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Floral nectar: fifty years of new ecological perspectives beyond pollinator reward

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Abstract

Floral nectar is central to ecology, since it mediates interactions with pollinators, flower-visiting antagonists and microbes through its chemical composition. Here we review how historical assumptions about its ecological meaning were first challenged, then modified and expanded since the discovery of secondary metabolites in nectar. We then explore the origin of specific neuroactive nectar compounds known to act as important insect neurotransmitters, and how advances in the field of bee cognition and plant-microbe-animal interactions challenge such historical views. As all actors involved in the latter interactions are under simultaneous reciprocal selective pressures, their coexistence is characterized by conflicts and trade-offs, the evolutionary interpretation of which suggests exciting new perspectives in one of the longest studied aspects of plant-pollinator interactions.

Graphic abstract



Keywords: nectar biogenic amines, plant-microbe-pollinator interactions, manipulation hypothesis, pollinator fidelity hypothesis, nectar ecology

Fifty years of ecological perspectives

The scientific definition of nectar, first given by Linnaeus in 1735, sees nectar as the secretion of specific organs, the nectaries. Specifically, the secretion of *floral* nectar is associated with the plant reproductive structures (Nepi 2017), and rewards animals that may perform pollination while visiting the flower (Nepi et al. 2018). Whether pollinator attraction was the primary driver leading to nectar appearance is a difficult question to answer (e.g. Sprengel 1793, Caspary 1848, Bonnier 1878). Nevertheless, its centrality in mediating plant-animal interaction is nowadays undeniable, and was somehow recognized as early as the first century BC, when the poet Virgil (Georgics part IV, 149-227) used the term "nectar" to refer to the substance that honeybees collect from the fields and store in combs as honey.

By virtue of its carbohydrate and amino acid content, nectar has been considered a readily absorbable, cost-effective, alimentary reward offered by plants in exchange for the pollination service mediated by animals (Nicolson 2007, González-Teuber and Heil 2009, Heil 2011). Up to 90% of its dry weight consists of sugars (Lüttge 1977), while the other 10% includes a plethora of other components such as amino acids, inorganic ions, proteins, lipids and organic acids (Nicolson and Thornburg 2007). Amino acids and inorganic ions are the most abundant classes after that of sugars (e.g. Lüttge 1961; Mostowska 1965; Göttlinger et al. 2019). Though much less abundant than in pollen, the role of amino acids in enhancing nectar nutritional value (e.g. Jervis and Boggs 2005), contributing to its taste and attractiveness (e.g. Gardener and Gillman 2002, Carter et al. 2006), and affecting the foraging choices of pollinators (e.g. Seo et al. 2019) is well known. A historical listing of European

scientists who reported amino acids in nectar between the 1950s and the 1970s is given by Baker and Baker (1975). The inorganic ion content of nectar, on the contrary, is often overlooked, in spite of the fact that minerals and ions may play a crucial role in maintaining salt balance in nectar feeding animals (e.g. Hiebert and Calder 1983, Nicolson and Fleming 2003).

If discussion is limited to these main classes of nectar chemicals, the ecological significance of nectar could easily be confined to its being an alimentary reward involved exclusively in pollinator nutrition and attraction. However, since the early 1970s the discovery of a series of nectar chemicals (Barberis et al. 2023a and references therein) not involved in primary metabolic pathways (Baker and Baker 1977, 1986), and therefore considered secondary metabolites (Pichersky and Gang 2000), has challenged this traditional view. Back then, most advances in nectar chemistry sprang from the pioneering work of Baker and Baker, who were also the first to speculate on the function of these so-called "unfavourable substances" (Baker and Baker 1975). They postulated what is nowadays known as the "pollinator fidelity" hypothesis, which states that secondary metabolites in nectar discourage flower-inconstant insects from visiting the flowers, while favouring specialist visitors (Adler 2000). This hypothesis relies on the assumption that specialists are more effective pollinators than generalists and deliver more intraspecific pollen (Rhoades and Bergdahl 1981, Adler 2000 and references therein). Since the 1970s, the main traditional alimentary function of floral nectar has therefore been flanked by a second crucial role suddenly recognized by scholars: that of discouraging those nectar consumers who contribute little or nothing to the pollination service.

This hypothesis was later expanded by Janzen (1977) and Baker (1978), who proposed that nectar secondary metabolites might deter nectar thieves, such as ants. This further interpretation, which many researchers agree to consider an extension of the "pollinator fidelity" hypothesis, was later called the "nectar robber" hypothesis (Adler 2000). The results of Stephenson (1982) showed that the floral nectar of *Catalpa speciosa*, which contains iridoid glycosides, is only moderately consumed by nectar thieves, despite its abundance and accessibility. Shortly after consuming the nectar, thieves

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appear intoxicated and behave abnormally (Stephenson 1981), whereas insects acting as legitimate pollinators do not show any effect after ingestion. While some studies seem to confirm the hypothesis, others show that in most cases the deterrent effect against ants is conferred rather by mechanical adaptations (Feinsinger and Swarm 1978, Schubart and Anderson 1978, Guerrant and Fiedler 1981).

In line with the idea that secondary metabolites prevent nectar exploitation by inefficient floral visitors, it was then postulated that they may prevent nectar waste by microbes (e.g. Hagler and Buchmann 1993, Verpoorte and Schripsema 1994). Indeed, by virtue of its sugar and amino acid composition, nectar has long been recognized as a potentially rich medium for microbial growth (e.g. Boutroux 1884, Schuster and Úlehla 1913, Grüss 1917, Schoellhorn 1919). Curiously, nectar proteins – compounds nowadays known to protect nectar against proliferation of microorganisms and plant tissues against infection by pathogens (Carter et al. 2007, Hillwig et al. 2010, Nepi 2017 and references therein) – were discovered more than 90 years ago (Buxbaum 1927), but were initially thought to be nutritional, supplying nectar consumers with organic nitrogen (Lüttge 1961, Heil 2011). Thus the so-called "antimicrobial" hypothesis (Adler 2000) received more support some years later, when more experiments conducted in this direction provided evidence of it. For example, Montenegro et al. (2012) described that honey obtained from *Quillaja saponaria* exerts biological action against pathogens associated with the phenols found in the floral nectar of the plant, suggesting an antimicrobial effect of these compounds.

Initially considered "unfavourable" (Baker and Baker 1975), nectar secondary metabolites were often later regarded as toxic. With reference to nectar non-protein amino acids, Baker and Baker were again the first to suggest that some nectar secondary metabolites could be toxic to certain flower-visitors (1977). In the same decade, a series of studies demonstrated the potential toxicity of several secondary metabolites for animal consumers in plant-herbivore interactions. Such toxicity was described to have deleterious post-ingestive effects on growth (e.g. Blau et al. 1978, Isman and Duffey 1982), organ function (e.g. Berenbaum 1988) and nutrient uptake (e.g. Slansky 1992). These findings led to

reconsideration of the early assumption that the benefits of nectar secondary metabolites must outweigh their cost, and the idea of direct selection of nectar toxins was momentarily obscured by the conjecture that their presence was due to prior selection pressures or pleiotropic constraints and that there were no adaptive functions in relation to pollination, or only after defense (Adler 2000, Stevenson et al. 2017). In this pleiotropic perspective, nectar chemistry is originally determined by co-evolutionary interactions with herbivores, and the occurrence of "unfavourable" substances in floral nectar is accidental and, all things considered, a small cost to pay in exchange for defence of the plant against the major threat of herbivory. Though it cannot be excluded, this theory does not account for the significant differences in chemical composition often observed between nectar and phloem sap (Roy et al. 2017). This consideration is just one of several aspects that gradually advanced the formulation of new hypotheses.

Since the studies conducted to assess the toxicity of the secondary metabolites found in nectar were performed with concentrations equal to those found in vegetative tissues (generally greater than those found in nectar, e.g. Adler et al. 2006, Wiese et al. 2018), a new interpretation of the ecological role of these secondary metabolites emerged in the 1980s. This new interpretation was based on the hypothesis that nectar-like concentrations of these potentially toxic secondary metabolites could indeed prove toxic for insect pathogens but not for their presumably less susceptible hosts. A series of pioneering studies conducted in this direction gave rise to the conviction that their ingestion by nectar consumers may improve the health and life expectancy of the latter (e.g. Price et al. 1980, Berenbaum 1988). This new perspective attributing curative benefits to secondary metabolites revives the concept that the presence of these compounds in nectar must outweigh the costs associated with their consumption.

In support of this latter view, it is now well established that the putative toxicity of certain secondary metabolites in nectar often depends on the sensitivity of the nectar consumer (e.g. Tiedeken et al. 2016), or may be greater for introduced species that are not native pollinators of the plants in question (e.g. Zhang et al. 2022). Moreover, as stated above, nectar concentrations are generally lower than

those found in other plant tissues (e.g. Cook et al. 2013, Palmer-Young et al. 2019), and the combined effects of two or more compounds may enhance or erase their separate effects (e.g. Muth et al. 2022). In general, pollinators may therefore benefit from consuming nectars rich in secondary metabolites which may reduce their pathogen loads, enhance their immune response or even enrich their gut microbiota (Gunasekaran et al. 2020, Baracchi et al. 2022), in line with what may be called the "medication" hypothesis. A growing number of recent studies supports this view. For example, nectar alkaloids such as gelsemine, anabasine, and nicotine benefit pollinators by increasing their resistance to parasites and pathogens (Manson et al. 2010, Richardson et al. 2015, Thornburn et al. 2015), and the idea that bees may actively seek alkaloid-enriched nectar to keep pathogens at bay (Gherman et al. 2014) has become popular. At least until recently, such active search beaviour has been explained by homeostasis, an impulse to seek a certain compound occurring when levels of the compound are low in the animal's its body (Samorini 2013). The fact that consumption of a potentially curative compound does not produce immediate healing induced Samorini (2013) to reject the idea that ingestion of curative substances is ruled exclusively by homeostasis, but rather suggests some degree of "awareness" or "intention".

Nowadays, it has finally been established that vertebrates and invertebrates self-medicate, and a growing number of studies provide evidence of this (Hutchings et al. 2003, de Roode et al. 2013, Abbott 2014, and references therein). Self-medication implies that the exposure of healthy animals to secondary metabolites has a cost, compensated by its beneficial effects in reducing symptoms or clearing infections (Clayton and Wolfe 1993, Lozano 1998, Abbott 2014 and references therein). For example, Singer et al. (2009) found that when parasitized caterpillars of the woolly bear moth *Gramnia incorrupta* ingest plant pyrrolizidine alkaloids, it improves their survival by conferring resistance to tachinid flies, a lethal endoparasite. On the contrary, when unparasitized caterpillars consume excessive amounts of these toxins, it reduces their survival, in line with the theory.

