Washington University School of Medicine

Digital Commons@Becker

2020-Current year OA Pubs

Open Access Publications

3-13-2023

Clinicopathological classification of immune checkpoint inhibitorassociated myocarditis: Possible refinement by measuring macrophage abundance

Jesus Jimenez

Nicolas Kostelecky

Joshua D Mitchell

Kathleen W Zhang

Chieh-Yu Lin

See next page for additional authors

Follow this and additional works at: https://digitalcommons.wustl.edu/oa_4

Part of the Medicine and Health Sciences Commons

Please let us know how this document benefits you.

Authors

Jesus Jimenez, Nicolas Kostelecky, Joshua D Mitchell, Kathleen W Zhang, Chieh-Yu Lin, Daniel J Lenihan, and Kory J Lavine

RESEARCH Open Access

Clinicopathological classification of immune checkpoint inhibitor-associated myocarditis: possible refinement by measuring macrophage abundance

Jesus Jimenez^{1,2*†}, Nicolas Kostelecky^{3†}, Joshua D. Mitchell², Kathleen W. Zhang², Chieh-Yu Lin³, Daniel J. Lenihan² and Kory J. Lavine^{1,4}

Abstract

Background Immune checkpoint inhibitor (ICI) myocarditis is associated with high morbidity and mortality. While endomyocardial biopsy (EMB) is considered a gold standard for diagnosis, the sensitivity of EMB is not well defined. Additionally, the pathological features that correlate with the clinical diagnosis of ICI-associated myocarditis remain incompletely understood.

Methods We retrospectively identified and reviewed the clinicopathological features of 26 patients with suspected ICI-associated myocarditis based on institutional major and minor criteria. Seventeen of these patients underwent EMB, and the histopathological features were assessed by routine hematoxylin and eosin (H&E) staining and immunohistochemical (IHC) staining for CD68, a macrophage marker.

Results Only 2/17 EMBs obtained from patients with suspected ICI myocarditis satisfied the Dallas criteria. Supplemental IHC staining and quantification of CD68⁺ macrophages identified an additional 7 patients with pathological features of myocardial inflammation (> 50 CD68⁺ cells/HPF). Macrophage abundance positively correlated with serum Troponin I (P = 0.010) and NT-proBNP (N-terminal pro-brain natriuretic peptide, P = 0.047) concentration. Inclusion of CD68 IHC could have potentially changed the certainty of the diagnosis of ICI-associated myocarditis to definite in 6/17 cases

Conclusions While the Dallas criteria can identify a subset of ICI-associated myocarditis patients, quantification of macrophage abundance may expand the diagnostic role of EMB. Failure to meet the traditional Dallas Criteria should not exclude the diagnosis of myocarditis.

Keywords Cardio-oncology, Immune checkpoint inhibitors, Myocarditis, Macrophage

*Correspondence:
Jesus Jimenez
jesus.jimenez@wustl.edu
Full list of author information is available at the end of the article



[†]Jesus Jimenez and Nicolas Kostelecky are co-first authors.

Jimenez et al. Cardio-Oncology (2023) 9:14 Page 2 of 11

Introduction

The revolutionary new class of cancer immunotherapy, known as immune checkpoint inhibitors (ICIs), have resulted in dramatic improvements in outcomes for previously treatment-resistant cancers. A high proportion of cancer patients receive ICIs as either standard clinical practice or through clinical trials that study ICIs as primary therapy or in combination with traditional treatment regimens [1]. Several monoclonal antibodies have been developed to target specific immune checkpoint components. Examples include nivolumab and pembrolizumab that target the programmed cell death protein 1 (PD-1) receptor, atezolizumab that targets programmed cell death ligand 1 (PD-L1), and ipilimumab that targets the cytotoxic T lymphocyte-associated protein 4 (CTLA-4) receptor [2-4]. When activated, these receptors inhibit the intrinsic immune response to cancer cells, allowing cancer cells to avoid detection [5]. By inhibiting PD-1/ PD-L1 and CTLA-4 signaling, ICIs permit activation of the cytotoxic immune response against tumor cells.

