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Bile acid signalling and its role in anxiety disorders

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Anxiety disorder is a prevalent neuropsychiatric disorder that afflicts 7.3%~28.0% of the world's population. Bile acids are synthesized by hepatocytes and modulate metabolism via farnesoid X receptor (FXR), G protein-coupled receptor (TGR5), etc. These effects are not limited to the gastrointestinal tract but also extend to tissues and organs such as the brain, where they regulate emotional centers and nerves. A rise in serum bile acid levels can promote the interaction between central FXR and TGR5 across the blood-brain barrier or activate intestinal FXR and TGR5 to release fibroblast growth factor 19 (FGF19) and glucagon-like peptide-1 (GLP-1), respectively, which in turn, transmit signals to the brain via these indirect pathways. This review aimed to summarize advancements in the metabolism of bile acids and the physiological functions of their receptors in various tissues, with a specific focus on their regulatory roles in brain function. The contribution of bile acids to anxiety via sending signals to the brain via direct or indirect pathways was also discussed. Different bile acid ligands trigger distinct bile acid signaling cascades, producing diverse downstream effects, and these pathways may be involved in anxiety regulation. Future investigations from the perspective of bile acids are anticipated to lead to novel mechanistic insights and potential therapeutic targets for anxiety disorders.

KEYWORDS

anxiety disorders, bile acid signaling, FXR, TGR5, FGF19, GLP-1

1 Introduction

As is well documented, anxiety disorder is a common neuropsychiatric disorder affecting 7.3% to 28.0% of the world's population (1, 2). Its incidence has increased by 2.3 times in recent years due to the impact of COVID-19 (3), making it the sixth-largest disability disease globally. Of note, it is associated with an increase in the risk of suicidal behavior and cardiovascular disease (4). To date, the pathogenesis of anxiety disorders remains elusive.

However, metabolic disorders, encompassing bile acid (BA) disorders, have recently garnered extensive attention in anxiety research (5). Bile acids are multifunctional endocrine factors that govern lipid and energy metabolism by coordinating the activation of FXR and TGR5 to modulate cellular signaling. Additionally, bile acids also communicate with the

central nervous system, traversing the blood-brain barrier (BBB) to bind to their cognate receptors within the brain parenchyma (6).

Emerging evidence from clinical and preclinical studies suggests that bile acid dysregulation may contribute to the development of mental health disorders like anxiety. Earlier clinical investigations have established a direct correlation between bile acid concentration and anxiety symptoms. Moreover, this relationship extends beyond anxiety, as bile acids have been shown to influence anxiety-like behavior in patients with irritable bowel syndrome and other gastrointestinal disorders (7, 8). Earlier studies have also described that bile acids transmit signals to the brain through both direct or indirect pathways and participate in the development of brain diseases, thereby implying a correlation between bile acids and mental illnesses. Elevated serum bile acids have been proven to increase BBB permeability and transmit signals to the central nervous system via FXR, TGR5, etc., eventually elevating the risk of mental disorders (9).However, the underlying mechanisms by which bile acid signals affect the occurrence of anxiety behavior have not been fully elucidated (10). The article is divided into two main sections. The first section provides an overview of the metabolism of bile acids and their different functions and corresponding receptors. The second section consolidates the available evidence on bile acids exerting their effects and transmitting signals through distinct pathways and tissues/organs to influence anxiety, offering a direction for the development of novel treatment strategies from the perspective of bile acids.

2 Synthesis, metabolism, and circulation of bile acids

Primary bile acids such as cholic acid and chenodeoxycholic acid are synthesized from cholesterol in hepatocytes by two pathways, namely the classic or neutral pathway and the alternative or acidic pathway (11) (Figure 1). The former is



FIGURE 1

Classic and alternative pathways for bile acid synthesis. Cholesterol undergoes multiple enzymatic reactions to form primary bile acids CA and CDCA. The solubility of these bile acids is increased, and their cell toxicity is reduced by coupling with glycine or taurine. The primary bile acids are then stored in the gallbladder and secreted into the intestine after food consumption. In the intestine, they are further metabolized into secondary bile acids by the intestinal flora. While approximately 95% of bile acids enter the enterohepatic circulation and return to the liver, the remaining 5% are excreted through feces.



FIGURE 2

Bile acids entering different regions of the brain directly regulate anxiety by binding to FXR and TGR5. (A) CA activates FXR in the prefrontal cortex, which inhibits the expression of NLRP3 inflammasomes and restores AMPAR levels, thereby affecting LTP and synaptic plasticity; (B) CDCA activates FXR on hippocampal neurons and targets CREB and BDNF through a CREB-dependent mechanism, reducing neural function. TUDCA activates TGR5 on hippocampal neurons to relieve neuroinflammation by lowering NF-xB and increasing BDNF levels, thus exerting neuroprotective effects; (C) TUDCA activates TGR5 in astrocytes in the cerebral cortex, upregulates AKT expression, and decreases NF-KB levels; (D) INT-777 activates TGR5/ cAMP/PKA on microglia to counteract neuroinflammation and promote CREB phosphorylation.

initiated by cholesterol 7α -hydroxylase (CYP7A1) in the liver, while the latter is triggered by sterol 27-hydroxylase (CYP27A1) in the liver, macrophages and adrenal glands, and cytochrome P450 46A1 (CYP46A1) in the brain (12). In the classic pathway, CYP7A1 catalyzes the conversion of cholesterol to 7a-hydroxycholesterol, which is then converted to 7α -hydroxy-4-cholesten-3-one by 3β hydroxysteroid dehydrogenase type 7 (HSD3B7). The latter is thereupon converted to cholic acid (CA) by 12a-hydroxylase (CYP8B1) or to chenodeoxycholic acid (CDCA) by Aldo-keto reductase Family 1 Member D1 (AKR1D1). In the alternative pathway, CYP27A1 generates 3β-hydroxy-5-cholenoic acid, which is subsequently converted to CDCA by 7α-hydroxylase (CYP7B1) (11). CDCA is also generated under the action of Cytochrome P450 Family 39 Subfamily A Member 1(CYP39A1) by CYP46A1 (12). Bile acid production is strictly controlled by cytochrome CYP8B1, which determines the amount of cholic acid and chenodeoxycholic acid in the bile acid pool. It is worthwhile emphasizing that in mice, CDCA is converted to ursodeoxycholic acid (UDCA), α-muricholic acid (α -MCA), and β -muricholic acid (β -MCA) by the cytochrome P450 2C70 (CYP2C70) enzyme (13).

Bile acids are conjugated with glycine (primarily in humans) or taurine (primarily in mice) by bile acid-CoA synthase (BACS) and bile acid CoA:amino acid N-acyltransferase (BAAT) before secretion in the liver increases solubility and reduce cellular toxicity (12). This process involves the conjugation of CA to generate glycocholic acid (GCA) or taurocholic acid (TCA) and the coupling of CDCA to yield glycochenodeoxycholic acid (GCDCA) or taurochenodeoxycholic acid (TCDCA). In mice, coupling generates tauro-\alpha-muricholic acid (T\alphaMCA), tauro-\betamuricholic acid (TBMCA), and tauro-ursodeoxycholic acid (TUDCA), which are then transported and stored in the gallbladder by the bile salt export pump (BSEP) (12).

FIGURE 3



Bile acids modulate anxiety by indirectly transmitting signals to the brain via the FXR-FGF19 and TGR5-GLP-1 pathways upon entering the intestine. Bile acids enter the intestine and activate FXR in ileal enterocytes to upregulate FGF19 expression. In addition to returning to the liver through the portal vein, a portion of FGF19 crosses the BBB to interact with FGFRs in the brain and affect the BDNF-TrkB signaling pathway. Bile acids can also stimulate intestinal L cells to release GLP-1. A proportion of GLP-1 reaches the portal vein, and a smaller amount reaches the brain to interact with GLP-1R, thereby affecting the CREB/BDNF signaling pathway.

