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Serum protein biomarkers for HCC risk prediction in HIV/HBV co-infected people: a clinical proteomic study using mass spectrometry

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Background: HBV coinfection is frequent in people living with HIV (PLWH) and is the leading cause of hepatocellular carcinoma (HCC). While risk prediction methods for HCC in patients with HBV monoinfection have been proposed, suitable biomarkers for early diagnosis of HCC in PLWH remain uncommon.

Methods: Liquid chromatography-tandem mass spectrometry (LC-MS/MS) was used to examine serum protein alterations in HCC and non-HCC patients with HIV and HBV co-infection. Gene Ontology (GO), Kyoto Encyclopedia of Genes and Genomes (KEGG), and Disease Ontology (DO) enrichment analysis were performed on the differentially expressed proteins (DEPs). The risk prediction model was created using five-cross-validation and LASSO regression to filter core DEPs.

Results: A total of 124 DEPs were discovered, with 95 proteins up-regulated and 29 proteins down-regulated. Extracellular matrix organization and membrane component were the DEPs that were most abundant in the categories of biological processes (BP) and cellular components (CC). Proteoglycans in cancer were one of the top three DEPs primarily enriched in the KEGG pathway, and 60.0% of DEPs were linked to various neoplasms in terms of DO enrichment. Eleven proteins, including GAPR1, PLTP, CLASP2, IGHV1-69D, IGLV5-45, A2M, VNN1, KLK11, ANPEP, DPP4 and HYI, were chosen as the core DEPs, and a nomogram was created to predict HCC risk.

Conclusion: In HIV/HBV patients with HCC, several differential proteins can be detected in plasma by mass spectrometry, which can be used as screening markers for early diagnosis and risk prediction of HCC. Monitoring protease expression differences can help in the diagnosis and prognosis of HCC.

KEYWORDS

hepatocellular carcinoma, human immunodeficiency virus, hepatitis B, golgi-associated plant pathogenesis-related protein 1, phospholipid transfer protein

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Hepatocellular carcinoma (HCC) is the sixth most common cancer and the second leading cause of cancer-related deaths worldwide, killing over 700,000 people each year (1, 2). In contrast to most solid tumors, the prevalence of HCC and HCCrelated mortality has risen in recent decades. Chronic hepatitis B virus (HBV) and chronic hepatitis C virus (HCV) infection are the most important causes of HCC. Overall, HCC occurs lifelong in 15% to 40% of chronic HBsAg carriers in the absence of antiviral treatment (3). Human immunodeficiency virus (HIV) is typically spread by sexual, parenteral, and neonatal transmission, which explains why HIV-HBV co-infection is relatively common. HBV infection among HIV-infected patients is estimated to be 5% to 15% (3, 4). Thus, HBV co-infection affects 3 million of the 35 million people living with HIV worldwide (3, 4). When compared to HBVmonoinfected individuals, HIV-HBV co-infected patients have a lower chance of spontaneous HBeAg and HBsAg clearance, and serum HBV-DNA levels are higher, which may contribute to a faster progression to end-stage liver disease and HCC, which is characteristic of HIV-HBV co-infected patients (5-7). As a result, HCC continues to be one of the leading causes of non-AIDS deaths among people living with HIV (PLWH) (8).

The traditional marker used for HCC risk monitoring is AFP (alpha-fetoprotein), although it has largely failed in the early diagnosis of HCC (9). Other alternative biomarkers have been proposed, such as AFP-L3 (Lens culinaris agglutinin-reactive fraction of alphafetoprotein or highly sensitive -hs-AFP-L3) (10) and PIVKA-II (Protein induced by vitamin K absence or antagonist-II) (11), but the levels of these two proteins typically increase as HCC develops and progresses to portal vein invasion (11). Other indicators, such as the Mac-2-binding protein glycosylation isomer (M2BPGi), have been used to predict hepatocarcinogenesis in patients receiving nucleoside analogs (12, 13). Some indexes, such as aMAP (14), PAGE-B index (15, 16), and GALAD score (17), developed from different statistical models, demonstrated superior specificity and predictive value for HCC evaluation and monitoring. Several novel predictive biomarkers for HCC, such as methylation status of HOXA1, TSPLY5, and PFKP from cell-free DNA (18, 19), and Telomerase Reverse Transcriptase (TERT) promoter mutations (C228T and C250T) (20), are frequently detected in HCC and have a good predictive value for hepatic oncogenesis. None of these biomarkers or scores, however, have been studied in HIV-HBV co-infected patients. Appropriate biomarkers for the early identification of HCC in patients with HIV and HBV co-infection are still lacking. There is a practical need to create non-invasive, cost-effective, and easy serological diagnostic markers for HCC early detection, particularly in those living with HIV and HBV co-infection in resource-limited settings.

Mass spectrometry (MS)-based proteomics is useful for identifying and measuring protein post-translational changes. The utility of this technique has improved dramatically in recent years. Many research have used mass spectrometry proteomics to screen and identify serum potential biomarkers for illness diagnosis and prognosis evaluation in a variety of diseases, which has played a significant role in promoting clinical practice. However, proteomics research in PLWH is currently limited. This study focused on HIV/HBV co-infected individuals who were separated into HCC and non-HCC groups based on their clinical diagnoses. First, we tried to screen out prospective biomarkers in the assessment of HCC by detecting the expression of differentially expressed proteins (DEPs) related to HCC. Then, we intend to provide new clues for the in-depth investigation of HCC pathogenesis from the standpoint of proteomics by doing a functional enrichment analysis of DEPs between HCC patients and non-HCC patients. Furthermore, we attempted to assess the probability of HCC development in individuals at high risk of developing end-stage liver disease by establishing an HCC risk prediction model.

Methods

Participants

From 578 HIV/HBV co-infected patients, 13 pairs of HCC patients and non-HCC patients with matched information were chosen for the proteomic investigation. Figure 1 depicts the flow chart of study object selection. The following were the inclusion criteria: (1) be 30-70 years old; (2) have complete HIV clinical data, including ART status and current CD4+ T lymphocyte counts; (3) have complete HBV clinical data, including hepatitis B indicators, HBV DNA levels, and liver imaging evaluation findings; and (4) have an HCC definition or exclusion diagnosis. Exclusion criteria included the following: (1) being pregnant or nursing; (2) an uncontrolled underlying condition, (3) organ dysfunction, (4)



liver illnesses caused by other hepatitis virus infections, (5) any cancer other than HCC, and (6) diseases other than viral hepatitis that may cause HCC, such as metabolic problems, substance abuse, and genetic conditions.

