Check for updates

OPEN ACCESS

EDITED BY Akira Sugawara, Tohoku University, Japan

REVIEWED BY Filippo Biscarini, National Research Council (CNR), Italy Thomas Kaiser, University of Minnesota Twin Cities, United States

CORRESPONDENCE Huawei Zhang Slyyzhw@163.com; Zhanghuawei@sdfmu.edu.cn Qian Wang Wangqian122411@126.com

RECEIVED 26 July 2023 ACCEPTED 23 October 2023 PUBLISHED 10 November 2023

CITATION

Liu X, Liu J, Zhang T, Wang Q and Zhang H (2023) Complex relationship between gut microbiota and thyroid dysfunction: a bidirectional two-sample Mendelian randomization study. *Front. Endocrinol.* 14:1267383. doi: 10.3389/fendo.2023.1267383

COPYRIGHT

© 2023 Liu, Liu, Zhang, Wang and Zhang. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Complex relationship between gut microbiota and thyroid dysfunction: a bidirectional two-sample Mendelian randomization study

Xiao Liu, Jingyu Liu, Tongxin Zhang, Qian Wang* and Huawei Zhang*

Department of Ultrasound, Shandong Provincial Hospital Affiliated to Shandong First Medical University, Jinan, Shandong, China

Background: Many studies have reported the link between gut microbiota and thyroid dysfunction. However, the causal effect of gut microbiota on thyroid dysfunction and the changes in gut microbiota after the onset of thyroid dysfunction are not clear.

Methods: A two-sample Mendelian randomization (MR) study was used to explore the complex relationship between gut microbiota and thyroid dysfunction. Data on 211 bacterial taxa were obtained from the MiBioGen consortium, and data on thyroid dysfunction, including hypothyroidism, thyroid-stimulating hormone alteration, thyroxine deficiency, and thyroid peroxidase antibodies positivity, were derived from several databases. Inverse variance weighting (IVW), weighted median, MR-Egger, weighted mode, and simple mode were applied to assess the causal effects of gut microbiota on thyroid dysfunction. Comprehensive sensitivity analyses were followed to validate the robustness of the results. Finally, a reverse MR study was conducted to explore the alteration of gut microbiota after hypothyroidism onset.

Results: Our bidirectional two-sample MR study revealed that the genera *Intestinimonas, Eubacterium brachy group, Ruminiclostridium5,* and *Ruminococcaceae UCG004* were the risk factors for decreased thyroid function, whereas the genera *Bifidobacterium* and *Lachnospiraceae UCG008* and phyla Actinobacteria and Verrucomicrobia were protective. The abundance of eight bacterial taxa varied after the onset of hypothyroidism. Sensitivity analysis showed that no heterogeneity or pleiotropy existed in the results of this study.

Conclusion: This novel MR study systematically demonstrated the complex relationship between gut microbiota and thyroid dysfunction, which supports the selection of more targeted probiotics to maintain thyroid–gut axis homeostasis and thus to prevent, control, and reverse the development of thyroid dysfunction.

KEYWORDS

Mendelian randomization, thyroid dysfunction, hypothyroidism, gut microbiota, thyroid-gut axis

Introduction

Thyroid dysfunction stands as one of the prevailing endocrine diseases, and hypothyroidism is one of its main types, affecting approximately 5% of the general population (1). Thyroid hormones exert their influence selectively on multiple organs and tissues. Hypothyroidism, a representation of thyroid dysfunction, exhibits strong associations with cardiovascular diseases (2), diabetes (3), and thromboembolism (4), among other conditions. Recent epidemiological studies globally have unveiled a significant correlation between hypothyroidism and elevated mortality rates (5). Hypothyroidism can arise due to factors such as iodine deficiency, medications, radiation therapy, immune system abnormalities, and pregnancy. Once established, reversing hypothyroidism poses significant challenges. Among these triggers, autoimmune hypothyroidism proves especially problematic, given its enduring nature. Hence, exploring autoimmune hypothyroidism demands comprehensive analysis from diverse angles. It necessitates in-depth research to uncover treatments beyond lifelong thyroxine administration. These treatments should not only impede the disease's progression but also enhance patients' quality of life significantly.

The human gut microbiota boasts unparalleled diversity and complexity among all human organs, with its composition intricately linked to ethnicity, dietary habits, and geographic location (6). Variations in gut microbiota among healthy individuals from diverse backgrounds pose a challenge in establishing a definitive healthy baseline. This microbiota actively participates in the fundamental physiological functions and diseases within the human body, including nutrient production, metabolic balance, immune response, brain behavior, and inflammatory reactions (7). Moreover, it plays a pivotal role in several endocrine diseases, notably diabetes (8) and polycystic ovary syndrome (9). The evolving recognition of the interconnection between gut microbiota and thyroid function is denoted as the thyroid-gut axis (10, 11). Dysbiosis in the gut microbiota has been evidenced to disrupt the absorption of iodine and to impact the synthesis and release of thyroid hormones (12). Researchers have explored this relationship, achieving compelling results by modulating gut microbiota in mouse models and clinical cases of thyroid dysfunction (13-16). For instance, hyperthyroidism correlates with increased Actinobacteria and decreased Bacteroidetes, whereas specific strains of *Bifidobacterium* and *Lactobacillus* were found to interact with human autoantibodies, disrupting thyroid function (14, 17). Clinical study has also linked severe hypothyroidism to higher instances of small intestinal bacterial overgrowth, contributing to gastrointestinal symptoms (18).

