CS, Costa, LDMS, Saragiotto BT. Understanding and interpreting confidence and credible intervals around effect estimates. Braz J Phys Ther. 2019; 23(4): 290–301.
- **140.** Chang S, Berger JO. Comparison of Bayesian and Frequentist multiplicity correction for testing mutually exclusive hypotheses under data dependence. Bayesian Anal. 2021; 16(1): 111–128.
- **141.** Bradley BP, Mogg K, Millar NH. Covert and overt orienting of attention to emotional faces in anxiety. Cogn Emot. 2000; 14(6): 789–808.
- 142. Lee HY, Chang HS, Ham BJ, Kang SG, Hahn SW, Lee MS. Association of the serotonin transporter intron 2 VNTR polymorphism with treatment response to mirtazapine. Eur Neuropsychopharmacol. 2009; 19(3): S231.
- **143.** Sharma S, Nair RB, Khess CRJ. "Why am I so anxious?" A study of attention bias in obsessive compulsive disorder. Psychol Stud. 2011; 56: 247–257.
- 144. Obsessive Compulsive Cognition Working Group. Cognitive assessment of obsessive-compulsive disorder. Behav Res Ther. 1997; 35: 667–681.
- 145. Turner CM. Cognitive—behavioural theory and therapy for obsessive—compulsive disorder in children and adolescents: Current status and future directions. Clin Psychol Rev. 2006; 26: 912–938. https://doi.org/10.1016/j.cpr.2005.10.004 PMID: 16624461
- 146. Rouel M, Smith E. Attentional bias and its modification in contamination OCD Symptomatology. Cognit Ther Res. 2018; 42: 686–698.
- 147. Wang M, Han B, Liu Q, Liu C, Shuai Teng W, Du H, et al. Attentional bias of subliminal emotional faces in adolescents with obsessive-compulsive disorder. Neurocase. 2021; 27(1): 22–29. https://doi.org/10.1080/13554794.2020.1861303 PMID: 33378225
- 148. Schneier FR, Kimeldorf MB, Choo TH, Steinglass JE, Wall MM, Fyer AJ, et al. Attention bias in adults with anorexia nervosa, obsessive-compulsive disorder, and social anxiety disorder. J Psychiatr Res. 2016; 79: 61–69. https://doi.org/10.1016/j.jpsychires.2016.04.009 PMID: 27174402
- 149. Iacoviello BM, Wu G, Abend R, Murrough JW, Eder A, Fruchter E, et al. Attention bias variability and symptoms of posttraumatic stress disorder. J Trauma Stress. 2014; 27: 232–239. https://doi.org/10.1002/jts.21899 PMID: 24604631
- 150. Hong EP, Park JW. Sample size and statistical power calculation in genetic association studies. Genomics Inform. 2012; 10(2): 117–122. https://doi.org/10.5808/GI.2012.10.2.117 PMID: 23105939
- 151. Button KS, Ioannidis JPA, Mokrysz C, Nosek BA, Flint J, Robinson ASJ, et al. Power failure: why small sample size undermines the reliability of neuroscience. Neurosci. 2013; 14: 365–376. https://doi.org/10.1038/nrn3475 PMID: 23571845
- 152. Ferguson B, Street SL, Wright H, Pearson C, Jia Y, Thompson SL, et al. Single nucleotide polymorphisms (SNPs) distinguish Indian-origin and Chinese-origin rhesus macaques (*Macaca mulatta*). BMC Genomics. 2007; 8: 43.
- 153. McCormack K, Newman TK, Higley JD, Maestripieri D, Sanchez MM. Serotonin transporter gene variation, infant abuse, and responsiveness to stress in rhesus macaque mothers and infants. Horm Behav. 2009; 55: 538–547. https://doi.org/10.1016/j.yhbeh.2009.01.009 PMID: 19470363
- 154. Watson KK, Ghodasra JH, Platt ML. Serotonin transporter genotype modulates social reward and punishment in rhesus macaques. PLoS ONE. 2009; 4(1): e4156. https://doi.org/10.1371/journal.pone. 0004156 PMID: 19142220
- 155. Ebitz RB, Watson KK, Platt ML. Oxytocin blunts social vigilance in the rhesus macaque. Proc Natl Acad Sci U S A. 2013; 110(28): 11630–11635. https://doi.org/10.1073/pnas.1305230110 PMID: 23798448
- 156. Kalin NH, Shelton SE. Defensive behaviors in infant rhesus monkeys: environmental cues and neurochemical regulation. Science. 1989; 243: 1718–1721. https://doi.org/10.1126/science.2564702 PMID: 2564702

- 157. Kalin NH, Shelton SE, Rickman M, Davidson RJ. Individual differences in freezing and cortisol in infant and mother rhesus monkeys. Behav Neurosci. 1998; 112: 251–254. https://doi.org/10.1037//0735-7044.112.1.251 PMID: 9517832
- 158. Poirier C, Oliver C, Castellano Bueno J, Flecknell P, Bateson M. Pacing behaviour in laboratory macaques is an unreliable indicator of acute stress. Sci Rep. 2019; 9: 7476. https://doi.org/10.1038/s41598-019-43695-5 PMID: 31097776
- **159.** Elder CM, Menzel CR. Dissociation of cortisol and behavioural indicators of stress in orangutan (*Pongo pygmaeus*) during a computerized task. Primates. 2001; 42(4): 345–357.
- Lutz CK, Coleman K, Hopper LM, Novak MA, Perlman JE, Pomerantz O. Nonhuman primate abnormal behaviour: Etiology, assessment and treatment. Am J Primatol. 2022; 84: e23380.
