



## Essential oils, chemical composition, and biological activities of *Eucalyptus oleosa* F. Muell. : A review

Souda Belaid<sup>1,2\*</sup>, Imen Chemlali<sup>1,2</sup>, Sonia Ben Rabeh<sup>1,2</sup>, Saousan Chamali<sup>2</sup>, Chokri Ben Romdhane<sup>3</sup>, Walid Elfalleh<sup>2</sup> & Ezzeddine Saadaoui<sup>3</sup>

<sup>1</sup>University of Gabes, Faculty of sciences of Gabes, Gabes 6072, Tunisia.

<sup>2</sup>University of Gabes, National Engineering School of Gabes, Energy, Water, Environment and Process Laboratory, (LR18ES35), Gabes 6072, Tunisia.

<sup>3</sup>University of Carthage, National Institute for Rural Engineering, Water and Forests (INRGREF), LGVRF 6011, Tunisia.

### Article info

#### Article history:

Received: 30 September 2023

Accepted: 15 November 2023

Keywords: Biological activities, Essential oil, yield, *Eucalyptus oleosa*, Composition.



Copyright©2023 JOASD

#### \*Corresponding author

saad\_ezz@yahoo.fr

**Conflict of Interest:** The authors declare no conflict of interest.

### Abstract

Many *Eucalyptus* species are growing in the border of oasis areas. *Eucalyptus* sp. are generally known for their richness in essential oils and their virtues and economic interests. However, the great taxonomic diversity affects the quantity and quality of these oils. This study is designed to summarize the chemical composition of *Eucalyptus oleosa* and their biological activities. The yield of essential oils in the leaves of this species varies from 0.45% to 6.7%. These oils contain many chemical compounds of which 1,8-cineole is the main component (15.31% - 89.4%) followed by  $\alpha$ -pinene (1%- 24.7%). *Eucalyptus oleosa* essential oils exhibited antioxidant, antibacterial, anti-fungal and insecticidal activities with high variability. This variability is associated to many factors such as subspecific diversity, geographical location, part of plant and essential oil's extraction method.

## 1. INTRODUCTION

*Eucalyptus* is a large genus of the Myrtaceae family (Grattapaglia et al., 2012) that includes 900 species and subspecies of which more than 300 species contain volatile essential oil in their leaves (Al-Snafi, 2017), and about twenty of these species are known for their richness in 1,8-cineole, used in pharmaceutical and cosmetic industries (Pino et al., 2002; Dhakad et al., 2018). Although *Eucalyptus* is widely grown in many countries all over the world (Booth, 2013), Australia is probably the only one where such a single group of plants dominate most of the landscape (Crisp et al., 2011). Worldwide production of *Eucalyptus* essentials oils is around 3000 tons, and the major producers are China, Spain, Portugal, South Africa, and Chile (Ghaffar et al., 2015). It has a long history of use in traditional medicine in the treatment of cold,

fever, flu, and general sickness (Salehi et al., 2019). In addition to its richness in essential oils, *Eucalyptus* is used for ornamentation, afforestation, and to obtain timber, gum, pulp and paper and it's known by its cosmetic and medicinal values (Saadaoui et al., 2017). Furthermore, Green synthesis of silver nanoparticles is possible to use the aqueous extract of *E. oleosa* as a green synthesis procedure without any catalyst, template, or surfactant. Colloidal gold and silver nanoparticles are synthesized with *Eucalyptus* leaf extract at non-photomediated conditions (Pourmortazavi et al., 2015; Pourmortazavi et al., 2017). In addition to these biological activities, *Eucalyptus* oils have ecological roles; they act as herbivore deterrents, alleviate ozone toxicity, and mitigate the temperature throughout the time of fires (Sawalha et al.,

2021). In food, the efficiency of *Eucalyptus* essential oils as a natural beverage preservative is verified; an effective and potent inhibitor of spoilage fungi and their *in vitro* antimicrobial effect is assessed against 17 food spoilage microorganisms (Boukhatem et al., 2020).

This study focused on *Eucalyptus oleosa*, a xeric species, resistant to environmental stresses including salinity and drought; and grows in very low rainfall (Merchant et al., 2006; Hobbs et al., 2009). The choice was made essentially for its high content of 1,8-cineole more than 80% (Jaymand et al., 2009) and its diversified biological activities. In terms of chemical composition, essential oils of *E. oleosa* were complex mixtures of substances generally terpenes and terpenoids (Marzoug et al., 2011). Considering the versatility of *Eucalyptus* essential oils in terms of bioactivities as well as their industrial importance, the purpose of this study is to provide the readers with the latest information concerning the essential oils yields, the chemical composition, and biological activities of *E. oleosa*.

## 2. YIELD OF *E. OLEOSA* ESSENTIAL OILS

The *Eucalyptus* essential oils were extracted by using different methods such as supercritical CO<sub>2</sub>, microwave and by solvents. Hydrodistillation is typically the most used method to obtain volatile compounds produced by plants (Richter and Schellenberg, 2007). A considerable variation in the yields of essential oils extracted from *E. oleosa* leaf has been detected and the values were ranging from 0.06 to 7 % (Elaissi et al., 2007). Previous studies have reported that the essential oils yield from plants collected in Iran was 6.7% (Ebadollahi et al., 2013; Rahimi-Nasrabadi et al., 2013), whereas others reported that the yield of *E. oleosa* essential oils varied from 2.31% to 3.2% collected from the same country (Jaymand et al., 2009; Ebadollahi et al., 2017). Relatively essential oils yield extracted from *E. oleosa* harvested in Tunisia was 4.90% (Marzoug et al., 2011) and similar results was detected with *E. oleosa* volatile oils from Australia that contained 4.60% of essential oils (Bignell et al., 1995). The detected values were more important than that reported in other species in Tunisia (*E. maideni*, *E. astrengens*, *E. cinerea*, *E. leucoxydon*, *E. lehmani*, *E. sideroxydon* and *E. bicostata*) in which the yields were ranging from 1.2% to 3% (Sebei et al., 2015). According to the literature, the yields of essential oils varies significantly between species of the genus *Eucalyptus* such as 0.29% in

