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1 **Community coalescence altered the potential of horizontal gene transfers**

2 **within the native soil microbiome**

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9 **Abstract**

10 Microbial community coalescence, which refers to the mixing of microbial communities,
11 frequently shapes the assemblage of soil microbiomes in natural ecosystems. It can exert
12 selective pressure on the coalescent taxa, leading to ecological changes in microbial
13 community structure or microbial evolutionary changes via horizontal gene transfer (HGT).
14 However, the influence of community coalescence on the potential of HGTs within native
15 communities, particularly in soil ecosystems, remains poorly understood. Here, we
16 experimentally quantified the potential evolutionary consequences of soil coalescence. We
17 achieved that by subjecting microcosms containing natural soil to invasion by several
18 microbial communities and profiling mobile genetic elements (MGEs) and adaptive genes of
19 microbial communities up to 60 days after coalescence. Our findings revealed both specific
20 and common responses of MGEs to coalescences over time. Specific effects differed across
21 invasive communities and were particularly pronounced in the early stages. Common effects
22 were associated with an increased abundance of insertion sequences (ISs) across different
23 treatments, suggesting that ISs played a crucial role in promoting diversification at the
24 community level. In summary, we showed that changing MGE profiles are an intrinsic
25 response of the soil microbial community to coalescence-imposed pressure. Our study
26 provides new insights into the modulation of adaptability in soil microbial communities by
27 utilizing community coalescences to address global challenges.

28 **Introduction**

29 Microbial community coalescence, which refers to merging different microbial communities,
30 is a fundamental process on our planet (Rillig et al., 2015). It influences the assemblage of
31 microorganisms, leading to the fundamental changes in composition, succession, and
32 functionalities of the resulting coalescent community (Rillig et al., 2016; Diaz-Colunga et al.,
33 2022; Liu and Salles, 2023). This phenomenon carries significant implications for our
34 understanding of microbial ecology and ecosystem functioning, from the exploitation of
35 beneficial microbial consortia for biofertilization and bioremediation to the investigation of
36 how microbiomes respond to disturbances caused by biotic factors in the context of global
37 changes (Ramoneda et al., 2021; Liu et al., 2023). Therefore, revealing the mechanisms and
38 consequences of community coalescences from various perspectives can provide valuable
39 insights into this biotic disturbance.

40 Microbial communities respond rapidly to disturbances through ecological and
41 evolutionary processes (Chase et al., 2021; Arnold et al., 2022). From an ecological
42 perspective, previous research has shown that coalescence primarily exerts competitive
43 pressure for resources, which drives compositional and functional changes in the resident
44 community (Liu and Salles, 2023). Specifically, while invasive communities face significant
45 selective pressure, resulting in low survival rates for the invaders (less than 1%), this process
46 can still suppress certain native species, leading to changes in community structure and
47 function (Liu and Salles, 2023). One subsequent consequence is that the surviving taxa
48 benefit from coalescence, exhibiting increased abundance compared to the uninvaded control
49 (Liu and Salles, 2023). However, the evolutionary responses of resident communities to
50 coalescence that poses biotic pressure remain largely unexplored.

51 Living microorganisms can acquire adaptive traits in changing environments through

52 many ways, such as mutation, gene expression regulation, horizontal gene transfer (HGT),
53 and epigenetic and phenotypic changes (Douglas and Langille, 2019; Pimpinelli and
54 Piacentini, 2020). Among these processes, HGT is a primary mechanism through which
55 microorganisms exchange genetic information with the external environment, connecting
56 them to other individuals. Through the utilization of mobile genetic elements (MGEs) such as
57 plasmids, insertion sequences (ISs), and integrative and conjugative elements (ICEs), HGT
58 facilitates the dissemination of advantageous traits, including antibiotic resistance and
59 metabolic capabilities, which enhance the survival and competitiveness of microorganisms
60 within their respective ecosystems (Woods et al., 2020; Arnold et al., 2022; Horne et al.,
61 2023). Empirical evidence has shown that the rapid evolution of microbes mediated by HGT
62 can be stimulated by abiotic stresses such as antibiotics, pesticides, and irradiation (Pasternak
63 et al., 2010; Hawkins et al., 2019; Pärnänen et al., 2019; Yao et al., 2022). Community
64 coalescence events will likely intensify the selective pressure (Huet et al., 2023; Liu and
65 Salles, 2023), fostering increased interactions and genetic exchange among diverse microbial
66 populations. As a result, these events provide opportunities for transferring genes and genetic
67 elements.

68 In this study, we aimed to capture indications of microbial gene transfer at a
69 community level during community coalescence. Monitoring evolutionary events and rates in
70 microbes in situ is still challenging at the community level and in the complex context of
71 soils (Brito, 2021; Arnold et al., 2022). We thus primarily employed shotgun metagenomic
72 sequencing and focused on assessing the relative abundance of MGEs in soil, which serves as
73 a proxy for characterizing the potential for horizontal gene transfer (HGT) within
74 communities. Here, we asked the following fundamental question: How does community
75 coalescence influence the MGE profile and the adaptive functions of the resident community
76 over time? We hypothesize that community coalescence will increase HGTs, which will be

77 especially relevant when the resident communities are resilient to the biotic disturbance or are
78 not experiencing an invasion meltdown.

79 **Methods**

80 *The setup of coalescence experiments*

81 The present study is based on a previous coalescence experiment. In brief, we created nine
82 invasive communities differing in composition and diversity by inoculating diluted soil
83 suspensions (A: 10^{-1} , B: 10^{-3} , and C: 10^{-6}) extracted from three soils (E, M, and L, collected
84 from a salt marsh ecosystem showing a significant difference in composition) in sterile soil
85 for 28 days to allow for soil colonization. Then, coalescence experiments were performed by
86 introducing nine invasive communities adjusted to the same bacterial density (~5% of the
87 resident community) into a natural soil (i.e., the original/resident community). The details of
88 experiments and soil microbial communities were described in the previous study (Liu and
89 Salles, 2023). In this study, we selected six treatments (E-A, M-A, L-A, L-B, L-C, and
90 uninvaded control) at dates 0, 5, and 60 to evaluate how community coalescences influence
91 the functional traits of the soil microbial community.

92 *Shotgun metagenomic sequencing and bioinformatic processing*

93 Fifty-four total genomic DNA samples were submitted for library construction and shotgun
94 sequencing at BGI TECH SOLUTIONS (HONGKONG) CO., LIMITED, Hong Kong. The
95 Short-Insert library was used and sequenced on a 2×150 bp DNBseq platform. Raw reads
96 were preprocessed by removing adaptor sequences, contamination, and low-quality reads
97 with SOAPnuke (Chen et al., 2018) to obtain clean reads (39,615,462 reads per sample on
98 average). Detailed information on the sequence data is summarized in Supplementary Table
99 S1.

