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4	Occurrence and courses of natural and anthronogenic
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6	lipid tracers in surface soils from arid urban areas of
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43 ABSTRACT

Soil particles contain a variety of natural and anthropogenic organic components, 44 and in urban areas can be considered as local collectors of pollutants. Surface soil 45 samples were taken from ten urban areas in Riyadh during early winter of 2007. They 46 were extracted with dichloromethane-methanol mixture and the extracts were analyzed 47 by gas chromatography-mass spectrometry. The major compounds were unresolved 48 49 complex mixture (UCM), plasticizers, n-alkanes, carbohydrates, n-alkanoic acids, hopanes, n-alkanols, and sterols. Vegetation detritus was the major natural source of 50 organic compounds (24.0+15.7%) in samples from areas with less human activities and 51 included n-alkanes, n-alkanoic acids, n-alkanols, sterols and carbohydrates. Vehicular 52 emission products and discarded plastics were the major anthropogenic sources in the soil 53 particles $(53.3\pm21.3\%)$ and $22.7\pm10.7\%$, respectively). The anthropogenic tracers were 54 UCM, plasticizers, n-alkanes, hopanes and traces of steranes. Vegetation and human 55 activities control the occurrence and distribution of natural and anthropogenic extractable 56 57 organic matter in this arid urban area.

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59 Capsule:

60 This work shows that human activities are critical factors that influence the 61 characteristics and distribution of EOM in soils of arid urban regions.

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63 Key words: soils, biomarkers, lipids, petroleum residues, plasticizers.

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1. Introduction

Biogenic and anthropogenic organic matter (OM) is a major carbonaceous 66 67 fraction in different ecosystems (e.g. Hansell and Carlson, 2014; Lischke et al., 2014; Schmidt et al., 2011). The occurrence and concentrations of OM in urban and 68 rural/remote region soils vary spatially and seasonally (Fraser et al., 1998; Kononova, 69 70 2013; Oros et al., 2002; Trendel et al., 2010). One of the primary sources of OM in soil is material from plant litter. Other sources include anthropogenic organic chemicals such as 71 pesticides and fossil fuel residues from vehicles (Al-Mutlag et al., 2005, 2007; Rushdi et 72 73 al., 2005, 2013; Yang et al., 2013). The composition, properties and distribution of this OM controls the formation of soil organic matter (SOM) and humification processes 74 (Kononova, 2013; Scholes et al., 1997; Schmidt et al., 2011). Microbial activity and 75 76 biomass are also important factors in SOM formation and alteration (Schmidt et al., 2011). Therefore, SOM varies in composition and properties. The major studies on soil 77 OM have focused on bulk properties (Kögel-Knabner, 2000; Martin and Haider, 1986), 78 79 whereas extractable organic matter (lipid) studies are relatively few (Otto and Simpson, 2005; Rogge et al., 2012; Rushdi et al., 2005, 2006). Less attention has been paid to the 80 lower molecular weight OM (Kögel-Knabner, 2000). 81

Lipid compounds usually comprise 1-5% of the total SOM. They are derived from plants, fungi, bacteria and mesofauna, and generally consist of <u>n</u>-alkanes, <u>n</u>-alkanols, wax esters, fatty acids, steroids, terpenoids and acyl glycerols (Oros et al., 2002). These products are a result of decomposition and exudation of OM from various sources (Miller and Donahue, 1995). Lipids can be found either free in the soil matrix, chemically bound in humic and fulvic material, or absorbed to soil particles (Kononova, 2013; Schmidt et
al., 2011).

Thus, characterization and identification of organic compounds and OM contents of surface soils are important for understanding the OM composition and compound sources that may be introduced to the different environmental compartments, such as for example: (1) absorption into the food chain, (2) advection into the atmosphere via dust resuspension, machinery and/or human activities, and (3) wash-in to water reservoirs. It will also aid to better understand the various effects on human health and environmental assessment processes.

