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ARTICLE

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# Direct links between the vernalization response and other key traits of cereal crops

Weiwei Deng<sup>1</sup>, M. Cristina Casao<sup>1,†</sup>, Penghao Wang<sup>1</sup>, Kazuhiro Sato<sup>2</sup>, Patrick M. Hayes<sup>3</sup>, E. Jean Finnegan<sup>1</sup> & Ben Trevaskis<sup>1</sup>

Transcription of the *VERNALIZATION1* gene (*VRN1*) is induced by prolonged cold (vernalization) to trigger flowering of cereal crops, such as wheat and barley. *VRN1* encodes a MADS box transcription factor that promotes flowering by regulating the expression of other genes. Here we use transcriptome sequencing (RNA-seq) and chromatin immunoprecipitation sequencing (ChIP-seq) to identify direct targets of *VRN1*. Over 500 genomic regions were identified as potential *VRN1*-binding targets by ChIP-seq. *VRN1* binds the promoter of *FLOWERING LOCUS T-like 1*, a promoter of flowering in vernalized plants. *VRN1* also targets *VERNALIZATION2* and *ODDSOC2*, repressors of flowering that are downregulated in vernalized plants. RNA-seq identified additional *VRN1* targets that might play roles in triggering flowering. Other targets of *VRN1* include genes that play central roles in low-temperature-induced freezing tolerance, spike architecture and hormone metabolism. This provides evidence for direct regulatory links between the vernalization response pathway and other important traits in cereal crops.

<sup>1</sup>CSIRO, Agriculture, Canberra, Australian Capital Territory 2601, Australia. <sup>2</sup>Institute of Plant Science and Resources, Okayama University, Kurashiki 710-0046, Japan. <sup>3</sup>Department of Crop and Soil Science, Oregon State University, Corvallis, Oregon 97331, USA. † Present Address: Max Planck Institute for Plant Breeding Research, Carl-von-Linné-Weg 10, 50829 Cologne, Germany. Correspondence and requests for materials should be addressed to B.T. (email: Ben.Trevaskis@csiro.au).

Many plants from temperate regions require prolonged exposure to low temperatures, or vernalization, to flower. Vernalization was originally defined in temperate cereals, such as wheat, barley and rye<sup>1</sup>. In these annual crops, vernalization is required to promote the transition to reproductive development, so that without vernalization plants will grow vegetatively for extended periods producing only leaves. Following vernalization, inflorescence development begins and will then proceed rapidly if plants are exposed to long day lengths, which accelerate flowering<sup>2</sup>.

The *VERNALIZATION1* gene (*VRN1*) encodes a MADS box transcription factor that is a central regulator of vernalization-induced flowering in cereals<sup>3–6</sup>. *VRN1* is expressed at low levels before vernalization and this limits the rate of progression towards flowering<sup>3–6</sup>. Prolonged exposure to low temperatures induces the transcription of *VRN1* (refs 3–6). Induction of *VRN1* begins rapidly with the onset of cold but the initial expression is low and several weeks at low temperatures are required for *VRN1* transcripts to accumulate to a level that will promote rapid flowering<sup>3–8</sup>. After vernalization, *VRN1* transcript levels remain elevated in leaves and at the shoot apex<sup>5,8</sup>. The expression of *VRN1* at the shoot apex is likely to promote inflorescence meristem identity, whereas the expression of *VRN1* in leaves is required for the long-day flowering response<sup>8–12</sup>.

Some alleles of *VRN1* are actively expressed without prior cold treatment<sup>3–6</sup>. These alleles have mutations in the proximal promoter or deletions/insertions in the first intron of *VRN1*, regions that appear to be required to maintain low levels of *VRN1* transcription before vernalization<sup>5,13</sup>. Chromatin in the first intron of *VRN1* has histone modifications that are thought to maintain an inactive transcriptional state, potentially explaining the role of the first intron in maintaining repression of *VRN1* before winter<sup>14</sup>. Sequences in the first intron might also regulate processing of the *VRN1* transcript<sup>15</sup>. Several active alleles of *VRN1* with deletions or insertions in the first intron have been identified in barley<sup>16–19</sup>. These reduce the vernalization requirement to different extents and have been used to breed varieties that flower without vernalization, which are grown where winters are warm or where crops are sown in spring<sup>19,20</sup>. In addition to flowering without vernalization, varieties of wheat or barley that carry active alleles of *VRN1* typically have reduced freezing tolerance<sup>21,22</sup>.

Molecular analyses have identified potential downstream regulatory targets of the *VRN1* gene. These include *VRN2* and *VRN3*, which, like *VRN1*, influence vernalization requirement<sup>20</sup>. *VRN2* is a repressor of flowering that is expressed in long days before vernalization, but is downregulated in vernalized plants or in plants that carry active alleles of *VRN1* (refs 9,10,23–25). *VRN3* encodes the cereal orthologue of *FLOWERING LOCUS T* (hereafter referred to as *FT1*) (ref. 26). In *Arabidopsis*, the *FLOWERING LOCUS T* protein is expressed in leaves in long days and then transported to the shoot apex to accelerate inflorescence development<sup>27</sup>. It seems likely that *FT1* has a similar function in cereals<sup>28</sup>. *FT1* is expressed at low levels before vernalization, irrespective of day length, but is induced by long days in vernalized plants, or in plants that have active alleles of *VRN1* (refs 10,26). Thus, the expression of *VRN1* is a prerequisite for long-day induction of *FT1* in cereals. Another regulatory target of *VRN1* is a second MADS box transcription factor, *ODDSOC2*, which represses flowering but is downregulated by vernalization or in plants that have active alleles of *VRN1* (refs 29,30). In addition, a series of *C-REPEAT BINDING FACTOR* (*CBF*) genes, which are induced by low temperatures to increase freezing tolerance<sup>31</sup>, show reduced expression in lines that carry active alleles of *VRN1* (refs 32,33). Whether *VRN1* directly regulates any of these potential targets is unclear.

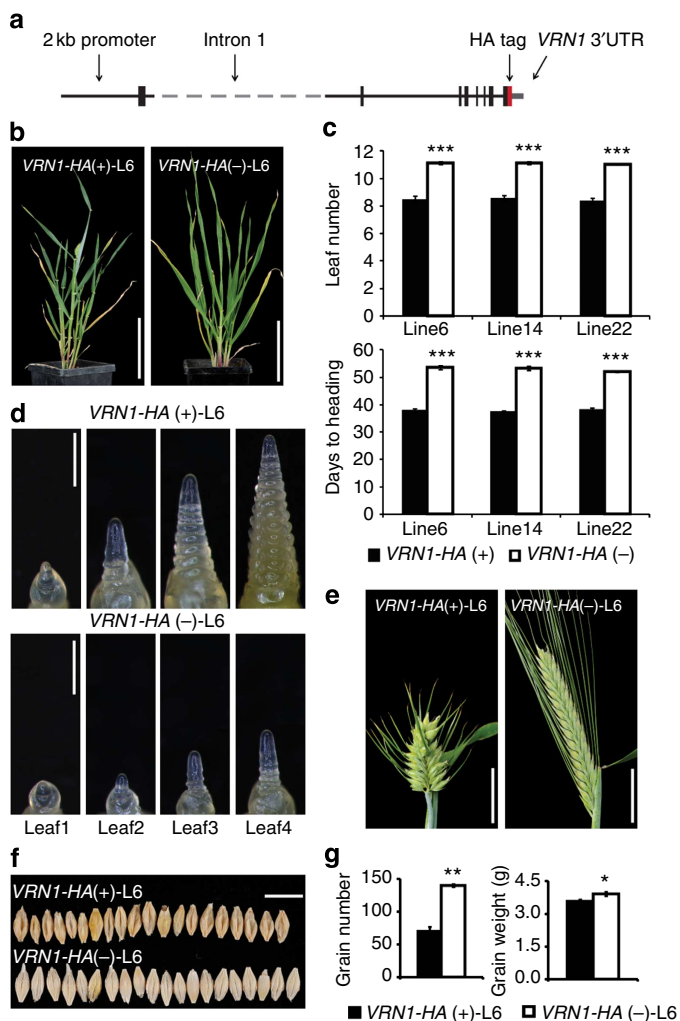
Optimal seasonal timing of flowering and grain production is critical to adapt cereals to temperate climates. The timing and duration of inflorescence development also influences key components of yield in cereal crops. For these reasons, *VRN1* is a major target for selection in cereal breeding. Understanding how *VRN1* functions can provide important insights into crop biology and inform future cereal breeding strategies. In this study, we used chromatin immunoprecipitation followed by deep sequencing (ChIP-seq) to identify sequences bound by the *VRN1* protein in barley, a model for temperate cereals and related grasses. This identified genes that act downstream of *VRN1* in the vernalization response and also revealed direct connections between *VRN1* and pathways that control other key traits.