Although these studies have indicated associations between gut microbiota and thyroid dysfunction, causality remains elusive in traditional observational studies due to confounding and reverse causation, which can lead to biased conclusions. A bidirectional two-sample Mendelian randomization (MR) study presents an innovative method for studying the causal effects between environmental exposures and diseases, or between two diseases, akin to a randomized controlled trial (RCT) (19). MR employs single-nucleotide polymorphisms (SNPs) in genes as instrumental variables (IVs), effectively circumventing confounding factors, and provides statistically significant causal effects by combining SNPs from genome-wide association studies (GWASs) of exposure and outcome (20, 21). It allows for better modeling of random assignment, thereby reducing bias in observational studies and better inferring causality. Although MR has been applied broadly to investigate the gut microbiota's role in diseases like cancers and autoimmune disorders, its application to thyroid dysfunction remains unexplored (22-24). Therefore, we conducted this bidirectional two-sample MR study utilizing thyroid functionrelated GWAS data from the international consortia, including the MiBioGen consortium, FinnGen consortium, MRC IEU OpenGWAS project, and ThyroidOmics. In this study, the application of two-sample MR involved the utilization of two distinct, independent European samples for analysis. This approach was employed to mitigate the potential biases associated with a single sample, thereby enhancing the reliability and generalizability of our findings. Furthermore, we used the bidirectional MR that simultaneously considered two different causal directions between the gut microbiota and autoimmune hypothyroidism, thus providing for a more nuanced understanding of the bidirectional causal relationship between them. This comprehensive MR analysis, to our knowledge, marks the first exploration of the association between gut microbiota and thyroid function. These results can determine the bidirectional causal effect between the gut microbiota and thyroid dysfunction and thus guide the regulation of the thyroid-gut axis.

Methods

Study design

The study's flowchart is depicted in Figure 1. To investigate the interplay between gut microbiota and thyroid dysfunction, we selected gut microbiota as the exposure and thyroid function–related factors as outcomes. MR study adhered to three key assumptions: (1) IVs selected from datasets were linked to exposure; (2) they were unrelated to unidentified exposure confounders; and (3) they influenced outcomes exclusively through exposure pathways (25). SNPs served as valid IVs in the MR study to evaluate the bidirectional causal relationship between exposure and outcome.

Data sources

All data utilized in this study were sourced from diverse GWASs. Regarding thyroid function, the GWASs focused on hypothyroidism, thyroid-stimulating hormone (TSH), thyroxine deficiency, and thyroid peroxidase antibodies (TPOAb) positivity except for gut microbiota data. For the gut microbiota analysis, we extracted the relevant genetic IVs from a comprehensive GWAS dataset provided by the MiBioGen consortium. This dataset was composed of 18,340 participants, primarily of European descent (n = 13,266), and comprised a total of 5,717,754 SNPs after imputation (21). The MiBioGen GWAS amalgamated outcomes from 16S ribosomal RNA gene sequencing effectively eliminated potential batch effects. In total, the dataset contained information on 211 taxa, including 131 genera, 35 families, 20 orders, 16 classes, and nine phyla. As for the thyroid function investigation, the genetic IVs were acquired from the FinnGen consortium (26), MRC IEU OpenGWAS project (27), and ThyroidOmics consortium (28), respectively. The details of each database are summarized in Table 1.

Instrumental variables selection

To infer an accurate and realistic bidirectional causal effect between gut microbiota and hypothyroidism risk, we performed a rigorous quality control to select the best IVs. First, we screened SNPs associated with each bacterial taxon using a mild p-value of 1×10^{-5} . Then, linkage disequilibrium analysis was used to select independent IVs for each bacterial taxon to prevent biased causal estimates. We selected the most suitable parameters for clustering ($\mathbb{R}^2 < 0.01$ and clustering distance = 500 kb) to evaluate the linkage disequilibrium among the included SNPs. The population was framed as European. Missing SNPs from bacterial taxa in thyroid dysfunction–related datasets were replaced with proxy SNPs ($\mathbb{R}^2 > 0.8$). Palindromic SNPs were excluded to prevent coding distortions. The F-statistic was calculated to detect the presence of weak IV instrument bias. When the F-statistic is greater than 10, it indicates that there is no weak instrument bias.

MR analysis

Five widely used MR methods were employed to detect the bidirectional causal relationships between exposure and outcome, encompassing inverse variance weighting (IVW), weighted median, MR-Egger, weighted mode, and simple mode (29-32). IVW method estimates the causal effect of exposure on the outcome by integrating ratio estimates for each SNP, and it was chosen as the primary method because it can provide a robust and unbiased causal effect when no polymorphism or heterogeneity is found (33). The weighted median method correctly estimates causality when up to 50% of the IVs are invalid (30). MR-Egger method is based on the assumption that instrument strengths are independent of direct effects and thereby allows calibration of pleiotropy and calculation of causal inferences, even if all genetic variants are polymorphic (34). If this assumption is violated, then the weighted model method would have greater power to detect causal effects and produce less bias than the MR-Egger method (34). Finally, the simple model method is an unweighted model of the experienced density function for causal estimation (35). A positive causal effect was affirmed if the IVW results were significant (p < 0.05) and beta values from other methods concurred in direction. Wald radio method was used to estimate the causal effect of exposure on the outcome when there was only one SNP, which was the simplest calculation method (36). Then, we further visualized the results of the five MR methods. The



