



Quercetin from methanol extracts of the leaves of *Ficus benghalensis*

Hassan Abdalla Almahy Dafalla 

Department of Chemistry, Alkurmah University College, Taif University, Saudi Arabia;

Faculty of Science, Department of Chemistry, University of Bahari, Sudan

Suggested Citation

Dafalla, H.A.A. (2023). Quercetin from methanol extracts of the leaves of *Ficus benghalensis*. *European Journal of Theoretical and Applied Sciences*, 1(6), 185-190.
DOI: [10.59324/ejtas.2023.1\(6\).19](https://doi.org/10.59324/ejtas.2023.1(6).19)

Abstract:

Flavonoids are ubiquitous in photosynthesising cells and are commonly found in fruit, vegetables, nuts, seeds, stems, flowers, tea, wine, propolis and honey. For centuries, preparations containing these compounds as the principal physiologically active constituents have been used to treat human diseases. Increasingly, this class of natural products is becoming the subject of anti-infective research and many groups have isolated and identified the structures of flavonoids. In this study we concerned on determination flavonoids

content from methanolic extracts of *Ficus benghalensis* using catechol and quercetin as standard. The result shows high content of quercetin in methanolic extracts.

Keywords: *flavonoids, quercetin, moraceae, extracts.*

Introduction

Quercetin belongs to a group of plant pigments called flavonoids that give many fruits, flowers and vegetables their colors. Flavonoids, such as quercetin, are antioxidants. They scavenge particles in the body known as free radicals which damage cell membranes, tamper with DNA and even cause cell death. Antioxidants can neutralize free radicals. They may reduce or even help prevent some of the damage free radicals cause. Quercetin may help protect against heart disease and cancer and it can also help stabilize the cells that release histamine in the body and thereby have an anti-inflammatory and antihistamine effect. It is categorized as a flavonol, one of the six subclasses of flavonoid compounds. The name has been used since 1857 and is derived from quercetum. It is a naturally occurring polar auxin transport inhibitor (Davis et al., 2009; Aguirre et al., 2011; Fischer et al., 1997). The common forms of quercetin were shown in Figure 1.

Quercetin is an aglycone lacking an attached sugar. A quercetin glycoside is formed by attaching a glycosyl group (a sugar such as glucose, rhamnose or rutinose) as a replacement for one of the OH groups (commonly at position 3). The attached glycosyl group can change the solubility absorption and *in vivo* effects. As a general rule of thumb, the presence of a glycosyl group (quercetin glycoside) results in increased water solubility compared to quercetin aglycone (Ross & Kasum, 2002; Hollman et al., 1999). Quercetin-type flavonols (primarily as quercetin glycosides), the most abundant of the flavonoid molecules are widely distributed in plants. They are found in a variety of foods including apples, grapes, onions, shallots, tea and tomatoes, as well as many seeds, nuts, flowers, barks and leaves. It is also found in medicinal botanicals, including *Ginkgo biloba*, *Hypericum perforatum* and *Sambucus canadensis* (Häkkinen et al., 1999; Williamson & Manach, 2005; Wiczkowski et al., 2008). In red onions, higher concentrations of quercetin occur in the



outermost rings and in the part closest to the root, the latter being the part of the plant with the highest concentration (Smith et al., 2003). One study found that organically grown tomatoes had 79% more quercetin than chemically grown fruit (Mitchell et al., 2007). Quercetin is present in various kinds of honey from different plant sources (Petrus, Schwartz, & Sontag, 2011). Food-based sources of

quercetin include vegetables, fruits, berries, nuts, beverages and other products of plant origin (Tutel'ian & Lashneva, 2013). In the determined food, the highest concentration is 234 mg/100 g of edible portion in capers (raw), the lowest concentration is 2 mg/100 g of edible portion in black or green tea (Bhagwat, Haytowits & Holden, 2011).

