



Epigenetic regulation of CXCR4 signaling in cancer pathogenesis and progression

Reem Khaled M.E. Alsayed^a, Abdul Q. Khan^a, Fareed Ahmad^{a,b,c}, Abdul Wahid Ansari^{a,b,c}, Majid Ali Alam^{a,b,c}, Jorg Buddenkotte^{a,b,c}, Martin Steinhoff^{a,b,c,d,e}, Shahab Uddin^{a,b,f}, Aamir Ahmad^{a,b,c,*}

^a Translational Research Institute, Academic Health System, Hamad Medical Corporation, Doha 3050, Qatar

^b Dermatology Institute, Academic Health System, Hamad Medical Corporation, Doha 3050, Qatar

^c Department of Dermatology and Venereology, Rumailah Hospital, Hamad Medical Corporation, Doha 3050, Qatar

^d Weill Cornell Medicine-Qatar, Medical School, Doha 24144, Qatar

^e Department of Dermatology, Weill Cornell Medicine, New York, NY 10065, USA

^f Laboratory Animal Research Center, Qatar University, Doha 2713, Qatar

ARTICLE INFO

Keywords:

CXCR4
SDF-1/CXCL12
Epigenetic
Non-coding RNA
Methylation/acetylation

ABSTRACT

Signaling involving chemokine receptor CXCR4 and its ligand SDF-1/CXCL12 has been investigated for many years for its possible role in cancer progression and pathogenesis. Evidence emerging from clinical studies in recent years has further established diagnostic as well as prognostic importance of CXCR4 signaling. CXCR4 and SDF-1 are routinely reported to be elevated in tumors, distant metastases, which correlates with poor survival of patients. These findings have kindled interest in the mechanisms that regulate CXCR4/SDF-1 expression. Of note, there is a particular interest in the epigenetic regulation of CXCR4 signaling that may be responsible for up-regulated CXCR4 in primary as well as metastatic cancers. This review first lists the clinical evidence supporting CXCR4 signaling as putative cancer diagnostic and/or prognostic biomarker, followed by a discussion on reported epigenetic mechanisms that affect CXCR4 expression. These mechanisms include regulation by non-coding RNAs, such as, microRNAs, long non-coding RNAs and circular RNAs. Additionally, we also discuss the regulation of CXCR4 expression through methylation and acetylation. Better understanding and appreciation of epigenetic regulation of CXCR4 signaling can invariably lead to identification of novel therapeutic targets as well as therapies to regulate this oncogenic signaling.

1. Introduction: receptor CXCR4

The Chemokine receptor, C-X-C chemokine receptor 4 (CXCR4) is a G-protein-coupled receptor (GPCR) [1], that is also known by its other names fusin/LESTR and cluster of differentiation-184 (CD184) [2,3]. The most well-characterized ligand that binds and activates CXCR4 is stromal cell-derived factor-1 (SDF-1) [4]. Ligand SDF-1 is also known by its other name, CXCL12 [5]. For many years, SDF-1 was believed to be the exclusive ligand for CXCR4 before ubiquitin was reported as another legitimate ligand for CXCR4 [6]. Even though the two ligands activate CXCR4, leading to similar activation of downstream signaling, there are subtle differences as well, such as the relatively weaker chemotactic activity in case of ubiquitin-activated CXCR4 [7]. Additionally, the cytokine MIF (macrophage migration inhibitory factor) has also been

suggested as a ligand for CXCR4 [8]. Thus, CXCR4 can be activated by a cytokine (MIF), a chemokine (SDF-1) as well as a regulatory protein, ubiquitin. Chemokines are a sub-class of cytokines, also referred to as chemotactic cytokines, and, once secreted, they serve as chemo-attractants inducing directional movement of different cell types, including many different immune cells. The important role of both cytokines and chemokines in cancer pathogenesis has been recognized [9, 10], in addition to their role in many other diseases as well [11,12], which is primarily because of their profound effect on inflammation and immune responses [9,11,13,14].

CXCR4 activates G protein-dependent signaling pathways, such as, AKT, PI3K, mTOR and EGFR [15,16] as well as G protein-independent signaling pathways, that include, JAK/STAT, p53 MAPK and ERK [1, 15]. Under normal conditions, CXCR4 signaling plays a role in

* Correspondence to: Translational Research Institute, Dermatology Institute, Academic Health System, Hamad Medical Corporation, Doha 3050, Qatar.
E-mail address: aahmad9@hamad.qa (A. Ahmad).

<https://doi.org/10.1016/j.semcan.2022.03.019>

Received 1 February 2022; Received in revised form 18 March 2022; Accepted 21 March 2022

Available online 26 March 2022

1044-579X/© 2022 The Author(s). Published by Elsevier Ltd. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>).

embryogenesis, tissue repair, hematopoiesis, organogenesis and immune response [1,17]. Normal cells express little to no CXCR4; however, dysregulated and aberrant CXCR4 expression has been observed and reported in a plethora of tumors [18,19], with serious consequences on cancer progression, especially, cancer cell differentiation, proliferation, invasion, metastasis and angiogenesis [18,20,21]. In this article, the mechanisms of CXCR4 epigenetic dysregulation will be reviewed which could, in turn, affect the influence of CXCR4 signaling on cancer progression and prognosis.

2. CXCR4 signaling in cancer

The GPCR family of proteins, to which receptor CXCR4 belongs, is the largest class of integral membrane proteins [22]. Upon binding of their cognitive ligand, GPCRs are activated via rearrangement of transmembrane helices [22]. The early interest in CXCR4 signaling was related to its identification as the receptor needed for the entry of human immunodeficiency virus-1 (HIV-1) into T-cells [23,24]. The realization that CXCR4 signaling is critically involved in cancer aggressiveness, has fueled the interest in elucidating the various mechanisms by which it influences cancer progression.

2.1. CXCR4 effect on cancer cell proliferation, invasion and metastasis

The ability of cancer cells to proliferate, followed by invasion and metastasis, is central to their propagation and spread. It is, therefore, not surprising that numerous studies have documented a role of CXCR4 signaling in these fundamental processes associated with cancer progression. It is beyond the scope of this article to provide a detailed overview of this topic. The topic has been reviewed extensively [17, 25–31] and we provide only an overview here before discussing the clinical manifestations of CXCR4 in human cancer patients in the next section.

Receptor CXCR4 plays an important role in the induction of cancer proliferation, which often requires activation of cellular signaling such as MAPK or PI3K/Akt signaling pathways, among many others [15,21]. Similarly, overexpression of CXCR4 has been linked to increased invasion and migration, through regulation of multiple pathways [16,32, 33]. Further, angiogenesis is another important phenomenon exhibited by cancer cells. Angiogenesis through CXCR4 activation, in cancer cells, is triggered by platelet-secreted SDF-1, as well as VEGF [4]. As CXCR4 has an associated function in tissue regeneration, it was observed that the SDF-1 is released upon tissue damage [4]. Specifically, platelets in the blood release SDF-1 when a blood vessel is injured, and cancer cells utilize this pathway to trigger angiogenesis. Once released into the microenvironment, SDF-1 activates MMP-9 and soluble kit-ligand release (sKitL). Not only does sKitL has positive feedback on SDF-1 production, but it also aids in the movement of cells showing aberrant CXCR4 expression into the bloodstream. Cells overexpressing CXCR4 extravasate into the tumor microenvironment, which contains high levels of ligand SDF-1, and the homing of these cells leads to cancer cells' metastasis.

3. Clinical significance of CXCR4 in different cancers

The studies *in vitro* are invariably an important first step in the overall process of establishing the credibility of any potential diagnostic and/or prognostic biomarker. However, the clinical data from actual patient-derived samples provides the critical validation. In case of CXCR4 signaling, there is a wealth of clinical data supporting its role in pathogenesis of various human cancers. One of the most important phenomenon responsible for cancer-associated mortality is cancer metastasis. For example, the brain metastasis from primary breast cancers is associated with significantly reduced survival and there is evidence that ligand SDF-1 secreted by cancer-associated fibroblasts (CAFs) plays a role in attracting breast cancer cells to brain, resulting in brain

metastases [34]. The important role of CXCR4-SDF-1 signaling in cancer progression and metastasis is increasingly being realized and in the following few sub-sections, we discuss the recent literature supporting a role of CXCR4 and/or SDF-1 as diagnostic/prognostic biomarkers and their role in cancer metastasis. The inclusion criteria for the studies discussed in this section is that they all have data from human patient-derived samples.

3.1. Diagnostic biomarker

A plethora of studies have evaluated CXCR4 and/or SDF-1 expression levels in cancer tissue(s) obtained from cancer patients which reported elevated levels; thus, supporting the possibility of exploiting CXCR4 signaling as a valid diagnostic biomarker (Table 1). In lung cancer, the number one cancer in terms of deaths in the United States [35] as well as globally [36], a possible use of CXCR4 as diagnostic biomarker has been suggested with the observation that CXCR4 is upregulated in lung cancer tissues [37]. Even in the relatively rare lung adeno-squamous carcinoma, CXCR4 is a possible diagnostic biomarker [38] with seventy out of seventy-eight patients expressing CXCR4 and forty-five out of seventy-eight patients expressing particularly higher levels of CXCR4. In esophageal cancer, CXCR4 gene expression was reported higher in 52 esophageal squamous cell carcinoma patients, compared to normal esophageal mucosae, in one study [39] and in 101 esophageal squamous cell carcinoma patients in another study [40]. In gastric cancer patient samples, CXCR4 was reported to be significantly increased, compared to distant and adjacent non-cancer tissues [41,42]. In head and neck adenoid cystic carcinoma, CXCR4 positivity was reported in 81% of patients suggesting its possible diagnostic importance [43]. In vulvar squamous cell carcinoma, while CXCR4 was virtually absent in tumor tissues, a strong immunostaining was apparent in tumor tissues [44]. Also, CXCR4 is abundantly expressed in papillary thyroid cancer, compared to normal thyroids or nonmalignant tissues [45,46].

Pancreatic cancer is one of the highly aggressive human cancers and there seems to be data supporting a diagnostic role of CXCR4 in this cancer. In a meta-analysis that included eleven studies and an impressive 1439 pancreatic ductal adenocarcinoma patients, it was found that CXCR4 is significantly highly expressed in pancreatic tumors, relative to normal pancreatic tissues and that the CXCR4 associates with tumor progression with the levels significantly increased in higher grades pancreatic tumors [47]. In colorectal cancer, which happens to be a relatively better studied one in terms of the evaluation of CXCR4 [48, 49], a study [50] found immunoreactivity of 89% of 186 colorectal adenocarcinoma patients for CXCR4. As an interesting observation in this study, CXCR4 was reported high in tumors < 5 cm than those \geq 5 cm and in patients with tumor grade 1–2 than grade those with grade 3 tumors [50]. Another study reported elevated CXCR4 in all tumor tissues from 32 patients, relative to the levels in normal colon tissues [51]. Similarly, CXCR4 was significantly higher in 42 colorectal cancer samples, relative to 27 normal controls [52]. Thus, there seems to be data from multiple human cancers supporting a diagnostic importance of CXCR4 significance which can probably be exploited in clinics.

3.2. Metastasis

CXCR4 has also been linked to cancer metastasis [53]. In an evaluation of tumors from cervical cancer patients, CXCR4 was not only high in cancer patients, its levels were also higher in patients with lymph node metastases [54]. In lung cancer as well, CXCR4 is a risk factor for lymph node metastasis [55] with 84.8% patients with lymph node metastasis expressing CXCR4 in metastatic lymph nodes, which was higher than the expression of CXCR4 in non-metastatic lymph nodes. Similarly, CXCR4 and SDF-1 levels associated with TNM staging and cervical lymph node metastases of nasopharyngeal carcinoma [56]. CXCR4 has been associated with metastasis in colorectal cancer patients [52], pancreatic cancer patients [47], melanoma [57,58], lung

Table 1
Studies supporting diagnostic importance of CXCR4 signaling.

Cancer	Number of patients	Methodology	Observations	Study
Breast	40 Luminal B –	IHC, Gene array IHC	High CXCR4 in lymph nodes High SDF-1 in lymph nodes	Raschioni 2018 [62] Gadalla 2019 [63]
Cervical	48	IHC	Higher CXCR4 in patients with lymph node metastasis	Dai 2017 [54]
Colorectal	186 32 49	IHC qRT-PCR	89% of patients exhibit CXCR4 expression Higher CXCR4 in cancer tissues	Saka 2017 [50] Yoshuantari 2018 [51] Mitchell 2019 [52]
Esophageal Squamous Cell Carcinoma	52 101	qRT-PCR	Higher CXCR4 mRNA in patient samples	Goto 2017 [39] Yang 2020 [40]
Gastric	120 589	IHC IHC	High CXCR4 in cancer tissues and metastases Higher CXCR4 in cancer tissues	Yu 2018 [41] Chen 2021 [42]
Lung	185 110 78 (Adeno-squamous carcinoma)	IHC qRT-PCR IHC	Elevated CXCR4 levels in lung adenocarcinoma tissues CXCR4 is high in metastatic lymph nodes 89.7% patients positive for CXCR4 with 57.7% exhibiting high levels	Cong 2017 [37] Bi 2017 [55] Zhu 2020 [38]
Melanoma	35 656	IHC Meta-analysis	High CXCR4 in patients with metastasis CXCR4 associates with lymph node metastasis	Ipenburg 2019 [57] Alimohammadi 2021 [58]
Nasopharyngeal	102	qRT-PCR, western blot	Higher expression of CXCR4 and SDF-1 in cancer tissues	Li 2017 [56]
Oral	51	IHC	Higher CXCR4 and CD133 in lymph node metastasis	Gokulan 2021 [60]
Osteosarcoma	50	IHC	CXCR4 and CD133 associate with lung metastases	Mardani 2020 [59]
Pancreatic	1439	Meta-analysis	Higher CXCR4 in tumor tissues and metastases	Ding 2019 [47]
Thyroid Cancer	54 (Follicular) 74 (Papillary)	IHC IHC	CXCR4 determines tumor size and distant metastases CXCR4 is high in patient samples	Werner 2018 [61] Sirakriengkrai 2021 [45]
Vulvar	115 (Papillary) 46	IHC IHC	CXCR4 is high in patients CXCR4 high in tumor samples	Cao 2021 [46] Rusetska 2021 [44]

adenosquamous carcinoma [38] and papillary thyroid cancer [46]. In osteosarcoma, a correlation of CXCR4 with lung metastasis was found, but only in association with the other stem cell marker CD133 [59] and, interestingly, a similar correlation between CXCR4 and CD133 but in lymph node metastases from primary oral squamous cell carcinoma has also been reported [60]. Elevated levels of CXCR4 have also been associated with TNM staging, lymph node metastases as well as distant metastases in gastric cancer [41] and follicular thyroid cancer [61]. In samples from patients with luminal B breast cancer, CXCR4 levels are high in lymph nodes suggesting a role of CXCR4 in lymph node metastasis [62]. Another study reported significantly high immunostaining of SDF-1 in patients with positive lymph nodes, compared to those with negative lymph nodes [63]. In summary, CXCR4 expression seems to correlate with metastasis, particularly lymph node metastasis in many different cancers.