Data	Trait	Year	Consortium	Ancestry	Sample size	Number of SNPs
Gut microbiomes	Gut microbiota abundance	2021	MiBioGen consortium	European	13,266	5,717,754
Hypothyroidism	Hypothyroidism, strict autoimmune	2021	FinnGen consortium	European	198,472	16,380,353
TSH alteration	Thyroid-stimulating hormone	2018	MRC IEU OpenGWAS project	European	3,301	10,534,735
Thyroxine deficiency	Medication code: levothyroxine sodium	2018	MRC IEU OpenGWAS project	European	462,933	9,851,867
TPOAb positivity	Thyroid peroxidase antibodies' positivity	2014	ThyroidOmics consortium	European	18,297	10,485,757

TABLE 1 Data sources used in the bidirectional two-sample MR study.

bidirectional causal effect was expressed as an odds ratio (OR) calculated from MR analysis. Notably, to obtain more IVs, we did not reach the traditionally strict significance threshold ($p < 5 \times 10^{-8}$) for exposure to gut microbiota. Thus, a false discovery rate (FDR) was corrected for multiple comparisons by the Benjamin–Hochberg method to limit the possibility of false positives, and the threshold was set at 0.05. The bidirectional causal effect was considered significant when p < 0.05 and FDR < 0.05 (37).

Sensitivity analyses

Several basic sensitivity analyses were used to validate the results. Cochran's Q statistic was employed to assess heterogeneity among IVs. The heterogeneity should be noted if it exists between different IVs (p < 0.05) (38). Horizontal pleiotropy indicates that IVs are associated with the outcome through pathways other than causal effects, potentially leading to false positive results (p < 0.05). MR pleiotropy residual sum and outlier (MR-PRESSO) analysis was also used to validate the potential pleiotropy of the direct effect between the selected IVs and the outcome. Subsequently, we applied the leave-one-out method to exclude each SNP from the IVs, evaluatingwhether individual SNPs significantly influenced causal effects using the IVW method. All the above analyses were done using two-sample MR and MR-PRESSOR R packages (35, 39).

Reverse MR analysis

To investigate alterations of gut microbiota following the onset of hypothyroidism, we conducted a reverse MR analysis, treating hypothyroidism as the exposure and gut microbiota as the outcome. The threshold of significance level for IVs was adjusted to be more accurate ($p < 5 \times 10^{-8}$). The procedure for the reverse MR analysis was the same as the MR analysis described above.

Results

Causal effects of gut microbiota on thyroid dysfunction

A total of 14,405 SNPs associated with the gut microbiota were meticulously curated following a rigorous screening process. Through robust linkage disequilibrium clustering and coordination, each bacterial taxon was linked to a varied range of IVs, spanning from 3 to 22. Notably, the F-statistic for all SNPs surpassed 10, indicating the absence of weak instrumental bias in the dataset.

After FDR correction, a meticulous analysis targeted three phyla and seven genera of bacteria taxa, revealing their causal effects on thyroid dysfunction via MR analysis. Six were linked to hypothyroidism, one to alterations in TSH levels, and three to TPOAb positivity (Figure 2). The IVW estimate pointed toward the genera Intestinimonas (OR = 1.120, p = 0.014) and Ruminiclostridium5 (OR = 1.189, p = 0.011) as the risk factors for hypothyroidism. In contrast, genera Bifidobacterium (OR = 0.877, p = 0.011) and Lachnospiraceae UCG008 (OR = 0.871, p = 0.002) and phyla Actinobacteria (OR = 0.827, p = 0.001) and Verrucomicrobia (OR = 0.876, p = 0.012) emerged as the protective factors against hypothyroidism. Moreover, phylum Bacteroidetes displayed an association with reduced TSH levels (OR = 0.711, p = 0.010). Three bacterial genera exhibited potential causal effects on TPOAb positivity: genus Anaerotruncus (OR = 0.291, p = 0.025) reduced TPOAb positivity, whereas genera Eubacterium brachy group (OR = 1.744, p = 0.034) and Ruminococcaceae UCG004 (OR = 2.193, p = 0.033) increased TPOAb positivity, as indicated by the IVW method. Intriguingly, no bacteria demonstrated a significant causal relationship with thyroxine deficiency. Rigorous sensitivity analyses revealed the absence of heterogeneity and pleiotropy in the identified causal effects (Supplementary Table 1). Furthermore, the leave-one-out method demonstrated the absence of outlier SNPs in all selected bacteria taxa. Regrettably, this method could not be performed due to the limited number of SNPs associated with TPOAb as the outcome (Figure 3).