96 The extractable organic matter (EOM) of surface soil has not been fully characterized for the Arabian Peninsula region. The preliminary results have shown that 97 soil and sand dusts from Kuwait and Saudi Arabia contained a mixture of natural and 98 99 anthropogenic OM (Al-Mutlag et al., 2002, 2007; Rushdi et al., 2005, 2006). Based on 100 aliphatic hydrocarbon contents of soil from Riyadh, the natural and anthropogenic 101 sources ranged from 20 to 57% and from 30 to 55% of the total lipid tracers, respectively 102 (Rushdi et al., 2005). Soil and samples from the vicinity of Kuwait City showed that 103 the natural sources ranged from 15 to 78% and the anthropogenic sources ranged from 8 to 88% of the total lipid compounds (Rushdi et al., 2006). 104

The research hypotheses of this work are: (1) the EOM sources in soils of an urban arid region is mainly from traffic emissions; and (2) the sources of natural EOM are relatively low and are primarily from soil microbiota with lesser contributions from vegetation. Therefore, the main objectives of this work are to: (1) determine the chemical composition of the solvent EOM in soils from arid urban areas in Saudi Arabia, (2) identify the main sources of the EOM, and (3) examine the relative changes in the source

abundances of the EOM. Consequently, surface soil samples were collected from various
locations such as public parks, streets, and agricultural fields in the city of Riyadh to
investigate the occurrence of natural biogenic and anthropogenic lipid compounds.

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2. Experimental Methods

116 2.1. Sampling and Extraction Procedure

117 Surface oil samples were collected in November 2007 from different metropolitan 118 areas of the city of Riyadh in Saudi Arabia to characterize their solvent extractable lipid contents by gas chromatography-mass spectrometry (GC-MS) analysis. The samples 119 were taken by scraping the uppermost layer (~1cm) of the soil in a 30 cm² area of 120 121 exposed surface. The locations of the samples were chosen to represent urban areas in and around Riyadh, ranging from highly populated, with different human activities such 122 as public markets (OL), governmental site (NZ), industrial area (MN), rural areas with 123 124 limited human impact (OG), and areas with agriculture (AQ, JZ, DR) (Fig. 1).

After air drying, each sample was sieved to obtain the fine particles ($<125\mu m$) 125 before extraction. The extraction was performed twice by adding a mixture of 126 dichloromethane/methanol (40 mL 3:1 v/v) to about 5 g of the particles of each sample, 127 ultra-sonicating for 20 min, and then filtering through pre-extracted glass microfiber 128 filters (Whatman[®], GF/A filters). Each total extract was concentrated under nitrogen 129 blow-down at room temperature to approximately 1.0-1.5 mL before GC-MS analysis. A 130 131 50 μ L aliquot of each total extract was also derivatized with silvlating reagent [N,Obis(trimethylsilyl)trifluoroacetamide, BSTFA, Pierce Chemical Co] by the standard 132 procedure before analysis by GC-MS. This derivatizing agent replaces the H in hydroxyl 133

- 134 groups with a trimethylsilyl [(CH₃)₃Si, i.e., TMS] group for better GC resolution of polar
- 135 compounds.



Figure 1. Map showing the sampling sites and locations of the soil samples.

137 2.2. Instrumental Analysis

The analyses of the total and silylated extracts were carried out by GC-MS, using an Agilent 6890 GC coupled to a 5973 Mass Selective Detector with a DB-5 (*Agilent*) fused silica capillary column (30 m x 0.25 mm i.d., 0.25 μ m film thickness) and helium as the carrier gas. The GC was temperature programmed from 65°C (2 min initial time) to 300°C at 6°C min⁻¹ (isothermal for 20 min final time). The MS was operated in the electron impact mode at 70 eV ion source energy. The data were acquired and processed with an Agilent ChemStation. Compounds were identified by GC retention index and comparison of mass spectra with those of authentic standards, literature and library data, or characterized mixtures. Unknown compounds were characterized by interpretation of the fragmentation pattern of their mass spectra. The concentrations of the compounds in ng g^{-1} sample were obtained from ratios of peak areas in the total ion current traces of the samples versus the peak areas of a mixture of quantitative standards.