## Results

### Epitope-tagged *VRN1* protein accelerates flowering of barley.

A transgene construct was designed to express the barley *VERNALIZATION1* protein (*VRN1*) fused to six copies of the haemagglutinin epitope tag (amino acid sequence YPYDVPDYA). The *VRN1-HA* gene construct was derived from the genomic sequence of *VRN1*, is driven by the endogenous promoter and has the 3' UTR of *VRN1*, but lacks most of the large first intron (Fig. 1a). This construct was transformed into the cultivar Golden Promise, a spring barley amenable to *Agrobacterium* transformation. Transgenic barley plants that carry the *VRN1-HA* construct flowered earlier than either wild-type parent line or sibling null lines, which were descended from the same transformation events but did not inherit the transgene (Fig. 1b–d and Supplementary Fig. 1). The earliest flowering lines expressing the *VRN1-HA* transgene also had reduced final height and fewer grains per spike, with altered grain morphology (Fig. 1e–g and Supplementary Fig. 1). The epitope-tagged version of the *VRN1* protein was detectable by immunoblot analysis using an anti-haemagglutinin monoclonal antibody (Fig. 2a). Compared with sibling nulls, plants carrying the *VRN1-HA* construct also had higher total transcript levels of *VRN1* (endogenous gene plus *VRN1-HA* transgene; Fig. 2b).

**Altered gene expression in lines expressing *VRN1-HA*.** To examine how the *VRN1-HA* construct triggers early flowering, gene expression was assayed by transcriptome sequencing (RNA-seq) at the second leaf stage when the transgenic lines approach inflorescence initiation. DEseq and EdgeR (see Methods) were applied to identify genes that are differentially expressed between a transgenic line (Line 6) and a sibling null control line descended from the same transformation event. These methods detected 678 and 1,283 differentially expressed transcripts, respectively, with 663 genes in common (Supplementary Data 1). Among the overlap between the two methods of analysis were 321 genes that were upregulated and 78 downregulated by twofold or more ( $P < 0.05$ , adjusted  $P$  value, DEseq or EdgeR). The upregulated genes include a number of genes that potentially promote floral development including *VRN1* and *VRN1*-like genes (*Barley MADS 3*, 8), and also four other MADS box genes including a *PANICLE ARCHITECTURE2*-like gene. In addition, *FT1* and *FT2* showed elevated transcript levels in the *VRN1-HA* positive line. Downregulated genes include a *SOC1*-like MADS box gene, a MYB transcription factor, histidine kinase genes and a cytokinin dehydrogenase. Quantitative reverse transcriptase-PCR (qRT-PCR) was used to further verify the expression data for *VRN1* and *FT1*, and a selection of other differentially expressed genes including *Barley MAD3* (Supplementary Fig. 2).



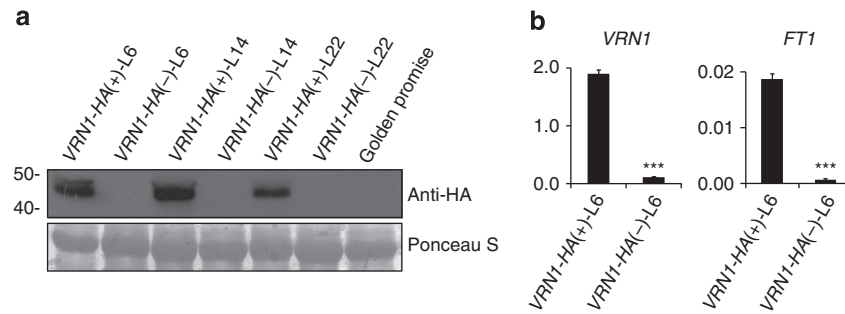
**Figure 1 | A VRN1-HA fusion accelerates the reproductive development in transgenic barley plants.** (a) Schematic representation of the VRN1-HA fusion construct, which fuses the VRN1 genomic sequence (minus most of first intron) to the coding sequence for six repeats of the haemagglutinin epitope tag (HA). (b) Homozygous plants carrying the VRN1-HA construct, from line 6 (VRN1-HA (+)-L6) compared with sibling null controls (VRN1-HA (-)-L6) that did not inherit the transgene. Scale bar, 10 cm. (c) The final number of leaves and days to heading of three transgenic lines versus sibling null control plants ( $n = 8$ ). (d) Apex development of VRN1-HA (+)-L6 and VRN1-HA (-)-L6 from the time of first leaf emergence until the fourth leaf emerged. Scale bar, 500  $\mu$ m. (e) Spike morphology of plants carrying the VRN1-HA construct versus a sibling null. Scale bar, 25 mm. (f) Twenty grains of VRN1-HA (+)-L6 compared with a sibling null control. Scale bar, 1 cm. (g) Grain number per plant and the weight of 100 grains of VRN1-HA (+)-L6 plant versus a sibling null control. Stars indicate statistical significance by Student's *t*-test: \* $P < 0.05$ , \*\* $P < 0.01$ , \*\*\* $P < 0.001$ .