For proteomic analysis, age, gender, clinical routine test indexes, CD4+ T lymphocyte counts, HBeAg status, liver function findings, duration of HBV infection, HIV and HBV replication levels, and other known characteristics influencing HCC progression had to be well matched. Following the fulfillment of all of the parameters for proteomic analysis, 26 patients with HIV and HBV coinfection who were successfully matched were divided into two groups for future proteome research: HCC (n=13) and non-HCC (n=13). The HCC patients were all in stage I.

This study was approved by the institutional review board of the Zhongnan Hospital of Wuhan University(2021022). An informed consent form was signed by all patients.

Plasma sample preparation

Blood samples were collected from patients living with HIV and HBV co-infection between March 1, 2021 and December 31, 2022. For one-step reduction and alkylation, the samples were treated for 1 hour at 60°C with 1% SDC/100 mM Tris-HCL/10 mM TCEP/40 mM CAA (pH=8.5). SDC was diluted to less than 0.5% using ddH2O. Trypsin was added at a 1:50 (enzyme:protein, w/w) ratio for overnight digestion at 37°C. The following day, TFA was added to drop the pH to 6.0 and end the digestion. The supernatant was centrifuged for 15 minutes at 16000g before peptide purification using a self-made SDB desalting column. The peptide eluate was vacuum dried and stored at -20°C for later use.

LC-MS/MS analysis

Tryptophan was dissolved in solvent A, which contained 0.1% formic acid. After that, the reverse-phase column is installed. The progressive gradient is constructed of solvent B containing 0.1% formic acid. Using the EASY-nLC system, the ascending gradient is filled with a mixture of formic acid concentrations ranging from 6% to 23%, and the flow rate is 400nL/min. The sample is subjected to a series of MS/MS assays on Thermo Scientific's QExactiveTM Plus platform after the initial nano spray ionization process. The voltage of the electrospray is set to 2.0 kV. It is utilized for peptide activation and has a resolution of 70,000 to 1800. Make use of NCE Settings For MS/ MS testing, peptides should be used. Orbitrap was then used to carry out a resolution. Data-dependent apps are executed dynamically on different scans. The fixed mass was then set to 100 m/z.

Database search

Briefly, the raw MS data were analyzed with the DIA-NN program (V1.7.16). Tandem MS data were thoroughly referred to

the SwissProt human protein sequences database. The deep learning algorithm in DIA-NN predicted a spectrum library, and the original DIA data were retrieved by the anticipated spectrum library and the spectrum library acquired by MBR function to obtain protein quantitative information. As a consequence, the parent ion and protein levels were tested using 1% FDR. Following screening, the quantitative information from the proteome was used for further analysis.

Biological information analysis

The function databases employed in this work were Gene Ontology (www.geneontology.org) (GO), Kyoto Encyclopedia of Genes and Genomes (http://www.genome.jp/kegg/) (KEGG), and Disease Ontology (https://disease-ontology.org/) (DO). Annotation information for GO, KEGG, and DO was collected by mapping identified proteins to matching databases downloaded from the official website. Then, using CytoScape software and the "CytoHubba" plug-in, network maps of DEPs were created using protein-protein interaction (PPI) data from the STRING database, and the hub proteins were further filtered.

Statistical analysis

In the descriptive statistical analysis of the study subjects' demographic and clinical features, means and standard deviations were utilized for continuous variables, while proportions or rates were provided for categorical data. The fold change and P value less than 0.05 were chosen as the key screening criteria for biomarkers. The penalty term was chosen using five-fold cross-validation, and the parameter for LASSO regression analysis with the smallest model fitting error was utilized to filter the variables. The model was tested in three ways: differentiation using ROC area AUC, calibration and consistency using calibration curve, and clinical effect using DCA clinical decision curve analysis or nomogram. The R programming language was utilized for statistical analysis and model development.

Results

Demographic and clinical characteristics

Table 1 shows the demographic and clinical features of the patients with HIV and HBV co-infection who participated in this investigation. In terms of age, gender, fraction of cART treatment, and anti-HBV therapy, no significant differences were found between the HCC and non-HCC groups. None of the clinical indicators, including HIV-RNA and HBV-DNA levels, HBeAg status, blood routine test results, and liver function test findings, were substantially different between the HCC and non-HCC groups.

	People living with HIV/HBV co-infection		Statistical value	P-value
	HCC group (n=13)	Non-HCC group (n=13)		
Age, mean ± SD, years	48.7 ± 10.8	48.8 ± 12.7	0.017	0.987
Gender (Male), n(%)	13(100.0)	13(100.0)	/	/
cART treatment (n,%)	6(46.2)	2(15.4)	1.625	0.202
Anti-HBV therapy (n,%)	6(46.2)	2(15.4)	1.625	0.202
CD4 ⁺ T cell count (cells/µl)	129 ± 33	147 ± 64	0.222	0.827
Log ₁₀ HIV-RNA (mean ± SD)	4.55 ± 0.33	4.61 ± 0.24	0.102	0.919
HBeAg (n,%)	1(7.7)	2(15.4)	0.377	0.539
Log ₁₀ HBV-DNA (mean ± SD)	3.15 ± 0.46	3.24 ± 0.32	0.550	0.587
Blood routine test (mean <u>+</u> SD)				
White blood cells (10 ⁹ /L)	4.08 ± 0.22	4.22 ± 0.42	0.506	0.617
Hemoglobin (g/L)	110.8 ± 10.5	123.5 ± 15.4	0.726	0.475
Platelets (10 ⁹ /L)	120.5 ± 15.8	127.7 ± 20.6	0.862	0.397
Liver function test (mean <u>+</u> SD)				
ALT (U/L)	38.6 ± 4.2	40.5 ± 3.7	1.025	0.315
AST (U/L)	38.0 ± 5.0	41.2 ± 6.1	1.002	0.326
Tbil (µmol/L)	11.8 ± 1.9	10.5 ± 2.3	0.908	0.373
Alb(g/L)	38.6 ± 2.8	43.2 ± 3.0	1.321	0.198

TABLE 1 Demographic and clinical characteristics of people living with HIV and HBV co-infection in this study.

