# Activation and Regulation of NLR Immune Receptor Networks

Jiorgos Kourelis[1], Hiroaki Adachi[2*]

[1]The Sainsbury Laboratory, University of East Anglia, Norwich Research Park, NR4 7UH, Norwich, UK
[2] Laboratory of Crop Evolution, Graduate School of Agriculture, Kyoto University, Mozume, Muko, Kyoto 617-0001, Japan

[*]Correspondence: adachi.hiroaki.3s@kyoto-u.ac.jp

## ABSTRACT

**Plants have many types of immune receptors that recognize diverse pathogen molecules and activate the innate immune system. The intracellular immune receptor family of nucleotide-binding domain leucine-rich repeat–containing proteins (NLRs) perceive translocated pathogen effector proteins and execute a robust immune response, including programmed cell death. Many plant NLRs have functionally specialized to sense pathogen effectors (sensor NLRs) or to execute immune signalling (helper NLRs). Sub-functionalized NLRs form a network-type receptor system known as the NLR network. In this review, we highlight the concept of NLR networks, discussing how they are formed, activated, and regulated. Two main types of NLR networks have been described in plants: the ADR1/NRG1 network and the NRC network. In both networks, multiple helper NLRs function as signalling hubs for sensor NLRs and cell surface–localized immune receptors. Additionally, the networks are regulated at the transcriptional and posttranscriptional levels, as well as being modulated by other host proteins to ensure proper network activation and prevent autoimmunity. Plant pathogens in turn have converged on suppressing NLR networks, thereby facilitating infection and disease. Understanding the NLR immune system at the network level could inform future breeding programs by highlighting the appropriate genetic combinations of immunoreceptors to use while avoiding deleterious autoimmunity and suppression by pathogens.**

*Keywords: autoimmunity // host–pathogen interaction // immune receptor network // NLR integrated domain // nucleotide-binding domain and leucine-rich repeat–containing protein // pathogen effector*

*Short running head: activation and regulation of plant NLR networks*

## INTRODUCTION

Plants have an effective innate immune system, which can be activated by several types of immune receptors upon recognition of diverse pathogen molecules. These immune receptors are located either at the cell surface or in the nucleocytoplasm of plant cells (Lu and Tsuda, 2021). Cell-surface receptors can recognize pathogen-associated molecular patterns (PAMPs) or pathogen-secreted proteins, known as effectors, in the extracellular apoplastic space. In

addition to secreting pathogen effectors into the apoplast, many plant pathogens also translocate effectors into the host nucleocytoplasm, thereby altering host physiological processes and facilitating infection. In response, plants have evolved a diverse repertoire of intracellular immune receptors, predominantly belonging to the nucleotide-binding domain and leucine-rich repeat–containing protein (NLR) family, to recognize these translocated pathogen effectors (Jones *et al.*, 2016). Upon recognition of their cognate ligands by either cell-surface or NLR receptors, both classes of immune receptors activate common signalling pathways to trigger defence responses (Lu and Tsuda, 2021). The activation of some cell-surface receptors and most NLRs results in the induction of a localized programmed cell death response, known as the hypersensitive response (HR), in the infected cells. This cellular suicide machinery is thought to restrict the spread of pathogens from the infection site to neighbouring cells (Balint-Kurti, 2019). In this manner, plants can fight off invading pathogens and prevent disease while maintaining their health at the tissue level.

Our understanding of how NLRs are activated and induce downstream signalling and immunity has expanded tremendously in recent years. NLR immune receptors are important components of the innate immunity of plants and animals (Jones *et al.*, 2016). Plant NLRs share a multidomain architecture, characterized by a central nucleotide-binding domain shared with APAF-1, various R proteins, and CED-4 (NB-ARC) domain, and a C-terminal leucine-rich repeat (LRR) domain (Kourelis, Sakai, *et al.*, 2021). The C-terminal LRR domain is typically involved in effector recognition, while the NB-ARC domain mediates the intramolecular activation of the NLR protein presumably by exchanging adenosine diphosphate (ADP) for adenosine triphosphate (ATP) in the nucleotide-binding pocket. In addition to the conserved NB-ARC and LRR domains, most plant NLRs have a variable N-terminal domain, which can be used to broadly classify these proteins into four subgroups: Toll/Interleukin-1 Receptor (TIR)-type NLRs (TIR-NLR or TNL), coiled-coil (CC)-type NLRs (CC-NLR or CNL), RESISTANCE TO POWDERY MILDEW 8 (RPW8)-type CC-NLRs ($CC_R$-NLR or RNL), and the more recently described G10-type CC-NLRs ($CC_{G10}$-NLR) (Lee *et al.*, 2021). The N-terminal domains, TIR, CC, $CC_R$, and $CC_{G10}$, are generally thought of as the signalling domain that executes downstream immune responses upon ligand recognition.

Effector recognition by plant NLRs can be direct or indirect. The structural elucidation of both the direct and indirect recognition mechanisms has benefited from developments in biophysics and cryo-electron microscopy (cryo-EM). Both the *Arabidopsis thaliana* TIR-NLR RECOGNITION OF PERONOSPORA PARASITICA 1 (RPP1) and *Nicotiana benthamiana* TIR-NLR RECOGNITION OF XOPQ 1 (Roq1) are activated upon the direct binding of their cognate effector ligands (Ma *et al.*, 2020; Martin *et al.*, 2020). The RPP1 and Roq1 structures reveal that effector binding is mediated by two distinct interaction surfaces: the LRR and a post-LRR C-terminal jelly roll and Ig-like (C-JID) domain (Ma *et al.*, 2020; Martin *et al.*, 2020). Similar to direct effector binding by TIR-NLRs, the wheat (*Triticum monococcum*) CC-NLR Sr35 also directly binds its cognate effector AvrSr35, which is mediated by the LRR domain (Förderer *et al.*, 2022). By contrast, the structure of the Arabidopsis CC-NLR HOPZ-ACTIVATED RESISTANCE 1 (ZAR1) reveals how indirect recognition can function (Wang, Wang, *et al.*, 2019; Wang, Hu, *et al.*, 2019). ZAR1 indirectly recognizes multiple bacterial

effectors through host receptor-like cytoplasmic kinases (RLCKs). One such partner RLCK is RESISTANCE-RELATED KINASE 1 (RKS1), which interacts with the ZAR1 LRR domain and recruits RLCK PBS1-LIKE PROTEIN 2 (PBL2) upon its uridylylation by the *Xanthomonas campestris* pv. *campestris* (*Xcc*) effector AvrAC (Wang *et al.*, 2015). Finally, as an additional mode of recognition, the 'integrated-decoy' model was proposed based on functional analyses of several unusual NLRs that carry noncanonical integrated domains (IDs) required for effector perception (Césari, Bernoux, *et al.*, 2014). These NLRs are referred to as 'NLR-IDs'. These receptors use IDs as a bait to directly or indirectly recognize effectors. Given that these effectors appear to exert their virulence activity by targeting host proteins containing the same domains as the IDs, NLR-IDs may represent fusions of effector target genes with NLR genes (Białas *et al.*, 2017).

Upon effector recognition, activated NLRs form a high-order 'resistosome' complex. The first example of a resistosome was revealed by elucidating the structure of the Arabidopsis CC-NLR ZAR1 (Wang, Hu, *et al.*, 2019). The ZAR1–RKS1 complex associates with uridylylated PBL2 ($PBL2^{UMP}$, a form of PBL2 modulated by the *Xcc* effector AvrAC), inducing a conformational change in monomeric ZAR1, leading to the replacement of ADP by ATP or deoxyadenosine triphosphate (dATP) in the ZAR1 NB-ARC domain *in vitro* (Wang, Wang, *et al.*, 2019; Wang, Hu, *et al.*, 2019). This ADP–ATP switch is required for the oligomerization of activated ZAR1 monomers into a pentameric resistosome structure. In the activated ZAR1–RKS1–$PBL2^{UMP}$ resistosome structure, the first N-terminal α helix (α1 helix) of the CC domain of the ZAR1 monomers is exposed and form a funnel-shaped structure (Wang, Hu, *et al.*, 2019). The exposed funnel on the ZAR1 resistosome is thought to insert into the plasma membrane to form a pore (Wang, Hu, *et al.*, 2019). As such, the ZAR1 resistosome functions as a calcium ion ($Ca^{2+}$) channel on the plasma membrane, which induces $Ca^{2+}$ influx and subsequent hypersensitive cell death (Bi *et al.*, 2021). Supporting this model, substitutions in the ZAR1 α1 helix, impairing $Ca^{2+}$ influx, lead to the loss of the hypersensitive cell-death response and immunity to *Xcc*, although they do not affect the formation of the ZAR1 resistosome *in vivo* (Wang, Hu, *et al.*, 2019; Hu *et al.*, 2020; Bi *et al.*, 2021). Additionally, the CC-NLR Sr35 also forms a similar pentameric resistosome structure which also acts as a $Ca^{2+}$ channel (Förderer *et al.*, 2022).

The cryo-EM structures of activated RPP1 and Roq1 reveal two examples of tetrameric resistosomes formed by TIR-NLRs (Ma *et al.*, 2020; Martin *et al.*, 2020). The RPP1 resistosome is bound by ADP, although it might be that the switch of ADP for ATP is crucial for oligomerization (Ma *et al.*, 2020). Activated RPP1 and Roq1 oligomerize and their N-terminal TIR domains form two active centres for $NAD^+$ cleaving activity (Ma *et al.*, 2020; Martin *et al.*, 2020). The enzymatic activity of these TIR domains results in the release of a variant of cyclic-ADP-ribose (v-cADPR) and this enzymatic activity is required for the induction of hypersensitive cell death (Horsefield *et al.*, 2019; Wan *et al.*, 2019). In addition, plant TIR proteins appear to form a distinct structure in which the TIR domain displays 2′,3′-cAMP/cGMP synthetase activity via the hydrolysis of RNA/DNA, and this catalytic activity is also required for the induction of hypersensitive cell death (Yu *et al.*, 2022). The N-terminal

domains on the NLR resistosomes therefore directly mediate immune responses in distinct ways.