To be fully in line with the key criteria defining self-medication, an animal must however also modify its dietary preferences, addressing its foraging to a source containing "nonnutritive" antimicrobial

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compounds when parasitized (Karban and English-Loeb 1997). Again, taking the study case of Singer et al. (2009) as example, parasitized are more likely than unparasitized caterpillars to ingest large amounts of pyrrolizidine alkaloids, indicating that infection indeed induces a change in their feeding behaviour.

Some of the secondary metabolites found in nectar share structural similarity with important neurotransmitters (Verpoorte 2005); this observation supports the hypothesis that their presence in nectar outweighs any costs associated with their consumption. As early as the 1970s, the idea that compounds such as alkaloids, glycosides and phenols could have a significant effect on the central nervous system of flower visitors had already been suggested (Baker and Baker 1975, 1977). If a certain chemical can modulate neuron signal transduction, the concentrations of neurotransmitters and the activity or expression of their receptors may vary, radically changing animal behaviour (Wink 2018). When ingestion of nectar secondary metabolites has pharmacological effects on the brain of nectar consumers, these substances have sometimes been considered drugs. For example, Wright et al. (2013), considered the nectar alkaloid caffeine in their study in this manner, providing the first evidence of its capacity to alter a pollinator's memory of reward pharmacologically. Indeed, honeybees treated with caffeine were shown to be three times more likely to remember a learned floral scent than those rewarded with sucrose alone.

It is a complex task to define a certain substance as a drug. Historically, drugs have been referred to as "nervous foods" (Mantegazza 1871), bringing two aspects into focus: i) they often interfere with animal nervous systems at various levels, and ii) it is difficult to draw a distinct line between food and drugs. Various criteria can be used for this definition. For example, the sharpening of specific senses or the onset of addiction (Samorini 2013), the only aspect commonly shared being changes in animal behaviour (Wink 2018). Addiction can manifest in various ways, but generally implies that the consumer craves the chemical once its serum levels drop (Wink 2018). Often it also implies consumption despite adverse consequences and perceptual changes in reward strength (Koob 2015,

Fattore and Diana 2016). The key drivers of addiction are reflected in altered expressions of motivation and learning, capacities that emerged early in the Precambrian (Menzel and Benjamin 2013), so that recent views frame addiction as a phenomenon with deep evolutionary roots, wide spread among invertebrates (van Staaden et al. 2018).

Cases in which consumption of nectar secondary metabolites implies a cost for the nectar-feeding animal – contrary to the instinct of conservation – were initially considered incidental, but observation of repeated feeding on inebriating sources raised the question of how "intentional" the ingestion of these compounds is. The hawkmoth *Manduca quinquemaculata*, for example, feeds on nectar of *Datura meteloides*, a plant belonging to the family Solanaceae, the nectar of which probably contains the hallucinogenic substances also found in the plant's other tissues (Grant 1983). These compounds intoxicate insects, making them sluggish and disoriented (Grant 1983). When moths remain on the ground, they are more exposed to predation (Grant 1983). Bees exposed to hallucinogenic or narcotic substances offered by orchids such as *Epipactis helleborine* are a similar case (Jakubska et al. 2005). An ecological explanation could be that inebriating compounds in floral nectar may enhance the chance of pollination by detaining the animals (Jakubska et al. 2005).

A recent study by Galpayage Dona et al. (2022) provided first evidence that bumblebees engage in activities not directly aimed at satisfying a primary need. Despite of the absence of external incentives, bees repeatedly engaged in rolling wooden balls, suggesting that this activity – fully ascribed to play – is rewarding in itself, an aspect in line with the criteria defining play. This finding, along with a series of other studies, marks a breakthrough in the field of insect behaviour, since it provides additional evidence of the existence of a form of sentience in bumblebees (e.g. Bateson 2014, Held and Špinka 2011, Solvi et al. 2016, Birch 2020). This in turn raises the question of whether the search for hallucinogenic/inebriating substances may also be rewarding in itself, and whether returning to such nectar sources may be dictated exclusively by the insurgence of physical dependence.

Other coercive mechanisms not necessarily implying intoxication are known, for instance that of offering nectar containing nicotine: after experiencing such nectar, bees keep returning to the food source even when it becomes suboptimal compared to other available rewards (e.g. Baracchi et al. 2017). In line with the prediction of Rhoades and Bergdahl (1981), detaining the pollinator in this case may increase mobilization of conspecific pollen, promoting the plant's reproductive fitness.

The above examples suggest potential harmful behaviour of floral visitors as a consequence of nectar ingestion. Such cases frame the concept of pollinator manipulation, a term that researchers have used since the early 2000s (e.g. Biernaskie and Cartar 2004, Bayleis et al. 2007), and which gained full recognition after formal introduction of the "manipulation" hypothesis by Pyke (2016), later structured by Nepi et al. (2018). However, to be fair, with reference to various nectar secondary metabolites, as early as 1981 Rhoades and Bergdahl wrote: "though at first sight the presence of these toxic substances seems incompatible with the reward function of nectar, they probably represent a mechanism to manipulate pollinator behaviour to the advantage of the plant and to exclude nectar thieves". They guessed that a combination of rewarding and defensive chemicals could model insect visiting patterns to favor plant fitness beyond the pollinator fidelity hypothesis.

Along with those secondary metabolites that have strong biological activities due to their structural relationship with animal neurotransmitters (Verpoorte 2005), an additional case concerns nectar chemicals that are environmental sources of invertebrate neurotransmitters *per se*. This is the case of biogenic amines (Roeder 1999, Blenau and Baumann 2001, Scheiner et al. 2006, Farooqui 2012), a class of compounds only recently reported in floral nectar (Muth et al. 2022, Barberis et al. 2023b, Barberis et al. 2023c). The two biogenic amines reported in floral nectar to date are tyramine and octopamine, the invertebrate counterparts of the vertebrate adrenergic transmitters, ruling the so-called fight or flight response, which is to say rapid adaptation to energy-demanding situations (Roeder 2005). They are decarboxylation products of the amino acid tyrosine and although tyramine is the biological precursor of octopamine, the twoare considered to act as independent

neurotransmitters (Roeder 2005). Their consumption can modulate a plethora of behavioural traits such as motivation (e.g. Farooqui 2012), reward-seeking (e.g. Schulz and Robinson 2001, Peng et al. 2020), locomotion (e.g. Fussnecker et al. 2006, Hardie et al. 2007), learning (e.g. Mercer and Menzel 1982, Hammer and Menzel 1998) and social communication (e.g. Barron et al. 2007, Linn et al. 2020).

Regarding their effects on food-source communication and exploitation, for example, octopamine was demonstrated to increase the likelihood of honeybee dancing (Barron et al. 2007), while lowering the probability that bees heed social information from other foragers (Linn et al. 2020). This means that even if the food source is poor, bees are more likely to retain their personal information than to heed indications of a richer source. This evidence supports the hypothesis that nectar octopamine increases bee faithfulness to a plant species favouring the latter's reproductive success. Octopamine was also demonstrated to regulate foraging behaviour in honeybee colonies, inducing a change in the type of food source to which foragers direct their collection activity, with a trend towards directing their efforts to less valuable resources (Giray et al. 2003). This phenomenon may be due to the fact that consumption of octopamine and tyramine lowers the sucrose concentration necessary to elicit the proboscis extension reflex; in other words it increases bee perception of the value of a food source (e.g. Pankiw and Page 2003, Mc Cabe et al. 2017). Regarding locomotion, an exploratory study investigating the effect of nectar-like concentrations of tyramine on bumblebee flower visits showed that bees fed tyramine-enriched solutions spent less time foraging on a single flower than those fed control solution, suggesting that their behaviour was on the whole more dynamic (Barberis et al. 2023b).

Beyond pollinator reward: the role of microbial interaction

So far, nectar-mediated interactions have been described as a bipartite phenomenon between plants and floral visitors, whereas the way secondary metabolites wind up in nectar has been neglected. This aspect is in fact still largely unclear (Heil 2011 and references therein). Along with the discovery of

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nectar secondary metabolites back in the 1970s, this question has resurfaced. One of the main hypotheses explaining the presence of secondary metabolites in floral nectar stated that nectaries secrete almost unmodified substances that flow directly or indirectly via the vascular tissues by passive diffusion (Lüttge 1977, Fahn 1988). Today, several examples have shown that the chemical composition of nectar is usually quite different from that of phloem (e.g. Bertazzini and Forlani 2016), and different mechanisms of fine-tuned nectar secretion have been described (e.g. Radhika et al. 2010, Lin et al. 2014). However, most of the models of floral nectar secretion, for example the so-called apoplastic, merocrine and eccrine models, focus mainly on the alternative processes of secretion of the nectar sugar component (Roy et al. 2017 and references therein), while the specific mechanisms ruling transport and secretion of other metabolites are still largely unknown.

Beyond this aspect of the knowledge gap, it is now well established that the chemical composition of floral nectar may not only be shaped by phylogenetic constraints but also by ecological drivers (e.g. Nepi et al. 2010, Bogo et al. 2021). Among these it is worth mentioning, for example, interactions with specific guilds of pollinators that may drive selection towards convergent nectar chemistry in unrelated taxa (e.g. Pozo et al. 2015), or interactions with different habitat types (at least in species with wide ecological ranges) (e.g. Farkas et al. 2012) and the influence of human-driven landscape changes such as urbanization, habitat fragmentation and land use (e.g. Tew et al. 2021, Biella et al. 2022). As habitat type and landscape can impart specific local microclimatic characteristics and influence animal communities, both can extensively affect nectar availability and chemistry, not only at the secretion stage, but also through post-secretion modifications, presumably influenced by meteorological conditions (e.g. Corbet et al. 1979, Plowright 1981, Chalcoff et al. 2017, Parachnowitsch et al. 2019) and interaction with floral visitors (e.g. Bogo et al. 2021).

Nowadays, animal visitors are recognized as the principal vectors of bacteria, fungi and other microorganisms to and between flowers (e.g. Brysch-Herzberg 2004, Herrera et al. 2010, Belisle et al. 2012). However, in many cases, not even when flowers have just opened can they be considered sterile (Aleklett et al. 2014 and references therein). Even before the bud opens, floral nectar often

contains bacteria and fungi (e.g. Shade et al. 2013, von Arx et al. 2019), the abundance of which increases over time in individual flowers (e.g. Pusey et al. 2009, von Arx et al. 2019, Morris et al. 2020). Besides bacteria and fungi commonly found in air, soil and other habitats – generally the first to be detected when the flower opens (e.g. Brysch-Herzberg 2004, Morris et al. 2020) – another commonly found group of microbes is that of flower specialists, which exhibit a range of traits that may be adaptations to nectar environments (e.g. Dhami et al. 2016, Herrera et al. 2010, Pozo and Jacquemyn 2019). Moreover, irrespective of continent or habitat type, microbial colonization was recently demonstrated to occur more frequently than previously believed. For example, Herrera et al. (2009) conducted a quantitative survey to assess how frequently floral nectar contains yeasts and found them in up to 44% of samples, while even greater percentages were obtained by other authors (see Brysch-Herzberg 2004 and references therein for a list of studies).

