



# BOOK OF ABSTRACTS

XXI EUROFOODCHEM

22-24 November 2021

On-line conference

**TITLE**

Book of Abstracts of the XXI EuroFoodChem Congress

**EDITORS**

Joana S. Amaral, Cristina Todasca, Michael Murkovic, Marco Arlorio, Tanja Cirković Veličković, Hans-Jacob Skarpeid, Karel Cejpek, Irena Vovk, Livia Simon Sarkadi, Małgorzata Starowicz, Matthias Wüst, Robert Tincu, Vuk Filipovic.

**EDITION**

Sociedade Portuguesa de Química  
Av. Da República, 45 – 3º Esq  
1050-187 Lisboa – Portugal

**DATE**

November 2021

**ISBN**

ISBN 978-989-8124-34-0



9 789898 124340

@ Sociedade Portuguesa de Química

All rights reserved.

The editors state that the content of scientific abstracts is of the responsibility of their respective authors.

## XXI EUROFOODCHEM CONFERENCE

### Scientific Committee

Joana Amaral (Portugal) – Chair, FCD-EuChemS  
Cristina Todasca (Romania) – Secretary, FCD-EuChemS  
Michael Murkovic (Austria) – Treasurer, FCD-EuChemS  
Marco Arlorio (Italy) – past-Chair, FCD-EuChemS  
Tanja Cirković Veličković (Serbia)  
Celestino Santos-Buelga (Spain)  
Hans-Jacob Skarpeid (Norway)  
Irena Vovk (Slovenia)  
Karel Cejpek (Czech Republic)  
Lillian Barros (Portugal)  
Livia Simon Sarkadi (Hungary)  
M. Beatriz P.P. Oliveira (Portugal)  
Małgorzata Starowicz (Poland)  
Manuel Coimbra (Portugal)  
María J. Cantalejo (Spain)  
Matthias Wüst (Germany)  
Michael Granvogl (Germany)  
Reto Battaglia (Switzerland)  
Slavica Ražić (Serbia, Division of Analytical Chemistry - EuChemS)  
Wiesław Wiczkowski (Poland)  
Zuzana Ciesarová (Slovakia)

### Organizing Committee

Joana Amaral (Portugal)  
Cristina Todasca (Romania)  
Michael Murkovic (Austria)  
Marco Arlorio (Italy)  
Tanja Cirković Veličković (Serbia)  
Hans-Jacob Skarpeid (Norway)  
Irena Vovk (Slovenia)  
Karel Cejpek (Czech Republic)  
Livia Simon Sarkadi (Hungary)  
Małgorzata Starowicz (Poland)  
Matthias Wüst (Germany)  
Robert Tincu (Romania)  
Vuk Filipovic (Serbia)

**Conference organized under the auspices of the Food Chemistry Division of the European Chemical Society (FCD-EuChemS), the Portuguese Chemical Society (SPQ) and the Serbian Chemical Society.**

## Extraction and quantification of tropomyosin in selected samples of shellfish

**Mirjana Radomirović<sup>1</sup>, Nikola Gligorijević<sup>2</sup>, Dragana Stanić-Vučinić<sup>1</sup>, Andreja Rajković<sup>3</sup>,  
Tanja Ćirković Veličković<sup>1,3,4,5</sup>**

<sup>1</sup> University of Belgrade - Faculty of Chemistry, Center of Excellence for Molecular Food Sciences and Department of Biochemistry, Belgrade, Serbia

<sup>2</sup> Institute for the Application of Nuclear Energy, Department of Metabolism, University of Belgrade, Belgrade, Serbia

<sup>3</sup> Faculty of Bioscience Engineering, Ghent University, Ghent, Belgium

<sup>4</sup> Ghent University Global Campus, Incheon, Korea

<sup>5</sup> Serbian Academy of Sciences and Arts, Belgrade, Serbia

\*tcirkov@chem.bg.ac.rs

Food allergies affect up to 10% of the general population and represent an important health problem in the field of food safety in industrialized countries. Hence, developing reliable, specific, and sensitive methods for detecting and quantifying allergens in food products is of high importance. Shellfish have been recognized as one of the eight most common sources of allergens, with tropomyosin (TPM) being considered a major heat-stable allergen, having a highly conserved amino acid sequence among different shellfish species. Allergenicity of TPM may change during food processing, such as cooking. The objective of this study was to develop an enzyme-linked immunosorbent assay (ELISA) for the detection and quantification of shellfish tropomyosin in food samples.

Two different extraction buffers - phosphate-buffered saline (PBS) and PBS containing 1 M sodium-chloride (PBSN), were compared for their ability to recover proteins from pre-cooked frozen Mediterranean mussel (*Mytilus galloprovincialis*) and fresh frozen razor mud shrimp (*Solenocera melanthero*). The samples were additionally cooked according to the manufacturer's instruction and analyzed as such. The protein content was quantified using Bradford protein assay, and the protein components of soluble extracts were profiled using SDS-PAGE. TPM presence was confirmed using Western blot. Sandwich ELISA was developed using a monoclonal anti-TPM antibody as a capture antibody, while polyclonal anti-TPM antibody served as a detection antibody and was coupled to the biotinylated secondary antibody and streptavidin-alkaline phosphatase conjugate. Tropomyosin was quantified using highly purified natural shrimp tropomyosin as standard.

The profile of extracted proteins was changed when using PBSN instead of PBS. A higher concentration of proteins was recovered from raw shrimp using PBSN instead of PBS. At the same time, the type of extraction buffer did not affect protein recovery either from heated shrimp or pre-cooked/heated mussels. Significantly fewer proteins were extracted from cooked shrimp sample compared to the raw shrimp, while cooking showed no effect on the extraction of proteins from mussels. Cooking did not affect TPM recognition in Western blot. TPM was quantified in shrimp samples in sandwich ELISA. However, developed ELISA could not quantify mussel's TPM, indicating that this approach may distinguish mussels and shrimp TPM.

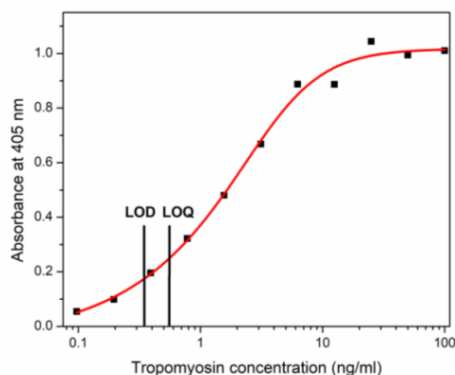


Fig. 1. Quantification of tropomyosin using sandwich ELISA.

**Acknowledgments:** This research work was supported by the Ministry of Education, Science and Technological Development of the Republic of Serbia, contract number: 451-03-9/2021-14/200168; the Science Fund of the Republic of Serbia, Program DIASPORA, #6504499, ShellPCR, and the European Commission, under the Horizon2020, FoodEnTwin Project, GA No. 810752.