**Identification of VRN1-binding targets.** The VRN1-HA protein was used for chromatin immunoprecipitation sequencing (ChIP-seq). Fragments of chromatin bound to the VRN1-HA protein were purified for DNA sequencing on the Illumina HiSeq 2000 sequencing platform. A VRN1-HA transgenic line (Line 6) was compared with a sibling line descended from the same original transgenic plant that did not inherit the transgene (null control). The fold enrichment of sequence reads mapped to the available barley genomic sequence for positive versus sibling null plants was then used to identify sequences bound by the VRN1-HA fusion protein. A total of 514 binding peaks, regions with a bias towards sequence reads in the samples from the positive versus

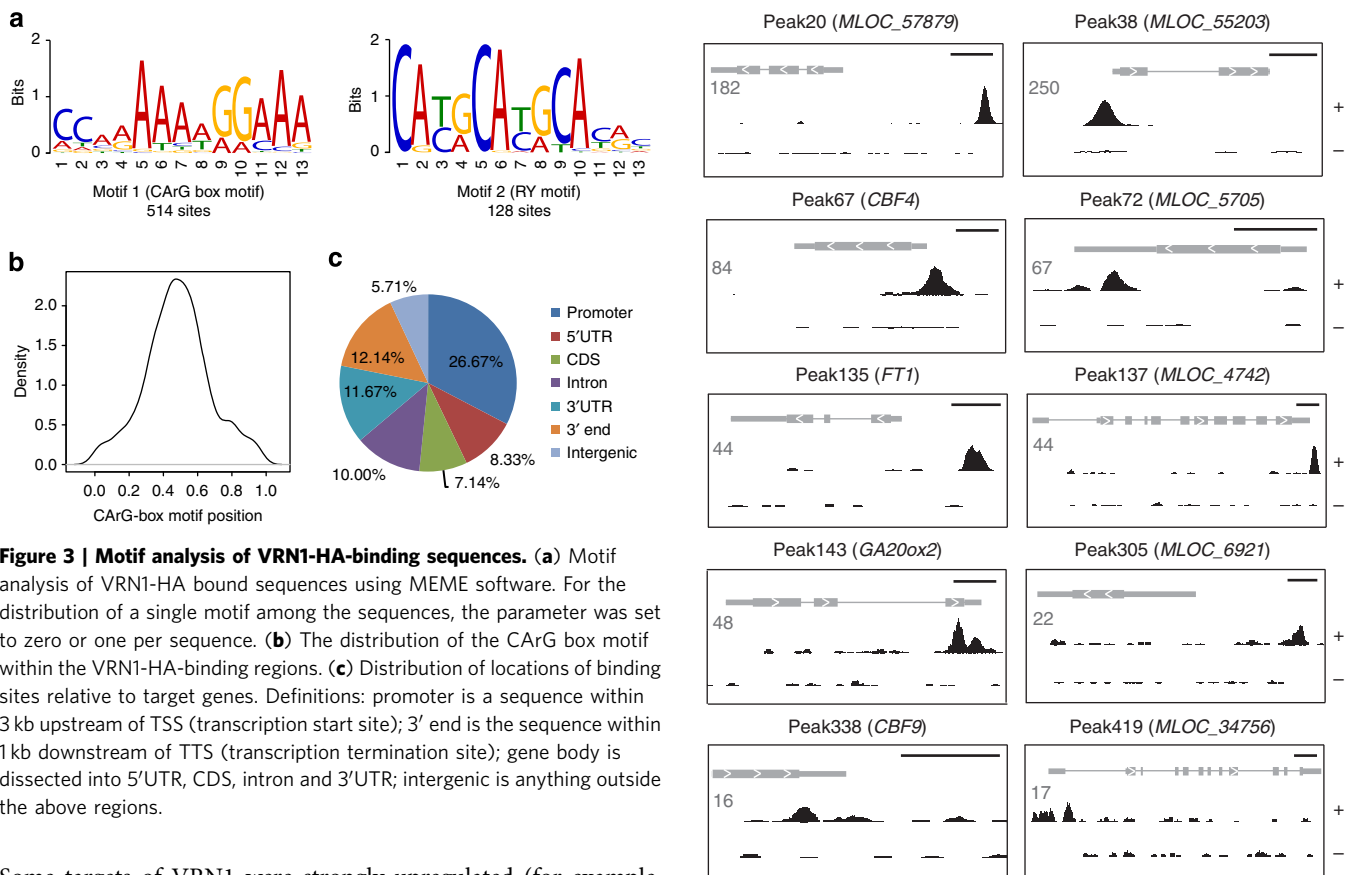
null lines, were identified using 20-fold enrichment as a cutoff value that was chosen on the basis of enrichment of false positives in the null sample ( $P < 0.05$ , model-based analysis of ChIP-seq<sup>34</sup>, Supplementary Data 2). This dropped to 379 with a 25-fold enrichment limit and 146 with 50-fold enrichment (Supplementary Data 2). Enriched sequence reads were distributed across binding peaks spanning regions from 220 to 1,300 bp in length. Motif analysis of enriched sequences identified two over-represented sequence motifs within the enriched sequences (Fig. 3a). One is similar to the CARG box motif (CC[A/T]<sub>6</sub>GG), the typical binding site for MADS box transcription factors. CARG box motifs were found in all the binding peaks, typically located at the centre of peak read distribution (Fig. 3b), though there was some sequence variation within the motifs compared with the canonical CARG box sequence (Fig. 3a). A second motif, a RY element (CATGCATG), was identified in a subset of 128 of the binding peaks (Fig. 3a).

The putative 514 VRN1-binding sites were located on 511 sequenced contigs of the fragmented barley genome assembly, with three having two potential binding sites. A total of 289 of these contigs included predicted genes and associated transcribed sequences. Many (~20%) correspond to transcription factor genes, encoding MYB, B3, Zinc Finger, B-ZIP, MADS box, homeodomain or Leucine Zipper DNA-binding proteins (Supplementary Data 2). Among these were low-temperature-induced *C-REPEAT BINDING FACTORS*; CBF2 (Peak 10), CBF4 (Peak 67) and CBF9 (Peak 338; Supplementary Data 2). Other potential targets are genes involved in hormone metabolism or regulators of hormone responses. For example, genes encoding enzymes involved in the biosynthesis or breakdown of gibberellins (GA20-oxidase 2, Peak 143), abscisic acid (ABA 8' Hydroxylase 2, Peak 346) or jasmonic acid (Allene oxide synthase, Peak 269; Supplementary Data 2). A number of binding peaks occurred near genes that are likely to regulate reproductive development. One of these was *FT1* (Peak 135). Others correspond to *CONSTANS*-like genes (Peaks 20, 38, 305), *SIX ROW SPIKE (VRS1)* (Peak 414), a *SUPPRESSOR OF OVER-EXPRESSION OF CONSTANS*-like MADS box gene (Peak 241) and an *ODDSOC2*-like MADS box gene (Peak 154; Supplementary Data 2). Mapping the locations of binding sites relative to target genes shows that the locations for putative VRN1-binding sites vary from the promoter to the gene body or the 3' ends of transcribed sequences (Figs 3c and 4). ChIP-PCR was used to verify the binding of the VRN1-HA protein to a subset of candidate target genes. The majority showed enrichment by both ChIP-PCR and ChIP-seq (Fig. 5a, Supplementary Fig. 3). In addition, ChIP-PCR was used to verify the binding of VRN1-HA to 20 targets that showed a range of different enrichment levels in ChIP-seq (Supplementary Fig. 3).

**VRN1-binding targets that potentially regulate flowering.** The potential for binding of VRN1 to influence transcription of nearby genes was examined by comparing ChIP-seq and RNA-seq data at the second leaf stage. A total of 33 differentially expressed transcripts identified by DEseq corresponded to predicted genes located on contigs that have potential VRN1-binding sites. An additional four transcripts identified as differentially expressed by EdgeR were derived from predicted genes located on contigs with VRN1-binding sites. Thus, 37 potential VRN1-binding sites are located near regions that encode transcripts that are differentially expressed at the second leaf stage in lines with elevated VRN1 levels (~13% of annotated transcripts located near VRN1-binding sites). These differentially expressed transcripts include *FT1* and also genes encoding zinc finger, B3, zinc finger and MYB transcription factors (Supplementary Data 3).