100 Megahit v. 1.2.9 was used to perform *de novo* assembly for each sample with the k-
101 mer length increasing from 21 to 149 in steps of 20 (Li et al., 2015). Assembled contigs over
102 500 bp were submitted to Prodigal v.2.6 to predict the protein-coding genes (Hyatt et al.,
103 2010). After discarding genes shorter than 100 bp, genes from all 54 samples were clustered
104 at $\geq 95\%$ identity and $\geq 90\%$ overlap with MMseqs2 (Steinegger and Söding, 2017), resulting
105 in a catalog containing 36,890,547 non-redundant genes. Paired-end reads of each sample
106 were mapped to the gene catalog using Salmon v.1.9.0 (Patro et al., 2017) to obtain the
107 relative abundance (estimated as Transcripts Per Million, TPM) of each gene, and the relative
108 abundance was used in downstream analyses.

109 ***Identification of mobile genetic elements, antibiotic resistance genes, and CNPS-cycling*** 110 ***genes***

111 The non-redundant gene sequences constructed from metagenomic data were used to identify
112 MGEs. The ORFs were aligned against corresponding databases for annotating plasmids
113 (PLSDB, (Galata et al., 2019)), insertion sequences (ISs, ISfinder (Siguier et al., 2006)),
114 prophages (PHASTER, (Arndt et al., 2016)), transposons (VRprofile2, (Wang et al., 2022)),
115 integrons (INTEGRALL (Moura et al., 2009)), integrative and conjugative elements (ICEs,
116 ICEberg 2.0 (Liu et al., 2019)), and integrative and mobilizable elements (IME, ICEberg 2.0
117 (Liu et al., 2019)), respectively. Moreover, a manually curated database of MGEs
118 (mobileOG-db, (Brown et al., 2022)) was used to obtain high-quality and functional
119 annotations. The MGE genes are categorized into five functional divisions: integration and
120 excision (IE), phage-related processes (Phage), replication/recombination/repair (RRR),
121 stable/transfer/defense (STD), and transfer (T). The filtering parameters for these annotations
122 were identity $\geq 70\%$, alignment length ≥ 25 bp, and e-value $\leq 10e-10$.

123 To identify potential antibiotic resistance genes (ARGs), the representative sequences

124 were aligned against the Comprehensive Antibiotic Resistance Database (CARD) (Alcock et
125 al., 2020) using Diamond Blastp (Buchfink et al., 2021). For the functional genes related to
126 carbon, nitrogen, phosphorus, and sulfur cycling, we aligned ORFs against the Carbohydrate-
127 Active EnZymes database (CAZy) (Cantarel et al., 2009), NCycDB (Tu et al., 2019),
128 PCycDB (Zeng et al., 2022), SCycDB (Yu et al., 2021). The cut-off of these annotations was
129 identity $\geq 70\%$, alignment length ≥ 25 bp, and e-value $\leq 10e-10$. The composition and
130 abundance of these genes in invasive and original resident communities are shown in
131 Supplementary Figure S1-4.

132 *Identifying the number of neighbors of intermediate contigs*

133 The number of neighbors of intermediate contigs refers to the count of other contigs that
134 share overlapping regions or have alignment relationships with a particular intermediate
135 contig. Thus, the number of neighbors of intermediate contigs in different sequenced
136 genomes may indicate the comparable complexity caused by repeated or duplicated regions
137 due to HGT when the sequencing depth and assembly error were similar across samples. In
138 this study, we developed an approach based on contig networks to calculate the ratio of
139 neighbors (number of neighbors per contig) for each sample based on an intermediate
140 assembly graph outputted from Megahit v. 1.2.9 (Li et al., 2015) to further analyze the HGT
141 processes. In brief, the FASTG files were first created from intermediate contigs constructed
142 from 41-mers with the core function “contig2fastg” in Megahit v. 1.2.9. The connection
143 information of assembly graphs was then extracted for neighbors counting and ratio
144 calculation.

145 *Statistical analysis*

146 The total relative abundance of MGEs across treatments was shown as a Z-score, and the

147 difference in total relative abundance related to the uninvaded control was estimated using a
148 t-test combining an ANOVA test for global variance. To reveal the compositional difference
149 of functional traits (e.g., MGEs, ARGs, and CNPS-cycling genes) between different
150 treatments, we performed PERMANOVA using the Adonis test (999 permutations) with R
151 package “Vegan”. The relative abundance difference of each MGE family compared to the
152 uninvaded control was estimated using the R package “edgeR” and $p < 0.05$ as the significant
153 level.

154 Spearman’s correlation-based network analysis revealed the potential relationship
155 between MGEs and other elements such as ARGs, CNPS-cycling genes, and ASVs on Days 5
156 and 60. The strict cut-offs (Spearman’s $r > 0.7$ or < -0.7 and $p < 0.001$) were used for these
157 analyses. Only nodes and edges significantly positively correlated with MGEs were kept in
158 the network. Networks were visualized with Gephi 0.9 (Bastian et al., 2009). Moreover, we
159 employed the Erdos-Renyi model (function *erdos.renyi.game* in R package “igraph”) to
160 construct two random networks according to the corresponding empirical networks. The
161 difference in degree distribution between random and empirical networks was estimated with
162 the Kolmogorov-Smirnov test.

163 We assessed the initial functional (nutrient-cycling genes) similarity between invasive
164 and resident communities before coalescences by calculating Bray-Curtis dissimilarity (1-
165 dissimilarity) with the R package “Vegan.” The Bray-Curtis dissimilarity in MGE
166 composition between coalescent community and uninvaded control was used to infer
167 coalescence impact on MGE profile.

168 **Results**

169 *Effects of coalescence on community functional traits*

170 The functional traits of coalescent communities depended on time and treatments (Figure 1).
171 Time was the main factor influencing the composition of the overall functional trait (Figure
172 1a), which was consistent with the bacterial community composition over two sampling times
173 (Figure 1b; $p < 0.001$, Procrustes analysis). The treatment (i.e., invasive communities) only
174 significantly influenced the overall functional trait on Day 5 ($p < 0.01$, Adonis), whereas the
175 functional trait on Day 60 showed no significant differences ($p > 0.05$, Adonis). Such
176 phenomenon was also observed for three specific functional traits, including CNPS-cycling
177 genes, MGEs, and ARGs (Supplementary Figure S5).

178 *Community coalescences altered the profile of MGEs*

179 In the analyzed dataset, mobile genetic elements (MGEs) represented a relative abundance of
180 4.07-4.81% of all genes (Figure 2). Among the MGEs, plasmids were the most abundant,
181 accounting for an average relative abundance of 3.64% of all coalescent communities (Figure
182 2). This was followed by ISs at 0.45%, ICEs at 0.29%, prophages at 0.078%, IMEs at 0.020%,
183 transposons at 0.013%, and integrons at 0.0056%.