*E. microtheca* (Hashemi-Moghaddam et al., 2013), 0.5% in *E. camaldulensis* (Ndiaye et al., 2018), 3.9% in *E. sargentii* (Fathi and Sefidkon, 2012), and 1.8% in *E. globulus* (Damjanović-Vratnica et al., 2011). Moreover, other investigation reported variable oil content (0.45%–1.12%) from different aerial parts of *E. oleosa* originating from Tunisia (Marzoug et al., 2011). Other *Eucalyptus* species analyzed from Morocco (*E. cinerea*, *E. baueriana*, *E. smithii*, *E. bridgesiana*, *E. microtheca*, *E. foecunda*, *E. propinqua* and *E. erythrocoris*) have similar oil yields ranging from 0.2% to 1.15% (Zrira et al., 2004). As previously stated, the observed variability not only might have been derived from harvest time, local, climatic, and seasonal factors but also it could be greatly depending upon the different parts of the plants extracted. Other study reported the effect of the extraction methods on the yield (Chamali et al., 2019).

## 3. CHEMICAL COMPOSITION

The *E. oleosa* essential oils were analyzed using GC-MS to identify their components. Generally, monoterpenes are the major components of *Eucalyptus* essential oils (Ohara et al., 2010). The essential oils composition of *E. oleosa* leaves showed by some studies that all of them contained 1,8-cineole, the highest content is (89.4%) (Jaymand et al., 2009) followed by (57.89%) (Safaei-Ghomi et al., 2009), (52.04%) (Bignell et al., 1995), (45.1%) (Rahimi-Nasrabadi et al., 2013) and (31.96%) (Marzoug et al., 2011), while some studies gave presented the lowest rate (15.31% and 22.94%) (Chamali et al., 2019; Ben Hassine et al., 2012). These data supported previous published results and confirmed that 1,8-cineole is the major one compound in leaf essential oils of *E. oleosa* and the most important volatile component in most of *Eucalyptus* species (Fadel et al., 1999; Vilela et al., 2009; Maghsoodlou et al., 2015; Vivekanandhan et al., 2020). Besides, other investigation confirmed that the main component of the essential oils of all parts of the *E. oleosa* (stems, adult leaves, immature flowers, and fruits) is 1,8-cineole (31.5%, 8.7%, 47.0% and 29.1%, respectively). For the other molecules there is a high diversity between plant tissues (Marzoug et al., 2011). The other species, especially *E. camaldulensis*, had relatively low monoterpene contents. In the case of *E. camaldulensis*,  $\alpha$ -pinene (22.52%) and 1,8-cineole (9.48%) were the predominant compounds, as indicated by Sebei et al. (2015).

Other studies reported on *E. microtheca* showed the following composition:  $\alpha$ -pinene (6.752 %) and  $\beta$ -pinene (5.006 %) as a major compound (Maghsoodlou et al., 2015). Thus, for many *Eucalyptus* species, several factors may influence monoterpenes synthesis, especially seasonal and diurnal emission activity cycles (He et al., 2000). The second major class in *E. oleosa* identified also with high rates is represented essentially by

$\alpha$ -pinene (1.0%- 24.7%),  $\beta$ -pinene (1.2%- 11.36%),  $\alpha$ -thujene (0.1%- 11.42%) and p-cymene (0.38%- 10.91%).

The third class in *E. oleosa* essential oils is the oxygenated sesquiterpenes constituted by spathulenol, thymol and borneol. The chemical composition of the essential oils of *E. oleosa* has been evaluated in many studies and they were presented in Table 1.

**Tableau 1.** Major components present in essential oils of *Eucalyptus oleosa* samples.

compounds	RI	Chamali et al. 2019	Ben Hassine et al. 2010	Rhimi et al. 2013	Jiamand et al. 2009	Marzoug et al. 2011	Safaei-Ghomi et al. 2009	Bignell et al. 1995	Ebadollahi et al. 2017
$\alpha$ thujene	931	--	11.42	0.1	--	--	--	--	2.42
$\alpha$ -pinene	936	12.28	10.91	14.5	1.0	21.80	11.19	24.7	15.25
$\beta$ -pinene	982	11.36	--	1.5	1.2	--	1.62	2.59	2.22
p-cymene	1026	--	10.91	--	--	3.30	5.86	3.39	0.38
m-cymene	1028	17.02	--	--	--	--	--	--	--
1,8-Cineole	1036	15.31	22.94	45.1	89.4	41.20	57.89	52.04	31.96
$\gamma$ -Terpinene	1060	--	--	0.7	0.70	3.00	--	--	--
soamyl valerianate	1106	--	--	1.9	--	0.30	--	--	--
$\alpha$ campholenal	1126	--	3.66	0.2	--	--	--	--	--
Trans-pinocarveol	1139	--	0.86	3.9	0.6	5.70	7.79	0.63	3.59
(E)-verbenol	1155	8.86	--	--	--	--	--	--	--
Pinocarvacrone	1163	--	1.58	--	--	--	3.18	--	--
Pinocarvone	1165	--	--	1.1	--	1.80	--	0.31	1.19
Borneol	1169	--	--	0.9	--	0.90	--	--	--
4-Terpineol	1177	--	--	1.9	--	--	--	--	--
p-cymén-8-ol	1185	--	--	--	--	1.70	--	--	--
$\alpha$ -Terpineol	1189	--	--	4.3	--	--	1.17	--	5.33
verbenone	1211	13.70	--	0.1	--	0.20	--	--	--
Isodihydrocarveol	1222	4.44	--	--	--	--	--	--	--
cuminaldéhyde	1237	--	--	--	--	0.90	--	--	--
Thymol	1310	--	2.04	0.1	--	--	--	--	0.73
Spathulenol	1578	--	--	0.4	--	--	0.34	0.83	0.36
Viridiflorol	1593	--	--	0.3	--	1.10	--	--	--
$\gamma$ -eudesmol	1635	0.34	--	--	--	5.30	--	--	0.57
$\beta$ -Eudesmol	1651	--	--	2.2	--	--	0.44	0.25	1.36
<b>Total</b>		99.97	99.10	98.00	93.50	97.60	99.64	97.44	99.87
<b>Monoterpene hydrocarbons (%)</b>		41.47	26.3	18.70	2.2	28.20	18.67	38.04	23.05
<b>Oxygenated monoterpenes (%)</b>		46.89	33.95	68.60	91.3	50.70	78.67	54.17	66.08
<b>Sesquiterpene hydrocarbons (%)</b>		2.54	38.88	0.50	0	4.10	0.34	2.27	0.77
<b>Oxygenated sesquiterpenes (%)</b>		6.01	0	6.50	0	7.70	1.05	2.54	3.69
<b>Others (%)</b>		3.44	0	3.70	0	6.50	0.91	0.42	6.28