**Figure 2 | Detection of VRN1-HA fusion protein in transgenic barley.** (a) Immunoblot detection of the 6xHA-tagged VRN1 protein using a monoclonal anti-HA antibody in plants carrying the VRN1-HA construct, from three independent lines (6, 14 and 22) descended from independent transformation events, compared with a sibling null control (line 6) and non-transformed wild-type cv. Golden Promise. Relative positions of 40 and 50 kilodalton markers are indicated. (b) Expression level of VRN1 and FT1, relative to ACTIN, in VRN1-HA(+)–L6 compared with a sibling null control. Data are average of three biological replicates, error bar indicates s.e. \*\*\* $P < 0.001$  (Student's  $t$ -test).



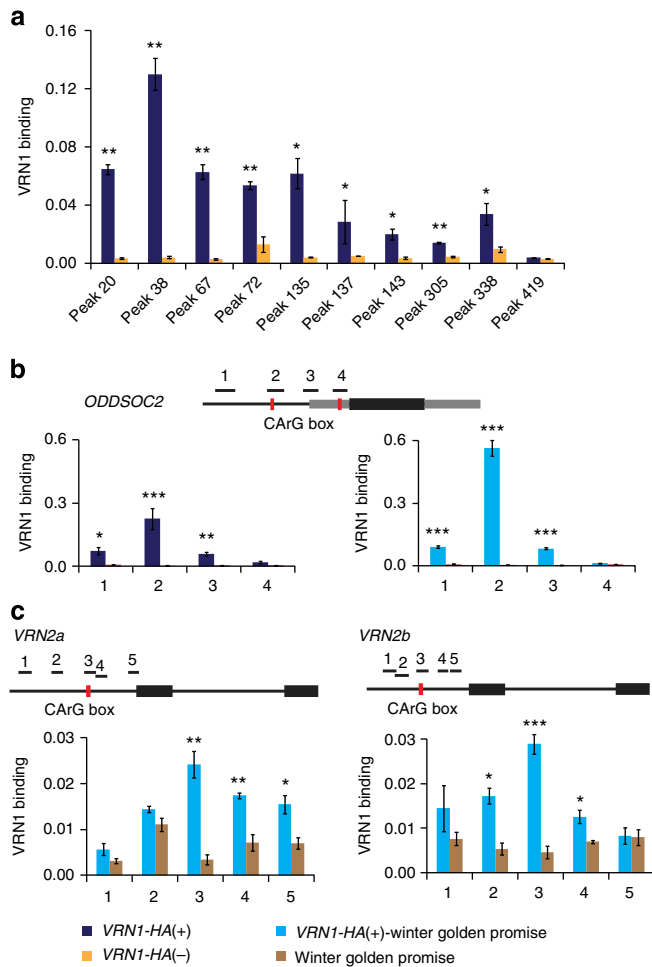
**Figure 3 | Motif analysis of VRN1-HA-binding sequences.** (a) Motif analysis of VRN1-HA bound sequences using MEME software. For the distribution of a single motif among the sequences, the parameter was set to zero or one per sequence. (b) The distribution of the CArG box motif within the VRN1-HA-binding regions. (c) Distribution of locations of binding sites relative to target genes. Definitions: promoter is a sequence within 3 kb upstream of TSS (transcription start site); 3' end is the sequence within 1 kb downstream of TTS (transcription termination site); gene body is dissected into 5'UTR, CDS, intron and 3'UTR; intergenic is anything outside the above regions.

Some targets of VRN1 were strongly upregulated (for example, FT1), whereas others were repressed (for example, CBF1) suggesting that the binding of VRN1 can activate or repress transcription of target genes (Supplementary Data 3).

**VRN1-HA binds to the promoters of ODDSOC2 and VRN2.** Some potential targets of VRN1 are absent from current barley genome sequence data sets, either due to incomplete genome sequence assembly (ODDSOC2) or due to naturally occurring deletions in the cultivar targeted for sequencing (VRN2). These cannot be identified by the generic ChIP-seq approach used here. Instead qPCR quantification of target DNA in VRN1-HA co-immunoprecipitated chromatin was performed (ChIP-PCR). For ODDSOC2 this was performed using the same Golden Promise transgenic line that was used for ChIP-seq. There was strong enrichment at three sites assayed in the promoter of this gene when chromatin was enriched by immunoprecipitation from the

**Figure 4 | Mapping of VRN1-HA-binding sites in putative target genes.** Visualization of VRN1-HA binding peaks in 10 selected candidate target genes. Maps were generated using IGV2.3 software. Target gene structures are shown in grey. Thick line indicates exons, medium line indicates untranslated regions and thin lines indicate the location of introns. Overlaid white arrowheads indicate the direction of transcription. The peak height, shown on the left side of each graph, indicates the maximum read number aligning to any region in the target gene. Peak heights for both positive (+) and negative control (–) are shown on the same scale, relative to the maximum number of sequence reads. Black scale bars indicate 500 bp.

positive versus null control line (Fig. 5b). Then, to allow ChIP-PCR analysis of VRN2, the VRN2 locus was introgressed into Golden Promise, along with a full-length version of VRN1 (requires cold to be actively transcribed) and a functional allele of



**Figure 5 | Verification of putative VRN1-HA-binding sites by ChIP-PCR.**

(a) ChIP-PCR was used to verify 10 potential VRN1-HA-binding sites using VRN1-HA line 6 and a sibling null control line. Peak numbers correspond to those in Supplementary Data 2. (b) ChIP-PCR data from multiple sites of the promoter of *ODDSOC2* in lines carrying the VRN1-HA construct versus sibling null controls. Assay was performed in cv. Golden Promise and in a vernalization-responsive version of the same cultivar, ‘Winter Golden Promise’. Target region is shown in grey. Thick line indicates first exon, medium line indicates transcribed regions and thin lines indicate 5' upstream sequence. The location of potential MADS box transcription factor-binding sites is indicated in red (CArG box). (c) The binding of VRN1-HA to *VRN2a* and *VRN2b* (duplicate genes at the *VRN2* locus of barley) assayed by ChIP-PCR. VRN1-HA binding was assayed in the Winter Golden Promise background. VRN1 binding indicates relative enrichment, immunoprecipitated DNA versus input chromatin, for three biological repeats. Error bars show s.e. Stars indicate statistical significance by Student's *t*-test: \* $P < 0.05$ , \*\* $P < 0.01$ , \*\*\* $P < 0.001$ .

*PHOTOPERIOD1*. As expected, this genotype requires vernalization to flower (hereafter referred to as ‘Winter Golden Promise’) but flowers rapidly when the VRN1-HA construct is present (Supplementary Fig. 4). The binding function of the VRN1-HA fusion in this background was verified by ChIP-PCR using a series of target genes identified by ChIP-seq, plus binding sites from the promoter region of *ODDSOC2*. All targets showed strong enrichment in this genotype (Fig. 5b and Supplementary Fig. 5). There was significant enrichment at sites in the promoters for both the duplicate copies of *VRN2*, including a region –480 to –360 bp relative to the ATG that is centred around a putative CArG box motif (Fig. 5c).

## Discussion

Plants that carry the *VRN1-HA* fusion construct have elevated *VRN1* transcript levels and flower earlier than wild-type plants (Figs 1 and 2 and Supplementary Fig. 1). Expression of the transgene is driven by the same promoter as the endogenous gene, so the elevated expression of *VRN1* in transgenic plants is likely due to increased overall *VRN1* gene copy number (endogenous plus transgene) and possibly the large intron deletion (8.9 kb) that is a feature of the transgene. The early flowering phenotype shows that the VRN1-HA fusion is biochemically active and has the expected biological function. Other phenotypes, such as reduced height and reduced spike length, were observed only in the earliest flowering lines and not in lines with more moderate acceleration of flowering (Supplementary Fig. 1) or in the Winter Golden Promise genotype (Supplementary Fig. 4). Similar phenotypes occur in some non-transgenic early flowering barleys (Supplementary Fig. 6) and are likely a consequence of rapid inflorescence development. Some barley mutants also have similar shortened spikes, which arise from altered rates of inflorescence development. For example, mutants that overexpress *HvAPETALA2* (ref. 35).

