

1  
2  
3  
4  
5  
6  
7  
8  
9  
10  
11  
12  
13  
14  
15  
16  
17  
18  
19  
20  
21  
22  
23  
24

**THERMOGRAPHIC STUDIES OF COCURRENT AND MIXED FLOW  
SPRAY DRYING OF HEAT SENSITIVE BIOACTIVE COMPOUNDS**

Obón<sup>\*</sup> J.M.<sup>a</sup>, Luna-Abad J.P.<sup>b</sup>, Bermejo B.<sup>a</sup>, Fernández-López, J.A.<sup>a</sup>

(a) Departamento de Ingeniería Química y Ambiental. (b) Departamento de Ingeniería Térmica y de Fluidos. Universidad Politécnica de Cartagena. Spain

**\*Corresponding author e-mail:** josemaria.obon@upct.es (José María Obón)

**Contact details:** Departamento de Ingeniería Química y Ambiental  
Universidad Politécnica de Cartagena  
Paseo Alfonso XIII, 52, E-30203 Cartagena (Murcia), Spain  
Phone: + 34 968 32 55 64 Fax: + 34 968 32 55 55

**Authors e-mail:** jp.lunabad@upct.es (Juan Pedro Luna-Abad)  
bartolo.bh@gmail.com (Bartolomé Bermejo)  
josea.fernandez@upct.es (José Antonio Fernández-López)

**Submitted to: Journal of Food Engineering**

**Short title:** Thermography of spray drying bioactive compounds

**Keywords:** cocurrent spray drying; mixed-flow spray drying; thermography; enzymes;  
natural colorants; microencapsulation

25 **Abstract**

26 Production of powders of heat sensitive bioactive compounds in a bench scale spray  
27 drier was studied under cocurrent and mixed flow pattern conditions using two inlet air  
28 temperatures of 200 and 120 °C. Two natural colorants (riboflavin-5-phosphate and red  
29 beet) and an enzyme (alpha-amylase), were selected for experimentation.  
30 Thermographic studies showed interesting asymmetric profiles of temperatures outside  
31 and inside of the drying chamber, because turbulent air flow conditions and thermal  
32 trajectories of atomized drops during its drying process were dependent on flow  
33 patterns. Powders of natural colorants maintained its color strength, and alpha-amylase  
34 powders retain more than 82.9% of its enzyme activity even at the highest air  
35 temperature of 200 °C and using mixed flow. This work concludes that spray drying  
36 under cocurrent and mixed-flow patterns of heat labile bioactive compounds is feasible,  
37 influencing drying yields and the properties of powders obtained.

## 38 **1. Introduction**

39 Spray drying is a well-known technique to formulate food powders (Murugesan & Orsat,  
40 2012; Ray et al., 2016; Fang & Bhandari, 2017). It plays an important role in  
41 microencapsulation of natural ingredients as antioxidants, colorants, enzymes or cells  
42 used for the food industry. Microencapsulation protects bioactive compounds from light,  
43 temperature or oxygen during storage, and can affect its release behavior during  
44 digestion (Gharsallaoui et al., 2007; Barbosa & Teixeira, 2017; Guerin et al., 2017;  
45 Wang et al., 2018).

46 Spray drying process comprises three phases: the atomization of the liquid stream, the  
47 drying phase with the formation of solid product particles and the collection of powders.  
48 All these phases are important but the drying in the chamber has a huge effect on the  
49 drying process efficacy and the final product properties. In the chamber, liquid atomized  
50 droplets move through different trajectories due to the turbulent drying air gas flow  
51 inside, so each droplet loses its water content under different conditions of temperature  
52 and humidity. Contact patterns of atomized liquid droplets and drying air gas flow inside  
53 the drying chamber can be classified as cocurrent, countercurrent and mixed-flow. In  
54 mixed-flow pattern atomized droplets passes countercurrent and then cocurrent with  
55 drying air gas flow. Cocurrent spray dryers are the most widely used in comparison with  
56 countercurrent, and mixed-flow dryers are a good option to dry thermostable products  
57 (Cal & Sollohub, 2010; Keshani et al. 2015). Selection of contact pattern in drying  
58 chamber and spray drying conditions are critical to avoid damage or inactivation during  
59 the spray drying process of natural compounds such as vitamins, enzymes or cells that  
60 are thermolabile (Yoshii et al., 2008; Schutyser et al., 2012).

61 There is a high number of studies that optimize spray drying conditions using factorial  
62 experimental design in order to improve powder properties and to maximize product

63 yields (Singh & Singh-Hathan, 2017; Focaroli et al., 2019; Nair et al., 2019). In all cases  
64 a good knowledge of spray drying fluid dynamics would help researchers to select  
65 properly the operation conditions of the equipment.

66 The aim of this work was to evaluate the spray drying of thermolabile bioactive  
67 compounds under cocurrent and mixed flow conditions, working with a worldwide used  
68 bench-scale Mini Spray dryer. This study selected three commercial bioactive  
69 preparations: two water-soluble natural colorants and an enzyme. Riboflavin-5-  
70 phosphate or flavin mononucleotide (FMN) is the phosphorylated form of vitamin B<sub>2</sub>  
71 (riboflavin), an enzyme cofactor, and it is commercialized pure as an orange colorant  
72 powder (Choe et al., 2005; Sheraz et al., 2014). Red beet is a red-purple colorant  
73 extract and its color is due to betalains, a mix of red-purple betacyanins and yellow  
74 betaxanthins. This colorant is moderately stable, and it is commercialized as a  
75 microencapsulated powder obtained by spray drying from a blend of red beet juice and  
76 maltodextrin as drying aid (Fernández-López et al., 2013; Khan, 2016). Alpha-amylase  
77 (1,4- $\alpha$ -D-Glucan-glucanohydrolase; EC 3.2.1.1) from *Aspergillus oryzae* is an enzyme  
78 widely used in the food industry that breaks down soluble starch (Gupta et al., 2003). A  
79 special emphasis was paid to the effect of temperature; as different spray patterns offer  
80 diverse thermal degradation of bioactive compounds. The study measure for the first  
81 time to our knowledge simultaneous temperature profiles in the surface (thermographic  
82 images) and inside of the drying chamber (thermocouples) under cocurrent and mixed  
83 flow conditions.

## 84 **2. Materials and methods**

### 85 *2.1. Materials*

86 The natural colorants riboflavin-5-phosphate (E-101) and red beet powder (E-162)  
87 were kindly supplied by PROQUIMAC PFC, Barcelona, Spain. Crude  $\alpha$ -amylase from

88 *Aspergillus oryzae* (30 U/mg) and dinitrosalicylic acid were purchased from SIGMA-  
89 ALDRICH, Madrid, Spain. Soluble starch, maltose 1-hydrate, and tartrate sodium-  
90 potassium were supplied by PANREAC QUÍMICA, S.A., Barcelona, Spain.

## 91 2.2. *Spray drying equipment set-up*

92 A bench-scale Mini Spray Dryer B-290 (Büchi Labortechnik AG, Flawil, Switzerland)  
93 was modified to be operated under cocurrent or mixed flow. An innovative glass  
94 cylinder (50 x 15 cm) was made with the same geometry that the original but bottom  
95 zone was adapted to locate a spray nozzle to spray the liquid upwards (Figure 1A).  
96 Working in cocurrent flow liquid was atomized with the upper spray nozzle and hot air  
97 travels downwards in the same direction, however, in mixed flow liquid was atomized  
98 with the bottom spray nozzle traveling upwards in the opposite direction of hot air first,  
99 and then in the same direction (Figure 1A). An atomization angle of 30° was calculated  
100 from photographs taken during water atomization and drawn in Figure 1A. Height of  
101 glass cylinder was graduated in centimetres from top (Figure 1A) and its circumference  
102 perimeter numbered from 1 to 8 (Figure 1B,1C) to facilitate drying chamber  
103 descriptions. Drying air enters from the top of the drying chamber through two  
104 symmetric semi-circular crowns, because in the middle is located the spray nozzle  
105 (Figure 1B). Drying air travelling through the semi-circular crown located in position 3 is  
106 divided in two because of the spray nozzle position, while air travelling through position  
107 7 it is not divided. Inlet drying air travels faster through position 3. Outlet of drying air is  
108 located at 23° of plane 1-5 (Figure 1C). Temperatures on the surface of the glass  
109 cylinder were recorded during the spray drying process using a high resolution infrared  
110 FLIR T400 thermographic camera (Portland, Oregon, USA). Due to equipment  
111 configuration, thermographic images could only be taken from positions 1 to 5 (Figure  
112 1C). Surface temperatures were followed in position 3 at four cylinder lengths H1-H4

113 (Figure 1A), and also in the cyclone upper zone. To measure the temperature inside  
114 the glass cylinder thermocouple tips were located at 0.5, 2, 4 and 6 cm from the centre,  
115 every 5 cm at ten different positions from the top (dots shown in Figure 1A and 1C).  
116 Four thermocouples were rotated together around central axis and moved vertically  
117 through drying chamber to measure all points. A total of 80 measurements were done  
118 in the plane of drying air flow exit (Figure 1C). A four channel temperature datalogger  
119 (4KDatalog, TC S.A., Madrid, Spain) acquired data from four stainless steel  
120 thermocouples of 1 mm diameter (Type K, class 1) done in the same set. Temperatures  
121 were measured as the average of three waves obtained in steady state, and data used  
122 to create filled contour plots with SigmaPlot for Windows v10.0 (Systat Software, Inc.,  
123 USA).