ICI's have a wide array of immune-related adverse events including tachyarrhythmias, heart failure, vasculitis, and myocardial infarction [6]. However, fulminant myocarditis is one of the most serious adverse events, with an incidence up to 1% and with fatality rates among the highest of any reported adverse event [7, 8]. The American Society of Clinical Oncology (ASCO) and National Comprehensive Cancer Network (NCCN) have published clinical practice guidelines to aid in diagnosis, including a myocarditis grading system to guide management of patients with ICI myocarditis [9, 10]. However, a definitive diagnosis is often elusive and frequently requires a multi-modal approach [11–13]. From a Cardio-Oncology perspective, considerable effort has been made to standardize the definition of ICI-associated myocarditis by grouping the certainty of the diagnosis into categories of Possible, Probable, and Definite myocarditis [14, 15].

Confirming the diagnosis of ICI myocarditis is particularly challenging given the limited availability of endomyocardial biopsy (EMB) and strict pathological criteria to support this diagnosis. The Dallas criteria for the pathologic diagnosis of myocarditis by EMB requires cardiomyocyte damage in conjunction with the presence of an inflammatory infiltrate that may include lymphocytes, macrophages, eosinophils, neutrophils or a mixture of these immune cells. While these findings can have high specificity for myocarditis, the Dallas criteria have been associated with poor sensitivity in part due to tissue sampling bias and intra-observer variability with interpretation [16]. The diagnostic yield of EMB can be improved with the utilization of immunohistochemistry (IHC) to more readily identify inflammatory infiltrates such as

macrophages, which comprise the bulk of tissue resident cardiac immune cells [17, 18]. In healthy hearts, there are roughly 17 macrophages (CD68⁺) per high powered field (HPF), with similar number of macrophages in the early phase following acute myocardial infarction [19, 20]. Previous studies have demonstrated that > 30 CD68⁺ macrophages/HPF are associated with high grade ICI-associated myocarditis [21].

Given the emergence of specific treatments for ICI myocarditis, the ability to make an accurate and expeditious diagnosis represents an important gap in our ability to care for these patients [22–24]. Here, we tested the possibility that quantifying macrophage abundance by IHC may improve the yield of EMB in establishing the diagnosis of ICI-associated myocarditis.

Methods

Patient selection

In this single-center retrospective case series, 26 patients with a suspected clinical diagnosis of ICI-associated myocarditis were identified from November 2017 through March 2022. ICI myocarditis was suspected in the setting of symptoms such as dyspnea, palpitations, and chest pain in conjunction with suggestive cardiac biomarkers, ECG, and/or TTE abnormalities not attributable to an alternate non-myocarditis diagnosis [14]. Each case was independently reviewed by two cardiologists and the clinical diagnosis of ICI-associated myocarditis was categorized as: Definite (EMB positive OR CMR positive plus 2 minor criteria), Probable (CMR positive plus 1 minor criterion OR≥3 minor criteria and EMB/ CMR not obtained), or Possible (CMR positive without minor criteria OR < 2 minor criteria) [15]. By consensus among institutional experts and integrating ASCO and NCCN guidelines, major criteria for ICI-associated myocarditis included: 1) positive EMB (based on the Dallas Criteria), and 2) positive CMR (by updated Lake Louise criteria [25]). Minor criteria included: 1) ECG changes (significant T wave changes indicating ischemia, any new conduction abnormality or new supraventricular or ventricular tachyarrhythmias), 2) elevated cardiac troponin above institutional normal value, 3) TTE abnormality (wall motion abnormality, left ventricular (LV) dysfunction with EF<50%, or new/worsening pericardial effusion), and 4) inflammation such as myositis, myasthenia gravis or other major organ inflammation related to recent ICI use.

Demographics, medical comorbidities, cancer treatment history, cardiovascular (CV) risk factors, and medications at the time of first ICI treatment were extracted from the electronic medical record. The CMR report, cardiac related biomarkers, TTEs and ECGs were obtained at the time of suspected myocarditis. TTEs and

Jimenez et al. Cardio-Oncology (2023) 9:14 Page 3 of 11

ECGs were retrieved from digital archive and re-analyzed in accordance with published guidelines [26]. The study was approved by the Washington University in St. Louis institutional review board (ID #201,909,079) and the requirement for written informed consent was waived.