Following food intake, the duodenum secretes cholecystokinin (CCK), which promotes gallbladder contraction and the release of bile acids into the intestine. Bile salt hydrolase (BSH) secreted by specific intestinal bacteria oxidizes and epimerizes specific hydroxyl groups on bile acids, affecting their physicochemical properties and biological toxicity, and converts primary bile acids to secondary bile acids, including deoxycholic acid (DCA), UDCA, and lithocholic acid (LCA), which promote the metabolism of fat and fat-soluble vitamins (14). Approximately 95% of bile acid molecules remain unbound and proceed to the distal end of the ileum, whereby they are reabsorbed by the ileal epithelium via the apical sodiumdependent transporter (ASBT). The reabsorbed BAs are transported through intestinal epithelial cells to organic solute transporter alpha/beta (OST α/β) on the sinusoidal membrane and are transported back to the liver via sodium (Na+)-taurocholate co-transporting polypeptide (NTCP) and organic anion transporters (OATP) through the portal vein to complete the enterohepatic circulation. Noteworthily, a small proportion of BAs (up to 10%) escape into the systemic circulation without undergoing hepatic reabsorption (15) and reach the brain by simple diffusion (16) or active transport (17) across the BBB.

Bile acid synthesis and enterohepatic circulation primarily occur in the liver, gallbladder, intestine, and brain. Evidence of

bile acid production in the brain has also emerged. In addition to those taken up from systemic circulation, BAs synthesized by the brain have been detected in rodent and human brains. For instance, CA, CDCA, and deoxycholic acid (DCA) were detected in rat brain tissue cytoplasm (18), whereas other bile acids were identified in the cortex of Alzheimer's disease patients, including glycocholic acid (GCA), LCA, and UDCA, among others (19). However, the pathological and physiological processes involved in brainsynthesized bile acids are currently unclear, making it difficult to describe their function.

3 Various bile acids and corresponding receptors are involved in systemic metabolism and immune regulation

The metabolism of bile acids is contingent upon the interaction between bile acids and their receptors, which are expressed in the liver, intestine, brain, etc. Nuclear receptors include FXR, pregnane X receptor (PXR), vitamin D receptor (VDR), and constitutive androstane receptor (CAR), while membrane receptors include TGR5 and sphingosine-1-phosphate receptor 2 (S1PR2) (20). They strictly govern the synthesis, absorption, and excretion of bile acids. Variations in the source and structure of bile acids impart unique physicochemical properties and functional characteristics that naturally translate into diverse effects on their corresponding receptors. To explore these differences, the functions of various bile acids were detailed based on their receptor targets, as illustrated in Table 1.

3.1 FXR

FXR is a ligand-activated transcription factor that plays a crucial role in regulating bile acid homeostasis and participates in enterohepatic circulation. FXR can be activated by several bile acids, with hydrophobic bile acid CDCA being the most effective ligand for FXR. The order of affinity of bile acids to FXR is as follows: CDCA > DCA > LCA > CA (41). FXR is largely expressed in the liver, intestine, kidney, adrenal gland, etc. (42), as well as in cortical neurons of the brain (43, 44).

Intestinal FXR is a crucial regulatory factor that maintains physiological enterohepatic circulation. Indeed, FXR acts as a

sensor for elevations in bile acid levels (especially CDCA). Specifically, intestinal FXR stimulates fibroblast growth factor 15 (FGF15) in mice or FGF19 in humans located at the distal portion of the ileum. After crossing the portal vein, it activates the liver fibroblast growth factor receptor (FGFR) to inhibit cholesterol 7 α -hydroxylase (CYP7A1) and mediate bile acid synthesis in liver cells (21). This is the primary pathway of bile acid negative feedback. Interestingly, FGF15/19 has been found to enter the systemic circulation and cross the blood-brain barrier to reach the brain, where it binds with FGFR4 in the brain and regulates brain function (6). Meanwhile, FGFR4 has been detected in the hypothalamus and cholinergic neurons located in the nucleus accumbens (45). The bile acid-mediated FXR-FGF15/19 pathway not only establishes a connection between the liver and intestine but also extends to various brain regions.

Hepatic FXR is a key molecule for synthesizing bile acids. Under physiological conditions, activation of FXR by the CDCA and CA primary bile acids plays a decisive role in maintaining hepatic bile acid homeostasis. This process involves hepatic FXR inhibition of 7-ketocholesterol synthesis through the small heterodimer partner (SHP) α Hydroxylase enzyme, resulting in the suppression of bile

TABLE 1 Functional involvement of different bile acids and their corresponding receptors.

Bile Acid Ligands	Receptor	Cellular Localization	Main Function	References
CDCA, DCA, LCA, CA		Intestine	Controlling bile acid synthesis by inhibiting CYP7A1 via FGFs/FGFRs	(21)
CDCA, DCA, LCA, CA	-	Liver	Suppressing CYP7A1 and CYP8B1 via SHP-induced inhibition of bile acid synthesis	(22)
CA, TCA	FYD		Regulation of lipid metabolism by SHP inhibition of SREBP-1c	(23)
CDCA, DCA	FAR		Promoting depression by inhibiting BDNF/TrkB	(24)
CDCA	-	Hippocampus	Regulation of glucose metabolism by enhancing insulin sensitivity in the brain	(25)
CDCA	-	Prefrontal cortex	Relieving depression through the NLRP3/GluA1 signaling pathway	(26, 27)
CA		Kupffer cells	Mediating the inflammatory response through JNK-dependent pathways	(28)
Taurine	_	Sinusoidal Preventing oxidative stress by inducing ENOS Endothelial Cell		(29)
НСА	-	Intestine	Stimulating the secretion of GLP-1 by intestinal L cells and regulating glucose metabolism	(30, 31)
TUDCA	TGR5	Neuron	Regulating Sirtuin3 signaling to protect against apoptosis	(32)
TUDCA	-	Microglia	Upregulating cAMP expression and inhibiting microglial activation	(33)
TCDCA	-	Astrocyte	Through AKT/NF κ B signaling pathway inhibits neuroinflammation	(34)
TLCA	-	Ventricle	Increase fat oxidation and regulate lipid metabolism	(35)
DCA, TDCA, TCA	-	Hypothalamus Activating SNS to promote energy metabolism and reduce fat content		(36)
	PXR, VDR	Liver	Alleviating liver toxicity of LCA through enzymes that regulate bile acid metabolism	(37)
LCA	CAR, PXR, VDR	Intestine	Maintaining bile acid homeostasis by inhibiting CYP7A1 through FGF15/FGF19	(38)
TCA, GCA, GDCA, TDCA, TUDCA	S1PR2	Liver	Promoting liver inflammation through ERK1/2/NF- κ B/COX-2	(39)
TCA		Neuron	Upregulating CCL2 expression and aggravating neuroinflammation	(40)

acid synthesis and prevention of hepatic bile acid accumulation (22). This negative feedback loop involving bile acids and FXR represents the second bile acid regulatory pathway. CA and TCA have also been determined to play key roles in the regulation of lipid metabolism. In other words, these bile acids activate the FXR-SHP pathway to inhibit Sterol Regulatory Element Binding Protein-1c (SREBP-1c), a transcription factor that controls hepatic lipid biosynthesis (23).

FXR expression has been detected in cortical neurons of both humans and mice. *In vitro* cultured neurons express FXR in their nuclei, whereas *in vivo* neurons express FXR in the cytoplasm (43). Moreover, FXR is present in various brain tissues, such as the hippocampus, cerebellum, and frontal cortex. Experimental data indicate that CDCA and DCA may downregulate hippocampal FXR expression, which in turn increases the level of brain-derived neurotrophic factor (BDNF), thereby exerting anti-depressant effects (24). However, CDCA exerts anti-depressant actions in the mouse prefrontal cortex by upregulating FXR expression to inhibit NOD-like receptor protein 3 (NLRP3) inflammasome activation and increase GluA1 levels (26, 27), highlighting the diverse regional effects of bile acid-FXR interactions in the brain. Additionally, CDCA binding to FXR in the hippocampus of AlCl3-treated rats can also enhance insulin sensitivity (25).