### 124 *2.3. Spray drying experiments*

125 Liquid feeds were prepared dissolving powders of colorants or  $\alpha$ -amylase in  
126 deionised water at 2% w/v. Inlet liquid was thermostated at 22 °C, and feed at a fixed  
127 rate of 0.5 L/h (0.14 g/s) through a two-fluids atomizing nozzle working with a spray air  
128 flow-rate of 0.47 m<sup>3</sup>/h. Atomization cone angle was 30° (Figure 1A). Drying air flow-rate  
129 was fixed to 18 m<sup>3</sup>/h (80% of scale). Two inlet air temperatures were studied 120 and  
130 200 °C. Spray drying equipment was in a room thermostated at 25 °C. Inlet air was  
131 from outside of the building and outlet air was also conducted outside of the building.  
132 Powders were collected with a high-performance cyclone in a collection vessel.  
133 Powders were placed in closed flasks, weighted, and characterized as soon as  
134 possible. Meantime, samples were stored in a fridge (4 °C).

135 Drying process efficacy was evaluated by drying yield, retention of color strength or  
136 enzyme activity. Powders were weighted with an electronic balance Sartorius AX224  
137 (Sartorius, Madrid, Spain), and drying yield of spray drying experiments calculated as:

$$\text{Drying yield (\%)} = \frac{\text{grams of powder obtained}}{\text{grams of initial solid of feed}} \times 100$$

138 Thermostability of colorants during spray drying experiments was evaluated as color  
139 strength retention:

$$\text{Color strength retention (\%)} = \frac{\text{final color strength}}{\text{initial color strength}} \times 100$$

140 Thermostability of  $\alpha$ -amylase in spray drying experiments was evaluated as enzyme  
141 activity retention:

$$\text{Enzyme activity retention (\%)} = \frac{\text{final } \alpha - \text{amylase activity}}{\text{initial } \alpha - \text{amylase activity}} \times 100$$

142 All different spray drying experimental conditions were done by triplicate.

#### 143 2.4. Characterization of bioactive compound powders

144 *Measurement of color strength.* Colorant powders were prepared 0.005% w/v  
145 (riboflavin-5-phospahte) and 0.4% w/v (red beet) with deionised water, and its  
146 absorbance measured at 420 nm (riboflavin-5-phosphate) or 535 nm (red beet), versus  
147 a blank cell filled with distilled water, and within the linear range of the  
148 spectrophotometer (Agilent 8453, Waldbronn, Germany). Solids weight was considered  
149 in wet basis. Color strength was calculated as the absorbance of a 1% w/v dissolution  
150 measured at the maximum wavelength of the colorant.

151 *Measurements of  $\alpha$ -amylase activity.* One unit (U) of  $\alpha$ -amylase activity was defined  
152 as the amount of enzyme that liberates 1  $\mu$ mol of maltose per minute at pH 6.0 and 25  
153  $^{\circ}$ C. Assay was adapted from Keharom et al. 2016, as follow: a reaction mixture of 4 ml  
154 starch solution (1% w/v) and 1 ml of enzyme solution were incubated at 25  $^{\circ}$ C. Samples  
155 of 0.5 ml were withdrawn every 30 seconds for 5 min and mixed in test tubes with 0.5  
156 ml of DNS reagent (Miller, 1959). Samples were incubated at 85  $^{\circ}$ C for 10 min, diluted  
157 with 2 ml of water, and absorbance measured spectrophotometrically at 540 nm versus

158 a blank cell filled with a sample incubated with water and DNS. Maltose was the  
159 standard of the calibration curve. Activity was calculated as:

160  $\text{Activity } (\mu\text{mol}/\text{min}) = \Delta\text{Absorbance}/\text{min} \times 5 \text{ (ml)} / \text{Calibration curve slope (Abs/mM)}$

161 *Measurements of water content.* Determinations were done following reduction in  
162 weight of 5 g of the samples dried at 90 °C in an oven (Digitheat, Selecta, Spain), until  
163 obtaining a constant value. Weights were measured with an electronic balance  
164 Sartorius AX224 of 0.1 mg precision (Sartorius, Madrid, Spain). Results were  
165 calculated in wet basis and expressed as percentage in mass.

166 *Measurements of bulk density.* Bulk density was determined as tapped density by  
167 weighting 3 g of a sample into a 10 ml graduated cylinder. Cylinder was vibrated  
168 approximately 1500 taps to obtain a near optimum packing (Autotap, Quantachrome,  
169 USA) (Miravet et al., 2016). When a steady volume was reached, volume was  
170 measured, and density calculated as  $\text{Kg}/\text{m}^3$ .

171 *Dissolution test.* Dissolution test consisted in dissolving 2.50 g of powder in 50 ml of  
172 water (solution 5%) by magnetic agitation (Agimatic S, Selecta, Spain) until obtaining a  
173 clear solution (Miravet et al., 2016). Magnetic stirring was established at 1000 rpm, with  
174 a 20 mm cylindrical stir bar and a 100 mL Erlenmeyer flask. The time to fully  
175 reconstitute the powders and obtain a visually clear solution was measured in seconds.

176 *Particle morphology analysis.* A Scanning Electronic Microscopic HITACHI High  
177 Technologies (S-3500N, Tokyo, Japan) was used to evaluate morphology and particle  
178 size of spray-dried powders. Samples were fixed in adhesive tape placed into SEM  
179 stubs before visualization. The sample surface was covered during 120 seconds by a  
180 thin gold layer, being conductive. SEM worked with a voltage of 5 kV. Lens was placed  
181 at 8 mm from the sample. Images were obtained from representative zones of samples  
182 at a 5000 magnification.



183 *Statistical analysis.* All spray-drying experiments and analytical measurements were  
184 done by triplicate. The mean values, standard deviations, and analysis of variance  
185 (ANOVA) were calculated with Minitab statistic software, version 13.2 (Minitab Inc.,  
186 State College, PA, USA). Mean comparisons were performed using Tukey's test ( $p$   
187  $\leq 0.05$ ).

188

### 189 **3. Results and discussion**

#### 190 *3.1. Temperature profiles of spray dryer operation with only drying air (no atomization)*

191 Initial studies to evaluate temperature profiles outside and inside of spray dryer were  
192 run with no atomization using inlet air at 200 or 120 °C. A thermographic camera  
193 located perpendicular to position 3 of the drying chamber (Figure 1C) measured  
194 surface temperatures from the beginning of inlet air heating until reaching steady state  
195 conditions. Figure 2ABC shows thermographic images of surface temperatures after 5,  
196 15 or 30 minutes of operation, using drying air at 200 °C. In Figure 2C drying chamber  
197 at steady state showed a first upper cold zone from top to about 6 cm height. It is cold  
198 because hot air flow enters only through the central part of the cylinder. Then a hot  
199 second zone is observed due to the impact of hot air flow into the wall, and finally there  
200 is a third zone with a descendent gradient of temperature as air flow travels to the exit.  
201 The highest surface temperatures in the equipment were always measured in the  
202 cyclone surface and the lowest in the collection vessel. Zoom images of the top of  
203 drying chamber taken with the camera perpendicular to different positions from 1 to 4,  
204 showed that temperatures in this zone were different at the same height (Figure 2DEF).  
205 There were two hot dots in positions 2 and 3, which means that two inlet air flows hit at  
206 similar rates at a cylinder height of 10 cm (H1). Positions 4 and 5 were cooler, and  
207 position 1 was the coolest. It can be explained by the geometry of top air inlet (Figure

208 1B). These findings suggest that three main air jets process about the central axis and  
209 two of them are directed to the left part of the cylinder, opposite to air exit (positions 2-  
210 3). Turbulent flow is responsible of the location of the different recirculation zones  
211 between the main jets and the chamber wall observed in the thermographic images.

212 Figure 2G shows the evolution of surface temperatures on drying chamber at positions  
213 H1-H4 and cyclone. Steady-state temperatures were achieved after 25 minutes. Time  
214 to reach steady state temperatures were similar at the different heights of the drying  
215 chamber, however cyclone stabilize its surface temperatures sooner due to its higher  
216 air flow rates. The small radii of the cyclone chamber and exit duct cause a greater  
217 resistance to air flow through the cyclone and hence a greater pressure drop across it  
218 (Maury et al., 2005).

219 Experiments done using an air inlet temperature of 120 °C showed similar images of  
220 surface temperatures, and upper hot dots in the same positions. Steady state  
221 conditions were achieved a bit later, after 30 minutes. The gradient of final surface  
222 temperatures from positions H1 to H4 was lower (11.6 °C, from 65.8 to 54.2 °C) than  
223 obtained previously with air at 200 °C (22.4 °C, from 95.7 to 73.3 °C). An increase of air  
224 inlet temperature of 80 °C resulted in a 20-30 °C increase on surface temperatures.  
225 Heat loss can explain that the wall temperatures of drying chamber were not as high as  
226 could be expected when using the inlet drying air temperature of 200 °C.