SD, standard deviation; cART, Combination antiretroviral therapy; ALT, alanine aminotransferase; AST, aspartate aminotransferase; Tbil, total bilirubin; Alb, albumin.

Protein identification

Using a data-independent acquisition (DIA) technique, plasma proteome profiles were obtained from a total of 26 patients living with HIV and HBV co-infection. Furthermore, 26 proteomes were examined in the entire cohort using a DIA technique, yielding a total of 653 proteins and 8200 precursors (Figures 2A, B). 124 proteins were expressed significantly differently in HCC patients than in non-HCC patients. There were 95 proteins that were up-regulated and 29 proteins that were down-regulated (Figure 2C). The PCA plot showing the intensities of the 124 differential proteins demonstrated a strong distinction of HCC patients and non-HCC patients (Figure 2D). The expression of differential proteins (DEPs) was used to create a heatmap (Figure 2E), demonstrating that DEP expression was significantly different between the HCC group and the control group.

Functional enrichment analysis of differentially expressed proteins

Figure 3A depicts the findings of GO keywords with statistical differences in enrichment analysis hypergeometric tests. Three of the five different GO keywords assigned to biological processes were related to extracellular matrix organization. In terms of cellular component, all five enriched terms were membrane components. The top four enriched terms in terms of molecular function were signaling receptor activity,

molecular transducer activity, transmembrane signaling receptor activity, and metallopeptidase activity. Proteoglycans were one of the top three DEPs in cancer that were primarily enriched in the KEGG pathway (Figure 3B). In terms of DO enrichment analysis, 60.0% of the top 15 enriched terms (shown in Figure 3C) were related to various neoplasms.

Protein-protein interaction network analysis of DEPs

Network maps of DEPs with protein–protein interaction (PPI) data from the STRING database were established (Figure 4A). The top 10 proteins in four algorithms (DEGREE, DMNC, MCC and MNC) were selected as hub proteins, respectively (Figures 4B–E). Moreover, by cross-analyzing and mapping to Venn diagram, CD44 antigen(CD44), intercellular adhesion molecule 1(ICAM1), transferrin receptor protein 1(TFRC), neural cell adhesion molecule 1(NCAM1), apolipoprotein A-IV(APOA4), haptoglobin (HP), low affinity immunoglobulin gamma Fc region receptor II-a (FCGR2A) were identified as hub proteins (Figure 4F).

LASSO regression model

Given the relatively small sample size, the LASSO model was developed to screen the 124 DEPs for core proteins that are closely



connected to HCC diagnosis in patients living with HIV and HBV co-infection. Finally, 11 proteins were chosen for additional logistic regression analysis (Figure 5). They were golgi-associated plant pathogenesis-related protein 1 (GAPR1), phospholipid transfer protein (PLTP), cytoplasmic junction associated protein 2 (CLASP2), immunoglobulin heavy variable 1-69D (IGHV1-69D), immunoglobulin lambda variable 5-45 (IGLV5-45), alpha-2-macroglobulin (A2M), pantetheinase (VNN1), kallikrein-11 (KLK11), aminopeptidase N (ANPEP), dipeptidyl peptidase 4 (DPP4) and putative hydroxypyruvate isomerase (HYI), and the last five were protease members.

Model validation

Based on the 11 core proteins screened by the previous part of the LASSO regression model, R programming language was performed for

five-cross-validation following the principle of random combination of less than 5 proteins, and the optimal AUC value was used to construct the clinical diagnostic model. Finally, GAPR1 and IGHV1-69D were chosen as the best DEPs combination, and a clinical risk model was built. The model was tested, and the discrimination was satisfactory (Figure 6A). The AUC (95% CI) for the train cohort was 0.96 (0.89-0.96) and 0.89 (0.58-0.89) for the test cohort. Furthermore, the findings of these feature combinations' confusion matrices demonstrated great accuracy for identifying distinct groups (Figure 6B). The calibration curve demonstrated that the model performed well when compared to an ideal model (Figure 6C). The decision curve analysis revealed favorable results (Figure 6D). Using the model data, a nomogram was created (Figure 6E).

The sensitivity and specificity of this clinical risk model were 76.9%(10/13) and 92.0%(115/125), respectively. The positive and negative prediction values were 71.4%(10/14) and 97.6%(121/124), respectively.



FIGURE 3

GO, KEGG and DO analysis for 124 DEPs. (A) GO-based enrichment analysis of DEPs(two-sided hypergeometric test, $p \le 0.05$), GO terms were sorted by *p*-value, and top 5 terms of each category were displayed. (B) KEGG-based enrichment analysis of DEPs(two-sided hypergeometric test, $p \le 0.05$), KEGG terms were sorted by *p*-value, and top 3 terms were displayed. (C) DO-based enrichment analysis of DEPs(two-sided hypergeometric test, $p \le 0.05$), DO terms were sorted by *p*-value, and top 15 terms were displayed.

Discussion

In HIV/HBV co-infected patients, the incidence rate of hepatocellular carcinoma can reach 2/1000 person-years (21, 22). The tumor microenvironment alters as a result of HCC. According to the findings of this study, 60% of the differential proteins enriched in DO entries are connected with distinct malignancies, implying that different types of tumors have varied pathophysiology but frequently have similar microenvironment modifications. Extracellular matrix (ECM) is a critical component of the tumor

microenvironment. Extracellular proteases directly govern and change the tumor microenvironment, as well as indirectly stimulating tumor-cell proliferation, apoptosis, angiogenesis, invasion, and metastasis. In this study, it was found that extracellular matrix organization and membrane component were significantly enriched in GO entries when comparing the differential proteins in the serum of HCC and non-HCC patients, implying that the occurrence and development of HCC can be accompanied by changes in the composition of basement membrane. LG2m, a basement membrane component that is not



PPI network analysis of DEPs. (A) Network graph depicting the correlation of proteins derived from DEPs. (B) The hub proteins calculated under DEGREE. (C) The hub proteins calculated under DMNC. (D) The hub proteins calculated under MCC. (E) The hub proteins calculated under MNC. (F) Venn diagram based on cross-analysis under four algorithms (DEGREE, DMNC, MCC and MNC). Circles indicated the gene symbol of protein, in which red and green indicated up-regulated and down-regulated proteins, respectively. The thickness of the lines represented the strength of the interaction. The darkness of the colors represented the magnitude of the discrepancy.

found in normal tissues, has been shown to be a predictive biomarker for the development of metastatic HCC (23). Elevated serum LG2m is associated with an increased risk of HCC in CHC patients who have achieved sustained virological response. The structural dynamics of basement membrane components in the process of hepatocarcinogenesis remain unknown, and more research is required.