Between 146 and 514 direct targets of VRN1 were identified by ChIP-seq, depending on the fold enrichment used as a cutoff value (50 versus 20-fold). Even at the least stringent enrichment limit (20-fold), all the target regions identified contain a potential CArG box motif identified by MEME (Multiple EM for Motif Elicitation<sup>36</sup>; Supplementary Data 2). The false discovery rate values for peaks with more than 20-fold enrichment were generally below 5% (Supplementary Data 2). Thus, 20-fold enrichment was selected as an arbitrary cutoff point for subsequent discussion, although potentially some genuine targets have been omitted. It is important to note that the current genome sequence reference data sets do not represent the entire barley genome<sup>37</sup>, so some targets present in the ChIP-seq data sets will not be identified by the subsequent bioinformatic analysis. The ChIP-seq data set can be realigned to future genomic reference sequences of barley to address this limitation. Binding of VRN1 to putative targets was retested with a second antibody-epitope tag combination using a VRN1::GFP fusion construct<sup>12</sup>, showing that the enrichment of VRN1-binding sites by ChIP is not dependent on the HA-tag/antibody (Supplementary Figs 3 and 5).

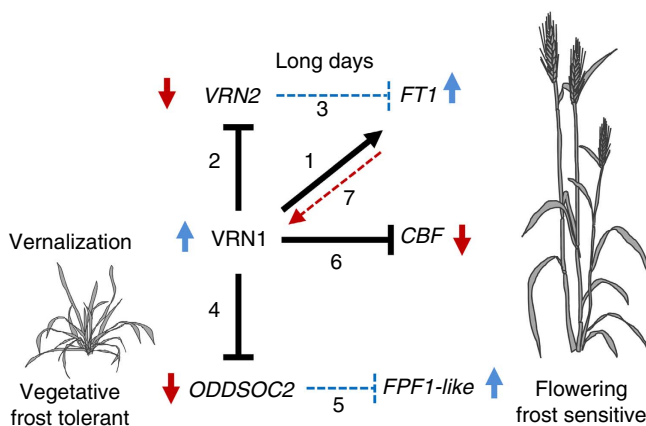
A key question for this study is which genes are targeted by VRN1 to promote rapid flowering after vernalization? Putative targets of VRN1 identified by ChIP-seq and ChIP-PCR include known regulators of flowering, such as *FT1*, *VRN2* and *ODDSOC2*. These genes are regulated by vernalization and by active alleles of *VRN1*, consistent with the idea that these genes are downstream targets of the *VRN1* gene (see Introduction). *SOCI*-like and *CONSTANS*-like genes, which regulate the reproductive development of rice<sup>38,39</sup>, were also identified as direct targets of VRN1. *FT1* was identified as a direct binding target of VRN1 and also shows altered expression during early development in the early flowering *VRN1-HA* transgenic plants (Figs 2 and 4). Genetic activation of *FT1* is sufficient to accelerate the flowering of barley<sup>26</sup>, consistent with the idea that the activation of *FT1* (resulting from elevated *VRN1* expression) plays a major role in eliciting the early flowering phenotype of VRN1-HA transgenic plants.

*ODDSOC2* expression did not differ in Golden Promise plants that carry the *VRN1-HA* transgene versus sibling null lines, despite being a direct target of VRN1 (Supplementary Fig. 7). Golden Promise already carries an active endogenous allele of *VRN1* (*HvVRN1-1*), so *ODDSOC2* might be downregulated by default in this genotype. Comparing the expression of *ODDSOC2*

in a barley that requires vernalization to flower (low *VRN1* activity) versus a near-isogenic line that carries a highly active allele of *VRN1* (*HvVRN1-7*; Supplementary Fig. 8) shows that this gene has reduced expression when comparisons are made between genotypes with strongly contrasting *VRN1* activity (Supplementary Fig. 9), as reported previously<sup>29</sup>. Similarly, *VRN2* expression differed in the near-isogenic lines (Supplementary Fig. 9).

The finding that *VRN1* directly binds to the promoter of *FT1* extends current models of the vernalization response of cereals by revealing a direct connection between the vernalization and photoperiod-response pathways (Fig. 6). A requirement for *VRN1* to bind to the promoter of *FT1* to activate expression of this key flowering regulator can explain why vernalization is a prerequisite for the acceleration of flowering by long days in temperate cereals. The data presented also support the hypothesized direct regulation of *VRN2* by *VRN1* (refs 9,40). Similarly, binding of *VRN1* to the promoter of *ODDSOC2* (and an additional *ODDSOC2*-like gene; Peak 154) supports a role for this gene in the vernalization response pathway.

More broadly, the direct targets of *VRN1* include genes involved in jasmonic acid, abscisic acid and gibberellin biosynthesis or breakdown (Supplementary Data 2). The regulation of hormone levels by *VRN1* might influence flowering or other aspects of plant development, such as final height. For example, gibberellins accelerate inflorescence development during the long-day flowering response, downstream of *VRN1* (refs 41,42). *VRN1* also binds to the promoters of *CBF* genes that play critical roles in low-temperature induction of freezing tolerance and to *VRS1* (Supplementary Data 2), which regulates spike architecture<sup>43</sup>.



**Figure 6 | A revised model of the molecular network controlling vernalization-induced flowering in cereals.** Prolonged exposure to low temperatures (vernalization) induces transcription of *VRN1* during winter. Then the *VRN1* protein binds to the promoter of *FT1/VRN3* (1). This enhances the expression of *FT1* in long days. *VRN1* also binds to the promoters of *VRN2* (2), blocking the expression of this floral repressor and allowing activation of *FT1* (3). *VRN1* binds to the promoter of *ODDSOC2* and downregulates this gene (4), allowing the increased expression of other floral promoters (5). Binding of *VRN1* to the promoters of *CBF* genes (6) contributes to decreased cold acclimation potential as plants develop towards flowering. Indirect regulation of *VRN1* by *FT1* can further activate the expression of the *VRN1* gene in leaves, in long days (7) and can trigger the floral initiation in some genetic backgrounds, but does not play a primary role in vernalization-induced flowering. Direct interactions are indicated by solid black connectors. Regulatory interactions that could be direct or indirect (status unknown) are indicated by thin blue dashed lines. Indirect feedback regulation of *VRN1* by *FT1* is indicated by a thin red dashed line<sup>56</sup>. Blue upwards arrows indicate increased expression, red arrows facing downwards indicate decreased expression.

Thus, in addition to controlling flowering, *VRN1* directly targets genes in pathways that control other key traits such as frost tolerance.

The expression analyses presented in this study focussed on early stages of reproductive development, when a subset of direct targets of *VRN1* show altered expression in a spring barley transformed with the *VRN1*-HA construct (Supplementary Data 3). These genes include several transcription factors of unknown function that could potentially play roles in floral development. Other genes that show altered expression during early development but are not bound by the *VRN1* protein might be indirectly regulated by increased activity of the *VRN1* gene. For example, the expression of *Barley MADS3*, an *APETALA*-like (*API*) MADS box gene, is possibly activated by increased *FT1* activity, since *FT*-like genes are known to activate the expression of *API*-like MADS box genes that play critical roles during later stages inflorescence development in cereal<sup>44,45</sup>.