227 Measurements of air temperatures inside drying chamber are closely related with  
228 the surface temperatures of drying chamber previously explained. They can offer  
229 interesting information about expected air flow pattern and its effect on bioactive  
230 compounds thermoinactivation. Figure 3 shows a filled contour plot of temperatures  
231 measured with thermocouples located inside the drying chamber in the plane of air  
232 outlet. Equipment run with inlet air at 200 °C, and temperatures were measured once

233 steady state was reached. The highest temperature values were obtained in the upper  
234 central part close to the inlet of hot air. It had only a 10 cm length, due to the turbulent  
235 flow inside the chamber. This is an important hot zone for drying. Contour plot also  
236 revealed that temperatures were not symmetric with central axis at the same height of  
237 cylinder, being lower in the half cylinder zone of air outlet and confirmed the turbulent  
238 air pattern inside the drying chamber. Presence of different zones of temperatures  
239 close to wall were quite coincident with the temperature map obtained in previous  
240 thermographic study. Figure 4 shows the evolution of temperatures measured in the  
241 half cylinder zone of air exit, working with inlet air at 200 °C and 120 °C. In both cases,  
242 temperatures located at inner positions of cylinder (0.5, 2 cm) decrease when air  
243 descend to its exit. This is the position of the hot inlet air jets. However, in the middle  
244 and outer positions (4, 6 cm) temperatures raise up to 15-20 cm (hot zone), and then  
245 stabilized up to 39 cm. Temperature profiles agrees with the expected complex  
246 dynamic of turbulent air flow inside the drying chamber. The values of temperature at  
247 44 cm close to air exit were around 151 and 90 °C using air at 200 and 120 °C,  
248 respectively. These are like air outlet temperatures of 152 and 91 °C measured with the  
249 outlet thermopar of the spray drier working with air at 200 and 120 °C, respectively.

250 Air flow patterns and presence of significant air flow instabilities has been explained  
251 with turbulence models using computational fluid dynamics (CFD) (Kuriakose &  
252 Anandharamakrishnan, 2010). Initial CFD studies with a Büchi B-290 bench-scale  
253 spray dryer reported that the k-epsilon turbulence model was the most suitable to  
254 predict flow behavior in the dryer (Pin et al., 2014). A more complete work of Pinto et  
255 al., 2014 showed CFD simulations of air flow patterns in a Büchi B-290 spray dryer with  
256 recirculation zones fluctuating in size between the main central jet and the chamber  
257 walls. These authors described central asymmetry, so that half cylinder located close to

258 air exit had a different pattern than its opposite half cylinder. While half cylinder close to  
259 air exit had a main recirculation zone up to H3, and a second until H4, its opposite half  
260 had a first zone located approximately up to a 4 cm height, a second between H1-H2, a  
261 third between H2-H3, a fourth between H3-H4, and a fifth from H4 until the end.  
262 Temperatures profiles and thermographic studies obtained in this work during drying  
263 chamber operation at steady state agrees well with CFD simulations results of Pinto et  
264 al., 2014.

265

### 266 *3.2. Temperature profiles of spray dryer operation with water in cocurrent and mixed* 267 *flow*

268 Atomizing cone angles were measured changing the values of atomizing air and water  
269 flow rates. Cone angle depends mainly on atomizing air flow rate, it changes from 80°  
270 at 0.25 m<sup>3</sup>/h (2 cm), to 50° at 0.36 m<sup>3</sup>/h (3 cm), 30° at 0.47 m<sup>3</sup>/h (4 cm), and 28° at 0.60  
271 m<sup>3</sup>/h (5 cm). Measurements in cm are related with the height of the rotameter scale  
272 used in the equipment. So, its suggested to use flow rates in the interval 0.47-0.6 m<sup>3</sup>/h,  
273 because atomization with cone angles higher than 30 °C would hit the wall of drying  
274 chamber (Figure 1). No remarkable effect of drying air flow rate on cone angle was  
275 observed.

276 In the following experiments, water was sprayed under cocurrent or mixed flow  
277 conditions using constant values of 0.47 m<sup>3</sup>/h of air flow rate and 0.5 L/h of water flow  
278 rate. Equipment working with inlet air temperatures of 200 or 120 °C reached steady  
279 state conditions after 20 minutes, then thermographic images were taken locating the  
280 camera perpendicular to position 3 of the drying chamber (Figure 5). In both cases is  
281 observed a descending gradient of temperature in the direction of air exit. Temperature  
282 gradients from H1 to H4 were smaller with cocurrent flow in comparison with mixed

283 flow. In mixed flow water enters from the bottom upwards and temperatures reached at  
284 top positions are slightly higher. The lowest gradient H1-H4 was 2.7 °C with cocurrent  
285 flow, and the highest 13 °C with mixed flow, both at 200 °C. Temperatures in lower  
286 zones are lower with mixed flow, thus cyclone surface temperatures are about 2.2 °C  
287 lower than those of cocurrent flow.

288 Temperature profiles were measured inside drying chamber in the half cylinder zone  
289 of air outlet. Figure 6 shows temperatures at steady state conditions obtained during  
290 spray drying of water under cocurrent or mixed flow conditions using inlet air  
291 temperatures of 200 or 120 °C. Water condensate in thermocouples located close to  
292 nozzle tip so these temperature values were not included in graphs. Working with  
293 cocurrent flow and 200 °C, temperatures measured inside the drying chamber ranged  
294 between 92 and 95 °C. These results are in good agreement with temperatures of glass  
295 surface where temperature gradient H1-H4 was only 2.7 °C. At 120 °C temperature of  
296 outer thermocouples were higher than central ones, and temperature range inside  
297 drying chamber was wider, in accordance with temperatures measured of glass  
298 surface. In case of mixed flow and using an inlet air at 200 °C, temperatures of  
299 thermocouples were higher in the central core in the top zone of the cylinder. However,  
300 from 14 cm onwards temperatures were lower in the center due to presence of sprayed  
301 water drops. A similar pattern was observed using inlet air at 120 °C. Temperature  
302 gradients H1-H4 were between 62 and 98 °C and are in good agreement with  
303 temperatures of glass surface where it was found the widest temperature difference  
304 between H1-H4. With mixed flow and air at 120 °C temperatures inside drying chamber  
305 were lower than 40 °C between H1-H4, and the lowest of the different conditions  
306 studied. These values also agree with the lowest surface temperatures measured with  
307 mixed flow and 120 °C.

308 Thermal studies show how air flow patterns inside drying chamber determine  
309 temperature profiles, and spray drying of bioactive compounds are expected to proceed  
310 at slightly higher temperatures with cocurrent flow versus mixed flow.

311

### 312 *3.3. Spray drying of bioactive compounds under cocurrent and mixed flow patterns*

313 Table 1 presents the results obtained of spray drying of bioactive compounds using  
314 cocurrent and mixed flow with inlet drying air at 200 and 120 °C. The drying yields were  
315 always significantly higher at 200 °C in comparison with 120 °C. In addition, spray  
316 drying at 200 °C led to particles of significant lower enzyme activity, higher bulk  
317 densities, lower water contents, and lower dissolution rates. These effects of  
318 temperature were independent of bioactive compound or flow pattern used. This  
319 dependence of temperature with drying yield is related to particle drying trajectories. If  
320 drying proceed insufficiently before impact with the wall, the particles can adhere, and  
321 hence drying yield is reduced. In case of spray drying trehalose (Maury et al., 2005) or  
322 soymilk (Nguyen et al., 2018) under cocurrent flow, higher drying yields and lower  
323 water contents were obtained increasing inlet temperatures.

324 Comparing the effect of pattern flow, cocurrent flow conditions presented with all  
325 bioactive compounds studied significant higher drying yields versus mixed flow,  
326 especially at 120 °C. The highest drying yield of 75.5% was achieved with riboflavin-5-  
327 phosphate with cocurrent flow and 200 °C, and the lowest 32% with the enzyme  $\alpha$ -  
328 amylase with mixed flow and 120 °C. Operation conditions using mixed flow at 120 °C  
329 were not adequate for spray drying because of the low drying yields obtained. During  
330 spray drying of the two colorants no color loss was observed, and color yields were  
331 close to 100% under all spray drying conditions. Published studies also showed no  
332 significant changes in color parameters of beetroot powders with maltodextrin working

333 under cocurrent flow at 160 °C using lab-scale or semi-industrial spray dryers (Koul et  
334 al., 2002; Janiszewska, 2014). In this work beetroot betalains were stable even under  
335 operation with mixed flow and air at 200 °C. However,  $\alpha$ -amylase enzyme activity was  
336 significantly affected by spray drying conditions. The best activity retention of 99.2%  
337 was achieved under cocurrent flow conditions using the inlet air temperature of 120 °C,  
338 while lowest activity retention was 82.9% using mixed flow and 200 °C. Operation at  
339 200 °C favor enzyme deactivation. Other authors spray dried  $\alpha$ -amylase with lab scale  
340 equipments using cocurrent flow, and found a preservation of 83% of the initial activity  
341 at 145 °C (de Jesus & Filho, 2014), and a range from 91.8 to 51.9% using air  
342 temperatures from 160 to 220 °C (Samborska & Witrowa-Rajchert, 2005). In our  
343 experimentation, spray drying was an adequate technique to obtain powders of  
344 bioactive compounds as far as residence times of drying particles were short enough to  
345 minimize the denaturing effect of high temperatures, even with mixed flow.