Thirty years of research on cloning plant disease resistance (*R*) genes has led to the identification of hundreds of *R* genes, which generally encode plant immune receptors (Kourelis and van der Hoorn, 2018; Ngou, Ding, *et al.*, 2022). Furthermore, as discussed above, the remarkable recent progress in plant NLR structural biology has dramatically advanced our understanding of how plant NLRs function at the molecular level. However, beyond the function of individual NLRs, a picture is now emerging in which intricate receptor network systems require multiple NLRs to function together to recognize diverse pathogen effectors and trigger immune signalling (Ngou, Jones, *et al.*, 2022). In this review, we discuss our current understanding of how NLR immune receptor networks form, become activated, and are regulated in plant immunity.

## NLR NETWORKS CONSIST OF SENSORS AND HELPERS

### NLR networks comprise sensor and helper NLRs

The conceptual basis of plant–microbe interactions was initially defined by the influential gene-for-gene model proposed by the plant pathologist Harold Flor (Flor, 1971). In this model, an *R* gene from the host plant evolves alongside a specific avirulence (*AVR*) gene from the pathogen. On a biochemical level, the gene-for-gene model dictates that plant NLR immune receptors (encoded by *R* genes) can recognize pathogen effector ligands, either directly or indirectly, and trigger an immune response as a single NLR unit. This means that plant NLRs are receptors that have both sensing and signalling functions, and are therefore referred to as 'singleton NLRs' (Adachi, Derevnina, *et al.*, 2019). The best described singleton NLR is ZAR1, since its sensing and signalling functions are described at the structural level. ZAR1 both recognizes its cognate effectors and executes immune signalling through homo-oligomerization and complex formation without relying on other NLRs (Wang, Wang, *et al.*, 2019; Wang, Hu, *et al.*, 2019).

In addition to the singleton NLRs, it is now clear that many plant NLRs have functionally specialized to either sense pathogen effectors (sensor NLRs) or execute immune signalling (helper NLRs, also known as executor NLRs). Sensor NLRs can recognize pathogen effectors either directly or indirectly by recognizing the modification of host target proteins, but they require helper NLRs to induce downstream immune signalling. Sensor and helper NLRs often work in pairs; for instance, The rice (*Oryza sativa*) CC-NLRs 'Sasanishiki' *RESISTANCE GENE ANALOG 5* (*SasRGA5*) and *PYRICULARIA ORYZAE RESISTANCE K-1* (*Pik-1*) CC-NLRs are sensor NLRs that are genetically linked to the CC-NLR genes, *SasRGA4* and *Pik-2*, respectively (Okuyama *et al.*, 2011; Ashikawa *et al.*, 2008). RGA4 and Pik-2 function as helper NLRs that form a heterocomplex with the corresponding sensor NLRs to trigger immune signalling (Césari, Kanzaki, *et al.*, 2014; Zdrzałek *et al.*, 2020).

In other cases, however, helper NLRs can function as signalling nodes for multiple sensor NLRs (Adachi, Derevnina, *et al.*, 2019). In this receptor system, many NLRs form a complex network architecture−an NLR network−beyond the one-to-one relationship of NLR pairs. As discussed in Adachi and Kamoun (2022), the potential benefits of NLR networks are evolvability and redundancy. For example, functional specialization of NLR receptors into sensors and helpers may allow sensor NLRs to diversify by diversifying selection, accumulating mutations or acquiring novel domain to recognize fast-evolving pathogen effectors. Helper NLRs instead maintain the ability to induce immune responses as signalling hubs, experiencing limited expansion and purifying selection. Redundancy in helper NLRs can allow the immune system to be more resilient from the suppression of central signalling nodes by pathogen effectors. The diversification and resilience in NLR receptor networks are distinct properties from other plant signalling networks and have presumably occurred in evolutionary arms-races with fast-evolving pathogen effectors which are not only perceived as signal inputs but also evolved to act as suppressors of these NLR networks (Katagiri, 2018; Adachi and Kamoun, 2022). Hereafter, we review examples of NLR immune receptor networks.

## The NRC network is an expanded helper/sensor clade in the Asterids

The NLR network model was first proposed upon the realization that a phylogenetic subclade of NLRs in the Solanaceae act as helper NLRs, which are differentially and redundantly required for the function of sensor NLRs (Wu *et al.*, 2017). In this network, CC-NLRs known as NLR-REQUIRED FOR CELL DEATH (NRC) proteins function as helper NLRs for multiple sensor NLRs to mediate immune responses (**Figure 1**); therefore, this immune receptor network is referred to as the NRC network. In the solanaceous model plant *Nicotiana benthamiana*, *NRC2*, *NRC3*, and *NRC4* act redundantly and with different specificities as helper NLRs for many sensor NLRs (Wu *et al.*, 2017; Witek *et al.*, 2021; Lin *et al.*, 2021); for example, the sensor NLR Rpi-blb2 specifically activates hypersensitive cell death and disease resistance to the oomycete potato blight pathogen *Phytophthora infestans* through *NRC4*, while the sensor NLR Prf-mediated response is dependent on *NRC2* and *NRC3* (Wu *et al.*, 2017; Wu *et al.*, 2016). All three helper NLRs redundantly contribute to sensor NLR Rx-mediated immunity against *potato virus X* (PVX) (Wu *et al.*, 2017).

Although NRC paralogs and NRC-dependent sensor NLRs are genetically unlinked and dispersed throughout the genomes of solanaceous plant species, they form a phylogenetically well-supported clade (NRC-helper clade) (Wu *et al.*, 2017). Interestingly, the NRC-helper clade phylogenetically clusters with a hugely expanded CC-NLR clade (NRC-sensor clade), which includes many sensor NLRs encoded by *R* genes from different solanaceous plant species. This phylogenetic relationship between helper and sensor NLRs in the NRC network indicates a common origin. Indeed, outside of the Asterid lineages, sugar beet (*Beta vulgaris*)—belonging to the Caryophyllales—has one NRC helper and two NRC sensors, which are genetically linked (Wu *et al.*, 2017). The NRC network components therefore presumably emerged as a sensor–helper gene cluster about 100 million years ago, before the Asterids and Caryophyllales linages split. Subsequently, this sensor–helper gene cluster massively expanded into the current NRC network through gene duplication and diversification in the Solanaceae

and several other Asterids; in some species, as much as 50% of all NLRs belong to this superclade of NRCs and their *R* sensors.

How do helper NRCs activate immune signalling? One clue is provided by the fact that the first 29 amino acids of NRC4 are sufficient to trigger a hypersensitive response (Adachi, Contreras, *et al.*, 2019). Notably, the N-termini of helper NRCs show a high sequence similarity to the N-terminal α1 helix of ZAR1, which forms the funnel and creates a pore at the plasma membrane for $Ca^{2+}$ influx. The consensus sequence motif for this N-terminus—MADAxVSFxVxKLxxLLxxEx— is called the 'MADA motif'. The MADA motif is present in about 20% of all CC-NLRs across flowering plant species, but it has degenerated in sensor CC-NLRs (Adachi, Contreras, *et al.*, 2019). The MADA motif of NRC4 can be functionally replaced by the N-terminal sequence of multiple other MADA-type CC-NLRs from both dicots and monocots (Adachi, Contreras, *et al.*, 2019). As in ZAR1, mutations of some hydrophobic residues in the NRC MADA motif ($NRC2^{L17E}$, $NRC3^{L21E}$, $NRC4^{L9E}$, $NRC4^{L13E}$, $NRC4^{L17E}$, $NRC4^{L9E/V10E/L14E}$) impair cell death activity (Adachi, Contreras, *et al.*, 2019; Kourelis, Contreras, *et al.*, 2021). Unlike in ZAR1, however, the E11A mutation in the MADA motif of NRC4 does not lead to loss of the hypersensitive cell-death response (Adachi, Contreras, *et al.*, 2019). Additionally, similar mutations of charged residues in the predicted α1 helix of Sr35 also do not abolish hypersensitive cell-death induction, while mutating hydrophobic residues ($Sr35^{L15E/L19E}$) does result in loss of hypersensitive cell-death induction (Förderer *et al.*, 2022). Therefore, hydrophobic residues in the ZAR1 MADA/α1 helix are likely involved in pore formation, while the negatively charged residue E11 could be essential for ZAR1 $Ca^{2+}$ channel activity, but not necessarily for other CC-NLRs (Wang, Hu, *et al.*, 2019; Hu *et al.*, 2020; Bi *et al.*, 2021; Adachi, Contreras, *et al.*, 2019; Förderer *et al.*, 2022). The helper NRCs may therefore function like ZAR1, forming a resistosome upon activation with a N-terminal funnel structure to make a pore on the plasma membrane (**Figure 1**); however, it remains unknown whether, like ZAR1, helper NRCs function as $Ca^{2+}$ channels.

## The ADR1/NRG1 network mediates TIR-NLR signalling

Another well-characterized NLR network is formed by the N REQUIREMENT GENE 1 (NRG1) and ACTIVATED DISEASE RESISTANCE 1 (ADR1) subfamilies of $CC_R$-type helper NLRs (**Figure 1**). $CC_R$-NLRs are required as helper NLRs for TIR-NLR-mediated immunity. Since the $CC_R$-NLR subfamily is found in the genomes of most flowering plant species and is smaller than the CC-NLR and TIR-NLR subfamilies, the $CC_R$-NLRs are considered to be conserved throughout angiosperm evolution as helper NLRs for sensor NLRs (Shao *et al.*, 2016; Baggs *et al.*, 2020; Liu *et al.*, 2021). Interestingly, the copy-number variation of the TIR-NLR and $CC_R$-NLR genes (primarily *NRG1*) is tightly correlated, reflecting an evolutionary association among these NLR subfamilies (Liu *et al.*, 2021). The TIR-NLR and NRG1 $CC_R$-NLR subfamily lineages have been lost in the monocots, although most monocot species do possess ADR1 subfamily $CC_R$-NLRs. This suggests that ADR1 is a helper subfamily not only for TIR-NLRs, but also for other types of sensors.