Histologic staining and evaluation

Among 17 patients who underwent EMB, three to five pieces of endomyocardial tissue were obtained during routine clinical practice for each patient. EMB specimens were examined microscopically via routine H&E staining as well as IHC for CD68 (KP-1 clone, Cell Marque, Rocklin CA). Staining was performed per manufacturer protocol in Clinical Laboratory Improvement Amendments (CLIA)-accredited clinical laboratories, with appropriate positive and negative controls. EMB specimens were retrospectively evaluated by a single pathologist.

H&E sections were evaluated for evidence of myocarditis using the Dallas criteria [16], and a diagnosis of myocarditis was rendered with the presence of cardiomyocyte damage/necrosis with associated inflammatory infiltrates that included a mixture of macrophages and lymphocytes. Cases were denoted as H&E positive if the pathologic sample met the Dallas criteria. Serial sequential IHC sections were evaluated for increased presence of CD68⁺ interstitial macrophages compared to the degree of scattered residential macrophages seen in the healthy heart and in patients with suspected ICI-associated myocarditis. Cases were denoted as IHC positive if there were greater than 50 interstitial CD68-stained macrophages/ hpf in hot spot regions (based on 1 standard deviation from 30 macrophages in high grade ICI-associated myocarditis and number of macrophages in healthy and postinjury hearts [19-21]).

The clinical diagnosis of ICI-associated myocarditis was re-assessed by the same two cardiologists using the supplemental IHC analyses, according to the diagnostic algorithm outlined above.

Statistical analysis

Continuous variables are reported as mean with standard deviation. Categorical variables are reported as total numbers and percentages. For continuous variables, data with 3 groups were compared using analysis of variance (ANOVA) with Bonferroni post-hoc analysis. For categorical variables, data were compared using Fisher's exact test. For correlation between cell count and biomarkers, Spearman correlation coefficient analysis was performed. For survival curves, data were compared using the Mantel-Cox log-rank test. Values of P < 0.05 were considered statistically significant.

Results

Histological analysis of ICI-associated myocarditis

In the 17 patients who underwent EMB, histological examination was performed with at least three levels of H&E-stained slides for assessment using the Dallas criteria. Two patients met the definition of myocarditis using routine H&E stain, having both cardiomyocyte injury/necrosis and focal inflammatory infiltrate comprised of macrophages and lymphocytes (Fig. 1B). Supplemental IHC staining for CD68 in serial sequential sections demonstrated interstitial macrophage infiltration in these two cases (Fig. 1E). In the remaining 15 EMBs that did not meet the Dallas criteria for myocarditis on H&E staining (Fig. 1A, C), supplemental IHC staining for CD68 demonstrated increased macrophage infiltration in 7 samples (Fig. 1F). EMBs demonstrating greater than 50 macrophages/HPF were considered positive by IHC criteria. Eight samples were classified as negative for IHC due to staining showing less than 50 macrophages/HPF (Fig. 1D).

Clinical myocarditis classification

In patients that underwent EMB, the initial clinical diagnosis was based on the major criteria for ICIassociated myocarditis, which requires a positive EMB based on the Dallas Criteria (see Methods). Based on positive CD68 IHC staining, we show potential clinical reclassification of patients from Possible or Probable to Definite myocarditis (Fig. 2). Two of the 8 patients originally classified as *Possible* myocarditis could be reclassified as Definite, given pathologic support of the diagnosis through evidence of cardiac inflammation with CD68 staining. Four of the 13 patients originally classified as *Probable* could be reclassified as *Definite* after confirming inflammation with CD68 staining of the EMB. In combination with the 2 patients who met both Dallas criteria and positive CD68 staining, a total of 9 patients could now meet Definite criteria through biopsy criteria. Two patients did not undergo biopsy but met Definite criteria through clinical criteria and imaging confirmation. Nine patients remained classified as Probable myocarditis and 6 remained classified as Possible myocarditis before and after CD68 staining. Collectively, the mean age for all patients was 64 ± 12 years and 38% were female (Table 1). Less than 50% of patients had prior coronary artery disease or cardiomyopathy and over 40% of patients were on at least one cardiovascular medication. No significant differences in clinical features were observed across the classifications of ICI-associated myocarditis.

