

The Effect of Human Serum Albumin and Hematocrit on the Cake
Collapse Temperature of Lyophilized Red Blood Cells

Running Title: Cake Collapse Temperature of RBCs

Daniel E. Runyon and Adam Z. Higgins

School of Chemical, Biological and Environmental Engineering, Oregon State University,
Corvallis, Oregon 97331-2702, USA

Address correspondence to:

Adam Higgins

School of Chemical, Biological and Environmental Engineering

Oregon State University

102 Gleeson Hall

Corvallis, OR 97331-2702, USA

Email: adam.higgins@oregonstate.edu

Phone: +1-541-737-6245

Fax: +1-541-737-4600

Abstract

Freeze-drying, or lyophilization, has shown great promise in addressing many of the logistical challenges of storing and preserving red blood cells (RBCs). A crucial part of any RBC lyophilization protocol is the primary drying temperature, which affects the sample drying rate and the dried cake's ability to form a stable glassy solid. Primary drying is most efficient just below the temperature at which the porous structure of the cake begins to collapse, known as the cake collapse temperature. In this short report we utilize freeze-drying microscopy to examine the effects of human serum albumin (HSA) and hematocrit on the cake collapse temperature. Increasing the hematocrit from 0% to 20% significantly raised the cake collapse temperature from -37.8 °C to -34.8 °C. Addition of 5% HSA to a 20% hematocrit RBC suspension further increased the cake collapse temperature to -20.4 °C. This data provides a basis for future study of the relationship between cake collapse and overall cell survival, with the object of building a clinically-viable RBC lyophilization protocol.

Introduction

Using current biopreservation methods, RBCs can be stored at 4°C for up to 6 weeks.¹ While generally sufficient for the needs of industrialized countries, reliable access to refrigeration is often unavailable in developing countries or conflict zones where this storage method limits access to life-saving transfusions. Freeze-drying, or lyophilization, may provide the ideal solution by enabling long-term storage at room temperature. While no fully effective protocol has been reported for RBCs, this study seeks to build off of recent promising results²⁻⁷ to explore the effect of buffer excipients and hematocrit on cake collapse temperature, a critical step in establishing the optimum primary drying temperature for any future RBC freeze-drying protocol.

Materials and Methods

All chemicals were purchased from VWR. Whole human blood was collected from healthy adult volunteers with informed consent and stored at 4 °C for less than 2 weeks. To prepare samples for experiments, RBCs were first isolated and washed 3 times using 1X Dulbecco's phosphate-buffered saline. The cells were then suspended at 30% hematocrit in a buffer

containing 800 mM trehalose, 100 mOsm ADSOL and 6.6 mM potassium phosphate, and incubated at 37° C for 7 hours to load trehalose into the cytoplasm, as described previously.^{2,3} After incubation, the cells were centrifuged at 2300 g for 1 minute to remove the loading buffer, and a lyophilization buffer containing 100 mOsm ADSOL, 100mM trehalose, 6.6 mM potassium phosphate, and up to 10% HSA was added to produce solutions of varying hematocrit and HSA concentration.

Freeze-drying microscopy was used to assess the effect of hematocrit and HSA concentration on cake collapse temperature. A 5 μ L volume of RBC suspension was pipetted onto a glass coverslip on the temperature-controlled silver block of a Linkam FDCS 196 Cryostage mounted to a Leica DM 2500 upright microscope. A second coverslip was placed on top of the sample separated by a spacer, carefully working to avoid air bubbles, and the sample area was sealed. The sample was then cooled to -50 °C at 50 °C/min. After confirmation that the sample was completely frozen, a vacuum pump was turned on which generated a pressure of about 0.07 mbar and the temperature was raised to -40 °C at 4 °C/min or until a drying front began to form and then raised slowly at 0.1 °C/min until the cake collapsed. The temperature of overall collapse was determined as described previously.^{8,9} The temperature probe was confirmed to be accurate to within 0.2 °C by measuring the eutectic temperature of 10% saline.

Results

The cake collapse temperature of RBC suspensions was determined from examination of microscopic images of a thin frozen sample while it was dried under vacuum using a freeze-drying microscope. Figure 1A-B shows representative images of a 20% hematocrit suspension just before and just after cake collapse. The dried cake appears as an opaque strip moving from left to right. Upon reaching the collapse temperature, which in this case is -35.2 °C, the dried material loses structural integrity and translucent voids become visible.