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2.3. *Quality Control*

153 Throughout the study, emphasis has been placed on the technical accuracy of the 154 research. This applied to sample collection, analytical chemistry, biomarker tracer determination, and geochemical interpretation. All solvents used in the analytical workup 155 procedure were tested for possible contaminants. Procedural blanks were analyzed to 156 157 provide a qualitative and quantitative assessment of background contamination introduced by laboratory procedures. Blank extracts were performed through the entire 158 project with batches of three samples. The contamination background levels were 159 160 insignificant in all procedural blanks. The recoveries of external standards (tetracosane for n-alkanes; hexadecanoic acid for n-alkanoic acids, alkyl alkanoates and n-alkanols; 161 sitosterol for triterpenoids; glucose for monosaccharides; and sucrose for disaccharides) 162 ranged from 72 to 105%. 163

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166 **3. Results and Discussion**

167 The total extract compositions of the soil samples from the metropolitan Riyadh 168 area are given in Table SM1 (Supplementary Material) and vary in contents depending on

the locations and human activities. This variation is illustrated by the data for samples from the city center (Fig. 2a, sample OL) and outside the city (Fig. 2b, sample OG). The EOM consists mainly of mixtures of <u>n</u>-alkanes, <u>n</u>-alkanoic acids, sterols, <u>n</u>-alkanols, carbohydrates hopanes, plasticizers, and UCM (unresolved complex mixture, mainly of branched and cyclic hydrocarbons from petroleum products, Simoneit, 1985).



Figure 2. GC-MS total ion current traces of total extracts of the soil samples from outside and in the city of Riyadh showing the major organic components: a) OL (center); b) OR (outside the city) (Numbers refer to the carbon chain length and symbols are: DIBP= diisobutyl phthalate, DBP= dibutyl phthalate and DEHP = diethylhexyl phthalate, H = hopanes, and UCM = unresolved complex mixture).

The <u>n</u>-alkanes were detected in all samples and ranged from C_{13} to C_{35} with a maximum concentration (C_{max}) at nonadecane, eicosane or heneicosane for some samples and heptacosane, nonacosane or hentriacontane for others (Fig. 3a,b; Table SM1). The concentrations of the total <u>n</u>-alkanes ranged from 0.10 to 4.28 µg g⁻¹ with an average value of 1.64 ± 1.53 µg g⁻¹.

The <u>n</u>-alkanoic acids ranged from C_6 to C_{18} with a C_{max} at octanoic and nonanoic acids in all samples, except sample JZ where the C_{max} was at tetradecanoic acid (Table SM1). Their total concentrations ranged from 0.06 to 2.29 µg g⁻¹ with an average value of 0.70±0.69 µg g⁻¹. The <u>n</u>-alkanols were also present in all samples and ranged from C_{10} to C_{30} with a C_{max} at tetradecanol, hexadecanol, octacdecanol, octacosanol or triacontanol. The total concentrations of <u>n</u>-alkanols ranged from 0.06 to 0.43 µg g⁻¹ with an average

The total concentrations of <u>n</u>-alkanols ranged from 0.06 to 0.43 μ g g⁻¹ with an average value of 0.17±0.12 μ g g⁻¹.

Sterols were found in all samples and included mainly cholesterol, brassicasterol, campesterol, stigmasterol and sitosterol. The maximum concentration was either cholesterol or sitosterol. The total concentration of sterols ranged from 0.03 to 0.35 μ g g⁻¹ (mean = 0.12±0.11 μ g g⁻¹). The carbohydrates in the soil samples included glucose, sucrose and mycose as well as glycerol (sugar alcohol). The maximum concentration was glucose, sucrose or mycose. The total concentrations of carbohydrates ranged from 0.18 to 3.22 μ g g⁻¹ with an average concentration of 0.73±0.76 μ g g⁻¹.





Figure 3. Examples of typical GC-MS key ion plots for various compound series found in the soil samples: a) OR and b) SH, *m/z* 85 for <u>n</u>-alkanes; c) MN, *m/z* 191 for hopanes; d) MN, *m/z* 217 and 218 for steranes, and e) MN *m/z* 149 for phthalate plasticizers.

Hopanes, biomarkers for petroleum residues (Peters and Moldowan, 1993; Simoneit, 1985), were found to range from C_{27} to C_{35} , with a C_{max} at 29 and 30 (e.g. Fig. 3c), and concentrations from 0.02 to 3.35 µg g⁻¹, mean 0.47±1.08 µg g⁻¹. The C_{27} to C_{29} steranes were only detectable in the sample from Manakh (MN) at a concentration of 7.36 µg g⁻¹ (Fig. 3d). Plasticizers, including diethyl-, dibutyl-, dioctyl-phthalates and dioctyl adipate, were significant components of these samples (Figs. 2 and 3e), with concentrations ranging from 0.37 to 8.03 µg g⁻¹ and mean of 2.49±2.40 µg g⁻¹. The UCM concentrations were high in some samples and ranged from 0.0 to 83.28 μ g g⁻¹. The average concentration of the UCM was 9.59±26.03 μ g g⁻¹.