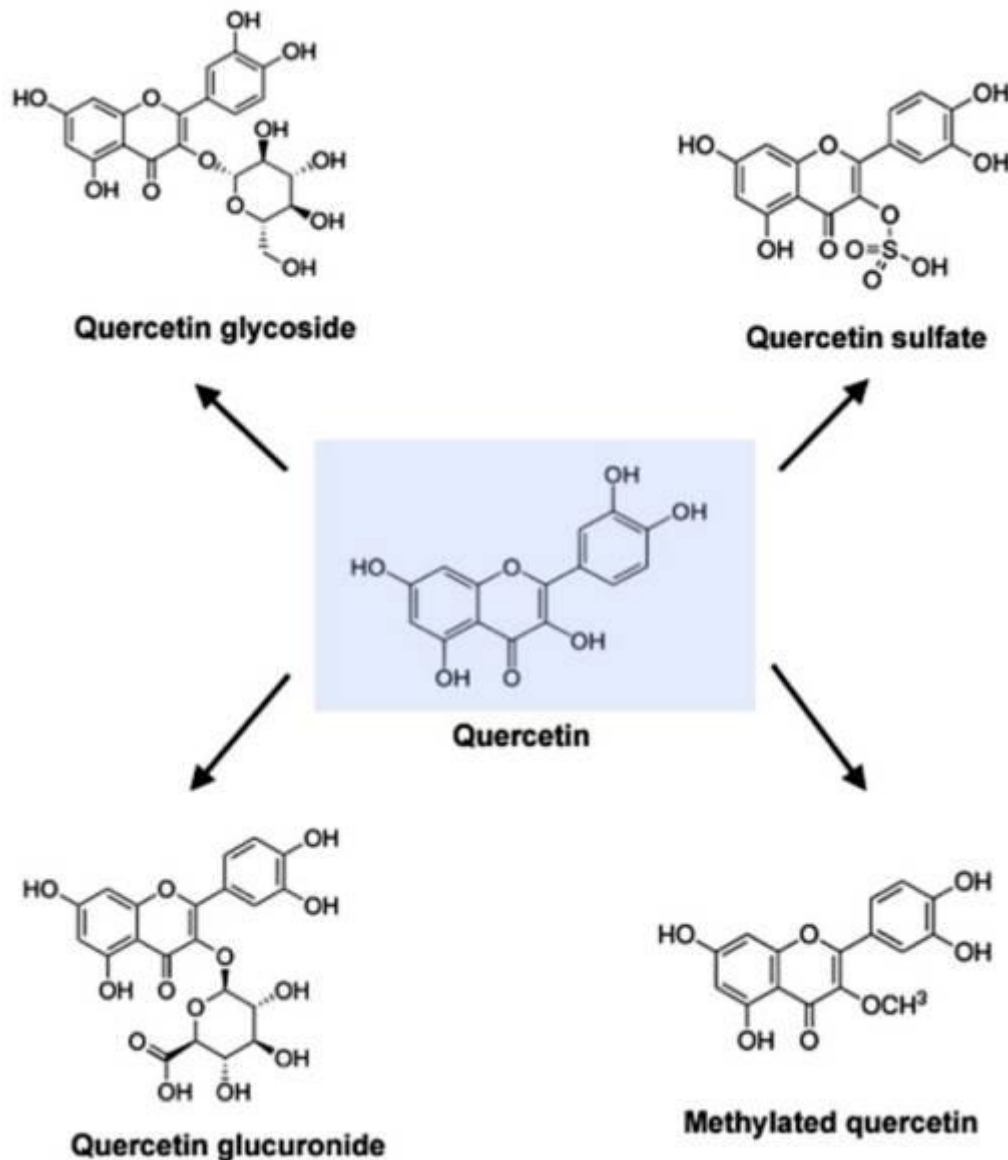


Figure 1. Flavonoids Compounds from *Petroselinum crispum* L.

Dietary intake of quercetin was different in several countries. The estimated flavonoid intake ranges from 50 to 800 mg/day (quercetin

accounts for 75%), mostly depending on the consumption of fruits and vegetables and the intake of tea (Chun, Chung & Song, 2007; Sun et

al., 2015; Zhang et al., 2010). Onions, tea and apples contained high amounts of quercetin (Sampson et al., 2002). In Japan, the average and median quercetin intakes were 16.2 and 15.5 mg/day, respectively, the quercetin intake by men was lower than that by women and the quercetin intakes showed a low correlation with age in both men and women. The estimated quercetin intake was similar during summer and winter. In Australia, black and green teas were the dominant sources of quercetin. Other sources included onion, broccoli, apple, grape and beans (Nishimuro et al., 2015; Somerset, S. & Johannot, 2008). In Spain, the average daily intake of quercetin is 18.48 mg/day, which is significantly higher than that in the United States (9.75 mg/day), based on sources like tea, citrus fruits and juice, beers and ales, wines, melon, apples, onions, berries and bananas (Zamora et al., 2010).



Figure 2. Fresh Leaves of *Ficus benghalensis*

Materials and methods

Plant Materials

The leaves of *Ficus benghalensis* were collected from Alkurmah State in the morning, then plant material was dried at room temperature in April 2023. The dried and finely ground samples of the leaves (48g) were extracted with petroleum ether (40- 80°C) for defatting of fatty acids then the dried residues was extracted with methanol solvents by maceration (48 hours). The solvents removed under vacuum at temperature below 50°C. Each extract was prepared just before the experiment to prevent any further degradation.