Arabidopsis possesses five full-length $CC_R$-NLR helpers, two NRG1 paralogs (*NRG1.1* and *NRG1.2*, also known as *NRG1A* and *NRG1B*, respectively), and three ADR1 paralogs (*ADR1*, *ADR1-L1*, and *ADR1-L2*). Saile *et al.* (2020) recently characterized the genetic requirement for *ADR1* and *NRG1* in immunity by comparing the phenotypes of *adr1 adr1-L1 adr1-L2* triple mutant, *nrg1.1 nrg1.2* double mutant, and the *nrg1.1 nrg1.2 adr1 adr1-L1 adr1-L2* '*helperless*' pentuple mutant lacking all full-length $CC_R$-NLR helpers. This comparison revealed that Arabidopsis *ADR1* genes are required for full resistance mediated by the TIR-NLRs RRS1/RPS4, RPP2, and RPP4 (Saile *et al.*, 2020). *NRG1* genes can partially substitute for *ADR1* function in resistance mediated by RRS1/RPS4 and RPP2; hence, the *helperless* mutant has a more susceptible phenotype than the *adr1* triple mutant (Saile *et al.*, 2020). In addition, the RRS1/RPS4-triggered hypersensitive cell-death response requires only NRG1s (Saile *et al.*, 2020). This reveals an unequal genetic redundancy between the *ADR1* and *NRG1* genes for TIR-NLR-mediated resistance and hypersensitive cell death. The CC-NLRs do not appear to require *ADR1* or *NRG1* for the activation of hypersensitive cell death and immunity, as the singleton CC-NLRs ZAR1 and RESISTANCE TO P. SYRINGAE PV MACULICOLA 1 (RPM1) do not require *ADR1* or *NRG1* (Saile *et al.*, 2020).

An elicitor-independent autoactive mutant of NRG1.1, $NRG1.1^{D485V}$, forms higher-order complexes and associates with the plasma membrane when it is expressed in *N. benthamiana*, while wild-type NRG1.1 does not (Jacob *et al.*, 2021). Furthermore, ADR1s can form self-associated complexes and localize to the plasma membrane through the interaction of their N-terminal $CC_R$ domain and the anionic lipid phosphatidylinositol-4-phosphate of the plant plasma membrane (Saile *et al.*, 2021). Interestingly, the N-terminal $CC_R$ domain of NRG1.1 is composed of a four-helical bundle structure like the ZAR1 CC domain (Jacob *et al.*, 2021), which implies that helper NRG1 and ADR1 form a ZAR1-type resistosome to execute immune signalling. Although the N-terminus of NRG1 and ADR1 do not share a similar sequence pattern to the ZAR1 MADA/α1 helix, their N-termini carry negatively charged residues similar to ZAR1, which are required for $Ca^{2+}$ influx and the initiation of cell death (Jacob *et al.*, 2021; Sun *et al.*, 2021). The activated helper NRG1 and ADR1 may therefore function like the ZAR1 resistosome on the plasma membrane, mediating the hypersensitive cell-death response by $Ca^{2+}$ influx (**Figure 1**).

## ACTIVATION OF NLR IMMUNE RECEPTOR NETWORKS

### Sensor NLRs activate helper NLRs

In the ADR1/NRG1 network, sensor TIR-NLRs form an enzymatically active resistosome upon effector recognition, and this enzymatic activity is required to induce the hypersensitive cell-death response and immunity (**Figure 1**). In addition to this catalytic activity, the hypersensitive cell-death response triggered by TIR-NLRs depends on lipase-like proteins belonging to the ENHANCED DISEASE SUSCEPTIBILITY 1 (EDS1) family (Lapin *et al.*, 2019; Gantner *et al.*, 2019). This family contains homologs of EDS1, SENESCENCE-ASSOCIATED GENE 101 (SAG101), and PHYTOALEXIN-DEFICIENT 4 (PAD4) (Lapin *et al.*, 2020). EDS1 forms mutually exclusive dimers with either SAG101 or PAD4 (Wagner

*et al.*, 2013), which are required for NRG1- or ADR1-mediated immunity, respectively (Sun *et al.*, 2021) (**Figure 1**). EDS1–SAG101–NRG1 and EDS1–PAD4–ADR1 physically associate during some stages of the immune signal transduction, but the hypersensitive cell death mediated by autoactive mutants of NRG1 does not require *EDS1* (Sun *et al.*, 2021; Jacob *et al.*, 2021).

Because *EDS1* is not required for the production of v-cADPR mediated by TIR proteins *in planta* (Wan *et al.*, 2019), it was proposed that v-cADPR may activate the EDS1–SAG101–NRG1 and/or EDS1–PAD4–ADR1 modules, in turn inducing the formation of the NRG1 and ADR1 resistosomes, which function as $Ca^{2+}$-permeable nonselective cation channels (**Figure 1**) (Saur *et al.*, 2021). TIR-NLRs thus indirectly induce a similar immune response to the CC-NLRs. The likely structure of plant TIR-domain produced v-cADPR was recently identified as 2'cADPR (independently identified and named 1′–2′ glycocyclic ADPR [gcADPR]) (Manik *et al.*, 2022; Leavitt *et al.*, 2022). 2'cADPR may serve as an intermediate in the synthesis of novel nucleosides associated with plant immunity (Manik *et al.*, 2022). Indeed, Huang *et al.*, (2022) show that TIR-NLRs catalyse the production of 2'-(5"-phosphoribosyl)-5'-adenosine mono-/di-phosphate (pRib-AMP/ADP), and that EDS1-PAD4 act as a receptor complex for pRib-AMP/ADP. Binding of pRib-ADP to EDS1-PAD4 results in a conformational change in the PAD4 C-terminal domain, thereby promoting interaction with ADR1s (Huang *et al.*, 2022). pRib-AMP can be directly derived from 2'cADPR by cleavage of its pyrophosphate bond (Manik *et al.*, 2022). EDS1-SAG101, instead, acts as a receptor complex for other TIR-catalysed second messengers, ADP-ribosylated ATP (ADPr-ATP) and ADPr-ADPR (di-ADPR), that induce EDS1-SAG101 association with NRG1.1 but not with ADR1-L1 (Jia *et al.*, 2022). These second messenger interactions with the EDS1-PAD4 and EDS1-SAG101 complexes then likely promote ADR1 and NRG1 resistosome formation and $Ca^{2+}$ channel activity. Finally, aside from their $NAD^+$ cleaving activity, the TIR domain of some TIR-NLRs displays 2′,3′-cAMP/cGMP synthetase activity via the hydrolysis of RNA/DNA (Yu *et al.*, 2022). While this activity is also required for the induction of hypersensitive cell-death, it is unclear which pathways these signalling molecules activate.

The mechanisms by which sensor NLRs activate helper NLRs in the NRC networks are the subject of ongoing investigation. A primary technical barrier for *in planta* biochemical and cell biological analyses of activated NRC network components is the cell death response elicited upon their activation. Contreras *et al.*, (2022) and Ahn *et al.*, (2022) took advantage of an NRC2 MADA motif mutant ($NRC2^{L9E/L13E/L17E}$) to abolish the cell death response without affecting resistosome assembly and plasma membrane localization, similar to what was previously shown for the MADA-type CC-NLRs ZAR1, NRC4 and Sr35 (Hu *et al.*, 2020; Duggan *et al.*, 2021; Förderer *et al.*, 2022). NRC2 oligomerization is induced upon effector-recognition by sensor NLRs Rx, Bs2, Rpi-amr1 or Rpi-amr3, resulting in molecular complexes in the ~720 to 1048 kDa range (Contreras *et al.*, 2022; Ahn *et al.*, 2022). Interestingly, the activated sensor NLRs do not appear to be incorporated in the NRC2 higher-order complex (Contreras *et al.*, 2022; Ahn *et al.*, 2022). Instead, the activated sensor NLRs are proposed to trigger homo-oligomerization of helper NRCs by an activation-and-release mechanism (**Figure 1**) (Contreras *et al.*, 2022; Ahn *et al.*, 2022). This activation also results in a subcellular relocalization of

helper NRCs. The helper NRC4, for example, localizes to the plasma membrane around the *P. infestans* invasion site where the effectors are secreted, and activation of NRC4 by the sensor Rpi-blb2, either by *P. infestans* secreting the Rpi-blb2 ligand AVR-blb2 or by co-expression of AVR-blb2, results in a punctate distribution of NRC4 (Duggan *et al.*, 2021). Similarly, activation of NRC2 by the sensor Rx, either upon PVX infection or by co-expressing the PVX coat protein ligand of Rx, results in the subcellular relocalization of NRC2 to plasma membrane localized puncta (Contreras *et al.*, 2022). In contrast, upon activation, the sensor NLR Rx does not form plasma membrane-associated puncta and remains cytosolic, further supporting an activation-and-release mechanism for NRC activation (Contreras *et al.*, 2022). This activation-and-release mechanism is distinct from the activation mechanism of the mammalian paired NLRs NLR neuronal apoptosis inhibitory protein (NAIP)/NOD-like receptor containing a caspase activating and recruitment domain 4 (NLRC4), which form a hetero-complex upon ligand perception (Zhang *et al.*, 2015; Tenthorey *et al.*, 2017). The exact mechanism by which sensor NLRs trigger oligomerization of helper NRCs, and whether this involves a transient interaction state or other components is currently not known.