Jimenez et al. Cardio-Oncology (2023) 9:14 Page 4 of 11

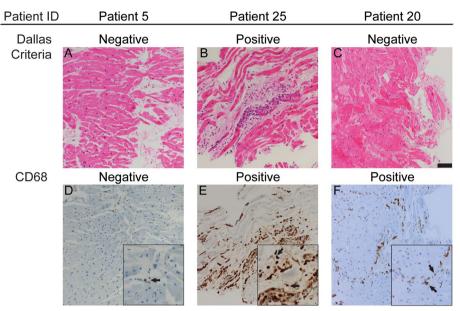


Fig. 1 Histology and immunohistochemistry (IHC) of endomyocardial biopsies from patients treated with immune checkpoint inhibitors. Representative areas of each respective biopsy sample that underwent staining from serial sequential sections are shown. The hematoxylin and eosin (H&E)-stained slides did not show definite evidence of myocarditis by Dallas criteria in either **A** Patient 5 or **C** Patient 20 but did demonstrate cardiomyocyte necrosis and loss with increased inflammatory infiltrates in **B** Patient 25. The CD68 stain showed an increased number of interstitial macrophages that overlap with the inflammatory cell infiltration and the cardiomyocyte necrosis perceived in H&E-stain in **E** Patient 25. CD68 macrophage stain also showed an increased number of interstitial macrophages in **F** Patient 20 but not in **D** Patient 5. Insets show magnified views with arrows highlighting representative CD68 stained macrophages. Scale bar = 100 μm. ID = identification

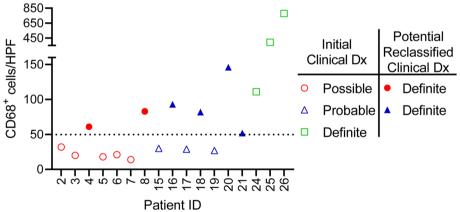


Fig. 2 Interstitial macrophage cell counts in endomyocardial biopsies (EMB). Plot of CD68 positive macrophages per high powered field (HPF) from EMBs of patients with clinical diagnosis of *Possible* (red circles), *Probable* (blue triangles), and *Definite* (green squares) myocarditis. The dotted line at 50 macrophages/HPF demarcates the CD68⁺ cell count threshold to consider a sample as immunohistochemistry positive and support reclassification of the initial clinical diagnosis (Dx)

Cardiac diagnostic testing

Overall, 16 patients underwent CMR and had an average left ventricular ejection fraction (LVEF) of 42%. Eight patients met Lake Louise Criteria for myocarditis (Table 2). Baseline TTE before ICI treatment was available for 20 patients with an average LVEF of 57%. The post ICI treatment TTE had an average LVEF of 43%

with a decrease in LVEF of 14% from baseline. Several abnormal findings were seen across all groups, including 1) reduction in LVEF, wall motion abnormalities, or new/worsening pericardial effusions in 18/26 patients (69%), 2) ECG abnormalities such as ischemic T wave changes, new conduction abnormality or new tachyarrhythmias in 20/26 patients (91%), 3) cardiac troponin

Jimenez et al. Cardio-Oncology (2023) 9:14 Page 5 of 11

Table 1 Patient characteristics – cardiovascular

	AII (n = 26)	Possible $(n=6)$	Probable $(n=9)$	Definite $(n = 11)$	P	P (Pos vs Pro)	P (Pos vs Def)	P (Pro vs Def)
Age at clinical diagnosis (yr)	64±12	65±8	63±9	65±16	0.904			
Sex								
Female	10 (38)	1 (17)	4 (44)	5 (45)		0.999	0.333	0.637
Male	16 (62)	5 (83)	5 (56)	6 (55)				
Race								
Black or African American	3 (12)	0	2 (22)	1 (9)		0.486	0.999	0.566
White, non-Hispanic	23 (88)	6 (100)	7 (78)	10 (91)				
Cardiovascular Risk Factors								
History of smoking	17 (65)	6 (100)	2 (22)	9 (82)		0.0070	0.515	0.0216
Hypertension	19 (73)	4 (67)	8 (89)	7 (64)		0.525	0.999	0.319
Diabetes mellitus	6 (23)	1 (17)	3 (33)	2 (18)		0.604	0.999	0.617
Coronary Artery Disease	12 (46)	3 (50)	2 (22)	7 (64)		0.329	0.645	0.092
Prior Cardiomyopathy (any)	8 (31)	3 (50)	1 (11)	4 (36)		0.559	0.999	0.282
Stroke	2 (8)	0	0	2 (18)		0.999	0.515	0.479
Cardiovascular Medications								
Statin	11 (42)	3 (50)	3 (33)	5 (45)		0.622	0.999	0.670
Aspirin	15 (58)	4 (67)	4 (44)	7 (64)		0.608	0.999	0.653
Beta-blockers	13 (50)	5 (83)	1 (11)	7 (64)		0.0110	0.600	0.0281
RAAS inhibitors	18 (69)	6 (100)	5 (56)	7 (64)		0.103	0.237	0.999