On colonization, microbes may therefore modify plant-provisioned nectar chemicals or impart their own by secreting metabolic by-products into the nectar (e.g. Canto and Herrera 2012, Vannette and Fukami 2018, Yang et al. 2019, Vannette and Fukami 2016, Rering et al. 2020). During sugar fermentation, for instance, different volatile organic compounds are released, and additional compounds can be added to the floral olfactory bouquet (Rering et al. 2018). Even in the case mentioned above, that of *Epipactis helleborine*, one of the potentially hallucinogenic/narcotic compounds offered in its floral nectar is ethanol (Løjtnant 1974, Müller 1988), which is thought to be of microbial origin (Ehlers and Olsen 1997, Kevan et al. 1998).

The influence of microbes on floral nectar is mainly regarded as detrimental for its quality (e.g. Eisikowitch et al. 1990, Herrera et al. 2008, Vannette et al. 2013), weakening or negatively interfering with plant-pollinator mutualism. For example, some studies have demonstrated that yeasts reduce the food value of floral nectar by decreasing sugar (Canto et al. 2011, de Vega and Herrera 2013) and amino acid concentrations (Pozo et al. 2014). In general, floral microbes are rarely believed to benefit plants (Vannette 2020 and references therein). Flower pathogens and some nectar bacteria can reduce plant fitness, either directly or by decreasing pollinator visits (e.g. Vannette et al. 2013). Other studies,

however, have demonstrated that in certain cases microorganisms may enhance pollination by producing volatiles that play a role in attracting pollinators, indirectly influencing plant fitness (e.g. Pozo et al. 2009, Herrera and Pozo 2010, Cullen et al. 2021). However, even in cases where nectar yeasts increase pollinator visits, this does not necessarily benefit plant fitness. For example, Herrera et al. (2013a) reported reduced seed set in yeast-colonized *Helleborus foetidus* plants despite increased pollinator attraction.

In addition, microbes in nectar not only alter pollinator attraction and visitation through volatile emissions or chemical modification (e.g. Raguso 2004, Rering et al. 2018, 2020), but their very presence seems to drive a preference for yeast-containing flowers in pollinators such as bumblebees, who were demonstrated to detect them in nectar (Herrera et al. 2013b, Schaeffer et al. 2014, Schaeffer et al. 2017). In this regard, it is suggested that nectar yeast cells supplement insects with important nutritional elements such as vitamins, sterols, and minerals (Vega and Dowd 2005, Stefanini 2018). Dharampal et al. (2019) was the first study in this direction and provided evidence of the benefits for honeybee larvae of the diverse communities of symbiotic microbes inhabiting the pollen surface. If pollen microbes are a crucial dietary resource for larval development, it is also likely that the microbial inhabitants of floral nectar are an important nutritional component as well (Jacquemyn et al. 2021). This view is in line with emerging evidence that the nectar microbiome, like that of pollen, may influence the health of pollinators (sensu López-Uribe et al. 2020) by modifying their nutritional landscape, altering foraging behaviour, and interacting with their symbionts and pathogens (Martin et al. 2022 and references therein). As consumption of nectar and pollen colonized by consortia of yeasts and bacteria (Pozo et al. 2012, Ambika Manirajan et al. 2016) may provide a regular supplement of the microorganisms that are part of a bee's gastrointestinal flora, it may also affect consumer fitness, and – in the case of social insects – colony development (Dharampal et al. 2019, Pozo et al. 2018, 2020). In their experiment, Pozo et al. (2020) found that yeasts in the bumblebee diet positively affected colony growth of Bombus terrestris, though the intensity of the effect depended on the yeast species. Another study by Pozo et al. (2021) showed that feeding bacterial

supplements to colonies of *Bombus terrestris* led to faster egg laying, greater brood size and increased production of workers, while combined yeasts and bacteria supplements induced less evident benefits. Examples such as the above provide preliminary evidence in support of the hypothesis that nectar microbes can benefit pollinator health. Further studies are needed to clarify certain aspects. For example, Pozo et al. (2020) also demonstrated that yeasts grown *in vitro* could suppress growth of the bumble bee gut pathogen *Crithidia bombi*; whether the same action is also exerted in the host gut remains to be assessed.

Since biogenic amines can be generated by microbial decarboxylation of free amino acids, it has been suggested that their presence in floral nectar could be imputed to yeast metabolism rather than to that of the plant (Nepi 2017, Nepi et al. 2018). To date, however, we still lack evidence supporting this hypothesis, so the conservative explanation that they are plant by-products stands. By virtue of endogenous enzyme production, tyramine, for instance, can be found in various plant parts or their derivatives (Vazquez y Novo et al. 1989, Preti et al. 2016, Gobbi et al. 2019), seeming ubiquitous and implicated in a number of metabolic pathways of which tyramine – precursor of many other pharmacologically active compounds – is the first product (Facchini et al. 2000). As tyramine can be the product of specific pathways activated in response to attack by various plant enemies (Servillo et al. 2017), the production of biogenic amines may be a general defensive response against pathogens or phytophages (Facchini et al. 2002, Macoy et al. 2015, Knolleberg et al. 2020, Shen et al. 2021, Plonka et al. 2022). In fermented foods and beverages of plant origin, however, its presence is associated with microbes with aminogenic activity (Trivedi et al. 2009). In addition, some biogenic amine-producing microbes found in wine have also been found in floral nectar (Landete et al. 2007, Pozo et al. 2016).

Knowledge gaps

Today the chemical complexity of nectar is well established, despite the fact that its composition was long assumed to be a constant trait of a species. This assumption encouraged a search for patterns, and justified pooling nectar samples when volumes were insufficient for analyses (Nicolson 2022). For decades, this approach masked the variability of nectar, its complex physiology and dependence on the environment, all of which make its study extremely challenging (Brandenburg et al. 2009 and references therein). Explanations of the ecological role of nectar in mediating plant-animal interactions are therefore less certain due to variable chemical expression (Stevenson et al. 2017). More insights into the molecular and genetic mechanisms ruling its secretion and composition are therefore needed.

A second level of complexity is represented by recent findings concerning animal cognition. Flower visiting involves perception, memory, expectation, and decision making (Waddington 2001), all tools known to be influenced by emotional states, at least in humans (e.g. Mathews and MacLeod 1994, Lerner and Keltner 2000). In recent years, the scientific community seems to have recognized the existence of emotions in vertebrates such as fish and birds (e.g. Rey et al. 2015, Valance et al. 2008), but also in invertebrates such as insects, for example bees and flies, turn out to fulfil the basic requirements of emotional behaviour (Baracchi et al. 2017 and references therein), as well as showing a form of sentience (Galpayage Dona et al. 2022). Several studies have established that insects have high levels of cognitive sophistication (e.g. Avarguès-Weber et al. 2011, Collett et al. 2013, Giurfa 2013, Klein et al. 2017). These important breakthroughs challenge the way we have been tackling the subject of how floral visitors exploit their floral nectar-landscape (Baracchi 2019 and references therein). Increasing evidence that insects self-medicate and engage in rewarding activities beyond their primary needs – for pleasure, one would say – is encouraging research in this direction.

Although the importance of having more information on wild pollinators for the purpose of their conservation has been acknowledged (Pegoraro et al. 2020), the effects of nectar secondary metabolites on the great majority of wild pollinators are largely unknown. In the case of wild bees, this is probably a consequence of our limited understanding of how to establish and maintain their

nests in laboratory conditions (Leonard and Harmon-Threatt 2019). So far, research has focused mainly on managed honeybees, bumblebees, and hummingbirds (e.g. Muth et al. and references therein, Stevenson et al. 2017 and references therein, Kessler et al. 2012), despite the fact that pollinators differ vastly in life cycle, sociality, dietary specialization (Muth et al. 2017) and other characteristics. For example, even a simple response, such as eliciting the proboscis extension reflex under laboratory conditions, seems profoundly influenced by the sociality of the bee species (Vorel and Pitts-Singer 2010). This stresses the importance (when possible) of coupling laboratory work with the study of wild pollinators under natural or semi-natural conditions, also in the light of recent reports that experiments conducted under controlled conditions may not always yield a realistic picture of animal behaviour (e.g. Mujagic and Erber 2009, Ayestaran et al. 2010).

It is even more important to fill up the knowledge gap concerning the synergic effects of the complex combinations of chemicals found in nectar on pollinator behaviour. Although foliar chemical ecology has highlighted the importance of synergistic effects (Richards et al. 2016) and recent studies have demonstrated that these can lead to unpredicted behaviours (e.g. Muth et al. 2022), nectar chemistry studies on the field of nectar chemistry generally involve the use of one substance at a time (e.g. Wright et al. 2013, Baracchi et al. 2017, Estravis-Barcala et al. 2021, Hernández et al. 2018, Marchi et al. 2021, Richman et al. 2022, Thorburn et al. 2015).

A better understanding of how nectar-like concentrations of combined co-occurring secondary metabolites affect animal behaviour is a prerequisite for assessing how human-induced dispersal of chemicals in the environment may affect plant-pollinator interactions. How nectar secondary metabolites interact with phytochemicals is also still largely unknown, but some first studies have shown that even a single acute exposure to a pesticide can reshape the interactions between plants and floral visitors mediated by nectar secondary metabolites (Richman et al. 2022). This highlights the importance of using realistic concentrations of chemicals, similar to those found in natural nectar.

So far pollinators have been regarded as the main source of selection leading to establishment of given concentrations of secondary metabolites in floral nectar (Stevenson et al. 2017), and current research has outlined how these can also be affected by floral microbes (e.g. McArt et al. 2014, Parachnowitsch et al. 2018, Rebolleda-Gomez et al. 2019, Rivest and Forrest 2020). Besides circumventing plant defensive mechanisms – such as high concentrations of reactive oxygen species (Thornburg et al. 2003) or proteins with antimicrobial properties (Schmitt et al. 2021 and references therein) – nectar specialized microbes need to colonize new spaces to maintain their populations (Morris et al. 2020), as the flowers where they live generally have short lifespans (e.g. Primack 1985). To do this, it has been suggested that microbes may affect flower traits and influence their own dispersal (Russell et al. 2019, Vannette 2020, Francis et al. 2021 and references therein). However, a part from a few examples like that of the fungal pathogen *Fusarium moniliforme*, which enhances bird visitation for spore dispersal (Lara and Ornelas 2003), there is little evidence that microbial species rely on floral visitors to maintain their populations. This means that further research is needed to verify what we may call the "manipulation-for-dispersal" hypothesis.