Exposure	Outcome	Method	nSNP		OR	p-
genus Bifidobacterium	Hypothyroidism	MR Egger	17		0.819 (0.620-1.082)	
		Weighted median	17	1.0-I	0.820 (0.731-0.921)	
		IVW	17	10 0 01	0.877 (0.793-0.970)	
		Simple mode	17		0.843 (0.687-1.035)	
		Weighted mode	17	1- 6 -4	0.815 (0.716-0.928)	
genus Intestinimonas	Hypothyroidism	MR Egger	16		1.045 (0.811-1.346)	
		Weighted median	16		1.096 (0.971-1.236)	
		IVW	16		1.120 (1.023-1.226)	
		Simple mode	16		1.059 (0.861-1.301)	
		Weighted mode	16		1.098 (0.891-1.352)	
genus Lachnospiraceae UCG008	Hypothyroidism	MR Egger	11		0.566 (0.370-0.867)	
		Weighted median	11		0.904 (0.805-1.016)	
		IVW	11	1- 6- 1	0.871 (0.798-0.950)	
		Simple mode	11		0.919 (0.744-1.136)	
		Weighted mode	11	p	0.926 (0.759-1.130)	
genus Ruminiclostridium5	Hypothyroidism	MR Egger	11	ı	1.053 (0.600-1.847)	
		Weighted median	11	I	1.262 (1.051-1.515)	
		IVW	11	····•	1.189 (1.040-1.361)	
		Simple mode	11	· · · · · · · · · · · · · · · · · · ·	1.323 (0.948-1.845)	
		Weighted mode	11	•	1.340 (0.948-1.894)	
phylum Actinobacteria	Hypothyroidism	MR Egger	17		0.560 (0.363-0.865)	
		Weighted median	17		0.748 (0.642-0.871)	
		IVW	17	10 0 00	0.827 (0.738-0.926)	
		Simple mode	17	I	0.744 (0.570-0.972)	
		Weighted mode	17	P	0.737 (0.608-0.892)	
phylum Verrucomicrobia	Hypothyroidism	MR Egger	12	BB	0.881 (0.672-1.155)	
		Weighted median	12	*··•·	0.889 (0.773-1.021)	
		IVW	12	+ • • •	0.876 (0.790-0.971)	
		Simple mode	12	I	0.943 (0.756-1.176)	
		Weighted mode	12		0.907 (0.747-1.102)	
phylum Bacteroidetes	TSH	MR Egger	12	p	0.562 (0.326-0.970)	
		Weighted median	12		0.701 (0.486-1.011)	
		IVW	12	F	0.711 (0.549-0.921)	
		Simple mode	12		0.633 (0.369-1.084)	
		Weighted mode	12		0.662 (0.419-1.047)	
genus Anaerotruncus	TPOAb	IVW	2		0.291 (0.099-0.859)	
genus Eubacterium brachy group	TPOAb	IVW	2	·	1.744 (1.042-2.918)	
genus Ruminococcaceae UCG004	TPOAb	IVW	2		2.193 (1.066-4.511)	

FIGURE 2

Forest plot for the causal effect of gut microbiota on thyroid dysfunction by the five MR methods. The error bar represents the 95% confidence interval of hazard ratio. The red dot represents an outcome of hypothyroidism, the green represents an outcome of TSH alteration, and the blue represents an outcome of TPOAb positivity. nSNP, number of SNPs; OR, odds ratio; MR, Mendelian randomization; IVW, inverse variance weighted.



Causal effects of thyroid dysfunction on the gut microbiota

In the reverse MR analysis, 56 SNPs linked with hypothyroidism were identified. Each bacterial taxon was linked to a minimum of 3 and a maximum of 36 IVs, all exhibiting robustness (F-statistic > 10).

Applying the IVW method in the MR analysis, it was observed that class Negativicutes (OR = 1.039, p = 0.048), family Christensenellaceae (OR = 1.065, p = 0.030), genera *Eubacterium ruminantium group* (OR = 1.057, p = 0.042) and *Ruminococcaceae UCG005* (OR = 1.047, p = 0.025), and order Selenomonadales (OR = 1.039, p = 0.048) exhibited upregulation following the onset of

hypothyroidism. Conversely, class Verrucomicrobiae (OR = 0.954, p = 0.029), family Verrucomicrobiaceae (OR = 0.954, p = 0.024), order Verrucomicrobiales (OR = 0.954, p = 0.029), phylum Verrucomicrobia (OR = 0.954, p = 0.024), and genera *Akkermansia* (OR = 0.954, p = 0.029) and *Erysipelotrichaceae UCG003* (OR = 0.906, p = 0.033) were downregulated subsequent to the onset of hypothyroidism (Figure 4). Rigorous sensitivity analyses confirmed the absence of heterogeneity or horizontal pleiotropy in the aforementioned causal effects (Supplementary Table 2). In addition, leave-one-out analysis did not reveal any SNPs driving the causal effect of hypothyroidism on gut microbiota (Figure 5).

Discussion

Thyroid dysfunction has emerged as a global public health concern. Advancements in scientific inquiry have delved deeper into gut microbiota research, particularly in its connection to thyroid function via the thyroid–gut axis, a burgeoning area of interest (1, 40). Hypothyroidism, a prevalent condition stemming from thyroid dysfunction, exhibits multifaceted and poorly understood causes. Beyond external causes such as iodine deficiency, medications, and surgery, autoimmune origins constitute the bulk of primary hypothyroidism, posing significant

challenges for explanation and management (41). Gut microbiota serves as the linchpin for stable intestinal lymphoid tissue function, acting as a vital shield in immune homeostasis, enhancing tolerance to autoantigens and non-pathogenic non-autoantigens (11). The interplay between gut microbiota and the host's innate and adaptive immunity potentially influences the susceptibility to autoimmune thyroid disease (42, 43). Moreover, reduced intestinal motility in patients with hypothyroidism might disrupt intestinal substrate utilization and physicochemical conditions, culminating in gut microbiota dysbiosis exacerbating the condition or giving rise to complications (44). To address these intricacies, we conducted a comprehensive bidirectional two-sample MR study utilizing multiple GWAS datasets. This investigation aimed to elucidate the bidirectional causal effects links between gut microbiota and thyroid dysfunction, shedding light on the pathogenesis of autoimmune hypothyroidism and informing strategies for prevention, delay, and reversal of thyroid dysfunction-associated health conditions.

Numerous clinical trials have identified disparities in gut microbiota composition between thyroid dysfunction patients and healthy populations. However, establishing the bidirectional causal relationship between gut microbiota alterations and thyroid dysfunction remained elusive. For instance, a cross-sectional study encompassing 97 cases revealed significantly lower levels of *Alistipes*,



FIGURE 4

Forest plot for the causal effect of hypothyroidism on gut microbiota by the five MR methods. The error bar represents the 95% confidence interval of the hazard ratio. nSNP, number of SNPs; OR, odds ratio; MR, Mendelian randomization; IVW, inverse variance weighted.