Proteoglycan is an important component of the extracellular matrix, which is linked to the development of atherosclerosis as well as the appearance and spread of malignancies. In this study, we discovered that proteoglycans in cancer are enriched in the KEGG pathway by analyzing biological information from HCC and non-HCC patients. It was discovered that aberrant proteoglycan expression has a significant impact in the formation and progression of HCC.

Impaired hepatic autophagy has been demonstrated to contribute to the pathogenesis of AALD, NAFLD, as well as HBV and HCV hepatitis (24–29), suggesting that deficient autophagy

may be a common mechanism for HCC development. Autophagy may also have a paradoxical role in liver cancer, preventing early tumor development while boosting the growth and aggressiveness of already developed liver tumors (30). In mammalian cells, Golgiassociated plant pathogenesis related protein 1 (GAPR-1) can interact with Beclin 1, a major component of the autophagosome nucleation complex, as a negative regulator of autophagy (31). Although the molecular mechanism of this relationship is unknown, GAPR-1 was identified in this investigation as a novel serum biomarker for predicting HCC. The outcome is quite crucial. It was proposed that inhibiting autophagy in combination with other chemotherapeutic medications is a promising therapeutic strategy for HCC.

GAPR-1 interacts with the autophagosome nucleation complex to adversely control autophagy. According to one study, the lipid content influences GAPR-1 binding to biological membranes (32). In this study, phospholipid transfer protein (PLTP) was identified as one of the main DEPs in HCC patients. Through its active form,



FIGURE 5

Dotted vertical lines were drawn at the optimal values by using the minimum criteria(min criteria) and the 1 standard error of the minimum criteria (1se criteria). λ value was chosen when the fitting error was minimum.

PLTP performs lipid transfer. The shuttling and redistribution of lipids between lipoproteins, which are ordinarily returned to the liver for metabolism via the reverse cholesterol transport pathway, is thought to be necessary for PLTP-mediated lipid transfer to highdensity lipoprotein (HDL). Elevated serum PLTP levels in cancer patients following treatment with topoisomerase I inhibitors may serve as a biomarker for tracking the progression of hypertriglyceridemia (33). We hypothesize that dyslipidemia, via the autophagy effect mediated by GAPR-1, may play a regulatory role in the etiology of HCC.

Furthermore, PLTP has been linked to the diagnostic and anticancer actions of specific malignancies. PLTP expression was found to be an appropriate marker for the course and prognosis of gastric cancer (GC) (34). Another study discovered that removing PLTP from U251 cells reduced the expression of epithelial mesenchymal transition markers and invasion molecules matrix metalloproteinases (35). As a result, PLTP was identified as a novel glioma-associated protein that may be involved in the progression of human glioma. In terms of anti-cancer properties, one study found that PLTP is essential in mediating the association of triacyl lipid A with lipoproteins, resulting in an extension of its residence time and magnification of its proinflammatory and anticancer properties (36). Furthermore, PLTP has been identified as a p53 target gene, and forced expression of PLTP reduces colony formation in human tumor cell lines (37). In this study, PLTP was found to be one of the core DEPs in patients with HCC and non-HCC in an HIV/HBV co-infected population, implying that PLTP can be used as a candidate protein to aid in the diagnosis of HCC in this population, and is expected to provide certain clues for the investigation of the mechanism of targeted therapy for HCC.

Five of the 11 key differential proteins identified by the LASSO model were protease members. Vanin-1 (VNN1) was one of the proteases involved. In a prior clinical microarray research, it was first described as a unique clinically derived biomarker of pancreatic cancer-associated new-onset diabetes (38). Another study found that pancreatic cancer cells overexpressing VNN1 aggravate paraneoplastic islet dysfunction by causing oxidative stress and βcell dedifferentiation (39). Pantetheinase VNN1 was discovered to be expressed differentially in HCC and non-HCC individuals, providing evidence to support the function of oxidative stress in HCC.

Kallikrein-11 (KLK11) was the second protease. The KLK gene family is the largest protease group in the human genome. KLK11 was first isolated from the human hippocampus and was found to cleave fewer substrates than other types of proteases, indicating a high degree of substrate specificity (40). KLK11 is thought to be a prostate, ovarian, and breast cancer biomarker (41-43). According to one study, KLK11 mRNA expression is a unique and independent biomarker in laryngeal cancer that can be used for diagnostic and prognostic purposes (44). Another study found that Kallikrein 11 was one of the contributing factors implicated in the rise of CAF-mediated oral squamous cell carcinoma invasion (45). According to one study, changes in KLK11 expression levels can be utilized as additional data to predict clinical outcome and prognosis in breast cancer, gastric cancer (46), and low rectal cancer (47). A Kaplan-Meier survival curve study demonstrated that elevated KLK11 expression was linked to a poor prognosis of cholangiocarcinoma (48). Stronger Kallikrein-11 expression appears to be associated with improved survival rates in nonsmall cell lung cancer (49). Although the preceding research established that KLK11 is associated with tumor diagnosis and prognosis, no studies have reported a link between KLK11 and HCC. The findings of this study help to expand the database of KLK11 and tumor associations.

