

Curcumin production by engineered *Escherichia coli* biofilms

Ana Azevedo^{1,2*}, Marta M. Ferreira^{1,2}, Luciana C. Gomes^{1,2}, Gabriel A. Monteiro³ and Filipe J. M. Mergulhão^{1,2}

1. LEPABE - Laboratory for Process Engineering, Environment, Biotechnology and Energy, Faculty of Engineering, University of Porto, Rua Dr. Roberto Frias, 4200-465 Porto, Portugal

2. ALiCE - Associate Laboratory in Chemical Engineering, Faculty of Engineering, University of Porto, Rua Dr. Roberto Frias, 4200-465 Porto, Portugal

3. iBB - Institute for Bioengineering and Biosciences, Department of Bioengineering, Instituto Superior Técnico, University of Lisbon, 1049-001 Lisboa, Portugal

* up201407568@edu.fe.up.pt

Introduction

Curcuminoids are polyphenolic compounds isolated from the rhizome of *Curcuma longa*, and are a mixture of curcumin, desmethoxycurcumin and bisdemethoxycurcumin.

Despite being famous for their therapeutic properties, pure curcuminoids are very expensive and rare because the plant is seasonal and grows slowly.

The production of curcuminoids has been described using an artificial pathway in *E. coli* [1]. A 4-step synthetic pathway has already been tested in planktonic *E. coli* cells, being able to produce 563 mg/L of curcumin from ferulic acid [2].

The main goal of this work was to attempt the production of curcumin, an added-value compound, using biofilms as production platforms.

Material and Methods

E. coli JM109(DE3) cells transformed with a plasmid carrying the three genes encoding for the artificial pathway of curcumin was used. Figure 1 illustrates the workflow of this study.