Finally, although floral microbes are rarely believed to benefit plants, a few cases of plants with adaptations to promote microbial growth in their flowers are known. For example, Wiens et al. (2008) suggest that the palm *Eugeissona tristis* may encourage the growth of ethanol-producing yeasts by selecting mammal pollinators that consume fermented nectar while discouraging less specialized ones. Though still an untested hypothesis, if floral microbes could enhance plant fitness by promoting compounds such as exogenous insect neurotransmitters in nectar, then further research should examine the potential for plant-chemical adaptation to facilitate microbe colonization of nectar. In other words: may plants show chemical adaptations of nectar that promote microbial settlement in the flower that optimizes pollinator attraction?

Concluding remarks

The recently established role of microorganisms as third partners in nectar-mediated, plant-animal interactions adds considerable complexity to our attempts to elucidate the ecological functions of floral nectar (Stevenson et al. 2017, Nepi 2017). All actors involved in plant-microbe-pollinator interactions are under simultaneous reciprocal selective pressures (Figure 1). Plants must ensure pollinator visitants and protect their floral alimentary resources against microbial exploitation, while flower-specialized microbes employ nectivores to disperse between hostplants. Such conflicts and trade-offs, coupled with recent advances in chemical ecology and bioinformatics, open many exciting avenues for research in one of the longest studied aspects of plant-pollinator interactions.



Figure 1. Network of complex nectar-mediated plant-pollinator-microbe relationships.

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References

- Abbott J (2014). Self-medication in insects: current evidence and future perspectives. Ecol Entom. 39, 273–280. https://doi.org/10.1111/een.12110
- Adler LS (2000). The ecological significance of toxic nectar. Oikos. 91, 409–420. https://doi.org/10.1034/j.1600-0706.2000.910301.x
- Adler LS, Wink M, Distl M, Lentz A (2006). Leaf herbivory and nutrients increase nectar alkaloids. Ecol Lett. 9(8), 960–967. https://doi.org/10.1111/j.1461-0248.2006.00944.x
- Aleklett K, Hart M, Shade A (2014). The microbial ecology of flowers: an emerging frontier in phyllosphere research. Botany. 92, 253–266. https://dx.doi.org/10.1139/cjb-2013-0166
- Álvarez-Pérez S, Herrera CM, de Vega C (2012). Zooming-in on floral nectar: a first exploration of nectar-associated bacteria in wild plant communities. FEMS Microbiol Ecol. 80(3), 591–602. https://doi.org/10.1111/j.1574-6941.2012.01329.x
- Ambika Manirajan B, Ratering S, Rusch V, Schwiertz A, Geissler-Plaum R, Cardinale M, Schnell S (2016). Bacterial microbiota associated with flower pollen is influenced by pollination type, and shows a high degree of diversity and species-specificity. Environ. 18(12), 5161–5174. https://doi.org/10.1111/1462-2920.13524
- Ayestaran A, Giurfa M, de Brito Sanchez MG (2010). Toxic but drank: gustatory aversive compounds induce post-ingestional malaise in harnessed honeybees. PLoS ONE. 5, e15000. https://doi.org/10.1371/journal.pone.0015000
- Avarguès-Weber A, Deisig N, Giurfa M (2011). Visual cognition in social insects. Annu Rev Entomol. 56, 423–443. https://doi.org/10.1146/annurev-ento-120709-144855
- Bayleis S, Hargreaves A, Hechtenthal SD, Laird R, Latty T, Reid TG, Teucher A, Tindall J (2007). Empty flowers as a pollination-enhancement strategy. Evol Ecol Res. 9, 1245–1262.
- Baker HG (1978). Chemical aspects of the pollination of woody plants in the tropics. In:Tomlinson PB, Zimmerman M (eds.). Tropical trees as living systems. Cambridge UniversityPress, New York. pp 57–82.

Baker HG, Baker I (1975). The study of nectar-constitution and pollinator plant coevolution.In: Gilbert LE, Raven PH (eds), Coevolution of plants and animals. University of Texas Press, Austin. pp. 100–140.

- Baker HG, Baker I (1977). Intraspecific constancy of floral nectar amino acid complements. Bot. Gaz. 138, 183–191. https://doi.org/10.1086/336914
- Baker HG, Baker I (1986). The occurrence and significance of amino acids in floral nectars. Plant Syst Evol. 151, 175–186. https://doi.org/10.1007/BF02430273
- Baracchi D (2019). Cognitive ecology of pollinators and the main determinants of foraging plasticity. Curr Zool. 65(4), 421–424. https://doi.org/10.1093/cz/zoz036
- Baracchi D, Brown MFJ, Chittka L (2015). Weak and contradictory effects of self-medication with nectar nicotine by parasitized bumblebees. F100 Research. 4, 73. https://doi.org/10.12688/f1000research.6262.1
- Baracchi D, Marples A, Jenkins AJ, Leitch AR, Chittka L (2017). Nicotine in floral nectar pharmagologically influences bumblebee learning of floral features. Sci Rep. https://doi.org/10.1038/s41598-017-01980-1
- Baracchi D, Lihoreau M, Giurfa M (2017). Do insects have emotions? Some insights from bumble bees. Front Behav Neurosci. 11, 157. https://doi.org/10.3389/fnbeh.2017.00157
- Baracchi D, Brown MJ, Chittka L (2022). Behavioural evidence for self-medication in bumblebees? F1000Research. 4, 73. https://doi.org/10.12688/f1000research.6262.3
- Barberis M, Calabrese D, Galloni M, Nepi M (2023a). Secondary metabolites in nectar-mediated plant-pollinator relationships. Plants. 12(3), 550. https://doi.org/10.3390/plants12030550
- Barberis M, Bogo G, Bortolotti L, Guarnieri M, Nepi M, Felicioli A, Galloni M (2023b). Nectar tyramine decreases the duration of bumblebee visits on flowers. Arthropod-Plant Interactions. 17, 563–569. http://doi.org/10.1007/s11829-023-09976-7

Barberis M, Bogo G, Bortolotti L, Flaminio S, Giordano E, Nepi M, Galloni M (2023c). Nectar

chemistry and insect flower handling time change over the flowering season: results from an exploratory study. Acta Oecol. 120, 103937. https://doi.org/10.1016/j.acta.2023.103937

- Barron AB, Maleszka R, Vander Meer RK, Robinson GE (2007). Octopamine modulates honey bee dance behavior. PNAS. 104(5), 1703–1707. https://doi.org/10.1073/pnas.0610506104
- Bateson P (2014). Play, playfulness, creativity and innovation. Animal Behavior and Cognition, 2(2), 99. https://doi.org/10.12966/abc.05.02.2014
- Belisle M, Peay KG, Fukami T (2012). Flowers as islands: spatial distribution of nectar-inhabiting microfungi among plants of *Mimulus aurantiacus*, a hummingbird-pollinated shrub. Microb Ecol. 63, 711–718. https://doi.org/10.1007/s00248-011-9975-8
- Bell DS (1995). The motivation of addiction. Acta Neurochir. 132, 185–191. https://doi.org/10.1007/BF01809345
- Berenbaum MR (1988). Allelochemicals in insect-microbe-plant interactions; agents provocateurs in the coevolutionary arms race. Wiley, New York.
- Bertazzini M, Forlani G (2016). Intraspecific variability of floral nectar volume and composition in rapeseed (*Brassica napus* L. var. *oleifera*). Front Plant Sci. 7, 288. https://doi.org/10.3389/fpls.2016.00288
- Beutler R (1935). Nectar. Bee World. 24, 106–162.
- Biella P, Tommasi N, Guzzetti L, Pioltelli E, Labra M, Galimberti A (2022). City climate and landscape structure shape pollinators, nectar and transported pollen along a gradient of urbanization. J Appl Ecol. 59, 1586–1595. https://doi.org/10.1111/1365-2664.14168
- Biernaskie and Cartar (2004). Variation in rate of nectar production depends on floral display size: a pollinator manipulation hypothesis. Func Ecol. 18(1), 125–129. https://doi.org/10.1111/j.1365-2435.2004.00815.x

Birch J (2020). The search for invertebrate consciousness. Nous. 56(1), 133–153. https://doi.org/10.1111/nous.12351

Blau PA, Feeny P, Contardo L, Robson DS (1978). Allyglucosinolate and herbivorous caterpillars -

contrast in toxicity and tolerance. Science. 200, 1296-

1298.https://doi.org/10.1126/science.200.4347.1296

- Blenau W, Baumann A (2001). Molecular and pharmacological properties of insect biogenic amine receptors: lessons from *Drosophila melanogaster* and *Apis mellifera*. Arch Insect Biochem Physiol. 48, 13–38. https://doi.org/10.1002/arch.1055
- Bonnier MG (1878). Ètude sur l'anatomie et al physiologie des nectaries. Bulletin de la Société Botanique de France. 25(8). 262–271. https://doi.org/10.1080/00378941.1878.10827912
- Boutroux L (1884). Sur la conservation des ferments alcooliques dans la nature. Ann Sci Nat Sér IV Bot. 17, 145–209.
- Brandenburg A, Dell'Olivo A, Bshary R, Kuhlemeier C (2009). The sweetest thing: advances in nectar research. Curr Opin Plant Biol. 12, 486–490. https://doi.org/10.1016/j.pbi.2009.04.002
- Brysch-Herzberg M (2004). Ecology of yeasts in plant-bumblebee mutualism in Central Europe. FEMS Microbiol Ecol. 50, 87–100. https://doi.org/10.1016/j.femsec.2004.06.003
- Buxbaum F (1927). Zur Frage des Eiweißgehaltes des Nektars. Planta (Berlin) 4, 818–821. https://doi.org/10.1007/BF01911267
- Canto A, Herrera CM, Garcia IM, Pérez R, Vaz M (2011). Intraplant variation in nectar traits in *Helleborus foetidus* (Ranunculaceae) as related to floral phase, environmental conditions and pollinator exposure. Flora. 206, 668–675. https://doi.org/10.1016/j.flora.2011.02.003
- Canto A, Herrera CM (2012). Micro-organisms behind the pollination scenes: microbial imprint on floral nectar sugar variation in a tropical plant community. Ann Bot. 110, 1173–1183. https://doi.org/10.1093/aob/mcs183
- Carlson J, Harms KE (2006) The evolution of gender-biased nectar production in hermaphrodite plants. Bot Rev. 72, 179–205. https://doi.org/10.1663/0006-8101(2006)72[179:TEOGNP]2.0.CO;2