346 Effect of flow pattern on particle properties as bulk density, water content or  
347 solubility were mainly dependent on bioactive compound properties. Powders of a pure  
348 compound as riboflavin-5-phosphate have significant higher bulk densities, lower water  
349 content and lower dissolution rates, under mixed flow operation at a similar  
350 temperature. However, flow pattern has no significant effect on densities, water content  
351 and dissolution rates of red beet and  $\alpha$ -amylase powders. As an exception under mixed  
352 flow, red beet powder also shown lower water content and lower dissolution rates using  
353 drying air at 200 °C, and  $\alpha$ -amylase powders a higher bulk density at 120 °C. The  
354 higher the bulk density, the lower the solubility of a powder is referenced by Fazaeli et  
355 al., 2012 for black mulberry powders.

356 Figure 7 compares the effect of flow pattern on the morphology of bioactive  
357 compounds obtained at 200 °C. All particles have a spherical shape with diameters

358 ranged between 1-7  $\mu\text{m}$ . No general differences in size or morphology for riboflavin 5  
359 phosphate and red beet could be attributed to flow pattern used. Riboflavine 5  
360 phosphate particles are small hollow spheres that have collapsed. Red beet powders  
361 had smooth surfaces and higher particle sizes, probably due to stickiness problems  
362 caused by sugars with low glass transition temperatures that are present in this vegetal  
363 juice. This morphology is well described in bibliography (Fazaeli et al., 2012; Miravet et  
364 al., 2016). Particles of  $\alpha$ -amylase prepared by cocurrent flow are wrinkled, while  
365 prepared by the mixed flow appeared to be collapsed. The shape and size of particles  
366 are related to drying time and evaporation rate (Alamilla-Beltran et al., 2005), so it is  
367 feasible that particle trajectories during mixed flow facilitate water evaporation, resulting  
368 collapsed particles of lower water content.

369

## 370 **Conclusions**

371 Thermographic studies of the mini spray dryer working with hot air and without  
372 atomization showed asymmetric profiles of temperatures outside and inside of the  
373 drying chamber at steady state and corroborated the expected turbulent air flow  
374 conditions. There was a fast central core and different air recirculation zones inside the  
375 drying chamber related to the geometric position of inlet air flow and the direction of air  
376 exit. When water was spray dried in the equipment under cocurrent or mixed flow  
377 conditions thermal studies showed a predictable sharp decrease of temperature in the  
378 drying chamber, and slightly lower values were found with mixed flow. Water solutions  
379 of three heat labile bioactive compounds were spray dried using both flow patterns, and  
380 while colorant powders of riboflavin-5-phosphate and betalains maintained 100% its  
381 color,  $\alpha$ -amylase retain more than 82.9% of its enzyme activity. It means that even  
382 when running with a high air temperature of 200 °C it was feasible to produce powders



383 of heat labile compounds. Operation with cocurrent flow achieved higher drying yields  
384 versus mixed flow. Properties of bioactive powders were influenced by flow pattern.  
385 Bulk density was higher and water content lower with mixed flow. Results of this work  
386 encourage potential users to obtain novel food powders maintaining its bioactive  
387 compounds by spray drying.

388

### 389 **Funding**

390 This research did not receive any specific grant from funding agencies in the public,  
391 commercial, or not-for-profit sectors.

392

393

### 394 **References**

395 Alamilla-Beltrán, L., Chanona-Pérez, J.J., Jiménez-Aparicio, A.R., & Gutiérrez-López,  
396 G.F. (2005). Description of morphological changes of particles along spray drying.  
397 *Journal of Food Engineering*, 67, 179–184.

398 Barbosa, J., & Teixeira, P. (2017). Development of probiotic fruit juice powders by  
399 spray-drying: A review. *Food Reviews International*, 33, 1525-6103.

400 Cal, K., & Sollohub, K. (2010). Spray Drying Technique. I: Hardware and Process  
401 Parameters. *Journal of Pharmaceutical Sciences*, 99, 575–586.

402 Choe, E., Huanf, R., & Min, D.B. (2005). Chemical reactions and stability of riboflavin in  
403 foods. *Journal of Food Science*, 70, 28–36.

404 de Jesus, S.S., & Filho R.M. (2014). Drying of  $\alpha$ -amylase by spray drying and freeze-drying –  
405 a comparative study. *Brazilian Journal of Chemical Engineering*, 31(3), 625 - 631.

406 Fang, Z., & Bhandari, B. (2017). Spray Drying of Bioactives. In Y.H. Roos, Y.D. Livney  
407 (Eds.), *Engineering Foods for Bioactives Stability and Delivery* (pp. 261-284). Springer,  
408 New York.

409 Fernández-López, J.A., Angosto, J.M., Giménez, P.J., & León, G. (2013). Thermal  
410 Stability of Selected Natural Red Extracts Used as Food Colorants. *Plant Foods for*  
411 *Human Nutrition*, 68,11–17.

412 Fazaeli, M., Emam-Djomeh, Z., Ashtari, A.K., & Omid, M. (2012) Effect of spray drying  
413 conditions and feed composition on physical properties of black mulberry juice  
414 powder. *Food and Bioproducts Processing*, 90, 667-675.

415 Focaroli, S., Mah, P.T., Hastedt, J.E., Gitlin, I., Oscarson, S., Fahy, J.V., & Healy, A.M.  
416 (2019). A Design of Experiment (DoE) approach to optimise spray drying process  
417 conditions for the production of trehalose/leucine formulations with application in  
418 pulmonary delivery. *International Journal of Pharmaceutics*, 562, 228-240.

419 Gharsallaoui, A., Roudaut, G., Chambin, O., Voilley, A., & Saurel, R. (2007).  
420 Applications of spray-drying in microencapsulation of food ingredients: An overview.  
421 *Food Research International*, 40, 1107–1121.

422 Guerin, J., Petit, J., Burgain, J., Borges, F., Bhandari, B., Perroud, C., Desobry, S.,  
423 Scher, J., & Gaiani, C. (2017). *Lactobacillus rhamnosus* GG encapsulation by spray-  
424 drying: Milk proteins clotting control to produce innovative matrices. *Journal of Food*  
425 *Engineering*, 193, 10–19.

426 Gupta, R., Gigras, P., Mohapatra, H., Goswami, V.K., & Chauhan, B. (2003). Microbial  
427  $\alpha$ -amylases: a biotechnological perspective. *Process Biochemistry*, 38, 1599-1616.

428 Janiszewska, E. (2014). Microencapsulated beetroot juice as a potential source of  
429 betalain. *Powder Technology*, 264, 190-196.

430 Keharom, S., Mahachai, R., & Chanthai, S. (2016). The optimization study of  $\alpha$ -  
431 amylase activity based on central composite design-response methodology by  
432 dinitrosalicylic acid method. *International Food Research Journal*, 23, 10-17.

433 Keshani, S., Daud, W.R.W., Nourouzi, M.M., Namvar, F., & Ghasemi, M. (2015). Spray  
434 drying: An overview on wall deposition, process and modeling. *Journal of Food*  
435 *Engineering*, 146, 152–162.

436 Khan, M.I. (2016). Stabilizations of betalains: A review. *Food Chemistry*, 197, 1280-  
437 1285.

438 Koul, V.K., Jain, M.P., Koul, S., Sharma, V.K., Tikoo, C.L., & Jain, S.M. (2002). Spray  
439 drying of beet root juice using different carriers. *Indian Journal of Chemical*  
440 *Technology*, 9, 442-445.

441 Kuriakose, R., & Anandharamakrishnan, C. (2010). Computational fluid dynamics  
442 (CFD) applications in spray drying of food products. *Trends in Food Science &*  
443 *Technology*, 21, 383-398.

444 Maury, M., Murphy, K., Kumar, S., Shi, L. & Lee, G. (2005) Effect of process variables  
445 on the powder yield of spray-dried trehalose on a laboratory spray-dryer. *European*  
446 *Journal of Pharmaceutics and Biopharmaceutics*, 59, 565-573.

447 Miller, G.L. (1959). Use of dinitrosalicylic acid reagent for determination of reducing  
448 sugar. *Analytical Chemistry*, 31, 426-428.

449 Miravet, G., Alacid, M., Obón, J.M., & Fernández-López, J.A. (2016) Spray-drying of  
450 pomegranate juice with prebiotic dietary fibre. *International Journal of Food Science*  
451 *and Technology*, 51, 633-640.

452 Murugesan, R., & Orsat, V. (2012). Spray Drying for the Production of Nutraceutical  
453 Ingredients—A Review. *Food and Bioprocess Technology*, 5, 3–14.

454 Nair, A., Khunt, D., & Misra, M. (2019). Application of quality by design for optimization  
455 of spray drying process used in drying of Risperidone nanosuspension. *Powder*  
456 *Technology*, 342, 156-165.