The majority of potential binding targets of *VRN1* do not show altered expression at the second leaf stage (ChIP-seq versus RNA-seq data). One potential reason for this is that the fragmented barley genome sequence limits capacity to associate binding sites with potential transcripts. *VRN1*-binding sites are often present on short contigs (<5 kb) with no predicted genes. Many of these binding sites might nevertheless be located near transcribed regions. A more complete picture of the association of *VRN1*-binding sites and differentially expressed transcripts will become apparent as the barley reference genome sequence improves. In addition, some targets of *VRN1* are likely to show altered expression only under specific conditions. This is known to be the case for *CBF* genes targeted by *VRN1*; *HvCBF2*, *HvCBF4* and *HvCBF9* have reduced expression during low-temperature treatment in lines with elevated *VRN1* activity but are not expressed in normal glasshouse conditions<sup>33</sup>. A key objective for future studies will be global analyses of gene expression in specific organs and cell types, combined with matching ChIP-seq analysis of *VRN1* targets.

It is also possible that other factors are required to act with *VRN1* to alter the target gene expression. A second promoter motif, the RY element, was identified in a subset of binding targets. This motif is bound by B3 transcription factors, a large family of proteins that regulate many aspects of plant development and also hormone or stress responses<sup>46</sup>. The binding of *VRN1* and B3 transcription factors to common promoters could further modify the expression of some genes. Combinatorial action with other transcription factors through discrete binding sites could also explain why *VRN1* binding is associated with the repression of some target genes (for example, *VRN2*) but activation of others (for example, *FT1*). In addition to secondary binding motifs in *VRN1* target genes, *VRN1* is likely to interact with other MADS box transcription factors when binding to CArG motifs, as indicated by yeast two-hybrid assays<sup>47,48</sup>. These proteins could modify the impact of *VRN1* on target gene expression via cooperative binding to CArG motifs in target genes.

In conclusion, the identification of direct and indirect targets of *VRN1* provides deeper insights into how this transcription factor regulates the life cycle of cereal crops and influences other key traits. This knowledge of *VRN1* targets can be applied to future crop improvement. For example, mutation of *VRN1*-binding sites can potentially be used to modify the relationship between the vernalization response and other traits such as frost tolerance, spike architecture or plant height.

## Methods

**Plant growth conditions.** Plants were grown in glasshouses at an average temperature of 20°, daily oscillation  $\pm$  4°, sine wave. The day length was 16 h

(16 light/8 dark), consisting of natural light and artificial day length extension. The production of near-isogenic barley lines has been described previously<sup>7</sup>, as has the production of transgenic barley plants carrying the *VRN1-GFP* transgene<sup>12</sup>.

**Expression of the VRN1-HA protein in transgenic barley.** The 3' UTR of *VRN1* was amplified from a bacterial artificial chromosome (DQ249273, bp 16,923–17,220 relative to *VRN1* cv. Strider, AY750993). This fragment (Fragment 1, primers presented in Supplementary Table 2) was placed downstream of six copies of the haemagglutinin epitope sequence (6xHA) and the resulting fragment was then fused to a fragment spanning the distal end of the first intron to the penultimate codon in the eighth exon (Fragment 2, bp 12,190 to 16,911 relative to cv. Strider, DQ249273, primers described in Supplementary Table 2). Finally, the promoter, first exon, plus a small segment of the first intron of *VRN1* (bp 38 to 2,557 relative to cv. Strider) were subcloned upstream of the VRN1-6xHA-UTR segments. Amplification steps were performed with high-fidelity thermostable DNA polymerase (Accuprime Polymerase, Invitrogen). Coding regions were verified by resequencing.

Barley plants were transformed with the *VRN1-HA* transgene using *Agrobacterium* transformation of excised embryos of the variety 'Golden Promise', as described previously<sup>10</sup>. Golden Promise flowers without vernalization (genotype *HvVRN1-1*, *ΔHvVRN2*) and is photoperiod insensitive. To produce a vernalization-requiring 'winter' version of cv. Golden Promise, full-length versions of *VRN1* (that is, no intron deletion) and *VRN2* (that is, locus present), plus a functional copy of *PHOTOPERIOD1*, where introgressed into Golden Promise through three rounds of recurrent crossing with marker assisted selection. The second backcross generation was then crossed to Golden Promise lines carrying the *VRN1-HA* transgene (three rounds of backcrossing in total) and homozygous plants selected from the BC3F2 generation.

**Western blot analysis.** Protein was extracted from barley seedlings (minus roots) at the second leaf stage in homogenization buffer (10% SDS, 10 mM DTT, 20% v/v glycerol, 0.2 M Tris-HCl, 0.2 M PMSF (phenylmethylsulphonyl fluoride) and Protease inhibitor cocktail and then separated by denaturing polyacrylamide electrophoresis (SDS PAGE) and transferred to PVDF transfer membrane (PolyScreen PVDF Hybridization Transfer Membrane, PerkinElmer). VRN1-HA protein was detected using monoclonal anti-HA antibody (H9658, Sigma) in Tris-buffered saline with Tween 20 buffer (20 mM Tris-HCl, 150 mM NaCl and 0.1% v/v Tween 20, pH 7.6) with 0.2% I-block (Tropix, Bedford, MA) at 1/10,000 dilution. Horseradish peroxidase-conjugated goat anti-mouse antibody (PIE31430, Thermo Scientific) was used for secondary detection at 1/100,000 dilution and Supersignal West Dura Substrate was used for signal detection. Raw images of immunoblot results are presented in Supplementary Fig. 10.

**ChIP-Seq and ChIP-PCR.** Chromatin immunoprecipitation was performed as described previously<sup>49</sup> using barley seedlings (minus roots) at the second leaf stage. In brief, barley seedlings were cut into small pieces and crosslinked with 1% formaldehyde for 15 min. Nuclei were isolated and then chromatin was sonicated to shear DNA to a size range of 100–500 bp and immunoprecipitation was performed with monoclonal anti-HA antibody (H9658, Sigma) at a final dilution factor of 1/2,000. Immune complex was collected by protein G Agarose (Millipore 16–266) and DNA was purified by QIAquick PCR Purification kit (Qiagen 28106). Purified DNA samples were used to prepare libraries with the Illumina TruSeq ChIP sample preparation protocol and then sequenced by Illumina HiSeq 2000 with 90 bp single end reads by the Beijing Genomics Institute (BGI).

ChIP-Seq data was analysed with Galaxy<sup>50</sup>. Data quality was checked by FastQC and low-quality reads were trimmed. Then the total reads of *VRN1-HA* (+) and *VRN1-HA* (–) were mapped to Morex whole-genome shotgun sequence assembly<sup>37</sup> using Bowtie2 with default parameters (total reads and mapping percentage are shown in Supplementary Table 1). The mapped reads were applied to Model-based Analysis of ChIP-Seq<sup>34</sup> with default parameters to identify regions enriched in *VRN1-HA* (+) relative to *VRN1-HA* (–) sample and visualized by the Integrative Genomics Viewer (IGV) 2.3 (ref. 51). A cutoff fold enrichment of 20 was used to finalize the peak list. The sequences of the binding region were extracted and applied to motif analysis using the MEME 4.7.0 (ref. 36).

