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Songbirds as sentinels of mercury in terrestrial habitats of eastern North America

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Abstract Mercury (Hg) is a globally distributed environmental contaminant with a variety of deleterious effects in fish, wildlife, and humans. Breeding songbirds may be useful sentinels for Hg across diverse habitats because they can be effectively sampled, have well-defined and small territories, and can integrate pollutant exposure over time and space. We analyzed blood total Hg concentrations from 8,446 individuals of 102 species of songbirds, sampled on their breeding territories across 161 sites in eastern North America [geometric mean Hg concentration = 0.25 µg/g wet weight (ww), range <0.01–14.60 µg/g ww]. Our records span an important time period—the decade leading up to implementation of the USEPA Mercury and Air Toxics Standards, which will reduce Hg emissions from

coal-fired power plants by over 90 %. Mixed-effects modeling indicated that habitat, foraging guild, and age were important predictors of blood Hg concentrations across species and sites. Blood Hg concentrations in adult invertebrate-eating songbirds were consistently higher in wetland habitats (freshwater or estuarine) than upland forests. Generally, adults exhibited higher blood Hg concentrations than juveniles within each habitat type. We used model results to examine species-specific differences in blood Hg concentrations during this time period, identifying potential Hg sentinels in each region and habitat type. Our results present the most comprehensive assessment of blood Hg concentrations in eastern songbirds to date, and thereby provide a valuable framework for designing and evaluating risk assessment schemes using sentinel songbird species in the time after implementation of the new atmospheric Hg standards.

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Keywords Bioaccumulation · Mercury · Passeriformes · Sentinel · Songbird

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Introduction

Sentinel species historically used to assess contaminant risk to ecosystem and human health have ranged from terrestrial and aquatic invertebrates (Brown and Luoma 1995; Bundy et al. 2007) to marine megafauna (Bossart 2006). A suitable sentinel species has at least the following characteristics: (1) utilizes a restricted home range that overlaps the area of interest, (2) integrates contaminant exposure across a defined period of time and space, (3) is widespread and abundant, permitting extensive sampling and (4) is reflective of contaminant availability or risk to an endpoint of interest (Beeby 2001; Basu et al. 2007; Burger and Gochfeld 2004, National Research Council 1991). Additionally, we should have a well-developed understanding of the basic biology of a sentinel species and how the contaminant of interest impacts their physiology (van der Schalie et al. 1999; Basu et al. 2007). The definition of “sentinel species” and efficacy of their use has been debated (Stahl 2008), but for our purposes, we define a sentinel in the simplest terms: a species that has the ability to accumulate a contaminant and can provide a baseline for understanding contaminant risk in a community or ecosystem (Beeby 2001).

Mercury (Hg) is a persistent environmental contaminant with global implications for human, wildlife, and ecosystem health (Driscoll et al. 2013; Wiener 2013). Despite the global ubiquity of Hg, wildlife exposure and risk is often determined at the local scale because Hg must be methylated by microbes before it can become readily available to wildlife (Wren et al. 1995; Scheuhammer et al. 2007, 2012) and Hg methylation is a local-scale process. The higher methylation potential of aquatic ecosystems, and the human health focus on fish consumption, have resulted in most Hg sentinel species being fish or fish-eating wildlife, especially fish-eating birds (Burger and Gochfeld 2004; Evers 2006; Grove et al. 2009).

Informed by a recent appreciation for the porosity of terrestrial-aquatic ecotones, terrestrial species with indirect ties to aquatic food webs, such as snakes, spiders, songbirds, and bats, are increasingly being sampled for Hg bioaccumulation (Cristol et al. 2008; Drewett et al. 2013; Yates et al. 2014). These invertebrate-eating species (hereafter “invertivores”) appear to bioaccumulate and biomagnify MeHg through the terrestrial food web in a

manner similar to aquatic species (Cristol et al. 2008; Newman et al. 2011). They also are sensitive to the impacts of this Hg exposure, with adverse impacts to invertivore songbird species including endocrine, immunological and neurological disruption (Wada et al. 2009; Lewis et al. 2013; Scoville and Lane 2013), altered song quality (Halling et al. 2010; McKay and Maher 2012), decreased flight performance (Carlson et al. 2014) and reduced reproductive success (Brasso and Cristol 2008; Jackson et al. 2011a; Varian-Ramos et al. 2013).

Songbirds (avian order Passeriformes) are the most numerous avian order, with over 5,000 species, occurring in almost every terrestrial ecosystem on earth. As such, they may be suitable as potential sentinels of Hg contamination in diverse terrestrial ecosystems of eastern North America (Osborne et al. 2011; Evers et al. 2012). Whereas other sentinel species, such as mink or osprey, are singular species with a wide geographic distribution (Basu et al. 2007; Grove et al. 2009), “songbirds” as a group represent a diversity of taxonomic and distributional characteristics that can complicate interpretation of contaminants data or use of songbirds in risk assessment. Because of this, most songbird Hg studies have focused on a single species or suite of species living in a particular habitat type, including riparian corridors (Evers et al. 2005; Custer et al. 2007; Cristol et al. 2008, Jackson et al. 2011b), tidal estuaries (Warner et al. 2010; Lane et al. 2011; Winder and Emslie 2011), freshwater bogs (Edmonds et al. 2012), upland forests of the southern Appalachians (Keller et al. 2014) and high elevation forests (Rimmer et al. 2005, 2010; Townsend et al. 2014). Research that integrates data from these distinct habitats is lacking, and a lack of understanding of differences among habitats limits the effectiveness of using songbirds as sentinels of Hg across terrestrial ecosystems.

Eastern North America has a well-documented portfolio of point-source (e.g., historic chlor-alkali plants) and atmospheric Hg inputs (e.g., current coal-fired power plants) leading to biological hotspots of Hg exposure and risk (Evers et al. 2007). In response to documentation of elevated exposure in natural resources across the country, paired with recognition of Hg releases through fossil fuel combustion, the USEPA promulgated the Mercury and Air Toxics Standards (MATS) Rule in 2011 to in-part reduce Hg emissions by 90 % from United States coal-fired power plants by 2015–2016 (USEPA 2011). Therefore, understanding current baseline conditions across diverse ecosystems is important for documenting biological responses to these emission reductions. In collaboration with a number of active songbird monitoring and research activities, spanning four ecological regions (see below) across the eastern United States and Canada (Fig. 1), we here document the trends in Hg exposure occurring within