The results indicate that both natural biogenic and anthropogenic sources are the 206 main inputs of the EOM to the soil samples of the city of Riyadh. Therefore, the entire 207 data set was statistically analyzed by principal component analysis (PCA) techniques 208 using the SPSS (IBM-Statistical Package for the Social Sciences, version 21) software to 209 210 understand the similarity among the different sites. The output of the PCA analysis, explaining 85.1% of the variance, is shown in Figure 4a where two site groups were 211 212 recognized. The first group (C1 explains 47.1%) included the sites AQ, NZ, DR, OG, AZ, JZ and OL; and the second one (C2 explains 25.4%) included the AM, MN and SH 213 sites. The MN site was in an industrial area and the AM and SH sites were in an area 214 215 where large trucks usually gather and idle before entering the city. The rest of the sites were in different locations inside the city. This indicates that the locations and the type of 216 217 human activities have an impact on the characteristics and distributions of EOM in the region. 218



Figure 4. Plots showing the statistical outputs of principal component analysis (PCA) for: (a) entire compounds; (b) natural biogenic compounds and (c) anthropogenic compounds.

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221 3.1. Natural biogenic sources

The occurrence and distribution of vegetation wax lipids, such as <u>n</u>-alkanes (in part), <u>n</u>-alkanols, and sterols, validates the natural source input from higher plant biomass to the soils, as well as carbohydrates indicate input from plant detritus (e.g. roots) and microbial/fungal activity in the soils (Otto and Simpson, 2005; Simoneit, 2006; Trendel et al., 2010). Plant wax <u>n</u>-alkanes show a C_{max} in the range of 25 to 33, which depends on

the plant species, as well as the season and locality (e.g. Simoneit, 1977; Buch and 227 228 McInerney, 2013; Eglinton and Hamilton, 1967). A CPI (here the ratio of $\sum (C_{23}+C_{25}+C_{27}+C_{29}+C_{31}+C_{33}+C_{35})-\text{to}-\sum (C_{22}+C_{24}+C_{26}+C_{28}+C_{30}+C_{32}+C_{34})$ value of >1.0 229 230 is also an indicator of natural sources, whereas a CPI ~1.0 (Table SM1) supports a source from fossil fuel utilization (Bray and Evans, 1961; Mazurek and Simoneit, 1984). The 231 odd numbered n-alkanes were dominant in the NZ, DR, SH, AZ and JZ samples, with 232 CPI ranging from 1.4 and 2.5, whereas samples from AM, OG, MN and OL showed no 233 significant carbon number preference with CPI of 0.8 to 1.1 (Table SM1). This odd 234 carbon number preference (CPI >1) of the n-alkanes supported by the C_{max} mainly at 29 235 and/or 31 is distinctive of higher plant waxes (e.g. Table SM1, Fig. 3b, Buch and 236 McInerney, 2013; Carr et al., 2014). The contribution of wax n-alkanes were calculated 237 following the method developed by Simoneit et al. (1991) and found to range from 0.01 238 to 0.31 μ g g⁻¹ in these samples. 239

240 The presence of long chain n-alkanols and minor n-alkanoic acids ($>C_{20}$), with primarily only the even carbon number homologs in the series, is another indicator for 241 plant wax (e.g. Hitchcock and Nichols, 1971; Simoneit, 1989; Table SM1). The n-242 alkanoic acids (<C₂₀) with a strong even carbon number predominance and C_{max} at 16, 243 are from multiple sources including microbiota and vascular plant waxes, cutin and 244 suberin (Holloway, 1982; Simoneit, 1989). The absence of unsaturated n-alkanoic acids 245 indicates extensive oxidation and biodegradation of the biomass components in the soils. 246 The even carbon numbered n-alkanols ranging from C_{18} to C_{30} with a C_{max} at 28 and 30 247 are from vascular plant wax typical of desert environments (Simoneit, 1989). The 248 occurrence of high levels of short chain <u>n</u>-alkanols (<C₁₈) with a C_{max} at 10, 11 and 12 249

indicate biodegradation and oxidative processes of the plant wax as well as microbialactivities in soils.