3.2 TGR5

TGR5 is a G protein-coupled receptor implicated in bile acid metabolism (6)and is activated by hydrophobic bile acid LCA, which has the highest affinity for TGR5 among other bile acids such as DCA, CDCA, and CA (6). TGR5 is abundantly expressed in various tissues and cells, including the intestine, gallbladder, liver sinusoidal endothelial cells (SEC), etc. (46), and its expression in the brain has also been gradually recognized (47, 48). It affects glucose and energy metabolism and plays a role in immune regulation.

Activation of TGR5 signaling in Kupffer cells and SEC has been noted to promote anti-inflammatory responses. Specifically, CA principally plays a pro-inflammatory role in Kupffer cells, driving the production of pro-inflammatory cytokines in the liver through the TGR5/c-Jun N-terminal kinase (JNK)-dependent pathway (28). As a protective bile acid, taurine is more likely to bind to TGR5 in SECs to stimulate endothelial nitric oxide synthase (ENOS) and prevent oxidative stress-mediated inflammatory reactions (29).

TGR5 has a wide range of functions in the intestine. Hyocholic acid (HCA) activates TGR5, triggering the secretion of GLP-1 from intestinal L cells to regulate glucose metabolism (30). Notably, HCA has been proposed as a biomarker for glucose metabolism disorders in clinical trials (31). Additionally, only a quarter of GLP-1 enters the portal vein of the liver. Among them, 10-15% enter the body circulation to activate GLP-1 receptors (GLP-1R) located in the terminal bed nucleus, hippocampus, and paraventricular nucleus of

the hypothalamus (6) or transmit signals to CNS through the vagus nerve-brainstem-hypothalamus pathway (6).

TGR5 in the brain is chiefly expressed in cortical neurons, astrocytes, and microglia. Unlike brain FXR, the activation of TGR5 exerts a positive effect on brain function. Known for its neuroprotective effects, TUDCA has been shown to mitigate cell apoptosis through activation of the TGR5/Sirtuin3 signaling axis in neurons (32). Besides, the neuroprotective actions of TUDCA extend beyond its impact on neurons, as it has also been documented to regulate neuroinflammation by modulating the TGR5/cAMP pathway and prevent microglial activation (33). Similarly, taurocholic acid deoxycholic acid (TCDCA), another conjugated bile acid, has been observed to possess antiinflammatory and immune regulatory properties that mainly operate within astrocytes. Its actions are mechanistically linked to the TGR5/threonine kinase (AKT)/nuclear factor kB (NF-kB) signaling axis and suppressing neuroinflammation (34). Comparable to the CNS FXR receptor, TGR5 within the CNS also regulates lipid and energy metabolism. The administration of taurolithocholate (TLCA), the most potent natural TGR5 agonist, through intracerebroventricular infusion in mice has been found to increase fat oxidation and limit fat mass (35). Additionally, three bile acids, namely DCA, taurine deoxycholic acid (TDCA), and TCA, target hypothalamic TGR5 to promote energy metabolism through the sympathetic nervous system (SNS) to reduce fat content (36).

3.3 PXR, CAR, and VDR

Although there is no specific bile acid ligand that binds with CAR, PXR, CAR, and VDR can all promote the clearance of hepatotoxic LCA. Accumulation of LCA leads to activation of PXR and VDR, and CAR acts as an indirect sensor of bile acids to transcriptionally regulate the expression of bile acid-related enzymes and transporters (37). In the intestines, PXR, CAR, and VDR inhibition of CYP7A1 action occur through the FGF15/FGF19 pathway to maintain BA homeostasis (38).

3.4 S1PR2

S1PR2 has been essentially implicated in promoting inflammatory responses, with heightened expression levels observed in the liver. S1PR2 activation by ligands such as TCA, GCA, Glycodeoxycholic acid (GDCA), TDCA, and TUDCA leads to membrane trafficking and activation of ERK1/2 kinases, which results in the nuclear translocation of NF- κ B transcription factors. This nuclear translocation then drives the transcriptional upregulation of cyclooxygenase-2 (COX-2), an enzyme involved in hepatic inflammation (39). In addition to its hepatic expression, S1PR2 is also abundantly expressed in neurons within the CNS. Circulating TCA has been theorized to penetrate the CNS and bind S1PR2 on neurons, potentiating chemokine ligand 2 (CCL2)-induced neuroinflammation and microglia activation, resulting in hepatic encephalopathy that is exacerbated by hepatic dysfunction (40).

4 Signal transduction of bile acids and their receptors in patients with anxiety disorders

Clinical studies have reported that in patients with anxiety disorders, certain metabolically abnormal bile acids excessively activate FXR and promote the secretion of FGF19 in intestinal cells. The latter enters the brain through systemic circulation and participates in the neuroinflammatory process, thereby promoting anxiety (49). A large number of studies have consistently demonstrated that anxiety related to abnormal bile acid metabolism is frequently accompanied by intestinal diseases (7, 8, 10, 50), particularly irritable bowel syndrome (IBS). Therefore, it is essential to emphasize that changes in bile acids are closely related to anxiety disorders in IBS patients. Excessive CA and DCA affect the expression of TGR5 in colonic mucosa, activate the TGR5-JNK pathway of intestinal epithelial cells, and cause an abnormal number and distribution of cell connections, leading to damage to the intestinal mucosal barrier and increased permeability. On the other hand, activating the TGR5/5-hydroxytryptamine (5-HT) signaling axis of intestinal enterochromaffin cells conduces to visceral hypersensitivity, which is particularly relevant to the occurrence of IBS (51, 52). These effects of intestinal mucosal barrier impairment and visceral hypersensitivity due to the influence of the gut-brain axis ascend along the afferent nerves in the intestine to the central nervous system, affecting the release of 5-HT and γ -aminobutyric acid, thereby exacerbating anxiety in IBS patients. Indeed, compelling evidence suggests that bile acids affect anxiety through different pathways (53). As previously mentioned, FXR and TGR5 are the most extensively researched and distinctive receptors of bile acids. These two receptors are widely expressed and play a crucial role in the liver-gut axis and brain. In light of the discussions in Sections 3.1 and 3.2, this article exclusively focused on anxiety and offered a more comprehensive explanation of the mechanisms by which bile acids and their receptor signaling pathways influence anxiety.

4.1 Bile acids transmit signals that affect anxiety via the direct pathways

According to earlier studies, unconjugated bile acids such as CA, DCA, and CDCA (54) can diffuse across the BBB in a concentration-dependent manner. At the same time, experimental studies have identified a positive correlation between the levels of CA, CDCA, and DCA in the brain and their serum levels (18). At high concentrations (≥ 1.5 mM), bile acids can function as detergents to dissolve endothelial cell membranes and thus disrupt the lipid layer of the BBB (16). At low concentrations (0.2-1.5 mM), bile acids enhance BBB permeability by a rac1dependent phosphorylation mechanism related to tight junctionassociated proteins (55) and diffuse into the brain. Conjugated bile acids require active transport across the BBB with the help of transport proteins such as NTCP, OATP, OST α/β , and BSEP in the blood-brain barrier and choroid plexus. This is ascribed to the presence of both hydrophilic hydroxyl and carboxyl groups and hydrophobic methyl groups (17, 56, 57). BAs interact with FXR and TGR5 in the brain through the aforementioned mechanisms. Although the primary objective of this process is to transport excess cholesterol from the brain to the circulatory system (58), BAs also affect brain functions such as emotions (58). The regulation of emotions is fundamentally dependent on different parts of the brain cortex, such as the prefrontal cortex and hippocampus. Of note, different bile acid ligands and receptors have varying effects when acting on different parts (Table 2 and Figure 2).

CDCA activates FXR in hippocampal neurons after diffusing through the BBB. This activation inhibits the activity of the cAMP response element-binding protein (CREB) and the expression of BDNF (25). The former is a transcription factor that regulates gene transcription and is an important transcription element necessary for long-term memory and neuronal survival. Furthermore, it can regulate the transcription and expression of BDNF and induce anxiety-like behavior. Contrastingly, the latter is a protein with neurotrophic effects that can enhance learning and memory abilities. It is also a downstream molecule of CREB and affects anxiety and other mental illnesses. Studies have evinced that CREB and BDNF expression levels are down-regulated in the hippocampus of rats with anxiety-like behavior (59, 60), suggesting that the overactivation of FXR in hippocampal neurons by excessive CDCA may target and inhibit the CREB/ BDNF pathway via a CREB-dependent mechanism to impair neural function and promote anxiety.