3.3. Prognostic biomarker

In addition to the possible diagnostic importance of CXCR4 signaling, there has been emergence of consistent data supporting a possible prognostic importance of CXCR4 and SDF-1 (Table 2). In esophageal cancer, higher CXCR4 expression in the cytoplasm and nuclei correlated with poor cause-specific survival [39] while the patients with high levels of ligand SDF-1 tended to have worse overall survival and disease-free survival [64]. In follicular thyroid cancer, CXCR4 expression has been reported to associate with poor overall and recurrence-free survival [61]. A meta-analysis reported an association of CXCR4 with poor survival in patients with pancreatic ductal adenocarcinoma [47]. In osteosarcoma patients, an effect of CXCR4 on metastasis-free survival was reported, with a 72 months metastasis-free survival in patients without CXCR4, compared to just 14 months metastasis-free survival in patients positive for CXCR4 [65]. Similar association of CXCR4 with poor overall survival in esophageal squamous cell carcinoma [40], gastric cancer [42] and oral squamous cell carcinoma [60] has also been reported.

In a colorectal cancer study with 60 patients, the patients with CXCR4 expression had a worse 5-year survival, compared to CXCR4-negative patients [66]. This was further confirmed in a meta-analysis

wherein receptor CXCR4 and its ligand SDF-1 were found to predict reduced disease-free survival and overall survival [67]. Other studies have also suggested a prognostic importance of CXCR4 or SDF-1 in colorectal cancer patients [52,68,69]. Further, a role of CXCR4 polymorphism in colorectal cancer prognosis has also been suggested with a common variant, CXCR4 rs2228014 linked to poor progression-free survival [70]. Interestingly, a possible role of CXCR4 polymorphism in breast cancer prognosis has also been suggested [71].

In lung cancer, CXCR4, in combination with Notch1, correlated with nodal stage, tumor stage and lymphovascular invasion in addition to poor prognosis [37]. Also, high CXCR4 at primary lung site while high SDF-1 at metastatic lymph nodes are determinants of poor overall survival [72]. CXCR4 has prognostic importance in the rare lung adenosquamous carcinoma and correlates with decreased overall and disease-free survival [38]. In a head and neck squamous cell carcinoma study [73], while no association between CXCR4 or SDF-1 with metastasis free survival or overall survival could be established, SDF-1 was still found to be a negative prognostic biomarker for loco-regional control after postoperative radiochemotherapy. This, however, is different from the observations in a study comprising of head and neck adenoid cystic carcinoma patients wherein CXCR4 was reported as an independent prognosis biomarker for poor recurrence-free survival [43]. In nasopharyngeal carcinoma, however, patients with higher CXCR4 and SDF-1 had significantly reduced survival, compared to those with lower expression [56]. Taken together, all this evidence supports a negative correlation between CXCR4/SDF-1 expression and good prognosis of cancer patients, and should be something that the attending clinicians might look out for, at the end of regular therapy.

4. Epigenetic regulation of CXCR4 signaling

Epigenetic regulation is the way a cell controls the expression of its genes, and this regulation defines how the cell functions. There are many machineries a cell can deploy to manipulate the expression of genes. Commonly, epigenetics could be manipulated using non-coding RNAs, such as microRNAs (miRNAs) [74,75], long non-coding RNAs (lncRNAs) [76,77] and circular RNAs (circRNAs) [78,79]. Other examples of common types of epigenetic gene expression regulation include

Table 2
Prognostic importance of CXCR4 signaling, as revealed in clinical studies.

Cancer	Number of patient samples (n)	Methodology	Observations	Study
Colorectal Cancer	60	IHC	CXCR4 expression correlates with poor 5-year survival	Ogawa 2017 [66]
	874	Meta-analysis	CXCR4-SDF-1 predict poor overall survival	Li 2017 [67]
		PCR sequencing	CXCR4 variant rs2228014 predicts progression-free survival	Matsusaka 2017 [70]
	49	qRT-PCR	CXCR4 determines overall survival	Mitchell 2019 [52]
Esophageal Squamous Cell Carcinoma	78	IHC	CXCR4 predicts response to chemotherapy	Ottaiano 2020 [68]
	172	IHC	High CXCR4 correlated with poor cause-specific survival	Goto 2017 [39]
	101		CXCR4 associates with poor overall survival	Yang 2020 [40]
Head & Neck Cancer	55	IHC	High SDF-1 patients have worse overall and disease free survival	Goto 2021 [64]
	201	IF	SDF-1 negatively correlates with loco-regional control	De-Colle 2017 [73]
Lung Cancer	66	IHC	CXCR4 correlates with poor recurrence-free survival	Nulent 2020 [43]
	185	IHC	Correlates with poor prognosis	Cong 2017 [37]
	140		CXCR4 and SDF-1 predict poor prognosis	Katsura 2018 [72]
Nasopharyngeal Carcinoma	78		CXCR4 associates with decreased overall and disease-free survival	Zhu 2020 [38]
	102	IHC	CXCR4 and SDF-1 correlate with shorter survival	Li 2017 [56]
Osteosarcoma	73	IHC	CXCR4 significantly reduces recurrence-free survival	Gong 2020 [65]
Pancreatic Cancer	1439	Meta-analysis	CXCR4 associates with poor survival	Ding 2019 [47]
Thyroid Cancer (Follicular)	72	IHC	CXCR4 correlates with poor overall and recurrence-free survival	Werner 2018 [61]

IHC: Immunohistochemistry

methylation (which is commonly associated with gene silencing) and acetylation (which is commonly associated with gene activation). A number of studies have been reported that support epigenetic regulation of CXCR4 by non-coding RNAs, methylation or acetylation. The sub-sections to follow discuss such aspects of CXCR4 regulation.

In addition to direct epigenetic regulation of CXCR4, as mentioned above, there are some indirect mechanisms by which the epigenetics of CXCR4 gene could be influenced. Transcription factors often influence gene expression, and for CXCR4, nuclear respiratory factor-1 [1] and hypoxia-inducible factor 1-alpha [30] are responsible for the upregulation of the transcription of CXCR4. Furthermore, some growth factors are known to increase CXCR4, such as, hepatocyte growth factor [30]. Additionally, the extracellular conditions also have an ability to affect the expression of CXCR4, for example, hypoxia not only induces CXCR4 production in cancer tissues, but it also stabilizes CXCR4's expression [80]. However, some molecules, such as, acetyl-11-keto- β -boswellic acid reduce the expression of CXCR4 gene, which results in changes in the cancer behavior, including, reduced invasion and migration [81]. Hence, there are many ways, direct as well as indirect, through which the expression of CXCR4 can be epigenetically controlled.

4.1. Effect of non-coding RNAs on CXCR4 expression

4.1.1. Effect of miRNAs on CXCR4

miRNAs represent a major class of non-coding RNAs that play an essential part in regulating gene expression [82,83]. After being transcribed from the DNA, miRNAs typically bind to the 3' untranslated region of their target(s) mRNA(s) to tag them for degradation or to block translation [82]. However, in certain cases, miRNA can even upregulate certain genes [82,84]. Therefore, miRNA can epigenetic regulate the expression of genes in multiple ways.

The ability of miRNAs to control the expression of CXCR4 has been demonstrated in a range of different cancers. In many of these cases, miRNA inhibits the production of CXCR4. Examples of this include; miR-146 [85], miR-193-5p [86], miR-206 [87], and miR-622 [88] in colorectal cancer (CRC) cells; miR-622 in hepatocellular carcinoma [89]; miR-126 and miR-221 in lung cancer [90]; miR-139 in breast cancer [91]; miRNA-34a and miR-200c down regulate CXCR4 through targeting HIF1-a in breast cancer [92]; miR-155 indirectly reduces CXCR4 levels in glioblastoma [93], miR-126 in gastric cancer [94]; miR-381 in pancreatic ductal adenocarcinoma [95]; miR-143 inhibits CXCR4 expression in melanoma cancer cells by inhibiting proliferation and migration and facilitating apoptosis [96]; miR-143 inhibits the expression of CXCR4 in oral squamous cell cancer [97]; miR-330 decreases the expression of CXCR4 in melanoma [98]; miR-204 indirectly inhibits cxc4 expression through the NF- κ B signaling pathway in nasopharyngeal carcinoma [99]; miR-140-3p inhibits CXCR4 expression through targeting of its 3'UTR in colorectal cancer [100], and nasopharyngeal carcinoma [101]; miR-381-3p decreases the mRNA and protein expression of CXCR4 in non-small cell lung cancer cells [102]; miR-494-3p was proven to inhibit CXCR4 production post-transcriptionally in Synovial sarcoma, prostate and breast cancer cells [103]; miR-193a-5p negatively regulates the expression of CXCR4 mRNA in colon cancer cells, which decreases the migration and metastasis of these cells [104]; miR-133b down regulated the expression of CXCR4 in colon cancer cells SW-480 and SW-620, which inhibited invasion and migration and the proliferation of the cells [105]; miR-613 directly shuts off the expression of CXCR4 and has a tumor suppressor effect in inhibiting cancer phenotypes in osteosarcoma cells [106]; miR-302a was proven to decreased CXCR4 levels in breast cancer [107]; miR-128 was shown to reduce CXCR4 protein levels in human thyroid cancer cells [108]; miR-1246 directly inhibits CXCR4 expression, hence inhibiting the renal cell carcinoma cell's ability to proliferate and migrate [109], as well as it inhibits CXCR4 in lung cancer cells [110]; miR-9 is a tumor suppressor that works by inhibiting growth and metastasis of glioblastoma cells by inhibiting CXCR4 mRNA [111], and

evidence from acute myeloid leukemia has also proved that miR-9 is responsible for inhibiting invasion, migration, proliferation, and apoptosis resistance of these cells, through the inhibition of CXCR4 mRNA and protein expression [112]. Hence, multiple miRNAs play an important role in the epigenetic inhibition of CXCR4 expression in a variety of cancers.

In addition to the suppression of CXCR4, in a few cases, it has been recognized that miRNAs could have an up-regulatory effect on CXCR4. Examples of this include let-7 f in human mesenchymal stem cells that are associated with breast cancer [113]. The expression of CXCR4 was also positively regulated by miR-301a in osteosarcoma [114]. Not only does miR-410 upregulate CXCR4 in non-small cells lung cancer, but it also increases the stemness of these cells [80].

4.1.2. Effect of circRNAs on CXCR4

Circular RNAs are the relatively new class of non-coding RNAs that are attracting attention for their role in cancer pathogenesis [78,115,116]. In last few years, some reports have emerged detailing a role of circRNAs in regulating CXCR4 signaling (Table 3). For example, in an early report on a possible connection between circRNAs and CXCR4 in cancer, it was shown that the oncogenic circ_0056618 is overexpressed in gastric cancer tissues and it sponges miR-206 reducing this miRNA's levels to attenuate the repression of CXCR4 [117]. Thus, elevated levels of circ_0056618 correlate with increased CXCR4 levels in gastric cancer. A similar role circ_0056618 has since been observed in colorectal cancer as well [87]. Here also, circ_0056618 sponged miR-206 and thus activated CXCR4. In addition to CXCR4, this study found similar effects of circ_0056618-miR-206 on VEGF-A, which could explain the positive regulation of angiogenesis by circ_0056618-miR-206 axis [87].

A bioinformatics-based study, that analyzed expression profiles of pancreatic ductal adenocarcinoma from multiple GEO databases, concluded that CXCR4 is one of the five hub genes overexpressed in pancreatic cancer [118]. Further, the analysis revealed that CXCR4 is regulated by circ-UBAP2 and hsa-miR-494 and that CXCR4 levels correlate with levels of M2 macrophages, a macrophage subtype with

Table 3
CXCR4 signaling regulating circular RNAs.

Circular RNA	Cancer	miRNA	Pathological Effect	Reference
ABC10	Head & Neck	miR-588	ABC10 and CXCR4 promote tumor growth	[120]
circFGFR1	NSCLC	miR-381	circFGFR1 promotes tumor progression and immune evasion	[102]
	Glioma	miR-224	circFGFR1 supports glioma progression	[122]
circN4BP2L2	Colorectal	miR-340	Promotes tumor growth and metastasis	[123]
circ_SLIT3	Hepatocellular	miR-223	circ_SLIT3 promotes tumor growth	[121]
circ_0056616	Lung	NE	circRNA_0056616 and CXCR4 levels elevated in metastatic patients	[124]
circ_0056618	Gastric	miR-206	circ_0056618 highly expressed in tumor tissues and negatively correlates with survival	[117]
	Colorectal		circ_0056618 is elevated in tumor tissues and promotes angiogenesis	[87]
circ_0020710	Melanoma	miR-370-3p	circ_0020710 upregulates SDF-1 and promotes melanoma progression	[125]
circ-UBAP2	Pancreatic	miR-494	CXCR4 mediates immune responses within tumor microenvironment	[118]

NE: none evaluated

important role in tumor progression [119].