184 Different MGEs had varied responses to different invasive communities (Figure 2).
185 On Day 5, the abundance of ISs exhibited the highest sensitivity to coalescence, with all
186 treatments showing a significant increase in IS abundance compared to the uninvaded control
187 on Day 5. The abundance of ICEs was significantly increased in the L-A treatment, and
188 prophage abundance significantly increased in the E-A treatment on Day 5. On Day 60, a
189 significant decrease was observed in several treatments compared to the uninvaded control.
190 For example, ICE abundance decreased significantly in the L-A treatment, prophage

191 abundance reduced considerably in the E-A treatment, and integron abundance decreased
192 significantly in the M-A treatment. However, the abundance of all MGEs in the uninvaded
193 control treatment was not significantly different between D0 and D5 (Supplementary Figure
194 S6). A significant decrease in MGE abundance in the uninvaded control treatment between
195 D0 and D60 was observed for IS and IME (Supplementary Figure S6).

196 Identifying MGE genes using the MobileOG database allowed for functional
197 classification (Supplementary Figure S7). Among the five critical divisions of MobileOG, the
198 genes encoding proteins for integration and excision (IE) were the most abundant.
199 Furthermore, the total relative abundance of these IE genes showed a significant increase in
200 all coalescent communities compared to the uninvaded control on Day 5. The abundance of
201 the phage-related process genes (P) was significantly increased in the E-A treatment but
202 decreased in the M-A treatment. Other divisions, such as replication/recombination/repair
203 (RRR), stable/transfer/defense (STD), and Transfer (T), were not significantly affected by
204 coalescence on Day 5. For Day 60, the abundance of phage-related process genes decreased
205 in both L-A and L-B treatments. Gene abundance mediating element transfer significantly
206 increased in E-A and L-C treatments. In contrast, the abundance of the STD gene was only
207 observed to increase in the M-A treatment (Supplementary Figure S7). The abundance of all
208 functional groups of MGEs in the uninvaded control treatment was not significantly different
209 between D0 and D5. In contrast, a significant decrease in abundance was observed for all
210 functional groups between D0 and D60 (Supplementary Figure S8).

211 We further performed correlation analyses for the total abundance of MGEs in
212 invasive communities and the corresponding coalescent communities to assess whether the
213 invader-derived MGEs contributed to the abundance changes after invasions. The results
214 showed the non-significant or negative correlations between the abundance of seven MGEs in

215 invasive communities and coalescent communities for both Days 5 and 60, except plasmid on
216 Day 5 (Supplementary Figure S9). This indicate that the increases in MGEs abundance in
217 coalescent communities, especially for ISs (negative correlation, $p = 0.0046$, Spearman's
218 correlation, Supplementary Figure S9), were not mainly due to the introduction of invaders.

219 The total abundance of the integration and excision (IE) division on Day 5 was
220 significant and negatively correlated with its abundance in invasive communities ($p = 0.048$,
221 Spearman's correlation, Supplementary Figure S10), while the correlation for Day 60 was
222 non-significant ($p = 0.61$). Besides, a positive and significant correlation was found for the
223 stable/transfer/defense (STD) division on Day 5 ($p = 0.042$; the correlation for Day 60 is non-
224 significant with a p-value = 0.046).

225 *Initial functional similarity between invasive and resident communities predicted MGEs'*
226 *response.*

227 The impact of coalescence impacts on MGE profiles estimated with the Bray-Curtis
228 dissimilarities between the coalescent community and uninvaded control on Day 5 was
229 positively correlated to the initial functional similarities between invasive and resident
230 communities (Figure 3a). Specifically, among the MGEs, the highest correlation coefficient
231 was observed for IS ($R = 0.63$) and prophage ($R = 0.65$). From a functional taxonomic
232 perspective, the genes' similarity in the carbon and nitrogen cycles exhibited stronger
233 predictability of the post-coalescence effects on MGE profiles. However, such correlations on
234 Day 60 shifted to predominantly negative or non-significant relationships (Figure 3a).

235 We obtained the MGEs at the gene family level with significantly enriched or
236 depleted abundance compared to the uninvaded control (Figure 3b and Supplementary
237 Figures S11a and S11b). First, our results revealed that enriched MGEs under different

238 treatments exhibited higher uniqueness (i.e., fewer proportions shared among multiple
239 treatments) for both Days 5 and 60, compared to the depleted MGEs (Supplementary Figure
240 S11c). Besides, we found that IS was the most sensitive MGE type, and the number of
241 increased IS families was higher than that of other MGE types, such as plasmid and ICE
242 (Figure 3b).

243 The correlation analysis suggested that the total number of enriched MGEs on Day 5
244 was positively correlated to the initial functional similarities between invasive and resident
245 communities before coalescence ($R = 0.89$ and $p < 0.001$ based on C-cycling function
246 similarity, $R = 0.65$ and $p = 0.0081$ based on N-cycling function similarity, $R = 0.24$ and $p =$
247 0.4 based on P-cycling function similarity, $R = 0.56$ and $p = 0.03$ based on S-cycling function
248 similarity, Figure 3c). Such positive correlations were also observed for the total number of
249 depleted MGEs on Day 5 ($R = 0.80$ and $p < 0.001$ based on C-cycling function similarity, $R =$
250 0.59 and $p = 0.021$ based on N-cycling function similarity, $R = 0.13$ and $p = 0.64$ based on P-
251 cycling function similarity, $R = 0.49$ and $p = 0.063$ based on S-cycling function similarity).
252 However, the correlation between the total number of enriched/depleted MGEs on Day 60
253 with initial functional similarities was mostly insignificant or negatively significant (Figure
254 3c).

255 *ISs contributed to the number of neighbors of intermediate contigs*

256 We created the intermediate assembly graph ($Kmer = 41$) for each community and used the
257 ratio of neighbors per intermediate contig (ratio or ratio value hereafter) to infer the potential
258 HGT level in a community, with larger the ratio values indicating more HGT occurred. The
259 reason for using a Kmer size of 41 is that the sequence information can be largely preserved,
260 and the contigs can be obtained in relatively longer lengths (Supplementary Figure S12). The
261 results showed that the uninvaded control had a similar ratio of neighbors (0.23 on average)

262 between Days 5 and 60, indicating the constant status of the HGT event (Figure 4a). However,
263 the coalescent communities had a significantly higher ratio than the uninvaded control on
264 Day 5. M-A, L-A, L-B, and L-C had a similar ratio value and higher than that under the E-A
265 treatment (Figure 4a). On day 60, the ratios for all treatments returned to levels similar to the
266 uninvaded control (Figure 4a).