ChIP-PCR was performed using quantitative PCR to assay the degree of enrichment of target genomic DNA by comparing immunoprecipitated chromatin fractions to input chromatin. In all cases, ChIP-PCR comparisons were made between lines carrying the *VRN1-HA* transgene versus sibling null controls (no transgene). Mock immunoprecipitations (no antibody) were also performed as negative controls. Primer sequences are provided in Supplementary Table 2.

**Gene expression.** RNA was extracted using the method of Chang *et al.*<sup>52</sup> Total RNA (5 µg) was reverse transcribed with Super Script III reverse transcriptase (Invitrogen, www.invitrogen.com), according to the manufacturer's instructions. qRT-PCR was performed on a 7900HT Fast Real-Time PCR System (Applied Biosystems, http://www.appliedbiosystems.com) with SYBR green and Platinum DNA polymerase (Invitrogen). The barley *ACTIN* gene was used as reference and relative transcript levels of biosynthesis were calculated with the  $\Delta\Delta C_t$  method,

factoring in primer amplification efficiencies. Primers used are described in Supplementary Table 2.

RNA-seq was performed using three biological replicates of RNA extracted from whole plants (minus roots) at the second leaf stage. RNA was used to generate libraries with the Illumina TruSeq RNA v2 protocol and then sequenced using the Illumina HiSeq platform (100 bp paired end reads) at the Australian Genome Research Facility using standard protocols. The Illumina CASAVA1.8 pipeline was used to generate sequence data. Data quality was checked by FastQC and low-quality reads were trimmed. Sequences were then aligned to the complete CDS contigs from the barley sequencing consortium (<http://www.public.iastate.edu/>) using Biokanga (version 3.4.2). Differential expression analysis was performed using DESeq<sup>53</sup> (version 1.12.1) and edgeR<sup>54</sup> (version 3.2.4, Robinson 2010), with normalization by model-based scaling normalization or Trimmed Mean of M-values (TMM), respectively.

Gene copy number was assayed using the same protocol as qRT-PCR, but with primers targeting the transgene (Supplementary Table 2). The *CO2* gene, described previously as a single-copy reference gene<sup>55</sup>, was used for normalization (Supplementary Table 2).

## References

- Chouard, P. Vernalization and its relations to dormancy. *Annu. Rev. Plant Physiol.* **11**, 191–238 (1960).
- Trevaskis, B. The central role of the *VERNALIZATION1* gene in the vernalization response of cereals. *Funct. Plant Biol.* **37**, 479–487 (2010).
- Danyluk, J. *et al.* TaVRT-1, a putative transcription factor associated with vegetative to reproductive transition in cereals. *Plant Physiol.* **132**, 1849–1860 (2003).
- Trevaskis, B., Bagnall, D. J., Ellis, M. H., Peacock, W. J. & Dennis, E. S. MADS box genes control vernalization-induced flowering in cereals. *Proc. Natl Acad. Sci. USA* **100**, 13099–13104 (2003).
- Yan, L. *et al.* Positional cloning of the wheat vernalization gene *VRN1*. *Proc. Natl Acad. Sci. USA* **100**, 6263–6268 (2003).
- Murai, K., Miyamae, M., Kato, H., Takumi, S. & Ogihara, Y. *WAP1*, a wheat *APETALA1* homolog, plays a central role in the phase transition from vegetative to reproductive growth. *Plant Cell Physiol.* **44**, 1255–1265 (2003).
- Oliver, S. N., Deng, W., Casao, M. C. & Trevaskis, B. Low temperatures induce rapid changes in chromatin state and transcript levels of the cereal *VERNALIZATION1* gene. *J. Exp. Bot.* **64**, 2413–2422 (2013).
- Sasani, S. *et al.* The influence of vernalization and daylength on expression of flowering-time genes in the shoot apex and leaves of barley (*Hordeum vulgare*). *J. Exp. Bot.* **60**, 2169–2178 (2009).
- Trevaskis, B., Hemming, M. N., Peacock, W. J. & Dennis, E. S. *HvVRN2* responds to daylength, whereas *HvVRN1* is regulated by vernalization and developmental status. *Plant Physiol.* **140**, 1397–1405 (2006).
- Hemming, M. N., Peacock, W. J., Dennis, E. S. & Trevaskis, B. Low-temperature and daylength cues are integrated to regulate *FLOWERING LOCUS T* in barley. *Plant Physiol.* **147**, 355–366 (2008).
- Preston, J. C. & Kellogg, E. A. Discrete developmental roles for temperate cereal grass *VERNALIZATION1/FRUITFULL*-like genes in flowering competency and the transition to flowering. *Plant Physiol.* **146**, 265–276 (2008).
- Alonso-Peral, M. M., Oliver, S. N., Casao, M. C., Greenup, A. A. & Trevaskis, B. The promoter of the cereal *VERNALIZATION1* gene is sufficient for transcriptional induction by prolonged cold. *PLoS ONE* **6**, e29456 (2011).
- Fu, D. *et al.* Large deletions within the first intron in *VRN-1* are associated with spring growth habit in barley and wheat. *Mol. Genet. Genomics* **273**, 54–65 (2005).
- Oliver, S. N., Finnegan, E. J., Dennis, E. S., Peacock, W. J. & Trevaskis, B. Vernalization-induced flowering in cereals is associated with changes in histone methylation at the *VERNALIZATION1* gene. *Proc. Natl Acad. Sci. USA* **106**, 8386–8391 (2009).
- Xiao, J. *et al.* O-GlcNAc-mediated interaction between VER2 and TaGRP2 elicits TaVRN1 mRNA accumulation during vernalization in winter wheat. *Nat. Commun.* **5**, 4572 (2014).
- von Zitzewitz, J. *et al.* Molecular and structural characterization of barley vernalization genes. *Plant Mol. Biol.* **59**, 449–467 (2005).
- Cockram, J. *et al.* Haplotype analysis of vernalization loci in European barley germplasm reveals novel *VRN-H1* alleles and a predominant winter *VRN-H1/VRN-H2* multi-locus haplotype. *Theor. Appl. Genet.* **115**, 993–1001 (2007).
- Szucs, P. *et al.* Validation of the *VRN-H2/VRN-H1* epistatic model in barley reveals that intron length variation in *VRN-H1* may account for a continuum of vernalization sensitivity. *Mol. Genet. Genomics* **277**, 249–261 (2007).
- Hemming, M. N., Fieg, S., Peacock, W. J., Dennis, E. S. & Trevaskis, B. Regions associated with repression of the barley (*Hordeum vulgare*) *VERNALIZATION1* gene are not required for cold induction. *Mol. Genet. Genomics* **282**, 107–117 (2009).
- Takahashi, R. & Yasuda, S. in *Barley Genetics II* (ed. Nilan, R. A.) 388–408 (Washington State University Press, 1971).