The circRNA-CXCR4 axis has been reported in several other cancers as well. circRNA ABCB10 has an up regulatory effect on the expression of CXCR4 in head and neck laryngeal squamous cell carcinoma [120]. Suppression of this circRNA reduced invasion and miR-588 was identified as the miRNA sponged by ABCB10 that could, in turn, target CXCR4. Circ_SLIT3 can sponge miR-223 leading to CXCR4 upregulation and increased tumor growth in hepatocellular carcinoma [121]. circFGFR1 is a circRNA that regulates the expression of CXCR4, through the sponging/inhibition of miR 381-3p in NSCLC cells [102]. This circRNA is upregulated in NSCLC tissues and correlates with poor prognosis of patients. circFGFR1 promotes glioma progression as well [122] by sponging miR-224 and upregulating CXCR4. In colorectal cancer, circN4BP2L2 is another circRNA that promotes cancer growth [123]. This circRNA sponges miR-340 and upregulates CXCR4 to induce tumor growth and metastasis.

A possible regulation of CXCR4 by circular RNAs with implications on cancer metastasis has been suggested in lung cancer. Similar to many studies detailed above in the section on diagnostic importance of CXCR4, this study also observed elevated CXCR4 levels in lung cancer patients with lymph node metastasis [124]. Further, circRNA_0056616 levels, as evaluated in released exosomes, were also high in patients with metastatic disease [124]. circRNAs can not just regulate CXCR4 but its ligand SDF-1 as well. In a study on melanoma, circ_0020710 was firstly found to be elevated in melanoma tissues, as determined by qRT-PCR, thus establishing its oncogenic role in melanoma [125]. Mechanism-wise, circ_0020710 upregulated SDF-1 through its sponging of miR-370-3p.

4.1.3. Effect of lncRNAs on CXCR4

LncRNAs' role in cancer metastasis and progression is increasingly being realized [77,126–129]. It is also being realized that lncRNAs exhibit regulatory effects on CXCR4 gene expression and CXCR4-SDF-1 signaling (Table 4). In an early report on the topic, lncRNA MALAT1 was investigated in human hilar cholangiocarcinoma and found to be expressed at higher levels in the samples obtained from cholangiocarcinoma patients [130]. This lncRNA MALAT1 associated positively with tumor stage and was reported to interact with miR-204 leading to increased growth, migration and invasion of cholangiocarcinoma cells through the upregulated CXCR4. miR-204 is also sponged by another lncRNA, LINC00922, which is upregulated in lung cancer cells [131]. Additionally, LINC00922 is associated with poor prognosis, as it enhances proliferation, and inhibits the CXCR4 expression suppressor miR-204 by competitively binding to it [131]. Lung cancer progression is further affected by lncRNA NORAD, which is elevated in NSCLC patients tissues as well as cell lines [132]. Inhibition of NORAD negatively affects cell growth because of reduced CXCR4 and SDF-1.

LncRNA HOTAIR, down regulates the expression of miR-126, thereby increasing CXCR4 expression in gastric cancer [94]. Another lncRNA, HNRNPKP2, also positively affects CXCR4 expression which possibly correlates with increased proliferation, migration invasion and even hepatic metastases of gastric cancer [133]. LncRNA COL1A1-014 is elevated in gastric cancer tissues as well as cell lines and its overexpression leads to elevated mRNA levels of ligand SDF-1 and elevated protein levels of both CXCR4 and SDF-1 [134]. lncRNA FER1L4, on the contrary, is a tumor suppressor lncRNA in gastric cancer cells whose overexpression reduces CXCR4 and SDF-1 resulting in reduced cell proliferation, invasion and lymphatic metastasis [135].

LncRNA GAS5 was shown to positively correlate with proliferation and invasion of esophageal cancer cells but through downregulation of CXCR4 [136]. Such activity of GAS5 was through its sponging of miR-301a. LncRNA UCA1 can also sponge miR-301a and thus affect CXCR4 expression, as shown in a study performed in osteosarcoma cells [114]. Additionally, UCA1 has been reported to sponge miR-204 in prostate cancer cells leading to increased growth and invasion through

Table 4
CXCR4 signaling regulating long non-coding RNAs.

lncRNA	Cancer	miRNA sponged	Pathological effect	Reference
COL1A1-014	Gastric	miR-1273h	Promotes cell growth	[134]
DLEU1	Pancreatic	miR-381	Upregulates CXCR4	[95]
DUXAP8	Thyroid	miR-223	Upregulates CXCR4	[143]
FER1L4	Gastric	NE	Downregulates CXCR4 and SDF-1	[135]
FEZF1-AS1	Osteosarcoma	miR-144	Promotes Warburg effect and proliferation	[138]
GAS5	Esophageal	miR-301a	Downregulates CXCR4	[136]
HILAR	Renal	miR-613	Induces cancer metastasis	[144]
HNRNP2	Gastric	NE	Induces cell proliferation, invasion and hepatic metastasis	[133]
HOTAIR	Gastric	miR-126	Upregulates CXCR4	[94]
LINC00922	Lung	miR-204	Upregulates CXCR4	[131]
LSINCT5	Ovarian	NE	Induces proliferation, migration and invasion through upregulated CXCR4	[139]
MALAT1	Cholangiocarcinoma	miR-204	Induces cell growth, migration and invasion through elevated CXCR4	[130]
	Leukemia	miR-146a	Sponges miR-146a to inhibit CXCR4	[141]
NEAT1	Retinoblastoma	miR-204	Correlates positively with CXCR4 in vitro and in vivo	[140]
NORAD	Lung	NE	Upregulates CXCR4 and SDF-1	[132]
TUG1	Tongue squamous	miR-133b	Determines resistance against cisplatin	[145]
UCA1	Osteosarcoma	miR-301a	Supports cell growth	[114]
	Prostate	miR-204	Induces cancer cell growth and metastasis	[137]
	Leukemia	NE	Upregulates CXCR4	[142]

NE: none evaluated

elevated CXCR4 [137]. In osteosarcoma cells, lncRNA FEZF1-AS1 sponges miR-144 and induces CXCR4 along with regulation of Warburg effect and suppression of apoptosis [138].

In ovarian cancer patients, lncRNA LSINCT5 was reported to be expressed at higher levels, relative to the normal ovarian tissue [139], especially in lymphatic metastases. Silencing of LSINCT5 reduced cell proliferation, migration and invasion through downregulation of CXCR4. lncRNA NEAT1 also correlated positively with CXCR4 expression in retinoblastoma tissues and cells [140]. NEAT1 sponged miR-204, a suppressor of CXCR4, thereby positively regulating CXCR4 expression in vitro as well as in vivo.

In pancreatic ductal adenocarcinoma, lncRNA DLEU1 has an oncogenic function, as it was shown to sponge miR-381, a CXCR4 targeting miRNA [95]. Hence, DLEU1 has an indirect role in the upregulation of

CXCR4. In acute myeloid leukemia, MALAT1 inhibits CXCR4 expression through the sponging of miR-146a [141], while UCA1 induces CXCR4 expression [142]. In papillary thyroid cancer, DUXAP8 lncRNA upregulates CXCR4 through targeting of miR-223 [143] while in renal cancer, lncRNA HILAR promotes renal cancer metastasis through sponging of miR-613 and upregulation of Notch and CXCR4 signaling [144].

lncRNAs have also been found to regulate CXCR4 signaling with resulting effects on sensitivity to therapy. For example, in a study that recruited twenty-one cisplatin sensitive vs. resistant tongue squamous cell carcinoma patients [145], higher lncRNA TUG1 levels were found in the resistant patients. TUG1 targeted miR-133b and therefore overexpression of miR-133b suppressed cisplatin resistance. miR-133b, in turn, targeted CXCR4 which was elevated in resistant cells with increased TUG1. In summary, evidence for the regulation of CXCR4 by lncRNAs is slowly but surely emerging and in most of the reports lncRNAs-mediated regulation of CXCR4 involves sponging of specific miRNAs, a mechanism also seen in circRNAs-mediated CXCR4 regulation.

4.2. Effect of methylation on CXCR4 expression

An important epigenetic modification is the methylation of genes. Methylation takes place when a methyl group (CH₃) covalently binds to the 5'-carbon of a cytosine in the DNA, and this process is catalyzed by DNA methyltransferases (DNMTs) [146]. Targets of methylation in the genome that are associated with gene expression regulation are CpG islands, CpG shores, and first exons [146]. Methylation is typically viewed as a gene silencer as it prevents the DNA from interacting with transcription factors and chromatin proteins [147]. In cancer, DNA methylation or demethylation (removal of methyl groups) plays a large role in the activation or silencing of tumor related genes [148]. The state of methylation of specific cancer-related genes can be used as a tumor biomarker [146]. The methylation of cancer-related genes is specific and different across different cancers [146]. Hypermethylation is often observed in the promoter regions of tumor suppressor genes in cancer, and protection against methylation is observed in the promoters of oncogenes [146].

The gene expression of CXCR4 is highly affected by methylation [149]. In a breast cancer study, the CXCR4 gene was observed to be significantly hypomethylated, and this provides an epigenetic explanation to the overexpression of CXCR4 in these cells [149]. Compared to regular breast tissue (48%), the hypomethylation was abundant in cancerous breast tissue (78%), which makes methylation a possible marker for breast cancer. In another breast cancer study, it was seen that the methylation status of CXCR4 directly correlated with metastatic status of the tumors [150]. For example, metastasis to the lymph nodes was associated with the lack of CXCR4 promoter methylation. Furthermore, another study about breast cancer showed that in 67% of the breast cancer samples analyzed, there was a loss of methylation in the promoter region of CXCR4 gene, and this observation was associated with different phenotypes including tumor size, metastatic potential, tumor stage and patient survival [151]. Therefore, the methylation status of the CXCR4 gene influences breast cancer tumor progression.

In colorectal cancer, an increased expression of CXCR4 associated with high levels of 5-hydroxymethylcytosine (5hmC) in the CXCR4 gene [152]. Generally, 5hmC is an epigenetic marker that can represent an active demethylation intermediate. Therefore, in colorectal cancer, it is suggested that CXCR4 is overexpressed through its tagging for increased transcription by 5hmC. However, in pancreatic cancer cell lines, wherein the expression of CXCR4 was downregulated, hypermethylation was observed [153]. Whereas, these observations, on one hand document a role of methylation in regulation of CXCR4 expression in pancreatic cancer, but, on the other hand also suggest that not all pancreatic cancers might rely on the CXCR4 signaling pathway for the activation of cancer phenotypes and/or cancer progression. This also makes case for the argument that the use of CXCR4 inhibitors might not

prove an effective form of therapy against certain cancers or cancer subtypes. Additionally, a study about melanoma utilized a demethylating agent to investigate the effect of epigenetic modification on CXCR4 gene expression [154]. The inhibitor 5-Aza-2-deoxycytidine (5-Aza) was used for 4 melanoma cell lines. In three of four cell lines, 5-Aza had an inhibitory effect on CXCR4 gene expression, and, in one of the cell lines, where CXCR4 is typically hypermethylated, the demethylation using 5-Aza led to increase in the expression of CXCR4. Therefore, methylation plays a role in the expression of CXCR4 in melanoma.

4.3. Effect of histone acetylation on CXCR4 expression

Histone acetylation is another mechanism for the epigenetic regulation of genes. Histone acetylation involves the addition of an acetyl group on a ϵ -amino lysine residue in a histone [155]. This allows the chromatin structure to relax, and separates the DNA slightly from the histone, creating space for transcription regulators to access the DNA [156]. The effect that histone acetylation has on transcription is that of activation, and the process is carried out by acetyltransferases [157]. The enzyme histone deacetylase (HDAC) is responsible for removing acetyl groups, and HDAC inhibitors (such as *n*-butyrate) have been extensively used to study the effect of acetylation on cancer progression [158–160]. In cancer, the epigenetic regulation of cancer related genes (oncogenes or tumor suppressors) can affect the tumorigenesis or progression of the cancer [161]. Generally, there is a dysregulation of the histone acetylation of tumor suppressors and oncogenes in a way that favors tumor progression. Additionally, patient prognosis can be predicted using the loss of acetylation biomarkers. Hence, it is important to study histone acetylation in the activation of oncogenes and the suppression of tumor suppressors in human cancers, using appropriate models. The importance of acetylation in cancers is supported by the approval of four HDAC inhibitors, namely, vorinostat, romidepsin, panobinostat and belinostat by United States Food and Drug Administration (US-FDA) [162,163].

An important molecule in suppressing immunity and inducing skin cancer is platelet-activating factor (PAF) [164]. This mediator of inflammation in skin cancer upregulates the expression of CXCR4 through histone acetylation near the promoter region [164]. Hyperacetylation of the promoter region of CXCR4 is associated with migration in mast cells in skin cancer, making acetylation an important aspect of melanoma progression. In advanced prostate cancer, the transcription factor Ac-KLF5 is associated with the differentiation of osteoclasts and inducing metastatic bone lesions [165]. As a mechanism, Ac-KLF5 upregulates the expression of CXCR4 through histone acetylation of the promoter region of the CXCR4 gene [165]. Additionally, this study showed that upon the inhibition or knockdown of CXCR4 there was an abolition of advanced prostate cancer metastasis to bones and osteoclast differentiation. Hence, the effect of histone acetylation of the CXCR4 promoter is essential in metastasis of advanced prostate cancer.

In a breast cancer study, acetylation proved to indirectly manipulate the epigenetic expression of CXCR4 [156]. The HDAC inhibitor used was trichostatin A (TSA), and it worked by inhibiting the deacetylation on the promoter of the miRNA gene MIR146A of miR-146a, which promoted its upregulation. Since miR-146a has an inhibitory effect on the transcription and subsequent expression of CXCR4, an inhibitory effect on the production of CXCR4 was seen upon TSA addition. Breast cancer cells have a lower expression of miR-146a, and this relates to poor prognosis of patients. However, it could be inferred that with reversal of this effect i.e. upregulation of miR-146a through acetylation, the prognosis could be improved. Thus, histone acetylation affects the expression of CXCR4 which subsequently positively impacts breast cancer progression and prognosis.