General Material and Methods

Column chromatography and analytical TLC were carried out using silica gel, Merck 230-400 mesh and Merck 70-230 mesh. Melting points were determined on a Kofler hot-stage microscope melting point apparatus and were uncorrected. ¹H and ¹³C-NMR were recorded on JEOL ANM-GSX 400 spectrometer. Mass spectra were obtained using GCMS-QP 5050 A Shimadzu mass spectrometer.

Extraction and Isolation of Quercetin

The leaves of *Ficus benghalensis* were air-dried, ground and subjected to extraction with petroleum ether (40-80°C) and methanol successively. The solvents were then removed under reduced pressure to give the following extracts as shown in table 1.

Table 1. The Extracts of the Leaves of *Ficus benghalensis*

Material (g)	Solvent	Colour and nature of extracts	Yield of extracts (g)
Leaves (2400)	petroleum ether	yellowish-brown, semi-solid	14.8
	methanol	dark-brown, solid	48.4

Isolation of Quercetin

The crude methanol extract (24.4g) was subjected to silica gel column chromatography using gradient solvent mixtures of 100% petroleum ether, petroleum ether/chloroform (1:1), 100% chloroform and chloroform / methanol (5:2) to give 15 fractions. Fractions 8 and 14 which gave the same values (0.35,

chloroform:methanol, 3:1) on TLC were combined together and resubjected to column chromatography with chloroform and methanol (1:1) to give quercetin as yellow color (42.3 mg) with melting point 314°C (Scholz and Williamson 2007, 316°C) (Scholz & Williamson, 2007).

Results and Discussion

The present study has shown the compound obtained from methanol extracts of the leaves of *Ficus benghalensis* was being quercetin according to interpretation of the spectral data of $^1\text{H-NMR}$ and $^{13}\text{C-NMR}$ as shown in figure 3 and figure 4 respectively.

In $^1\text{H-NMR}$ spectrum there were five protons in the aromatic region appearing at singlets at δ 7, 7.1, 7.4, 8.2 and 12 assigned for proton at C-

3, C-3, C-4, C-7 and C-5 respectively. The two sharp singlets in the downfield region at δ 8.2 and 12 were assigned to the two chelated hydroxyl groups at positions 5 and 7. The $^{13}\text{C-NMR}$ spectrum indicated the presence of fifteen carbons in the structure. The mass spectrum gave a molecular ion peak at m/z 302 which corresponded to the molecular formula $\text{C}_{15}\text{H}_{10}\text{O}_7$. On comparison the spectral with published data (Day et al., 2000) suggested the structure to be 3, 3', 4', 5, 7-pentahydroxyflvanone (quercetin).