457 Nguyen, D.Q., Nguyen, T.H., Mounir, T.S. & Allaf, K. (2018) Effect of feed  
458 concentration and inlet air temperature on the properties of soymilk powder obtained  
459 by spray drying, *Drying Technology*, 36, 817-829.

460 Pin, C.S., Rashmi, W., Khalid, M., Chong, C.H., Woo, M.W., & Tee, L.H. (2014).  
461 Simulation of spray drying on *Piper betle* Linn extracts using computational fluid  
462 dynamics. *International Food Research Journal*, 21, 1089-1096.

463 Pinto, M., Kemp, I., Bermingham, S., Hartwig, T., & Bisten, A. (2014). Development of  
464 an axisymmetric population balance model for spray drying and validation against  
465 experimental data and CFD simulations. *Chemical Engineering Research and*  
466 *Design*, 9(2), 619–634.

467 Ray, S., Raychaudhuri, U., & Chakraborty, R. (2016). An overview of encapsulation of  
468 active compounds used in food products by drying technology. *Food Bioscience*, 13,  
469 76–83.

470 Samborska, K., & Witrowa-Rajchert, D. (2005). Spray-drying of  $\alpha$ -amylase-The Effect  
471 of Process Variables on the Enzyme Inactivation. *Drying Technology*, 23, 941–953.

472 Schutyser, M.A.I., Perdana, J., & Boom, R.M. (2012). Single droplet drying for optimal  
473 spray drying of enzymes and probiotics. *Trends in Food Science & Technology*, 27,  
474 73-82.

475 Sheraz, A.M., Kazi, S.H., Ahmed, S., Anwar, Z., & Ahmad, I. (2014). Photo, thermal  
476 and chemical degradation of riboflavin. *Belstein Journal of Organic Chemistry*, 10,  
477 1999-2012.

- 478 Singh, B., & Singh-Hathan, B. (2017). Process optimization of spray drying of beetroot  
479 Juice. *Journal Food Science and Technology*, 54(8), 2241–2250.
- 480 Wang, S., Shi, Y., & Han, L. (2018). Development and evaluation of microencapsulated  
481 peony seed oil prepared by spray drying: oxidative stability and its release behavior  
482 during in-vitro digestion. *Journal of Food Engineering*, 231, 1-9.
- 483 Yoshii, H., Buche, F., Takeuchi, N., Terrol, C., Ohgawara, M., & Furuta, T. (2008).  
484 Effects of protein on retention of ADH enzyme activity encapsulated in trehalose  
485 matrices by spray drying. *Journal of Food Engineering*, 87, 34–39.
- 486