- Carter C, Sharoni S, Yehonatan L, Palmer RG, Thornburg R (2006). A novel role for proline in plant floral nectars. Naturwissenschaften. 93, 72–79. https://doi.org/10.1007/s00114-005-0062-1
- Carter C, Healy R, O'Tool NM, Naqvi SSM, Ren G, Park S, Beattie GA, Horner HT, Thornburg RW (2007). Tobacco nectaries express a novel NADPH oxidase implicated in the defense of floral reproductive tissues against microorganisms. Plant Physiol. 143, 389–399. https://doi.org/10.1104/pp.106.089326

Caspary R de (1848). De nectariis. Elberfeld

- Chalcoff VR, Gleiser G, Ezcurra C, Aizen MA (2017). Pollinator type and secondarily climate are related to nectar sugar composition across the angiosperms. Evol Ecol. 31, 585–602. https://doi.org/10.1007/ s10682-017-9887-2
- Clayton DH, Wolfe ND (1993). The adaptive significance of self-medication. Trends Ecol Evol. 8, 60–63. https://doi.org/10.1016/0169-5347(93)90160-Q
- Collett M, Chittka L, Collett TS (2013). Spatial memory in insect navigation. Curr Biol. 23, R789-R800. https://doi.org/10.1016/j.cub.2013.07.020
- Cook D, Manson JS, Gardner DR, Welch KD, Irwin RE (2013). Norditerpene alkaloid concentrations in tissues and floral rewards of larkspurs and impacts on pollinators. Biochem Syst Ecol. 48, 123–131. https://doi.org/10.1016/j.bse.2012.11.015
- Corbet SA, Willmer PG, Beament JWL, Unwin DM, Prŷs-Jones OE (1979). Post-secretory determinants of sugar concentration in nectar. Plant Cell Environ. 2, 293– 308. https://doi.org/10.1111/j.1365-3040.1979.tb00084.x
- Cullen N, Fetters A, Ashman TL (2021). Integrating microbes into pollination. Curr Opin Insect Sci. 44, 48–54. https://doi.org/10.1016/j.cois.2020.11.002
- Dhami MK, Hartwig T, Fukami T (2016). Genetic basis of priority effects: insights from nectar yeasts. Proc R Soc B. 238, 20161455. https://doi.org/10.1098/rspb.2016.1455
- Dharampal PS, Carlson C, Currie CR, Steffan SA (2019). Pollen-borne microbes shape bee fitness. Proc Royal Soc B. 286, 20182894. https://doi.org/10.1098/rspb.2018.2894

- de Vega C, Herrera CM (2013). Microorganisms transported by ants induce changes in floral nectar composition of an ant-pollinated plant. Am J Bot. 100(4), 792-800. https://doi.org/10.3732/ajb.1200626
- de Vega C, Albaladejo RG, Álvarez-Pérez S, Herrera CM (2022). Contrasting effects of nectar yeasts on the reproduction of Mediterranean plant species. Am J Bot. 109, 393–405. https://doi.org/10.1002/ajb2.1834
- Eisikowitch D, Kevan PG, Lachance MA (1990). The nectar-inhabiting yeasts and their effect on pollen germination in common milkweed, *Asclepias syriaca* L. Israel J Bot. 39, 217–225. https://doi.org/10.1080/0021213X.1990.10677145
- Ehlers BK, Olesen JM (1997). The fruit-wasp route to toxic nectar in *Epipactis* orchids? Flora. 192, 223–229. https://doi.org/10.1016/S0367-2530(17)30787-9
- Estravis-Barcala MC, Palottini F, Farina WM (2021). Learning of a mimic odor combined with nectar nonsugar compounds enhances honeybee pollination of a commercial crop. Sci Rep. 11, 23918. https://doi.org/10.1038/s41598-021-03305-9
- Facchini PJ, Huber-Allanach KL, Tari LW (2000). Plant aromatic L-amino acid decarboxylases: evolution, biochemistry, regulation, and metabolic engineering applications. Phytochem. 54, 121–138 https://doi.org/10.1016/s0031-9422(00)00050-9
- Facchini PJ, Hagel JM, Zulak KG (2002). Hydroxycinnamic acid amide metabolism: physiology and biochemistry. Can J Bot. 80, 577–589. https://doi.org/10.1139/b02-065
- Fahnn A (1988). Secretory tissues in vascular plants. New Phytol. 108, 229–258. https://doi.org/10.1111/j.1469-8137.1988.tb04159.x
- Farkas A, Molnár R, Morschhauser T, Hahn I (2012). Variation in nectar volume and sugar concentration of *Allium ursinum* L. spp. *ucrainicum* in three habitats. Sci World J. 2012(3), 138579. https://doi.org/10.1100/2012/138579
- Farooqui T (2012). Review of octopamine in insect nervous systems. Open Access Insect Physiol. 4, 1–17. https://doi.org/10.2147/OAIP.S20911

- Fattore L, Diana M (2016).Drug addiction: an affective cognitive disorder in need of a cure. Neuroscienceand Biobehavioral Reviews. 65, 341–361. https://doi.org/10.1016/j.neurobiorev.2016.04.006
- Feinsinger P, Swarm LA (1978). How common are ant-repellent nectars? Biotropica. 10, 238. https://doi.org/10.2307/2387911

Francis JS, Tatarko AR, Richman SK, Vaudo AD, Leonard AS (2021). Microbes and pollinator behavior in the floral marketplace. Curr Opin Insect Sci. 44, 16–22. https://doi.org/10.1016/j.cois.2020.10.003

- Fussnecker BL, Smith BH, Mustard JA (2006). Octopamine and tyramine influence the behavioural profile of locomotor activity in the honey bee (*Apis mellifera*). J Insect Physiol. 52(10), 1083– 1092. https://doi.org/10.1016/j.jinsphys.2006.07.008
- Galpayage Dona HS, Solvi C, Kowalewska A, Makela K, MaBouDi HD, Chittka L (2022). Do bumble bees play? https://doi.org/10.1016/j.anbehav.2022.08.013
- Gardener M, Gillman MP (2002). The taste of nectar a neglected area of pollination ecology. Oikos. 98(3), 552–557. https://doi.org/10.1034/j.1600-0706.2002.980322.x
- Gherman BI, Denner A, Bobiş O, Dezmirean DS, Mărghitaş LA, Schlüens H, Moritz RFA, Erler S (2014). Pathogen-associated self-medication behavior in the honeybee *Apis mellifera*. Behav Ecol Sociobiol. 68, 1777-1784. https://doi.org/10.1007/s00265-014-1786-8
- Giray T, Galindo-Cardona A, Oskay D (2003). Octopamine influences honey bee foraging preference. J Insect Phys. 53, 691–698. https://doi.org/10.1016/j.jinsphys.2007.03.016
- Giurfa M (2013). Cognition with few neurons: higher-order learning in insects. Trends Neurosci. 36, 285–294. https://doi.org/10.1016/j.tins.2012.12.011
- Gobbi L, Ciano S, Rapa M, Ruggieri R (2019). Biogenic amines determination in "Plant Milks". Beverages. 5(2), 40. https://doi.org/10.3390/beverages5020040

González-Teuber M, Heil M (2009). Nectar chemistry is tailored for both attraction of

mutualists and protection from exploiters. Plant Signal Behav. 4, 809-813.

https://doi.org/10.4161/psb.4.9.9393

- Göttlinger T, Schwerdtfeger M, Tiedge K, Lohaus G (2019). What do nectarivorous bats like? Nectar composition in Bromeliaceae with special emphasis on bat-pollinated species. Front Plant Sci. 10. https://doi.org/10.3389/fpls.2019.00205
- Gowan E, Lewis BA, Turgeon R (1995). Phloem transport of antirrhinoside, an iridoid glycoside, in *Asarina scandens* (Scrophulariaceae). J Chem Ecol. 21, 1781–1788. https://doi.org/10.1007/BF02033676
- Grant V (1983). Behavior of hawkmoths on flowers of *Datura meteloides*. Bot Gaz. 144, 280–284. https://doi.org/10.1086/337374
- Grüss J (1917). Die anpassung eines pilzes (*Anthomyces reukaufii*) an den blütenbau und den bienenrüssel. Berichte der Deutschen Botanischen Gesellschaft. 35, 746–761.
- Guerrant EO, Fiedler P (1981). Flower defenses against nectar-pilferage by ants. Biotropica. 13(2), 25–33. https://doi.org/10.2307/2388067
- Gunasekaran M, Trabelcy B, Izhaki I, Halpern M (2021). Direct evidence that sunbirds' gut microbiota degrades floral nectar's toxic alkaloids. 12, 639808. https://doi.org/10.3389/fmicb.2021.639808
- Hagler JR, Buchmann SL (1993). Honey bee (Hymenoptera: Apidae) foraging responses to phenolic-rich nectars. J Kansas Entomol Soc. 66, 223–230. https://doi.org/10.1093/jee/96.5.1510
- Hammer M, Menzel R (1998). Multiple sites of associative odor learning as revealed by local brain microinjections of octopamine in honeybees. Learn Mem. 5, 146–156.
- Hardie SL, Zhang JX, Hirsh J (2007). Trace amines differentially regulate adult locomotor activity, cocaine sensitivity, and female fertility in Drosophila melanogaster. Dev Neurobiol. 67(10), 1396–1405. https://doi.org/10.1002/dneu.20459

- Heil M (2011). Nectar: generation, regulation and ecological functions. Trends Plant Sci. 16(4), 191–200. https://doi.org/10.1016/j.tplants.2011.01.003
- Held SDE, Špinka M (2011). Animal play and animal welfare. Animal Behaviour, 81(5), 891e899. https://doi.org/10.1016/j.anbehav.2011.01.007
- Hernández IG, Palottini F, Macri I, Galmarini CR, Farina WM (2018). Appetitive behavior of the honey bee *Apis mellifera* L. in response to phenolic compounds naturally found in nectars. J Exp Biol. https://doi.org/10.1242/jeb.189910
- Herrera CM, de Vega C, Canto A, Pozo MI (2009). Yeasts in floral nectar: a quantitative survey. Ann Bot. 103, 1415–1423. http://doi.org/10.1093/aob/mcp026
- Herrera CM, Canto A, Pozo MI, Bazaga P (2010). Inhospitable sweetness: nectar filtering of pollinator-borne inocula leads to impoverished, physiologically clustered yeast communities. Proc Royal Soc London B Biol Sci. 277, 747–754. https://doi.org/10.1098/rspb.2009.1485
- Herrera CM, Pozo MI (2010). Nectar yeasts warm the flowers of a winter blooming plant. Proc Royal Soc B. 277, 1827–1834. https://doi.org/10.1098/rspb.2009.2252
- Herrera CM, Pozo MI, Medrano M (2013a). Yeasts in nectar of an early-blooming herb: sought by bumble bees, detrimental to plant fecundity. Ecology. 94, 273–279. https://doi.org/10.1890/12-0595.1
- Herrera CM, Pozo MI, Medrano M (2013b). Bumble bees prefer nectar containing yeasts. Bulletin of the Ecological Society of America. 94, 186–189. https://doi.org/10.1890/0012-9623-94.2.186
- Hiebert SM, Calder WA Jr (1983). Sodium, potassium, and chloride in floral nectars: energy-free contributions to refractive index and salt balance. Ecology. 64(2), 399–402. https://doi.org/10.2307/1937086
- Hillwig MS, Liu X, Liu G, Thornburg RW, MacIntosh GC (2010). *Petunia* nectar proteins have ribonuclease activity. J Experim Bot. 61, 2951–2965. https://doi.org/10.1093/jxb/erq119