(--) no identified, RI: Retention Index

All these variations might be due to the influence of geographical differences, environmental and growing conditions, physiological and biochemical states of plants, different extraction and analytical procedures, and genetic factors (Kokkini et al., 2004; Hassanpouraghdam et al., 2011). Furthermore, such variation can be attributed to several factors including plant age, climate, vegetative cycle stage, harvest time, geographical location, part plant used and genetic variation (Barra, 2009; Ben Hassine et al., 2012; Barbosa et al., 2016; Dorsaf et al., 2016; Almas et al., 2018). Several clinical studies indicate that Eucalyptol, the monoterpene, due to different medicinal properties including antioxidant (Ciftci et al., 2011), antimicrobial (Schürmann et al., 2019), anti-inflammatory

(Zhao et al., 2014) and respiratory disorder treatments (Sudhoff et al., 2015). On other hand, the richness in 1,8-cineole revealed several potential applications; as an insect repellent (Aldoghaim et al., 2018). Furthermore, it is often used as a flavoring agent for food products (Santos and Rao, 2001).

#### 4. BIOLOGICAL ACTIVITIES

Traditionally, the *Eucalyptus* oils are used to treat fever, bronchitis, asthma, and pulmonary diseases via inhalation (Horváth and Ács, 2015). Previous research found that *Eucalyptus* essential oils have numerous biological properties such as antioxidant, anti-bacterial, fungicide, anti-allergic, anti-cancer, anti-inflammatory, insecticidal and herbicidal effects

(Silva et al., 2003; Batish et al., 2006; Gilles et al., 2010; Salem et al., 2015; Vuong et al., 2015; Nakamura et al., 2020; Sharma, 2020; Pinto et al., 2021). Indeed, *Eucalyptus* essential oils play a central role in these biological functions by their active chemical substances (Barbosa et al., 2016; Migacz et al., 2018). Similarly, many essential oils produced by *E. oleosa* have been reported for their antioxidant, antimicrobial and insecticidal activities (Ebadollahi et al., 2013; Rahimi-Nasrabadi et al., 2013; Marzoug et al., 2015). Moreover, *E. oleosa* terpenes such as  $\alpha$ -Terpineol, p-cymene,  $\alpha$ - and  $\beta$ -pinene show much wider therapeutic uses antimicrobial, antiviral, antihyperglycemic, anti-inflammatory, antioxidant, antiparasitic and immunomodulatory (Upadhyay, 2022).

#### 4.1. Antioxidant activities

Antioxidants play an important role in food preservation by inhibiting oxidation processes and contributing to the health promotion provided by many dietary supplements, nutraceuticals, and functional food ingredients (Shahidi and Zhong, 2015). Furthermore, natural antioxidants are in demand for pharmaceuticals products (Brewer, 2011). Therefore, in recent years, considerable attention has been detected towards the identification of plants with antioxidant activity (Moon and Shibamoto, 2009). Previous studies revealed that *E. oleosa* leaf essential oils from Tunisia exhibited high antioxidant potential in both assays (DPPH and ABTS), the IC<sub>50</sub> were 52.8 ± 0.7 mg/mL and 176.5 ± 3.1mg/mL respectively (Ben Marzoug et al., 2010). In addition, the antioxidant capacity of four parts of *E. oleosa* essential oils (stems, adult leaves, fruits, and immature flowers) showed moderate antioxidant activities in which the best IC<sub>50</sub> is found for the adult leaves essential oil (0.013 ± 0.0006 mg/mL) in the ABTS assay (Marzoug et al., 2011). Whereas another study showed that the leaf essential oils of *E. oleosa* from Iran did not show any antioxidant activity (Rahimi-Nasrabadi et al., 2013). This moderate antioxidant activity is probably due to the low content of phenolic compounds presents that is related to the extraction technical used and origin of plant.

#### 4.2. Antimicrobial activities

The antimicrobial activity of the *E. oleosa* essential oils has been studied by several researchers and discussed in the text; the essential oils exhibit toxicity against a wide range of microbes, including bacteria, fungi,

yeast but the bioactivity against virus has not investigated (Table 2).