267 The total abundance of ISs in coalescent communities was significantly and positively
268 correlated to the ratio value ($R^2 = 0.64$, $p < 0.0001$, Figure 4b), which indicated the
269 importance of ISs in response to the coalescence. Importantly, we found that such a ratio
270 value was negatively correlated to the IS abundance in the corresponding invasive
271 communities on Day 5 ($R^2 = 0.38$, $p = 0.0077$) and not significantly correlated to that on Day
272 60 ($p = 0.45$, Figure 4c). This suggests that the increase in ratio values in coalescent
273 communities was not due to the introduction of ISs derived from invaders.

274 We further compared the ratio value across all contigs (General), contigs containing
275 ISs (All_IS), and contigs containing enriched IS families (Enriched_IS). We found that
276 contigs containing enriched ISs had the highest average ratio value (Figure 4d). Specifically,
277 the average ratio value of the Enriched_IS group was significantly higher than that of the
278 other two groups on Day 5 (Figure 4d; $p < 0.05$, paired t-test). For Day 60, the average ratio
279 value of all treatments was assessed with a non-significant difference between enriched_IS
280 and all_IS groups with a p -value = 0.054 (paired t-test). However, the ratio values of
281 enriched_IS contigs under L-A and L-B treatments were significantly higher than that of the
282 other two groups ($p < 0.05$, ANOVA with Tukey test) for both Days 5 and 60 (Figure 4d).

283 The overall contribution of MGEs to the general ratio of contigs' neighbors was
284 estimated with Spearman's correlation between the ratio value and MGE abundance (Figure
285 4e). The result showed that ISs had the highest coefficient ($R^2 = 0.64$) with the ratio value,

286 followed by ICE ($R^2 = 0.48$), plasmid ($R^2 = 0.23$), and integron ($R^2 = 0.15$). The correlations
287 were non-significant for IME ($R^2 = 0.11$) and prophage ($R^2 = 0.03$) and negative for
288 Transposon ($R^2 = 0.20$).

289 *Network analysis revealing potential associations to MGEs*

290 Two correlation-based networks revealed the difference in the potential linkages between
291 MGEs and ASVs, ARGs, and CNPS-cycling genes on Days 5 (Network D5) and 60
292 (Network D60) (Figure 5a). The degree distribution of these two networks was estimated to
293 significantly differ from the corresponding random networks ($p < 0.001$, Kolmogorov-
294 Smirnov test, Supplementary Figure S13). For networks D5 and D60, the total number of
295 positive links was 5320 and 4726, the average degree was 1.29 and 1.18, the average path
296 length was 10.38 and 12.59, and the maximum degree of the node was 34 and 17,
297 respectively (Figure 5b).

298 The node distribution of four components was similar between the two networks,
299 although the total number of nodes was higher in the D5 than in the D60 network (Figure 5c).
300 The MGE-ARG and MGE-CNPS links were 575 and 3850 on Day 5, respectively (Figure 5d).
301 These were higher than those on Day 60 (411 and 3209, respectively) under the same
302 network construction approach. For example, the total positive correlations between MGEs
303 and the multidrug group of ARGs accounted for 201 (3.8% of total positive edges) and 118
304 (2.5% of total positive edges) on Days 5 and 60, respectively. However, the total number of
305 positive links between MGEs and ASVs was observed with a different phenomenon, while
306 such links for network D5 (895) were lower compared to network D60 (1106) (Figure 5d).

307 Plasmids had the highest positive correlations with other components, while such
308 proportion slightly decreased across time (70.6% in D5 and 70.1% in D60, Figure 5e). The

309 proportion of positive edges linked with ISs accounted for the second, which was 20.1% in
310 D5 and 21.2% in D60. By comparing the ratio of positive edges in networks D5 and D60, we
311 found that the proportion increase in D60 was observed for ISs, and this phenomenon did not
312 occur in other major components, such as plasmid, prophage, and ICE. This result may
313 indicate that the importance of IS-mediated associations increased over time after
314 coalescence.

315 The sub-networks centered with ISs were constructed and showed distinct patterns
316 between D5 and D60 (Figure 5f). Our results suggested that more ARGs and CNPS-cycling
317 genes were correlated with ISs in network D5. For instance, 15 types of ARGs (regarding
318 drug resistance) were found in sub-network D5, compared to sub-network D60 (13 types).
319 Moreover, CNPS-IS edges were 13.7% higher in sub-network D5 (749) than in sub-network
320 D60 (659). However, 11 phyla of microbes (212 in total) correlated with ISs in sub-network
321 D5, which was higher than that in sub-network D60 (14 phyla, 255 total). For instance, taxa
322 affiliated with Proteobacteria were the most frequently present in networks where the number
323 of edges was 52 and 115 in sub-networks D5 and D60, respectively.

324 **Discussion**

325 Microbial communities are sensitive and adaptive to environmental changes. Due to their fast
326 reproduction, such ecological and evolutionary responses could happen quickly, even within
327 several hours or days (Jain et al., 2003; Bosshard et al., 2017; Döbelmann et al., 2017). The
328 previous attention had been put mainly on either ecological changes of microbial
329 communities or the evolution of specific individuals under abiotic selections. However, the
330 assessment of the microbial evolutionary response under the community coalescence, which
331 represents a common biotic pressure in the soil, is lacking. This study provides vital insight in
332 this respect.

333 *Specific and common impacts of coalescences on MGEs*

334 Horizontal gene transfer (HGT) is one of the most critical processes in microbial evolution
335 that allows a remarkable increase in genome innovation that significantly exceeds anything
336 that could have been accomplished by clonal evolution alone (Jain et al., 2003). By assessing
337 the relative abundance and composition of mobile genetic elements (MGEs), we
338 demonstrated that coalescences led to alterations in MGE profiles, with the most pronounced
339 changes observed on Day 5 after coalescence. In contrast, the profiles on Day 60 showed
340 resilience compared to the uninvaded control. These findings provide evidence of
341 coalescence-induced modifications in HGT potentials within the soil community.

342 The influence of coalescence on MGE profiles varied among different treatments. For
343 instance, treatment E-A showed an increase in prophage abundance, while treatments L-A
344 exhibited an increased total abundance of ICE (Figure 2 and Supplementary Figure S9),
345 indicating that the mechanisms for coalescence regulating MGEs are complicated and
346 dependent on invasive communities. Notably, we introduced invaders at a low invasion rate,
347 approximately 5% of the resident taxa density. This suggests that MGE abundance is less
348 likely to change due to direct accumulation by introduced invaders. Therefore, the change in
349 prophage abundance under E-A treatment is expected due to the more extensive input derived
350 from invasive communities with a significantly higher quantity of prophages than other
351 invasive communities and the resident community (Supplementary Figures S1 and S9). The
352 introduced prophages were viable and capable of lysing their hosts and reproducing within
353 the coalescent community during environmental changes [39]. In this case, the coalescence
354 process could be the source of various stresses for invaders, such as nutritional competitions
355 between invaders and residents, antibiotics produced by residents, and quorum sensing
356 derived from residents, thereby leading to phage excision.