TABLE 2	Different	bile	acids	transmitting	signals	to	affect	anxiety.
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Bile Acid Ligands	Cellular Localization	Pathways	Mechanisms	References
CDCA	Hippocampal neurons	FXR/CREB/BDNF	CREB dependency	(25, 59, 60)
СА	Prefrontal cortex	FXR/NLRP3/AMPARs	Synaptic plasticity	(27, 61, 62)
INT-777	Microglia	TGR5/cAMP/PKA/CREB	Microglia activation	(63–66)
TUDCA	Hippocampal neurons	TGR5/NF-ĸB/BDNF	Neuroprotection	(67–71)
	Astrocyte	TGR5/AKT/NFĸB	Neuroinflammation	(34, 72–75)

CA, a natural ligand of FXR (76), can activate FXR in the prefrontal cortex (PFC) and suppress the expression of NLRP3 inflammasome, thereby restoring the level of α -amino-3-hydroxy-5-methyl-4-isoxazole-propionic acid receptors (AMPARs) (27). NLRP3 is an inflammasome sensor protein that aggravates anxiety-like behavior by inducing the activation of microglia in PFC (61, 62). It is noteworthy that NLRP3 also participates in longterm potentiation (LTP) mediated by AMPARs (77), which are tetramers composed of four homologous core subunits. The dynamic expression of AMPARs in the postsynaptic membrane is associated with LTP. Their synaptic transmission efficiency is a crucial factor in the LTP process that is responsible for synaptic plasticity. The weakening of this efficiency can impair LTP (78), reducing synaptic activity and quantity, especially in the PFC, which can cause anxiety (79, 80). It is evident that CA activates the prefrontal cortex FXR and dynamically balances anxiety via the NLRP3 inflammasome/AMPARs signaling pathway.

INT-777 is a modified bile acid analog that acts as a specific TGR5 agonist. The activation of the TGR5-cAMP-protein kinase A (PKA) axis exerts an anti-inflammatory effect on microglia, which in turn promotes the phosphorylation of the target protein CREB. The cAMP/PKA/CREB signaling pathway is closely associated with anxiety (63–66). The anti-anxiety effect of INT-777 is elicited through the activation of microglial cells, which modulate the TGR5/cAMP/PKA/CREB axis.

TUDCA is an agonist of TGR5 and confers neuroprotective effects on the central nervous system. It activates TGR5 in hippocampal neurons to mediate NF- κ B/BDNF signaling and mitigate neuronal apoptosis (67). NF- κ B is a transcription factor that swiftly responds to harmful stimuli and is highly expressed in inflammatory reactions (68). Its expression is negatively correlated with BDNF expression in hippocampal neurons and impairs neuronal development and synaptic plasticity (69–71), resulting in anxiety-like behavior.

TUDCA is known to activate the TGR5/AKT/NF-κB signaling pathway in astrocytes in the cerebral cortex (34). AKT, also referred to as protein kinase B, is a widely expressed protein in the cerebral cortex with a molecular weight of approximately 60 kDa (72). Its upregulation in astrocytes is associated with anxiety-like behavior in mice (73). Moreover, it can lead to NF-κB phosphorylation and participate in the occurrence and development of anxiety disorders (74, 75). TUDCA regulates the TGR5/AKT/NFκB signaling pathway through astrocyte-mediated inflammatory processes, thus playing a role in anxiety.

4.2 Bile acids transmit signals affecting anxiety via indirect pathways

In addition to direct pathways, bile acids can also affect anxiety via indirect pathways. As aforestated, bile acids initiate the FXR-FGF19 and TGR5-GLP-1 pathways following their release into the intestine to transmit signals to the central nervous system. This section aimed to focus on the mechanisms by which these two pathways affect anxiety in the brain (Figure 3).

Activation of FXR by fasting serum CDCA has been shown to independently modulate FGF19 in clinical studies (81, 82). FGF19 is a member of the FGF family that is predominantly expressed in the intestinal epithelial cells located at the terminal ileum (83, 84). FGFs are widely distributed throughout the central nervous system and are known to play a central role in neuronal function, development, and metabolism. Recent studies have revealed that FGFs have a significant impact on the occurrence and development of mental illnesses such as anxiety disorders, depression, schizophrenia, bipolar affective disorder, etc., and are anticipated to serve as novel biomarkers for the diagnosis and prognosis of mental illnesses (85, 86). The intestinal FXR-FGF19 complex partially diffuses into the hepatic portal vein and binds with the auxiliary receptor β -Klotho in the liver to activate FGFRs, thereby mediating the negative feedback pathway of bile acids. The remaining part of the complex subsequently circulates throughout the body and binds with FGFRs of the brain. Among the four types of FGFRs, FGFR4 is mainly distributed in the hypothalamus (87) and cholinergic neurons (45) in the medial habenula, with a lower proportion of β-Klotho receptors (88). FGFR4 mainly participates in emotional regulation (89). An unbalanced FGFs/FGFRs system can cause glucose metabolism disorders, neural inflammation, hypothalamic-pituitary-adrenal axis hyperfunction, BBB damage, neuroplasticity reduction, neuronal apoptosis, etc., which affect the structure and function of the cerebral cortex, hippocampus, hypothalamus, pituitary and other tissues, ultimately resulting in emotional disorder (89). Among them, aberrant glucose metabolism is closely related to anxiety; that is, insulin resistance may play a pivotal role in the development of emotional disorders (90-92). Similarly, HPA axis dysfunction caused by endocrine disorders is an inducing or aggravating factor for anxiety (93-95). Recent studies have shown that regulating FGFR can affect BDNF expression (96) and that the BDNF-TrkB signaling pathway plays an important role in anxiety (96, 97). The overexpression of serum CDCA over-activates intestinal FXR receptors, resulting in an imbalance in the FGFs/FGFRs system that impacts the BDNF-TrkB signal pathway, which finally culminates in anxietylike behavior.

Several studies have indicated that bile acid can stimulate intestinal TGR5 to release GLP-1, with LCA being the most potent agent in this process (98–101). GLP-1 is a type of enterotropic insulin produced by specific intestinal endocrine cells (L cells) and secreted in large quantities after food intake. It can regulate glucose uptake and insulin resistance in the brain, thereby improving neuroinflammation and neurogenesis (102). Additionally, GLP-1 plays a crucial role in synaptic plasticity and emotion regulation (103) and may be a potential key regulator of anxiety behavior (104–106). The indirect TGR5-GLP-1 signaling pathway can interact with GLP-1R in the brain through systemic circulation to affect central nervous system function (6). This interaction can concurrently stimulate CREB/ BDNF to modify brain synaptic plasticity and participate in anxiety regulation (106). Alternatively, it can regulate anxiety emotions through the vagus nerve-brainstem-hypothalamus pathway. The vagus nerve is a regulator of mental illness (107–109), and its incoming fibers stimulate the monoaminergic brain system in the brainstem (107), which then plays a key role in anxiety emotions in the hypothalamus (110).

5 Conclusion and perspectives

Anxiety may be associated with bile acids and their signaling pathways. Different bile acids and their corresponding receptors participate in systemic activities, including metabolic and immune regulation and radiating to tissues and organs such as the brain, liver, and intestines. Recent years have witnessed a growing body of research into the effect of bile acids on brain function, providing valuable insights and opening up new avenues of investigation. Bile acids can directly bind to bile acid receptors in the brain through the blood-brain barrier to induce anxiety-like behavior. They can also form receptor-hormone complexes through the enterohepatic circulation to enter the systemic circulation and bind to corresponding hormone receptors in the brain, inducing anxiety. The mechanism of anxiety disorder is complex and is regulated at various levels besides brain lesions. Targeting bile acids may be an efficient approach for exploring the pathological mechanism underlying anxiety disorders. Despite only specific bile acid signaling pathways being related to anxiety, this approach can broaden our understanding of anxiety for the development of new treatment strategies.