##### 4.2.1. Antibacterial activities

It has been reported that *E. oleosa* is active against Gram+ strains *Enterococcus faecalis* and *Staphylococcus aureus* with minimal inhibitory concentration which was situated between 0.028-0.056 mg/ml. In addition, the bactericidal dose against all organisms tested was ranged between 28-56 µg/ml (Ben Hassine et al., 2012). These results agreed with the study showed that the *E. oleosa* essential oils of different parts (stems, adult leaves, fruits, and immature flowers) appeared more active against the tested Gram+ such as *Staphylococcus aureus* and *Listeria monocytogenes* than Gram negative bacteria, although the immature flowers presented a larger prevalence of activity 0.39-3.72 mg/mL (Srinivasan et al., 2001). But another study revealed that the essential oil of *E. oleosa* exhibited high antibacterial activity against Gram negative ones, with highest inhibition zone 19.0 mm diameter and lowest MIC value 0.062 mg/ml against *E. coli* which shows that this microorganism is sensitive to *E. oleosa* essential oil (Rahimi-Nasrabadi et al., 2013). Hence, the activity against both types of bacteria Gram+ and Gram-, may be indicative of presence of broad spectrum antibiotic compounds or simply general metabolic toxins (Srinivasan et al., 2001).

##### 4.2.2. Antifungal activities

An antifungal activity of the different part (stems, adult leaves, fruits, and immature flowers) of *E. oleosa* essential oils were tested against three pathogenic fungi *Aspergillus ochraceus*, *Mucor ramannianus* and *Fusarium culmorum* and demonstrated that immature flowers and stems had strongest antifungal activity with minimal inhibition concentration value between 2.79-3.88 mg/ml (Srinivasan et al., 2001). Additionally, these findings were consistent with another study performed by Kouki et al. (2023) who reported that *E. oleosa* EOs exhibited a significant antifungal activity against five *Fusarium* spp.

Generally, *E. oleosa* essential oils showed variable antimicrobial activity against the different test organisms. This variability could be related to several factors such as chemical composition of essential oils and geographic location of the plant material, also the sensitivity of the bacterial strains and its nature (Sabo and Knezevic, 2019).

**Table 2.** Antimicrobial effects of *Eucalyptus oleosa* EOs investigated on the pathogenic microorganisms.

<i>Eucalyptus oleosa</i> EOs origin	Part used	Inhibited microorganisms	References
Gabes (Tunisia)	Arial parts	<i>Bacillus subtilis</i> , <i>Staphylococcus aureus</i> , <i>Listeria monocytogenes</i> , <i>Pseudomonas aeruginosa</i> , <i>Salmonella enterica</i> , <i>Escherichia coli</i> , <i>Klebsiella pneumoniae</i> , <i>Saccharomyces cerevisiae</i> , <i>Candida albicans</i> , <i>Aspergillus ochraceus</i> , <i>Mucor ramamnianus</i> , <i>Fusarium culmorum</i>	Marzoug et al., 2011
Gabes (Tunisia)	leaves	<i>Listeria monocytogenes</i> , <i>Klebsiella pneumoniae</i> , <i>Saccharomyces cerevisiae</i> , <i>Candida albicans</i> , <i>Mucor ramamnianus</i> , <i>Aspergillus ochraceus</i>	Ben Marzoug et al., 2010
Kashan (Iran)	Leaves	<i>Staphylococcus aureus</i> , <i>Staphylococcus epidermidis</i> , <i>Escherichia coli</i> , <i>Klebsiella pneumoniae</i> , <i>Bacillus subtilis</i> , <i>Bacillus cereus</i> , <i>Salmonella typhimurium</i>	Rahimi-Nasrabadi et al., 2013
Monastir (Tunisia)	Leaves	<i>Escherichia coli</i> , <i>Enterococcus feacalis</i> , <i>Staphylococcus aureus</i> , <i>Listeria monocytogens</i> , <i>Salmonella anatum</i> , <i>Salmonella enteritidis</i>	Ben Hassine et al., 2012

#### 4.2.3. Insecticidal/Acaricidal activities

The intense application of insecticides leads to the development of insecticide resistance in insect pest populations worldwide (Pittendrigh et al., 2008) and resulted in an increased risk of pesticides resistance, toxicological implications for human health and environmental pollution (Batish et al., 2006; Mahmood et al., 2016; Alengebawy et al., 2021). Thus, there has been a growing interest in research concerning the possible use of plant extracts as alternatives to synthetic pesticides (Ghosh et al., 2012). Recently, the insecticidal activity elicited by certain essential oils indicates that these botanical compounds could be used as alternative tools (Isman, 2000). Furthermore, essential oils are applied similarly to other pesticides and their biological activity is manifested both by exposure to their vapors and by topical application (Isman, 2000; Tarelli et al., 2009). Further, it has been reported that *Eucalyptus* species essential oils have significant pesticidal potentials (Maciel et al., 2010; Alzogaray et al., 2011; Pant et al., 2014; Filomeno et al., 2017; Ainane et al., 2019). Insecticidal activity of *E. oleosa* oils from Iran are assessed against American white moth, *Hyphantria cunea* Drury 1773 (Lepidoptera: Arctiidae) at different concentrations (0.1, 0.21, 0.45, 0.95 and 2%) used at three times (24h, 48h and 72h) in which found the LC<sub>50</sub> (Lethal Concentration to kill 50% of insects) values were estimated as 0.36% at a shorter duration (24h) (Ebadollahi et al., 2013). In addition, the

acaricidal effects of *E. oleosa* essential oils against *Tetranychus urticae* Koch (Acarina: Tetranychidae) have been reported (Ebadollahi et al., 2017). Hence, volatile oils from *E. oleosa* show strong phytotoxicity towards *H. cunea* and *T. urticae*. Indeed, it could provide opportunities for new biodegradable products for pest control considering their noticeable effects at low applied concentrations and short times of exposure. Indeed, the toxicity of the essential oils tested varies widely depending on the nature of the essential oil, the concentration used and the duration of the treatment. Studies have revealed that monoterpenes have insecticidal activities against the stored-product insects (Rajendran and Sriranjini, 2008; Papachristos et al., 2004).