357 Alternatively, the changes in the total abundance of other MGEs, such as plasmids,
358 ICE, and IS, may primarily rely on shifts in microbial communities, including establishments
359 of specific invaders and the thriving or suppression of particular resident taxa. Importantly,
360 we observed a common effect across treatments: the significant increase in the total
361 abundance of IS after coalescence. This phenomenon is supported by the increased
362 abundance of “IE division” genes associated with integration and excision, which coincides
363 with the function of the ISs (Brown et al., 2022). Additionally, the significant negative
364 correlation between the abundance of ISs in invasive and coalescent communities on Day 5
365 suggests that ISs did not directly contribute to increased ISs in coalescent communities
366 (Supplementary Figures S9 and S10). Moreover, the abundance of ISs enriched under each
367 treatment related to uninvaded control is significantly lower in invasive communities than in
368 uninvaded control and coalescent communities, let alone the low invasion rate
369 (approximately 5% of the density of resident taxa) (Supplementary Figure S14) we applied.
370 Our results suggest that the increase in IS abundance is an intrinsic response of the resident
371 community following coalescence.

372 ***Competitive stress triggers changes in the MGE profile after coalescence.***

373 Competition for resources between invaders and residents is a common interaction after
374 coalescence. Our results suggest that the response of MGEs is significantly correlated to the
375 probability of such competition between invaders and residents. Two noteworthy phenomena
376 emerged from this study: First, positive correlations were predominantly observed on Day 5,
377 suggesting higher competitive stress at the early coalescence stage. This aligns with our
378 previous observations that the survival rate of invaders was lower, and the impact on
379 community composition was less severe at the late stages of the coalescence (Liu and Salles,
380 2023). Second, relatively stronger correlations were found with carbon-cycling genes

381 compared to nitrogen, phosphorus, and sulfur-related genes, indicating a more intensive
382 competition for carbon sources.

383 These two points are further supported by network analyses of potential association
384 patterns between MGEs and genes related to microbial antibiotic resistance and nutrient
385 cycling. MGE-mediated transfers of antibiotic resistance genes and resource utilization genes
386 are the main ways microorganisms enhance their fitness (Toft and Andersson, 2010).
387 Especially in terms of harnessing energy, microorganisms may have the opportunity to utilize
388 novel carbon sources after acquiring genes from other microorganisms through HGT, thereby
389 gaining survival advantages in resource-limited or contaminated environments (Jain et al.,
390 2003). The network analyses suggest that MGEs exhibited more intensive and positive
391 correlations with these genes on Day 5 than Day 60, indicating the more intensive adaptive
392 pattern of coalescent communities at the early stage after coalescence. Furthermore, within
393 these associations, genes involved in nutrient cycling, especially carbon-cycling genes,
394 appeared more susceptible to MGE influences on Day 5. These findings underscore the
395 significance of invader-triggered competitions and nutrient-cycling genes for communities
396 responding to such disturbances.

397 *The critical role of ISs in HGTs*

398 Our findings support the notion that the increased abundance of ISs is likely a direct outcome
399 of horizontal gene transfers (HGTs) within the community by evaluating the neighbors of
400 intermediate contigs containing IS fragments. Given that IS fragments are relatively short in
401 length (typically 0.7–2.5 kb long (Siguier et al., 2014)), they are more likely to be assembled
402 into a single contig in the assembly graph, enabling a more accurate analysis of the number of
403 contig neighbors to represent the frequency of ISs in different genomes at the community
404 level. The dissemination of IS fragments occurs through multiple mechanisms, including

405 vectors like plasmids via conjugation or through their host genomes via replication and
406 translocation (Preston et al., 2004). This can lead to repetitive sequences in the meta-genome
407 graph. Our results show that the increased abundance of ISs was significantly positively
408 correlated with the increased contig connectivity in the assembly graph, and the coefficient
409 was higher for ISs than other MGEs such as ICEs, plasmids, and integrons.

410 The Network analysis unveiled notable associations between ISs and genes related to
411 nutrient cycling and antibiotic resistance. Interestingly, the proportion of IS-related links in
412 the network increased on Day 60, suggesting a potentially more pronounced role of ISs at this
413 stage. This increase in IS-related links was accompanied by a decrease in the proportion of
414 links associated with other MGEs. The enhanced connectivity of ISs with nutrient-cycling
415 and antibiotic genes underscores their potential role in shaping community functions and
416 adaptation. The association of ISs with nutrient-cycling genes highlights their relevance in
417 facilitating resource utilization and metabolic versatility within the coalescent communities.
418 Furthermore, the links between ISs and antibiotic genes indicate their potential involvement
419 in disseminating antibiotic resistance traits, which can enhance the survival and
420 competitiveness of microorganisms in resource-limited or contaminated environments.
421 However, the insertion sites of ISs may be approximately random, and the transfer may
422 activate or inactivate nearby functional genes (Vandecraen et al., 2017). In addition, the
423 fragments transferred by ISs do not necessarily result in better host fitness (Vandecraen et al.,
424 2017). Therefore, the genetic diversification suggested by increased IS abundance does not
425 necessarily imply better fitness or adaptivity of microbial communities. Indeed, we found that,
426 on Day 60, a large proportion of ISs that enriched on Day 5 was not varied from the
427 uninvaded control, and the total abundance of ISs was recovered to a similar level across
428 treatments. These reflect that ISs can promote and constrain microbial mutation and
429 adaptivity (Consuegra et al., 2021). Nevertheless, on Day 60, the network analysis ISs

430 displayed more connections with potential hosts, represented by ASVs. These observations
431 suggest that a portion of ISs carrying adaptive genes via horizontal gene transfer may
432 facilitate the acquisition of adaptations by the host and establish a broader presence within
433 different hosts at the late stage of coalescence. Overall, our results highlight the importance
434 of ISs in community genetic diversity and adaptation following coalescence.

435 It is worth noting that further investigations are needed to unravel the specific
436 mechanisms underlying the interactions between ISs and host genomes, as well as the
437 functional implications of these associations. To achieve this, long-length sequencing
438 techniques can provide a more accurate assessment of the relationships between IS fragments
439 and adjacent functional elements and their respective host genomes (Zhou et al., 2020; Che et
440 al., 2021). Besides, by adopting targeted metagenomic sequencing approaches (Dunon et al.,
441 2018), it becomes easier to track the dynamic patterns of specific ISs throughout the process
442 of community succession in the context of community coalescence. This targeted approach is
443 a powerful tool for characterizing the precise role of ISs in driving community dynamics and
444 adaptation.