Values are expressed as mean \pm standard deviation or n (%). Fisher's exact test was used when comparing categorical variables and Student's t-test or analysis of variance when comparing continuous variables. P values < 0.05 were considered significant. Possible, Probable, and Definite are based on the proposed reclassified myocarditis patient cohorts

RAAS Renin-angiotensin-aldosterone system, yr years

I elevation in 16/24 patients (67%) and 4) inflammation including myositis or other major organ inflammation in 4/26 patients (15%). NT-proBNP was elevated in all patients at the time of clinical diagnosis and CRP was elevated in 12/18 patients (67%). We identified a significant correlation between CD68⁺ macrophage abundance and serum biomarkers of cardiac injury including troponin I (P=0.026) and NT-proBNP (P=0.047) (Fig. 3A, B).

Cancer characteristics and clinical outcomes

The most common cancer types were non-small cell lung cancer, renal cell carcinoma, and melanoma (Table 3). Patients often receive ICIs following or in conjunction with standard chemotherapy and/or radiation. Fifty-eight percent of patients underwent radiation of which 15% received chest radiation. Eighty-eight percent of patients had prior cancer treatment with 7% receiving anthracyclines and 27% vascular endothelial growth factor (VEGF) inhibitors. The majority of patients were treated with ICI monotherapy that included nivolumab (31%) and pembrolizumab (31%) while 27% were treated with combination therapy comprised of nivolumab and ipilimumab.

Under the potential reclassification, the total number of ICI treatment cycles until the clinical diagnosis of myocarditis would be significantly different among the three cohorts (P = 0.001). Post-hoc analysis revealed that patients reclassified into the *Probable* (P=0.002)and Definite (P=0.002) groups may present at earlier time points compared to the Possible group (Fig. 4A). In evaluating the number of months between cycle 1 to clinical diagnosis of myocarditis, there was a significant difference among the three groups (P=0.005) with 2 group analysis showing that patients reclassified into the Probable (P=0.047) and Definite (P=0.003) groups may present after fewer cycles compared to the *Possible* group (Fig. 4B). For all 26 patients including those with potential reclassification, the time from cycle 1 of ICI treatment to clinical diagnosis occurred within 9 months for 100% of patients in both the *Probable* and *Definite* groups compared to only 25% for the *Possible* group. There was a trend (P=0.08) in survival probability at 15 months among the three groups from the time following cycle 1 of ICI treatment that suggests potential reduced survival in the Probable and Definite groups (Fig. 4C). Almost 41% of patients died within 8 weeks of initial diagnosis despite steroid treatment (Table 4).