#### 5. CONCLUSION

*Eucalyptus oleosa* shows high subspecific variability and resistance to arid conditions. It showed a great richness in essential oils and their chemical composition indicated that the most abundant component is 1,8-cineole, followed by  $\alpha$ -pinene. These characteristics represent advantages for the use of the species in afforestation and the valorization of these oils. Furthermore, *E. oleosa* essential oils possess a broad spectrum of biological effects, such as antioxidant, antibacterial, anti-fungal and anti-insecticidal activities. *E. oleosa* deserve to be deepened by studies on the relation subspecies, essential oils, and activities for a selection of promising subspecies for their aridity tolerance and their oils quantity, quality, and activities.

## REFERENCES

- Ainane, A., Khammour, F., Charaf, S., Elabboubi, M., Elkouali, M., Talbi, M., Benhima, R., Cherroud, S., Ainane, T. (2019). Chemical composition and insecticidal activity of five essential oils: *Cedrus atlantica*, *Citrus limonum*, *Rosmarinus officinalis*, *Syzygium aromaticum* and *Eucalyptus globules*. *Materials Today: Proceedings* 13, 474-485.
- Aldoghaim, F. S., Flematti, G. R., Hammer, K. A. (2018). Antimicrobial activity of several cineole-rich Western Australian *Eucalyptus* essential oils. *Microorganisms* 6, 122.
- Alengebawy, A., Abdelkhalek, S.T., Qureshi, S.R., Wang, M.Q. (2021). Heavy metals and pesticides toxicity in agricultural soil and plants: Ecological risks and human health implications. *Toxics* 9, 42.
- Almas, I., Innocent, E., Machumi, F., Kisinza, W. (2018). Effect of Geographical location on yield and chemical composition of essential oils from three *Eucalyptus* species growing in Tanzania. *Asian Journal of Traditional Medicines* 14, 1-12.
- Al-Snafi, A.E. (2017). The pharmacological and therapeutic importance of *Eucalyptus* species grown in Iraq. *IOSR Journal of Pharmacy* 7, 72-91.
- Alzogaray, R.A., Lucia, A., Zerba, E.N., Masuh, H.M. (2011). Insecticidal activity of essential oils from eleven *Eucalyptus* spp. and two hybrids: lethal and sublethal effects of their major components on *Blattella germanica*. *Journal of economic entomology* 104, 595-600.
- Barbosa, L.C.A., Filomeno, C.A., Teixeira, R.R. (2016). Chemical variability and biological activities of *Eucalyptus* spp. essential oils. *Molecules* 21, 1671.
- Barra, A. (2009). Factors affecting chemical variability of essential oils: a review of recent developments. *Natural product communications* 4.
- Batish, D.R., Singh, H.P., Setia, N., Kaur, S., Kohli, R.K. (2006). Chemical composition and phytotoxicity of volatile essential oil from intact and fallen leaves of *Eucalyptus citriodora*. *Zeitschrift für Naturforschung C* 61, 465-471.
- Ben Hassine, D., Ben Ismail, H., Jribi, C., Khouja, M.L., Abderrabba, M. (2012). *Eucalyptus oleosa* F. Muell essential oil: extraction, chemical composition and antimicrobial activity. In *International symposium on Medicinal and Aromatic Plants-SIPAM 997*, 77-82.
- Ben Marzoug, H.N., Bouajila, J., Ennajar, M., Lebrihi, A., Mathieu, F., Couderc, F., Abderraba, M., Romdhane, M. (2010). *Eucalyptus (gracilis, oleosa, salubris and salmonophloia)* essential oils: their chemical composition and antioxidant and antimicrobial activities. *Journal of medicinal food* 13, 1005-1012.
- Bignell, C.M., Dunlop, P.J., Brophy, J.J., Jackson, J.F. (1995). Volatile leaf oils of some South-western and Southern Australian species of the genus *Eucalyptus*. Part V. subgenus symphyomyrtus, section bisectaria, series oleosae. *Flavour and fragrance journal* 10, 313-317.
- Booth, T.H. (2013). Eucalypt plantations and climate change. *Forest ecology and management* 301, 28-34.
- Boukhatem, M.N., Boumaiza, A., Nada, H.G., Rajabi, M., Mousa, S.A. (2020). *Eucalyptus globulus* essential oil as a natural food preservative: Antioxidant, antibacterial and antifungal properties in vitro and in a real food matrix (orangina fruit juice). *Applied Sciences* 10, 5581.
- Brewer, M.S. (2011). Natural antioxidants: sources, compounds, mechanisms of action, and potential applications. *Comprehensive reviews in food science and food safety* 10, 221-247.
- Chamali, S., Bendaoud, H., Saadaoui, E., Elfalleh, W., Romdhane, M. (2019). A new process for green extraction of essential oil from *Eucalyptus oleosa*: Microwave-assisted hydro distillation. *Arabian Journal of Medicinal and Aromatic Plants* 5, 35-46.
- Ciftci, O., Ozdemir, I., Tanyildizi, S., Yildiz, S., Oguzturk, H. (2011). Antioxidative effects of curcumin,  $\beta$ -myrcene and 1, 8-cineole against 2, 3, 7, 8-tetrachlorodibenzo-p-dioxin-induced oxidative stress in rats liver. *Toxicology and industrial health* 27, 447-453.
- Crisp, M.D., Burrows, G.E., Cook, L.G., Thornhill, A.H., Bowman, D.M. (2011). Flammable biomes dominated by eucalypts originated at the Cretaceous-Palaeogene boundary. *Nature Communications* 2, 1-8.
- Damjanović-Vratnica, B., Đakov, T., Suković, D., Damjanović, J. (2011). Antimicrobial effect of essential oil isolated from *Eucalyptus globulus* Labill. from Montenegro. *Czech Journal of Food Sciences* 29, 277-284.
- Dhakad, A.K., Pandey, V.V., Beg, S., Rawat, J.M., Singh, A. (2018). Biological, medicinal and toxicological significance of *Eucalyptus* leaf essential oil: a review. *Journal of the Science of Food and Agriculture* 98, 833-848.
- Dorsaf, B.H., Hanen, B.I., Chokri, J., Larbi, K.M., Manef, A. (2016). Chemical composition of some Tunisian *Eucalyptus* essential oils as