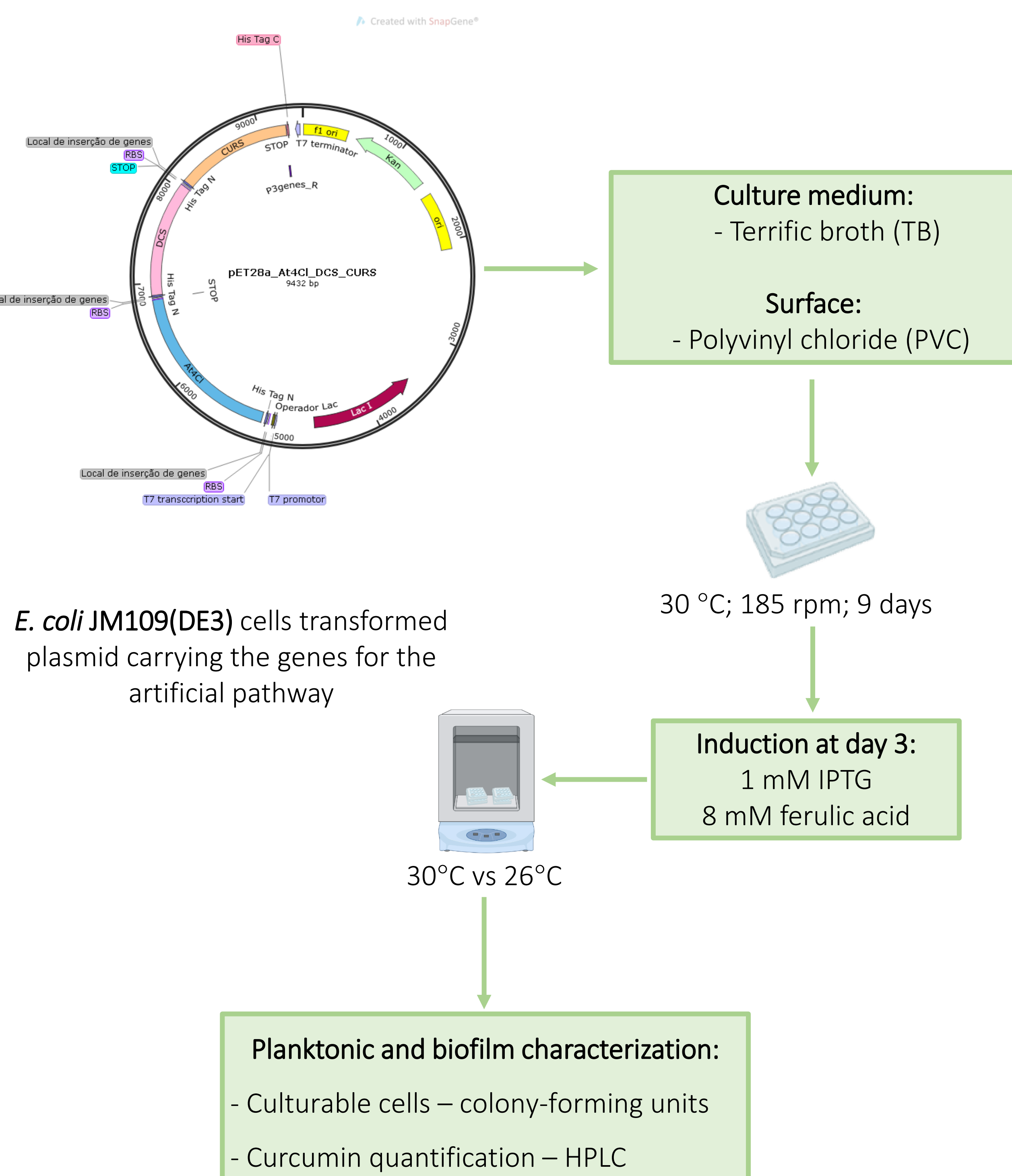


Fig. 1 – Diagram of the materials and methodology used in these assays

Results

Regarding planktonic fraction (fig.2-A) at day 4, induced cultures at 30°C presented 14% more culturable cells than at 26°C. This difference became more evident on days 6 and 7, with 73% and 85% more culturable cells, respectively, at 30°C than at 26°C ($p < 0.05$).

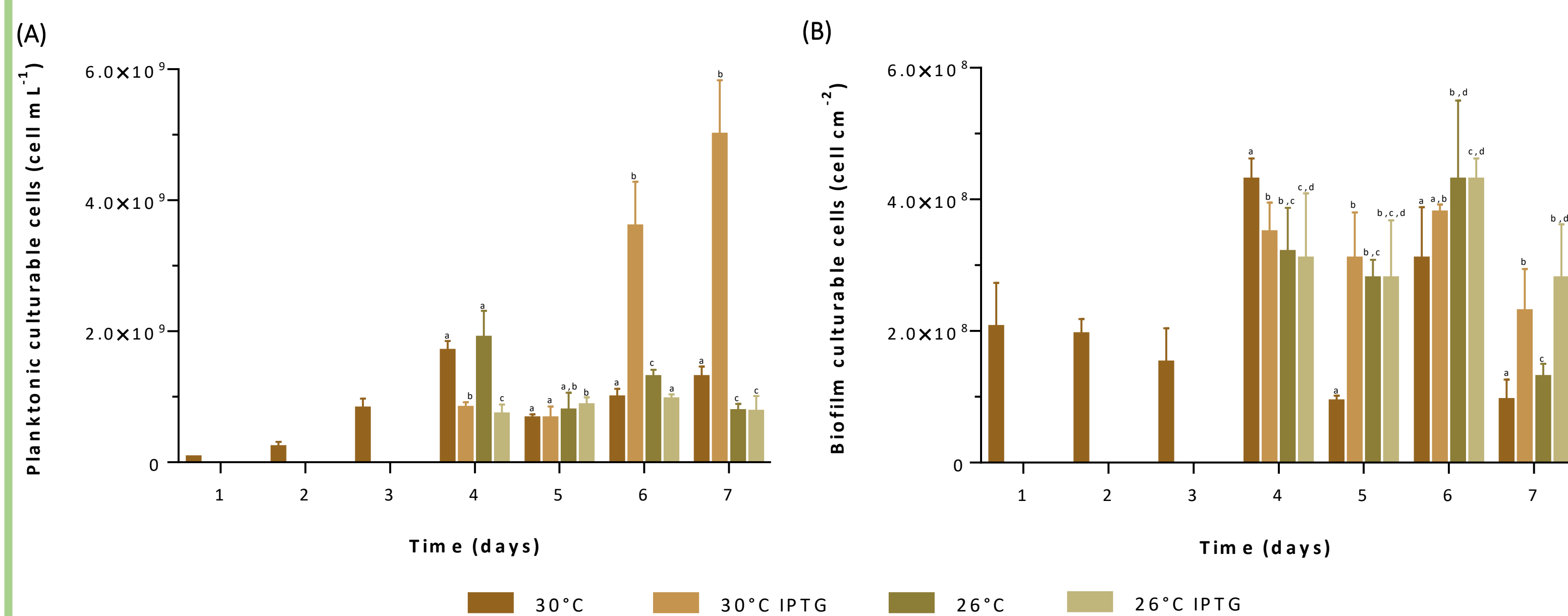


Fig. 2 – Planktonic (A) and biofilm culturable cells (B) during a 7-day experiment. The means and standard deviations (SDs) of three independent experiments are presented. Statistically significant differences on each time point were considered, and different lowercase letters indicate statistically significant differences between the conditions on each day ($p < 0.05$).

Regarding biofilm fraction (fig.2-B), biofilms grown at 30°C presented a similar pattern to that observed in the planktonic state. After day 4, viable cells at 30°C showed a decrease, maintaining levels below those observed for biofilms grown at 26°C, with and without induction. For biofilms incubated at 26°C, induction did not promote an increase in the number of biofilm culturable cells (except on day 7).

Curcumin production was about 14-fold higher in the biofilm state than in the planktonic state at days 4 and 5 (fig. 3), with production values of on average of 0.14 and 1.77 fg.cell⁻¹ on planktonic and biofilm states, respectively.

Regarding the biofilm state, 30 °C seemed to slightly benefit curcumin production.