Jimenez et al. Cardio-Oncology (2023) 9:14 Page 6 of 11

Table 2 Patient characteristics – diagnostic testing

All	Possible	Probable	Definite	P	P (Pos vs Pro)	P (Pos vs Def)	P (Pro vs Def)
42 ± 20	37 ± 13	47 ± 20	41 ± 22	0.756			
8 (57)	2 (50)	2 (67)	4 (57)		0.999	0.999	0.999
10 (63)	3 (75)	3 (60)	4 (57)		0.999	0.999	0.999
phy							
57 ± 13	44 ± 16	61 ± 7	61 ± 10	0.0382	0.066	0.065	0.999
43 ± 19	38 ± 14	44 ± 21	45 ± 19	0.778			
-15±17	-4±14	-19±18	-17 ± 16	0.419			
24 ± 8	25 ± 9	26±9	23 ± 7	0.575			
7 (29)	2 (33)	3 (43)	2 (18)		0.999	0.517	0.617
20 (91)	3 (75)	7 (88)	10 (100)		0.999	0.286	0.444
6 (27)	2 (50)	1 (13)	3 (30)		0.236	0.580	0.588
12 (55)	1 (25)	4 (50)	7 (70)		0.576	0.245	0.631
13 (59)	1 (25)	6 (75)	6 (60)		0.222	0.559	0.638
5 (19)	1 (17)	3 (33)	1 (9)		0.604	0.999	0.285
4.4 ± 17.4	0.2 ± 0.2	12 ± 29	0.9 ± 0.9	0.357			
9,896 ± 18,841	1,786 ± 969	17,267 ± 24,549	8,791 ± 16,928	0.398			
	42±20 8 (57) 10 (63) phy 57±13 43±19 -15±17 24±8 7 (29) 20 (91) 6 (27) 12 (55) 13 (59) 5 (19) 4.4±17.4	42 ± 20 37 ± 13 $8 (57)$ $2 (50)$ $10 (63)$ $3 (75)$ 10 (63) $3 (75)$ 10 (63) $3 (75)$ 10 (64) $3 (75)$ 10 (65) $3 (75)$ 11 (16) $3 (75)$ 12 (17) $3 (75)$ 12 (18) $3 (75)$ 13 (18) $3 (75)$ 14 (18) $3 (75)$ 15 (18) $3 (75)$ 16 (18) $3 (75)$ 17 (18) $3 (75)$ 18 (19) $3 (75)$ 19 (10) $3 (75)$ 10 (17) $3 (75)$ 11 (17) $3 (75)$ 11 (17) $3 (75)$ 12 (18) $3 (75)$ 13 (19) $3 (75)$ 14 (17) $3 (75)$ 15 (19) $3 (75)$ 16 (27) $3 (75)$ 17 (28) $3 (75)$ 18 (29) $3 (75)$ 19 (29) $3 (75)$ 10 (21) $3 (75)$ 11 (25) $3 (75)$ 11 (25) $3 (75)$ 12 (25) $3 (75)$ 13 (25) $3 (75)$ 14 (25) $3 (75)$ 15 (17) $3 (75)$ 16 (27) $3 (75)$ 17 (28) $3 (75)$ 18 (29) $3 (75)$ 19 (20) $3 (75)$ 10 (21) $3 (75)$ 11 (25) $3 (75)$ 11 (25) $3 (75)$ 11 (25) $3 (75)$ 12 (25) $3 (75)$ 13 (25) $3 (75)$ 14 (25) $3 (75)$ 15 (17) $3 (75)$ 16 (27) $3 (75)$ 17 (28) $3 (75)$ 18 (29) $3 (75)$ 19 (20) $3 (75)$ 20 (20) $3 (75)$ 21 (25) $3 (75)$ 22 (25) $3 (75)$ 23 (25) $3 (75)$ 24 (25) $3 (75)$ 25 (26) $3 (75)$ 26 (27) $3 (75)$ 27 (28) $3 (75)$ 28 (28) $3 (75)$ 29 (29) $3 (75)$ 20 (29) $3 (75)$ 20 (29) $3 (75)$ 20 (29) $3 (75)$ 21 (25) $3 (75)$ 22 (25) $3 (75)$ 23 (25) $3 (75)$ 24 (25) $3 (75)$ 25 (26) $3 (75)$ 26 (27) $3 (75)$ 27 (28) $3 (75)$ 28 (28) $3 (75)$ 29 (29) $3 (75)$ 20 (29) $3 (75)$ 20 (29) $3 (75)$ 20 (29) $3 (75)$ 20 (29) $3 (75)$ 20 (29) $3 (75)$ 21 (25) $3 (75)$ 22 (25) $3 (75)$ 23 (25) $3 (75)$ 24 (27) $3 (75)$ 25 (28) $3 (75)$ 26 (28) $3 (75)$ 27 (29) $3 (75)$ 28 (28) $3 (75)$ 29 (29) $3 (75)$ 20 (29) $3 (75)$ 20 (29) $3 (75)$ 20 (29) $3 (75)$ 20 (29) $3 (75)$ 20 (29) $3 (75)$ 21 (25) $3 (75)$ 22 (25) $3 (75)$ 23 (25) $3 (75)$ 24 (27) $3 (75)$ 25 (28) $3 (75)$ 26 (28) $3 (75)$ 27 (29) $3 (75)$ 28 (28) $3 (75)$ 29 (29) $3 (75)$ 20 (29) $3 (75)$ 20 (29) $3 (75)$ 20 (29) $3 (75)$ 