- Hutchings MR, Athanasiadou S, Kyriazakis I, Gordon IJ (2003). Can animals use foraging behaviour to combat parasites? Proc Nutr Soc. 62, 361–370.
- Isman MB, Duffey SS (1982). Toxicity of tomato phenolic compounds to the fruitworm *Heliothis zea*. Entomol Exp Appl. 31, 370–376. https://doi.org/10.1111/j.1570-7458.1982.tb03162.x
- Jacquemyn H, Pozo MI, Álvarez-Pèrez S, Lievens B, Fukami T (2021). Yeast-nectar interactions: metacommunities and effects on pollinators. Curr Opin Insect Sci. 44, 35–40. https://doi.org/10.1016/j.cois.2020.09.014
- Jakubska A, Przado D, Steininger M, Aniol-Kwiatkowska J, Kadej M (2005). Why do pollinators become "sluggish"? Nectar chemical constituents from *Epipactis helleborine* (L.) Crantz (Orchidaceae). Applied Ecol Environ Research. 3(2), 29–38. https://doi.org/10.15666/aeer/0302_029038
- Janzen DH (1977). Why don't ants visit flowers? Biotropica 9, 252–1252. https://doi.org/10.2307/2388142
- Jervis MA, Boggs CL (2005). Linking nectar amino acids to fitness in female butterflies. Trends Ecol Evol. 20(11), 585–587. https://doi.org/10.1016/j.tree.2005.08.015
- Karban R, English-Loeb G (1997). Tachinid parasitoids affect host plant choice by caterpillars to
- increase caterpillar survival. Ecology. 78, 603-611. https://doi.org/10.1890/0012-
- 9658(1997)078[0603:TPAHPC]2.0.CO;2
- Kessler D, Bhattacharya S, Diezel C, Rothe E, Gase K, Schöttner M, Baldwin IT (2012). Unpredictability of nectar nicotine promotes outcrossing by hummingbirds in *Nicotiana attenuate*. Plant J. 71, 529–538. https://doi.org/10.1111/j.1365-313X.2012.05008.x
- Kevan PG, Eisikowitsch D, Fowle S, Thomas K (1998). Yeast-contaminated nectar and its effects on bee foraging. J Apicult Res. 27, 26–29.

https://doi.org/10.1080/00218839.1988.11100777

Koob GF (2015). The dark side of emotion: the addiction perspective. Eur J Pharmacol. 753, 73–87. 15(753), 73–87. https://doi.org/10.1016/j.ejphar.2014.11.044

- Knolleberg BJ, Li G-X, Lambert JD, Maximova SN, Guiltinan MJ (2020). Clocamide, a hydroxycinnamic acid amide, is a resistance factor against *Phytophthora spp*. in *Theobroma cacao*. Front Plant Sci. 11, 617520. https://doi.org/10.3389/fpls.2020.617520
- Klein S, Cabirol A, Devaud JM, Barron AB, Lihoreau M (2017). Why bees are so vulnerable to environmental stressors. Trends Ecol Evol. 32, 268–278. https://doi.org/10.1016/j.tree.2016.12.009
- Landete JM, de Las Rivas B, Marcobal A, Muñoz R (2007). Molecular methods for the detection of biogenic amine-producing bacteria on foods. Int J Food Microbiol. 117(3), 258–269. https://doi.org/10.1016/j.ijfoodmicro.2007.05.001
- Lara C, Ornelas JF (2003). Hummingbirds as vectors of fungal spores in *Moussonia deppeana* (Gesneriaceae): taking advantage of ephemeral flowers of a mutualism? Am J Bot. 90, 262–269. https://doi.org/10.3732/ajb.90.2.262
- Lefèvre T, Chiang A, Kelavkar M, Li H, Li J, de Castillejo CLF, Oliver L, Potini Y, Hunter MD, de Roode JC (2012). Behavioural

resistance against a protozoan parasite in the monarch butterfly. J Animal Ecol. 81, 70–79. https://doi.org/10.1111/j.1365-2656.2011.01901.x

- Lerner JS, Keltner D (2000). Beyond valence: toward a model of emotion-specific influences on judgement and choice. Cogn Emot. 14, 473–493. https://doi.org/10.1080/026999300402763
- Leonard RJ, Harmon-Threatt AN (2019). Methods for rearing ground-nesting bees under laboratory conditions. Apidologie. 50(5), 689–703. https://doi.org/10.1007/s13592-019-00679-8
- Linn M, Glaser SM, Peng T, Grüter C (2020). Octopamine and dopamine mediate waggle dance following and information use in honeybees. Proc R Soc B. 287, 20201950. https://doi.org/10.1098/rspb.2020.1950

Linnaeus C (1735). Nectarium, pars mellifica floripropria. Syst naturae et Phil bot.

Løjtnant B (1974). Toxic nectar, "drunken" wasps and orchids. Kaskelot 15, 3-7.

- López-Uribe M, Ricigliano V, Simone-Finstrom M (2020). Defining pollinator health: assessing bee ecological, genetic, and physiological factors at the individual, colony, and population levels. Annu Rev Anim Biosci. 8, 269–294. https://doi.org/10.1146/annurev-animal-020518-115045
- Lozano GA (1998). Parasitic stress and self-medication in wild animals. Adv Study Behav. 27, 291–317. https://doi.org/10.1016/S0065-3454(08)60367-8
- Lüttge U (1961). Über die Zusammensetzung des Nektars und den Mechanismus seiner Sekretion. I. Planta. 56, 189–212. https://doi.org/10.1007/BF01914720
- Lüttge U (1977). Nectar composition and membrane transport of sugars and amino acids: a review on the present state of nectar research. Apidologie. 8, 305–319. https://doi.org/10.1051/APIDO:19770402
- Macoy DM, Kim W-Y, Lee SY, Kim MG (2015). Biotic Stress Related Functions of Hydroxycinnamic Acid Amide in Plants. J Plant Biol. 58, 156–163. https://doi.org/10.1007/s12374-015-0104-y
- Manson JS, Otterstatter MC, Thomson JD (2010). Consumption of a nectar alkaloid reduces pathogen load in bumble bees. Oecologia. 162, 81–89. https://doi.org/10.1007/s00442-009-1431-9
- Manson JS, Cook D, Gardner DR, Irwin RE (2013). Dose-dependent effects of nectar alkaloids in a montane plant-pollinator community. J Ecol. 101(6), 1604–1612. https://doi.org/10.1111/1365-2745.12144

Mantegazza P (1871). Quadri della natura umana. Feste ed ebbrezze, vol. II. Brigola, Milano.

Marchi IL, Palottini F, Farina WM (2021). Combined secondary compounds naturally found in nectars enhance honeybee cognition and survival. J Exp Biol. 224, jeb239616. https://doi.org/10.1242/jeb.239616

- Martin VN, Schaeffer RN, Fukami T (2022). Potential effects of nectar microbes on pollinator health. Phil Trans R Soc B. 377, 20210155. https://doi.org/10.1098/rstb.2021.0155
- Mathews A, MacLeod C (1994). Cognitive approaches to emotion and emotional disorders. Annu Rev Psychol. 45, 25–50. 10.1146/annurev.ps.45.020194.000325
- McArt SH, Koch H, Irwin RE, Adler LS (2014). Arranging the bouquet of disease: floral traits and the transmission of plant and animal pathogens. Ecol Lett. 17(5), 624–636. https://doi.org/10.1111/ele.12257
- Mc Cabe SIM, Ferro MWB, Farina WM, Hrncir M (2017). Dose- and time-dependent effects of oral octopamine treatments on the sucrose responsiveness in stingless bees (*Melipona scutellaris*).
 Apidologie. 48, 204–210. https://doi.org/10.1007/s13592-016-0442-x

Menzel R, Benjamin PR (2013). Invertebrate learning and memory. Amsterdam: Elsevier Science.

- Mercer AR, Menzel R (1982). The effect of biogenic amines on conditioned and unconditioned responses to olfactory stimuli in the honeybee *Apis mellifera*. J Comp Physiol A. 145, 363– 368. https://doi.org/10.1007/BF00619340
- Merritt SZ (1996). Within-plant variation in concentrations of amino acids, sugar, and sinigrin in phloem sap of black mustard, *Brassica nigra* (L.) Koch (Cruciferae). J Chem Ecol. 22, 1133–1145. https://doi.org/10.1007/BF02027950
- Lin IW, Sosso D, Chen LQ, Gase K, Kim SG, Kessler D, Klinkenberg PM, Gorder MK, Hou BH, Qu XQ, Carter CJ, Baldwin IT, Frommer WB (2014). Nectar secretion requires sucrose phosphate synthases and the sugar transporter SWEET9. Nature. 508, 546–549. https://doi.org/10.1038/nature13082
- Montenegro G, Dìaz-Forestier J, Fredes C, Rodriguez S (2013). Phenolic profiles of nectar and honey of *Quillaja saaponaria* Mol. (Quillajaceae) as potential chemical markers. 46(2), 177–182. https://doi.org/10.4067/S0716-97602013000200009

Morris M, Frixione N, Burkert A, Dinsdale E, Vannette RL (2020). Microbial abundance,

composition, and function in nectar are shaped by flower visitor identity. FEMS Microbiol Ecol. 96, fiaa003. https://doi.org/10.1093/femsec/fiaa003

- Mostowska I (1965). Aminokwasy w nektarach i jednogatunkowych miodach. Zesz. Nauk. Wyzsz. Szk. Roln. Olsztyn. 20(3), 417–432.
- Mujagic S, Erber J (2009). Sucrose acceptance, discrimination and proboscis responses of honey bees (*Apis mellifera* L.) in the field and the laboratory. J Comp Physiol A, 195, 325– 339. https://doi.org/10.1007/s00359-008-0409-0
- Müller I (1988). Vergleichende blütenökologische untersuchungen an der orchideen gattung *Epipactis*. Mitt. Bl. AHO Baden-Württemberg. 20, 701–803.
- Mustard JA (2020). Neuroactive nectar: compounds in nectar that interact with neurons. Arthropod Plant Interact. 14, 151–159. https://doi.org/10.1007/s11829-020-09743-y
- Muth F, Philbin CS, Jeffrey CS, Leonard AS (2022). Discovery of octopamine and tyramine in nectar and their effects on bumblebee behavior. *ISCIENCE*. https://doi.org/10.1016/j.isci.2022.104765
- Nepi M (2017). New perspectives in nectar evolution and ecology: simple alimentary reward or a complex multiorganism interaction? Acta Agrobot. 70(1), 1704. https://doi.org/10.5586/aa.1704
- Nepi M, Selvi F, Pacini E (2010). Variation in nectar-sugar profile of *Anchusa* and allied genera (Boraginaceae). Bot J Linn Soc. 162(4), 616–627. https://doi.org/10.1111/j.1095-8339.2010.01036.x
- Nepi M, Grasso DA, Mancuso S (2018). Nectar in plant–insect mutualistic relationships: from food reward to partner manipulation. Front Plant Sci. 9, 1063. https://doi.org/10.3389/fpls.2018.01063
- Nicolson SW (2007). Nectar consumers. In: Nicolson SW, Nepi M, Pacini E (eds), Nectaries and nectar. Springer, Dordrecht, pp 289–342.