**Autoactive sensor NLR mutants require helper NLRs in immune signalling**

In addition to effector recognition, amino acid insertions or substitutions in NLR proteins often result in autoimmunity. In Arabidopsis, some alleles of TIR-NLRs, such as *suppressor of npr1-1, constitutive 1* (*snc1*), *chilling-sensitive mutant 1* (*chs1*), and *chs3-2D*, result in an autoimmune phenotype (Zhang *et al.*, 2003; Bi *et al.*, 2011; Wang *et al.*, 2013). These alleles encode TIR-NLR proteins with gain-of-function mutations and the autoimmunity is dependent on *EDS1* (Zhang *et al.*, 2003; Wang *et al.*, 2013), and *NRG1* and *ADR1* with different strengths (Wu *et al.*, 2019; Castel *et al.*, 2019).

Furthermore, substitutions in the conserved MHD motif in the NB-ARC domain are commonly used to generate autoactive versions of NLRs. The MHD motif is located in a position binding to ADP in the ZAR1 structure, suggesting that the MHD motif-ADP interaction may have a role in intramolecular regulation of NLR proteins (Wang, Wang, *et al.*, 2019). The first example of such a MHD autoactive mutant was identified through the random mutagenesis of the NRC sensor Rx (Bendahmane *et al.*, 2002). The MHD mutant $Rx^{D460V}$ can induce the autoimmune cell-death response in the absence of the cognate pathogen ligand (Bendahmane *et al.*, 2002). Similarly, MHD mutants of helper NLRs, such as $NRC1^{D481V}$, $NRC2^{H480R}$, $NRC3^{D480V}$, $NRC4^{D478V}$, $NRG1.1^{D485V}$, and $ADR1\text{-}L2^{D484V}$, are also autoactive (Gabriëls *et al.*, 2007; Roberts *et al.*, 2013; Derevnina *et al.*, 2021; Jacob *et al.*, 2021). The autoactive $NRG1.1^{D485V}$ mutant forms a high-order complex *in vivo* (Jacob *et al.*, 2021), indicating that autoactive mutants could be used to analyse the biochemical and biophysical properties of sensor and helper NLRs in their activated state. Finally, autoactive sensor NLR mutants are useful tools for dissecting the NLR network specificity in the absence of a known pathogen ligand, as this autoactivity requires helper NLRs (Derevnina *et al.*, 2021).

**Cell-surface receptors can signal through NLR networks**

Finally, it is becoming increasingly clear that helper NLRs are also required for signalling mediated by cell-surface receptors. Cell-surface receptors are typically divided into two categories: the receptor-like kinases (RKs, also known as RLKs) and the receptor-like proteins (RPs, also known as RLPs) (Saijo *et al.*, 2018). Upon ligand recognition, many leucine-rich repeat (LRR)-RKs involved in immunity hetero-oligomerize with the LRR-RK co-receptor SOMATIC EMBRYOGENESIS RECEPTOR-LIKE KINASE 3 (SERK3, also known as BRI1-associated kinase 1 [BAK1]). Similarly, most LRR-RPs constitutively interact with the LRR-RK SUPPRESSOR OF BAK1-INTERACTING RECEPTOR-LIKE KINASE 1 (BIR1) 1 (SOBIR1), and hetero-oligomerize with SERK3 upon ligand binding. This in turn activates downstream RLCKs, which together relay the immune response.

In Arabidopsis, the EDS1–PAD4–ADR1 and, to a lesser extent, EDS1–SAG101–NRG1 modules are genetically required for a subset of the immune responses triggered by LRR-RPs and LRR-RKs (Pruitt *et al.*, 2021; Tian *et al.*, 2021) (**Figure 1**). For example, the LRR-RP RECEPTOR LIKE PROTEIN 23 (RLP23)-mediated immunity is dependent on SOBIR1, the RLCK-VII subfamily protein AVRPPHB SUSCEPTIBLE 1 (PBS)-LIKE 31 (PBL31), and the EDS1–PAD4–ADR1 module (Pruitt *et al.*, 2021). Protein–protein interaction analyses suggest that a cell-surface receptor complex including SOBIR1 and PBL31 associates with the EDS1–PAD4–ADR1 module in a ligand-independent manner (Pruitt *et al.*, 2021); therefore, upon ligand perception, LRR-RPs and LRR-RKs converge to activate NLR networks for a subset of their immune functions, possibly by protein kinase–mediated phosphorylation.

In addition to $CC_R$-type helper NLRs, a helper NLR in the NRC network is also involved in cell-surface receptor–mediated immune responses (**Figure 1**). In virus-induced gene silencing experiments, the helper NRC1 was previously implicated as a key component in the cell death mediated by the LRR-RPs Cf-4 (Gabriëls *et al.*, 2006), LeEIX2 (Gabriëls *et al.*, 2007), and Ve1 (Fradin *et al.*, 2009). Recently, the precise contribution of helper NRCs to the hypersensitive response mediated by Cf-4 has been validated using *N. benthamiana* CRISPR mutants of various NRCs (Kourelis, Contreras, *et al.*, 2021). This showed that the Cf-4-mediated hypersensitive cell death in response to the recognition of the *Cladosporium fulvum* (syn. *Passalora fulva*) effector Avr4 is lost in a *nrc2/3* CRISPR line, which could be restored by expressing *NRC3* (Kourelis, Contreras, *et al.*, 2021). Furthermore, a functional MADA motif in NRC3 is required for the hypersensitive cell-death response (Kourelis, Contreras, *et al.*, 2021). This implies the function of a signalling pathway downstream of the cell-surface receptor Cf-4, which activates NRC3 to trigger hypersensitive cell death, presumably through a ZAR1 resistosome–type mechanism. The RLCK-VII member Avr9/Cf-9 induced kinase 1 (ACIK1) was identified as a downstream component in Cf-4- and Cf-9-mediated hypersensitive cell death (Rowland *et al.*, 2005). Although the exact signalling components and the molecular mechanism by which the LRR-RP signals are transduced into the NRC network are currently unknown, the phosphorylation of helper NLRs by cell-surface receptor complexes such as RLCKs might be a key of helper activation. Indeed, the function of the TIR-NLR RRS1 is regulated by the phosphorylation of its C terminus by unknown protein kinase(s) (Guo *et al.*, 2020).

## NLR NETWORKS ARE REGULATED AT MULTIPLE LEVELS

### Transcriptional and posttranscriptional regulation of NLR networks

Plant NLRs are regulated at the transcriptional, posttranscriptional, and posttranslational levels to prevent the autoimmune fitness costs associated with the inappropriate activation of immune signalling. Our current understanding is that many NLR genes are expressed at a low basal level but are amplified upon the activation of immunity. For example, at the transcriptional level, many plant NLRs are subject to the premature termination of transcription mediated by the RNA-binding protein FPA, thereby regulating NLR protein levels (Parker *et al.*, 2021) (**Figure 2**).

The activation of cell-surface receptors leads to transcriptional upregulation of immune-related genes, including the NLRs, which is required for the induction of NLR-mediated hypersensitive cell death and immunity (Ngou *et al.*, 2021; Yuan *et al.*, 2021). Notably, the activation of the TIR-NLR pair RRS1/RPS4 by AvrRps4 (Ngou *et al.*, 2021), and the $CC_{G10}$-NLRs RPS2 and RPS5 by AvrRpt2 (Yuan *et al.*, 2021; Ngou *et al.*, 2021) and AvrPphB (Ngou *et al.*, 2021), requires the cell-surface receptor–mediated potentiation of signalling to trigger hypersensitive cell death.

At the posttranscriptional level, microRNA-mediated gene silencing has been shown to regulate NLR genes in NLR immune receptor networks (**Figure 2**); for example, miR-n033 regulates a large number of CC-NLR genes in Solanaceae species (Seo *et al.*, 2018). Most of the miR-n033 targets belong to the NRC sensor superfamily, including the *R* genes *Rpi-blb2*, *Mi-1.2*, and *Hero*. Additionally, homologs of the Solanaceous NRC-sensor *R* genes *Rx1*, *R2*, and *R1* are targeted by the microRNAs stu-miR6024, stu-miR482d, and nta-miR6025a, respectively (Li *et al.*, 2012). In addition to the NRC network, the ADR1/NRG1 network is also regulated by microRNAs; for example, in *N. benthamiana*, nta-miR6019 and nta-miR6020 lead to the cleavage of transcripts from the TIR-NLR *R* gene *N*, which encodes a sensor NLR in the ADR1/NRG1 network (Li *et al.*, 2012). In addition to the sensor NLRs, transcripts of the LRR-RP genes are also regulated by microRNAs. In tomato (*Solanum lycopersicum*) and pepper (*Capsicum annuum*), sly-miR6022, sly-miR6023, and miR-n026 target LRR-RPs belonging to the *Homologs of Cladosporium fulvum resistance 9* (*Hcr9*) clade, which are homologs of the *Cf-9 R* gene (Li *et al.*, 2012; Seo *et al.*, 2018). MicroRNAs presumably regulate the transcript levels of diverse sensor NLRs and RPs in NLR networks, thereby preventing autoimmunity.

### Non-NLR host proteins modulate NLR networks

In addition to transcriptional and posttranscriptional regulation, plant NLR networks are regulated at the posttranslational level. One such mechanism is the ubiquitin–proteasome degradation pathway (**Figure 2**). In the ADR1/NRG1 network, the Arabidopsis TIR-NLR SNC1 and the related proteins SIDEKICK SNC1 1/2/3 (SIKIC1/2/3) are targeted for protein degradation by SKP1–CULLIN1–F-box (SCF) E3 complex and the RING-type E3 ligases

MUTANT and SNC1-ENHANCING 1/2 (MUSE1/2) (Cheng *et al.*, 2011; Dong *et al.*, 2018). A recent study identified the novel E3 ligases *SNC1*-INFLUENCING PLANT E3 LIGASE REVERSE 1/2 (SNIPER1/2), which broadly regulate protein levels of sensor NLRs in Arabidopsis (Wu *et al.*, 2020). In *N. benthamiana*, the putative E3 ubiquitin ligase UBR7 modulates the protein levels of the TIR-NLR N (Zhang *et al.*, 2019).