252 Steroids can be derived from vascular higher plants, microbial activity in the soil, 253 and cooking in urban areas (Huang and Meinschein, 1976; Rogge et al., 1991). The 254 phytosterols, mainly campesterol, stigmasterol and sitosterol, in the samples from outside the city are derived from vegetation (Simoneit et al., 1983). The presence of cholesterol 255 256 at relatively high concentrations in most of the soil samples (e.g. AQ, AM, AZ, MN and OL; Table SM1) indicates microbial and/or cooking activities in the area (Rogge et al., 257 1991; Thompson al., 2013). The ratio of cholesterol-to-258 et 259 (campesterol+stigmasterol+sitosterol) was used to estimate the dominant sources of sterols in soils (Table SM1). The sites of AQ AM and MN were found to have higher 260 ratios indicating that microbial activities and possibly cooking as a result of evening 261 262 picnics in these areas were the major sources of sterols.

The carbohydrates are comprised mainly of primary saccharides produced by 263 photosynthesis and utilized by biota for energy, with minor metabolites. Complex 264 carbohydrate biopolymers (e.g. cellulose) in soils undergo enzymatic and hydrolytic 265 degradation to simple sugars (i.e., glucose) (Gleixner et al., 2002). Glycerol, the simplest 266 reduced sugar, is derived from the hydrolysis of glyceride lipids and is a significant 267 carbohydrate component of the soils (mean concentration 0.17 μ g g⁻¹, Table SM1). The 268 total saccharide concentrations are low in these soils (0.16 to 3.22 µg g⁻¹, mean 0.90 µg g⁻¹ 269 ¹, Table SM1), compared to an example from a temperate region (3.77 to 23.71 μ g g⁻¹; 270 271 Medeiros et al., 2006). Also mycose (a.k.a. trehalose), the recognized fungal saccharide (Niederer et al., 1989), is low compared to sucrose or glucose (Table SM1). This 272 273 indicates that fungal activity in these soils is minor. The ratio of sucrose-to-mycose was used to explore the microbial/fungal activities in these soil samples. The ratios were low
in the sites of DR, OL and MN due greater fungal activity there. Moreover, the small
sugar levels in these samples confirm low primary production and alteration of soil
biomass.

The results of these natural compounds were also analyzed by principal 278 component analysis (PCA) to understand the similarity among these sites in terms of 279 natural components. The PCA analysis output (explaining 98.5% of the variance with an 280 Eigen value of > 1) is shown in Figure 4b where two groups were recognized. The first 281 group included the AQ, AM, NZ, SH, AZ, MN and OL sites (explained by 53.4% as C1 282 283 dominant) and the second group included the DR, OG and JZ sites (explained by 45.1% as C2 dominant). The former sites are mainly located in areas with less vegetation and 284 relatively high human activities, where the latter sites are located in the city outskirts of 285 286 Riyadh in areas of relatively low human activities and more vegetation. This indicates that the vegetation cover has an impact on the characteristics and distributions of the 287 natural EOM in the region. 288

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3.2. Anthropogenic sources

Fossil fuel components such as <u>n</u>-alkanes, biomarkers (e.g. hopanes and steranes, Fig. 3c,d and Table SM1), and UCM, as well as the plasticizers (Table SM1 and Fig 3e) indicate anthropogenic inputs from traffic and litter (plastics) (Al-Mutlaq et al., 2002; Rushdi et al., 2005). The <u>n</u>-alkanes with a carbon number predominance of about unity (i.e., CPI ~ 1.0, Bray and Evans, 1961), after subtraction of the wax <u>n</u>-alkane contribution (Simoneit et al., 1991), suggest a source from vehicle/traffic emissions. Crude petroleum, the primary source of gasoline and diesel fuels, and lubricating oils, contains n-alkanes ranging up to about C_{35} with essentially no carbon number predominance (CPI ~ 1) (Simoneit, 1984) and with lower carbon number maxima in the range of C_{22} to C_{25} (Simoneit, 1984). These results indicate that fossil fuel utilization is a major input of organic matter to the soil samples, quite obvious for the AM and MN sites (Table SM1). The contribution of total petroleum residues ranges from 0.48 to 98.86 µg g⁻¹ of the total EOM.