- obtained by hydrodistillation using Clevenger type apparatus. *Biosciences Biotechnology Research Asia* 7, 647-656.
- Ebadollahi, A., Rahimi-Nasrabadi, M., Batooli, H., Geranmayeh, J. (2013). Evaluation of the insecticidal activities of three *Eucalyptus* species cultivated in Iran, against *Hyphantria Cunea* Drury (Lepidoptera: Arctiidae). *Journal of Plant Protection Research* 53.
- Ebadollahi, A., Sendi, J.J., Maroufpoor, M., Rahimi-Nasrabadi, M. (2017). Acaricidal potentials of the terpene-rich essential oils of two Iranian *Eucalyptus* species against *Tetranychus urticae* Koch. *Journal of oleo science* 66,307-314.
- Elaissi, A., Chraif, I., Bannour, F., Farhat, F., Salah, M. B., Chemli, R., Khouja, M.L. (2007). Contribution to the qualitative and quantitative study of seven *Eucalyptus* species essential oil harvested of Hajeb's Layoun arboreta (Tunisia). *Journal of Essential Oil Bearing Plants* 10, 15-25.
- Fadel, H., Mar, F., El-Sawy, A., El-Ghorab, A. (1999). Effect of extraction techniques on the chemical composition and antioxidant activity of *Eucalyptus camaldulensis* var. *brevirostris* leaf oils. *Zeitschrift für Lebensmitteluntersuchung und-Forschung A* 208, 212-216.
- Fathi, E., Sefidkon, F. (2012). Influence of drying and extraction methods on yield and chemical composition of the essential oil of *Eucalyptus sargentii*. *Journal of Agricultural Science and Technology* 14, 1035-1042.
- Filomeno, C.A., Barbosa, L.C.A., Teixeira, R.R., Pinheiro, A.L., de Sá Farias, E., de Paula Silva, E.M., Picanço, M.C. (2017). *Corymbia* spp. and *Eucalyptus* spp. essential oils have insecticidal activity against *Plutella xylostella*. *Industrial crops and products* 109, 374-383.
- Ghaffar, A., Yameen, M., Kiran, S., Kamal, S., Jalal, F., Munir, B., Salim, S., Rafiq, N., Ahmad, A., Saba, I., Jabbar, A. (2015). Chemical composition and in-vitro evaluation of the antimicrobial and antioxidant activities of essential oils extracted from seven *Eucalyptus* species. *Molecules* 20, 20487-20498.
- Ghosh, A., Chowdhury, N., Chandra, G. (2012). Plant extracts as potential mosquito larvicides. *The Indian journal of medical research* 135, 581.
- Gilles, M., Zhao, J., An, M., Agboola, S. (2010). Chemical composition and antimicrobial properties of essential oils of three Australian *Eucalyptus* species. *Food Chemistry* 119, 731-737.
- Grattapaglia, D., Vaillancourt, R.E., Shepherd, M., Thumma, B.R., Foley, W., Külheim, C., Potts, B.M., Myburg, A.A. (2012). Progress in Myrtaceae genetics and genomics: *Eucalyptus* as the pivotal genus. *Tree Genetics & Genomes* 8, 463-508.
- Hashemi-Moghaddam, H., Kalatejari, A., Afshari, H., Ebadi, A.H. (2013). Microwave accelerated distillation of essential oils from the leaves of *Eucalyptus microtheca*: Optimization and comparison with conventional hydrodistillation. *Asian Journal of Chemistry* 25, 5423-5427.
- Hassanpouraghdam, M.B., Akhgari, A.B., Aazami, M.A., Emarat-Pardaz, J. (2011). New menthone type of *Mentha pulegium* L. volatile oil from Northwest Iran. *Czech Journal of Food Sciences* 29, 285-290.
- He, C., Murray FLYons, T. (2000). Monoterpene and isoprene emissions from 15 *Eucalyptus* species in Australia. *Atmospheric Environment* 34, 645-655.
- Ho, J.C. (2010). Chemical composition and bioactivity of essential oil of seed and leaf from *Alpinia speciosa* grown in Taiwan. *Journal of the Chinese Chemical Society* 57, 758-763.
- Hobbs, T.J., Bennell, M., Bartle, J. (2009). Developing species for woody biomass crops in lower rainfall southern Australia. *Flora Search 3a*. Report to the Joint Venture Agroforestry Program (JVAP) and Future Farm Industries CRC. Publication 242.
- Horváth, G., Ács, K. (2015). Essential oils in the treatment of respiratory tract diseases highlighting their role in bacterial infections and their anti-inflammatory action: a review. *Flavour and Fragrance Journal* 30, 331-341.
- Isman, M.B. (2000). Plant essential oils for pest and disease management. *Crop protection* 19, 603-608.
- Jaymand, K., Rezaei, M.B., Naderi, H.B.M. (2009). Volatile oil constituents of the *Eucalyptus viridis* RT Baker and *Eucalyptus oleosa* F. Muell. leaves from Iran. *Journal of Medicinal Plants* 8, 105-108.
- Kokkini, S., Hanlidou, E., Karousou, R., Lanaras, T. (2004). Clinal variation of *Mentha pulegium* essential oils along the climatic gradient of Greece. *Journal of Essential Oil Research* 16, 588-593.
- Kouki, H., Amri, I., Souihi, M., Pieracci, Y., Trabelsi, I., Hamrouni, L., Flamini, G., Hirsch, A.M., Mabrouk, Y. (2023). Chemical composition, antioxidant, herbicidal and antifungal activities of leaf essential oils from