445 **Conclusions**

446 This study provided insights into the evolutionary perspective of soil microbial communities
447 under the coalescence scenario. On the one hand, our results confirm that competition for
448 resources could be the primary mechanism underlying coalescences in soil. This has
449 implications for understanding soil microbial community dynamics and regulating
450 inoculating beneficial consortia in agroecosystems. Specifically, predicting the consequences
451 of community coalescences requires consideration of a lose-lose competitive situation (Liu
452 and Salles, 2023), and in agricultural management, taking measures to reduce competition for
453 inoculum will increase inoculation benefits (Liu et al., 2022). On the other hand, our results

454 suggest a rapid response of MGE profiles and increased HGT potential triggered by
455 coalescence, where ISs may play a vital role in promoting genomic diversification in soil
456 communities. The artificially manipulated and improved adaptation of microbial communities
457 to drought, heat, or cold stress may enhance our ability to predict and manage the adaptability
458 of ecosystems to changing climates (Allsup et al., 2023). Although our results are limited to
459 profile functional traits at the community level, they point to a need for further investigation
460 of specific evolution within coalescent communities using new technologies such as third-
461 generation sequencing and the coalescence impact on community fitness against future
462 disturbances and subsequent ecosystem functions.

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468 **Competing interests**

469 The authors declare that they have no competing interests.

470 **Availability of data and materials**

471 All the raw sequencing data were deposited in the National Center for Biotechnology
472 Information Sequence Read Archive under the accession number PRJNA843110.

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- 626

1 **Community coalescence altered the potential of horizontal gene transfers**

2 **within the native soil microbiome**

3 **Xipeng Liu, Thomas Hackl, Joana Falcão Salles[#]**

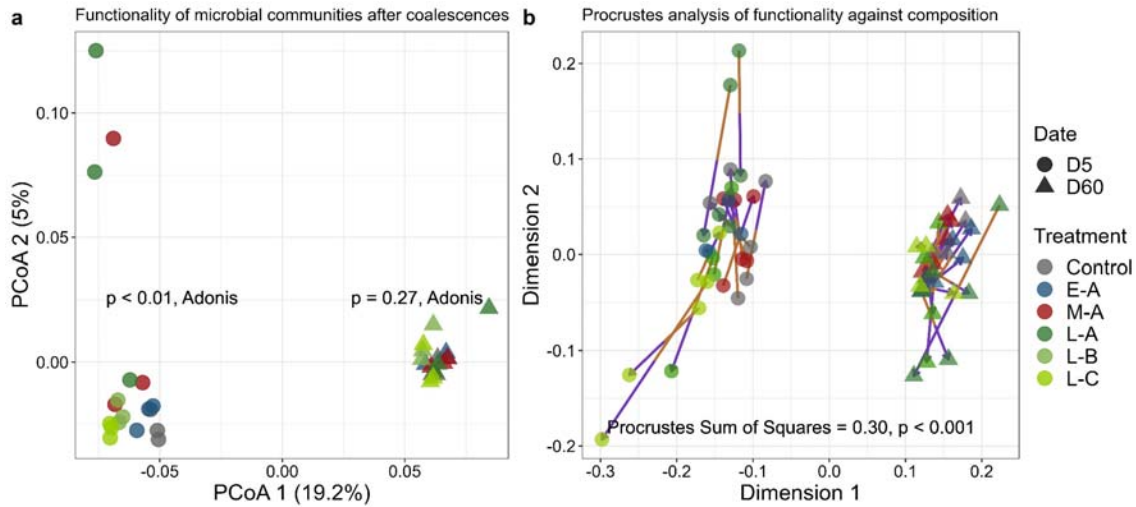
4 Microbial Ecology cluster, Genomics Research in Ecology and Evolution in Nature

5 (GREEN), Groningen Institute for Evolutionary Life Sciences (GELIFES), University of

6 Groningen, 9747 AG Groningen, The Netherlands

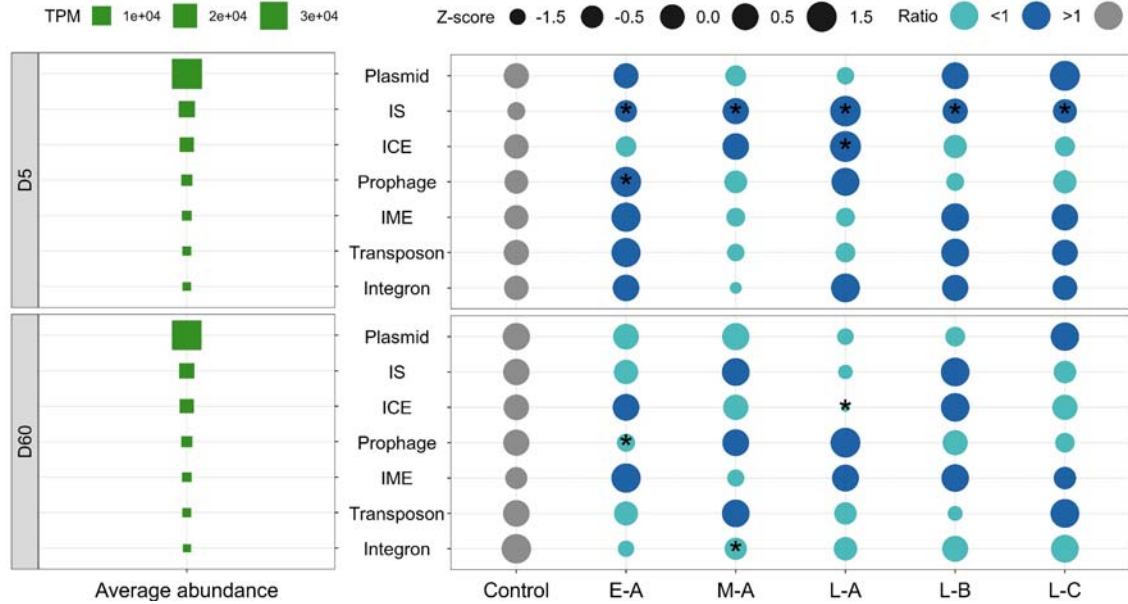
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9

10 **Figure 1. The functionality of microbial communities after coalescence is primarily**
11 **influenced by time. (a)** Principal Coordinates Analysis (PCoA) showing the overall
12 functionality (including genes related to nutrient cycling, MGEs, and ARGs) of coalescent
13 communities was mainly changed at the early stage after coalescences. The p -value of
14 Permutational Multivariate Analysis of Variance (Adonis) close to different points indicated
15 the functional variance for each date. **(b)** Procrustes analysis of functionality against
16 microbial community composition (based on weighted Unifrac distance) showed that changes
17 in functionality over time were synchronized with community changes. The purple end of
18 each line connects to the 16S rRNA gene data for the sample, while the green end is
19 connected to the overall functionality data.