Figure 1

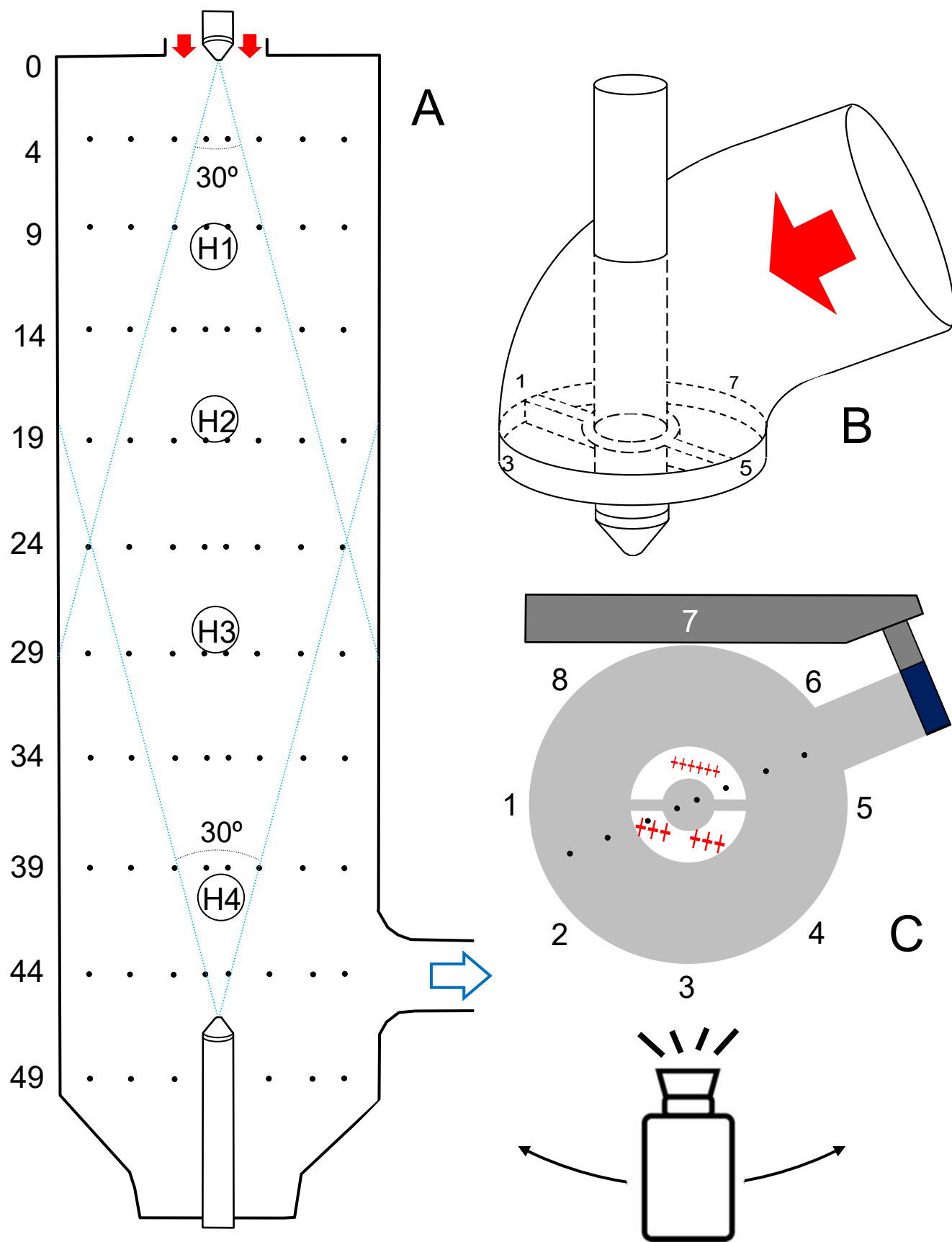


Figure 1

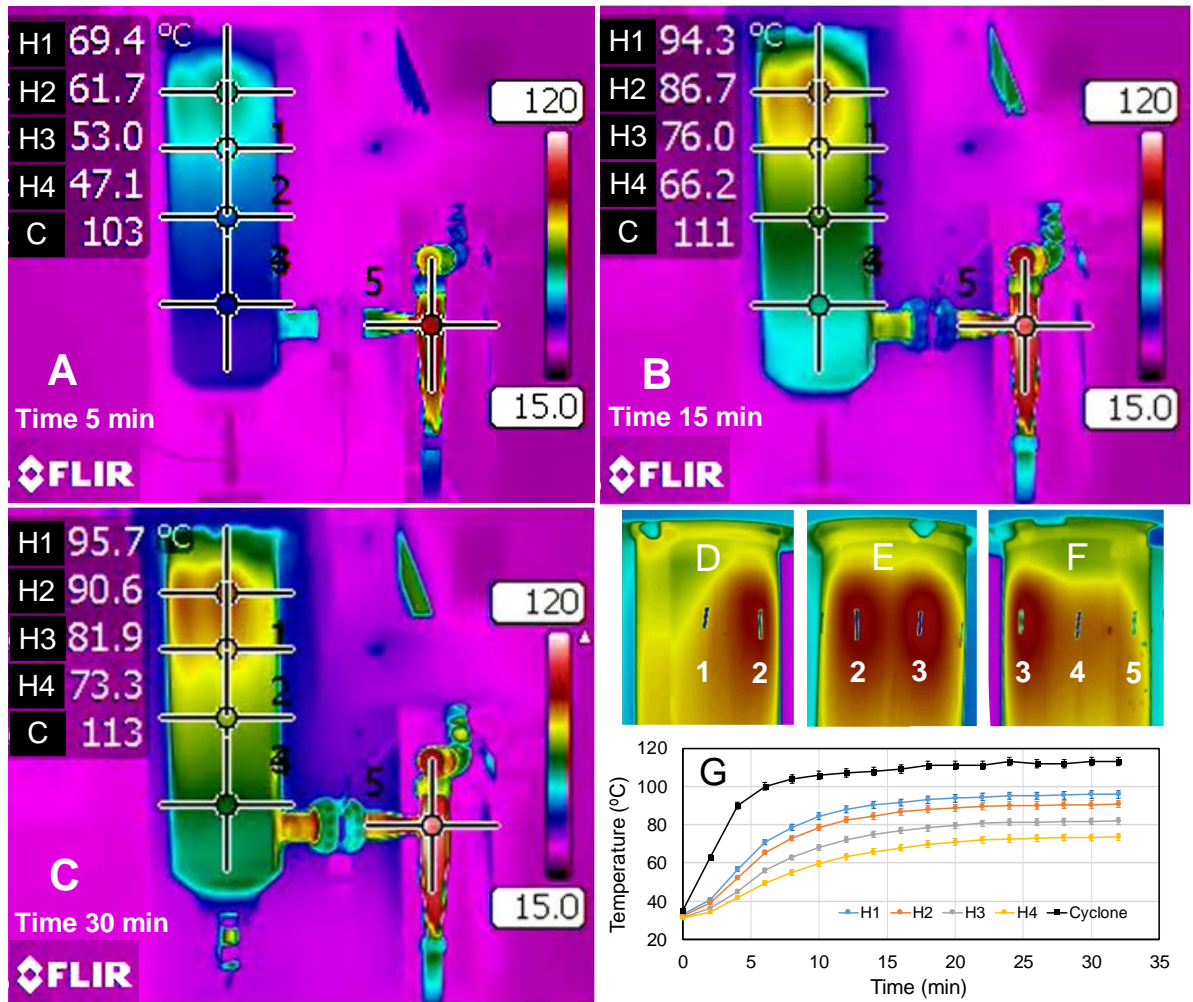


Figure 2

Figure 3

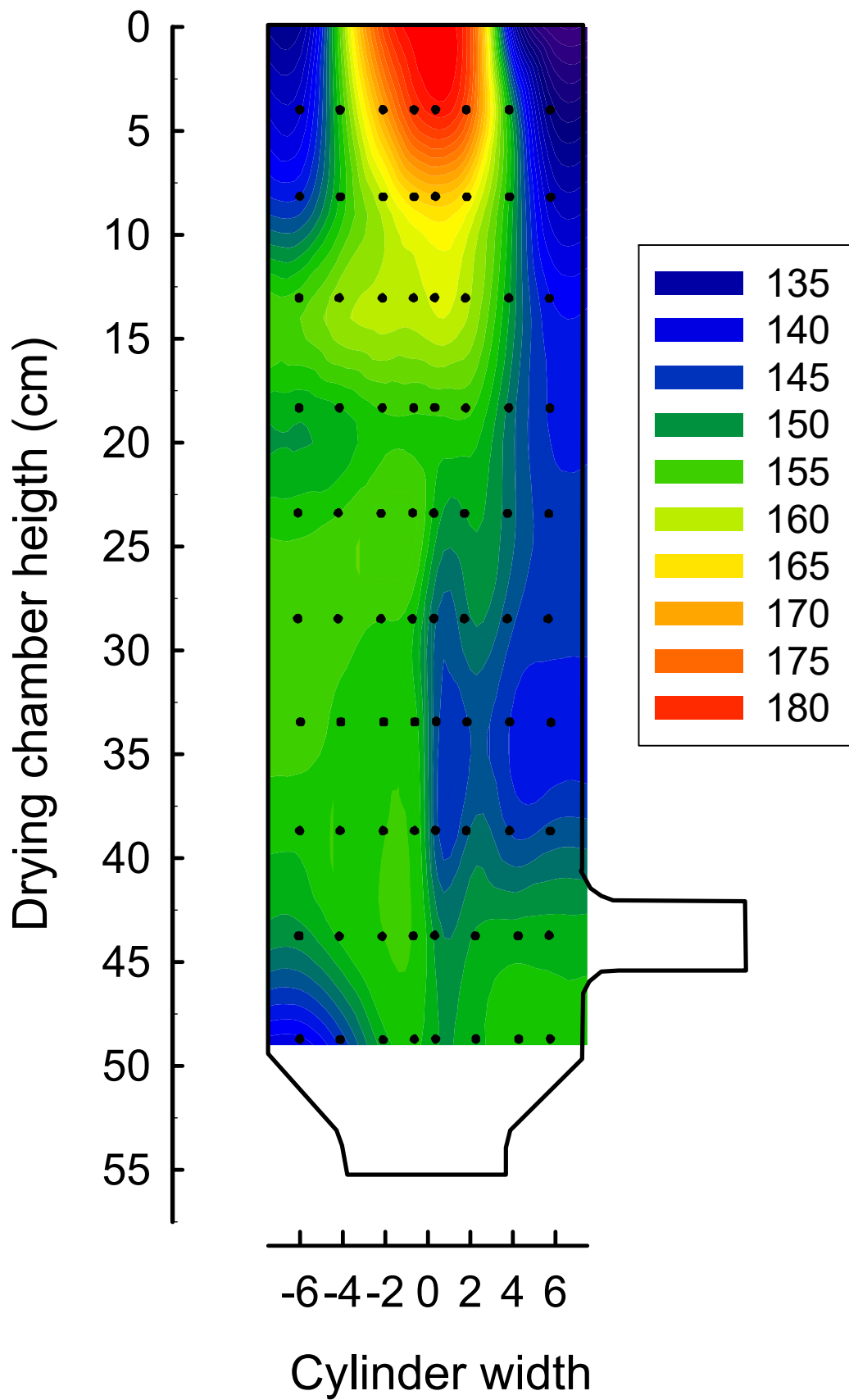
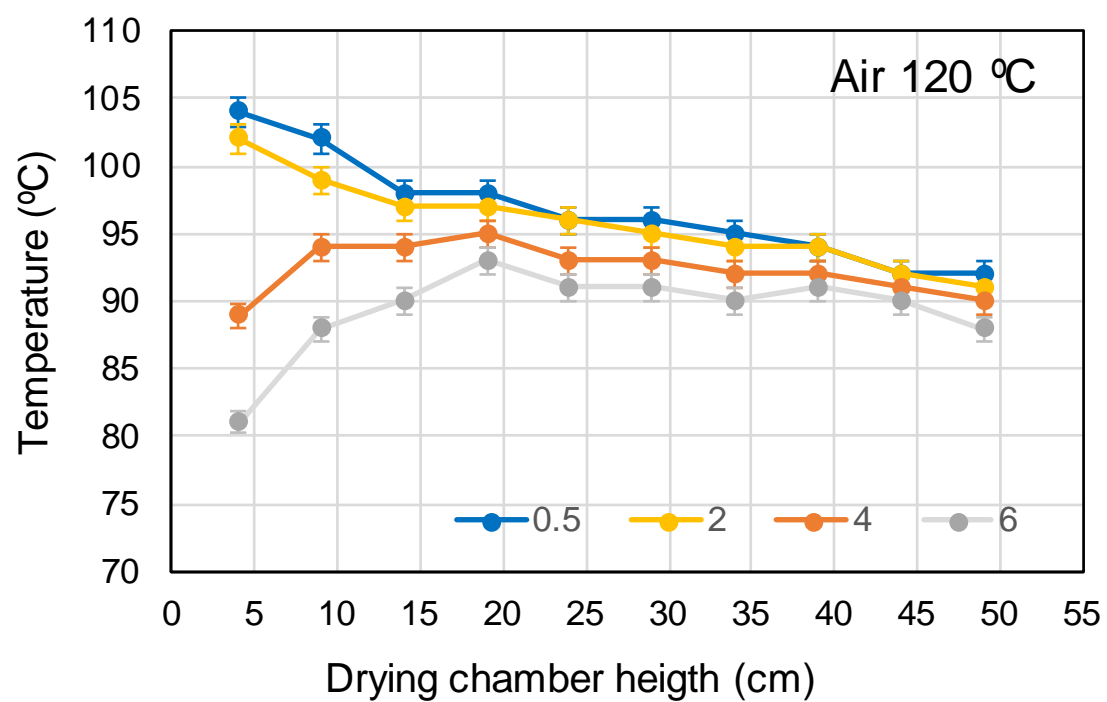
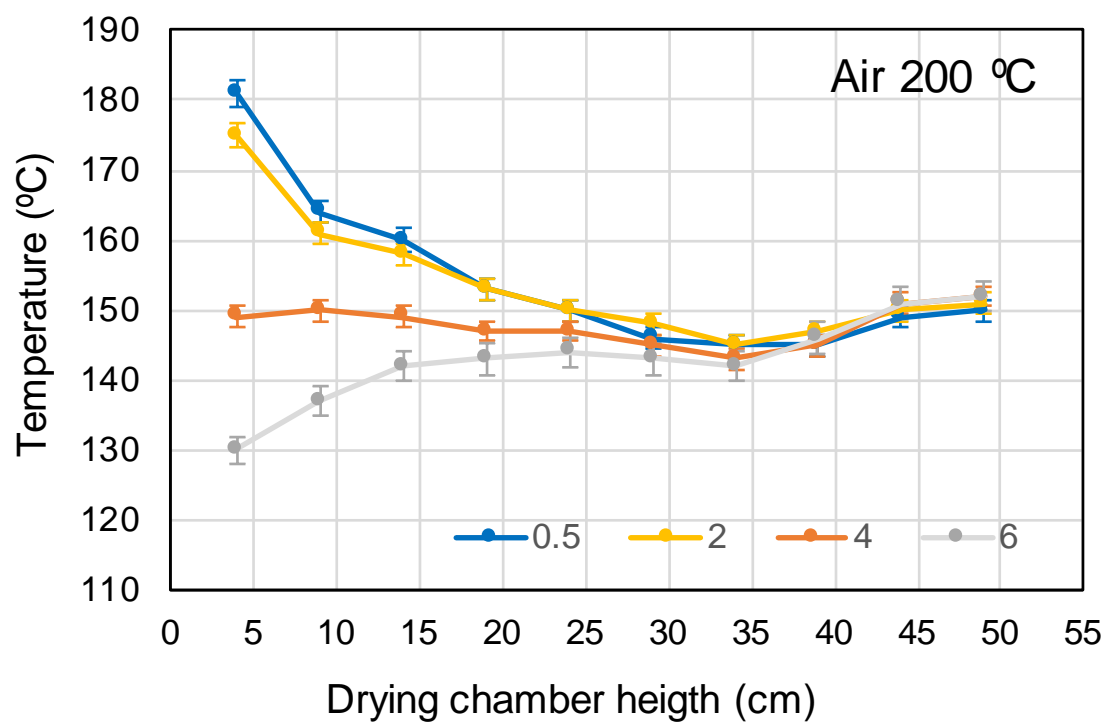