- three Tunisian *Eucalyptus* species. *Journal of Plant Diseases and Protection* 1-12.
- Maciel, M.V., Morais, S.M., Bevilaqua, C.M.L., Silva, R.A., Barros, R.S., Sousa, R.N., Sousa, R.N., Brito, E.S., Souza-Neto, M.A. (2010). Chemical composition of *Eucalyptus* spp. essential oils and their insecticidal effects on *Lutzomyia longipalpis*. *Veterinary parasitology* 167, 1-7.
- Maghsoodlou, M.T., Kazemipoor, N., Valizadeh, J., Seifi, M.F.N., Rahnesan, N. (2015). Essential oil composition of *Eucalyptus microtheca* and *Eucalyptus viminalis*. *Avicenna journal of Phytomedicine* 5, 540.
- Mahmood, I., Imadi, S.R., Shazadi, K., Gul, A., & Hakeem, K.R. (2016). Effects of pesticides on environment. *Plant, soil and microbes: volume 1: implications in crop science* 253-269.
- Marzoug, H.N.B., Romdhane, M., Lebrihi, A., Mathieu, F., Couderc, F., Abderraba, M., Khouja, M.A., Bouajila, J. (2011). *Eucalyptus oleosa* essential oils: chemical composition and antimicrobial and antioxidant activities of the oils from different plant parts (stems, leaves, flowers and fruits). *Molecules* 16, 1695-1709.
- Merchant, A., Tausz, M., Arndt, S.K., Adams, M.A. (2006). Cyclitols and carbohydrates in leaves and roots of 13 *Eucalyptus* species suggest contrasting physiological responses to water deficit. *Plant, Cell & Environment* 29, 2017-2029.
- Migacz, I.P., Raeski, P.A., Almeida, V.P.D., Raman, V., Nisgoski, S., Muniz, G.I.B.D., Farago, P.V., Khan, I.A., Budel, J.M. (2018). Comparative leaf morpho-anatomy of six species of *Eucalyptus* cultivated in Brazil. *Revista Brasileira de Farmacognosia* 28, 273-281.
- Moon, J.K., Shibamoto, T. (2009). Antioxidant assays for plant and food components. *Journal of agricultural and food chemistry* 57, 1655-1666.
- Nakamura, T., Yoshida, N., Yamanoi, Y., Honryo, A., Tomita, H., Kuwabara, H., Kojima, Y. (2020). *Eucalyptus* oil reduces allergic reactions and suppresses mast cell degranulation by downregulating IgE-FcεRI signalling. *Scientific reports* 10, 1-15.
- Ndiaye, E.H.B., Diop, M.B., Gueye, M.T., Ndiaye, I., Diop, S.M., Fauconnier, M.L., Lognay, G. (2018). Characterization of essential oils and hydrosols from senegalese *Eucalyptus camaldulensis* Dehnh. *Journal of essential oil research* 30, 131-141.
- Ohara, K., Matsunaga, E., Nanto, K., Yamamoto, K., Sasaki, K., Ebinuma, H., & Yazaki, K. (2010). Monoterpene engineering in a woody plant *Eucalyptus camaldulensis* using a limonene synthase cDNA. *Plant biotechnology journal* 8, 28-37.
- Pant, M., Dubey, S., Patanjali, P.K., Naik, S.N., Sharma, S. (2014). Insecticidal activity of *Eucalyptus* oil nanoemulsion with karanja and jatropha aqueous filtrates. *International Biodeterioration & Biodegradation* 91, 119-127.
- Papachristos, D.P., Karamanoli, K.I., Stamopoulos, D.C., Menkissoglu-Spiroudi, U. (2004). The relationship between the chemical composition of three essential oils and their insecticidal activity against *Acanthoscelides obtectus* (Say). *Pest Management Science: Formerly Pesticide Science* 60, 514-520.
- Pino, J.A., Marbot, R., Quert, R., García, H. (2002). Study of essential oils of *Eucalyptus resinifera* Smith, *E. tereticornis* Smith and *Corymbia maculata* (Hook.) KD Hill & LAS Johnson, grown in Cuba. *Flavour and fragrance journal* 17, 1-4.
- Pinto, M., Soares, C., Martins, M., Sousa, B., Valente, I., Pereira, R., Fidalgo, F. (2021). Herbicidal effects and cellular targets of aqueous extracts from young *Eucalyptus globulus* Labill. Leaves. *Plants* 10, 1159.
- Pittendrigh, B.R., Margam, V.M., Sun, L., Huesing, J.E. (2008). Resistance in the postgenomics age. *Insect resistance management: Biology, economics and prediction* 39-68.
- Pourmortazavi, S.M., Rahimi-Nasrabadi, M., Aghazadeh, M., Ganjali, M.R., Karimi, M.S., Norouzi, P. (2017). Synthesis, characterization and photocatalytic activity of neodymium carbonate and neodymium oxide nanoparticles. *Journal of Molecular Structure* 1150, 411-418.
- Pourmortazavi, S.M., Taghdiri, M., Makari, V., Rahimi-Nasrabadi, M. (2015). Procedure optimization for green synthesis of silver nanoparticles by aqueous extract of *Eucalyptus oleosa*. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy* 136, 1249-1254.
- Rahimi-Nasrabadi, M., Nazarian, S., Farahani, H., Fallah Koohbijari, G.R., Ahmadi, F., Batooli, H. (2013). Chemical composition, antioxidant, and antibacterial activities of the essential oil and methanol extracts of *Eucalyptus largiflorens* F. Muell. *International Journal of Food Properties* 16, 369-381.
- Rajendran, S., Sriranjini, V. (2008). Plant products as fumigants for stored-product insect control. *Journal of stored products Research* 44, 126-135.