Aminopeptidase N (APN, commonly known as CD13) was the third protease. It is a transmembrane protein that is widely expressed. APN was commonly elevated in HCC tumor tissues and cell lines with strong metastatic potential (50). It has a distinct canalicular pattern when expressed in normal and neoplastic liver tissue (CD13(can)). CD13(can) has been shown to be more sensitive than AFP in distinguishing between HCC and non-HCC (51). APN is engaged in numerous sorts of tumor cellular processes and is a possible anti-cancer therapeutic target. According to the findings of one study, APN causes BCKDKS31 phosphorylation and



(D) Decision curve analysis curve. (E) Nomogram for predicting the risk of liver cirrhosis.

activates its downstream pathway to enhance HCC proliferation and metastasis (50). The new APN inhibitors 4cc and 24F were shown to be anti-cancer agents for HCC by activating mitochondrial apoptosis pathways and inhibiting HCC cell invasion and angiogenesis (52, 53). Furthermore, CD13 is likely to be a promising target for reducing HCC resistance (54). These findings imply that aminopeptidase N (APN)/CD13 is linked to HCC, and our work provides information on the difference in serum APN expression between HCC patients and non-HCC patients in an HIV/HBV co-infected community. It is proposed that APN/CD13 may be a candidate noninvasive serological marker to aid in the diagnosis of HCC.

Dipeptidyl peptidase 4 (DPP4) was the fourth protease. It has pro-apoptotic effects as a regulatory protease. It has been discovered to be improperly regulated in patients with HCC, similar to the findings in this investigation (55–57). As a result, its therapeutic importance is worth summarizing and delving into. According to one study, the DPP4 gene family's mRNA expression is

substantially increased in human HCC and is related with poor survival in HCC patients (58). As a result, targeting the DPP4 enzyme family could be a unique and effective strategy for promoting anti-tumor immunity in HCC. DPP-4 inhibitors were observed to be related with a decreased incidence of HCC in individuals with Type 2 Diabetes Mellitus (DM) with chronic HCV or HBV infection in two Taiwanese studies (59, 60). A mouse investigation revealed that DPP4 suppresses HCC growth in addition to its anti-diabetic properties (61). It is worthwhile to study the association between the usage of DPP4 inhibitors and the occurrence of HCC in HIV/HBV co-infected individuals.

Notably, the expression of putative hydroxypyruvate isomerase (HYI), the fifth protease, was shown to be elevated in HCC patients. The current understanding of HYI is rather limited. So yet, we have not retrieved studies on HYI. However, our findings imply that HYI is a new protease linked to HCC.

The findings of this investigation in an HIV-infected population revealed that cytoplasmic junction associated protein 2 (CLASP2) was differently upregulated in HCC. According to the research, HIV uses a variety of specific microtubule plus-end tracking proteins to drive the production of stable microtubules shortly after virus entrance and boost the early phases of infection (62). CLASP2 has been discovered as a host component that allows HIV to induce microtubule stability and boost early infection in natural target cell types (63). We hypothesized that the oncogenesis and process of HIV/HBV co-infection in HCC patients are exceedingly complex, and CLASP2 is one of the numerous pathogenic elements of HCC. This could be the difference between the factors impacting HCC in the PLWH group and the non-HIV infected population.

A mass spectrometry analysis discovered that IGLV5-45, located in the variable region of the immunoglobulin Lambda light chain, is the light chain peptide with the greatest variation between active and stable SLE (63). It was found to be associated with anhedonia/retardation (64). In HCC patients, the expression of immunoglobulin lambda variable 5-45 (IGLV5-45) and immunoglobulin heavy variable 1-69D (IGHV1-69D) in serum was enhanced, indicating B cell hyperactivity and an inflammatory condition. The immunoglobulin lambda light chain variable region, which is strongly associated to antigenantibody interactions and therapeutic target development, gave rise to IGLV5-45 molecules. It was identified as DEPs, indicating that HCC progression was tightly linked to immune response.

We acknowledge that there are some limitations with this study. First, since all of the participants were male, the results of this study are limited to a certain extent, and whether the results are applicable to female patients requires more examination. Second, to strengthen and promote the application value of predictive indicators of HCC, validation study would be done in a relatively larger population, including both HBV mono-infected patients and HIV/HBV co-infected patients. Third, the sample size of this study is relatively small. In the future, conditions will be created to supplement appropriate samples to further improve the experimental results and analysis. However, despite these limitations, the bio-informatics analysis results in this study could provide valuable clues for future study about early diagnosis and pathogenesis of HCC.

Conclusions

This is a proteomics-based study in an HIV/HBV co-infected group. A total of 124 DEP signatures were detected in patients with and without HCC, with 95 DEPs being up-regulated and 29 DEPs being down-regulated. The functional examination of DEPs indicates that the majority of them are abundant in extracellular matrix organization and membrane components that comprise the tumor microenvironment. Proteoglycan is a component of the extracellular matrix that is abundant in the KEGG pathways in HCC condition. The LASSO model screened eleven core differential proteins, including GAPR1, PLTP, CLASP2, IGHV1-69D, IGLV5-45, A2M, VNN1, KLK11, ANPEP, DPP4 and HYI, and the last five belonged to protease members. The findings could aid in the discovery of novel biomarkers for HCC diagnosis in patients living with HIV and HBV co-infection. Further research into the molecular processes underpinning how these DEPs influence HCC formation and progression may aid in the development of better therapeutic options for HCC prevention and therapy.

Data availability statement

The data presented in the study are deposited in the ProteomeXchange Consortium via the iProX repository, accession number PXD046369. IPX0007140003 in PXD046369 is the original sequence information of this study.

Ethics statement

The studies involving humans were approved by the institutional review board of the Zhongnan Hospital of Wuhan University. The studies were conducted in accordance with the local legislation and institutional requirements. Written informed consent for participation in this study was provided by the participants' legal guardians/next of kin.