Figure 1C shows cake collapse temperatures for RBC suspensions with hematocrits ranging from 0% to 20%. Statistical analysis revealed a significant effect of hematocrit on cake collapse temperature ($p = 0.0002$). Collapse temperatures increased from -37.8 ± 0.1 °C in cell-free

lyophilization buffer to -34.8 ± 0.2 °C in the 20% hematocrit solution. We also attempted to measure the cake collapse temperature at 40% hematocrit, but this solution was too opaque to determine visually whether a cake had formed or collapsed using the current microscopy method.

Addition of HSA to the lyophilization buffer also had a statistically significant effect on the cake collapse temperature ($p = 0.0001$), as illustrated in Figure 1D. Even adding minimal amounts of HSA significantly increased the cake collapse temperature from -34.8 ± 0.2 °C without HSA to -25.4 ± 1.2 °C with just 2.5% HSA. Further increasing the HSA concentration to 5% provided a smaller, yet significant increase to -20.4 ± 1.5 °C. However, increasing the HSA concentration to 10% did not significantly affect the overall collapse temperature.

Discussion

This study used freeze-drying microscopy to demonstrate that both HSA and hematocrit increase the cake collapse temperature of trehalose-loaded RBCs. The effect of hematocrit in particular may have implications for selection of the primary drying temperature for RBC freeze-drying. Whereas most previous studies of RBC freeze-drying have used relatively low cell densities,³⁻⁷ a clinical protocol would likely involve a hematocrit of 40% or higher. Such a high cell density would be expected to increase the cake collapse temperature by several degrees and potentially enable the use of a higher primary drying temperature.

Importantly, our results provide a framework for examining the relationship between cake collapse, primary drying temperature and RBC survival. It has previously been reported that hemolysis increases substantially when the primary drying temperature exceeds -35 °C.¹⁰ This is consistent with the cake collapse temperatures observed in the present study, suggesting a link between cake collapse and hemolysis. Future studies will enable more rigorous investigation of this idea.

Acknowledgements

We are grateful to the volunteer blood donors and to the Oregon State University Student Health Center for performing blood collections.

Disclosure Statement

No competing financial interests exist.

References

1. Hess J. An update on solutions for red blood cell storage. *Vox Sanguinis* 2006 Jul;91(1): 13-19.
2. Satpathy G, Torok Z, Bali R, et al. Loading red blood cells with trehalose: a step towards biopreservation. *Cryobiology* 2004;49: 123-136.
3. Torok Z, Satpathy G, Banerjee M, et al. Preservation of Trehalose-Loaded RBC by Lyophilization. *Cell Preservation Technology* 2005;3(2): 96-111
4. Kheirrolomoom A, Satpathy G, Torok Z, et al. Phospholipid Vesicles Increase the Survival of Freeze-Dried Human Red Blood Cells. *Cryobiology* 2005;51: 290-305.
5. Arav A, Natan D. Freeze drying of red blood cells: the use of directional freezing and a new radio frequency lyophilization device. *Biopreservation and Biobanking* 2012;10(4): 386-394.
6. Han Y, Quan GB, Liu XZ, Ma EP, Liu A, Jin P, Cao W. Improved preservation of human red blood cells by lyophilization. *Cryobiology* 2005;51: 152-164.
7. Zhou XL, Yuan J, Liu JF, Liu BL. Loading trehalose into red blood cells by electroporation and its application in freeze-drying. *Cryoletters* 2010;31(2): 147-156.
8. Fonesca F, Passot S, Cunin O and Marin M. Collapse Temperature of Freeze-Dried *Lactobacillicus Bulgaricus* Suspensions and Protective Media. *Biotechnology Progress* 2004;20: 229-238.
9. Yang G, Gilstrap K, Zhang A, et al. Collapse Temperature of Solutions Important for Lyopreservation of Living Cells at Ambient Temperature. *Biotechnology and Bioengineering* 2010;106(2): 247-259.
10. Rindler V, Luneberger S, Schwindke P, et. al. Freeze Drying of Red Blood Cells at Ultra-Low Temperatures. *Cryobiology* 1999;38: 2-15.