- Nicolson SW, Fleming PA (2003). Nectar as food for birds: the physiological consequences of drinking dilute sugar solutions. Plant Syst Evol. 238(1/4), 139–153. https://doi.org/10.1007/s00606-003-0276-7
- Nicolson SW, Thornburg RW (2007). Nectar chemistry. In: Nicolson SW, Nepi M, Pacini E (eds), Nectaries and nectar. Springer, Dordrecht, pp 215–264.
- Nicolson SW (2022). Sweet solutions: nectar chemistry and quality. Phil Trans R Soc B. 377, 20210163. https://doi.org/10.1098/rstb.2021.0163
- Palmer-Young EC, Farrell IW, Adler LS, Milano NJ, Egan PA, Junker RR, Irwin RE, Stevenson PC (2019). Chemistry of floral rewards: intra- and interspecific variability of nectar and pollen secondary metabolites across taxa. Ecol Monogr. 89(1), e01335. https://doi.org/10.1002/ecm.1335
- Pankiw T, Page RE Jr. (2003). Effect of pheromones, hormones, and handling on sucrose response thresholds of honey bees (*Apis mellifera* L.). J Comp Physiol A. 189, 675–684. https://doi.org/10.1007/s00359-003-0442-y
- Parachnowitsch AL, Manson JS, Sletvold N (2019). Evolutionary ecology of nectar. Ann Bot. 123, 247–261. https://doi.org/10.1093/aob/mcy132
- Pegoraro L, Hidalgo O, Leitch I, Pellicer J, Barlow S (2020). Automated video monitoring of insect pollinators in the field. Emerg Top Life Sci. 4(1), 87–97. https://doi.org/10.1042/ETLS20190074
- Peng T, Schroeder M, Grüter C (2020). Octopamine increases individual and collective foraging in a neotropical stingless bee. Biol Lett. 16, 20200238. http://dx.doi.org/10.1098/rsbl.2020.0238
- Pichersky E, Gang DR (2000). Genetics and biochemistry of secondary metabolites in plants: an evolutionary perspective. Trends Plant Sci. 5, 439–445
- Pyke GH (2016). Floral nectar: pollinator attraction or manipulation? Trends Ecol Evol. 31(5), 339– 341. https://doi.org/10.1016/j.tree.2016.02.013

- Płonka J, Barchańska H, Kokoszka K, Krzyżanowska A (2022). Effect of herbicide stress on the content of tyramine and its metabolites in japanese radish sprouts (*Raphanus sativus*). J Food Compos Anal. 106, 104301. https://doi.org/10.1016/j.jfca.2021.104301
- Plowright RC (1981). Nectar production in the boreal forest lily *Clatonia borealis*. Can J Bot. 59, 156–160. https://doi.org/10.1139/b81-025
- Preti R, Bernacchia R, Vinci G (2016). Chemometric evaluation of biogenic amines in commercial fruit juices. Eur Food Res Technol. 242, 2031-2039. https://doi.org/10.1007/s00217-016-2701-5
- Price PW, Bouton CE, Gross O, McPheron BA, Thompson JN, Weis AE (1980). Interactions among 3 trophic levels-influence of plants on interactions between insect herbivores and natural enemies. Annu Rev Ecol Syst. 11, 41–65. https://doi.org/10.1146/annurev.es.11.110180.000353
- Primack RB (1985). Longevity of individual flowers. Annu Rev Ecol Syst. 16, 15–37. https://doi.org/10.1146/annurev.es.16.110185.000311
- Pozo MI, de Vega C, Canto A, Herrera CM (2009). Presence of yeasts in floral nectar is consistent with the hypothesis of microbial- mediated signaling in plant-pollinator interactions. Plant Signal Behav. 4(11), 1102–1104. https://doi.org/10.4161/psb.4.11.9874
- Pozo MI, Lachance M-A, Herrera CM (2012). Nectar yeasts of two southern Spanish plants: the roles of immigration and physiological traits in community assembly. FEMS Microbiol Ecol. 80, 281–293. https://doi.org/10.1093/femsec/fiz090
- Pozo MI, Lievens B, Jacquemym H (2014). Impact of microorganisms on nectar chemistry, pollinator attraction and plant fitness. In: Peck RL (ed), Nectar: production, chemical composition and benefits to animals and plants. Nova Science Publishers: New York, NY, USA. pp. 1–40.

Pozo MI, Herrera C, Van der Ende W (2015). The impact of nectar chemical features on phenotypic variation in two related nectar yeasts. FEMS Microbiol Ecol. 91(6). https://doi.org/10.1093/femsec/fiv055

- Pozo MI, Herrera CM, Lachance MA, Verstrepen K, Lievens B, Jacquemyn H (2016) Species coexistence in simple microbial communities: unravelling the phenotypic landscape of cooccurring *Metschnikowia* species in floral nectar. Environ Microbiol. 18(6),1850–1862. https://doi.org/10.1111/1462-2920.13037
- Pozo MI, Van Kemenade G, Van Oystaeyen A, Aledòn-Català T, Benavente A, Wäckers F, Jacquemyn H (2020). The impact of the yeast presence in nectar on bumble bee behavior and fitness. Ecol Monogr. 90, e01393. https://doi.org/10.1002/ecm.1393
- Pozo MI, Mariën T, van Kemenade G, Wäckers F, Jacquemyn H (2021). Effects of pollen and nectar inoculation by yeasts, bacteria or both on bumblebee colony development. Oecologia. 195(3), 689–703. https://doi.org/10.1007/s00442-021-04872-4
- Pusey PL, Stockwell VO, Mazzola M (2009). Epiphytic bacteria and yeasts on apple blossoms and their potential as antagonists of *Erwinia amylovora* on apple flower stigmas. Phytopathology. 101, 1234–1241. https://doi.org/10.1094/PHYTO-99-5-0571
- Radhika V, Kost C, Boland W, Heil M (2010). The role of jasmonates in floral nectar secretion. PLoS One., e9265. https://doi.org/10.1371/journal.pone.0009265Raguso RA (2004). Why are some floral nectars scented? Ecology. 85(6), 1486–1494. https://doi.org/10.1890/03-0410
- Rebolleda-Gómez M, Forrester NJ, Russell AL, Wei N, Fetters AM, Stephens JD, Ashman TL (2019). Gazing into the anthosphere: considering how microbes influence floral evolution. New Physiol. 224(3), 1012–1020. https://doi.org/10.1111/nph.16137
- Rey S, Huntingford FA, Boltana S, Vargas R, Knowles TG, Mackenzie S (2015). Fish can show emotional fever: stress-induced hyperthermia in zebrafish. Proc R Soc B Biol Sci. 282:20152266. 10.1098/rspb.2015.2266

- Rering CC, Beck JJ, Hall GW, McCartney MM, Vannette RL (2018). Nectar-inhabiting microorganisms influence nectar volatile composition and attractiveness to a generalist pollinator. Nee Phytol. 220, 750–759. https://doi.org/10.1111/nph.14809
- Rering CC, Vannette RL, Schaeffer RN, Beck JJ (2020). Microbial co-occurrence in floral nectar affects metabolites and attractiveness to a generalist pollinator. J Chem Ecol. 46, 659–667.

Rhoades DF, Bergdahl JC (1981). Adaptive significance of toxic nectar. Am Nat. 117, 798–803.

- Richards LA, Glassmire AE, Ochsenrider KM, Smilanich AM, Dodson CD, Jeffrey CS, Dyer LA (2016). Phytochemical diversity and synergistic effects on herbivores. Phytochem Rev. 15, 1153–1166. https://doi.org/10.1007/s11101-016-9479-8
- Richardson LL, Adler LS, Leonard AS, Andicoechea J, Regan KH, Anthony WE, Manson JS, Irwin RE (2015). Secondary metabolites in floral nectar reduce parasite infections in bumblebees. Proc Biol Sci. 282(1803), 20142471. https://doi.org/10.1098/rspb.2014.2471
- Richman SK, Maalouf IM, Smilanich AM, Marquez Sanchez D, Miller SZ, Leonard AS (2022). A neonicotinoid pesticide alters how nectar chemistry affects bees. Func Ecol. 36, 1063–1073. https://doi.org/10.1111/1365-2435.14016
- Rivest S, Forrest JRK (2020). Defense compounds in pollen: why do they occur and how do they affect the ecology and evolution of bees? New Phytol. 225(3), 1053–1064. https://doi.org/10.1111/nph.16230

Roeder T (1999). Octopamine in invertebrates. Prog Neurobiol. 59, 533–561. https://doi.org/10.1016/S0301-0082(99)00016-7

- Roeder T (2005). Tyramine and octopamine: ruling behavior and metabolism. Annu Rev Entomol 50, 447–477. https://doi.org/10.1146/annurev.ento.50.071803.130404
- de Roode JC, Lefèvre T, Hunter MD (2013). Self-medication in animals. Science. 340, 150– 151. https://doi.org/10.1126/science.1235824

Roy R, Schmitt AJ, Thomas JB, Carter CJ (2017). Review: nectar biology: from molecules to

ecosystems. Plant Sci. 262, 148–164. https://dx.doi.org/10.1016/j.plantsci.2017.04.012