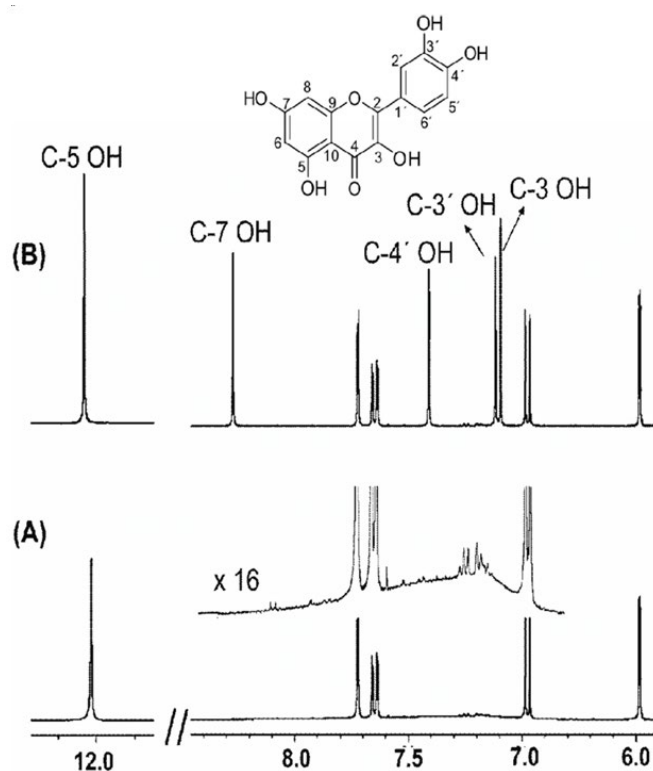


Figure 3. $^1\text{H-NMR}$ for quercetin

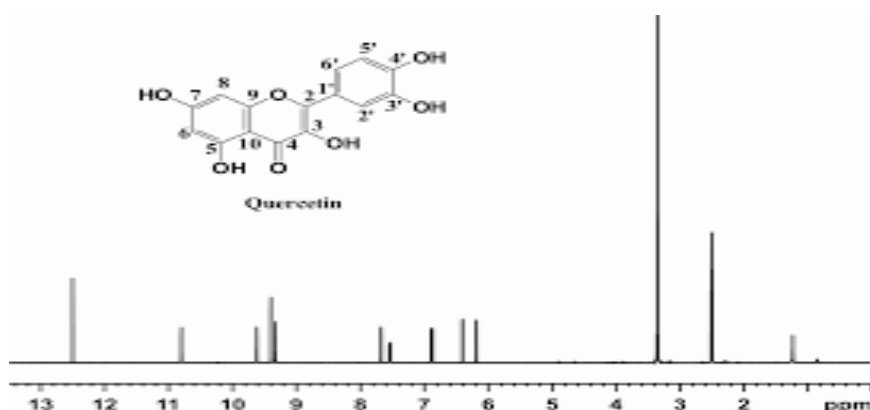


Figure 4. $^{13}\text{C-NMR}$ for quercetin

Conclusions

Isolation of phytochemical compounds often leads to possible synthesis in the laboratory and on the industrial scale. The methanol extract of leaves shows higher flavonoids content. Therefore, the plant is an excellent source of quercetin according to the percentage yield.

References

- Aguirre, L., Arias, N., Macarulla, M.T., Gracia, A., & Portillo, M.P. (2011). Beneficial Effects of Quercetin on Obesity and Diabetes. *The Open Nutraceuticals Journal*, 4, 189-198. <https://doi.org/10.2174/1876396001104010182>
- Bhagwat, S., Haytowitz, D.B., & Holden, J.M. (2011). *USDA Database for the Flavonoid Content of Selected Foods, Release 3*. U.S. Department of Agriculture; Beltsville, MD, USA.
- Chun, O. K., Chung, S. J., & Song, W. O. (2007). Estimated dietary flavonoid intake and major food sources of U.S. adults. *The Journal of nutrition*, 137(5), 1244–1252. <https://doi.org/10.1093/jn/137.5.1244>
- Crespy, V., Morand, C., Manach, C., Besson, C., Demigne, C., & Remesy, C. (1999). Part of quercetin absorbed in the small intestine is conjugated and further secreted in the intestinal lumen. *The American journal of physiology*, 277(1), G120–G126. <https://doi.org/10.1152/ajpgi.1999.277.1.G120>
- Davis, J. M., Murphy, E. A., & Carmichael, M. D. (2009). Effects of the dietary flavonoid quercetin upon performance and health. *Current sports medicine reports*, 8(4), 206–213. <https://doi.org/10.1249/JSR.0b013e3181ae8952>
- Day, A. J., Bao, Y., Morgan, M. R., & Williamson, G. (2000). Conjugation position of quercetin glucuronides and effect on biological activity. *Free radical biology & medicine*, 29(12), 1234–1243. [https://doi.org/10.1016/S0891-5849\(00\)00416-0](https://doi.org/10.1016/S0891-5849(00)00416-0)
- Fischer, C., Speth, V., Fleig-Eberenz, S., & Neuhaus, G. (1997). Induction of Zygotic Polyembryos in Wheat: Influence of Auxin Polar Transport. *The Plant cell*, 9(10), 1767–1780. <https://doi.org/10.1105/tpc.9.10.1767>
- Häkkinen, S. H., Kärenlampi, S. O., Heinonen, I. M., Mykkänen, H. M., & Törrönen, A. R. (1999). Content of the flavonols quercetin, myricetin, and kaempferol in 25 edible berries. *Journal of agricultural and food chemistry*, 47(6), 2274–2279. <https://doi.org/10.1021/jf9811065>
- Hollman, P. C., Bijlsman, M. N., van Gameren, Y., Cnossen, E. P., de Vries, J. H., & Katan, M. B. (1999). The sugar moiety is a major determinant of the absorption of dietary flavonoid glycosides in man. *Free radical research*, 31(6), 569–573. <https://doi.org/10.1080/10715769900301141>
- Mitchell, A. E., Hong, Y. J., Koh, E., Barrett, D. M., Bryant, D. E., Denison, R. F., & Kaffka, S. (2007). Ten-year comparison of the influence of organic and conventional crop management practices on the content of flavonoids in tomatoes. *Journal of agricultural and food chemistry*, 55(15), 6154–6159. <https://doi.org/10.1021/jf070344+>
- Nishimuro, H., Ohnishi, H., Sato, M., Ohnishi-Kameyama, M., Matsunaga, I., Naito, S., Ippoushi, K., Oike, H., Nagata, T., Akasaka, H., Saitoh, S., Shimamoto, K., & Kobori, M. (2015). Estimated daily intake and seasonal food sources of quercetin in Japan. *Nutrients*, 7(4), 2345–2358. <https://doi.org/10.3390/nu7042345>
- Petrus, K., Schwartz, H., & Sontag, G. (2011). Analysis of flavonoids in honey by HPLC coupled with coulometric electrode array detection and electrospray ionization mass spectrometry. *Analytical and bioanalytical chemistry*, 400(8), 2555–2563. <https://doi.org/10.1007/s00216-010-4614-7>
- Ross, J. A., & Kasum, C. M. (2002). Dietary flavonoids: bioavailability, metabolic effects, and safety. *Annual review of nutrition*, 22, 19–34.