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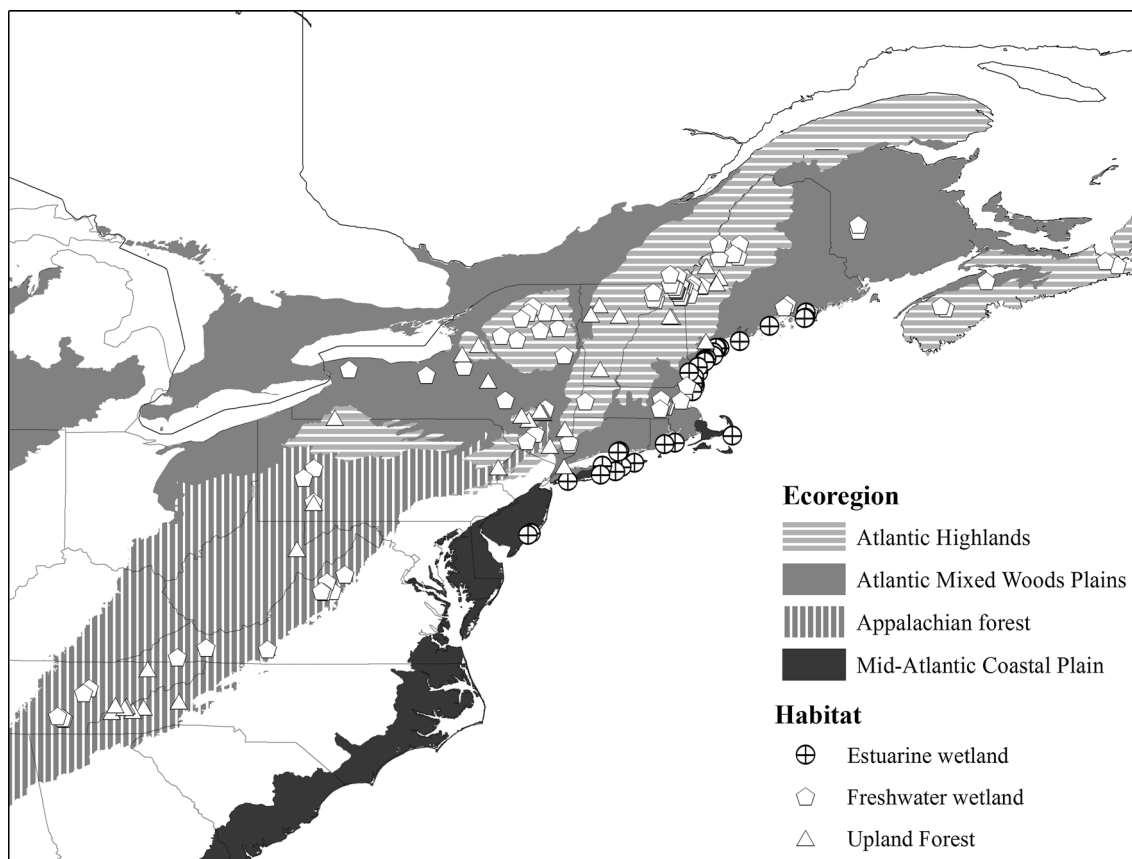


Fig. 1 Songbird capture sites during 1999–2012, illustrating differences in habitat types (freshwater wetlands, estuarine wetlands, and upland forests) and ecoregion (Mid-Atlantic Coastal Plain, Atlantic

Mixed Wood Plains, Atlantic Highlands, and Appalachian Forests). Detailed descriptions of sites are available in Supplemental Table 1

songbirds in the decade prior to implementation of the MATS rule. Songbirds were sampled in three broad habitat types—freshwater wetlands (including palustrine and lacustrine), tidal or estuarine wetlands, and upland forests. Our goals were twofold. First, to elucidate the factors affecting bioaccumulation of Hg in songbirds, we modeled Hg exposure as a function of age, foraging guild and habitat across all species in all ecoregions. Second, we used information from our model to highlight potential sentinel species of terrestrial Hg in future risk assessments. We provide summary information for blood Hg concentrations in these species, to aid in future comparisons after the anticipated MATS Hg emission reductions.

Materials and methods

Samples were collected opportunistically from 1999 to 2012 in association with a variety of projects across a wide geographic area (Fig. 1). Site locations and characteristics are outlined in Supplementary Table 1. Because we were interested in linking habitat characteristics of breeding

territories with recent Hg exposure, we selected blood, which is strongly influenced by recent local exposure, as the tissue of most relevance. Blood was collected only during breeding seasons. At the southern sites (Virginia, Tennessee, North Carolina, and West Virginia) we classified breeding season as beginning on 20 March, while the more northerly sites were sampled only after 1 May to account for the later nesting. By restricting sampling to the breeding season, we focused on the life history stage at which most species of songbirds are restricted to small breeding territories and Hg in their blood is likely to have accumulated locally.

All mist netting, banding and sampling followed appropriate state and federal permits. Although specific sample collection methods varied among researchers, in general, birds were captured using mesh mist nets at each site and species, age, and sex were determined by external characteristics. Because of the difficulty in determining age at high resolution for many species, we limited age classification to either juvenile (hatch-year) or adult (after hatch-year). We were unable to include sex as a factor in this analysis because we would have had to remove over

1,900 records to exclude birds of unknown sex. Samples for which sex was known consisted of 2,900 females and 3,629 males.

Blood samples were collected by puncturing the cutaneous ulnar vein with a sterile 26- to 28-gauge needle and placing a heparinized capillary tube at the puncture site to gather the blood. Between 30 and 150 μ l of blood was collected depending on the size of the bird, which was always less than 1 % of the bird's weight, as recommended (Fair et al. 2010); birds were released unharmed. Once the blood was collected, researchers sealed tubes at both ends with Crito-caps[®] and placed them in labeled 10 ml glass vacutainers (BD, Franklin Lakes, NJ, USA) and a zipped plastic bag to prevent breakage or desiccation. All samples were kept on ice after sampling, and then frozen at approximately -20°C later the same day for preservation until analysis.

Data used for this multi-year synthesis were generated at a number of laboratories. Although there were some differences in sample preparation and analytical methods, all analyses included quality control samples to allow evaluation of accuracy and precision, and all laboratories utilized atomic absorption spectroscopy to measure total Hg concentrations. Labs used either USEPA method 7473 (Trace Elements Research Lab at Texas A&M, Biodiversity Research Institute, College of William and Mary, Acadia University, Center for Environmental Systems Engineering at Syracuse University) or USEPA method 1631 modified (Brooks Rand, CEBAM). Total Hg was measured as a proxy for MeHg as bird blood had previously been found to be $>90\%$ MeHg (Rimmer et al. 2005; Edmonds et al. 2010). Although the specific analytical protocols may have varied at each lab, all sample runs included, at minimum, analytical blanks, sample replicates, and certified reference materials (DORM and/or DOLT) with every batch of 20–30 samples. All labs and analyses reported their relative percent difference for replicates to be $<10\%$. All samples were above the minimum detection limits and recovery of standard reference material was 90 to 110 %.

We assigned each of the 102 species sampled to one of two trophic categories (invertivore or omnivore) based upon individual species accounts of their diets during the breeding season (Poole and ed 2013). The 59 species classified as invertivores were those that consume $>90\%$ insect and invertebrate prey during the breeding season, whereas the 43 omnivores consumed 30–90 % invertebrates during the breeding season with the remainder comprised of plant matter (Table 1; includes scientific names of all species).