Lachnospiraceae, Intestinimonas, Ruminococcus, and Subdoligranulum in hypothyroid patients, whereas Phascolarctobacterium and Bacteroidetes were more abundant (45). Another study comparing 29 hypothyroid patients with 11 healthy individuals highlighted a higher prevalence of Actinobacteria and Enterobacteriaceae and significantly diminished counts of Bifidobacteria and Ruminococcaceae in the hypothyroid group (46). Our MR analysis not only validated these findings but also established the bidirectional causal relationship between altered gut microbiota and diminished thyroid function. Notably, the genera Intestinimonas and Ruminiclostridium5 were linked to reduced thyroid function, whereas the genera Bifidobacterium and Lachnospiraceae UCG008 and phylum Actinobacteria mitigated this decline. Furthermore, only after the onset of hypothyroidism did the genera Eubacterium ruminantium group, Ruminococcaceae UCG005, and Erysipelotrichaceae UCG003 exhibit changes (47, 48). Previous controlled studies also revealed a significant decrease in Bacteroidetes in hyperthyroid patients with elevated TSH levels, with MR analysis corroborating Bacteroidetes' role in decreasing TSH (49). In an RCT, decreased Eubacterium post-probiotic intervention ameliorated complications in patients not treated with radioiodine after thyroidectomy, aligning with our study's identification of Eubacterium as a risk factor for TPOAb positivity (50).

Several mechanisms underpinning gut microbiota's impact on thyroid function via the thyroid-gut axis have been elucidated. The equilibrium between pathogenic and probiotic bacteria is pivotal for maintaining gut barrier function. Bifidobacterium, a widely used probiotic, confers several physiological benefits to humans, rendering it a protective genus against hypothyroidism in our study (51). In addition, phylum Actinobacteria and genus Lachnospiraceae UCG008, both protective in this study, are implicated in human sugar and protein metabolism. Phylum Actinobacteria participates in the biosynthesis of phenylalanine, tyrosine, and tryptophan, whereas Lachnospiraceae UCG008 is associated with alanine, aspartate, and glutamate metabolism (52-54). Our results also identified Ruminiclostridium5 as a risk for promoting hypothyroidism. Although limited information is available on Ruminiclostridium5, its increased abundance is linked to systemic inflammation and a negative correlation with secondary and conjugated bile acids (55, 56). Consistent with prior studies, phylum Verrucomicrobia emerged as a protective factor against hypothyroidism in our result. This phylum comprises diverse beneficial bacteria for the gut, whose outer membrane proteins effectively safeguard interactions with other cells (57). Genus Akkermansia, a member of the phylum Verrucomicrobia,

plays a significant role in enhancing host metabolic function and immune responses. Intriguingly, its abundance significantly decreases after hypothyroidism onset (58). Despite these insights, in-depth *in vivo* and *in vitro* experiments are imperative to explore the effects and mechanisms of gut microbiota as delineated in our findings.

The bidirectional two-sample MR study offers significant advantages in investigating the bidirectional causal relationship between gut microbiota and hypothyroidism. First, the sample size of clinical trials often lacks the representativeness necessary for generalizability. In contrast, our study utilized gut microbiome data from 13,266 samples. Thyroid function-related data were sourced from four datasets in three databases, encompassing hundreds of thousands of samples, each mutually exclusive. This vast dataset enhances the present results' representativeness and credibility. Moreover, IV analysis grounded in effectively mitigates confounding factors and eliminates the outcome's interference with the exposure's reverse effect. Utilizing the MR-PRESSO approach and a comprehensive set of sensitivity analyses eliminates study pleiotropy and heterogeneity. The non-duplication of GWAS datasets for both exposure and outcome samples significantly reduces and avoids bias. However, it is crucial to acknowledge the study's limitations. Gut microbiota data were derived from the GWAS meta-analysis rather than raw data, precluding subgroup analyses. In addition, the study's inclusion was limited to individuals of European ancestry, restricting the applicability of our findings to broader populations. Furthermore, the lowest taxonomic level for gut bacteria is the genus, precluding an indepth exploration of gut microbiota's causal effects on hypothyroidism at the species level. Notably, some results, with a limited number of SNPs, necessitate cautious interpretation. It is pivotal to underscore that RCTs remain the gold standard for treatment development and establishing causal relationships in biological contexts. They offer unparalleled control and evidence. Mendelian randomization analysis serves as a potent supplementary tool, especially in scenarios where conducting RCTs proves challenging.

Conclusions

In summary, this study firstly and comprehensively provides evidence to support the effects of various gut microbiota on thyroid dysfunction, including hypothyroidism, TSH, thyroxine, and TPOAb, and further reveals alterations in gut microbiota following hypothyroidism. Genera *Intestinimonas, Eubacterium brachy group, Ruminiclostridium5*, and *Ruminococcaceae UCG004* are found to be risk factors for decreased thyroid function, whereas genera *Bifidobacterium* and *Lachnospiraceae UCG008* and phyla Actinobacteria and Verrucomicrobia were protective. Eight types of gut microbiota are thought to show altered abundance after the onset of hypothyroidism. This bidirectional two-sample MR study provides fairly strong evidence for the thyroid–gut axis theory to select more targeted probiotics to reverse the disturbed immune system and control the progression of thyroid dysfunction.

Data availability statement

The datasets presented in this study can be found in online repositories. The names of the repository/repositories can be found in the article.