References

1. Szuhany KL, Simon NM. Anxiety disorders: A review. JAMA (2022) 328:2431–45. doi: 10.1001/jama.2022.22744

2. Kim SY, Jeon SW, Lim WJ, Oh KS, Shin DW, Cho SJ, et al. The relationship between serum vitamin d levels, c-reactive protein, and anxiety symptoms. *Psychiatry Investig* (2020) 17:312–19. doi: 10.30773/pi.2019.0290

3. Huang Y, Zhao N. Corrigendum to generalized anxiety disorder, depressive symptoms and sleep quality during COVID-19 outbreak in china: a web-based cross-sectional survey [Psychiatry research, 288 (2020) 112954]. *Psychiatry Res* (2021) 299:113803. doi: 10.1016/j.psychres.2021.113803

4. DeMartini J, Patel G, Fancher TL. Generalized anxiety disorder. *Ann Intern Med* (2019) 170(7):ITC49–64doi: 10.7326/AITC201904020

5. MahmoudianDehkordi S, Bhattacharyya S, Brydges CR, Jia W, Fiehn O, Rush AJ, et al. Gut microbiome-linked metabolites in the pathobiology of major depression with or without anxiety-a role for bile acids. *Front Neurosci* (2022) 16:937906. doi: 10.3389/fnins.2022.937906

6. Mertens KL, Kalsbeek A, Soeters MR, Eggink HM. Bile acid signaling pathways from the enterohepatic circulation to the central nervous system. *Front Neurosci* (2017) 11:617. doi: 10.3389/fnins.2017.00617

7. Feng L, Zhou N, Li Z, Fu D, Guo Y, Gao X, et al. Co-occurrence of gut microbiota dysbiosis and bile acid metabolism alteration is associated with psychological disorders in crohn's disease. *FASEB J* (2022) 36:e22100. doi: 10.1096/fj.202101088RRR

8. BouSaba J, Sannaa W, McKinzie S, Vijayvargiya P, Chedid V, Wang XJ, et al. Impact of bile acid diarrhea in patients with diarrhea-predominant irritable bowel syndrome on symptoms and quality of life. *Clin Gastroenterol Hepatol* (2022) 20:2083– 90. doi: 10.1016/j.cgh.2021.11.035

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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9. Ren ZL, Li CX, Ma CY, Chen D, Chen JH, Xu WX, et al. Linking nonalcoholic fatty liver disease and brain disease: Focusing on bile acid signaling. *Int J Mol Sci* (2022) 23:13045. doi: 10.3390/ijms232113045

10. Shin A, Camilleri M, Vijayvargiya P, Busciglio I, Burton D, Ryks M, et al. Bowel functions, fecal unconjugated primary and secondary bile acids, and colonic transit in patients with irritable bowel syndrome. *Clin Gastroenterol Hepatol* (2013) 11:1270–75. doi: 10.1016/j.cgh.2013.04.020

11. Russell DW. The enzymes, regulation, and genetics of bile acid synthesis. *Annu Rev Biochem* (2003) 72:137–74. doi: 10.1146/annurev.biochem.72.121801.161712

12. Chiang J, Ferrell JM. Bile acids as metabolic regulators and nutrient sensors. Annu Rev Nutr (2019) 39:175–200. doi: 10.1146/annurev-nutr-082018-124344

13. Takahashi S, Fukami T, Masuo Y, Brocker CN, Xie C, Krausz KW, et al. Cyp2c70 is responsible for the species difference in bile acid metabolism between mice and humans. *J Lipid Res* (2016) 57:2130–37. doi: 10.1194/jlr.M071183

14. Chand D, Avinash VS, Yadav Y, Pundle AV, Suresh CG, Ramasamy S. Molecular features of bile salt hydrolases and relevance in human health. *Biochim Biophys Acta Gen Subj* (2017) 1861:2981–91. doi: 10.1016/j.bbagen.2016.09.024

15. Angelin B, Bjorkhem I, Einarsson K, Ewerth S. Hepatic uptake of bile acids in man. fasting and postprandial concentrations of individual bile acids in portal venous and systemic blood serum. *J Clin Invest* (1982) 70:724–31. doi: 10.1172/jci110668

16. Greenwood J, Adu J, Davey AJ, Abbott NJ, Bradbury MW. The effect of bile salts on the permeability and ultrastructure of the perfused, energy-depleted, rat blood-brain barrier. *J Cereb Blood Flow Metab* (1991) 11:644–54. doi: 10.1038/jcbfm.1991.116

17. Nizamutdinov D, DeMorrow S, McMillin M, Kain J, Mukherjee S, Zeitouni S, et al. Hepatic alterations are accompanied by changes to bile acid transporter-

expressing neurons in the hypothalamus after traumatic brain injury. *Sci Rep* (2017) 7:40112. doi: 10.1038/srep40112

18. Higashi T, Watanabe S, Tomaru K, Yamazaki W, Yoshizawa K, Ogawa S, et al. Unconjugated bile acids in rat brain: Analytical method based on LC/ESI-MS/MS with chemical derivatization and estimation of their origin by comparison to serum levels. *Steroids* (2017) 125:107–13. doi: 10.1016/j.steroids.2017.07.001

19. Pan X, Elliott CT, McGuinness B, Passmore P, Kehoe PG, Holscher C, et al. Metabolomic profiling of bile acids in clinical and experimental samples of alzheimer's disease. *Metabolites* (2017) 7:28. doi: 10.3390/metabo7020028

20. Ticho AL, Malhotra P, Dudeja PK, Gill RK, Alrefai WA. Bile acid receptors and gastrointestinal functions. *Liver Res* (2019) 3:31–9. doi: 10.1016/j.livres.2019.01.001

21. Jia B, Zou Y, Han X, Bae JW, Jeon CO. Gut microbiome-mediated mechanisms for reducing cholesterol levels: implications for ameliorating cardiovascular disease. *Trends Microbiol* (2023) 31:76–91. doi: 10.1016/j.tim.2022.08.003

22. Chiang J, Ferrell JM. Discovery of farnesoid x receptor and its role in bile acid metabolism. *Mol Cell Endocrinol* (2022) 548:111618. doi: 10.1016/j.mce.2022.111618

23. Huang C, Wang J, Hu W, Wang C, Lu X, Tong L, et al. Identification of functional farnesoid x receptors in brain neurons. *FEBS Lett* (2016) 590:3233–42. doi: 10.1002/1873-3468.12373

24. McMillin M, Frampton G, Quinn M, Ashfaq S, de Los SMR, Grant S, et al. Bile acid signaling is involved in the neurological decline in a murine model of acute liver failure. *Am J Pathol* (2016) 186:312–23. doi: 10.1016/j.ajpath.2015.10.005

25. Lundasen T, Galman C, Angelin B, Rudling M. Circulating intestinal fibroblast growth factor 19 has a pronounced diurnal variation and modulates hepatic bile acid synthesis in man. *J Intern Med* (2006) 260:530–36. doi: 10.1111/j.1365-2796.2006.01731.x

26. Miyake A, Itoh N. Rat fibroblast growth factor receptor-4 mRNA in the brain is preferentially expressed in cholinergic neurons in the medial habenular nucleus. *Neurosci Lett* (1996) 203:101–04. doi: 10.1016/0304-3940(95)12272-9

27. Goodwin B, Jones SA, Price RR, Watson MA, McKee DD, Moore LB, et al. A regulatory cascade of the nuclear receptors FXR, SHP-1, and LRH-1 represses bile acid biosynthesis. *Mol Cell* (2000) 6:517–26. doi: 10.1016/s1097-2765(00)00051-4

28. Clifford BL, Sedgeman LR, Williams KJ, Morand P, Cheng A, Jarrett KE, et al. FXR activation protects against NAFLD *via* bile-acid-dependent reductions in lipid absorption. *Cell Metab* (2021) 33:1671–84. doi: 10.1016/j.cmet.2021.06.012