In chronic lymphocytic leukemia, HDAC inhibitor suberoylanilide hydroxamic acid (SAHA) was used to study the expression of CXCR4 [166]. This study revealed that upon exposure of cells to SAHA, the gene expression level of CXCR4 was decreased, which reduced the migration

of the leukemia cells. Even though the exact mechanism for the blocking of CXCR4 expression by HDACi is not fully established, accumulating data indicates that HDAC inhibitor TSA reduces CXCR4 expression and might thus be a candidate for cancer therapy.

5. Conclusions and future perspectives

Based on the studies discussed in this article, it is apparent that CXCR4 signaling plays a critical role in the pathogenesis of various human malignancies and is involved in proliferation, invasion, angiogenesis and metastasis of tumors (Fig. 1). This is the collective conclusion from a plethora of studies that include *in vitro*, *in vivo* as well as clinical studies/trials. Additionally, a case has been made for possible diagnostic as well as prognostic importance of CXCR4 signaling. This calls for design of further studies to cement the clinical importance of CXCR4 signaling in order to help shape future therapeutic strategies for the benefit of cancer patients. To certain extent, initial evaluations for feasibility are already in progress. For example, in a breast cancer study, the diagnostic performance of CXCR4-directed PET imaging has been evaluated in patients given the encouraging data from both clinical and pre-clinical studies. This evaluation revealed that CXCR4-targeted PET imaging does not confer advantage as a general diagnostic tool for breast cancer imaging, particularly when compared with established techniques for tumors detection [167]. The study was a relatively small one with limited number of patients. Additionally, there needs to be a word of caution because CXCR4 targeting might be beneficial for only a subset within a patient population, or a cancer subtype. For instance, again in breast cancer, it was reported that inhibition of CXCR4 signaling, through the use of specific inhibitors AMD3100 and TN14003, could suppress tumor growth and metastasis in both therapy-sensitive and therapy-resistant HER2-overexpressing breast cancers represented by patient-derived xenografts [168]. This is an encouraging news supporting the logic behind targeting CXCR4 even in therapy-refractory cancers. However, the same study also noted that inhibition of CXCR4 in another subset of breast cancer, namely, triple negative breast cancer, could actually be counter-productive and may result in increased metastatic spread [168]. On a positive note, in a phase IIa study, antagonizing CXCR4 through the use of BL-8040/ motixafortide expanded the benefit of chemotherapy in pancreatic ductal adenocarcinoma patients, especially when combined with inhibitor of programmed cell death-1 (PD-1) [169]. All these observations suggest that identifying the right patient cohort/ cancer subtype as well as the combination treatments might be the strategy moving forward. The notion is further supported by a prognostic role of CXCR4 in response to chemotherapy [68] and the observation that SDF-1 is high in platinum-treated NSCLC patients with progressive disease and worse clinical prognosis [170].

Additionally, the epigenetic regulation of CXCR4 is evident at many levels with non-coding RNAs and methylation/acetylation, as discussed in this article, playing an important role in the eventual expression of CXCR4. It would therefore be interesting for the future studies to elucidate if HDAC inhibitors (such as the US-FDA approved HDAC inhibitors vorinostat, romidepsin, panobinostat and belinostat) can directly or indirectly affect receptor CXCR4 and/or ligand SDF-1 expression. Further, the post-translational modifications of CXCR4 such as phosphorylation, ubiquitination, glycosylation and sulfation can also potentially affect receptor-ligand interactions, and a better understanding of these can lead to novel strategies to regulate CXCR4 signaling. Moving forward, a more patient-centric and precision medicine-based approach combined with placebo-controlled, large scale clinical trials might be the best way to further evaluate and exploit the clinical value of CXCR4 signaling in cancer patients, and as a tool for imaging and targeted therapy.

CRedit authorship contribution statement

AA: Conceptualization, Supervision. RKA, AQK, FA, AWA and AA:

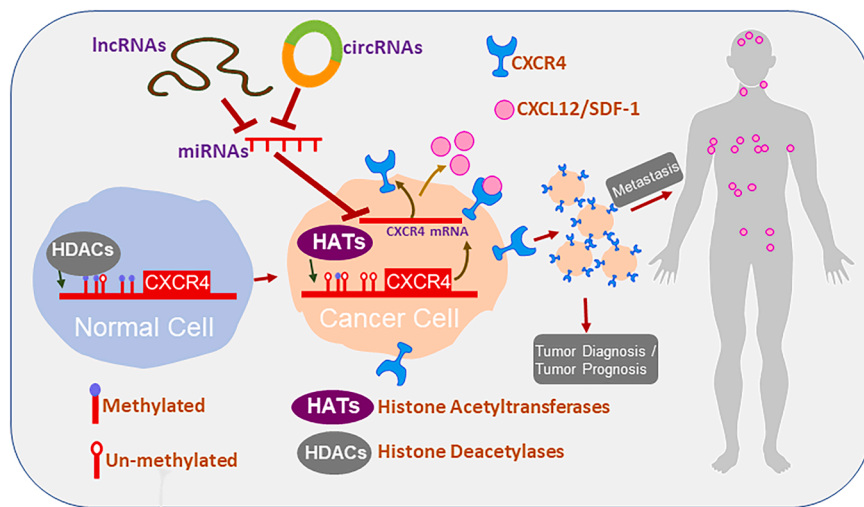


Fig. 1. CXCR4 in cancer progression. Expression of receptor CXCR4 can be induced by hypomethylation or acetylation leading to its recruitment at cell membrane. Additionally, non-coding RNAs, such as, microRNAs (miRNAs), long non-coding RNAs (lncRNAs) and circular RNAs (circRNAs) can affect CXCR4 transcription regulation. Ligand CXCL12/SDF-1 can bind to CXCR4 thereby activating the signaling resulting in increased proliferation, invasion, angiogenesis etc. Increased SDF-1 release at various distant organs can attract cancer cells (with CXCR4 expression), resulting in homing of cancer cells at distant sites. Elevated CXCR4/SDF-1 levels have been proposed as promising diagnostic and/or prognostic biomarkers.

Writing – original draft. MAA, JB, MS, SU and AA: Writing – review & editing.

Acknowledgment

Open Access funding for this article has been provided by the Qatar National Library.

Conflict of Interest Statement

None of the authors have any conflict of interest to report.

References

- [1] J.M. Busillo, J.L. Benovic, Regulation of CXCR4 signaling, *Biochim. Biophys. Acta* 1768 (4) (2007) 952–963, <https://doi.org/10.1016/j.bbamec.2006.11.002>.
- [2] K. Tachibana, T. Nakajima, A. Sato, K. Igarashi, H. Shida, H. Iizasa, N. Yoshida, O. Yoshie, T. Kishimoto, T. Nagasawa, CXCR4/fusin is not a species-specific barrier in murine cells for HIV-1 entry, *J. Exp. Med.* 185 (10) (1997) 1865–1870, <https://doi.org/10.1084/jem.185.10.1865>.
- [3] M. Burger, A. Glodek, T. Hartmann, A. Schmitt-Graff, L.E. Silberstein, N. Fujii, T. J. Kipps, J.A. Burger, Functional expression of CXCR4 (CD184) on small-cell lung cancer cells mediates migration, integrin activation, and adhesion to stromal cells, *Oncogene* 22 (50) (2003) 8093–8101, <https://doi.org/10.1038/sj.onc.1207097>.
- [4] I. Petit, D. Jin, S. Rafii, The SDF-1-CXCR4 signaling pathway: a molecular hub modulating neo-angiogenesis, *Trends Immunol.* 28 (7) (2007) 299–307, <https://doi.org/10.1016/j.it.2007.05.007>.
- [5] B.A. Teicher, S.P. Fricker, CXCL12 (SDF-1)/CXCR4 pathway in cancer, *Clin. Cancer Res.* 16 (11) (2010) 2927–2931, <https://doi.org/10.1158/1078-0432.CCR-09-2329>.
- [6] V. Saini, A. Marchese, M. Majetschak, CXC chemokine receptor 4 is a cell surface receptor for extracellular ubiquitin, *J. Biol. Chem.* 285 (20) (2010) 15566–15576, <https://doi.org/10.1074/jbc.M110.103408>.
- [7] V. Saini, D.M. Staren, J.J. Ziarek, Z.N. Nashaat, E.M. Campbell, B.F. Volkman, A. Marchese, M. Majetschak, The CXC chemokine receptor 4 ligands ubiquitin and stromal cell-derived factor-1alpha function through distinct receptor interactions, *J. Biol. Chem.* 286 (38) (2011) 33466–33477, <https://doi.org/10.1074/jbc.M111.233742>.
- [8] J. Bernhagen, R. Krohn, H. Lue, J.L. Gregory, A. Zerneck, R.R. Koenen, M. Dewor, I. Georgiev, A. Schober, L. Leng, T. Kooistra, G. Fingerle-Rowson, P. Ghezzi, R. Kleemann, S.R. McColl, R. Bucala, M.J. Hickey, C. Weber, MIF is a noncognate ligand of CXC chemokine receptors in inflammatory and atherogenic cell recruitment, *Nat. Med.* 13 (5) (2007) 587–596, <https://doi.org/10.1038/nm1567>.
- [9] S. Ahmad, S. Manzoor, S. Siddiqui, N. Mariappan, I. Zafar, A. Ahmad, A. Ahmad, Epigenetic underpinnings of inflammation: connecting the dots between pulmonary diseases, lung cancer and COVID-19, *Semin. Cancer Biol.* (2021), <https://doi.org/10.1016/j.semcancer.2021.01.003>.
- [10] K. Patil, S. Kuttikrishnan, A.Q. Khan, F. Ahmad, M. Alam, J. Buddenkotte, A. Ahmad, M. Steinhoff, S. Uddin, Molecular pathogenesis of Cutaneous T cell Lymphoma: role of chemokines, cytokines, and dysregulated signaling pathways, *Semin. Cancer Biol.* (2021), <https://doi.org/10.1016/j.semcancer.2021.12.003>.
- [11] M.D. Turner, B. Nedjai, T. Hurst, D.J. Pennington, Cytokines and chemokines: at the crossroads of cell signalling and inflammatory disease, *Biochim. Biophys. Acta* 1843 (11) (2014) 2563–2582, <https://doi.org/10.1016/j.bbamec.2014.05.014>.
- [12] A. Ahmad, S. Banerjee, Z. Wang, D. Kong, A.P. Majumdar, F.H. Sarkar, Aging and inflammation: etiological culprits of cancer, *Curr. Aging Sci.* 2 (3) (2009) 174–186, <https://doi.org/10.2174/1874609810902030174>.
- [13] J.M. Zhang, J. An, Cytokines, inflammation, and pain, *Int. Anesthesiol. Clin.* 45 (2) (2007) 27–37, <https://doi.org/10.1097/AIA.0b013e318034194e>.
- [14] B. Moser, K. Willmann, Chemokines: role in inflammation and immune surveillance, *Ann. Rheum. Dis.* 63 (Suppl 2) (2004) ii84–ii89, <https://doi.org/10.1136/ard.2004.028316>.
- [15] M.E. Bianchi, R. Mezzapelle, The chemokine receptor CXCR4 in cell proliferation and tissue regeneration, *Front. Immunol.* 11 (2020) 2109, <https://doi.org/10.3389/fimmu.2020.02109>.
- [16] J. Zuo, M. Wen, S. Li, X. Lv, L. Wang, X. Ai, M. Lei, Overexpression of CXCR4 promotes invasion and migration of non-small cell lung cancer via EGFR and MMP-9, *Oncol. Lett.* 14 (6) (2017) 7513–7521, <https://doi.org/10.3892/ol.2017.7168>.
- [17] G.D. Luker, J. Yang, A. Richmond, S. Scala, C. Festuccia, M. Schottelius, H. J. Wester, J. Zimmermann, At the bench: pre-clinical evidence for multiple functions of CXCR4 in cancer, *J. Leukoc. Biol.* 109 (5) (2021) 969–989, <https://doi.org/10.1002/JLB.2BT1018-715RR>.
- [18] H. Zhao, L. Guo, H. Zhao, J. Zhao, H. Weng, B. Zhao, CXCR4 over-expression and survival in cancer: a system review and meta-analysis, *Oncotarget* 6 (7) (2015) 5022–5040, <https://doi.org/10.18632/oncotarget.3217>.
- [19] B. Furusato, A. Mohamed, M. Uhlen, J.S. Rhim, CXCR4 and cancer, *Pathol. Int.* 60 (7) (2010) 497–505, <https://doi.org/10.1111/j.1440-1827.2010.02548.x>.
- [20] X. Sun, G. Cheng, M. Hao, J. Zheng, X. Zhou, J. Zhang, R.S. Taichman, K.J. Pienta, J. Wang, CXCL12 / CXCR4 / CXCR7 chemokine axis and cancer progression, *Cancer Metastasis Rev.* 29 (4) (2010) 709–722, <https://doi.org/10.1007/s10555-010-9256-x>.
- [21] Y. Shi, D.J. Riese 2nd, J. Shen, The role of the CXCL12/CXCR4/CXCR7 chemokine axis in cancer, *Front. Pharmacol.* 11 (2020), 574667, <https://doi.org/10.3389/fphar.2020.574667>.
- [22] M.P. Wescott, I. Kufareva, C. Paes, J.R. Goodman, Y. Thaker, B.A. Puffer, E. Berdoug, J.B. Rucker, T.M. Handel, B.J. Doranz, Signal transmission through the CXC chemokine receptor 4 (CXCR4) transmembrane helices, *Proc. Natl. Acad. Sci. USA* 113 (35) (2016) 9928–9933, <https://doi.org/10.1073/pnas.1601278113>.
- [23] C.C. Bleul, L. Wu, J.A. Hoxie, T.A. Springer, C.R. Mackay, The HIV coreceptors CXCR4 and CCR5 are differentially expressed and regulated on human T lymphocytes, *Proc. Natl. Acad. Sci. USA* 94 (5) (1997) 1925–1930, <https://doi.org/10.1073/pnas.94.5.1925>.
- [24] S. Liu, Q. Wang, X. Yu, Y. Li, Y. Guo, Z. Liu, F. Sun, W. Hou, C. Li, L. Wu, D. Guo, S. Chen, HIV-1 inhibition in cells with CXCR4 mutant genome created by CRISPR-Cas9 and piggyBac recombinant technologies, *Sci. Rep.* 8 (1) (2018) 8573, <https://doi.org/10.1038/s41598-018-26894-4>.
- [25] É. Midavaine, J. Côté, P. Sarret, The multifaceted roles of the chemokines CCL2 and CXCL12 in osteophilic metastatic cancers, *Cancer Metastasis Rev.* 40 (2) (2021) 427–445, <https://doi.org/10.1007/s10555-021-09974-2>.
- [26] Y. Lavrovsky, Y.A. Ivanenkov, K.V. Balakin, D.A. Medvedeva, A.V. Ivachtchenko, CXCR4 receptor as a promising target for oncolytic drugs, *Mini Rev. Med. Chem.* 8 (11) (2008) 1075–1087, <https://doi.org/10.2174/138955708785909907>.
- [27] L. Portella, A.M. Bello, S. Scala, CXCL12 signaling in the tumor microenvironment, *Adv. Exp. Med. Biol.* 1302 (2021) 51–70, https://doi.org/10.1007/978-3-030-62658-7_5.
- [28] M. Cojoc, C. Peitzsch, F. Trautmann, L. Polishchuk, G.D. Telegeev, A. Dubrovskaya, Emerging targets in cancer management: role of the CXCL12/CXCR4 axis, *Onco Targets Ther.* 6 (2013) 1347–1361, <https://doi.org/10.2147/ott.S36109>.