Author contributions

XL: Data curation, Methodology, Software, Writing – original draft, Investigation. JL: Methodology, Writing – original draft, Formal Analysis. TZ: Software, Writing – original draft, Data curation. QW: Conceptualization, Validation, Writing – review & editing, Funding acquisition, Methodology, Supervision, Visualization. HZ: Supervision, Writing – review & editing, Project administration.

Funding

The author(s) declare financial support was received for the research, authorship, and/or publication of this article. The work was supported by the Natural Science Foundation of Shandong Province (grant number ZR2021QH047) and the Clinical Science and Technology Innovation Development Program of Jinan (grant number 202134036).

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fendo.2023.1267383/ full#supplementary-material

References

1. Su X, Zhao Y, Li Y, Ma S, Wang Z. Gut dysbiosis is associated with primary hypothyroidism with interaction on gut-thyroid axis. *Clin Sci (Lond)* (2020) 134 (12):1521–35. doi: 10.1042/cs20200475

2. Paschou SA, Bletsa E, Stampouloglou PK, Tsigkou V, Valatsou A, Stefanaki K, et al. Thyroid disorders and cardiovascular manifestations: an update. *Endocrine* (2022) 75(3):672–83. doi: 10.1007/s12020-022-02982-4

3. Roa Dueñas OH, van der Burgh AC, Ittermann T, Ligthart S, Ikram MA, Peeters R, et al. Thyroid function and the risk of prediabetes and type 2 diabetes. *J Clin Endocrinol Metab* (2022) 107(6):1789–98. doi: 10.1210/clinem/dgac006

4. Han F, Zhang C, Xuan M, Xie Z, Zhang K, Li Y. Effects of hyperthyroidism on venous thromboembolism: A Mendelian randomization study. *J Immunol Res* (2022) 2022:2339678. doi: 10.1155/2022/2339678

5. Chen YL, Tian S, Wu J, Li H, Li S, Xu Z, et al. Impact of thyroid function on the prevalence and mortality of metabolic dysfunction-associated fatty liver disease. *J Clin Endocrinol Metab* (2023) 108(7): e434–e43. doi: 10.1210/clinem/dgad016

6. Durack J, Lynch SV. The gut microbiome: relationships with disease and opportunities for therapy. J Exp Med (2019) 216(1):20-40. doi: 10.1084/jem.20180448

7. Cryan JF, Dinan TG. Mind-altering microorganisms: the impact of the gut microbiota on brain and behaviour. *Nat Rev Neurosci* (2012) 13(10):701–12. doi: 10.1038/nrn3346

8. Weninger SN, Ding A, Browne EN, Frost ML, Schiro G, Laubitz D, et al. Longitudinal characterization of the gut microbiota in the diabetic zdsd rat model and therapeutic potential of oligofructose. *Metabolites* (2023) 13(5):660. doi: 10.3390/ metabol3050660

9. Sun Y, Gao S, Ye C, Zhao W. Gut microbiota dysbiosis in polycystic ovary syndrome: mechanisms of progression and clinical applications. *Front Cell Infect Microbiol* (2023) 13:1142041. doi: 10.3389/fcimb.2023.1142041

10. Knezevic J, Starchl C, Tmava Berisha A, Amrein K. Thyroid-gut-axis: how does the microbiota influence thyroid function? *Nutrients* (2020) 12(6):1769. doi: 10.3390/nu12061769

11. Köhling HL, Plummer SF, Marchesi JR, Davidge KS, Ludgate M. The microbiota and autoimmunity: their role in thyroid autoimmune diseases. *Clin Immunol* (2017) 183:63–74. doi: 10.1016/j.clim.2017.07.001

12. Virili C, Centanni M. Does microbiota composition affect thyroid homeostasis? Endocrine (2015) 49(3):583-7. doi: 10.1007/s12020-014-0509-2

13. Virili C, Stramazzo I, Centanni M. Gut microbiome and thyroid autoimmunity. *Best Pract Res Clin Endocrinol Metab* (2021) 35(3):101506. doi: 10.1016/ j.beem.2021.101506

14. Biscarini F, Masetti G, Muller I, Verhasselt HL, Covelli D, Colucci G, et al. Gut microbiome associated with graves disease and graves orbitopathy: the indigo multicenter European study. *J Clin Endocrinol Metab* (2023) 108(8):2065–77. doi: 10.1210/clinem/dgad030

15. Moshkelgosha S, Verhasselt HL, Masetti G, Covelli D, Biscarini F, Horstmann M, et al. Modulating gut microbiota in a mouse model of Graves' orbitopathy and its impact on induced disease. *Microbiome* (2021) 9(1):45. doi: 10.1186/s40168-020-00952-4

16. Masetti G, Moshkelgosha S, Köhling HL, Covelli D, Banga JP, Berchner-Pfannschmidt U, et al. Gut microbiota in experimental murine model of Graves' orbitopathy established in different environments may modulate clinical presentation of disease. *Microbiome* (2018) 6(1):97. doi: 10.1186/s40168-018-0478-4

17. Kiseleva EP, Mikhailopulo KI, Sviridov OV, Novik GI, Knirel YA, Szwajcer Dey E. The role of components of bifidobacterium and lactobacillus in pathogenesis and serologic diagnosis of autoimmune thyroid diseases. *Benef Microbes* (2011) 2(2):139–54. doi: 10.3920/bm2010.0011

18. Lauritano EC, Bilotta AL, Gabrielli M, Scarpellini E, Lupascu A, Laginestra A, et al. Association between hypothyroidism and small intestinal bacterial overgrowth. J Clin Endocrinol Metab (2007) 92(11):4180–4. doi: 10.1210/jc.2007-0606