- **161.** Mazumder J, Kaburu SSK. Object manipulation and tool use in Nicobar long-tailed macaques (*Macaca fascicularis umbrosus*). Int J Primatol. 2020; 41: 141–159.
- 162. Heintz MR, Santymire RM, Parr LA, Lonsdorf EV. Validation of a cortisol enzyme immunoassay and characterization of salivary cortisol circadian rhythm in chimpanzees (Pan troglodytes). Am J Primatol. 2011; 73(9): 903–908. https://doi.org/10.1002/ajp.20960 PMID: 21538448
- 163. Ruys JD, Mendoza SP, Capitanio JP, Mason WA. Behavioral and physiological adaptation to repeated chair restraint in rhesus macaques. Physiol Behav. 2004; 82: 205–213. https://doi.org/10.1016/j.physbeh.2004.02.031 PMID: 15276782
- 164. Heistermann M, Palme R, Ganswindt A. Comparison of different enzyme immunoassays for assessment of adrenocortical activity in primates based on fecal analysis. Am J Primatol. 2006; 68: 257–273. https://doi.org/10.1002/ajp.20222 PMID: 16477600
- **165.** Winterborn AN, Bates WA, Feng C, Wyatt JD. The efficacy of orally dosed ketamine and ketamine/ medetomidine compared with intramuscular ketamine in rhesus macaques (*Macaca mulatta*) and the effects of dosing route on haematological stress markers. J Med Primatol. 2008; 37(3): 116–127.
- 166. NC3Rs. Captive management. 2020. https://macaques.nc3rs.org.uk/captive-management/.
- 167. Meyer JS, Hamel AF. Models of stress in nonhuman primates and their relevance for human psychopathology and endocrine dysfunction. ILAR J. 2014; 55(2): 347–360. https://doi.org/10.1093/ilar/ilu023 PMID: 25225311
- 168. Southwick CH, Beg MA, Siddiqi MR. Rhesus monkeys in North India. In: DeVore I, editor. Primate Behavior: Field Studies in Monkeys and Apes. New York: Holt, Rinehart and Winston; 1965. pp. 111– 159
- 169. Lindburg DG. The rhesus monkey in North India: an ecological and behavioural study. In: Rosenblum LA, editor. Primate Behavior: Development in Field and Laboratory Research (Vol. 2). New York: Academic Press; 1971. pp. 1–106.
- 170. Harvey PH, Martin RD, Clutton-Brock TH. Life histories in comparative perspective. In: Smuts BB, Cheyney DL, Seyfarth RM, Wrangham RW, Struhsaker TT, editors. *Primate Societies*. Chicago: University of Chicago Press; 1987. pp. 181–186.
- 171. Prescott MJ, Nixon ME, Farningham DAH, Naiken S, Griffiths M-A. Laboratory macaques: When to wean? Appl Anim Behav Sci. 2012; 137(3–4): 194–207.
- 172. NC3Rs. Weaning in macaques. 2021. https://www.nc3rs.org.uk/our-portfolio/weaning-macaques.
- 173. Weisstaub NV, Zhou M, Lira A, Lambe E, González-Maeso J, Hornung JP, et al. Cortical 5-HT2A receptor signaling modulates anxiety-like behaviors in mice. Science. 2006; 313: 536–540. https://doi.org/10.1126/science.1123432 PMID: 16873667
- 174. Shelton RC, Sanders-Bush E, Manier DH, Lewis DA. Elevated 5-HT2A receptors in postmortem prefrontal cortex in major depression is associated with reduced activity of protein kinase A. Neurosci. 2009; 158: 1406–1415. https://doi.org/10.1016/j.neuroscience.2008.11.036 PMID: 19111907
- 175. Isosaka T, Matsuo T, Yamaguchi T, Funabiki K, Nakanishi S, Kobayakawa R, et al. Htr2a-expressing cell in the central amygdala control the hierarchy between innate and learned fear. Cell. 2015; 163(5): 1153–1164.
- 176. Sartori SB, Hauschild M, Bunck M, Gaburro S, Landgraf R, Singewald N. Enhanced fear expression in a psychopathological mouse model of trait anxiety: pharmacological interventions. PLoS ONE. 2011; 6(2): e16849. https://doi.org/10.1371/journal.pone.0016849 PMID: 21386891
- Quah SKL, McIver L, Roberts AC, Santangelo AM. Trait anxiety mediated by amygdala serotonin transporter in the common marmoset. J Neurosci. 2020; 40(24): 4739–4749. https://doi.org/10.1523/JNEUROSCI.2930-19.2020 PMID: 32393533
- 178. Myers AJ, Williams L, Gatt JM, McAuley-Clark EZ, Dobson-Stone C, Schofield PR, et al. Variation in the oxytocin receptor gene is associated with increased risk for anxiety, stress and depression in individuals with a history of exposure to early life stress. J Psychiatr Res. 2014; 59: 93–100. https://doi.org/10.1016/j.jpsychires.2014.08.021 PMID: 25262417

- 179. Zeev-Wolf M, Levy J, Ebstein RP, Feldman R. Cumulative risk on oxytocin-pathway genes impairs default mode network connectivity in trauma-exposed youth. Front Endocrinol. 2020; 11: 335. https:// doi.org/10.3389/fendo.2020.00335 PMID: 32528417
- 180. Avinun R, Israel S, Shalev I, Gritsenko I, Bornstein G, Ebstein RP, et al. AVPR1A variant associated with preschoolers' lower altruistic behavior. PLoS ONE. 2011; 6: e25274. https://doi.org/10.1371/journal.pone.0025274 PMID: 21980412