- [29] A. Ahmad, W.A. Sakr, K.M. Rahman, Novel targets for detection of cancer and their modulation by chemopreventive natural compounds, *Front. Biosci.* 4 (2012) 410–425, <https://doi.org/10.2741/388>.
- [30] S. Chatterjee, B. Behnam Azad, S. Nimmagadda, The intricate role of CXCR4 in cancer, *Adv. Cancer Res.* 124 (2014) 31–82, <https://doi.org/10.1016/B978-0-12-411638-2.00002-1>.
- [31] F. Guo, Y. Wang, J. Liu, S.C. Mok, F. Xue, W. Zhang, CXCL12/CXCR4: a symbiotic bridge linking cancer cells and their stromal neighbors in oncogenic communication networks, *Oncogene* 35 (7) (2016) 816–826, <https://doi.org/10.1038/ncr.2015.139>.
- [32] H. Zhang, P. Wang, X. Zhang, W. Zhao, H. Ren, Z. Hu, SDF1/CXCR4 axis facilitates the angiogenesis via activating the PI3K/AKT pathway in degenerated discs, *Mol. Med. Rep.* 22 (5) (2020) 4163–4172, <https://doi.org/10.3892/mmr.2020.11498>.
- [33] T. Kou, D. Sha, F. Wang, T. He, X. He, The novel target of colorectal carcinoma: TRIM44 regulates cell migration and invasion via activation of CXCR4/NF-kappaB signaling, *Cell. Biochem. Biophys.* 79 (1) (2021) 113–121, <https://doi.org/10.1007/s12013-020-00955-w>.
- [34] B. Chung, A.A. Esmaili, S. Gopalakrishna-Pillai, J.P. Murad, E.S. Andersen, N. Kumar Reddy, G. Srinivasan, B. Armstrong, C. Chu, Y. Kim, T. Tong, J. Waisman, J.H. Yim, B. Badie, P.P. Lee, Human brain metastatic stroma attracts breast cancer cells via chemokines CXCL16 and CXCL12, *NPJ Breast Cancer* 3 (2017) 6, <https://doi.org/10.1038/s41523-017-0008-8>.
- [35] R.L. Siegel, K.D. Miller, H.E. Fuchs, A. Jemal, Cancer statistics, 2022, *CA Cancer J. Clin.* 72 (1) (2022) 7–33, <https://doi.org/10.3322/caac.21708>.
- [36] H. Sung, J. Ferlay, R.L. Siegel, M. Laversanne, I. Soerjomataram, A. Jemal, F. Bray, Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries, *CA Cancer J. Clin.* 71 (3) (2021) 209–249, <https://doi.org/10.3322/caac.21660>.
- [37] Z. Cong, H. Wu, Z. Guo, T. Qin, Y. Xu, H. Jing, Y. Wang, Y. Shen, High expression of C-X-C chemokine receptor 4 and Notch1 is predictive of lymphovascular invasion and poor prognosis in lung adenocarcinoma, *Tumour Biol.* 39 (6) (2017), <https://doi.org/10.1177/1010428317708698>.
- [38] Q. Zhu, R. Luo, J. Gu, Y. Hou, Z. Chen, F. Xu, L. Wang, W. Mao, C. Lu, D. Ge, High CXCR4 expression predicts a poor prognosis in resected lung adenocarcinoma, *J. Cancer* 11 (4) (2020) 810–818, <https://doi.org/10.7150/jca.36498>.
- [39] M. Goto, T. Yoshida, Y. Yamamoto, Y. Furukita, S. Inoue, S. Fujiwara, N. Kawakita, T. Nishino, T. Minato, Y. Yuasa, H. Yamai, H. Takechi, J. Seike, Y. Bando, A. Tangoku, CXCR4 expression is associated with poor prognosis in patients with esophageal squamous cell carcinoma, *Ann. Surg. Oncol.* 24 (3) (2017) 832–840, <https://doi.org/10.1245/s10434-015-4974-5>.
- [40] X. Yang, Q. Lu, Y. Xu, C. Liu, Q. Sun, Clinicopathologic significance of CXCR4 expressions in patients with esophageal squamous cell carcinoma, *Pathol. Res. Pract.* 216 (1) (2020), 152787, <https://doi.org/10.1016/j.prp.2019.152787>.
- [41] S. Yu, T. Wu, J. Wang, C. Cheng, J. Wang, L. Sun, C. Liu, G. Cao, T. Hu, Combined evaluation of expression of CXCR4 and Nrf2 as prognostic factor for patients with gastric carcinoma, *Anticancer Agents Med. Chem.* 18 (3) (2018) 388–393, <https://doi.org/10.2174/187152061766617110312019>.
- [42] G. Chen, Z. Zhou, J. Jin, Y. Zhou, Y. Liu, W. Wang, CXCR4 is a prognostic marker that inhibits the invasion and migration of gastric cancer by regulating VEGF expression, *Oncol. Lett.* 22 (2) (2021) 587, <https://doi.org/10.3892/ol.2021.12848>.
- [43] T.J.W. Klein Nulent, R.J.J. van Es, M.H. Valstar, L.E. Smeel, L.A. Smit, R. Klein Gunnewiek, N.P.A. Zuihthoff, B. de Keizer, R. de Bree, S.M. Willems, High CXCR4 expression in adenoid cystic carcinoma of the head and neck is associated with increased risk of locoregional recurrence, *J. Clin. Pathol.* 73 (8) (2020) 476–482, <https://doi.org/10.1136/jclinpath-2019-206273>.
- [44] N. Rusetska, K. Kowalski, K. Zalewski, S. Zięba, M. Bidziński, K. Goryca, B. Kotowicz, M. Fuksiewicz, J. Kopczyński, E. Bakula-Zalewska, A. Kowalik, M. Kowalewska, CXCR4/ACKR3/CXCL12 axis in the lymphatic metastasis of vulvar squamous cell carcinoma, *J. Clin. Pathol.* (2021), <https://doi.org/10.1136/jclinpath-2020-206917>.
- [45] K. Sirakriengkrai, S. Tepmongkol, S. Keelawat, U. Techavijit, Clinical association of CXCR4 in primary tumor of papillary thyroid cancer and response to iodine-131 treatment, *Nucl. Med. Commun.* 42 (4) (2021) 396–401, <https://doi.org/10.1097/mnm.0000000000001340>.
- [46] X. Cao, J. Zhu, X. Li, Y. Ma, Q. He, Expression of CXCR4 and CXCR7 in papillary thyroid carcinoma and adjacent tissues and their relationship with pathologic indicators of tumor aggressiveness, *Endocr. J.* (2021), <https://doi.org/10.1507/endoerj.EJ21-0076>.
- [47] Y. Ding, Y. Du, Clinicopathological significance and prognostic role of chemokine receptor CXCR4 expression in pancreatic ductal adenocarcinoma, a meta-analysis and literature review, *Int. J. Surg.* 65 (2019) 32–38, <https://doi.org/10.1016/j.ijss.2019.03.009>.
- [48] J. Urosevic, M.T. Blasco, A. Llorente, A. Bellmunt, A. Berenguer-Llargo, M. Guiu, A. Cañellas, E. Fernandez, I. Burkov, M. Clapés, M. Cartanà, C. Figueras-Puig, E. Batlle, A.R. Nebreda, R.R. Gomis, ERK1/2 signaling induces upregulation of ANGPT2 and CXCR4 to mediate liver metastasis in colon cancer, *Cancer Res.* 80 (21) (2020) 4668–4680, <https://doi.org/10.1158/0008-5472.Can-19-4028>.
- [49] D.D. Cave, X. Hernando-Mombolona, M. Sevillano, G. Minchiotti, E. Lonardo, Nodal-induced L1CAM/CXCR4 subpopulation sustains tumor growth and metastasis in colorectal cancer derived organoids, *Theranostics* 11 (12) (2021) 5686–5699, <https://doi.org/10.7150/thno.54027>.
- [50] B. Saka, O. Ekinici, A. Dursun, N. Akyurek, Clinicopathologic and prognostic significance of immunohistochemical expression of HIF-1 α , CXCR4 and CA9 in colorectal carcinoma, *Pathol. Res. Pract.* 213 (7) (2017) 783–792, <https://doi.org/10.1016/j.prp.2017.04.001>.
- [51] N. Yosuantari, D.S. Heriyanto, S.H. Hutajulu, J. Kurnianda, A. Ghozali, Clinicopathologic significance of CXCL12 and CXCR4 expressions in patients with colorectal cancer, *Gastroenterol. Res. Pract.* 2018 (2018), 9613185, <https://doi.org/10.1155/2018/9613185>.
- [52] A. Mitchell, S.L. Hasanali, D.S. Morera, R. Baskar, X. Wang, R. Khan, A. Talukder, C.S. Li, M. Manoharan, A.R. Jordan, J. Wang, R.J. Bollag, N. Singh, D. Albo, S. Ghosh, V.B. Lokeshwar, A chemokine/chemokine receptor signature potentially predicts clinical outcome in colorectal cancer patients, *Cancer Biomark.* 26 (3) (2019) 291–301, <https://doi.org/10.3233/cbm-190210>.
- [53] P. Yang, Y. Hu, Q. Zhou, The CXCL12-CXCR4 signaling axis plays a key role in cancer metastasis and is a potential target for developing novel therapeutics against metastatic cancer, *Curr. Med. Chem.* 27 (33) (2020) 5543–5561, <https://doi.org/10.2174/0929867326666191113113110>.
- [54] Y. Dai, R. Tong, H. Guo, T. Yu, C. Wang, Association of CXCR4, CCR7, VEGF-C and VEGF-D expression with lymph node metastasis in patients with cervical cancer, *Eur. J. Obstet. Gynecol. Reprod. Biol.* 214 (2017) 178–183, <https://doi.org/10.1016/j.ejogrb.2017.04.043>.
- [55] M.M. Bi, B. Shang, Z. Wang, G. Chen, Expression of CXCR4 and VEGF-C is correlated with lymph node metastasis in non-small cell lung cancer, *Thorac. Cancer* 8 (6) (2017) 634–641, <https://doi.org/10.1111/1759-7714.12500>. PubMed PMID: 28925100; PMCID: PMC5668524.
- [56] Y.L. Li, Y.F. Li, H.F. Li, H.Q. Lv, D.Z. Sun, Role of SDF-1 α /CXCR4 signaling pathway in clinicopathological features and prognosis of patients with nasopharyngeal carcinoma, *Biosci. Rep.* 37 (4) (2017), <https://doi.org/10.1042/bsr20170144>.
- [57] J.A. van Ipenburg, N.E. de Waard, N.C. Naus, M.J. Jager, D. Paridaens, R. M. Verdijk, Chemokine receptor expression pattern correlates to progression of conjunctival melanocytic lesions, *Investig. Ophthalmol. Vis. Sci.* 60 (8) (2019) 2950–2957, <https://doi.org/10.1167/iovs.19.27162>.
- [58] M. Alimohammadi, A. Rahimi, F. Faramarzi, R. Alizadeh-Navaei, A. Rafiei, Overexpression of chemokine receptor CXCR4 predicts lymph node metastatic risk in patients with melanoma: a systematic review and meta-analysis, *Cytokine* 148 (2021), 155691, <https://doi.org/10.1016/j.cyto.2021.155691>.
- [59] A. Mardani, E. Gheyanchi, S.H. Mousavie, Z. Madjd Jabari, T. Shooshtarizadeh, Clinical significance of cancer stem cell markers CD133 and CXCR4 in osteosarcomas, *Asian Pac. J. Cancer Prev.* 21 (1) (2020) 67–73, <https://doi.org/10.31557/apjcp.2020.21.1.67>.
- [60] R. Caspa Gokulan, H. Devaraj, Stem cell markers CXCR-4 and CD133 predict aggressive phenotype and their double positivity indicates poor prognosis of oral squamous cell carcinoma, in: *Cancers*, 13, 2021, <https://doi.org/10.3390/cancers13235895>.
- [61] T.A. Werner, C.M. Forster, L. Dizdar, P.E. Verde, K. Raba, M. Schott, W.T. Knoefel, A. Krieg, CXCR4/CXCR7/CXCL12-axis in follicular thyroid carcinoma, *J. Cancer* 9 (6) (2018) 929–940, <https://doi.org/10.7150/jca.23042>.
- [62] C. Raschioni, G. Bottai, A. Sagona, V. Errico, A. Testori, W. Gatzemeier, F. Corsi, C. Tinterri, M. Roncalli, L. Santarpia, L. Di Tommaso, CXCR4/CXCL12 signaling and proinflammatory macrophages in primary tumors and sentinel lymph nodes are involved in luminal B breast cancer progression, *Dis. Markers* 2018 (2018), 5018671, <https://doi.org/10.1155/2018/5018671>.
- [63] R. Gadalla, H. Hassan, S.A. Ibrahim, M.S. Abdullah, A. Gaballah, B. Greve, S. El-Deeb, M. El-Shinawi, M.M. Mohamed, Tumor microenvironmental plasmacytoid dendritic cells contribute to breast cancer lymph node metastasis via CXCR4/SDF-1 axis, *Breast Cancer Res. Treat.* 174 (3) (2019) 679–691, <https://doi.org/10.1007/s10549-019-05129-8>.
- [64] M. Goto, Y. Shibahara, C. Baciu, F. Allison, J.C. Yeung, G.E. Darling, M. Liu, Prognostic impact of CXCR7 and CXCL12 expression in patients with esophageal adenocarcinoma, *Ann. Surg. Oncol.* 28 (9) (2021) 4943–4951, <https://doi.org/10.1245/s10434-021-09775-5>.
- [65] C. Gong, K. Sun, H.H. Xiong, T. Sneh, J. Zhang, X. Zhou, P. Yan, J.H. Wang, Expression of CXCR4 and MMP-2 is associated with poor prognosis in patients with osteosarcoma, *Histol. Histopathol.* 35 (8) (2020) 863–870, <https://doi.org/10.14670/hh-18-219>.
- [66] M. Ogawa, M. Watanabe, T. Hasegawa, K. Ichihara, K. Yoshida, K. Yanaga, Expression of CXCR-4 and IDO in human colorectal cancer: an immunohistochemical approach, *Mol. Clin. Oncol.* 6 (5) (2017) 701–704, <https://doi.org/10.3892/mco.2017.1207>.
- [67] Y.P. Li, J. Pang, S. Gao, P.Y. Bai, W.D. Wang, P. Kong, Y. Cui, Role of CXCR4 and SDF1 as prognostic factors for survival and the association with clinicopathology in colorectal cancer: a systematic meta-analysis, *Tumour Biol.* 39 (6) (2017), <https://doi.org/10.1177/1010428317706206>.
- [68] A. Ottaiano, S. Scala, N. Normanno, G. Botti, F. Tatangelo, A. Di Mauro, M. Capozzi, S. Facchini, S. Tafuto, G. Nasti, Prognostic and predictive role of CXCR4 chemokine receptor 4 in metastatic colorectal cancer patients, *Appl. Immunohistochem. Mol. Morphol.* 28 (10) (2020) 755–760, <https://doi.org/10.1097/pai.0000000000000828>.
- [69] S. Kim, M.K. Yeo, J.S. Kim, J.Y. Kim, K.H. Kim, Elevated CXCL12 in the plasma membrane of locally advanced rectal cancer after neoadjuvant chemoradiotherapy: a potential prognostic marker, *J. Cancer* 13 (1) (2022) 162–173, <https://doi.org/10.7150/jca.64082>.
- [70] S. Matsusaka, S. Cao, D.L. Hanna, Y. Sunakawa, M. Ueno, N. Mizunuma, W. Zhang, D. Yang, Y. Ning, S. Stintzing, A. Sebilo, S. Stremtizer, S. Yamauchi, A. Parekh, S. Okazaki, M.D. Berger, R. El-Khoukiry, A. Mendez, W. Ichikawa, F. Loupakis, H.J. Lenz, CXCR4 polymorphism predicts progression-free survival in metastatic colorectal cancer patients treated with first-line bevacizumab-based