Figure 3





**Figure 4**

Figure 5

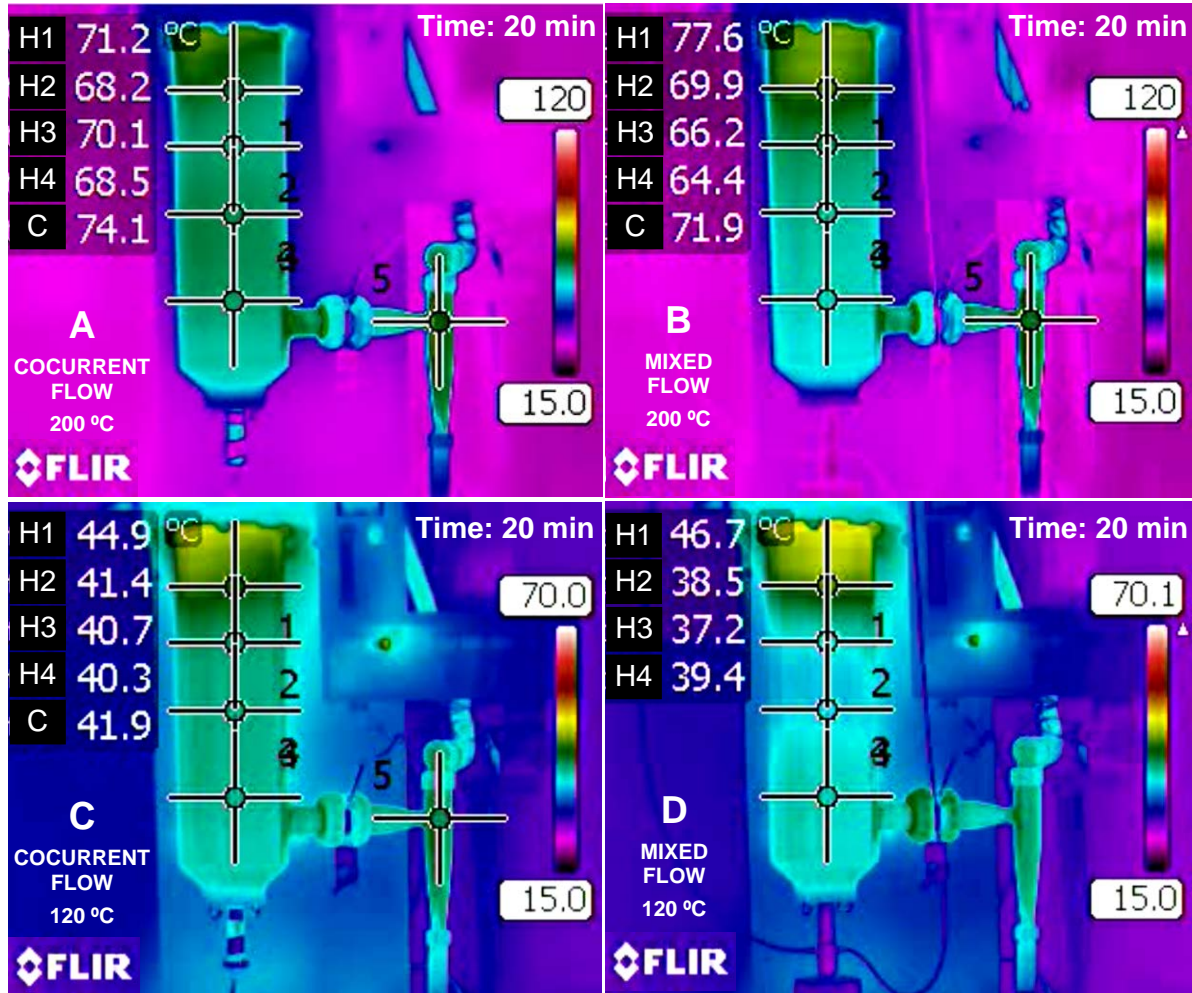


Figure 5

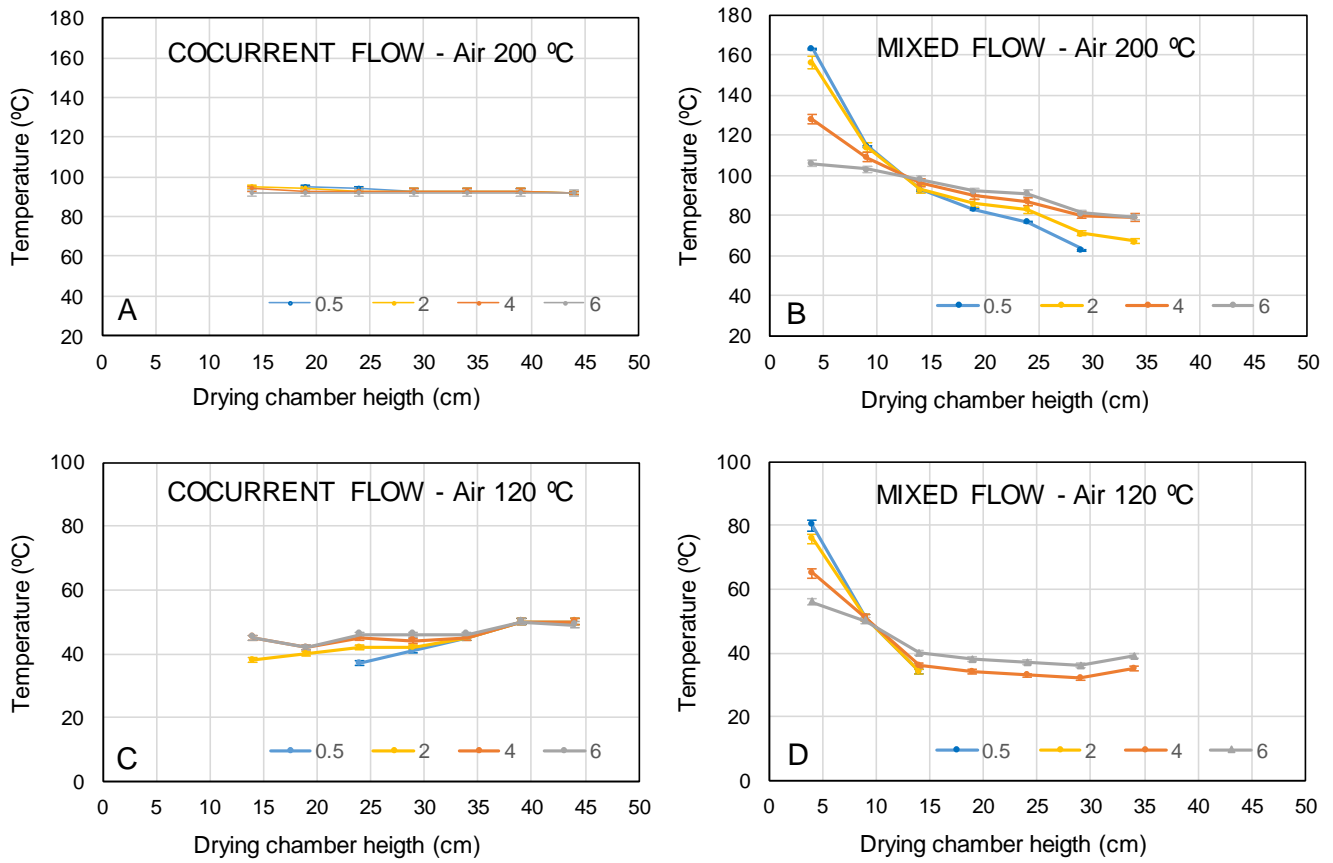


Figure 6

Figure 7

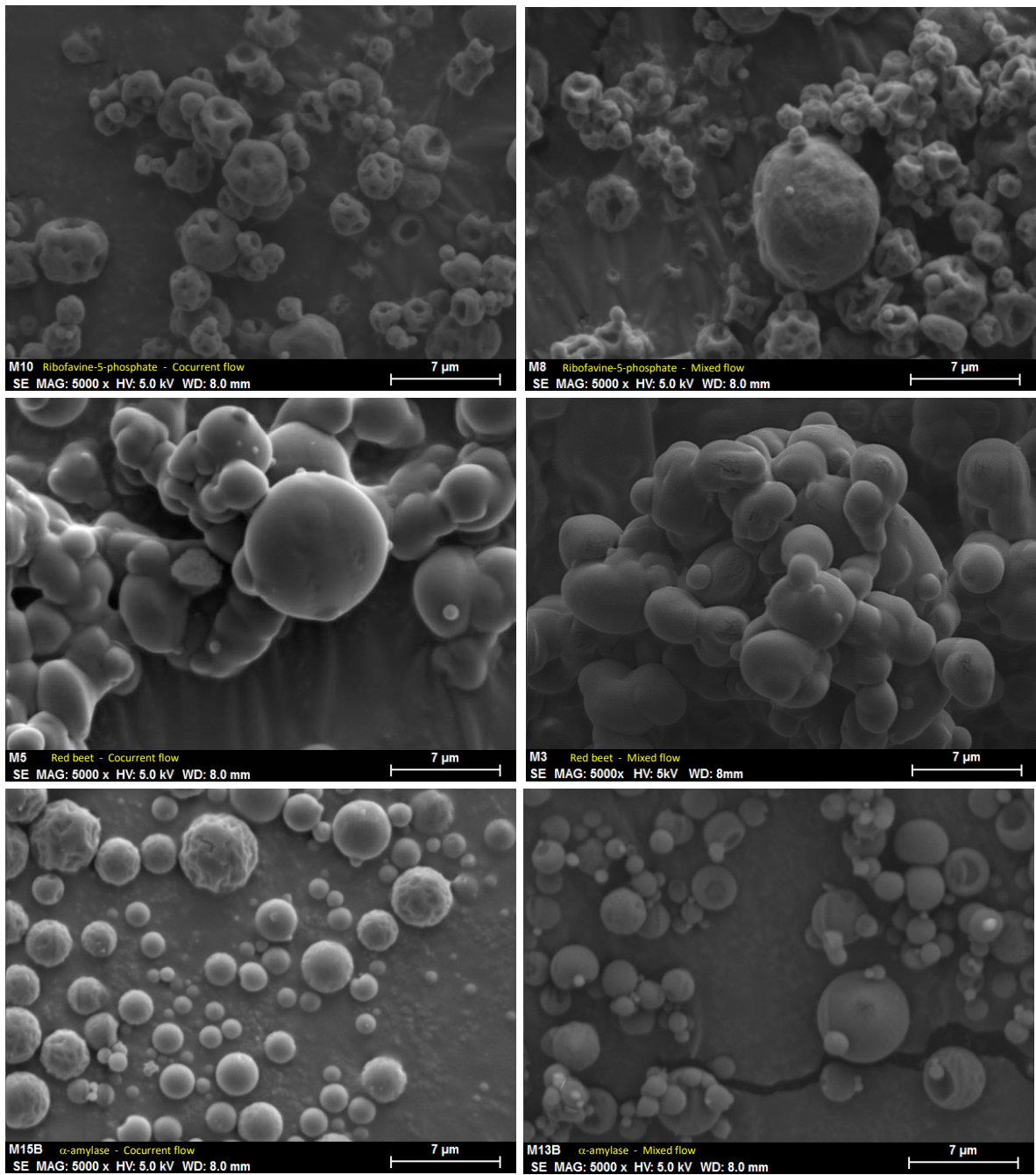


Figure 7

**1 Figure Legends**

2 **Figure 1.** Schematic drawing of the lab-scale spray drying equipment. A. Drying chamber  
3 working with cocurrent or mixed flow, B. Inlet of drying air located in the upper part of the  
4 chamber, C. Up-view of equipment showing inlet of drying air (+) and locations of  
5 temperature thermocouples (●).

6 **Figure 2.** Thermographic images at different times of the spray drying equipment during  
7 operation with inlet air at 200 °C (A: 5 min, B: 15 min, C: 30 min). Zoom images of upper  
8 zone of drying chamber at steady state conditions taken from different camera positions  
9 (D,E,F). Evolution of surface temperatures in positions H1-H4 with time (G).

10 **Figure 3.** Filled contour plot of temperatures inside drying chamber during operation with  
11 inlet air at 200 °C.

12 **Figure 4.** Temperature profiles inside drying chamber at 0.5, 2, 4 and 6 cm of central  
13 axis during operation with inlet air at 200 °C and 120 °C.

14 **Figure 5.** Thermographic images at steady state conditions during spray drying of water  
15 at 200 and 120 °C, with cocurrent (A, C) and mixed flow (B,D), respectively.

16 **Figure 6.** Temperature profiles inside drying chamber at 0.5, 2, 4 and 6 cm of central  
17 axis during spray drying of water at 200 and 120 °C, with cocurrent (A, C) and mixed flow  
18 (B,D), respectively.

19 **Figure 7.** Morphology of bioactive compounds powders obtained by spray drying at 200  
20 °C with cocurrent (A, C, E) and mixed flow (B, D, F). Riboflavine-5-phosphate (A,B), red  
21 beet (C, D) and  $\alpha$ -amylase (E, F).

**Table 1.** Evaluation of bioactive compounds spray drying performance using cocurrent and mixed flow with drying air at 200 and 120 °C

Parameter	Cocurrent flow		Mixed flow	
	200 °C	120 °C	200 °C	120 °C
<b>Riboflavine-5-phosphate</b>				