- Richter, J., Schellenberg, I. (2007). Comparison of different extraction methods for the determination of essential oils and related compounds from aromatic plants and optimization of solid-phase microextraction/gas chromatography. *Analytical and bioanalytical chemistry* 387, 2207-2217.
- Saadaoui, E., Yahia, K.B., Dhahri, S., Jamaa, M.L.B., Khouja, M.L. (2017). An overview of adaptive responses to drought stress in *Eucalyptus* spp. *Forestry Studies* 67, 86-96.
- Sabo, V.A., Knezevic, P. (2019). Antimicrobial activity of *Eucalyptus camaldulensis* Dehn. plant extracts and essential oils: A review. *Industrial crops and products* 132, 413-429.
- Safaei-Ghomi, J., Shahroodi, F., Batooli, H. (2009). Volatile constituents of the flowers and leaves of *Eucalyptus oleosa* cultivated in central Iran. *Chemistry of natural compounds* 45, 106-107.
- Salehi, B., Sharifi-Rad, J., Quispe, C., Llaïque, H., Villalobos, M., Smeriglio, A., Trombetta, D., Ezzat, M.S., Salem, M.A., Zayed, A., Castillo, C.M.S., Yazdi, S.E., Sen, S., Acharya, K., Sharopov, F., Martins, N. (2019). Insights into *Eucalyptus* genus chemical constituents, biological activities and health-promoting effects. *Trends in Food Science & Technology* 91, 609-624.
- Salem, M.Z., Ashmawy, N.A., Elansary, H.O., El-Settawy, A.A. (2015). Chemotyping of diverse *Eucalyptus* species grown in Egypt and antioxidant and antibacterial activities of its respective essential oils. *Natural Product Research* 29, 681-685.
- Santos, F.A., Rao, V.S.N. (2001). 1, 8-cineol, a food flavoring agent, prevents ethanol-induced gastric injury in rats. *Digestive diseases and sciences* 46, 331-337.
- Sawalha, H., Abiri, R., Sanusi, R., Shaharuddin, N.A., Noor, A.A.M., Ab-Shukor, N.A., Abdul-Hamid, H., Ahmad, S.A. (2021). Toward a better understanding of metal nanoparticles, a novel strategy from *Eucalyptus* plants. *Plants* 10, 929.
- Schürmann, M., Oppel, F., Gottschalk, M., Büker, B., Jantos, C.A., Knabbe, C., Hutten, A., Kaltschmidt, B., Kaltschmidt, C., Sudhoff, H. (2019). The therapeutic effect of 1, 8-cineol on pathogenic bacteria species present in chronic rhinosinusitis. *Frontiers in microbiology* 10, 2325.
- Sebei, K., Sakouhi, F., Herchi, W., Khouja, M.L., Boukhchina, S. (2015). Chemical composition and antibacterial activities of seven *Eucalyptus* species essential oils leaves. *Biological research* 48, 1-5.
- Shahidi, F., Zhong, Y. (2015). Measurement of antioxidant activity. *Journal of functional foods* 18, 757-781.
- Sharma, A.D., Kaur, I. (2020). Eucalyptol (1, 8 cineole) from *Eucalyptus* essential oil a potential inhibitor of COVID 19 corona virus infection by molecular docking studies. *Preprints*, 2020030455.
- Silva, J., Abebe, W., Sousa, S.M., Duarte, V.G., Machado, M.I.L., Matos, F.J.A. (2003). Analgesic and anti-inflammatory effects of essential oils of *Eucalyptus*. *Journal of ethnopharmacology* 89, 277-283.
- Srinivasan, D., Nathan, S., Suresh, T., Perumalsamy, P.L. (2001). Antimicrobial activity of certain Indian medicinal plants used in folkloric medicine. *Journal of ethnopharmacology* 74, 217-220.
- Sudhoff, H., Klenke, C., Greiner, J.F., Müller, J., Brotzmann, V., Ebmeyer, J., Kaltschmidt, B., Kaltschmidt, C. (2015). 1, 8-Cineol reduces mucus-production in a novel human ex vivo model of late rhinosinusitis. *PLoS One* 10.
- Tarelli, G., Zerba, E.N., Alzogaray, R.A. (2009). Toxicity to vapor exposure and topical application of essential oils and monoterpenes on *Musca domestica* (Diptera: Muscidae). *Journal of economic entomology* 102, 1383-1388.
- Upadhyay, R.K. (2022). Therapeutic and insecticidal potential of plant terpenes: A review. *International Journal of Green Pharmacy* 16.
- Vilela, G.R., de Almeida, G.S., D'Arce, M.A.B.R., Moraes, M.H.D., Brito, J.O., da Silva, M.F.D.G.F., Silva, S.C., Piedade, S.M.D.F., Calori-Domingues, M.A., da Gloria, E.M. (2009). Activity of essential oil and its major compound, 1,8-cineole, from *Eucalyptus globulus* Labill., against the storage fungi *Aspergillus flavus* Link and *Aspergillus parasiticus* Speare. *Journal of Stored Products Research* 45, 108-111.
- Vivekanandhan, P., Usha-Raja-Nanthini, A., Valli, G., Subramanian, S.M. (2020). Comparative efficacy of *Eucalyptus globulus* (Labill) hydrodistilled essential oil and temephos as mosquito larvicide. *Natural product research* 34, 2626-2629.
- Vuong, Q.V., Chalmers, A.C., Jyoti, B.D., Bowyer, M.C., Scarlett, C.J. (2015). Botanical, phytochemical, and anticancer properties of the *Eucalyptus* species. *Chemistry & biodiversity* 12, 907-924.

Zhao, C., Sun, J., Fang, C., Tang, F. (2014). 1, 8-cineol attenuates LPS-induced acute pulmonary inflammation in mice. *Inflammation* 37, 566-572.

Zrira, S., Bessiere, J.M., Menut, C., Elamrani, A., Benjilali, B. (2004). Chemical composition of the essential oil of nine *Eucalyptus* species growing in Morocco. *Flavour and fragrance journal* 19, 172-175.