29. Ma C, Yuan D, Renaud SJ, Zhou T, Yang F, Liou Y, et al. Chaihu-shugan-san alleviates depression-like behavior in mice exposed to chronic unpredictable stress by altering the gut microbiota and levels of the bile acids hyocholic acid and 7-ketoDCA. *Front Pharmacol* (2022) 13:1040591. doi: 10.3389/fphar.2022.1040591

30. Bao H, Li H, Jia Y, Xiao Y, Luo S, Zhang D, et al. Ganoderic acid a exerted antidepressant-like action through FXR modulated NLRP3 inflammasome and synaptic activity. *Biochem Pharmacol* (2021) 188:114561. doi: 10.1016/j.bcp.2021.114561

31. Li H, Zhu X, Xu J, Li L, Kan W, Bao H, et al. The FXR mediated anti-depression effect of CDCA underpinned its therapeutic potentiation for MDD. *Int Immunopharmacol* (2023) 115:109626. doi: 10.1016/j.intimp.2022.109626

32. Bazzari FH, Abdallah DM, El-Abhar HS. Chenodeoxycholic acid ameliorates AlCl(3)-induced alzheimer's disease neurotoxicity and cognitive deterioration *via* enhanced insulin signaling in rats. *Molecules* (2019) 24:1992. doi: 10.3390/molecules24101992

33. Keitel V, Haussinger D. Perspective: TGR5 (Gpbar-1) in liver physiology and disease. *Clin Res Hepatol Gastroenterol* (2012) 36:412–19. doi: 10.1016/j.clinre.2012.03.008

34. Zuo G, Zhang T, Huang L, Araujo C, Peng J, Travis Z, et al. Activation of TGR5 with INT-777 attenuates oxidative stress and neuronal apoptosis *via* cAMP/ PKCepsilon/ALDH2 pathway after subarachnoid hemorrhage in rats. *Free Radic Biol Med* (2019) 143:441–53. doi: 10.1016/j.freeradbiomed.2019.09.002

35. Keitel V, Gorg B, Bidmon HJ, Zemtsova I, Spomer L, Zilles K, et al. The bile acid receptor TGR5 (Gpbar-1) acts as a neurosteroid receptor in brain. *Glia* (2010) 58:1794–805. doi: 10.1002/glia.21049

36. Lou G, Ma X, Fu X, Meng Z, Zhang W, Wang YD, et al. GPBAR1/TGR5 mediates bile acid-induced cytokine expression in murine kupffer cells. *PloS One* (2014) 9:e93567. doi: 10.1371/journal.pone.0093567

37. Guizoni DM, Vettorazzi JF, Carneiro EM, Davel AP. Modulation of endothelium-derived nitric oxide production and activity by taurine and taurine-conjugated bile acids. *Nitric Oxide* (2020) 94:48–53. doi: 10.1016/j.niox.2019.10.008

38. Zheng X, Chen T, Jiang R, Zhao A, Wu Q, Kuang J, et al. Hyocholic acid species improve glucose homeostasis through a distinct TGR5 and FXR signaling mechanism. *Cell Metab* (2021) 33:791–803. doi: 10.1016/j.cmet.2020.11.017

39. Zheng X, Chen T, Zhao A, Ning Z, Kuang J, Wang S, et al. Hyocholic acid species as novel biomarkers for metabolic disorders. *Nat Commun* (2021) 12:1487. doi: 10.1038/s41467-021-21744-w

40. Wu H, Yu N, Wang X, Yang Y, Liang H. Tauroursodeoxycholic acid attenuates neuronal apoptosis *via* the TGR5/ SIRT3 pathway after subarachnoid hemorrhage in rats. *Biol Res* (2020) 53:56. doi: 10.1186/s40659-020-00323-1

41. Yanguas-Casas N, Barreda-Manso MA, Nieto-Sampedro M, Romero-Ramirez L. TUDCA: An agonist of the bile acid receptor GPBAR1/TGR5 with antiinflammatory effects in microglial cells. *J Cell Physiol* (2017) 232:2231-45. doi: 10.1002/jcp.25742 42. Xu N, Bai Y, Han X, Yuan J, Wang L, He Y, et al. Taurochenodeoxycholic acid reduces astrocytic neuroinflammation and alleviates experimental autoimmune encephalomyelitis in mice. *Immunobiology* (2023) 228:152388. doi: 10.1016/j.imbio.2023.152388

43. Eggink HM, Tambyrajah LL, van den Berg R, Mol IM, van den Heuvel JK, Koehorst M, et al. Chronic infusion of taurolithocholate into the brain increases fat oxidation in mice. *J Endocrinol* (2018) 236:85–97. doi: 10.1530/JOE-17-0503

44. Castellanos-Jankiewicz A, Guzman-Quevedo O, Fenelon VS, Zizzari P, Quarta C, Bellocchio L, et al. Hypothalamic bile acid-TGR5 signaling protects from obesity. *Cell Metab* (2021) 33:1483–92. doi: 10.1016/j.cmet.2021.04.009

45. Thibaut MM, Bindels LB. Crosstalk between bile acid-activated receptors and microbiome in entero-hepatic inflammation. *Trends Mol Med* (2022) 28:223–36. doi: 10.1016/j.molmed.2021.12.006

46. Collins SL, Stine JG, Bisanz JE, Okafor CD, Patterson AD. Bile acids and the gut microbiota: metabolic interactions and impacts on disease. *Nat Rev Microbiol* (2023) 21:236–47. doi: 10.1038/s41579-022-00805-x

47. Wang Y, Aoki H, Yang J, Peng K, Liu R, Li X, et al. The role of sphingosine 1phosphate receptor 2 in bile-acid-induced cholangiocyte proliferation and cholestasisinduced liver injury in mice. *Hepatology* (2017) 65:2005–18. doi: 10.1002/hep.29076

48. McMillin M, Frampton G, Grant S, Khan S, Diocares J, Petrescu A, et al. Bile acid-mediated sphingosine-1-Phosphate receptor 2 signaling promotes neuroinflammation during hepatic encephalopathy in mice. *Front Cell Neurosci* (2017) 11:191. doi: 10.3389/fncel.2017.00191

49. Boehlig A, Gerhardt F, Petroff D, van Boemmel F, Berg T, Blank V, et al. Prevalence of pruritus and association with anxiety and depression in patients with nonalcoholic fatty liver disease. *Biomedicines* (2022) 10:451. doi: 10.3390/biomedicines10020451

50. Aziz I, Mumtaz S, Bholah H, Chowdhury FU, Sanders DS, Ford AC. High prevalence of idiopathic bile acid diarrhea among patients with diarrhea-predominant irritable bowel syndrome based on rome III criteria. *Clin Gastroenterol Hepatol* (2015) 13:1650–55. doi: 10.1016/j.cgh.2015.03.002

51. de Aguiar VT, Tarling EJ, Edwards PA. Pleiotropic roles of bile acids in metabolism. Cell Metab (2013) 17:657-69. doi: 10.1016/j.cmet.2013.03.013

52. Miyata M, Sakaida Y, Matsuzawa H, Yoshinari K, Yamazoe Y. Fibroblast growth factor 19 treatment ameliorates disruption of hepatic lipid metabolism in farnesoid x receptor (Fxr)-null mice. *Biol Pharm Bull* (2011) 34:1885–89. doi: 10.1248/bpb.34.1885

53. Hyland NP, Quigley EM, Brint E. Microbiota-host interactions in irritable bowel syndrome: epithelial barrier, immune regulation and brain-gut interactions. *World J Gastroenterol* (2014) 20:8859–66. doi: 10.3748/wjg.v20.i27.8859

54. Kamp F, Hamilton JA, Kamp F, Westerhoff HV, Hamilton JA. Movement of fatty acids, fatty acid analogues, and bile acids across phospholipid bilayers. *Biochemistry* (1993) 32:11074–86. doi: 10.1021/bi00092a017

55. Quinn M, McMillin M, Galindo C, Frampton G, Pae HY, DeMorrow S. Bile acids permeabilize the blood brain barrier after bile duct ligation in rats *via* Rac1-dependent mechanisms. *Dig Liver Dis* (2014) 46:527–34. doi: 10.1016/j.dld.2014.01.159