We distinguished sites with known Hg point sources nearby (hereafter “contaminated”) from those without known point sources (hereafter “uncontaminated”). Many

of the sites where point-source contamination is an issue have already been summarized elsewhere (Jackson et al. 2011a, b; Cristol et al. 2008; Folsom and Evers 2008; Franceschini et al. 2009). The designation of “contaminated” can be fairly arbitrary since sites in the Northeast with high atmospheric deposition are classified as “uncontaminated” even though Hg loading may be higher than sites with small or localized point sources (which would be classified as “contaminated”). The purpose of this analysis is not to assess differences between contaminated and uncontaminated sites, but to instead consider our questions of interest about how Hg varies between habitats, ages, and guilds. We, therefore, accounted for differences in Hg loading among sites, by nesting site within contamination regime as a random effect in all models. We include geometric mean Hg concentrations for contaminated and uncontaminated birds in Supplementary Table 2.

We used the USEPA Level II Ecoregions to group sites into one of four ecoregions: (1) Ozark, Ouachita-Appalachian Forest, hereafter Appalachian forest, (2) Mixed Woods Plains, where our samples were restricted to a subsection of this ecoregion, so more accurately described as Atlantic Mixed Woods Plains, (3) Mid-Atlantic Coastal Plain and (4) Atlantic Highlands (Commission for Environmental Cooperation Working Group 1997). Within each of these ecoregions, Hg cycling was hypothesized to vary based on proximity to aquatic habitat. We characterized the dominant habitat type associated with each site into one of three broad groups based on criteria that could be reliably distinguished from field notes, global positioning system (GPS) locations and site visits. Based on Cowardin et al. (1979), the three broad habitat groups to which we could reliably assign sites were (1) estuarine or tidal wetlands, (2) freshwater wetlands (including emergent, scrub–shrub, and forested wetlands bordering rivers, streams, lakes, ponds and bogs), and (3) uplands (forest or open habitats without dominant water features nearby) (Table 2).

Modeling effects of guild, age, and habitat

Although a strength of using songbirds as sentinels is that one species or another occurs in nearly every habitat type, few species were—or could have been—sampled across all habitats (Table 1). Because our modeling effort could not, therefore, focus on species differences, we combined species into foraging guilds, while accounting for species differences by nesting species within guild as a random effect. Similarly, we accounted for site and any known point source contamination by including a random effect of site nested within contamination status (contaminated or uncontaminated). Sites were also classified by EPA Level II Ecoregion, which was included as a random effect. We examined habitat type and individual bird age class as main

Table 1 Sample sizes for each species sampled in each habitat type

Species code ^a	Species common name ^a	Species scientific name ^a	Foraging guild	Total (N)	Estuarine wetland (N)	Freshwater wetland (N)	Upland forest (N)
ACFL	Acadian Flycatcher	<i>Empidonax vireescens</i>	Invertivore	16		16	
ALFL	Alder Flycatcher	<i>Empidonax alnorum</i>	Invertivore	2		2	
AMRE	American Redstart	<i>Setophaga ruticilla</i>	Invertivore	24		11	13
AMRO	American Robin	<i>Turdus migratorius</i>	Omnivore	41		26	15
BAOR	Baltimore Oriole	<i>Icterus galbula</i>	Invertivore	8		8	
BARS	Barn Swallow	<i>Hirundo rustica</i>	Invertivore	15	9	6	
BAWW	Black-and-White Warbler	<i>Mniotilta varia</i>	Invertivore	19		13	6
BCCH	Black-capped Chickadee	<i>Poecile atricapillus</i>	Omnivore	43		41	2
BGGN	Blue-gray Gnatcatcher	<i>Poliophtila caerulea</i>	Invertivore	6		6	
BHCO	Brown-headed Cowbird	<i>Molothrus ater</i>	Omnivore	3		3	
BHNU	Brown-headed Nuthatch	<i>Sitta pusilla</i>	Omnivore	1		1	
BHVI	Blue-headed Vireo	<i>Vireo solitarius</i>	Invertivore	17		9	8
BITH	Bicknell's Thrush	<i>Catharus bicknelli</i>	Invertivore	71			71
BLJA	Blue Jay	<i>Cyanocitta cristata</i>	Omnivore	7		7	
BLPW	Blackpoll Warbler	<i>Setophaga striata</i>	Invertivore	25		2	23
BOBO	Bobolink	<i>Dolichonyx oryzivorus</i>	Omnivore	4	4		
BOCH	Boreal Chickadee	<i>Poecile hudsonicus</i>	Omnivore	3		3	
BRCR	Brown Creeper	<i>Certhia americana</i>	Invertivore	2		2	
BRTH	Brown Thrasher	<i>Toxostoma rufum</i>	Omnivore	8		7	1
BTBW	Black-throated Blue Warbler	<i>Setophaga caerulescens</i>	Invertivore	16		8	8
BTNW	Black-throated Green Warbler	<i>Setophaga virens</i>	Invertivore	10		3	7
CACH	Carolina Chickadee	<i>Poecile carolinensis</i>	Omnivore	166	1	165	
CARW	Carolina Wren	<i>Thryothorus ludovicianus</i>	Invertivore	512		510	2
CAWA	Canada Warbler	<i>Cardellina canadensis</i>	Invertivore	21		21	
CERW	Cerulean Warbler	<i>Setophaga cerulea</i>	Invertivore	3		3	
CHSP	Chipping Sparrow	<i>Spizella passerina</i>	Omnivore	5	1	3	1
CLSW	Cliff Swallow	<i>Petrochelidon pyrrhonota</i>	Invertivore	16		16	
COGR	Common Grackle	<i>Quiscalus quiscula</i>	Omnivore	25		25	
COYE	Common Yellowthroat	<i>Geothlypis trichas</i>	Invertivore	56	3	51	2
CSWA	Chestnut-sided Warbler	<i>Setophaga pensylvanica</i>	Invertivore	4		2	2
DOWO	Downy Woodpecker	<i>Picoides pubescens</i>	Omnivore	8	1	6	1
EABL	Eastern Bluebird	<i>Sialia sialis</i>	Invertivore	756		756	
EAKI	Eastern Kingbird	<i>Tyrannus tyrannus</i>	Omnivore	7		7	
EAPH	Eastern Phoebe	<i>Sayornis phoebe</i>	Invertivore	32		29	3
EATO	Eastern Towhee	<i>Pipilo erythrophthalmus</i>	Omnivore	10		10	
EAWP	Eastern Wood-Pewee	<i>Contopus virens</i>	Invertivore	9		8	1
EUST	European Starling	<i>Sturnus vulgaris</i>	Omnivore	1		1	
FISP	Field Sparrow	<i>Spizella pusilla</i>	Omnivore	12		12	
GCFL	Great Crested Flycatcher	<i>Myiarchus crinitus</i>	Invertivore	16		15	1
GRCA	Gray Catbird	<i>Dumetella carolinensis</i>	Omnivore	99	5	91	3
GRSP	Grasshopper Sparrow	<i>Ammodramus savannarum</i>	Omnivore	2		2	
HAWO	Hairy Woodpecker	<i>Picoides villosus</i>	Omnivore	6		4	2
HETH	Hermit Thrush	<i>Catharus guttatus</i>	Omnivore	238		157	81
HOFI	House Finch	<i>Haemorhous mexicanus</i>	Omnivore	1		1	
HOWA	Hooded Warbler	<i>Setophaga citrina</i>	Invertivore	14		9	5
HOWR	House Wren	<i>Troglodytes aedon</i>	Invertivore	319	1	317	1
INBU	Indigo Bunting	<i>Passerina cyanea</i>	Omnivore	100		99	1