Author contributions

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Conflict of interest

Author YY is employed by SpecAlly Life Technology Co., Ltd. The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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References

1. Yang JD, Roberts LR. Hepatocellular carcinoma: A global view. Nat Rev Gastroenterol Hepatol (2010) 7(8):448-58. doi: 10.1038/nrgastro.2010.100

2. Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin* (2018) 68(6):394–424. doi: 10.3322/ caac.21492

3. Soriano V, de Mendoza C, Fernández-Montero JV, Labarga P, Barreiro P. Management and treatment of chronic hepatitis B in HIV-positive patients. *Ann Med* (2014) 46(5):290-6. doi: 10.3109/07853890.2014.899103

4. Konopnicki D, Mocroft A, de Wit S, Antunes F, Ledergerber B, Katlama C, et al. Hepatitis B and HIV: prevalence, AIDS progression, response to HAART and increased mortality in the EuroSIDA cohort. *AIDS* (2005) 19:2117–25. doi: 10.1097/01.aids.0000163936.99401.fe

5. Chen C, Yang H, Su J, Jen C, You S, Lu S, et al. Risk of hepatocellular carcinoma across biological gradient of serum hepatitis B virus DNA level. *JAMA* (2006) 295:65–3. doi: 10.1001/jama.295.1.65

6. Puoti M, Torti C, Bruno R, Filice G, Carosi G. Natural history of chronic hepatitis B in co-infected patients. *J Hepatol* (2006) 44(1 Suppl):S65–70. doi: 10.1016/ j.jhep.2005.11.015

7. Martín-Carbonero L, Teixeira T, Poveda E, Plaza Z, Vispo E, Gonzalez-Lahoz J, et al. Clinical and virological outcomes in HIV-infected patients with chronic hepatitis B on long-term nucleos(t)ide analogues. *AIDS* (2011) 25:73–9. doi: 10.1097/QAD.0b013e328340fde2

8. Ganesan M, Poluektova LY, Kharbanda KK, Osna NA. Human immunodeficiency virus and hepatotropic viruses co-morbidities as the inducers of liver injury progression. *World J Gastroenterol* (2019) 25(4):398–410. doi: 10.3748/ wjg.v25.i4.398

9. European Association for the Study of the Liver. EASL Clinical Practice Guidelines: Management of hepatocellular carcinoma. *J Hepatol* (2018) 69(1):182–236. doi: 10.1016/j.jhep.2018.03.019

10. Toyoda H, Kumada T, Tada T. Highly sensitive Lens culinaris agglutinin-reactive α -fetoprotein: a new tool for the management of hepatocellular carcinoma. *Oncology* (2011) 81 Suppl 1:61–5. doi: 10.1159/000333263

11. Toyoda H, Kumada T, Tada T, Sone Y, Kaneoka Y, Maeda A. Tumor markers for hepatocellular carcinoma: simple and significant predictors of outcome in patients with HCC. *Liver Cancer* (2015) 4(2):126–36. doi: 10.1159/000367735

12. Chuaypen N, Chittmittraprap S, Pinjaroen N, Sirichindakul B, Poovorawan Y, Tanaka Y, et al. Serum Wisteria floribunda agglutinin-positive Mac-2 binding protein level as a diagnostic marker of hepatitis B virus-related hepatocellular carcinoma. *Hepatol Res* (2018) 48(11):872–81. doi: 10.1111/hepr.13187

13. Kim SU, Heo JY, Kim BK, Park JY, Kim DY, Han KH, et al. Wisteria floribunda agglutinin-positive human Mac-2 binding protein predicts the risk of HBV-related liver cancer development. *Liver Int* (2017) 37(6):879–87. doi: 10.1111/liv.13341

14. Fan R, Papatheodoridis G, Sun J, Innes H, Toyoda H, Xie Q, et al. aMAP risk score predicts hepatocellular carcinoma development in patients with chronic hepatitis. *J Hepatol* (2020) 73(6):1368–78. doi: 10.1016/j.jhep.2020.07.025

15. Costa APM, da Silva MACN, Castro RS, Sampaio ALO, Alencar Júnior AM, da Silva MC, et al. PAGE-B and REACH-B predicts the risk of developing hepatocellular

carcinoma in chronic hepatitis B patients from northeast, Brazil. Viruses (2022) 14 (4):732. doi: 10.3390/v14040732

16. Berzigotti A, Tsochatzis E, Boursier J, Castera L, Cazzagon N, Friedrich-Rust M, et al. European Association for the Study of the Liver. EASL Clinical Practice Guidelines on non-invasive tests for evaluation of liver disease severity and prognosis - 2021 update. *J Hepatol* (2021) 75(3):659–89. doi: 10.1016/j.jhep.2021.05.025

17. Yang JD, Addissie BD, Mara KC, Harmsen WS, Dai J, Zhang N, et al. GALAD score for hepatocellular carcinoma detection in comparison with liver ultrasound and proposal of GALADUS score. *Cancer Epidemiol Biomarkers Prev* (2019) 28(3):531–38. doi: 10.1158/1055-9965.EPI-18-0281

18. Kisiel JB, Dukek BA VSR, Kanipakam R, Ghoz HM, Yab TC, Berger CK, et al. Hepatocellular carcinoma detection by plasma methylated DNA: discovery, phase I pilot, and phase II clinical validation. *Hepatology* (2019) 69(3):1180–92. doi: 10.1002/hep.30244

19. Chalasani NP, Ramasubramanian TS, Bhattacharya A, Olson MC, Edwards V DK, Roberts LR, et al. A novel blood-based panel of methylated DNA and protein markers for detection of early-stage hepatocellular carcinoma. *Clin Gastroenterol Hepatol* (2021) 19(12):2597–605.e4. doi: 10.1016/j.cgh.2020.08.065

20. Cevik D, Yildiz G, Ozturk M. Common telomerase reverse transcriptase promoter mutations in hepatocellular carcinomas from different geographical locations. *World J Gastroenterol* (2015) 21(1):311–7. doi: 10.3748/wjg.v21.i1.311

21. Sun J, Althoff KN, Jing Y, Horberg MA, Buchacz K, Gill MJ, et al. Trends in hepatocellular carcinoma incidence and risk among persons with HIV in the US and Canada, 1996-2015. *JAMA Netw Open* (2021) 4(2):e2037512. doi: 10.1001/jamanetworkopen.2020.37512

22. Kim HN, Newcomb CW, Carbonari DM, Roy JA, Torgersen J, Althoff KN, et al. Risk of HCC with hepatitis B viremia among HIV/HBV-coinfected persons in North America. *Hepatology* (2021) 74(3):1190–202. doi: 10.1002/hep.31839

23. Yamashita T, Koshikawa N, Shimakami T, Terashima T, Nakagawa M, Nio K, et al. Serum laminin γ 2 monomer as a diagnostic and predictive biomarker for hepatocellular carcinoma. *Hepatology* (2021) 74(2):760–75. doi: 10.1002/hep.31758