56. Klaassen CD, Aleksunes LM. Xenobiotic, bile acid, and cholesterol transporters: function and regulation. *Pharmacol Rev* (2010) 62:1–96. doi: 10.1124/pr.109.002014

57. Choudhuri S, Cherrington NJ, Li N, Klaassen CD. Constitutive expression of various xenobiotic and endobiotic transporter mRNAs in the choroid plexus of rats. *Drug Metab Dispos* (2003) 31:1337–45. doi: 10.1124/dmd.31.11.1337

58. McMillin M, DeMorrow S. Effects of bile acids on neurological function and disease. FASEB J (2016) 30:3658–68. doi: 10.1096/fj.201600275R

59. Abdo QE, Abdullah Z, Zakaria R, Long I. Minocycline attenuates lipopolysaccharide-induced locomotor deficit and anxiety-like behavior and related expression of the BDNF/CREB protein in the rat medial prefrontal cortex (mPFC). *Int J Mol Sci* (2022) 23:13474. doi: 10.3390/ijms232113474

60. Hosseinzadeh SO, Rezayof A, Ghasemzadeh Z, Alijanpour S, Rahimian S. Ameliorating effect offluoxetine on tamoxifen-induced memory loss: The role of corticolimbic NMDA receptors and CREB/BDNF/cFos signaling pathways in rats. *Brain Res* (2022) 1794:148058. doi: 10.1016/j.brainres.2022.148058

61. Li J, Wang H, Liu D, Li X, He L, Pan J, et al. CB2R activation ameliorates late adolescent chronic alcohol exposure-induced anxiety-like behaviors during withdrawal by preventing morphological changes and suppressing NLRP3 inflammasome activation in prefrontal cortex microglia in mice. *Brain Behav Immun* (2023) 110:60–79. doi: 10.1016/j.bbi.2023.02.001

62. Lei Y, Chen CJ, Yan XX, Li Z, Deng XH. Early-life lipopolysaccharide exposure potentiates forebrain expression of NLRP3 inflammasome proteins and anxiety-like behavior in adolescent rats. *Brain Res* (2017) 1671:43–54. doi: 10.1016/j.brainres.2017.06.014

63. Sadeghi MA, Nassireslami E, Yousefi ZM, Hosseini Y, Abbasian K, Chamanara M. Phosphodiesterase inhibitors in psychiatric disorders. *Psychopharmacol (Berl)* (2023) 240:1201–19. doi: 10.1007/s00213-023-06361-3

64. Ugolini M, Keil MF, Paradiso E, Wu J, Stratakis CA. Anxiety-like behavior and other consequences of early life stress in mice with increased protein kinase a activity. *Behav Brain Res* (2018) 348:22–30. doi: 10.1016/j.bbr.2018.04.001

65. Keil MF, Briassoulis G, Stratakis CA, Wu TJ. Protein kinase a and anxiety-related behaviors: A mini-review. *Front Endocrinol (Lausanne)* (2016) 7:83. doi: 10.3389/ fendo.2016.00083

66. Jin P, Deng S, Tian M, Lenahan C, Wei P, Wang Y, et al. INT-777 prevents cognitive impairment by activating takeda g protein-coupled receptor 5 (TGR5) and attenuating neuroinflammation *via* cAMP/ PKA/ CREB signaling axis in a rat model of sepsis. *Exp Neurol* (2021) 335:113504. doi: 10.1016/j.expneurol.2020.113504

67. Wu X, Liu C, Chen L, Du YF, Hu M, Reed MN, et al. Protective effects of tauroursodeoxycholic acid on lipopolysaccharide-induced cognitive impairment and neurotoxicity in mice. *Int Immunopharmacol* (2019) 72:166–75. doi: 10.1016/j.intimp.2019.03.065

68. Lawrence T. The nuclear factor NF-kappaB pathway in inflammation. Cold Spring Harb Perspect Biol (2009) 1:a1651. doi: 10.1101/cshperspect.a001651

69. Jang HM, Han SK, Kim JK, Oh SJ, Jang HB, Kim DH. Lactobacillus sakei alleviates high-Fat-Diet-Induced obesity and anxiety in mice by inducing AMPK activation and SIRT1 expression and inhibiting gut microbiota-mediated NF-kappaB activation. *Mol Nutr Food Res* (2019) 63:e1800978. doi: 10.1002/mnfr.201800978

70. Jang HM, Lee KE, Lee HJ, Kim DH. Immobilization stress-induced escherichia coli causes anxiety by inducing NF-kappaB activation through gut microbiota disturbance. *Sci Rep* (2018) 8:13897. doi: 10.1038/s41598-018-31764-0

71. Paltian JJ, Dos RA, de Oliveira RL, Da FC, Domingues WB, Dellagostin EN, et al. The anxiolytic effect of a promising quinoline containing selenium with the contribution of the serotonergic and GABAergic pathways: Modulation of parameters associated with anxiety in mice. *Behav Brain Res* (2020) 393:112797. doi: 10.1016/j.bbr.2020.112797

72. Rai SN, Dilnashin H, Birla H, Singh SS, Zahra W, Rathore AS, et al. The role of PI3K/Akt and ERK in neurodegenerative disorders. *Neurotox Res* (2019) 35:775–95. doi: 10.1007/s12640-019-0003-y

73. Jung EM, Ka M, Kim WY. Loss of GSK-3 causes abnormal astrogenesis and behavior in mice. *Mol Neurobiol* (2016) 53:3954–66. doi: 10.1007/s12035-015-9326-8

74. Hauger RL, Olivares-Reyes JA, Dautzenberg FM, Lohr JB, Braun S, Oakley RH. Molecular and cell signaling targets for PTSD pathophysiology and pharmacotherapy. *Neuropharmacology* (2012) 62:705–14. doi: 10.1016/j.neuropharm.2011.11.007

75. Carbone K, Gervasi F. An updated review of the genus humulus: A valuable source of bioactive compounds for health and disease prevention. *Plants (Basel)* (2022) 11:3434. doi: 10.3390/plants11243434

76. Mudaliar S, Henry RR, Sanyal AJ, Morrow L, Marschall HU, Kipnes M, et al. Efficacy and safety of the farnesoid x receptor agonist obeticholic acid in patients with type 2 diabetes and nonalcoholic fatty liver disease. *Gastroenterology* (2013) 145:574–82. doi: 10.1053/j.gastro.2013.05.042

77. Lu C, Wang Y, Furukawa K, Fu W, Ouyang X, Mattson MP. Evidence that caspase-1 is a negative regulator of AMPA receptor-mediated long-term potentiation at hippocampal synapses. *J Neurochem* (2006) 97:1104–10. doi: 10.1111/j.1471-4159.2006.03800.x

78. Diering GH, Huganir RL. The AMPA receptor code of synaptic plasticity. *Neuron* (2018) 100:314–29. doi: 10.1016/j.neuron.2018.10.018

79. Ma X, Li L, Li Z, Huang Z, Yang Y, Liu P, et al. eEF2 in the prefrontal cortex promotes excitatory synaptic transmission and social novelty behavior. *EMBO Rep* (2022) 23:e54543. doi: 10.15252/embr.202154543

80. Chen ZJ, Su CW, Xiong S, Li T, Liang HY, Lin YH, et al. Enhanced AMPARdependent synaptic transmission by s-nitrosylation in the vmPFC contributes to chronic inflammatory pain-induced persistent anxiety in mice. *Acta Pharmacol Sin* (2023) 44:954–68. doi: 10.1038/s41401-022-01024-z

81. Zhang J, Li H, Zhou H, Fang L, Xu J, Yan H, et al. Lowered fasting chenodeoxycholic acid correlated with the decrease of fibroblast growth factor 19 in chinese subjects with impaired fasting glucose. *Sci Rep* (2017) 7:6042. doi: 10.1038/ s41598-017-06252-6

82. Zweers SJ, de Vries EM, Lenicek M, Tolenaars D, de Waart DR, Koelfat KV, et al. Prolonged fibroblast growth factor 19 response in patients with primary sclerosing cholangitis after an oral chenodeoxycholic acid challenge. *Hepatol Int* (2017) 11:132– 40. doi: 10.1007/s12072-016-9769-7