20

21 **Figure 2. Coalescences changed the MEG profile of the soil microbial community.**

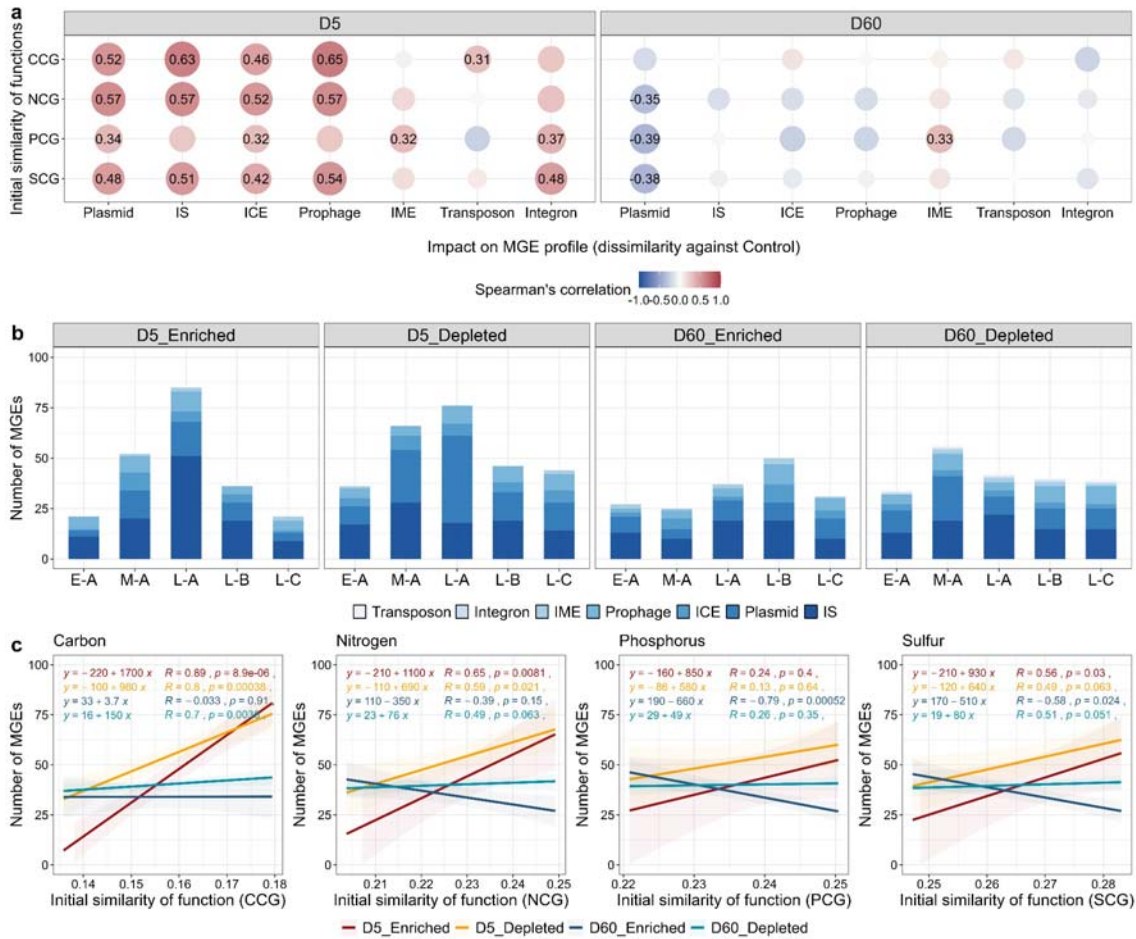
22 Increased abundance of ISs was the common effect across invasive communities. The

23 abundance (TPM) of MGEs was transformed with Z-score. The color indicates the ratio of

24 MGE abundance in relation to the uninvaded control. Asterisks suggest the significant

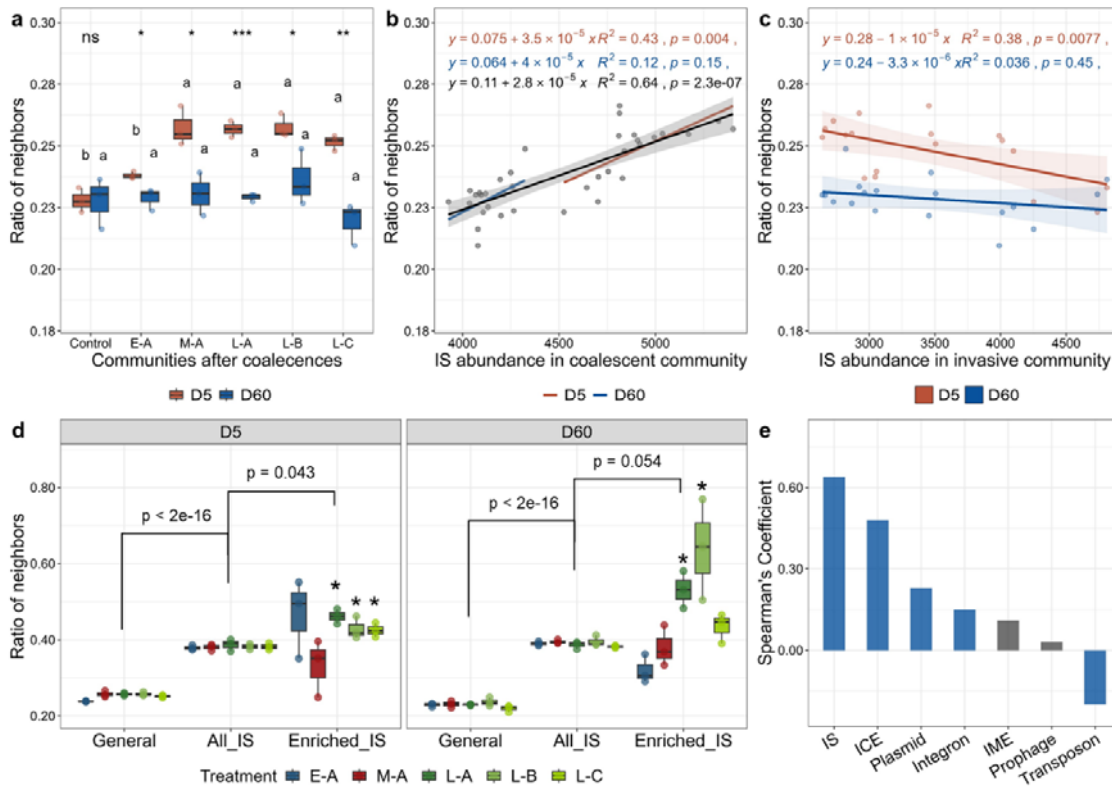
25 difference in MGEs' abundance between uninvaded control and coalescent communities with

26 a t-test ($p < 0.05$).