- chemotherapy, *Pharm. J.* 17 (6) (2017) 543–550, <https://doi.org/10.1038/tj.2016.59>.
- [71] A.L. Guembarovski, R.L. Guembarovski, B.K.B. Hirata, G.A.F. Vitiello, K. M. Suzuki, M.T. Enokida, M.A.E. Watanabe, E.M.V. Reiche, CXCL12 chemokine and CXCR4 receptor: association with susceptibility and prognostic markers in triple negative breast cancer, *Mol. Biol. Rep.* 45 (5) (2018) 741–750, <https://doi.org/10.1007/s11033-018-4215-7>.
- [72] M. Katsura, F. Shoji, T. Okamoto, S. Shimamatsu, F. Hirai, G. Toyokawa, Y. Morodomi, T. Tagawa, Y. Oda, Y. Maehara, Correlation between CXCR4/CXCR7/CXCL12 chemokine axis expression and prognosis in lymph-node-positive lung cancer patients, *Cancer Sci.* 109 (1) (2018) 154–165, <https://doi.org/10.1111/cas.13422>.
- [73] C. De-Colle, D. Mönnich, S. Welz, S. Boeke, B. Sipos, F. Fend, P.S. Mauz, I. Tinhof, V. Budach, J.A. Jawad, M. Stuschke, P. Balermphas, C. Rödel, A. L. Grosu, A. Abdollahi, J. Debus, C. Bayer, C. Belka, S. Pigorsch, S.E. Combs, F. Lohaus, A. Linge, M. Krause, M. Baumann, D. Zips, A. Menegakis, SDF-1/CXCR4 expression in head and neck cancer and outcome after postoperative radiochemotherapy, *Clin. Transl. Radiat. Oncol.* 5 (2017) 28–36, <https://doi.org/10.1016/j.ctro.2017.06.004>.
- [74] A.P. Ferragut Cardoso, M. Banerjee, A.N. Nail, A. Lykoudi, J.C. States, miRNA dysregulation is an emerging modulator of genomic instability, *Semin. Cancer Biol.* 76 (2021) 120–131, <https://doi.org/10.1016/j.semcancer.2021.05.004>.
- [75] G. Shommo, B. Apolloni, A holistic miRNA-mRNA module discovery, *Noncoding RNA Res.* 6 (4) (2021) 159–166, <https://doi.org/10.1016/j.ncrna.2021.09.001>.
- [76] S. Ahmad, M. Abbas, M.F. Ullah, M.H. Aziz, O. Beylerli, M.A. Alam, M.A. Syed, S. Uddin, A. Ahmad, Long non-coding RNAs regulated NF-kappaB signaling in cancer metastasis: micromanaging by not so small non-coding RNAs, *Semin. Cancer Biol.* (2021), <https://doi.org/10.1016/j.semcancer.2021.07.015>.
- [77] I. Gareev, Y. Gileva, A. Dzidzaria, O. Beylerli, V. Pavlov, M. Agavardiev, B. Mazorov, I. Biganyakov, A. Vardikyan, M. Jin, A. Ahmad, Long non-coding RNAs in oncurology, *Noncoding RNA Res.* 6 (3) (2021) 139–145, <https://doi.org/10.1016/j.ncrna.2021.08.001>.
- [78] A. Beilerli, I. Gareev, O. Beylerli, G. Yang, V. Pavlov, G. Aliev, A. Ahmad, Circular RNAs as biomarkers and therapeutic targets in cancer, *Semin. Cancer Biol.* (2021), <https://doi.org/10.1016/j.semcancer.2020.12.026>.
- [79] A.A. Farooqi, K. Gulnara, A.A. Mukhanbetzhanovna, U. Datkhayev, A. Z. Kussainov, A. Adylova, Regulation of RUNX proteins by long non-coding RNAs and circular RNAs in different cancers, *Noncoding RNA Res.* 6 (2) (2021) 100–106, <https://doi.org/10.1016/j.ncrna.2021.05.001>.
- [80] X. Ke, Y. Yuan, C. Guo, Y. Yang, Q. Pu, X. Hu, K. Tang, X. Luo, Q. Jiang, X. Su, L. Liu, W. Zhu, Y. Wei, MiR-410 induces stemness by inhibiting Gsk3beta but upregulating beta-catenin in non-small cell lung cancer, *Oncotarget* 8 (7) (2017) 11356–11371, <https://doi.org/10.18632/oncotarget.14529>.
- [81] B. Park, B. Sung, V.R. Yadav, S.G. Cho, M. Liu, B.B. Aggarwal, Acetyl-11-keto-beta-boswellic acid suppresses invasion of pancreatic cancer cells through the downregulation of CXCR4 chemokine receptor expression, *Int. J. Cancer* 129 (1) (2011) 23–33, <https://doi.org/10.1002/ijc.25966>.
- [82] J. O'Brien, H. Hayder, Y. Zayed, C. Peng, Overview of MicroRNA biogenesis, mechanisms of actions, and circulation, *Front. Endocrinol.* 9 (2018) 402, <https://doi.org/10.3389/fendo.2018.00402>.
- [83] A. Ahmad, Y. Li, B. Bao, D. Kong, F.H. Sarkar, Epigenetic regulation of miRNA-cancer stem cells nexus by nutraceuticals, *Mol. Nutr. Food Res.* 58 (1) (2014) 79–86, <https://doi.org/10.1002/mnfr.201300528>.
- [84] A. Valinezhad Orang, R. Safaralizadeh, M. Kazemzadeh-Bavili, Mechanisms of miRNA-mediated gene regulation from common downregulation to mRNA-specific upregulation, *Int. J. Genom.* 2014 (2014), 970607, <https://doi.org/10.1155/2014/970607>.
- [85] R. Afshar-Khamesh, A. Javeri, M.F. Taha, MiR-146a suppresses the expression of CXCR4 and alters survival, proliferation and migration rate in colorectal cancer cells, *Tissue Cell* 73 (2021), 101654, <https://doi.org/10.1016/j.tice.2021.101654>.
- [86] M. Azar, H. Aghazadeh, H.N. Mohammed, M.R.S. Sara, A. Hosseini, N. Shomali, R. Tamjidifar, S. Tarzi, M. Mansouri, S.P. Sarand, F. Marofi, M. Akbari, H. Xu, S. S. Shotorbani, miR-193a-5p as a promising therapeutic candidate in colorectal cancer by reducing 5-FU and Oxaliplatin chemoresistance by targeting CXCR4, *Int. Immunopharmacol.* 92 (2021), 107355, <https://doi.org/10.1016/j.intimp.2020.107355>.
- [87] X. Zheng, Y.F. Ma, X.R. Zhang, Y. Li, H.H. Zhao, S.G. Han, Circ_0056618 promoted cell proliferation, migration and angiogenesis through sponging with miR-206 and upregulating CXCR4 and VEGF-A in colorectal cancer, *Eur. Rev. Med. Pharmacol. Sci.* 24 (8) (2020) 4190–4202, https://doi.org/10.26355/eurrev_202004_20999.
- [88] Y. Fang, B. Sun, J. Wang, Y. Wang, miR-622 inhibits angiogenesis by suppressing the CXCR4-VEGFA axis in colorectal cancer, *Gene* 699 (2019) 37–42, <https://doi.org/10.1016/j.gene.2019.03.004>.
- [89] H. Liu, Y. Liu, W. Liu, W. Zhang, J. Xu, EZH2-mediated loss of miR-622 determines CXCR4 activation in hepatocellular carcinoma, *Nat. Commun.* 6 (2015) 8494, <https://doi.org/10.1038/ncomms9494>.
- [90] D. Di Paolo, F. Pontis, M. Moro, G. Centonze, G. Bertolini, M. Milione, M. Mensah, M. Segale, I. Petrarola, C. Borzi, P. Suatoni, C. Brignole, P. Perri, M. Ponzoni, U. Pastorino, G. Sozzi, C. Fortunato, Cotargeting of miR-126-3p and miR-221-3p inhibits PIK3R2 and PTEN, reducing lung cancer growth and metastasis by blocking AKT and CXCR4 signalling, *Mol. Oncol.* 15 (11) (2021) 2969–2988, <https://doi.org/10.1002/1878-0261.13036>.
- [91] C.W. Cheng, W.L. Liao, P.M. Chen, J.C. Yu, H.P. Shiau, Y.H. Hsieh, H.J. Lee, Y. C. Cheng, P.E. Wu, C.Y. Shen, MiR-139 modulates cancer stem cell function of human breast cancer through targeting CXCR4, in: *Cancers*, 13, 2021, <https://doi.org/10.3390/cancers13112582>.
- [92] B. Mansoori, N. Silvestris, A. Mohammadi, V. Khaze, E. Baghbani, A. Mokhtarzadeh, D. Shanebandi, A. Derakhshani, P.H.G. Duijf, B. Baradaran, miR-34a and miR-200c have an additive tumor-suppressive effect on breast cancer cells and patient prognosis, in: *Genes*, 12, 2021, <https://doi.org/10.3390/genes12020267>.
- [93] A. Singh, N. Srivastava, A. Yadav, B. Ateeq, Targeting AGTR1/NF-kappaB/CXCR4 axis by miR-155 attenuates oncogenesis in glioblastoma, *Neoplasia* 22 (10) (2020) 497–510, <https://doi.org/10.1016/j.neo.2020.08.002>.
- [94] J. Xiao, H. Lai, S.H. Wei, Z.S. Ye, F.S. Gong, L.C. Chen, lncRNA HOTAIR promotes gastric cancer proliferation and metastasis via targeting miR-126 to active CXCR4 and RhoA signaling pathway, *Cancer Med.* 8 (15) (2019) 6768–6779, <https://doi.org/10.1002/cam4.1302>.
- [95] S. Gao, Y. Cai, H. Zhang, F. Hu, L. Hou, Q. Xu, Long noncoding RNA DLEU1 aggravates pancreatic ductal adenocarcinoma carcinogenesis via the miR-381/CXCR4 axis, *J. Cell Physiol.* 234 (5) (2019) 6746–6757, <https://doi.org/10.1002/jcp.27421>.
- [96] S.A. Nabipoorashrafi, N. Shomali, L. Sadat-Hatamnezhad, M. Mahami-Oskouei, J. Mahmoudi, B. Sandoghchian Shotorbani, M. Akbari, H. Xu, Sandoghchian, S. Shotorbani, miR-143 acts as an inhibitor of migration and proliferation as well as an inducer of apoptosis in melanoma cancer cells in vitro, *IUBMB Life* 72 (9) (2020) 2034–2044, <https://doi.org/10.1002/iub.2345>.
- [97] A.H. Mesgarzadeh, M. Aali, F. Farhadi, S. Noorolyai, E. Baghbani, F. Mohammadnejad, B. Baradaran, Transfection of microRNA-143 mimic could inhibit migration of HN-5 cells through down-regulating of metastatic genes, *Gene* 716 (2019), 144033, <https://doi.org/10.1016/j.gene.2019.144033>.
- [98] N. Sehati, N. Sadeghie, B. Mansoori, A. Mohammadi, D. Shanebandi, B. Baradaran, MicroRNA-330 inhibits growth and migration of melanoma A375 cells: in vitro study, *J. Cell Biochem.* 121 (1) (2020) 458–467, <https://doi.org/10.1002/jcb.29211>.
- [99] G. Zong, J. Han, Z. Yue, Y. Liu, Z. Cui, L. Shi, Downregulation of miR-204 facilitates the progression of nasopharyngeal carcinoma by targeting CXCR4 through NF-kappaB signaling pathway, *J. BUON* 25 (2) (2020) 1098–1104.
- [100] Q. Zhao, J.Y. Li, J. Zhang, Y.X. Long, Y.J. Li, X.D. Guo, M.N. Wei, W.J. Liu, Role of visfatin in promoting proliferation and invasion of colorectal cancer cells by downregulating SDF-1/CXCR4-mediated miR-140-3p expression, *Eur. Rev. Med. Pharmacol. Sci.* 24 (10) (2020) 5367–5377, https://doi.org/10.26355/eurrev_202005_21320.
- [101] C. Yao, S. Huang, J. Wu, L. Yin, X. Jiang, C. Chen, W. Wu, J. Xu, X. He, MicroRNA-140 inhibits tumor progression in nasopharyngeal carcinoma by targeting CXCR4, *Int. J. Clin. Exp. Pathol.* 10 (7) (2017) 7750–7759.
- [102] P.F. Zhang, X. Pei, K.S. Li, L.N. Jin, F. Wang, J. Wu, X.M. Zhang, Circular RNA circGFR1 promotes progression and anti-PD-1 resistance by sponging miR-381-3p in non-small cell lung cancer cells, *Mol. Cancer* 18 (1) (2019) 179, <https://doi.org/10.1186/s12943-019-1111-2>.
- [103] L. Pazzaglia, S. Pollino, M. Vitale, E. Bientinesi, S. Benini, C. Ferrari, E. Palmerini, M. Gambarotti, P. Picci, M.S. Benassi, miR494.3p expression in synovial sarcoma: role of CXCR4 as a potential target gene, *Int. J. Oncol.* 54 (1) (2019) 361–369, <https://doi.org/10.3892/ijo.2018.4627>.
- [104] N. Shirafkan, N. Shomali, T. Kazemi, D. Shanebandi, M. Ghasabi, E. Baghbani, M. Ganji, V. Khaze, B. Mansoori, B. Baradaran, microRNA-193a-5p inhibits migration of human HT-29 colon cancer cells via suppression of metastasis pathway, *J. Cell Biochem.* (2018), <https://doi.org/10.1002/jcb.28164>.
- [105] F.T. Duan, F. Qian, K. Fang, K.Y. Lin, W.T. Wang, Y.Q. Chen, miR-133b, a muscle-specific microRNA, is a novel prognostic marker that participates in the progression of human colorectal cancer via regulation of CXCR4 expression, *Mol. Cancer* 12 (2013) 164, <https://doi.org/10.1186/1476-4598-12-164>.
- [106] Y. Zhu, L. Tang, S. Zhao, B. Sun, L. Cheng, Y. Tang, Z. Luo, Z. Lin, J. Zhu, W. Zhu, R. Zhao, B. Lu, H. Long, CXCR4-mediated osteosarcoma growth and pulmonary metastasis is suppressed by MicroRNA-613, *Cancer Sci.* 109 (8) (2018) 2412–2422, <https://doi.org/10.1111/cas.13653>.
- [107] M. Zhang, C.E. Gao, W.L. Chen, Y.Y. Tang, J.Y. Nie, L.D. Shen, X. Ma, D.D. Chen, Opposite response to hypoxia by breast cancer cells between cell proliferation and cell migration: a clue from microRNA expression profile, *Oncol. Lett.* 15 (3) (2018) 2771–2780, <https://doi.org/10.3892/ol.2017.7636>.
- [108] X. Zheng, S. Wang, S. Hong, S. Liu, G. Chen, W. Tang, Y. Zhao, H. Gao, B. Cha, CXCR4/RhoA signaling pathway is involved in miR-128-regulated proliferation and apoptosis of human thyroid cancer cells, *Int. J. Clin. Exp. Pathol.* 10 (9) (2017) 9213–9222.
- [109] H.T. Liu, W.X. Fan, MiRNA-1246 suppresses the proliferation and migration of renal cell carcinoma through targeting CXCR4, *Eur. Rev. Med. Pharmacol. Sci.* 24 (11) (2020) 5979–5987, https://doi.org/10.26355/eurrev_202006_21491.
- [110] X. Xu, L. Cao, Y. Zhang, H. Lian, Z. Sun, Y. Cui, MicroRNA-1246 inhibits cell invasion and epithelial mesenchymal transition process by targeting CXCR4 in lung cancer cells, *Cancer Biomark.* 21 (2) (2018) 251–260, <https://doi.org/10.3233/CBM-170317>.
- [111] Y.C. Chien, J.N. Chen, Y.H. Chen, R.H. Chou, H.C. Lee, Y.L. Yu, Epigenetic silencing of miR-9 promotes migration and invasion by EZH2 in glioblastoma cells, in: *Cancers*, 12, 2020, <https://doi.org/10.3390/cancers12071781>.
- [112] B. Zhu, X. Xi, Q. Liu, Y. Cheng, H. Yang, MiR-9 functions as a tumor suppressor in acute myeloid leukemia by targeting CX chemokine receptor 4, *Am. J. Transl. Res.* 11 (6) (2019) 3384–3397.
- [113] V. Egea, K. Kessenbrock, D. Lawson, A. Bartelt, C. Weber, C. Ries, Let-7f miRNA regulates SDF-1alpha- and hypoxia-promoted migration of mesenchymal stem