Table 1 continued

Species code ^a	Species common name ^a	Species scientific name ^a	Foraging guild	Total (N)	Estuarine wetland (N)	Freshwater wetland (N)	Upland forest (N)
KEWA	Kentucky Warbler	<i>Geothlypis formosa</i>	Invertivore	2		2	
LEFL	Least Flycatcher	<i>Empidonax minimus</i>	Invertivore	1			1
LISP	Lincoln's Sparrow	<i>Melospiza lincolni</i>	Omnivore	88		88	
LOWA	Louisiana Waterthrush	<i>Parkesia motacilla</i>	Invertivore	59		37	22
MAWA	Magnolia Warbler	<i>Setophaga magnolia</i>	Invertivore	26		19	7
MAWR	Marsh Wren	<i>Cistothorus palustris</i>	Invertivore	23	22	1	
MOWA	Mourning Warbler	<i>Geothlypis philadelphia</i>	Invertivore	1			1
MYWA	Myrtle Warbler	<i>Setophaga coronata</i>	Omnivore	19		13	6
NAWA	Nashville Warbler	<i>Oreothlypis ruficapilla</i>	Invertivore	31		29	2
NESP	Nelson's Sparrow	<i>Ammodramus nelsoni</i>	Invertivore	126	123	3	
NOCA	Northern Cardinal	<i>Cardinalis cardinalis</i>	Omnivore	91		91	
NOFL	Northern Flicker	<i>Colaptes auratus</i>	Omnivore	1		1	
NOPA	Northern Parula	<i>Setophaga americana</i>	Invertivore	5		5	
NOWA	Northern Waterthrush	<i>Parkesia noveboracensis</i>	Invertivore	9		9	
NRWS	Northern Rough-winged Swallow	<i>Stelgidopteryx serripennis</i>	Invertivore	34		34	
OROR	Orchard Oriole	<i>Icterus spurius</i>	Invertivore	7		7	
OVEN	Ovenbird	<i>Seiurus aurocapilla</i>	Invertivore	122		60	62
PAWA	Palm Warbler	<i>Setophaga palmarum</i>	Invertivore	66		66	
PIWA	Pine Warbler	<i>Setophaga pinus</i>	Invertivore	2		2	
PRAW	Prairie Warbler	<i>Setophaga discolor</i>	Invertivore	1		1	
PROW	Prothonotary Warbler	<i>Protonotaria citrea</i>	Invertivore	3		3	
PUFI	Purple Finch	<i>Haemorhous purpureus</i>	Omnivore	3		3	
RBGR	Rose-breasted Grosbeak	<i>Pheucticus ludovicianus</i>	Omnivore	1			1
RBNU	Red-breasted Nuthatch	<i>Sitta canadensis</i>	Omnivore	9		9	
RBWO	Red-bellied Woodpecker	<i>Melanerpes carolinus</i>	Omnivore	19		19	
REVI	Red-eyed Vireo	<i>Vireo olivaceus</i>	Invertivore	328		197	131
RUBL	Rusty Blackbird	<i>Euphagus carolinus</i>	Invertivore	75		75	
RWBL	Red-winged Blackbird	<i>Agelaius phoeniceus</i>	Omnivore	107	7	100	
SALS	Saltmarsh Sparrow	<i>Ammodramus caudacutus</i>	Invertivore	1,126	1,126		
SAVS	Savannah Sparrow	<i>Passerculus sandwichensis</i>	Omnivore	4	2	2	
SCJU	Slate-colored Junco	<i>Junco hyemalis</i>	Omnivore	65		1	64
SCTA	Scarlet Tanager	<i>Piranga olivacea</i>	Omnivore	13		10	3
SESP	Seaside Sparrow	<i>Ammodramus maritimus</i>	Invertivore	38	38		
SOSP	Song Sparrow	<i>Melospiza melodia</i>	Omnivore	370	20	348	2
SWSP	Swamp Sparrow	<i>Melospiza georgiana</i>	Omnivore	60	7	53	
SWTH	Swainson's Thrush	<i>Catharus ustulatus</i>	Omnivore	111		33	78
TRES	Tree Swallow	<i>Tachycineta bicolor</i>	Invertivore	2,155	116	2,038	1
TRFL	Traill's Flycatcher	<i>Empidonax spp.</i>	Invertivore	16		16	
TUTI	Tufted Titmouse	<i>Baeolophus bicolor</i>	Omnivore	52		49	3
VEER	Veery	<i>Catharus fuscescens</i>	Omnivore	106		27	79
WAVI	Warbling Vireo	<i>Vireo gilvus</i>	Invertivore	13		13	
WBNU	White-breasted Nuthatch	<i>Sitta carolinensis</i>	Omnivore	8		6	2
WEVI	White-eyed Vireo	<i>Vireo griseus</i>	Invertivore	9		7	2
WEWA	Worm-eating Warbler	<i>Helmitheros vermivorum</i>	Invertivore	20		20	
WIFL	Willow Flycatcher	<i>Empidonax traillii</i>	Invertivore	10		10	
WIWA	Wilson's Warbler	<i>Cardellina pusilla</i>	Invertivore	3		3	
WIWR	Winter Wren	<i>Troglodytes hiemalis</i>	Invertivore	3			3

Table 1 continued

Species code ^a	Species common name ^a	Species scientific name ^a	Foraging guild	Total (N)	Estuarine wetland (N)	Freshwater wetland (N)	Upland forest (N)
WOTH	Wood Thrush	<i>Hylocichla mustelina</i>	Omnivore	126		49	77
WTSP	White-throated Sparrow	<i>Zonotrichia albicollis</i>	Omnivore	67		32	35
YBCH	Yellow-breasted Chat	<i>Icteria virens</i>	Omnivore	2		2	
YBCU	Yellow-billed Cuckoo	<i>Coccyzus americanus</i>	Invertivore	1		1	
YBFL	Yellow-bellied Flycatcher	<i>Empidonax flaviventris</i>	Invertivore	3		1	2
YEWA	Yellow Warbler	<i>Setophaga petechia</i>	Invertivore	19		19	
YTVI	Yellow-throated Vireo	<i>Vireo flavifrons</i>	Invertivore	3		3	
YTWA	Yellow-throated Warbler	<i>Setophaga dominica</i>	Invertivore	5		5	