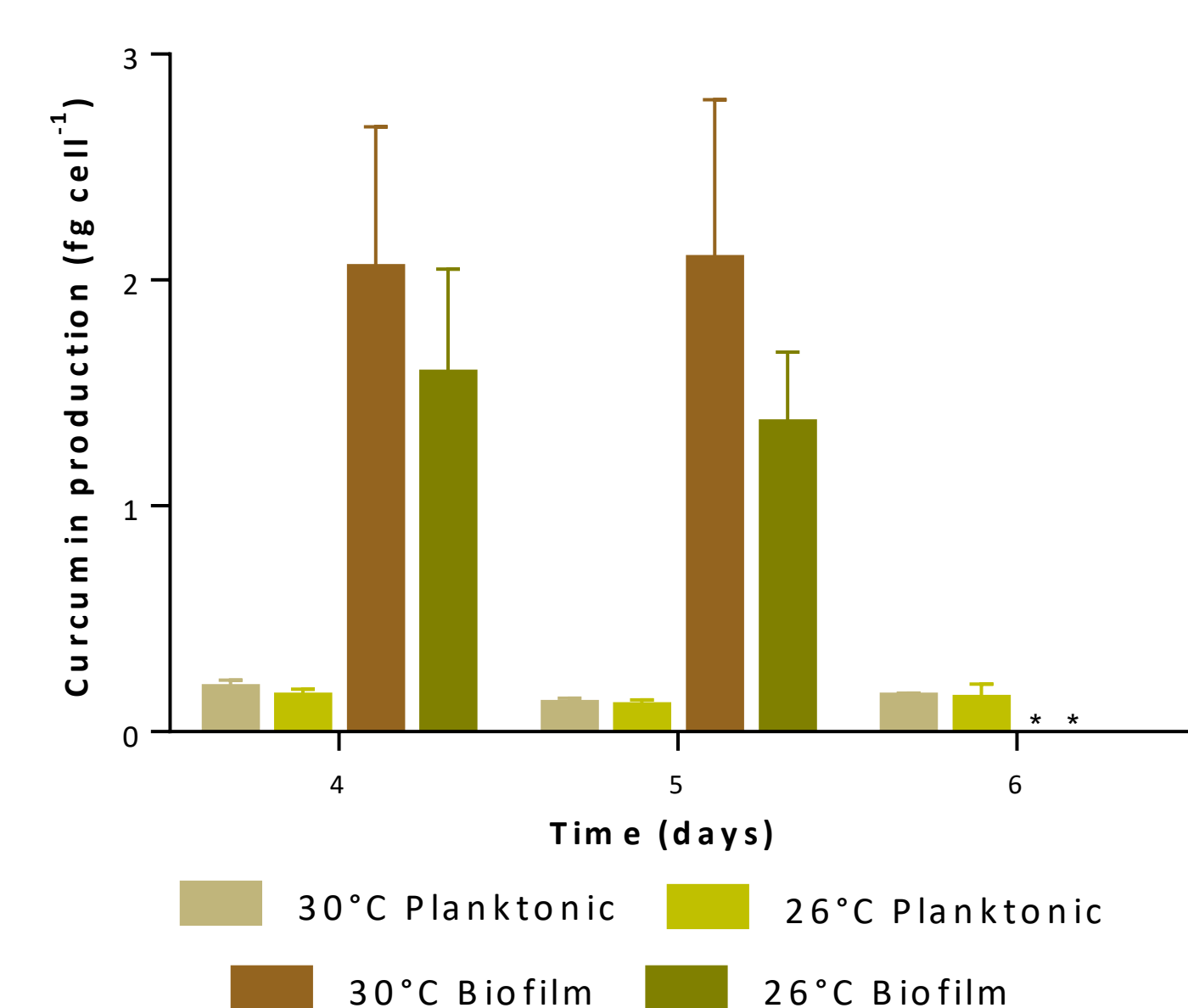


Fig. 3 – Curcumin production after induction at different temperatures. * no curcumin production was quantified.

Conclusion

In general, induction negatively affected the growth of planktonic and biofilm cultures, except at 30°C where the chemical induction promoted the planktonic growth.

Biofilm state tended to promote the production of curcumin compared to the planktonic state, and 30°C seemed to benefit the biofilm production.

Acknowledgements

This work was financially supported by: LA/P/0045/2020 (ALiCE) and UIDB/00511/2020 - UIDP/00511/2020 (LEPABE) funded by national funds through FCT/MCTES (PIDDAC); and by Project PTDC/BII-BIO/29589/2017 - POCI-01-0145-FEDER-029589 - funded by FEDER funds through COMPETE2020 - Programa Operacional Competitividade e Internacionalização (POCI) and by national funds (PIDDAC) through FCT/MCTES; SurfSAFE project funded by the European Union's Horizon 2020 research and innovation programme under grant agreement No. 952471. A. Azevedo acknowledges the receipt of a Ph.D. Grant from FCT (2020.07427.BD).

References

1. <https://doi.org/10.1099/mic.0.2008/018721-0>
2. <https://doi.org/10.3389/fbioe.2020.00059>