24. Chao X, Ding WX. Role and mechanisms of autophagy in alcohol-induced liver injury. *Adv Pharmacol* (2019) 85:109–31. doi: 10.1016/bs.apha.2019.01.008

25. Ma X, McKeen T, Zhang J, Ding WX. Role and mechanisms of mitophagy in liver diseases. *Cells* (2020) 9(4):837. doi: 10.3390/cells9040837

26. Yang L, Li P, Fu S, Calay ES, Hotamisligil GS. Defective hepatic autophagy in obesity promotes ER stress and causes insulin resistance. *Cell Metab* (2010) 11(6):467–78. doi: 10.1016/j.cmet.2010.04.005

27. Sir D, Ann DK, Ou JH. Autophagy by hepatitis B virus and for hepatitis B virus. *Autophagy* (2010) 6(4):548–9. doi: 10.4161/auto.6.4.11669

28. Sir D, Chen WL, Choi J, Wakita T, Yen TS, Ou JH. Induction of incomplete autophagic response by hepatitis C virus *via* the unfolded protein response. *Hepatology* (2008) 48(4):1054–61. doi: 10.1002/hep.22464

29. Chao X, Wang S, Zhao K, Li Y, Williams JA, Li T, et al. Impaired TFEB-mediated lysosome biogenesis and autophagy promote chronic ethanol-induced liver injury and steatosis in mice. *Gastroenterology* (2018) 155(3):865–79.e12. doi: 10.1053/j.gastro.2018.05.027

30. Chao X, Qian H, Wang S, Ding WX. Autophagy and liver cancer. Clin Mol Hepatol (2020) 26(4):606–17. doi: 10.3350/cmh.2020.0169

31. Sirati N, Shen Z, Olrichs NK, Popova B, Verhoek IC, Lagerwaard IM, et al. GAPR-1 interferes with condensate formation of beclin 1 in saccharomyces cerevisiae. *J Mol Biol* (2023) 435(4):167935. doi: 10.1016/j.jmb.2022.167935

32. Van Galen J, Van Balkom BW, Serrano RL, Kaloyanova D, Eerland R, Stüven E, et al. Binding of GAPR-1 to negatively charged phospholipid membranes: unusual binding characteristics to phosphatidylinositol. *Mol Membr Biol* (2010) 27(2-3):81–91. doi: 10.3109/09687680903507080

33. Saunders RA, Fujii K, Alabanza L, Ravatn R, Kita T, Kudoh K, et al. Altered phospholipid transfer protein gene expression and serum lipid profile by topotecan. *Biochem Pharmacol* (2010) 80(3):362–9. doi: 10.1016/j.bcp.2010.04.015

34. Huang K, Chen S, Xie R, Jiang P, Yu C, Fang J, et al. Identification of three predictors of gastric cancer progression and prognosis. *FEBS Open Bio* (2020) 10 (9):1891–9. doi: 10.1002/2211-5463.12943

35. Dong W, Gong H, Zhang G, Vuletic S, Albers J, Zhang J, et al. Lipoprotein lipase and phospholipid transfer protein overexpression in human glioma cells and their effect on cell growth, apoptosis, and migration. *Acta Biochim Biophys Sin (Shanghai)* (2017) 49(1):62–73. doi: 10.1093/abbs/gmw117

36. Gautier T, Paul C, Deckert V, Desrumaux C, Klein A, Labbé J, et al. Innate immune response triggered by triacyl lipid A is dependent on phospholipid transfer protein (PLTP) gene expression. *FASEB J* (2010) 24(9):3544–54. doi: 10.1096/fj.09-152876

37. Gnanapradeepan K, Indeglia A, Stieg DC, Clarke N, Shao C, Dougherty JF, et al. PLTP is a p53 target gene with roles in cancer growth suppression and ferroptosis. *J Biol Chem* (2022) 298(12):102637. doi: 10.1016/j.jbc.2022.102637

38. Kang M, Qin W, Buya M, Dong X, Zheng W, Lu W, et al. VNN1, a potential biomarker for pancreatic cancer-associated new-onset diabetes, aggravates paraneoplastic islet dysfunction by increasing oxidative stress. *Cancer Lett* (2016) 373(2):241–50. doi: 10.1016/j.canlet.2015.12.031

39. Qin W, Kang M, Li C, Zheng W, Guo Q. VNN1 overexpression in pancreatic cancer cells inhibits paraneoplastic islet function by increasing oxidative stress and inducing β -cell dedifferentiation. *Oncol Rep* (2023) 49(6):120. doi: 10.3892/ or.2023.8557

40. Ozturk T, Kain EZ, Tuzuner MB, Diren A, Batur S, Yilmaz-Aydogan H, et al. Kallikrein 11 down-regulation in breast carcinoma: correlation with prognostic parameters. *In Vivo* (2021) 35(6):3233–43. doi: 10.21873/invivo.12618

41. McIntosh MW, Liu Y, Drescher C, Urban N, Diamandis EP. Validation and characterization of human kallikrein 11 as a serum marker for diagnosis of ovarian carcinoma. *Clin Cancer Res* (2007) 13(15 Pt 1):4422–8. doi: 10.1158/1078-0432.CCR-06-2224

42. Mitsui S, Yamada T, Okui A, Kominami K, Uemura H, Yamaguchi N. A novel isoform of a kallikrein-like protease, TLSP/hippostasin, (PRSS20), is expressed in the human brain and prostate. *Biochem Biophys Res Commun* (2000) 272(1):205–11. doi: 10.1006/bbrc.2000.2761

43. Stavropoulou P, Gregorakis AK, Plebani M, Scorilas A. Expression analysis and prognostic significance of human kallikrein 11 in prostate cancer. *Clin Chim Acta* (2005) 357(2):190–5. doi: 10.1016/j.cccn.2005.03.026

44. Patsis C, Yiotakis I, Scorilas A. Diagnostic and prognostic significance of human kallikrein 11 (KLK11) mRNA expression levels in patients with laryngeal cancer. *Clin Biochem* (2012) 45(9):623–30. doi: 10.1016/j.clinbiochem.2012.03.005

45. Zhang X, Hwang YS. Cancer-associated fibroblast stimulates cancer cell invasion in an interleukin-1 receptor (IL-1R)-dependent manner. *Oncol Lett* (2019) 18(5):4645– 50. doi: 10.3892/ol.2019.10784