In addition to the ubiquitin–proteasome-mediated degradation of NLR proteins, other host components can negatively regulate NLR network-mediated immunity; for instance, RPS2-mediated hypersensitive cell death is suppressed by salicylic acid (SA) treatment (Zavaliev *et al.*, 2020). SA is a plant hormone that induces systemic acquired resistance through the master regulator NONEXPRESSOR OF PATHOGENESIS-RELATED GENES 1 (NPR1). Interestingly, the suppression of hypersensitive cell death is dependent on NPR1 and is not limited to RPS2, but also to the TIR-NLRs RPS4 and RPP1 (Zavaliev *et al.*, 2020). Spatiotemporal analyses of phytohormone responses during RPS2-mediated immunity reveal that the SA-signalling pathway is activated in the areas surrounding cells displaying hypersensitive cell death (Betsuyaku *et al.*, 2018; Zavaliev *et al.*, 2020). These findings suggest that the NPR1 pathway may regulate the ADR1/NRG1 network–mediated hypersensitive cell-death response and contribute to the survival of cells adjacent to the pathogen infection site.

**The ADR1/NRG1 network and NRC network are modulated by NRG1C and NRCX, respectively**

There are also cases where NLR proteins regulate NLR networks in plants (**Figure 2**); for example, in the ADR1/NRG1 network, the Arabidopsis TIR-NLR RRS1 associates with its genetically linked NLR partner RPS4, thereby negatively regulating its autoactivity (Williams *et al.*, 2014). The overexpression of *RPS4* results in the constitutive activation of immunity in tobacco and Arabidopsis, which is suppressed in the presence of RRS1 (Huh *et al.*, 2017). In contrast to this one-to-one regulation, Wu *et al.*, (2022) recently showed that NRG1C negatively regulates TIR-NLR-mediated immunity and autoimmunity of its paralog NRG1.1. NRG1C is a member of the $CC_R$-NLR family, forming a gene cluster with helper NRG1.1 and NRG1.2; however, unlike NRG1.1 and NRG1.2, NRG1C lacks the N-terminal $CC_R$ domain and has a severely truncated NB-ARC domain, suggesting that NRG1C has lost its signalling activity and the capacity to induce hypersensitive cell death. Protein–protein interaction analyses indicate that the negative regulation by NRG1C likely occurs through its interference with the EDS1–SAG101 complex rather than an interaction with its helper NLR NRG1.1 (Wu *et al.*, 2022).

Similarly, the NRC network is also regulated by other NLRs; for example, in *N. benthamiana*, systemic gene silencing of *NRCX* markedly impairs plant growth, resulting in a dwarf phenotype (Adachi *et al.*, 2021). Although NRCX is a member of the helper NRC family with a CC-NLR domain architecture, it lacks certain canonical features of helper NRCs, such as a functional N-terminal MADA motif and the capacity to trigger autoimmunity. The alteration of *NRCX* expression modulates the hypersensitive cell death mediated by NRC2 and NRC3, but not by NRC4 (Adachi *et al.*, 2021), although the molecular mechanism underpinning the

NRCX antagonism remains unknown. An emerging picture is that NRG1C and NRCX are atypical homologs of helper NLRs, which lost their cell death executor activity and instead evolved to modulate the signalling hubs of NLR networks.

### Pathogen effectors have evolved to suppress NLR networks

Because helper NLRs in NLR networks are central hubs in mediating immune responses, diverse plant pathogens have evolved effectors to suppress them and thereby establish infection (**Figure 2**). Derevnina *et al.* (2021) conducted a screen using effector libraries from pathogens of solanaceous plant species and identified five effectors suppressing hypersensitive cell death induced by NRC network components in *N. benthamiana*. Three of these effectors, SPRYSEC10 and SPRYSEC34 from the potato cyst nematode *Globodera rostochiensis* and PITG-15278 from the oomycete *P. infestans*, suppress the hypersensitive cell-death response mediated by the sensor NLR Rpi-blb2 (Derevnina *et al.*, 2021). By contrast, two other effectors, *G. rostochiensis* SPRYSEC15 and *P. infestans* AVRcap1b, suppress the helper NRCs NRC2 and NRC3, thereby preventing the recognition of effectors mediated by NRC2/3-dependent sensor NLRs (Derevnina *et al.*, 2021). Interestingly, SPRYSEC15 directly binds to the NB-ARC domain of NRC2 and NRC3, suggesting this direct association interferes with the helper NLR function. AVRcap1b, however, appears to indirectly suppress NRC2- and NRC3-mediated hypersensitive cell death by binding to the host protein Target of Myb 1-like protein 9a (TOL9a). The suppression of NRC2 and NRC3 by AVRcap1b is compromised when *TOL9a* expression is silenced by RNA interference.

In addition to the suppression of the NRC network, pathogens have evolved effectors to suppress NRG1 and ADR1 subfamily $CC_R$-type helper NLRs; for example, a *Phytophthora capsici* effector PcAvh103 associates with the lipase domain of EDS1 and promotes the dissociation of the EDS1–PAD4 interaction (Li *et al.*, 2020), thereby disrupting the function of the EDS1–PAD4–ADR1 network. Similarly, the AvrA1 effector from the bacterial pathogen *Pseudomonas syringae* interacts with the soybean (*Glycine max*) homologs of EDS1, and requires these proteins to exert its virulence function (Wang *et al.*, 2014). Finally, the *P. syringae* effector HopAM1 is a TIR-domain containing effector which can suppress cell-surface-mediated signalling and TIR-NLR mediated signalling (Eastman *et al.*, 2022; Manik *et al.*, 2022). HopAM1 serves to produce a distinct version of v-cADPR recently identified as 3'cADPR (Manik *et al.*, 2022). It appears that 3'cADPR and its derivatives can manipulate ADR1/NRG1 network signalling (Manik *et al.*, 2022). This host $NAD^+$ manipulation could be a conserved virulence mechanism of *P. syringae*, considering that 93% of the primary phylogroup *P. syringae* strains have at least one NADase effector (Hulin and Ma, 2022). Taken together, these findings indicate that plant pathogens have evolved effectors targeting NLR networks at multiple levels, thereby enabling them to establish infection and cause disease in the host.

## FUTURE PERSPECTIVES

In this review, we highlight major advances in our understanding of NLR biology and NLR immune receptor networks in plants. Recent discoveries of NLR protein structures have provided mechanistic insights into how plant NLRs are activated and initiate downstream signalling; however, there are many unanswered questions about NLR function in NLR networks. Two main types of NLR networks have been described thus far: 1) $CC_R$-NLRs acting as helper NLRs downstream of TIR-NLRs, and 2) the NRC network of phylogenetically related CC-NLRs, which diversified into helper and sensor NLRs in Asterid species. In addition, it is now evident that both types of networks are also activated during the activation of some cell-surface receptors. How are helper NLRs activated by sensor NLRs and cell-surface receptors? What is the determinant of the sensor NLR/helper NLR or cell-surface receptor/helper NLR connections?

In addition to the activation of helper NLRs, how are genetically scattered NLR components co-ordinately regulated at the transcriptional level? How do host modulators appropriately regulate massively expanded NLR components to maintain homeostasis in NLR receptor networks? How are NLR networks activated and regulated at the single-cell level during pathogen infection? Further studies combining molecular evolution, biochemistry, biophysics, and cell biology approaches are required to fully understand the activation and regulation of network-forming NLRs.

Most molecular breeding programs incorporate *R* gene–encoded sensor NLRs to generate disease resistance crops. Given that a large number of sensor NLRs can function together with one or more helper NLRs, incorporating knowledge of NLR networks in future breeding programs could ensure that proper combinations of sensor/helper and cell-surface receptor/helper NLRs are achieved. A further understanding of how NLR network homeostasis is maintained will provide new insights into breeding disease-resistant crops without the potential fitness costs and yield loss. Furthermore, given that plant pathogens have evolved effectors to target NLR networks and overcome host immunity, the molecular engineering of NLRs and cell-surface receptors would make the NLR receptor network system more resilient by avoiding suppression by effectors. In conclusion, understanding NLR networks at multiple levels is required to inform future plant breeding programs.

## AVAILABILITY OF DATA

No new datasets were generated or analysed in this study.


## ACKNOWLEDGEMENTS

We thank Dr. Chika Tateda and Dr. Shigetaka Yasuda for commenting on the figures, and Dr. Daniel Lüdke for comments on the manuscript. We also thank Jennifer Mach from Plant Editors for editing the manuscript.

## FUNDING

H.A. was funded by the Japan Society for the Promotion of Science (JSPS) and Japan Science and Technology Agency (JST) Precursory Research for Embryonic Science and Technology

(PRESTO). J.K. was funded by BASF Plant Science. The funders had no role in the study design, data collection and analysis, decision to publish, or preparation of the manuscript.

## COMPETING INTERESTS

JK receives funding from industry utilizing NLR biology.

## AUTHOR CONTRIBUTIONS

Conceptualization: J.K. and H.A.; Funding acquisition: J.K. and H.A.; Roles/Writing - original draft: J.K. and H.A.; Writing - review & editing: J.K. and H.A.