27

28 **Figure 3. Initial functional similarity between invasive and resident communities**
 29 **correlated with the impacts on MGE profiles suggesting the importance of invader-**
 30 **resident competition. (a)** Spearman's correlations between the initial functional similarities
 31 between invasive and original resident communities and the impacts on MGE profiles after
 32 coalescences. Correlation coefficients (R value) are shown above points, whereas the point
 33 without coefficient indicates a non-significant correlation. **(b)** The number of MGEs with
 34 significantly changed abundance compared to the uninvaded control on Days 5 and 60. **(c)**
 35 Spearman's correlations between the initial functional similarities between invasive and
 36 original resident communities and the number of MGEs with significantly changed
 37 abundance. Four functional traits include genes involved in carbon (CCG), nitrogen (NCG),
 38 phosphorus (PCG), and sulfur (SCG) cycling.



39

40 **Figure 4. ISs contributed to the increased potential of horizontal gene transfer during**

41 **coalescences. (a)** The general ratio of neighbors of contigs in the intermediate assembly

42 graph. Asterisks indicate the significant difference in the ratio between Days 5 and 60, while

43 different letters above the boxes indicate the significant difference among treatments. **(b)**

44 Spearman's correlation between the ratio of neighbors and IS abundance in coalescent

45 communities. **(c)** Spearman's correlation between the ratio of neighbors and IS abundance in

46 invasive communities. **(d)** The ratio of neighbors for three contig types: all contigs (General),

47 contigs containing ISs (all_IS), and contigs containing enriched IS families (Enriched_IS). P-

48 values (paired t-test) imply the difference in average ratio between groups, and the asterisks

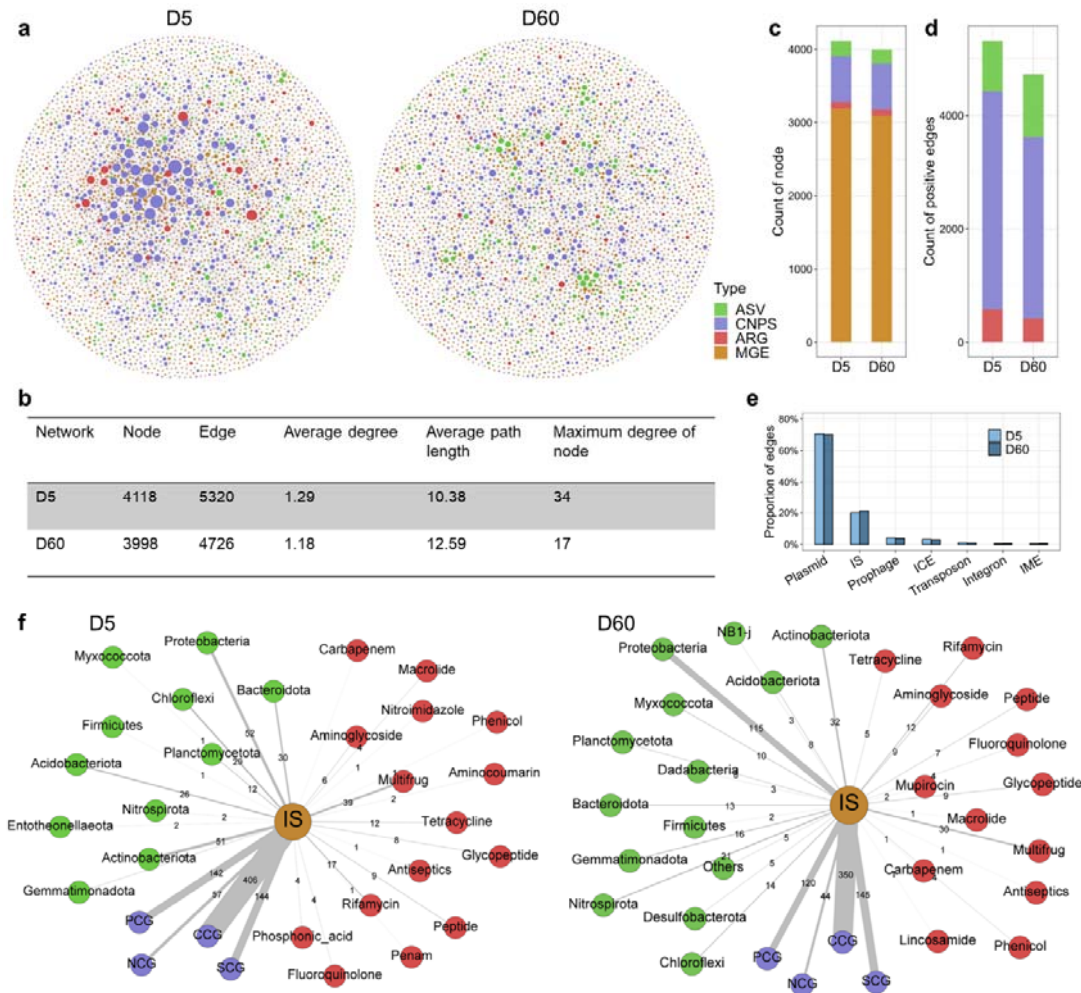
49 above boxes indicate the significantly higher proportion in the group (Enriched_IS) compared

50 to the other two groups (ANOVA with Tukey test, $p < 0.05$). **(e)** Spearman's correlation

51 coefficients of general ratio value and the abundance of each MGE type. The positive and

52 negative coefficients imply positive and negative correlations, respectively. The grey color of

53 the bar indicates a non-significant correlation ($p > 0.05$).



54

55 **Figure 5. Correlation-based network analyses on the potential associations to MGEs. (a)**
 56 Networks (for Days 5 and 60) based on four types of components: MGEs, nutrient-cycling
 57 genes (carbon, nitrogen, phosphorus, sulfur, CNPS), ARGs, and microbial taxa (ASVs). Only
 58 significant and positive correlations (Spearman's $r > 0.7$ and $p < 0.001$) are shown in the
 59 network. The nodes' size represents the degree number and is scaled to the maximum number
 60 of degrees. **(b)** Network metrics. **(c)** The total number of four components in two networks.
 61 **(d)** The total number of edges of ARGs, CNPS, and ASVs linked to MGEs. **(e)** The
 62 proportion of edges between MGEs and other components in a total number of edges. **(f)**
 63 Sub-networks show significant and positive associations with ISs. The width of the line

64 represents the association amount and is indicated by the number next to the line.