46. Unal D, Tasdemir A, Oguz A, Eroglu C, Cihan YB, Turak EE, et al. Is human kallikrein-11 in gastric cancer treated with surgery and adjuvant chemoradiotherapy associated with survival? *Pathol Res Pract* (2013) 209(12):779–83. doi: 10.1016/j.prp.2013.09.004

47. Yu X, Tang HY, Li XR, He XW, Xiang KM. Over-expression of human kallikrein 11 is associated with poor prognosis in patients with low rectal carcinoma. *Med Oncol* (2010) 27(1):40–4. doi: 10.1007/s12032-009-9167-2

48. Siriphak S, Chanakankun R, Proungvitaya T, Roytrakul S, Tummanatsakun D, Seubwai W, et al. Kallikrein-11, in association with coiled-coil domain containing 25, as a potential prognostic marker for cholangiocarcinoma with lymph node metastasis. *Molecules* (2021) 26(11):3105. doi: 10.3390/molecules26113105

49. Unal D, Eroglu C, Tasdemir A, Karaman H, Kurtul N, Oguz A, et al. Is human kallikrein 11 in non-small cell lung cancer treated chemoradiotherapy associated with survival? *Cancer Res Treat* (2016) 48(1):98–105. doi: 10.4143/crt.2014.364

50. Zhai M, Yang Z, Zhang C, Li J, Jia J, Zhou L, et al. APN-mediated phosphorylation of BCKDK promotes hepatocellular carcinoma metastasis and proliferation *via* the ERK signaling pathway. *Cell Death Dis* (2020) 11(5):396. doi: 10.1038/s41419-020-2610-1

51. Röcken C, Licht J, Roessner A, Carl-McGrath S. Canalicular immunostaining of aminopeptidase N (CD13) as a diagnostic marker for hepatocellular carcinoma. *J Clin Pathol* (2005) 58(10):1069–75. doi: 10.1136/jcp.2005.026328

52. Sun ZP, Zhang J, Shi LH, Zhang XR, Duan Y, Xu WF, et al. Aminopeptidase N inhibitor 4cc synergizes antitumor effects of 5-fluorouracil on human liver cancer cells through ROS-dependent CD13 inhibition. *BioMed Pharmacother* (2015) 76:65–72. doi: 10.1016/j.biopha.2015.10.023

53. Inagaki Y, Tang W, Zhang L, Du G, Xu W, Kokudo N. Novel aminopeptidase N (APN/CD13) inhibitor 24F can suppress invasion of hepatocellular carcinoma cells as well as angiogenesis. *Biosci Trends* (2010) 4(2):56–60.

54. Zhao Y, Wu H, Xing X, Ma Y, Ji S, Xu X, et al. CD13 induces autophagy to promote hepatocellular carcinoma cell chemoresistance through the P38/Hsp27/ CREB/ATG7 pathway. *J Pharmacol Exp Ther* (2020) 374(3):512–20. doi: 10.1124/ jpet.120.265637

55. Itou M, Kawaguchi T, Taniguchi E, Sata M. Dipeptidyl peptidase-4: a key player in chronic liver disease. *World J Gastroenterol* (2013) 19(15):2298–306. doi: 10.3748/ wjg.v19.i15.2298

56. Yu DM, Yao TW, Chowdhury S, Nadvi NA, Osborne B, Church WB, et al. The dipeptidyl peptidase IV family in cancer and cell biology. *FEBS J* (2010) 277(5):1126–44. doi: 10.1111/j.1742-4658.2009.07526.x

57. Kawaguchi T, Kodama T, Hikita H, Makino Y, Saito Y, Tanaka S, et al. Synthetic lethal interaction of combined CD26 and Bcl-xL inhibition is a powerful anticancer therapy against hepatocellular carcinoma. *Hepatol Res* (2015) 45(9):1023–33. doi: 10.1111/hepr.12434

58. Henderson JM, Xiang MSW, Huang JC, Wetzel S, Jiang L, Lai JH, et al. Dipeptidyl peptidase inhibition enhances CD8 T cell recruitment and activates intrahepatic inflammasome in a murine model of hepatocellular carcinoma. *Cancers (Basel)* (2021) 13(21):5495. doi: 10.3390/cancers13215495

59. Hsu WH, Sue SP, Liang HL, Tseng CW, Lin HC, Wen WL, et al. Dipeptidyl peptidase 4 inhibitors decrease the risk of hepatocellular carcinoma in patients with chronic hepatitis C infection and type 2 diabetes mellitus: A nationwide study in Taiwan. *Front Public Health* (2021) 9:711723. doi: 10.3389/fpubh.2021.711723

60. Chen TI, Lee FJ, Hsu WL, Chen YC, Chen M. Association of dipeptidyl peptidase-4 inhibitors use with reduced risk of hepatocellular carcinoma in type 2 diabetes patients with chronic HBV infection. *Cancers (Basel)* (2023) 15(4):1148. doi: 10.3390/cancers15041148

61. Kawakubo M, Tanaka M, Ochi K, Watanabe A, Saka-Tanaka M, Kanamori Y, et al. Dipeptidyl peptidase-4 inhibition prevents nonalcoholic steatohepatitisassociated liver fibrosis and tumor development in mice independently of its antidiabetic effects. *Sci Rep* (2020) 10(1):983. doi: 10.1038/s41598-020-57935-6

62. Mitra S, Shanmugapriya S, Santos da Silva E, Naghavi MH. HIV-1 exploits CLASP2 to induce microtubule stabilization and facilitate virus trafficking to the nucleus. *J Virol* (2020) 94(14):e00404-20. doi: 10.1128/JVI.00404-20

63. Jiang J, Zhao J, Liu D, Zhang M. Different roles of urinary light chains and serum light chains as potential biomarkers for monitoring disease activity in systemic lupus erythematosus. *PeerJ* (2022) 10:e13385. doi: 10.7717/peerj.13385

64. Lee H, Rhee SJ, Kim J, Lee Y, Kim H, Lee J, et al. Predictive protein markers for depression severity in mood disorders: A preliminary trans-diagnostic approach study. *J Psychiatr Res* (2021) 142:63–72. doi: 10.1016/j.jpsychires.2021.07.041