^a Based on American Ornithologists' Union Check-list of North American birds, 7th edition

Table 2 Blood mercury sample size for each all songbirds captured in each ecoregion and habitat type

Guild	Age	Atlantic highlands		Mid-Atlantic coastal plain	Atlantic mixed woods plains			Appalachian forests		Total
		Freshwater wetlands	Upland forest	Estuarine wetlands	Estuarine wetlands	Freshwater wetlands	Upland forest	Freshwater wetlands	Upland forest	
Invertivore	Adult	359	291	188	1,129	238	9	2,539	64	4,817
	Juvenile	52	11	9	112	110	0	1,211	12	1,517
Omnivore	Adult	344	328	10	14	184	21	799	67	1,767
	Juvenile	55	35	8	16	93	3	132	3	345
	Total	810	665	215	1,271	625	33	4,681	146	8,446

effects, because these have been hypothesized to affect Hg exposure in other avian taxa (Evers et al. 2005; Eagles-Smith et al. 2009). Blood Hg data were natural log-transformed to meet assumptions of parametric tests and to normalize residuals. All analyses were conducted in R (R Development Core Team 2012). Unless specified otherwise, all model-generated results are presented as back-transformed least squares mean blood Hg concentrations in $\mu\text{g/g}$ (or ppm, wet weight) with 95 % confidence intervals (R Package “lmerTest”, Kuznetsova et al. 2014).

In the first tier of this analysis, we evaluated the effects of habitat type (freshwater wetland, estuarine wetland, or upland), age (juvenile or adult), and foraging guild (invertivore or omnivore) using linear mixed-effects models, with species nested within guild, site nested within contamination status, and ecoregion as random effects (R Package “lme4”, Bates et al. 2013). We also included all two-way interactions (habitat \times age, habitat \times guild, age \times guild) in the global model. Models with significant interaction terms ($p < 0.05$) confounded our ability to properly interpret the importance of main effects. Therefore, we conducted a second tier analysis where we divided the data by guild and ran a mixed-effects model with main effects of habitat, age, and habitat \times age, and species nested within guild, site nested within contamination regime, and ecoregion were again included as random effects. Pairwise differences between ages and habitats

were assessed using Tukey's honest significant difference (HSD) test (R Package “lsmeans”, Lenth 2014).

Potential sentinels for Hg

Our second objective was to provide Hg concentrations in songbird sentinel species in each of the habitat and ecoregion we sampled. We are using the term sentinel in the broadest sense, to highlight songbird species with robust data in our database that can be used for future comparisons. To do this, we selected only sites that were not point-source contaminated—and were thus most likely to be influenced primarily by atmospheric emissions—in order to provide Hg concentrations for these species before the MATS rule goes into effect. Model results indicated that juvenile birds bioaccumulated Hg in blood at lower concentrations than adults; therefore, only the analysis results from data obtained from adult birds were used when considering sentinel candidates. After isolating species with robust sample sizes (i.e., sample sizes greater than 20) in each habitat-ecoregion pair, we ranked each species in terms of mean Hg concentration. We highlight the top ten highest mean Hg levels in freshwater wetland and upland habitats and the top three highest in estuarine habitats, where species richness was low. We include descriptive statistics of blood Hg concentrations for future comparisons to these sentinels in Table 3; descriptive statistics of

Hg concentrations for all birds used in the large model are shown in Supplementary Table 2.

Results

Modeling approach

Analysis of the mixed-effects model with species nested within guild and site as random effects revealed that blood Hg concentrations were significantly influenced by age ($F_{1,8444} = 536.3$, $p < 0.001$) and marginally influenced by habitat ($F_{2,8443} = 2.9$, $p = 0.067$) and guild ($F_{1,8444} = 3.5$, $p = 0.064$). However, the significant habitat \times guild ($F_{2,8443} = 5.8$, $p = 0.003$), habitat \times age ($F_{2,8443} = 58.7$, $p < 0.001$), and guild \times age ($F_{1,8444} = 384.6$, $p < 0.001$) interactions confounded our ability to interpret the main effects. Sample sizes for each guild, age and habitat are shown in Table 2.

We simplified our approach by separately analyzing invertivores and omnivores (Fig. 2). Invertivore songbird blood Hg concentrations were influenced by age ($F_{1,6332} = 425.6$, $p < 0.001$) and an interaction of habitat \times age ($F_{2,6331} = 79.4$, $p < 0.001$) but not habitat alone ($F_{2,6331} = 0.4$, $p = 0.67$). Adult invertivores in freshwater wetland exhibited 1.97 times higher blood Hg concentrations than adult invertivores in upland forests ($t = 4.4$, $p = 0.0003$) (Fig. 2a). Similarly, adult invertivores in estuarine wetland habitats had 1.65 times higher blood Hg concentrations than adult invertivores in upland forests; however, the difference was not significant ($t = 2.1$, $p = 0.3$) (Fig. 2a). Adult invertivores living in both wetland habitat types exhibited higher Hg concentrations than juvenile invertivores living in the same habitats; adult Hg levels were 7.01 times higher than juveniles in freshwater wetlands ($t = 81.7$, $p < 0.0001$) and 3.92 times higher than juveniles in estuarine wetlands ($t = 20.9$, $p < 0.0001$).

Hg levels in omnivores were influenced by age ($F_{1,2110} = 38.2$, $p < 0.001$) and a habitat \times age interaction ($F_{2,2109} = 3.9$, $p = 0.02$). Unlike adult invertivores, significant differences in blood Hg did not occur consistently among the wetland and upland habitats; adults in freshwater wetlands were 1.99 times higher than adults in upland forest ($t = 4.4$, $p = 0.0004$) but this trend was not significant for estuarine wetlands (Fig. 2b). There were no significant differences between adults and juveniles living in wetland habitats but adults in upland forest had significantly higher Hg concentrations than juveniles in upland forests ($t = 3.2$, $p = 0.02$).

Sentinel species

We identified ten sentinel species for freshwater wetlands in both Atlantic Highlands (Fig. 3a) and Appalachian

forests (Fig. 3b) but only four sentinel species in Atlantic Mixed Woods Plains (Fig. 3c). The candidate sentinel species for freshwater wetlands varied between ecoregion and we identified only one species, Tree Swallow, with suitable sample sizes across all ecoregions to be considered a cross-region common sentinel. Red-eyed Vireo was found in each ecoregion but limited sample size in Atlantic Mixed Woods plains precluded comparison. Song Sparrow and Hermit Thrush occurred in only two out of the three ecoregions and regionally unique species, such as the Carolina Wren in Appalachian forests and the Rusty Blackbird and Palm Warbler in Atlantic Highlands, cannot be compared to other ecoregions.