<https://doi.org/10.1146/annurev.nutr.22.1114.01.144957>

Sampson, L., Rimm, E., Hollman, P. C., de Vries, J. H., & Katan, M. B. (2002). Flavonol and flavone intakes in US health professionals. *Journal of the American Dietetic Association*, 102(10), 1414–1420.

[https://doi.org/10.1016/s0002-8223\(02\)90314-7](https://doi.org/10.1016/s0002-8223(02)90314-7)

Scholz, S., & Williamson, G. (2007). Interactions affecting the bioavailability of dietary polyphenols in vivo. *International journal for vitamin and nutrition research. Internationale Zeitschrift für Vitamin- und Ernährungsforschung. Journal international de vitaminologie et de nutrition*, 77(3), 224–235.

<https://doi.org/10.1024/0300-9831.77.3.224>

Smith, C.L., Lombard, K.A., Peffley, E.B., & Liu, W. (2003). Genetic Analysis of Quercetin in Onion (*Allium cepa* L.) 'Lady Raider'. *Texas Journal of Agriculture and Natural Resources*, 16, 24–28.

Somerset, S. M., & Johannot, L. (2008). Dietary flavonoid sources in Australian adults. *Nutrition and cancer*, 60(4), 442–449.

<https://doi.org/10.1080/01635580802143836>

Sun, C., Wang, H., Wang, D., Chen, Y., Zhao, Y., & Xia, W. (2015). Using an FFQ to assess intakes of dietary flavonols and flavones among female adolescents in the Suihua area of northern China. *Public health nutrition*, 18(4), 632–639.

<https://doi.org/10.1017/S1368980014000780>

Tutel'ian, V. A., & Lashneva, N. V. (2013). Biologically active substances of plant origin.

Flavonols and flavones: Prevalence, dietary sources and consumption. *Voprosy pitaniia*, 82(1), 4–22.

Wiczowski, W., Romaszko, J., Bucinski, A., Szawara-Nowak, D., Honke, J., Zielinski, H., & Piskula, M. K. (2008). Quercetin from shallots (*Allium cepa* L. var. *aggregatum*) is more bioavailable than its glucosides. *The Journal of nutrition*, 138(5), 885–888.

<https://doi.org/10.1093/jn/138.5.885>

Williamson, G., & Manach, C. (2005). Bioavailability and bioefficacy of polyphenols in humans. II. Review of 93 intervention studies. *The American journal of clinical nutrition*, 81(1 Suppl), 243S–255S.

<https://doi.org/10.1093/ajcn/81.1.243S>

Zamora-Ros, R., Andres-Lacueva, C., Lamuela-Raventós, R. M., Berenguer, T., Jakszyn, P., Barricarte, A., Ardanaz, E., Amiano, P., Dorronsoro, M., Larrañaga, N., Martínez, C., Sánchez, M. J., Navarro, C., Chirlaque, M. D., Tormo, M. J., Quirós, J. R., & González, C. A. (2010). Estimation of dietary sources and flavonoid intake in a Spanish adult population (EPIC-Spain). *Journal of the American Dietetic Association*, 110(3), 390–398.

<https://doi.org/10.1016/j.jada.2009.11.024>

Zhang, Y., Li, Y., Cao, C., Cao, J., Chen, W., Zhang, Y., Wang, C., Wang, J., Zhang, X., & Zhao, X. (2010). Dietary flavonol and flavone intakes and their major food sources in Chinese adults. *Nutrition and cancer*, 62(8), 1120–1127.

<https://doi.org/10.1080/01635581.2010.513800>