## Figure legends

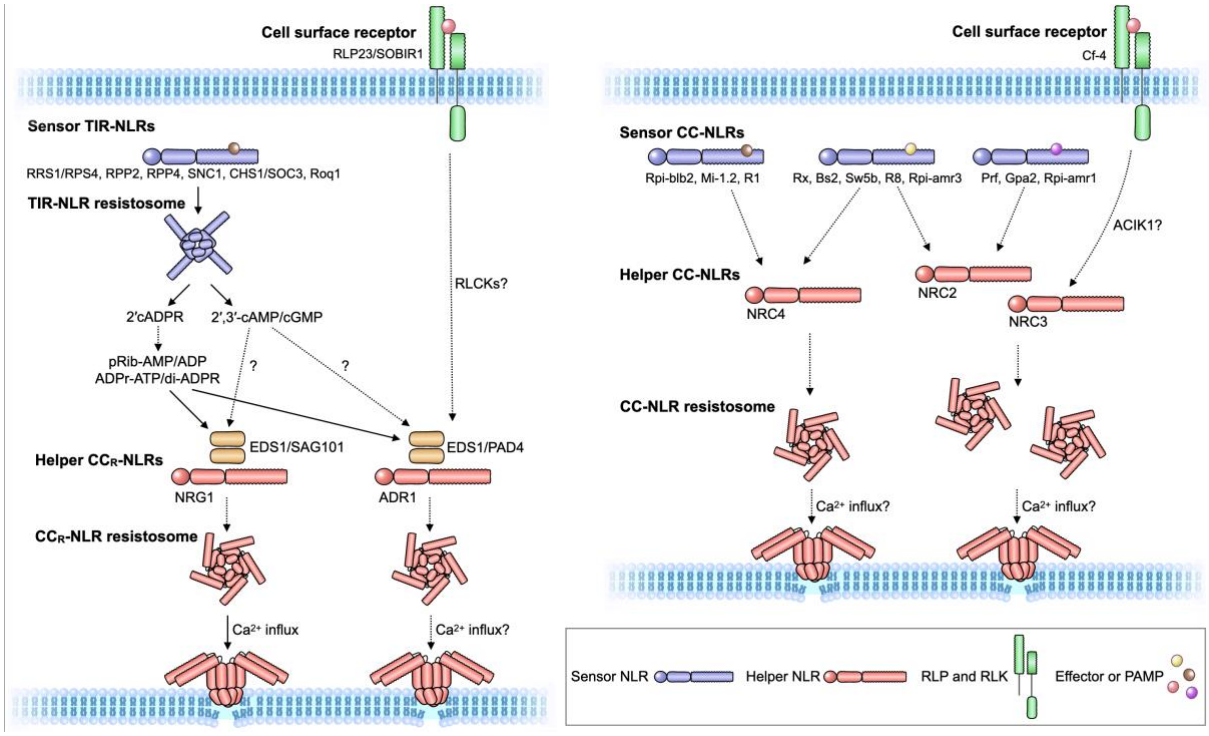


**Figure 1. Activation of NLR immune receptor networks.**
Pathogen ligand perception by cell-surface receptors and sensor NLRs leads to the activation of signalling pathways through helper NLRs. In the ADR1/NRG1 network (left), the EDS1–PAD4–ADR1 and EDS1–SAG101–NRG1 modules function downstream of the TIR-NLRs and some cell-surface RLP/RLKs. The activated TIR-NLR resistosome has enzymatic activity to produce v-cADPR and 2',3'-cAMP/cGMP, which likely activate the helper $CC_R$-NLRs through EDS1–PAD4 and EDS1–SAG101. Upon activation, ADR1 and NRG1 form a high-order complex ($CC_R$-NLR resistosome), acting as a $Ca^{2+}$ channel to induce immunity and hypersensitive cell death. In the NRC network (right), NRC helper subfamily members function downstream of phylogenetically linked sensor CC-NLRs and the cell-surface RLP(s). NRC2, NRC3, and NRC4 are functionally validated helpers for resistance gene–encoded sensors. Effector recognition by sensor NLRs results in NRC homo-oligomerization by an activation-and-release mechanism. The resulting homo-oligomerized NRC complex may function as a CC-NLR resistosome, inducing a $Ca^{2+}$ influx resulting in immunity and hypersensitive cell death. Solid lines indicate validated molecular mechanisms, while dashed lines indicate hypothetical models requiring further mechanistic elucidation.

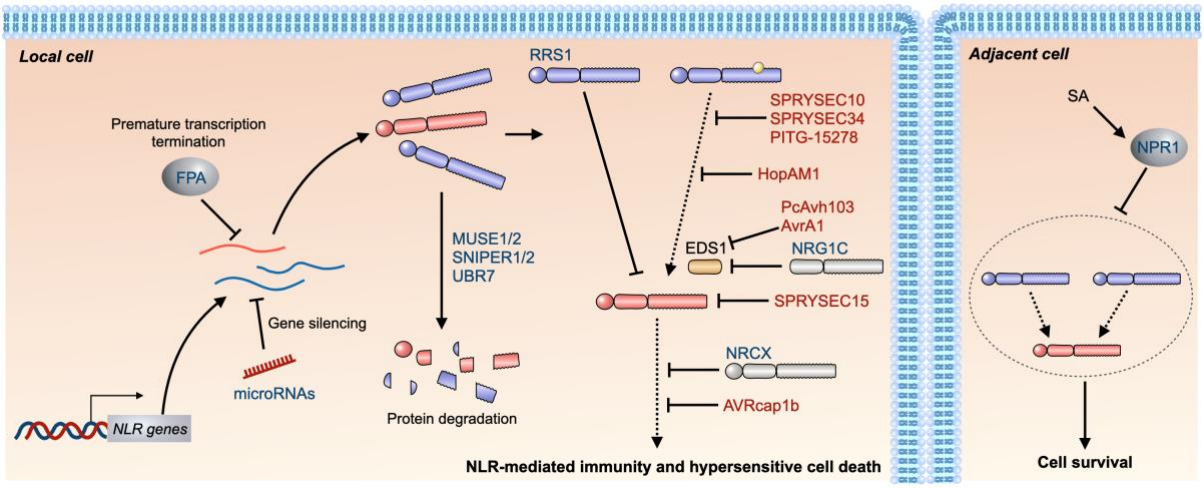


**Figure 2. Negative regulation of NLR immune receptor networks.**

NLR network components are tightly regulated at multiple levels. NLR transcripts are regulated by host premature transcription termination and microRNA-mediated silencing machineries. NLR networks are modulated by other host proteins, such as E3 ligases and NPR1, and by NLRs such as RRS1, NRG1C, and NRCX, which likely suppress the inappropriate activation of the networks. Plant pathogens have evolved effectors to target NLR networks or other key host proteins. Host regulators and pathogen effectors are shown in blue and red characters, respectively.

# REFERENCES


**Adachi, H., Contreras, M.P., Harant, A., et al.** (2019) An N-terminal motif in NLR immune receptors is functionally conserved across distantly related plant species. *eLife*, **8**, e49956.

**Adachi, H., Derevnina, L. and Kamoun, S.** (2019) NLR singletons, pairs, and networks: evolution, assembly, and regulation of the intracellular immunoreceptor circuitry of plants. *Curr. Opin. Plant Biol.*, **50**, 121–131.

**Adachi, H. and Kamoun, S.** (2022) NLR receptor networks in plants. *Essays Biochem.*, EBC20210075.

**Adachi, H., Sakai, T., Harant, A., Duggan, C., Bozkurt, T.O., Wu, C. and Kamoun, S.** (2021) An atypical NLR protein modulates the NRC immune receptor network. Available at: https://www.biorxiv.org/content/10.1101/2021.11.15.468391v1.

**Ahn, H.-K., Lin, X., Olave-Achury, A.C., Derevnina, L., Contreras, M.P., Kourelis, J., Kamoun, S. and Jones, J.D.G.** (2022) Effector-dependent activation and oligomerization of NRC helper NLRs by Rpi-amr3 and Rpi-amr1. Available at: https://www.biorxiv.org/content/10.1101/2022.04.25.489359v1.

**Ashikawa, I., Hayashi, N., Yamane, H., Kanamori, H., Wu, J., Matsumoto, T., Ono, K. and Yano, M.** (2008) Two adjacent nucleotide-binding site–leucine-rich repeat class genes are required to confer *Pikm*-specific rice blast resistance. *Genetics*, **180**, 2267–2276.

**Baggs, E.L., Monroe, J.G., Thanki, A.S., O'Grady, R., Schudoma, C., Haerty, W. and Krasileva, K.V.** (2020) Convergent loss of an EDS1/PAD4 signaling pathway in several plant lineages reveals coevolved components of plant immunity and drought response. *Plant Cell*, **32**, 2158–2177.

**Balint-Kurti, P.** (2019) The plant hypersensitive response: concepts, control and consequences. *Mol. Plant Pathol.*, **20**, 1163–1178.

**Bendahmane, A., Farnham, G., Moffett, P. and Baulcombe, D.C.** (2002) Constitutive gain-of-function mutants in a nucleotide binding site–leucine rich repeat protein encoded at the *Rx* locus of potato. *Plant J.*, **32**, 195–204.

**Betsuyaku, S., Katou, S., Takebayashi, Y., Sakakibara, H., Nomura, N. and Fukuda, H.** (2018) Salicylic acid and jasmonic acid pathways are activated in spatially different domains around the infection site during effector-triggered immunity in *Arabidopsis thaliana*. *Plant Cell Physiol.*, **59**, 8–16.

**Bi, D., Johnson, K., Huang, Y., Zhu, Z., Li, X. and Zhang, Y.** (2011) Mutations in an Atypical TIR-NB-LRR-LIM Resistance Protein Confer Autoimmunity. *Front. Plant Sci.*, **2**.

**Bi, G., Su, M., Li, N., et al.** (2021) The ZAR1 resistosome is a calcium-permeable channel triggering plant immune signaling. *Cell*, **184**, 3528-3541.e12.

**Białas, A., Zess, E.K., De la Concepcion, J.C., et al.** (2017) Lessons in effector and NLR biology of plant-microbe systems. *Mol. Plant. Microbe Interact.*, **31**, 34–45.