20 (29) $3 (75)$ 20 (29) $3 (75)$ 20 (29) $3 (75)$ 20 (29) $3 (75)$ 20 (29) $3 (75)$ 20 (29) $3 (75)$ 20 (29) $3 (75)$ 20 (20) $3 (75)$ 20 (20) $3 (75)$ 20 (20) $3 $	42 ± 20 37 ± 13 47 ± 20 $8 (57)$ $2 (50)$ $2 (67)$ $10 (63)$ $3 (75)$ $3 (60)$ 10 (63) $3 (75)$ $3 (60)$ 10 (63) $3 (75)$ $3 (60)$ 10 (63) $3 (75)$ $3 (60)$ 10 (63) $3 (75)$ $3 (60)$ 11 (12 (14 - 14 - 15 - 15 - 17 - 15 - 17 - 14 - 14 - 19 - 18 - 18 - 19 - 18 - 18 - 19 - 18 - 19 - 18 - 19 - 18 - 19 - 18 - 19 - 18 - 19 - 18 - 19 - 18 - 19 - 18 - 19 - 19	42±20 37±13 47±20 41±22 8 (57) 2 (50) 2 (67) 4 (57) 10 (63) 3 (75) 3 (60) 4 (57) 10 (63) 3 (75) 3 (60) 4 (57) 10 (63) 3 (75) 3 (60) 4 (57) 10 (63) 3 (75) 3 (60) 4 (57) 10 (63) 44±16 61±7 61±10 43±19 38±14 44±21 45±19 -15±17 -4±14 -19±18 -17±16 24±8 25±9 26±9 23±7 7 (29) 2 (33) 3 (43) 2 (18) 20 (91) 3 (75) 7 (88) 10 (100) 6 (27) 2 (50) 1 (13) 3 (30) 12 (55) 1 (25) 4 (50) 7 (70) 13 (59) 1 (25) 6 (75) 6 (60) 5 (19) 1 (17) 3 (33) 1 (9) 4.4±17.4 0.2±0.2 12±29 0.9±0.9 9,896±18,841 1,786±969 17,267±24,549 8,791±16,928	42±20 37 ± 13 47 ± 20 41 ± 22 0.756 8 (57) 2 (50) 2 (67) 4 (57) 10 (63) 3 (75) 3 (60) 4 (57) phy 57 ± 13 44 ± 16 61 ± 7 61 ± 10 0.0382 43 ± 19 38 ± 14 44 ± 21 45 ± 19 0.778 -15 ± 17 -4 ± 14 -19 ± 18 -17 ± 16 0.419 24 ± 8 25 ± 9 26 ± 9 23 ± 7 0.575 7 (29) 2 (33) 3 (43) 2 (18) 20 (91) 3 (75) 7 (88) 10 (100) 6 (27) 2 (50) 1 (13) 3 (30) 12 (55) 1 (25) 4 (50) 7 (70) 13 (59) 1 (25) 6 (75) 6 (60) 5 (19) 1 (17) 3 (33) 1 (9) 4.4 ± 17.4 0.2 ± 0.2 12 ± 29 0.9 ± 0.9 0.357 $9,896\pm18,841$ $1,786\pm969$ $17,267\pm24,549$ $8,791\pm16,928$ 0.398	42 ± 20 37 ± 13 47 ± 20 41 ± 22 0.756 $8(57)$ $2(50)$ $2(67)$ $4(57)$ 0.999 $10(63)$ $3(75)$ $3(60)$ $4(57)$ 0.999 $10(63)$ $3(75)$ $3(60)$ $4(57)$ 0.999 57 ± 13 44 ± 16 61 ± 7 61 ± 10 0.0382 0.066 43 ± 19 38 ± 14 44 ± 21 45 ± 19 0.778 -15 ± 17 -4 ± 14 -19 ± 18 -17 ± 16 0.419 24 ± 8 25 ± 9 26 ± 9 23 ± 7 0.575 $7(29)$ $2(33)$ $3(43)$ $2(18)$ 0.999 $20(91)$ $3(75)$ $7(88)$ $10(100)$ 0.999 $6(27)$ $2(50)$ $1(13)$ $3(30)$ 0.236 $12(55)$ $1(25)$ $4(50)$ $7(70)$ 0.576 $13(59)$ $1(25)$ $6(75)$ $6(60)$ 0.222 $5(19)$ $1(17)$ $3(33)$ $1(9)$ 0.357 $9.896\pm18,841$ $1,786\pm969$ $17,267\pm24,549$ $8,791\pm16,928$ 0.398	42 ± 20 37 ± 13 47 ± 20 41 ± 22 0.756 0.999 0.999 $8 (57)$ $2 (50)$ $2 (67)$ $4 (57)$ 0.999 0.999 0.999 $10 (63)$ $3 (75)$ $3 (60)$ $4 (57)$ 0.999 0.999 0.999 $10 (63)$ $3 (75)$ $3 (60)$ $4 (57)$ 0.999 0.999 0.999 $10 (63)$ $3 (75)$ $3 (60)$ $4 (57)$ 0.999 0.999 0.999 $10 (63)$ $3 (75)$ $3 (60)$ $4 (57)$ 0.0382 0.066 0.065 43 ± 19 $3 8\pm14$ 44 ± 21 45 ± 19 0.778 0.778 0.419 24 ± 8 25 ± 9 26 ± 9 23 ± 7 0.575 0.575 0.999 0.517 $7 (29)$ $2 (33)$ $3 (43)$ $2 (18)$ 0.999 0.286 0.576 0.286 0.580 $12 (55)$ $1 (25)$ $4 (50)$ $7 (70)$ 0.576 0.245 $13 (59)$ $1 (25)$ $4 (50)$ $7 (70)$ 0.576 0.222