19. Didelez V, Sheehan N. Mendelian randomization as an instrumental variable approach to causal inference. *Stat Methods Med Res* (2007) 16(4):309–30. doi: 10.1177/0962280206077743

20. Lawlor DA, Harbord RM, Sterne JA, Timpson N, Davey Smith G. Mendelian randomization: using genes as instruments for making causal inferences in epidemiology. *Stat Med* (2008) 27(8):1133–63. doi: 10.1002/sim.3034

21. Kurilshikov A, Medina-Gomez C, Bacigalupe R, Radjabzadeh D, Wang J, Demirkan A, et al. Large-scale association analyses identify host factors influencing human gut microbiome composition. *Nat Genet* (2021) 53(2):156–65. doi: 10.1038/s41588-020-00763-1

22. Long Y, Tang L, Zhou Y, Zhao S, Zhu H. Causal relationship between gut microbiota and cancers: A two-sample mendelian randomisation study. *BMC Med* (2023) 21(1):66. doi: 10.1186/s12916-023-02761-6

23. Xu Q, Ni JJ, Han BX, Yan SS, Wei XT, Feng GJ, et al. Causal relationship between gut microbiota and autoimmune diseases: A two-sample Mendelian randomization study. *Front Immunol* (2021) 12:746998. doi: 10.3389/fimmu.2021.746998

24. Cao J, Wang N, Luo Y, Ma C, Chen Z, Chenzhao C, et al. A cause-effect relationship between Graves' disease and the gut microbiome contributes to the thyroid-gut axis: A bidirectional two-sample Mendelian randomization study. *Front Immunol* (2023) 14:977587. doi: 10.3389/fmmu.2023.977587

25. Davey Smith G, Hemani G. Mendelian randomization: genetic anchors for causal inference in epidemiological studies. *Hum Mol Genet* (2014) 23(R1):R89–98. doi: 10.1093/hmg/ddu328

26. Kurki MI, Karjalainen J, Palta P, Sipilä TP, Kristiansson K, Donner KM, et al. Finngen provides genetic insights from a well-phenotyped isolated population. *Nature* (2023) 613(7944):508–18. doi: 10.1038/s41586-022-05473-8

27. Sun BB, Maranville JC, Peters JE, Stacey D, Staley JR, Blackshaw J, et al. Genomic atlas of the human plasma proteome. *Nature* (2018) 558(7708):73–9. doi: 10.1038/ s41586-018-0175-2

28. Medici M, Porcu E, Pistis G, Teumer A, Brown SJ, Jensen RA, et al. Identification of novel genetic loci associated with thyroid peroxidase antibodies and clinical thyroid disease. *PloS Genet* (2014) 10(2):e1004123. doi: 10.1371/journal.pgen.1004123

29. Burgess S, Butterworth A, Thompson SG. Mendelian randomization analysis with multiple genetic variants using summarized data. *Genet Epidemiol* (2013) 37 (7):658–65. doi: 10.1002/gepi.21758

30. Hartwig FP, Davey Smith G, Bowden J. Robust inference in summary data Mendelian randomization via the zero modal pleiotropy assumption. *Int J Epidemiol* (2017) 46(6):1985–98. doi: 10.1093/ije/dyx102

31. Bowden J, Davey Smith G, Burgess S. Mendelian randomization with invalid instruments: effect estimation and bias detection through Egger regression. *Int J Epidemiol* (2015) 44(2):512–25. doi: 10.1093/ije/dyv080

32. Bowden J, Davey Smith G, Haycock PC, Burgess S. Consistent estimation in Mendelian randomization with some invalid instruments using a weighted median estimator. *Genet Epidemiol* (2016) 40(4):304–14. doi: 10.1002/gepi.21965

33. Burgess S, Dudbridge F, Thompson SG. Combining information on multiple instrumental variables in Mendelian randomization: comparison of allele score and summarized data methods. *Stat Med* (2016) 35(11):1880–906. doi: 10.1002/sim.6835

34. Burgess S, Thompson SG. Erratum to: interpreting findings from Mendelian randomization using the Mr-Egger method. *Eur J Epidemiol* (2017) 32(5):391–2. doi: 10.1007/s10654-017-0276-5

35. Hemani G, Zheng J, Elsworth B, Wade KH, Haberland V, Baird D, et al. The Mr-Base platform supports systematic causal inference across the human phenome. *Elife* (2018) 7:e34408. doi: 10.7554/eLife.34408

36. Zhang Y, Zhang X, Chen D, Lu J, Gong Q, Fang J, et al. Causal associations between gut microbiome and cardiovascular disease: A Mendelian randomization study. *Front Cardiovasc Med* (2022) 9:971376. doi: 10.3389/fcvm.2022.971376

37. Storey JD, Tibshirani R. Statistical significance for genomewide studies. *Proc Natl Acad Sci U.S.A.* (2003) 100(16):9440–5. doi: 10.1073/pnas.1530509100

38. Bowden J, Holmes MV. Meta-analysis and Mendelian randomization: A review. *Res Synth Methods* (2019) 10(4):486–96. doi: 10.1002/jrsm.1346

39. Verbanck M, Chen CY, Neale B, Do R. Detection of widespread horizontal pleiotropy in causal relationships inferred from Mendelian randomization between complex traits and diseases. *Nat Genet* (2018) 50(5):693–8. doi: 10.1038/s41588-018-0099-7

40. Ejtahed HS, Angoorani P, Soroush AR, Siadat SD, Shirzad N, Hasani-Ranjbar S, et al. Our little friends with big roles: alterations of the gut microbiota in thyroid disorders. *Endocr Metab Immune Disord Drug Targets* (2020) 20(3):344–50. doi: 10.2174/1871530319666190930110605