**Castel, B., Ngou, P.-M., Cevik, V., Redkar, A., Kim, D.-S., Yang, Y., Ding, P. and Jones, J.D.G.** (2019) Diverse NLR immune receptors activate defence via the RPW8-NLR NRG1. *New Phytol.*, **222**, 966–980.

**Césari, S., Bernoux, M., Moncuquet, P., Kroj, T. and Dodds, P.** (2014) A novel conserved mechanism for plant NLR protein pairs: the 'integrated decoy' hypothesis. *Front. Plant Sci.*, **5**. Available at: https://www.frontiersin.org/article/10.3389/fpls.2014.00606.

**Césari, S., Kanzaki, H., Fujiwara, T., et al.** (2014) The NB-LRR proteins RGA4 and RGA5 interact functionally and physically to confer disease resistance. *EMBO J.*, **33**, 1941–1959.

**Cheng, Y.T., Li, Y., Huang, S., Huang, Y., Dong, X., Zhang, Y. and Li, X.** (2011) Stability of plant immune-receptor resistance proteins is controlled by SKP1-CULLIN1-F-box (SCF)-mediated protein degradation. *Proc. Natl. Acad. Sci.*, **108**, 14694–14699.

**Contreras, M.P., Pai, H., Tumtas, Y., et al.** (2022) Sensor NLR immune proteins activate oligomerization of their NRC helper. Available at: https://www.biorxiv.org/content/10.1101/2022.04.25.489342v1.

**Derevnina, L., Contreras, M.P., Adachi, H., et al.** (2021) Plant pathogens convergently evolved to counteract redundant nodes of an NLR immune receptor network. *PLOS Biol.*, **19**, e3001136.

**Dong, O.X., Ao, K., Xu, F., et al.** (2018) Individual components of paired typical NLR immune receptors are regulated by distinct E3 ligases. *Nat. Plants*, **4**, 699–710.

**Duggan, C., Moratto, E., Savage, Z., et al.** (2021) Dynamic localization of a helper NLR at the plant–pathogen interface underpins pathogen recognition. *Proc. Natl. Acad. Sci.*, **118**, e2104997118.

**Eastman, S., Smith, T., Zaydman, M.A., et al.** (2022) A phytobacterial TIR domain effector manipulates $NAD^+$ to promote virulence. *New Phytol.*, **233**, 890–904.

**Flor, H.H.** (1971) Current status of the gene-For-gene concept. *Annu. Rev. Phytopathol.*, **9**, 275–296.

**Förderer, A., Li, E., Lawson, A., et al.** (2022) A wheat resistosome defines common principles of immune receptor channels. Available at: https://www.biorxiv.org/content/10.1101/2022.03.23.485489v1.

**Fradin, E.F., Zhang, Z., Ayala, J.C.J., Castroverde, C.D.M., Nazar, R.N., Robb, J., Liu, C.-M. and Thomma, B.P.H.J.** (2009) Genetic dissection of *Verticillium* wilt resistance mediated by tomato *Ve1*. *Plant Physiol.*, **150**, 320–332.

**Gabriëls, S.H.E.J., Takken, F.L.W., Vossen, J.H., et al.** (2006) cDNA-AFLP combined with functional analysis reveals novel genes involved in the hypersensitive response. *Mol. Plant. Microbe Interact.*, **19**, 567–576.

**Gabriëls, S.H.E.J., Vossen, J.H., Ekengren, S.K., et al.** (2007) An NB-LRR protein required for HR signalling mediated by both extra- and intracellular resistance proteins. *Plant J.*, **50**, 14–28.

**Gantner, J., Ordon, J., Kretschmer, C., Guerois, R. and Stuttmann, J.** (2019) An EDS1-SAG101 complex is essential for TNL-mediated immunity in *Nicotiana benthamiana*. *Plant Cell*, **31**, 2456–2474.

**Guo, H., Ahn, H.-K., Sklenar, J., Huang, J., Ma, Y., Ding, P., Menke, F.L.H. and Jones, J.D.G.** (2020) Phosphorylation-regulated activation of the *Arabidopsis* RRS1-R/RPS4 immune receptor complex reveals two distinct effector recognition mechanisms. *Cell Host Microbe*, **27**, 769-781.e6.

**Horsefield, S., Burdett, H., Zhang, X., et al.** (2019) $NAD^+$ cleavage activity by animal and plant TIR domains in cell death pathways. *Science*, **365**, 793–799.

**Hu, M., Qi, J., Bi, G. and Zhou, J.-M.** (2020) Bacterial effectors induce oligomerization of immune receptor ZAR1 *in vivo*. *Mol. Plant*, **13**, 793–801.

**Huang, S., Jia, A., Song, W., et al.** (2022) Identification and receptor mechanism of TIR-catalyzed small molecules in plant immunity. Available at: https://www.biorxiv.org/content/10.1101/2022.04.01.486681v1.

**Huh, S.U., Cevik, V., Ding, P., Duxbury, Z., Ma, Y., Tomlinson, L., Sarris, P.F. and Jones, J.D.G.** (2017) Protein-protein interactions in the RPS4/RRS1 immune receptor complex. *PLOS Pathog.*, **13**, e1006376.

**Hulin, M.T. and Ma, W.** (2022) Pangenomics facilitated with structural analysis reveals host $NAD^+$ manipulation as a major virulence activity of bacterial effectors. Available at: https://www.biorxiv.org/content/10.1101/2022.06.07.495176v2.

**Jacob, P., Kim, N.H., Wu, F., et al.** (2021) Plant "helper" immune receptors are $Ca^{2+}$-permeable nonselective cation channels. *Science*, **373**, 420–425.

**Jia, A., Huang, S., Song, W., et al.** (2022) TIR-catalyzed ADP-ribosylation reactions produce signaling molecules for plant immunity. Available at: https://www.biorxiv.org/content/10.1101/2022.05.02.490369v1.

**Jones, J.D.G., Vance, R.E. and Dangl, J.L.** (2016) Intracellular innate immune surveillance devices in plants and animals. *Science*, **354**, aaf6395.

**Katagiri, F.** (2018) Review: Plant immune signaling from a network perspective. *Plant Sci.*, **276**, 14–21.

**Kourelis, J., Contreras, M.P., Harant, A., Adachi, H., Derevnina, L., Wu, C.-H. and Kamoun, S.** (2021) The helper NLR immune protein NRC3 mediates the hypersensitive cell death caused by the cell-surface receptor Cf-4. Available at: https://www.biorxiv.org/content/10.1101/2021.09.28.461843v1.

**Kourelis, J. and Hoorn, R.A.L. van der** (2018) Defended to the nines: 25 years of resistance gene cloning identifies nine mechanisms for R protein function. *Plant Cell*, **30**, 285–299.

**Kourelis, J., Sakai, T., Adachi, H. and Kamoun, S.** (2021) RefPlantNLR is a comprehensive collection of experimentally validated plant disease resistance proteins from the NLR family. *PLOS Biol.*, **19**, e3001124.

**Lapin, D., Bhandari, D.D. and Parker, J.E.** (2020) Origins and immunity networking functions of EDS1 family proteins. *Annu. Rev. Phytopathol.*, **58**, 253–276.

**Lapin, D., Kovacova, V., Sun, X., et al.** (2019) A coevolved EDS1-SAG101-NRG1 module mediates cell death signaling by TIR-domain immune receptors. *Plant Cell*, **31**, 2430–2455.

**Leavitt, A., Yirmiya, E., Amitai, G., Lu, A., Garb, J., Morehouse, B.R., Hobbs, S.J., Kranzusch, P.J. and Sorek, R.** (2022) Viruses inhibit TIR gcADPR signaling to overcome bacterial defense. Available at: https://www.biorxiv.org/content/10.1101/2022.05.03.490397v1.

**Lee, H.-Y., Mang, H., Choi, E., Seo, Y.-E., Kim, M.-S., Oh, S., Kim, S.-B. and Choi, D.** (2021) Genome-wide functional analysis of hot pepper immune receptors reveals an autonomous NLR clade in seed plants. *New Phytol.*, **229**, 532–547.

**Li, F., Pignatta, D., Bendix, C., Brunkard, J.O., Cohn, M.M., Tung, J., Sun, H., Kumar, P. and Baker, B.** (2012) MicroRNA regulation of plant innate immune receptors. *Proc. Natl. Acad. Sci.*, **109**, 1790–1795.

**Li, Q., Wang, J., Bai, T., Zhang, Ming, Jia, Y., Shen, D., Zhang, Meixiang and Dou, D.** (2020) A *Phytophthora capsici* effector suppresses plant immunity via interaction with EDS1. *Mol. Plant Pathol.*, **21**, 502–511.

**Lin, X., Olave-Achury, A., Heal, R., et al.** (2021) Rpi-amr3 confers resistance to multiple *Phytophthora* species by recognizing a conserved RXLR effector. Available at: https://www.biorxiv.org/content/10.1101/2021.06.10.447899v1.

**Liu, Y., Zeng, Z., Zhang, Y.-M., et al.** (2021) An angiosperm *NLR* Atlas reveals that *NLR* gene reduction is associated with ecological specialization and signal transduction component deletion. *Mol. Plant*, **14**, 2015–2031.

**Lu, Y. and Tsuda, K.** (2021) Intimate association of PRR- and NLR-mediated signaling in plant immunity. *Mol. Plant. Microbe Interact.*, **34**, 3–14.

**Ma, S., Lapin, D., Liu, L., et al.** (2020) Direct pathogen-induced assembly of an NLR immune receptor complex to form a holoenzyme. *Science*, **370**, eabe3069.

**Manik, M.K., Shi, Y., Li, S., et al.** (2022) Chemical structures of cyclic ADP ribose (cADPR) isomers and the molecular basis of their production and signaling. Available at: https://www.biorxiv.org/content/10.1101/2022.05.07.491051v1.