304 The presence of hopane and sterane biomarkers in the samples confirms the input from traffic emissions (Simoneit, 1984). Hopanes and steranes are present in exhaust 305 particles from both auto and diesel engines (Simoneit, 1985). This particulate matter falls 306 307 out and becomes entrained in urban soil. The C-22 S/(R+S) epimer ratios of the homohopanes (C_{31} and C_{32}), which have been used as maturity indices for petroleum 308 (Seifert and Moldowan, 1978), range between 0.36-0.65 in the soil samples from the city, 309 310 further indicating the anthropogenic petroleum product inputs (Table SM1). Evidently, crude oil products are one of the major inputs of the OM to the soils. The predominant 311 petroleum component is derived from engine lubricating oil in vehicle emissions, and that 312 313 with direct oil leakage from vehicles is represented by the UCM (Table SM1). These petroleum residues were detected in samples from areas where truck traffic and industrial 314 factories are major activities (e.g. AM, MN, OG, SH). The absence of levoglucosan and 315 PAHs indicates that organic compounds from biomass burning, i.e., soot fallout 316 317 (Simoneit et al., 1999) are not a detectable input.

Discarded plastics (e.g. shopping bags) are probably the major sources of the plasticizers in the soils of Riyadh, because we have not detected them in blank sample analyses of solvents or the analytical procedures. The contribution of plasticizers in the soil OM ranges from 0.37 to 8.03 μ g g⁻¹ of the total EOM.

The results of the anthropogenic compounds were also analyzed by the PCA 322 323 statistical analysis to investigate the similarity among the different sites based on the anthropogenic components. The PCA analysis output (explaining 89.0% of the variance 324 325 of an Eigen value of > 1) is shown in Figure 4c where two site groups were observed. The first group (C1 and explains 61.9% of variance) included the AQ, NZ, DR, OR, AZ, 326 JZ and OL sites, which are inside the city with relatively more vegetation and 327 328 landscaping. The second group (C2 and explains 27.1% of the variance) includes only sites AM, SH and MN, which are characterized by the presence of stop stations for large 329 trucks and industrial factories with little vegetation. This indicates that the types of 330 331 human activities have an impact on the characteristics and the distribution of 332 anthropogenic sources of the EOM in the area.

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3.3. Anthropogenic versus natural sources

It is obvious that the contribution of natural biogenic versus anthropogenic sources varies and depends on the locations of the soil samples (Fig. 5). The contributions from natural sources (mainly vegetation) ranged from 1.5 to 46.0% of the total EOM in the soils of the city. These values are relatively lower than those reported previously (20-57%, Rushdi et al., 2005). This is attributed to less vegetation and arid conditions in and around Riyadh.

The anthropogenic sources were found to range from 54 to 99% of the total EOM. This includes fossil fuel combustion and unintentional crude oil leaks and spills (31 to 99%, based on <u>n</u>-alkanes, hopanes, steranes and UCM, and market place and leisure related activities 6% to 39% based on plasticizers, Figs. 5 and 6). The results confirmed

- that traffic is the major source of detrital anthropogenic OM and pollutants in Riyadh.
- Littering is another major source of organic pollutants in soils of the city (Fig. 6).



Figure 5. Pi plots of the petroleum residue (black), plasticizers (white) and natural (black and white stripes) fractions at different sites in Riyadh.



Figure 6. Ternary diagram showing the EOM compound compositions from petroleum residues, plasticizers and natural inputs.

4. Conclusions

The analysis of soil samples from the Riyadh metropolitan area showed that both natural biogenic and anthropogenic sources contributed to their OM contents. The natural sources of organic compounds are primarily from higher plant wax (i.e., vegetation) and ranged from 1.5 to 46.0% of the total EOM of the soils. The natural components included <u>n</u>-alkanols, <u>n</u>-alkanoic acids, <u>n</u>-alkanes (in part), and sterols, with concentrations lower in the city compared to the surrounding rural areas. The presence of the natural compounds from vegetation as major sources refuted the second hypothesis of this research (*the* sources of natural EOM are relatively low and are primarily from soil microbiota with lesser contributions from vegetation.).

The presence of <u>n</u>-alkanes with CPI ~1, hopanes, steranes, and UCM from urban traffic emissions and plasticizers from plastic litter and detritus (e.g. shopping bags) characterized the anthropogenic sources, and comprised from 54 to 99% of the total EOM. This, reasonably validated the first hypothesis of this work (*the EOM sources in soils of an urban arid region is mainly from traffic emissions*). The abundances of anthropogenic organic compounds in the soils of the city depend on fallout/washout at the sampling location and the types of urban activities.

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