In the upland habitat type, we could identify ten sentinel species in the Atlantic Highlands (Fig. 4a) but only two in the Appalachian forests (Fig. 4b). Only Wood Thrush and Red-eyed Vireo were considered common across the Atlantic Highlands and Appalachian forests and regionally unique species included Bicknell's Thrush and Blackpoll Warbler in the Atlantic Highlands. Estuarine habitats were relatively species poor and only the Saltmarsh Sparrow met our criteria in both Mid-Atlantic Coastal Plain and Atlantic Mixed Woods Plains ecoregions (Fig. 5). Descriptive statistics for all adults in each of these sentinel species at uncontaminated sites are summarized in Table 3, while descriptive statistics for all 102 species included in our model (across contamination regime, ages, habitats and ecoregions) are summarized in Supplementary Table 2.

Discussion

Mercury pollution is expected to decrease incrementally in the next few years, following the implementation of the MATS rule, as the approximately 1,400 coal- and oil-fired electric generating units at the 600 power plants in the United States reduce emissions through the installation of updated control technologies (USEPA 2011). Our dataset, spanning the decade before the MATS Rule is implemented, shows that across all studied ecoregions, foraging guild, habitat, and age can influence Hg exposure in songbirds. As expected, ecoregions were inhabited by different species, and we used the dataset to generate a list of potential Hg sentinel species in each, along with baseline blood Hg concentrations to aid in future comparisons after the MATS Hg emissions reductions go into effect.

Our modeling results illustrate that, even at the coarse designations described here, foraging guild, habitat, and age were all related to songbird blood Hg concentrations and should be accounted for when comparing across species or locations. In general, invertivores had higher blood Hg concentrations than omnivores and habitat type was important for some age and guild groups; for example,

Table 3 Descriptive statistics for blood Hg concentrations (µg/g, ww) for adult sentinel species on uncontaminated sites in each habitat-ecoregion pair

Habitat	Ecoregion	Species	N	Arithmetic mean	Arithmetic SE	Geometric mean	Geometric SE ^a	Maximum
Freshwater wetlands	Atlantic highlands	Hermit Thrush	106	0.157	0.025	0.098	0.008	1.900
		Lincoln's Sparrow	80	0.232	0.020	0.168	0.016	0.821
		Nashville Warbler	28	0.094	0.013	0.074	0.010	0.254
		Ovenbird	36	0.068	0.005	0.058	0.006	0.124
		Palm Warbler	51	0.607	0.065	0.497	0.043	2.815
		Red-eyed Vireo	70	0.236	0.025	0.169	0.017	1.313
		Rusty Blackbird	47	0.642	0.044	0.573	0.042	1.544
	Appalachian forest	Swainson's Thrush	32	0.098	0.006	0.092	0.006	0.191
		Tree Swallow	28	0.338	0.022	0.320	0.020	0.640
		White-throated Sparrow	25	0.063	0.008	0.052	0.007	0.161
		Carolina Chickadee	20	0.071	0.008	0.062	0.007	0.140
		Carolina Wren	142	0.246	0.016	0.207	0.010	1.660
		Eastern Bluebird	98	0.102	0.006	0.089	0.004	0.418
		Gray Catbird	55	0.131	0.028	0.097	0.008	1.571
Atlantic mixed woods plains	House Wren	21	0.122	0.012	0.111	0.011	0.269	
	Indigo Bunting	22	0.138	0.075	0.040	0.012	1.670	
	Northern Cardinal	20	0.056	0.014	0.038	0.007	0.293	
	Red-eyed Vireo	27	0.148	0.035	0.102	0.015	0.923	
	Song Sparrow	93	0.119	0.008	0.092	0.007	0.373	
	Tree Swallow	723	0.147	0.003	0.136	0.002	1.290	
	Black-capped Chickadee	12	0.151	0.023	0.136	0.017	0.347	
	Common Yellowthroat	14	0.245	0.060	0.168	0.042	0.880	
	Hermit Thrush	21	0.177	0.024	0.141	0.023	0.382	
	Red-eyed Vireo	8	0.338	0.044	0.307	0.060	0.480	
	Red-winged Blackbird	47	0.212	0.027	0.157	0.017	0.821	
	Song Sparrow	31	0.166	0.026	0.114	0.018	0.522	
	Swamp Sparrow	5	0.419	0.119	0.366	0.092	0.864	
	Traill's Flycatcher	7	0.152	0.019	0.142	0.023	0.212	
Tree Swallow	67	0.275	0.026	0.192	0.021	0.996		
Yellow Warbler	6	0.094	0.042	0.050	0.028	0.288		

Table 3 continued

Habitat	Ecoregion	Species	N	Arithmetic mean	Arithmetic SE	Geometric mean	Geometric SE ^a	Maximum
Upland forest	Atlantic highlands	Bicknell's Thrush	64	0.097	0.006	0.087	0.005	0.286
		Blackpoll Warbler	23	0.058	0.003	0.056	0.003	0.082
		Hermit Thrush	64	0.064	0.004	0.058	0.003	0.173
		Ovenbird	52	0.047	0.003	0.043	0.003	0.096
		Red-eyed Vireo	94	0.079	0.005	0.066	0.004	0.266
		Slate-colored Junco	38	0.052	0.004	0.049	0.003	0.151
		Swainson's Thrush	76	0.088	0.005	0.080	0.004	0.238
		Veery	56	0.055	0.004	0.049	0.003	0.165
		White-throated Sparrow	34	0.041	0.004	0.036	0.003	0.153
		Wood Thrush	40	0.080	0.006	0.072	0.006	0.180
		American Redstart	4	0.062	0.008	0.060	0.009	0.072
		American Robin	4	0.196	0.134	0.106	0.063	0.597
		Black-and-White Warbler	2	0.226	0.086	0.208	0.084	0.312
		Black-throated Blue Warbler	4	0.054	0.006	0.053	0.006	0.067
		Blue-headed Vireo	3	0.119	0.023	0.114	0.024	0.153
		Ovenbird	2	0.090	0.037	0.082	0.036	0.127
Red-eyed Vireo	30	0.113	0.014	0.083	0.013	0.300		
Slate-colored Junco	16	0.044	0.006	0.039	0.005	0.088		
Veery	13	0.095	0.025	0.073	0.015	0.372		
Wood Thrush	24	0.099	0.008	0.091	0.008	0.204		
Marsh Wren	10	0.744	0.060	0.723	0.059	1.134		
Saltmarsh Sparrow	79	0.954	0.039	0.882	0.041	1.888		
Seaside Sparrow	12	1.060	0.058	1.041	0.060	1.328		
Tree Swallow	107	0.230	0.010	0.209	0.009	0.698		
Nelson's Sparrow	120	0.511	0.023	0.464	0.018	2.270		
Saltmarsh Sparrow	892	0.888	0.017	0.773	0.014	3.746		