**Martin, R., Qi, T., Zhang, H., Liu, F., King, M., Toth, C., Nogales, E. and Staskawicz, B.J.** (2020) Structure of the activated ROQ1 resistosome directly recognizing the pathogen effector XopQ. *Science*, **370**, eabd9993.

**Ngou, B.P.M., Ahn, H.-K., Ding, P. and Jones, J.D.G.** (2021) Mutual potentiation of plant immunity by cell-surface and intracellular receptors. *Nature*, 1–6.

**Ngou, B.P.M., Ding, P. and Jones, J.D.G.** (2022) Thirty years of resistance: zig-zag through the plant immune system. *Plant Cell*, koac041.

**Ngou, B.P.M., Jones, J.D.G. and Ding, P.** (2022) Plant immune networks. *Trends Plant Sci.*, **27**, 255–273.

**Okuyama, Y., Kanzaki, H., Abe, A., et al.** (2011) A multifaceted genomics approach allows the isolation of the rice *Pia*-blast resistance gene consisting of two adjacent NBS-LRR protein genes. *Plant J.*, **66**, 467–479.

**Parker, M.T., Knop, K., Zacharaki, V., et al.** (2021) Widespread premature transcription termination of *Arabidopsis thaliana* NLR genes by the spen protein FPA H. Yu, D. Weigel, C. Eunyoung, and B. C. Meyers, eds. *eLife*, **10**, e65537.

**Pruitt, R.N., Locci, F., Wanke, F., et al.** (2021) The EDS1–PAD4–ADR1 node mediates *Arabidopsis* pattern-triggered immunity. *Nature*, 1–5.

**Roberts, M., Tang, S., Stallmann, A., Dangl, J.L. and Bonardi, V.** (2013) Genetic requirements for signaling from an autoactive plant NB-LRR intracellular innate immune receptor. *PLOS Genet.*, **9**, e1003465.

**Rowland, O., Ludwig, A.A., Merrick, C.J., et al.** (2005) Functional analysis of *Avr9/Cf-9 rapidly elicited genes* identifies a protein kinase, ACIK1, that is essential for full Cf-9–dependent disease resistance in tomato. *Plant Cell*, **17**, 295–310.

**Saijo, Y., Loo, E.P. and Yasuda, S.** (2018) Pattern recognition receptors and signaling in plant–microbe interactions. *Plant J.*, **93**, 592–613.

**Saile, S.C., Ackermann, F.M., Sunil, S., et al.** (2021) Arabidopsis ADR1 helper NLR immune receptors localize and function at the plasma membrane in a phospholipid dependent manner. *New Phytol.*, **232**, 2440–2456.

**Saile, S.C., Jacob, P., Castel, B., Jubic, L.M., Salas-Gonzáles, I., Bäcker, M., Jones, J.D.G., Dangl, J.L. and Kasmi, F.E.** (2020) Two unequally redundant "helper" immune receptor families mediate *Arabidopsis thaliana* intracellular "sensor" immune receptor functions. *PLOS Biol.*, **18**, e3000783.

**Saur, I.M.L., Panstruga, R. and Schulze-Lefert, P.** (2021) NOD-like receptor-mediated plant immunity: from structure to cell death. *Nat. Rev. Immunol.*, **21**, 305–318.

**Seo, E., Kim, T., Park, J.H., Yeom, S.-I., Kim, S., Seo, M.-K., Shin, C. and Choi, D.** (2018) Genome-wide comparative analysis in Solanaceous species reveals evolution of microRNAs targeting defense genes in *Capsicum* spp. *DNA Res.*, **25**, 561–575.

**Shao, Z.-Q., Xue, J.-Y., Wu, P., Zhang, Y.-M., Wu, Y., Hang, Y.-Y., Wang, B. and Chen, J.-Q.** (2016) Large-scale analyses of angiosperm nucleotide-binding site-leucine-rich repeat genes reveal three anciently diverged classes with distinct evolutionary patterns. *Plant Physiol.*, **170**, 2095–2109.

**Sun, X., Lapin, D., Feehan, J.M., et al.** (2021) Pathogen effector recognition-dependent association of NRG1 with EDS1 and SAG101 in TNL receptor immunity. *Nat. Commun.*, **12**, 3335.

**Tenthorey, J.L., Haloupek, N., López-Blanco, J.R., et al.** (2017) The structural basis of flagellin detection by NAIP5: a strategy to limit pathogen immune evasion. *Science*, **358**, 888–893.

**Tian, H., Wu, Z., Chen, S., et al.** (2021) Activation of TIR signalling boosts pattern-triggered immunity. *Nature*, **598**, 500–503.

**Wagner, S., Stuttmann, J., Rietz, S., Guerois, R., Brunstein, E., Bautor, J., Niefind, K. and Parker, J.E.** (2013) Structural basis for signaling by exclusive EDS1 heteromeric complexes with SAG101 or PAD4 in plant innate immunity. *Cell Host Microbe*, **14**, 619–630.

**Wan, L., Essuman, K., Anderson, R.G., et al.** (2019) TIR domains of plant immune receptors are $NAD^+$-cleaving enzymes that promote cell death. *Science*, **365**, 799–803.

**Wang, G., Roux, B., Feng, F., et al.** (2015) The decoy substrate of a pathogen effector and a pseudokinase specify pathogen-induced modified-self recognition and immunity in plants. *Cell Host Microbe*, **18**, 285–295.

**Wang, J., Shine, M.B., Gao, Q.-M., Navarre, D., Jiang, W., Liu, C., Chen, Q., Hu, G. and Kachroo, A.** (2014) Enhanced disease susceptibility1 mediates pathogen resistance and virulence function of a bacterial effector in soybean. *Plant Physiol.*, **165**, 1269–1284.

**Wang, Jizong, Hu, M., Wang, Jia, et al.** (2019) Reconstitution and structure of a plant NLR resistosome conferring immunity. *Science*, **364**, eaav5870.

**Wang, Jizong, Wang, Jia, Hu, M., et al.** (2019) Ligand-triggered allosteric ADP release primes a plant NLR complex. *Science*, **364**, eaav5868.

**Wang, Y., Zhang, Y., Wang, Z., Zhang, X. and Yang, S.** (2013) A missense mutation in CHS1, a TIR-NB protein, induces chilling sensitivity in Arabidopsis. *Plant J.*, **75**, 553–565.

**Williams, S.J., Sohn, K.H., Wan, L., et al.** (2014) Structural basis for assembly and function of a heterodimeric plant immune receptor. *Science*, **344**, 299–303.

**Witek, K., Lin, X., Karki, H.S., et al.** (2021) A complex resistance locus in *Solanum americanum* recognizes a conserved *Phytophthora* effector. *Nat. Plants*, **7**, 198–208.

**Wu, C.-H., Abd-El-Haliem, A., Bozkurt, T.O., Belhaj, K., Terauchi, R., Vossen, J.H. and Kamoun, S.** (2017) NLR network mediates immunity to diverse plant pathogens. *Proc. Natl. Acad. Sci. U. S. A.*, **114**, 8113–8118.

**Wu, C.-H., Belhaj, K., Bozkurt, T.O., Birk, M.S. and Kamoun, S.** (2016) Helper NLR proteins NRC2a/b and

NRC3 but not NRC1 are required for Pto-mediated cell death and resistance in *Nicotiana benthamiana*. *New Phytol.*, **209**, 1344–1352.

**Wu, Z., Li, M., Dong, O.X., Xia, S., Liang, W., Bao, Y., Wasteneys, G. and Li, X.** (2019) Differential regulation of TNL-mediated immune signaling by redundant helper CNLs. *New Phytol.*, **222**, 938–953.

**Wu, Z., Tian, L., Liu, X., Huang, W., Zhang, Y. and Li, X.** (2022) The N-terminally truncated helper NLR *NRG1C* antagonizes immunity mediated by its full-length neighbors *NRG1A* and *NRG1B*. *Plant Cell*, **34**, 1621–1640.

**Wu, Z., Tong, M., Tian, L., Zhu, C., Liu, X., Zhang, Y. and Li, X.** (2020) Plant E3 ligases SNIPER1 and SNIPER2 broadly regulate the homeostasis of sensor NLR immune receptors. *EMBO J.*, **39**, e104915.

**Yu, D., Song, W., Tan, E.Y.J., et al.** (2022) TIR domains of plant immune receptors are 2′,3′-cAMP/cGMP synthetases mediating cell death. *Cell*, **185**, 2370-2386.e18.

**Yuan, M., Jiang, Z., Bi, G., et al.** (2021) Pattern-recognition receptors are required for NLR-mediated plant immunity. *Nature*, 1–5.

**Zavaliev, R., Mohan, R., Chen, T. and Dong, X.** (2020) Formation of NPR1 condensates promotes cell survival during the plant immune response. *Cell*, **182**, 1093-1108.e18.

**Zdrzałek, R., Kamoun, S., Terauchi, R., Saitoh, H. and Banfield, M.J.** (2020) The rice NLR pair Pikp-1/Pikp-2 initiates cell death through receptor cooperation rather than negative regulation. *PLOS ONE*, **15**, e0238616.

**Zhang, L., Chen, S., Ruan, J., et al.** (2015) Cryo-EM structure of the activated NAIP2-NLRC4 inflammasome reveals nucleated polymerization. *Science*, **350**, 404–409.

**Zhang, Y., Goritschnig, S., Dong, X. and Li, X.** (2003) A gain-of-function mutation in a plant disease resistance gene leads to constitutive activation of downstream signal transduction pathways in *suppressor of npr1-1, constitutive 1*. *Plant Cell*, **15**, 2636–2646.

**Zhang, Y., Song, G., Lal, N.K., et al.** (2019) TurboID-based proximity labeling reveals that UBR7 is a regulator of N NLR immune receptor-mediated immunity. *Nat. Commun.*, **10**, 3252.