- Russell AL, Rebolleda GM, Shaible TM, Ashman TL (2019). Movers and shakers: bumble bee foraging behavior shapes the dispersal of microbes among and within flowers. Ecosphere. 10, e02714. https://doi.org/10.1002/ecs2.2714
- Valance D, Boissy A, Després G, Arnould C, Galand C, Favreau A, Constantin P, Leterrier C (2008). Changes in social environment induce higher emotional disturbances than changes in physical environment in quail. App Anim Behav Sci. 112, 307–320. 10.1016/j.applanim.2007.08.002
- Vannette RL, Fukami T (2016). Nectar microbes can reduce secondary metabolites in nectar and alter effects on nectar consumption by pollinators. Ecology. 97, 1410–1419. https://doi.org/10.1890/15-0858.1
- Vannette RL, Gauthier M-PL, Fukami T (2013). Nectar bacteria, but not yeast, weaken a plant– pollinator mutualism. Proc Royal Soc B. 280, e20122601. https://doi.org/10.1098/rspb.2012.2601
- Vannette RL, Fukami T (2018). Contrasting effects of yeasts and bacteria on floral nectar traits. Ann Bot. 121(7), 1343–1349. https://doi.org/10.1093/aob/mcy032
- Vannette RL (2020). The floral microbiome: plant, pollinator and microbial perspectives. 51, 363–386. https://doi.org/10.1146/annurev-ecolsys-011720-013401
- van Staaden MJ, Scott Hall F, Huber R (2018). The deep evolutionary roots of addiction. J Ment Health Clin Psychol. 2(3), 8–13. https://doi.org/10.29245/2578-2959/2018/3.1135
- Vazquez y Novo SP, Wagner ML, Gurni AA (1989). Importancia toxicológica de la presencia de sustancias aminadas en ejemplares de *Ligaria cuneifolia* var. *cuneifolia* colectados en diferentes areas de la República Argentina. Acta Farm Bonaerense. 8(1), 23-9.
 http://sedici.unlp.edu.ar/handle/10915/7253

Verpoorte R (2005). Alkaloids. In: Worsfold P, Townshend A, Poole C (eds), Encyclopedia of

analytical scienceAmsterdam, Elsevier. pp 77–84. https://doi.org/10.1016/B0-12-369397-7/00010-8

Verpoorte R, Schripsema J (1994). Isolation, identification, and structure elucidation of alkaloids: a general review. In: Linskens HF, Jackson JF (eds), Modern methods of plant analysis. Vol. 15. Springer-Verlag, pp. 1–24.

Vega FE, Dowd PF (2005). The role of yeasts as insect endosymbionts. In: Vega FE, Blackwell M (eds), Insect-Fungal

Associations: Ecology and Evolution. Oxford University Press. pp. 211-243.

- von Arx M, Moore A, Davidowitz G, Arnold AE (2019). Diversity and distribution of microbial communities in floral nectar of two night-blooming plants of the Sonoran Desert. PLOS ONE. 14, e0225309. https://doi.org/10.1371/journal.pone.0225309
- Vorel CA, Pitts-Singer T (2010). The proboscis extension reflex not elicited in Megachilids bees. J Kansal Entom Soc. 83(1), 80–83. https://doi.org/10.2317/0022-8567-83.1.80

Samorini G (2013). Animali che si drogano. ShaKe Edizioni

- Schaeffer RN, Phillips CR, Duryea MC, Andicoechea J, Irwin RE (2014). Nectar yeasts in the tall larkspur *Delphinium barbeyi* (Ranunculaceae) and effects on components of pollinator foraging behavior. PLoS One. 9, e108214. https://doi.org/10.1371/journal.pone.0108214
- Schaeffer RN, Mei YZ, Andicoechea J, Manson JS, Irwin RE (2017). Consequences of a nectar yeast for pollinator preference and performance. Funct Ecol. 31, 613–621. https://doi.org/10.1111/1365-2435.12762
- Scheiner R, Baumann A, Blenau W (2006). Aminergic control and modulation of honeybee behaviour. Curr Neuropharmacol. 4, 259–276 https://doi.org/10.2174/157015906778520791
- Schmitt A, Roy R, Carter CJ (2021). Nectar antimicrobial compounds and their potential effects on pollinators. Curr Opin Insect Sci. 44, 55–63. https://doi.org/10.1016/j.cois.2021.03.004
- Schoelhorn K (1919). Sur la fermentation de quelques levures des nectars des plants d'hiver. Bulletin de la Société de Genève. 11, 154–190.

- Schubart H, Anderson A (1978). Why don't ants visit flowers? A reply to D. H. Janzen. Biotropica. 10, 310. https://doi.org/10.2307/2387686
- Schulz DJ, Robinson GE (2001). Octopamine influences division of labor in honey bee colonies. J Comp Physiol A. 187, 53–61.
- Schuster V, Úlehla V (1913). Studien über Nektarorganismen. Berichte der Deutschen Botanischen Geselschaft. 31, 129–139.
- Seo HJ, Song J, Yoon HJ, Lee KY (2019). Effects of nectar contents on the foraging activity of honeybee (*Apis mellifera*) on Asian pear (*Pyrus pyrifolia* Nakai). Sci Hortic. 245, 185–192. https://doi.org/10.1016/j.scienta.2018.10.009
- Servillo L, Castaldo D, Giovane A, Casale R, D'Onofrio N, Cautela D, Balestrieri ML (2017). Tyramine pathways in citrus plant defense: Glycoconjugates of tyramine and its Nmethylated derivatives. J Agric Food Chem. 65, 892–899. https://doi.org/10.1021/acs.jafc.6b04423
- Shade A, McManus PS, Handelsman J (2013). Unexpected diversity during community succession in the apple flower microbiome. mBio. 4, e00602-12. https://doi.org/10.1128/mBio.00602-12
- Shen S, Peng M, Fang H, Wang Z, Zhou S, Jing X, Zhang M, Yang C, Gou H, Li Y, Lei L Shi Y, Sun Y, Liu X, Xu C, Tohge T, Yuan M, Fernie AR, Ning Y, Wang GL, Luo J (2021). An *Oryza*-specific hydroxycinnamoyl tyramine gene cluster contributes to enhance disease resistance. Sci Bull. 66, 2369–2380. https://doi.org/10.1016/j.scib.2021.03.015
- Singer MS, Mace KC, Bernays EA (2009). Self-medication as adaptive plasticity: increased ingestion of plant toxins by parasitized caterpillars. PLoS ONE. 4, e4796. https://doi.org/10.1371/journal.pone.0004796

Slansky F (1992). Allelochemical-nutrient interactions in herbivore nutrient ecology, vol. II.

Ecological and Evolutionary processes. In: Rosenthal GA, Berenbaum MR (eds), Herbivores: their interactions with secondary plant metabolites, vol. II, 2nd edn. Academic Press, San Diego. pp. 135–176

- Solvi C, Baciadonna L, Chittka L (2016). Unexpected rewards induce dopaminedependent positive emotionelike state changes in bumblebees. Science, 353(6307), 1529e1531. https://doi.org/10.1126/science.aaf4454
- Sprengel CK (1793). Das entdeckte geheimniss der natur im bauund in der befruchtung der blumen. Vieweg, Berlin
- Stefanini I (2018). Yeast-insect associations: it takes guts. Yeast. 35, 315–330. https://doi.org/10.1002/yea.3309
- Stephenson AG (1981). Toxic nectar deters nectar thieves of *Catalpa speciosa* (Bignoniaceae). Am Midl Nat. 105, 381–383. https://doi.org/10.2307/2424757
- Stephenson AG (1982). Iridoid glycosides in the nectar of Catalpa speciosa are unpalatable to nectar thieves. J Chem Ecol. 8(7), 1025–1034. https://doi.org/10.1007/BF00987883
- Stevenson PC, Nicolson SW, Wright GA (2017). Plant secondary metabolites in nectar: impacts on pollinators and ecological functions. Funct Ecol. 31, 65–75. https://doi.org/10.1111/1365-2435.12761
- Stevenson PC (2019). For antagonists and mutualists: the paradox of insect toxic secondary metabolites in nectar and pollen. Phytochem Rev. 19, 603–614. https://doi.org/10.1007/s11101-019-09642-y
- Tew NE, Memmott J, Vaughan IP, Bird S, Stone GN; Potts SG, Baldock KCR (2021). Quantifying nectar production by flowering plants in urban and rural landscapes. J Ecol. 109, 1747–1757. https://doi.org/10.1111/1365-2745.13598

Thorburn LP, Adler LS, Irwin RE, Palmer-Young EC (2015). Variable effects of nicotine and anabasine on parasitized bumble bees. F1000Res. 4, 880. https://doi.org/10.12688/f1000research.6870.1

- Thornburg RW, Carter C, Powell A, Mittler R, Rizhsky L, Horner HT (2003). A major function of the tobacco floral nectary is defense against microbial attack. Plant Syst Evol. 238, 211–218.
- Tiedeken EJ, Egan PA, Stevenson PC, Wright GA, Brown MJF, Power EF, Farrell I, Matthews SM, Stout JC (2016). Nectar chemistry modulates the impact of an invasive plant on native pollinators. Funct Ecol. 30, 885–893. https://doi.org/10.1111/1365-2435.12588
- Trivedi K, Borkovcová I, Karpíšková R (2009). Tyramine production by Enterococci from various foodstuffs: a threat to the consumers. Czech J Food Sci. 27, S357–S360. https://doi.org/10.17221/934-CJFS
- Yang M, Deng G-C, Gong Y-B, Huang S-Q (2019). Nectar yeasts enhance the interaction between Clematis akebioides and its bumblebee pollinator. Plant Biol. 21, 732–737. https://doi.org/10.1111/plb.12957
- Waddington KD (2001) Subjective evaluation and choice behavior by nectar- and pollen-collecting bees, In: Chittka L, Thompson JD (eds.), Cognitive Ecology of Pollination: Animal Behavior and Floral Evolution. Cambridge University Press, Cambridge, pp. 41–60.
- Wiens F, Zitzmann A, Lachance MA, Yegles M, Pragst F, Wurst FM (2008). Chronic intake of fermented floral nectar by wild treeshrews. PNAS. 105, 10426–104231. https://doi.org/10.1073/pnas.0801628105
- Wink M (2018). Plant secondary metabolites modulate insect behavior-steps toward addiction? Front Physiol. 9, 364. https://doi.org/10.3389/fphys.2018.00364
- Wright GA, Baker DD, Palmer MJ, Stabler D, Mustard JA, Power EF, Borland AM, Stevenson PC (2013). Caffeine in floral nectar enhances a pollinator's memory of reward. Science. 339, 1202–1204. https://doi.org/10.1126/science.1228806
- Zhang J, Wang Z, Klett K, Qu Y, Tan K (2022). Higher toxin tolerance to triptolide, a terpenoid foraged by a sympatric honeybee. J Insect Physiol. 137, 104358. http://doi.org/10.1016/j.jinsphys.2022.104358

Declaration of Interest Statement

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Highlights:

- The series of hypotheses on the role of nectar is reviewed and collated together
- New discoveries challenge how we think floral visitors use their food environment
- Upon colonization, microbes introduce their own metabolites into the nectar
- Microbe byproducts may act as animal neurotransmitters and influence their behavior
- Future research should address the so called "manipulation for dispersal" hypothesis