21. Hayes, P. M. *et al.* Quantitative trait loci on barley (*Hordeum vulgare* L.) chromosome 7 associated with components of winterhardiness. *Genome* **36**, 66–71 (1993).
22. Koemel, J. E., Guenzi, A. C., Anderson, J. A. & Smith, E. L. Cold hardiness of wheat near-isogenic lines differing in vernalization alleles. *Theor. Appl. Genet.* **109**, 839–846.
23. Yan, L. *et al.* The wheat *VRN2* gene is a flowering repressor down-regulated by vernalization. *Science* **303**, 1640–1644 (2004).
24. Dubcovsky, J., Chen, C. & Yan, L. Molecular characterization of the allelic variation at the *VRN-H2* vernalization locus in barley. *Mol. Breeding* **15**, 395–407 (2005).
25. Loukoianov, A., Yan, L., Blechl, A., Sanchez, A. & Dubcovsky, J. Regulation of *VRN-1* vernalization genes in normal and transgenic polyploid wheat. *Plant Physiol.* **138**, 2364–2373 (2005).
26. Yan, L. *et al.* The wheat and barley vernalization gene *VRN3* is an orthologue of *FT*. *Proc. Natl Acad. Sci. USA* **103**, 19581–19586 (2006).
27. Corbesier, L. *et al.* FT protein movement contributes to long-distance signaling in floral induction of *Arabidopsis*. *Science* **316**, 1030–1033 (2007).
28. Turner, A., Beales, J., Faure, S., Dunford, R. P. & Laurie, D. A. The pseudo-response regulator *Ppd-H1* provides adaptation to photoperiod in barley. *Science* **310**, 1031–1034 (2005).
29. Greenup, A. G. *et al.* *ODDSOC2* is a MADS box floral repressor that is down-regulated by vernalization in temperate cereals. *Plant Physiol.* **153**, 1062–1073 (2010).
30. Greenup, A. G. *et al.* Transcriptome analysis of the vernalization response in barley (*Hordeum vulgare*) seedlings. *PLoS ONE* **6**, e17900 (2011).
31. Thomashow, M. F. Plant cold acclimation: freezing tolerance genes and regulatory mechanisms. *Annu. Rev. Plant Physiol. Plant Mol. Biol.* **50**, 571–599 (1999).
32. Stockinger, E. J., Skinner, J. S., Gardner, K. G., Francia, E. & Pecchioni, N. Expression levels of barley *Cbf* genes at the *Frost resistance-H2* locus are dependent upon alleles at *Fr-H1* and *Fr-H2*. *Plant J.* **51**, 308–321 (2007).
33. Dhillon, T. *et al.* Regulation of freezing tolerance and flowering in temperate cereals: the *VRN-1* connection. *Plant Physiol.* **153**, 1846–1858 (2010).
34. Zhang, Y. *et al.* Model-based analysis of ChIP-Seq (MACS). *Genome Biol.* **9**, R137 (2008).
35. Houston, K. *et al.* Variation in the interaction between alleles of *HvAPETALA2* and microRNA172 determines the density of grains on the barley inflorescence. *Proc. Natl Acad. Sci. USA* **110**, 16675–16680 (2013).
36. Bailey, T. L. & Elkan, C. Fitting a mixture model by expectation maximization to discover motifs in biopolymers. *Proc. Int. Conf. Intell. Syst. Mol. Biol.* **2**, 28–36 (1994).
37. Mayer, K. F. *et al.* A physical, genetic and functional sequence assembly of the barley genome. *Nature* **491**, 711–716 (2012).
38. Hayama, R., Yokoi, S., Tamaki, S., Yano, M. & Shimamoto, K. Adaptation of photoperiodic control pathways produces short-day flowering in rice. *Nature* **422**, 719–722 (2003).
39. Lee, S., Kim, J., Han, J. J., Han, M. J. & An, G. Functional analyses of the flowering time gene *OsMADS50*, the putative *SUPPRESSOR OF OVEREXPRESSION OF CO 1/AGAMOUS-LIKE 20 (SOC1/AGL20)* ortholog in rice. *Plant J.* **38**, 754–764 (2004).
40. Trevaskis, B., Hemming, M. N., Dennis, E. S. & Peacock, W. J. The molecular basis of vernalization-induced flowering in cereals. *Trends Plant Sci.* **12**, 352–357 (2007).
41. Pearce, S., Vanzetti, L. & Dubcovsky, J. Exogenous gibberellins induces wheat spike development under short days only in the presence of *VERNALIZATION1*. *Plant Physiol.* **163**, 1433–1445 (2013).
42. Boden, S. A. *et al.* *EARLY FLOWERING3* regulates flowering in spring barley by mediating gibberellin production and *FLOWERING LOCUS T* expression. *Plant Cell* **29**, 1557–1569 (2014).
43. Komatsuda, T. *et al.* Six-rowed barley originated from a mutation in a homeodomain-leucine zipper I-class homeobox gene. *Proc. Natl Acad. Sci. USA* **104**, 1424–1429 (2007).
44. Teper-Bamnlolker, P. & Samach, A. The flowering integrator *FT* regulates *SEPALLATA3* and *FRUITFULL* accumulation in *Arabidopsis* leaves. *Plant Cell* **17**, 2661–2675 (2005).
45. Kobayashi, K. *et al.* Inflorescence meristem identity in rice is specified by overlapping functions of three *API/FUL*-like MADS box genes and *PAP2*, a *SEPALLATA* MADS box gene. *Plant Cell* **24**, 1848–1859 (2012).
46. Peng, F. Y. & Weselake, R. J. Genome-wide identification and analysis of the B3 superfamily of transcription factors in Brassicaceae and major crop plants. *Theor. Appl. Genet.* **126**, 1305–1319 (2013).
47. Tardif, G. *et al.* Interaction network of proteins associated with abiotic stress response and development in wheat. *Plant Mol. Biol.* **63**, 703–718 (2007).
48. Cao, S. & Yan, L. Construction of a high-quality yeast two-hybrid (Y2H) library and its application in identification of interacting proteins with key vernalization regulator TaVRN-A1 in wheat. *BMC Res. Notes* **6**, 81 (2013).
49. Deng, W. *et al.* FLOWERING LOCUS C (FLC) regulates development pathways throughout the life cycle of *Arabidopsis*. *Proc. Natl Acad. Sci. USA* **108**, 6680–6685 (2011).
50. Giardine, B. *et al.* Galaxy: a platform for interactive large-scale genome analysis. *Genome Res.* **15**, 1451–1455 (2005).
51. Robinson, J. T. *et al.* Integrative genomics viewer. *Nat. Biotechnol.* **29**, 24–26 (2011).
52. Chang, S., Puryear, J. & Cairney, J. A simple and efficient method for isolating RNA from pine trees. *Plant Mol. Biol. Rep.* **11**, 113–116 (1993).
53. Anders, S. & Huber, W. Differential expression analysis for sequence count data. *Genome Biol.* **11**, R106 (2010).
54. Robinson, M. D., McCarthy, D. J. & Smyth, G. K. edgeR: a Bioconductor package for differential expression analysis of digital expression data. *Bioinformatics* **26**, 139–140 (2009).
55. Bartlett, J. G., Alves, S. C., Smedley, M., Snape, J. W. & Harwood, W. A. High-throughput *Agrobacterium*-mediated barley transformation. *Plant Methods* **4**, 22 (2007).
56. Li, C. & Dubcovsky, J. Wheat FT protein regulates *VRN1* transcription through interactions with FDL2. *Plant J.* **55**, 543–554 (2008).

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### Author contributions

W.D. and B.T. designed the research. W.D., M.C.C. and B.T. performed the experiments. P.W. analysed the RNA-Seq data sets. P.M.H. and K.S. provided sequence information for target genes. W.D., E.J.F. and B.T. prepared the manuscript.

### Additional information

**Accession codes:** ChIP-seq data has been submitted to the NCBI Sequence Read Archive (SRP041049). RNA-seq data was submitted to the NCBI Sequence Read Archive (SRP045121).

**Supplementary Information** accompanies this paper at <http://www.nature.com/naturecommunications>

**Competing financial interests:** The authors declare no competing financial interests.

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