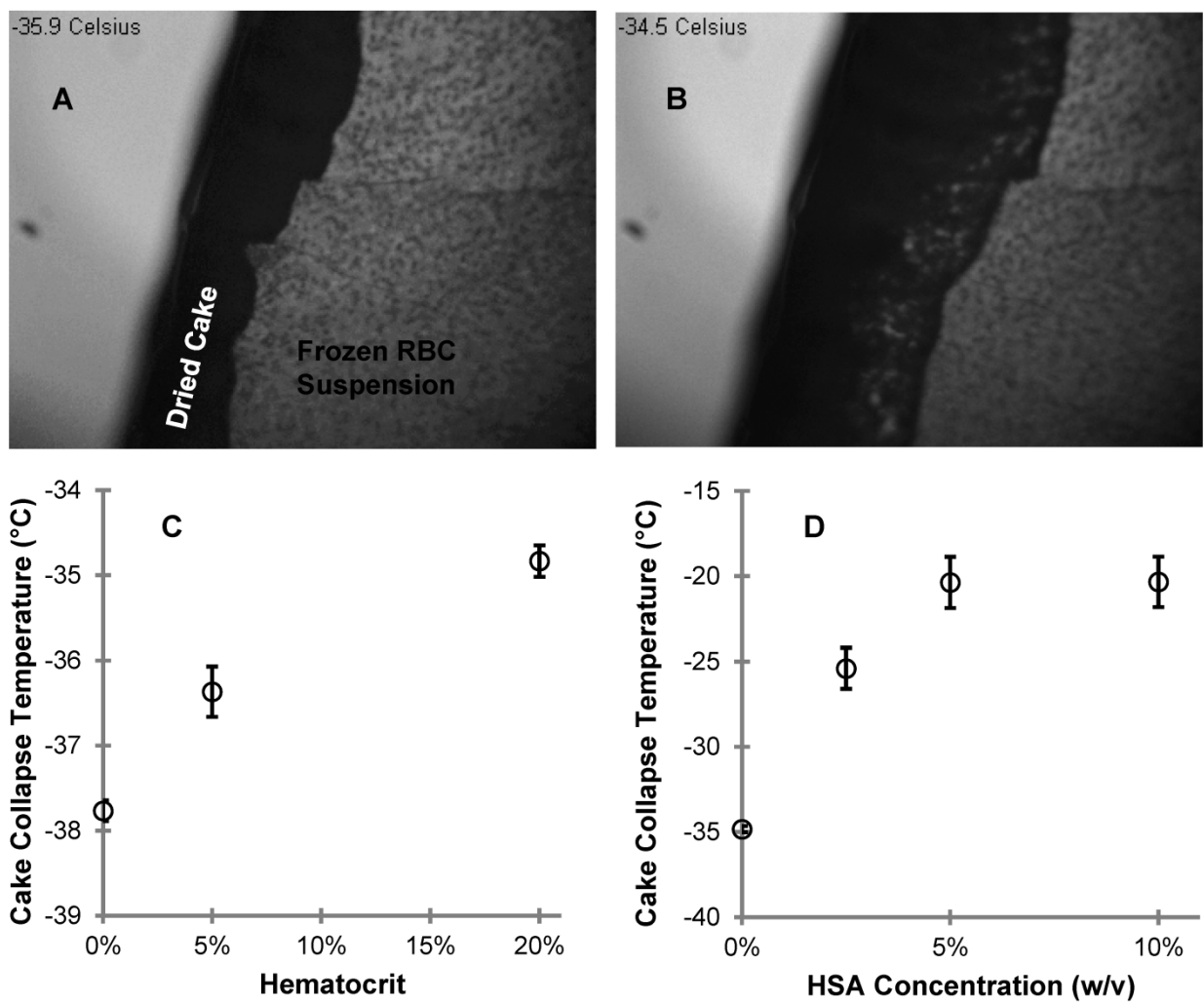


FIG. 1. (A) Representative image of a frozen RBC suspension (speckled region at the right) in which the dried cake (vertical black strip) is advancing left to right at $-35.9\text{ }^{\circ}\text{C}$, just before cake collapse. (B) Representative image of the same sample at $-34.5\text{ }^{\circ}\text{C}$, just after cake collapse, indicated by the white voids in the black strip of dried cake. (C) Effect of hematocrit on the overall cake collapse temperature of RBCs in HSA-free lyophilization buffer. (D) Effect of HSA concentration on the cake collapse temperature of 20% hematocrit RBC suspensions. Error bars represent the standard error of 3 replicates.