Scientific names are shown in Table 1

^a Calculated using the delta method

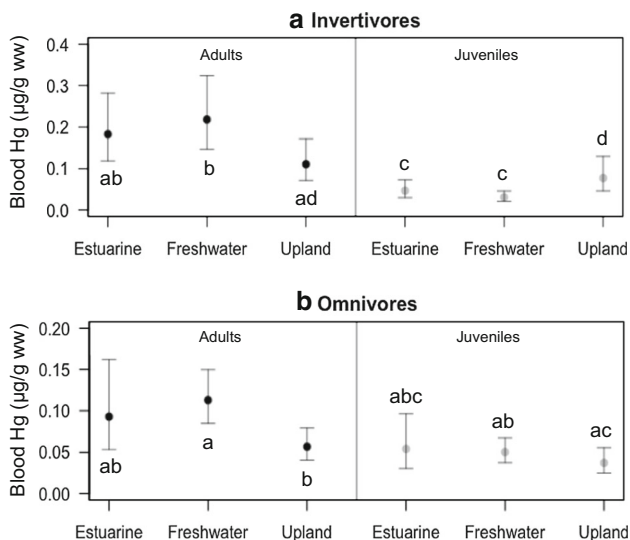


Fig. 2 Songbird blood Hg concentrations ($\mu\text{g/g}$, ww) for each age class in each habitat type for **a** invertivores and **b** omnivores [back-transformed $\mu\text{g/g}$ wet weight (ww), from the second tier mixed effects model, accounting for site and species effects]. Error bars represent 95 % confidence intervals; different letters indicate statistically significant Tukey HSD differences between pairwise groups

adult invertivores exhibited consistently higher blood Hg exposure in wetland habitats than in upland habitats. Generally, adults had higher blood Hg concentrations than juveniles within each habitat type and the strongest relationships between habitat and avian blood Hg were found in adult invertivores. Other studies have found similarly complex relationships of Hg exposure across habitats, but never across multiple species and broad geographic areas (Winder and Emslie 2011; Alberts et al. 2013).

Invertivores likely show elevated Hg concentrations over omnivores because a larger percentage of their diet comes from higher trophic levels in the food web (Poole 2013), thereby, providing more opportunity for biomagnification of MeHg in their prey. Invertivores in wetland habitats likely also exploit the aquatic subsidy of emergent aquatic insects that accumulate Hg directly from sediments (Gray 1993; Nakano and Murakami 2001). Because MeHg production occurs largely in aquatic-associated habitats (Driscoll et al. 2013), birds foraging on emergent aquatic insects rather than seeds and fruit are likely exposed to more MeHg (Jackson et al. 2011b). Birds do not need to feed directly on emergent aquatic insects to be exposed; spiders, and other predatory invertebrates also effectively transfer, and most importantly biomagnify, aquatic MeHg into the terrestrial food web, either from floodplain biota or sediment-dwelling aquatic emergent insects (Cristol et al. 2008). This link has also been demonstrated with other environmentally persistent contaminants, such as PCBs (Walters et al. 2008, 2010).

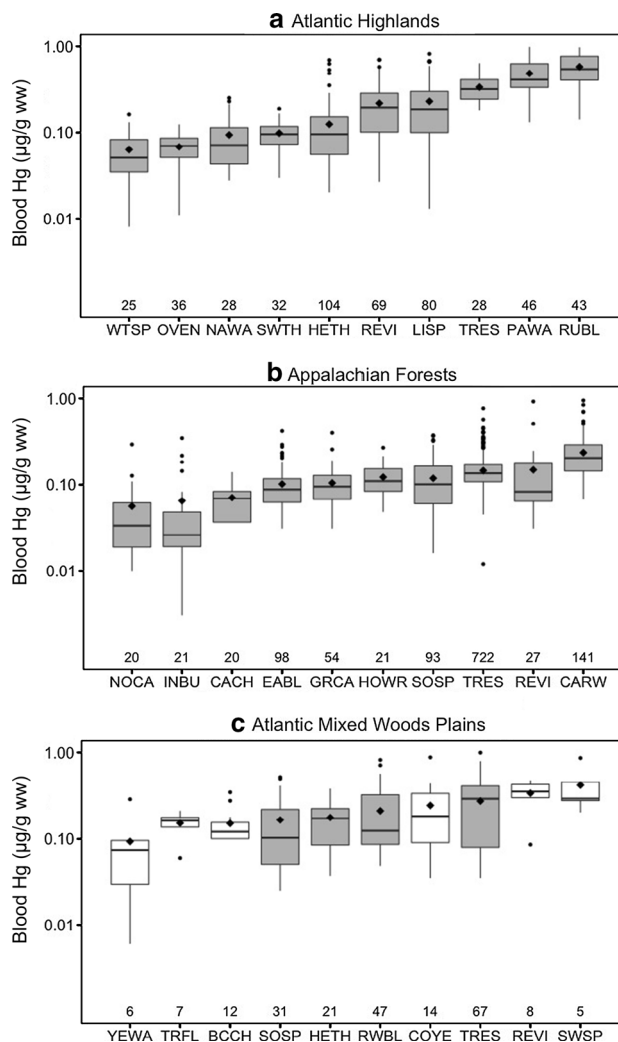


Fig. 3 Blood Hg concentrations ($\mu\text{g/g}$, ww) of adult freshwater wetland songbird sentinel species in **a** Atlantic Highlands, **b** Appalachian Forests, and **c** Atlantic Mixed Woods Plains, plotted on a log-scale. Gray shaded boxes identify suitable Hg sentinel species for each ecoregion ($N > 20$). Diamonds indicate mean blood Hg concentrations for each species. Bottom and top of boxes indicate first and third quartiles, respectively. Upper and lower whiskers indicate the highest and lowest values that are within 1.5 times the interquartile range. Data outside of this range are plotted as points. Common and scientific names of species are listed in Table 1

Adult and juvenile songbirds exhibited differences in Hg exposure across habitats and guilds. The disparity between adult and juvenile blood Hg concentrations is likely driven by the complex dynamics of blood MeHg concentrations in chicks as they reduce their blood burdens of MeHg through somatic growth and depuration through feather growth (Condon and Cristol 2009). In rapidly growing young birds, mass accretion can exceed or dampen MeHg accumulation, which manifests as lower body Hg concentrations compared to non-growing birds with identical Hg exposure (French et al. 2010). Although the juvenile birds