41. Ragusa F, Fallahi P, Elia G, Gonnella D, Paparo SR, Giusti C, et al. Hashimotos' Thyroiditis: epidemiology, pathogenesis, clinic and therapy. *Best Pract Res Clin Endocrinol Metab* (2019) 33(6):101367. doi: 10.1016/j.beem.2019.101367

42. Chow J, Lee SM, Shen Y, Khosravi A, Mazmanian SK. Host-bacterial symbiosis in health and disease. *Adv Immunol* (2010) 107:243–74. doi: 10.1016/b978-0-12-381300-8.00008-3

43. Suzuki K, Kawamoto S, Maruya M, Fagarasan S. Galt: organization and dynamics leading to iga synthesis. *Adv Immunol* (2010) 107:153–85. doi: 10.1016/ b978-0-12-381300-8.00006-x

44. Woting A, Blaut M. The intestinal microbiota in metabolic disease. *Nutrients* (2016) 8(4):202. doi: 10.3390/nu8040202

45. Liu S, An Y, Cao B, Sun R, Ke J, Zhao D. The composition of gut microbiota in patients bearing hashimoto's thyroiditis with euthyroidism and hypothyroidism. *Int J Endocrinol* (2020) 2020:5036959. doi: 10.1155/2020/5036959

46. Ishaq HM, Mohammad IS, Guo H, Shahzad M, Hou YJ, Ma C, et al. Molecular estimation of alteration in intestinal microbial composition in Hashimoto's thyroiditis patients. *BioMed Pharmacother* (2017) 95:865–74. doi: 10.1016/j.biopha.2017.08.101

47. Lu G, Yu X, Jiang W, Luo Q, Tong J, Fan S, et al. Alterations of gut microbiome and metabolite profiles associated with anabatic lipid dysmetabolism in thyroid cancer. *Front Endocrinol (Lausanne)* (2022) 13:893164. doi: 10.3389/fendo.2022.893164

48. Zhao F, Feng J, Li J, Zhao L, Liu Y, Chen H, et al. Alterations of the gut microbiota in Hashimoto's thyroiditis patients. *Thyroid* (2018) 28(2):175-86. doi: 10.1089/thy.2017.0395

49. Biscarini F, Masetti G, Muller I, Verhasselt HL, Covelli D, Colucci G, et al. Gut microbiome associated with Graves' disease and Graves' orbitopathy: the indigo* Multicentre European study. *J Clin Endocrinol Metab* (2023) 108(8):2065–77. doi: 10.1210/ clinem/dgad030

50. Lin B, Zhao F, Liu Y, Wu X, Feng J, Jin X, et al. Randomized clinical trial: probiotics alleviated oral-gut microbiota dysbiosis and thyroid hormone withdrawalrelated complications in thyroid cancer patients before radioiodine therapy following thyroidectomy. *Front Endocrinol (Lausanne)* (2022) 13:834674. doi: 10.3389/ fendo.2022.834674

51. Butel MJ. Probiotics, gut microbiota and health. *Med Mal Infect* (2014) 44(1):1–8. doi: 10.1016/j.medmal.2013.10.002

52. Karl JP, Margolis LM, Madslien EH, Murphy NE, Castellani JW, Gundersen Y, et al. Changes in intestinal microbiota composition and metabolism coincide with increased intestinal permeability in young adults under prolonged physiological stress. *Am J Physiol Gastrointest Liver Physiol* (2017) 312(6):G559–g71. doi: 10.1152/ajpgi.00066.2017

 $53.\,$ Wu M, Yang Y, Fan Y, Guo S, Li T, Gu M, et al. Characteristics of the intestinal flora of tpoab-positive women with subclinical hypothyroidism in the second trimester

of pregnancy: A single-center prospective cohort study. Front Cell Infect Microbiol (2022) 12:794170. doi: 10.3389/fcimb.2022.794170

54. Wu M, Chi C, Yang Y, Guo S, Li T, Gu M, et al. Dynamics of gut microbiota during pregnancy in women with tpoab-positive subclinical hypothyroidism: A prospective cohort study. *BMC Pregnancy Childbirth* (2022) 22(1):592. doi: 10.1186/ s12884-022-04923-5

55. Thompson RS, Gaffney M, Hopkins S, Kelley T, Gonzalez A, Bowers SJ, et al. Ruminiclostridium 5, parabacteroides distasonis, and bile acid profile are modulated by prebiotic diet and associate with facilitated sleep/clock realignment after chronic disruption of rhythms. *Brain Behav Immun* (2021) 97:150–66. doi: 10.1016/j.bbi.2021.07.006

56. Zhang B, Chen T, Cao M, Yuan C, Reiter RJ, Zhao Z, et al. Gut microbiota dysbiosis induced by decreasing endogenous melatonin mediates the pathogenesis of Alzheimer's disease and obesity. *Front Immunol* (2022) 13:900132. doi: 10.3389/fimmu.2022.900132

57. van Niftrik L, Devos DP. Editorial: planctomycetes-verrucomicrobia-chlamydiae bacterial superphylum: new model organisms for evolutionary cell biology. *Front Microbiol* (2017) 8:1458. doi: 10.3389/fmicb.2017.01458

58. Zhang T, Li Q, Cheng L, Buch H, Zhang F. Akkermansia muciniphila is a promising probiotic. *Microb Biotechnol* (2019) 12(6):1109-25. doi: 10.1111/1751-7915.13410