Drying yield (%)	75.5 ± 1.3 <sup>c</sup>	71.5 ± 1.2 <sup>b</sup>	72.0 ± 1.4 <sup>b</sup>	63.0 ± 1.8 <sup>a</sup>
Color yield (%)	99.8 ± 0.2 <sup>a</sup>	99.7 ± 0.3 <sup>a</sup>	99.6 ± 0.2 <sup>a</sup>	99.8 ± 0.2 <sup>a</sup>
Outlet temperature (°C)	86 ± 1 <sup>c</sup>	43 ± 1 <sup>b</sup>	84 ± 1 <sup>c</sup>	36 ± 1 <sup>a</sup>
Bulk density (Kg/m <sup>3</sup> )	482 ± 7 <sup>b</sup>	348 ± 5 <sup>a</sup>	532 ± 3 <sup>c</sup>	490 ± 8 <sup>b</sup>
Water content (%)	2.1 ± 0.2 <sup>b</sup>	2.9 ± 0.4 <sup>c</sup>	1.6 ± 0.3 <sup>a</sup>	1.9 ± 0.3 <sup>b</sup>
Dissolution test (s)	95 ± 1 <sup>b</sup>	85 ± 2 <sup>a</sup>	105 ± 1 <sup>d</sup>	100 ± 3 <sup>c</sup>
<b>Red beet</b>				
Drying yield (%)	74.5 ± 1.5 <sup>d</sup>	68.5 ± 1.6 <sup>c</sup>	60.0 ± 1.2 <sup>b</sup>	52.5 ± 1.7 <sup>a</sup>
Color yield (%)	99.6 ± 0.3 <sup>a</sup>	99.7 ± 0.2 <sup>a</sup>	99.8 ± 0.1 <sup>a</sup>	99.7 ± 0.3 <sup>a</sup>
Outlet temperature (°C)	89 ± 1 <sup>c</sup>	49 ± 1 <sup>b</sup>	87 ± 1 <sup>c</sup>	41 ± 1 <sup>a</sup>
Bulk density (Kg/m <sup>3</sup> )	558 ± 8 <sup>b</sup>	532 ± 5 <sup>a</sup>	552 ± 7 <sup>b</sup>	537 ± 9 <sup>a</sup>
Water content (%)	2.5 ± 0.3 <sup>b</sup>	2.8 ± 0.3 <sup>b</sup>	1.6 ± 0.3 <sup>a</sup>	2.7 ± 0.3 <sup>b</sup>
Dissolution test (s)	150 ± 3 <sup>c</sup>	75 ± 4 <sup>a</sup>	140 ± 1 <sup>b</sup>	70 ± 5 <sup>a</sup>
<b>α-amylase</b>				
Drying yield (%)	66 ± 0.9 <sup>c</sup>	58 ± 1.3 <sup>b</sup>	65 ± 1.1 <sup>c</sup>	32 ± 1.4 <sup>a</sup>
Activity retention (%)	90.0 ± 0.6 <sup>b</sup>	99.2 ± 0.2 <sup>c</sup>	82.9 ± 0.4 <sup>a</sup>	91.1 ± 0.3 <sup>b</sup>
Outlet temperature (°C)	87 ± 1 <sup>c</sup>	56 ± 0 <sup>b</sup>	85 ± 1 <sup>c</sup>	43 ± 0 <sup>a</sup>
Bulk density (Kg/m <sup>3</sup> )	533 ± 3 <sup>c</sup>	475 ± 7 <sup>a</sup>	544 ± 9 <sup>c</sup>	510 ± 3 <sup>b</sup>
Water content (%)	1.4 ± 0.3 <sup>a</sup>	2.5 ± 0.3 <sup>b</sup>	0.9 ± 0.3 <sup>a</sup>	2.4 ± 0.3 <sup>b</sup>
Dissolution test (s)	50 ± 1 <sup>b</sup>	40 ± 2 <sup>a</sup>	48 ± 1 <sup>b</sup>	35 ± 3 <sup>a</sup>

Means having different letters indicate significant differences within each row at  $p \leq 0.05$  (Tukey's test).