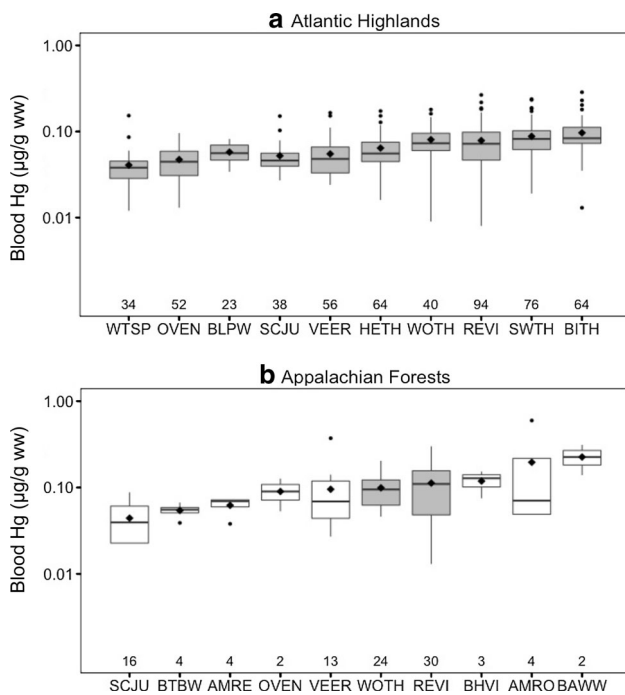


Fig. 4 Blood Hg concentrations ($\mu\text{g/g ww}$) of adult upland songbird sentinel species in **a** Atlantic Highlands and **b** Appalachian Forests, plotted on a log-scale. *Gray shaded boxes* identify suitable Hg sentinel species for each ecoregion ($N > 20$). *Diamonds* indicate mean blood Hg concentrations for each species. *Bottom and top of boxes* indicate first and third quartiles, respectively. *Upper and lower whiskers* indicate the highest and lowest values that are within 1.5 times the interquartile range. Data outside of this range are plotted as *points*. Common and scientific names of species are listed in Table 1

we sampled were full grown, early-life MeHg dynamics could influence Hg concentrations into the fledgling time period and beyond. Perhaps the most important mechanism responsible for the disparity between juvenile and adult birds is the strong influence of feather growth in young birds. As new feathers grow, MeHg from the blood and muscle tissue binds to the keratin in the growing feather, which is then permanently sequestered in feather tissue, effectively eliminating it from internal tissues such as blood and organs (Condon and Cristol 2009). Because songbirds experience multiple molts during their juvenile stage, many of the blood samples for juveniles are likely obtained from birds that recently depurated a large amount of Hg into growing feathers.

Our findings allow a qualitative examination of species and habitats at risk for Hg bioaccumulation in the eastern United States. The suite of well-sampled species from each habitat type offers a baseline for the variation seen in Hg concentrations at uncontaminated sites (i.e. those without a known point source of contamination). A wide range of variation among individuals of the same species was observed and average concentrations varied by an order of magnitude among some species within a habitat type. For

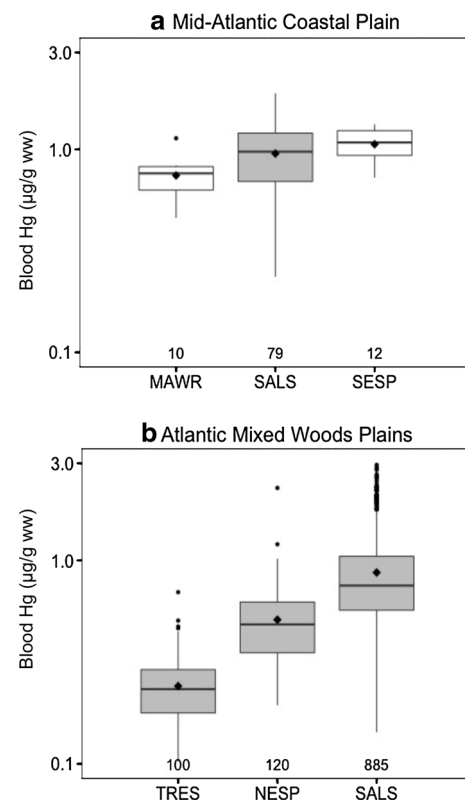


Fig. 5 Blood Hg concentrations ($\mu\text{g/g ww}$) of adult estuarine wetland songbird sentinel species in **a** Mid-Atlantic Coastal Plain and **b** Atlantic Mixed Woods Plains, plotted on a log-scale. *Gray shaded boxes* identify suitable Hg sentinel species for each ecoregion ($N > 20$). *Diamonds* indicate mean blood Hg concentrations for each species. *Bottom and top of boxes* indicate first and third quartiles, respectively. *Upper and lower whiskers* indicate the highest and lowest values that are within 1.5 times the interquartile range. Data outside of this range are plotted as *points*. Common and scientific names of species are listed in Table 1

the freshwater wetland habitat type, this variation might be driven by the wide diversity of vegetative structure and associated hydrology along with greater trophic complexity that may significantly influence methylation rates (Shanley and Bishop 2012).

In estuarine wetlands there is high potential for elevated methylation and transport through the food web to high trophic level species (Lane et al. 2011). These habitats have low species diversity compared to the range of species supported by diverse freshwater and upland habitats. While species diversity might be lower, multiple species are of particular concern because they are obligate estuarine specialists and are subject to a host of conservation issues ranging from sea level rise and urbanization to contaminant exposure. For example, Saltmarsh and Nelson's Sparrows show some of the highest blood Hg concentrations in our dataset and these species are also exhibiting population declines across their ranges (Lane et al. 2011).

Some invertivore songbirds of regional conservation concern that prefer wetland breeding habitats are conspicuously missing from our analysis. This is because they are rare or difficult to sample. In the future, we recommend also sampling wetland-dependent songbird species that are experiencing significant population declines, such as the Olive-sided Flycatcher (*Contopus cooperi*) and Canada Warbler (*Cardellina canadensis*), to expand upon the information presented here and to help determine if Hg is an important adverse stressor to their health, reproduction, and survival. Environmental Hg loads originating from the deposition of atmospheric Hg may already be causing widespread reproductive harm for some species of high conservation concern, such as the Rusty Blackbird (Edmonds et al. 2010, 2012).

Although we report the most comprehensive dataset on songbird Hg exposure to date, further investigations are needed to fully understand the impact of Hg exposure on songbirds in eastern North America. For example, we focused on breeding season Hg concentrations, but we should be concerned about Hg exposure throughout a bird's life cycle, especially for neotropical migrants that overwinter in tropical areas with elevated environmental Hg loads. Our sampling design does not provide an ability to connect songbird blood Hg concentrations to abiotic environmental Hg levels, an important step in understanding if songbirds can serve as bioindicators of Hg loading to an ecosystem. We lack a detailed understanding of the effects of Hg on songbirds at a population level. Although short-term fieldwork on wild songbirds has determined Hg effects on reproductive success (Jackson et al. 2011a; Brasso and Cristol 2008), obtaining Hg estimates from sites with long-term banding operations would provide a better understanding of potential population-level impacts of Hg exposure. Promulgation of the MATS rule by the USEPA is an important step toward reducing atmospherically generated loads of Hg across the North American landscape and therefore potentially reducing harmful body burdens in birds. Our findings provide a foundation from which to gauge songbird response to the new MATS regulations and an important step forward in assessing the risk of environmental Hg exposure to songbirds.

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Conflict of interest The authors declare that they have no conflict of interest.

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