- cells and attenuates mammary tumor growth upon exosomal release, *Cell Death Dis.* 12 (6) (2021) 516, <https://doi.org/10.1038/s41419-021-03789-3>.
- [114] G. Zhu, X. Liu, Y. Su, F. Kong, X. Hong, Z. Lin, Knockdown of urothelial carcinoma-associated 1 suppressed cell growth and migration through regulating miR-301a and CXCR4 in osteosarcoma MHCC97 cells, *Oncol. Res.* 27 (1) (2018) 55–64, <https://doi.org/10.3727/096504018X15201143705855>.
- [115] F. Li, Q. Yang, A.T. He, B.B. Yang, Circular RNAs in cancer: limitations in functional studies and diagnostic potential, *Semin. Cancer Biol.* 75 (2021) 49–61, <https://doi.org/10.1016/j.semcancer.2020.10.002>.
- [116] E. Arnaiz, C. Sole, L. Manterola, L. Iparraguirre, D. Otaegui, C.H. Lawrie, CircRNAs and cancer: biomarkers and master regulators, *Semin. Cancer Biol.* 58 (2019) 90–99, <https://doi.org/10.1016/j.semcancer.2018.12.002>.
- [117] H. Li, G. Yao, B. Feng, X. Lu, Y. Fan, Circ_0056618 and CXCR4 act as competing endogenous in gastric cancer by regulating miR-206, *J. Cell Biochem.* 119 (11) (2018) 9543–9551, <https://doi.org/10.1002/jcb.27271>.
- [118] R. Zhao, J. Ni, S. Lu, S. Jiang, L. You, H. Liu, J. Shou, C. Zhai, W. Zhang, S. Shao, X. Yang, H. Pan, W. Han, CircUBAP2-mediated competing endogenous RNA network modulates tumorigenesis in pancreatic adenocarcinoma, *Aging* 11 (19) (2019) 8484–8501, <https://doi.org/10.18632/aging.102334>.
- [119] A. Ahmad, Epigenetic regulation of immunosuppressive tumor-associated macrophages through dysregulated microRNAs, *Semin. Cell Dev. Biol.* (2021), <https://doi.org/10.1016/j.semcdb.2021.09.001>.
- [120] J. Zhao, X.D. Li, M. Wang, L.N. Song, M.J. Zhao, Circular RNA ABCB10 contributes to laryngeal squamous cell carcinoma (LSCC) progression by modulating the miR-588/CXCR4 axis, *Aging* 13 (10) (2021) 14078–14087, <https://doi.org/10.18632/aging.203025>. PubMed PMID: 34015764; PMCID: PMC8202875.
- [121] H. Si, H. Wang, H. Xiao, Y. Fang, Z. Wu, Anti-tumor effect of celastrol on hepatocellular carcinoma by the circ_SLIT3/miR-223-3p/CXCR4 axis, *Cancer Manag. Res.* 13 (2021) 1099–1111, <https://doi.org/10.2147/cmar.S278023>.
- [122] Q. Zhang, S. Chen, Y. Zhen, P. Gao, Z. Zhang, H. Guo, Y. Wang, Circular RNA circFGFR1 functions as an oncogene in glioblastoma cells through sponging to hsa-miR-224-5p, *J. Immunol. Res.* 2022 (2022), 7990251, <https://doi.org/10.1155/2022/7990251>.
- [123] K.D. Yang, Y. Wang, F. Zhang, B.H. Luo, D.Y. Feng, Z.J. Zeng, CircN4BP2L2 promotes colorectal cancer growth and metastasis through regulation of the miR-340-5p/CXCR4 axis, *Lab. Invest.* 102 (1) (2022) 38–47, <https://doi.org/10.1038/s41374-021-00632-3>.
- [124] F. He, X. Zhong, Z. Lin, J. Lin, M. Qiu, X. Li, Z. Hu, Plasma exo-hsa_circRNA_0056616: a potential biomarker for lymph node metastasis in lung adenocarcinoma, *J. Cancer* 11 (14) (2020) 4037–4046, <https://doi.org/10.7150/jca.30360>.
- [125] C.Y. Wei, M.X. Zhu, N.H. Lu, J.Q. Liu, Y.W. Yang, Y. Zhang, Y.D. Shi, Z.H. Feng, J. X. Li, F.Z. Qi, J.Y. Gu, Circular RNA circ_0020710 drives tumor progression and immune evasion by regulating the miR-370-3p/CXCL12 axis in melanoma, *Mol. Cancer* 19 (1) (2020) 84, <https://doi.org/10.1186/s12943-020-01191-9>.
- [126] A. Ahmad, P. Poltronieri, S. Uddin, Editorial: LncRNAs in cancer metastasis and therapy resistance, *Front. Oncol.* 11 (2021), 813274, <https://doi.org/10.3389/fonc.2021.813274>.
- [127] E. D'Angelo, M. Agostini, Long non-coding RNA and extracellular matrix: the hidden players in cancer-stroma cross-talking, *Noncoding RNA Res.* 3 (4) (2018) 174–177, <https://doi.org/10.1016/j.ncrna.2018.08.002>.
- [128] J. Samson, S. Cronin, K. Dean, BC200 (BCYRN1) - the shortest, long, non-coding RNA associated with cancer, *Noncoding RNA Res.* 3 (3) (2018) 131–143, <https://doi.org/10.1016/j.ncrna.2018.05.003>.
- [129] H. Helmsmoortel, C. Everaert, N. Lumen, P. Ost, J. Vandesompele, Detecting long non-coding RNA biomarkers in prostate cancer liquid biopsies: hype or hope, *Noncoding RNA Res.* 3 (2) (2018) 64–74, <https://doi.org/10.1016/j.ncrna.2018.05.001>.
- [130] X. Tan, Z. Huang, X. Li, Long non-coding RNA MALAT1 interacts with miR-204 to modulate human hilar cholangiocarcinoma proliferation, migration, and invasion by targeting CXCR4, *J. Cell Biochem.* 118 (11) (2017) 3643–3653, <https://doi.org/10.1002/jcb.25862>.
- [131] T. Liang, B. Wang, J. Li, Y. Liu, LINC00922 accelerates the proliferation, migration and invasion of lung cancer via the miRNA-204/CXCR4 axis, *Med. Sci. Monit.* 25 (2019) 5075–5086, <https://doi.org/10.12659/MSM.916327>.
- [132] Y. Wu, Q.W. Shen, Y.X. Niu, X.Y. Chen, H.W. Liu, X.Y. Shen, LncNORAD interference inhibits tumor growth and lung cancer cell proliferation, invasion and migration by down-regulating CXCR4 to suppress RhoA/ROCK signaling pathway, *Eur. Rev. Med. Pharmacol. Sci.* 24 (10) (2020) 5446–5455, <https://doi.org/10.26355/eurrev.202005.21329>.
- [133] Y. Zhang, Q. Zhang, M. Zhang, M. Yuan, Z. Wang, J. Zhang, X. Zhou, Y. Zhang, F. Lin, H. Na, S. Ren, Y. Zuo, DC - SIGNR by influencing the lncRNA HNRNPKP2 upregulates the expression of CXCR4 in gastric cancer liver metastasis, *Mol. Cancer* 16 (1) (2017) 78, <https://doi.org/10.1186/s12943-017-0639-2>.
- [134] X.Z. Dong, Z.R. Zhao, Y. Hu, Y.P. Lu, P. Liu, L. Zhang, LncRNA COL1A1-014 is involved in the progression of gastric cancer via regulating CXCL12-CXCR4 axis, *Gastric Cancer* 23 (2) (2020) 260–272, <https://doi.org/10.1007/s10120-019-01011-0>.
- [135] J. Xu, N. Li, W. Deng, S. Luo, Long noncoding RNA FER1L4 suppresses proliferation, invasion, migration and lymphatic metastasis of gastric cancer cells through inhibiting the Hippo-YAP signaling pathway, *Am. J. Transl. Res.* 12 (9) (2020) 5481–5495.
- [136] W. Li, W. Zhao, Z. Lu, W. Zhang, X. Yang, Long noncoding RNA GAS5 promotes proliferation, migration, and invasion by regulation of miR-301a in esophageal cancer, *Oncol. Res.* 26 (8) (2018) 1285–1294, <https://doi.org/10.3727/096504018x15166193231711>.
- [137] C. He, X. Lu, F. Yang, L. Qin, Z. Guo, Y. Sun, J. Wu, LncRNA UCA1 acts as a sponge of miR-204 to up-regulate CXCR4 expression and promote prostate cancer progression, *Biosci. Rep.* 39 (5) (2019), <https://doi.org/10.1042/bsr20181465>.
- [138] J. Liu, G. Feng, Z. Li, R. Li, P. Xia, Long non-coding RNA FEZF1-AS1 modulates CXCR4 to promote cell proliferation, warburg effect and suppress cell apoptosis in osteosarcoma by sponging miR-144, *Onco Targets Ther.* 13 (2020) 2899–2910, <https://doi.org/10.2147/ott.S235970>.
- [139] X. Long, L. Li, Q. Zhou, H. Wang, D. Zou, D. Wang, M. Lou, W. Nian, Long non-coding RNA LSINCT5 promotes ovarian cancer cell proliferation, migration and invasion by disrupting the CXCL12/CXCR4 signalling axis, *Oncol. Lett.* 15 (5) (2018) 7200–7206, <https://doi.org/10.3892/ol.2018.8241>.
- [140] W. Zhong, J. Yang, M. Li, L. Li, A. Li, Long noncoding RNA NEAT1 promotes the growth of human retinoblastoma cells via regulation of miR-204/CXCR4 axis, *J. Cell Physiol.* 234 (7) (2019) 11567–11576, <https://doi.org/10.1002/jcp.27812>.
- [141] X.F. Sheng, L.L. Hong, H. Li, F.Y. Huang, Q. Wen, H.F. Zhuang, Long non-coding RNA MALAT1 modulate cell migration, proliferation and apoptosis by sponging microRNA-146a to regulate CXCR4 expression in acute myeloid leukemia, *Hematology* 26 (1) (2021) 43–52, <https://doi.org/10.1080/16078454.2020.1867781>.
- [142] J. Li, Z. Li, X. Bai, X. Chen, M. Wang, Y. Wu, H. Wu, LncRNA UCA1 promotes the progression of AML by upregulating the expression of CXCR4 and CYP1B1 by affecting the stability of METTL14, *J. Oncol.* 2022 (2022), <https://doi.org/10.1155/2022/2756986>.
- [143] Y. Liu, H. Zhang, H. Wang, J. Du, P. Dong, M. Liu, Y. Lin, Long non-coding RNA DUXAP8 promotes the cell proliferation, migration, and invasion of papillary thyroid carcinoma via miR-223-3p mediated regulation of CXCR4, *Bioengineered* 12 (1) (2021) 496–506, <https://doi.org/10.1080/108021655979.2021.1882134>.
- [144] G. Hu, J. Ma, J. Zhang, Y. Chen, H. Liu, Y. Huang, J. Zheng, Y. Xu, W. Xue, W. Zhai, Hypoxia-induced lncHILAR promotes renal cancer metastasis via ceRNA for the miR-613/206/1-1-3p/Jagged-1/Notch/CXCR4 signaling pathway, *Mol. Ther.* 29 (10) (2021) 2979–2994, <https://doi.org/10.1016/j.jymthe.2021.05.020>.
- [145] K. Zhang, H. Zhou, B. Yan, X. Cao, TUG1/miR-133b/CXCR4 axis regulates cisplatin resistance in human tongue squamous cell carcinoma, *Cancer Cell Int.* 20 (2020) 148, <https://doi.org/10.1186/s12935-020-01224-9>.
- [146] C.J. Lee, J. Evans, K. Kim, H. Chae, S. Kim, Determining the effect of DNA methylation on gene expression in cancer cells, *Methods Mol. Biol.* 1101 (2014) 161–178, https://doi.org/10.1007/978-1-62703-721-1_9.
- [147] A. Razin, H. Cedar, DNA methylation and gene expression, *Microbiol. Rev.* 55 (3) (1991) 451–458, <https://doi.org/10.1128/mr.55.3.451-458.1991>.
- [148] J.C. Spainhour, H.S. Lim, S.V. Yi, P. Qiu, Correlation patterns between DNA methylation and gene expression in the cancer genome atlas, *Cancer Inf.* 18 (2019), <https://doi.org/10.1177/1176935119828776>.
- [149] A. Shirkevand, Z.N. Boroujeni, S.A. Aleyasin, Examination of methylation changes of VIM, CXCR4, DOK7, and SPDEF genes in peripheral blood DNA in breast cancer patients, *Indian J. Cancer* 55 (4) (2018) 366–371, https://doi.org/10.4103/ijc.1JC_100_18.
- [150] L. Alevizos, A. Katakaki, A. Derventzi, I. Gomatos, C. Loutraris, G. Gloustanou, A. Manouras, M.M. Konstadoulakis, G. Zografos, Breast cancer nodal metastasis correlates with tumour and lymph node methylation profiles of Caveolin-1 and CXCR4, *Clin. Exp. Metastasis* 31 (5) (2014) 511–520, <https://doi.org/10.1007/s10585-014-9645-6>.
- [151] E.A. Ramos, M. Grochoski, K. Braun-Prado, G.G. Seniski, I.J. Cavalli, E.M. Ribeiro, A.A. Camargo, F.F. Costa, G. Klassen, Epigenetic changes of CXCR4 and its ligand CXCL12 as prognostic factors for sporadic breast cancer, *PLoS One* 6 (12) (2011), e29461, <https://doi.org/10.1371/journal.pone.0029461>.
- [152] A.J. Stuckel, W. Zhang, X. Zhang, S. Zeng, U. Dougherty, R. Mustafa, Q. Zhang, E. Perreand, T. Khare, T. Joshi, D.C. West-Szymanski, M. Bissonnette, S. Khare, Enhanced CXCR4 expression associates with increased gene body 5-hydroxymethylcytosine modification but not decreased promoter methylation in colorectal cancer, *Int. J. Cancer*, 12, 2020, <https://doi.org/10.3390/cancers12030539>.
- [153] N. Sato, H. Matsubayashi, N. Fukushima, M. Goggins, The chemokine receptor CXCR4 is regulated by DNA methylation in pancreatic cancer, *Cancer Biol. Ther.* 4 (1) (2005) 70–76, <https://doi.org/10.4161/cbt.4.1.1378>.
- [154] T. Mori, J. Kim, T. Yamano, H. Takeuchi, S. Huang, N. Umetani, K. Koyanagi, D. S. Hoon, Epigenetic up-regulation of C-C chemokine receptor 7 and C-X-C chemokine receptor 4 expression in melanoma cells, *Cancer Res.* 65 (5) (2005) 1800–1807, <https://doi.org/10.1158/0008-5472.CAN-04-3531>.
- [155] A. Drazic, L.M. Myklebust, R. Ree, T. Arnesen, The world of protein acetylation, *Biochim. Biophys. Acta* 1864 (10) (2016) 1372–1401, <https://doi.org/10.1016/j.bbapap.2016.06.007>.
- [156] G. Li, Q. Xie, Z. Yang, L. Wang, X. Zhang, B. Zuo, S. Zhang, A. Yang, L. Jia, Sp1-mediated epigenetic dysregulation dictates HDAC inhibitor susceptibility of HER2-overexpressing breast cancer, *Int. J. Cancer* 145 (12) (2019) 3285–3298, <https://doi.org/10.1002/ijc.32425>.
- [157] T.R. Hebbes, A.W. Thorne, C. Crane-Robinson, A direct link between core histone acetylation and transcriptionally active chromatin, *EMBO J.* 7 (5) (1988) 1395–1402.
- [158] B. Biersack, S. Polat, M. Hopfner, Anticancer properties of chimeric HDAC and kinase inhibitors, *Semin. Cancer Biol.* (2020), <https://doi.org/10.1016/j.semcancer.2020.11.005>.
- [159] M. Farhan, M.F. Ullah, M. Faisal, A.A. Farooqi, U.Y. Sabitaliyevich, B. Biersack, A. Ahmad, Differential methylation and acetylation as the epigenetic basis of