83. Inagaki T, Choi M, Moschetta A, Peng L, Cummins CL, McDonald JG, et al. Fibroblast growth factor 15 functions as an enterohepatic signal to regulate bile acid homeostasis. *Cell Metab* (2005) 2:217–25. doi: 10.1016/j.cmet.2005.09.001

84. Ding L, Yang L, Wang Z, Huang W. Bile acid nuclear receptor FXR and digestive system diseases. *Acta Pharm Sin B* (2015) 5:135–44. doi: 10.1016/j.apsb.2015.01.004

85. Galvez-Contreras AY, Campos-Ordonez T, Lopez-Virgen V, Gomez-Plascencia J, Ramos-Zuniga R, Gonzalez-Perez O. Growth factors as clinical biomarkers of prognosis and diagnosis in psychiatric disorders. *Cytokine Growth Factor Rev* (2016) 32:85–96. doi: 10.1016/j.cytogfr.2016.08.004

86. Deng Z, Deng S, Zhang MR, Tang MM. Fibroblast growth factors in depression. Front Pharmacol (2019) 10:60. doi: 10.3389/fphar.2019.00060

87. Ryan KK, Kohli R, Gutierrez-Aguilar R, Gaitonde SG, Woods SC, Seeley RJ. Fibroblast growth factor-19 action in the brain reduces food intake and body weight and improves glucose tolerance in male rats. *Endocrinology* (2013) 154:9–15. doi: 10.1210/en.2012-1891

88. Kuro-O M. The klotho proteins in health and disease. Nat Rev Nephrol (2019) 15:27-44. doi: 10.1038/s41581-018-0078-3

89. Jianwen ZW. Research on the antidepressant mechanism of Kaixin Jieyu Pills in regulating bile acid metabolism in the "gut-liver-brain axis". *China Academy of Chinese Medical Sciences* (2023) 2. doi: 10.27658/d.cnki.gzzyy.2022.000159

90. Wu WL, Cheng CF, Sun WH, Wong CW, Chen CC. Targeting ASIC3 for pain, anxiety, and insulin resistance. *Pharmacol Ther* (2012) 134:127–38. doi: 10.1016/j.pharmthera.2011.12.009

91. Soto M, Cai W, Konishi M, Kahn CR. Insulin signaling in the hippocampus and amygdala regulates metabolism and neurobehavior. *Proc Natl Acad Sci USA* (2019) 116:6379–84. doi: 10.1073/pnas.1817391116

92. Zeng W, Yang F, Shen WL, Zhan C, Zheng P, Hu J. Interactions between central nervous system and peripheral metabolic organs. *Sci China Life Sci* (2022) 65:1929–58. doi: 10.1007/s11427-021-2103-5

93. Frankiensztajn LM, Elliott E, Koren O. The microbiota and the hypothalamuspituitary-adrenocortical (HPA) axis, implications for anxiety and stress disorders. *Curr Opin Neurobiol* (2020) 62:76–82. doi: 10.1016/j.conb.2019.12.003

94. Fiksdal A, Hanlin L, Kuras Y, Gianferante D, Chen X, Thoma MV, et al. Associations between symptoms of depression and anxiety and cortisol responses to and recovery from acute stress. *Psychoneuroendocrinology* (2019) 102:44–52. doi: 10.1016/j.psyneuen.2018.11.035

95. Salpekar JA, Basu T, Thangaraj S, Maguire J. The intersections of stress, anxiety and epilepsy. *Int Rev Neurobiol* (2020) 152:195–219. doi: 10.1016/bs.irn.2020.02.001

96. Tang L, Li S, Yu J, Zhang Y, Yang L, Tong D, et al. Nonylphenol induces anxietylike behavior in rats by regulating BDNF/TrkB/CREB signal network. *Food Chem Toxicol* (2022) 166:113197. doi: 10.1016/j.fct.2022.113197

97. Jiang YL, Wang XS, Li XB, Liu A, Fan QY, Yang L, et al. Tanshinone IIA improves contextual fearand anxiety-like behaviors in mice *via* the CREB/BDNF/TrkB signaling pathway. *Phytother Res* (2022) 36:3932–48. doi: 10.1002/ptr.7540

98. Hui S, Huang L, Wang X, Zhu X, Zhou M, Chen M, et al. Capsaicin improves glucose homeostasis by enhancing glucagon-like peptide-1 secretion through the regulation of bile acid metabolism *via* the remodeling of the gut microbiota in male mice. *FASEB J* (2020) 34:8558–73. doi: 10.1096/fj.201902618RR

99. Tian F, Xu W, Chen L, Chen T, Feng X, Chen J, et al. Ginsenoside compound k increases glucagon-like peptide-1 release and l-cell abundance in db/db mice through TGR5/YAP signaling. *Int Immunopharmacol* (2022) 113:109405. doi: 10.1016/j.intimp.2022.109405

100. Kuhre RE, Wewer AN, Larsen O, Jepsen SL, Balk-Moller E, Andersen DB, et al. Bile acids are important direct and indirect regulators of the secretion of appetite- and metabolism-regulating hormones from the gut and pancreas. *Mol Metab* (2018) 11:84– 95. doi: 10.1016/j.molmet.2018.03.007

101. Chaudhari SN, Luo JN, Harris DA, Aliakbarian H, Yao L, Paik D, et al. A microbial metabolite remodels the gut-liver axis following bariatric surgery. *Cell Host Microbe* (2021) 29:408–24. doi: 10.1016/j.chom.2020.12.004

102. Bae CS, Song J. The role of glucagon-like peptide 1 (GLP1) in type 3 diabetes: GLP-1 controls insulin resistance, neuroinflammation and neurogenesis in the brain. *Int J Mol Sci* (2017) 18:2493. doi: 10.3390/ijms18112493

103. Saglam C, Turan I, Ozacmak HS. The effect of glucagon like peptide-1 receptor agonist on behavioral despair and anxiety-like behavior in ovariectomized rats: Modulation of BDNF/CREB, Nrf2 and lipocalin 2. *Behav Brain Res* (2022) 435:114053. doi: 10.1016/j.bbr.2022.114053

104. Lopez-Ferreras L, Eerola K, Shevchouk OT, Richard JE, Nilsson FH, Jansson LE, et al. The supramammillary nucleus controls anxiety-like behavior; key role of GLP-1R. *Psychoneuroendocrinology* (2020) 119:104720. doi: 10.1016/j.psyneuen.2020.104720

105. Graham DL, Madkour HS, Noble BL, Schatschneider C, Stanwood GD. Longterm functional alterations following prenatal GLP-1R activation. *Neurotoxicol Teratol* (2021) 87:106984. doi: 10.1016/j.ntt.2021.106984

106. Sun B, Jia X, Yang F, Ren G, Wu X. CREB-mediated generation and neuronal growth regulates the behavioral improvement of geniposide in diabetes-associated depression mouse model. *Neurosci Res* (2021) 165:38-44. doi: 10.1016/j.neures.2020.05.003

107. Breit S, Kupferberg A, Rogler G, Hasler G. Vagus nerve as modulator of the brain-gut axis in psychiatric and inflammatory disorders. *Front Psychiatry* (2018) 9:44. doi: 10.3389/fpsyt.2018.00044

108. Magnon V, Dutheil F, Vallet GT. Benefits from one session of deep and slow breathing on vagal tone and anxiety in young and older adults. *Sci Rep* (2021) 11:19267. doi: 10.1038/s41598-021-98736-9

109. Yesiltepe M, Cimen B, Sara Y. Effects of chronic vagal nerve stimulation in the treatment of beta-amyloid-induced neuropsychiatric symptoms. *Eur J Pharmacol* (2022) 931:175179. doi: 10.1016/j.ejphar.2022.175179

110. Rogers GB, Keating DJ, Young RL, Wong ML, Licinio J, Wesselingh S. From gut dysbiosis to altered brain function and mental illness: mechanisms and pathways. *Mol Psychiatry* (2016) 21:738–48. doi: 10.1038/mp.2016.50