- resveratrol's anticancer activity, in: *Medicines*, 6, 2019, <https://doi.org/10.3390/medicines6010024>.
- [160] D. Ruzic, N. Djokovic, T. Srdic-Rajic, C. Echeverria, K. Nikolic, J.F. Santibanez, Targeting histone deacetylases: opportunities for cancer treatment and chemoprevention, *Pharmaceutics* 14 (1) (2022), <https://doi.org/10.3390/pharmaceutics14010209>.
- [161] M. Di Martile, D. Del Bufalo, D. Trisciuglio, The multifaceted role of lysine acetylation in cancer: prognostic biomarker and therapeutic target, *Oncotarget* 7 (34) (2016) 55789–55810, <https://doi.org/10.18632/oncotarget.10048>.
- [162] S. Yoon, G.H. Eom, HDAC and HDAC inhibitor: from cancer to cardiovascular diseases, *Chonnam Med. J.* 52 (1) (2016) 1–11, <https://doi.org/10.4068/cmj.2016.52.1.1>.
- [163] P. Autin, C. Blanquart, D. Fradin, Epigenetic drugs for cancer and microRNAs: a focus on histone deacetylase inhibitors, in: *Cancers*, 11, 2019, <https://doi.org/10.3390/cancers11101530>.
- [164] E. Damiani, N. Puebla-Osorio, B.M. Lege, J. Liu, S.S. Neelapu, S.E. Ullrich, Platelet activating factor-induced expression of p21 is correlated with histone acetylation, *Sci. Rep.* 7 (2017) 41959, <https://doi.org/10.1038/srep41959>.
- [165] B. Zhang, Y. Li, Q. Wu, L. Xie, B. Barwick, C. Fu, X. Li, D. Wu, S. Xia, J. Chen, W. P. Qian, L. Yang, A.O. Osunkoya, L. Boise, P.M. Vertino, Y. Zhao, M. Li, H.R. Chen, J. Kowalski, O. Kucuk, W. Zhou, J.T. Dong, Acetylation of KLF5 maintains EMT and tumorigenicity to cause chemoresistant bone metastasis in prostate cancer, *Nat. Commun.* 12 (1) (2021) 1714, <https://doi.org/10.1038/s41467-021-21976-w>.
- [166] B. Stamatopoulos, N. Meuleman, C. De Bruyn, A. Delforge, D. Bron, L. Lagneaux, The histone deacetylase inhibitor suberoylanilide hydroxamic acid induces apoptosis, down-regulates the CXCR4 chemokine receptor and impairs migration of chronic lymphocytic leukemia cells, *Haematologica* 95 (7) (2010) 1136–1143, <https://doi.org/10.3324/haematol.2009.013847>.
- [167] T. Vag, K. Steiger, A. Rossmann, U. Keller, A. Noske, P. Herhaus, J. Ettl, M. Niemeyer, H.J. Wester, M. Schwaiger, PET imaging of chemokine receptor CXCR4 in patients with primary and recurrent breast carcinoma, *EJNMMI Res.* 8 (1) (2018) 90, <https://doi.org/10.1186/s13550-018-0442-0>.
- [168] S. Lefort, A. Thuleau, Y. Kieffer, P. Sirven, I. Bieche, E. Marangoni, A. Vincent-Salomon, F. Mechta-Grigoriou, CXCR4 inhibitors could benefit to HER2 but not to triple-negative breast cancer patients, *Oncogene* 36 (9) (2017) 1211–1222, <https://doi.org/10.1038/ncr.2016.284>.
- [169] B. Bockorny, V. Semenisty, T. Macarulla, E. Borazanci, B.M. Wolpin, S. M. Stemmer, T. Golan, R. Geva, M.J. Borad, K.S. Pedersen, J.O. Park, R. A. Ramirez, D.G. Abad, J. Feliu, A. Muñoz, M. Ponz-Sarvisé, A. Peled, T.M. Lustig, O. Bohana-Kashtan, S.M. Shaw, E. Sorani, M. Chaney, S. Kadosh, A. Vainstein Haras, D.D. Von Hoff, M. Hidalgo, B.L.–8040, a CXCR4 antagonist, in combination with pembrolizumab and chemotherapy for pancreatic cancer: the COMBAT trial, *Nat. Med.* 26 (6) (2020) 878–885, <https://doi.org/10.1038/s41591-020-0880-x>.
- [170] G. Bertolini, V. Cancila, M. Milione, G. Lo Russo, O. Fortunato, N. Zaffaroni, M. Tortoreto, G. Centonze, C. Chiodoni, F. Facchinetti, G. Pollaci, G. Taiè, F. Giovino, M. Moro, C. Camisaschi, A. De Toma, C. D'Alterio, U. Pastorino, C. Tripodo, S. Scala, G. Sozzi, L. Roz, A novel CXCR4 antagonist counteracts paradoxical generation of cisplatin-induced pro-metastatic niches in lung cancer, *Mol. Ther.* 29 (10) (2021) 2963–2978, <https://doi.org/10.1016/j.ymthe.2021.05.014>.