Values are expressed as mean \pm standard deviation or n (%). Fisher's exact test was used when comparing categorical variables and Student's t-test or analysis of variance when comparing continuous variables. P values < 0.05 were considered significant. The n are defined as the following for All, Possible, Probable, and Definite, respectively: ${}^{a}n = 16, 4, 5, 7$; ${}^{b}n = 14, 4, 3, 7$; ${}^{c}n = 20, 5, 7, 8$; ${}^{d}n = 25, 5, 9, 11$; ${}^{e}n = 18, 4, 6, 8$; ${}^{f}n = 17, 2, 7, 8$; ${}^{g}n = 24, 4, 9, 11$; ${}^{h}n = 22, 4, 8, 10$; ${}^{h}n = 26, 6, 9, 11$; ${}^{h}n = 24, 5, 8, 10$; ${}^{h}n = 10$; h

CRP C-reactive protein, ECG Electrocardiogram, ICI Immune checkpoint inhibitors, LVEF Left ventricle ejection fraction, MRI Magnetic resonance imaging, NT-proBNP N-terminal pro-brain natriuretic peptide

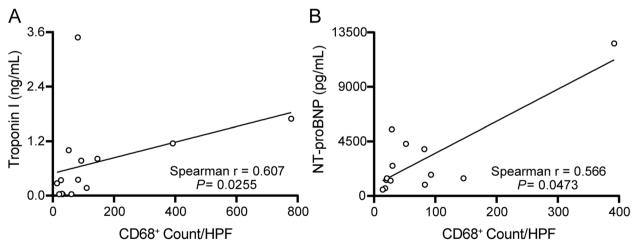


Fig. 3 Correlation of cardiac biomarkers and macrophage cell counts. **A** Cardiac troponin I and **B** N-terminal pro-brain natriuretic peptide (NT-pro BNP) levels plotted against corresponding CD68 positive macrophage cells counts per high powered field (HPF). Data were compared using Spearman correlation coefficient analysis with values of P < 0.05 being considered statistically significant

Jimenez et al. Cardio-Oncology (2023) 9:14 Page 7 of 11

Table 3 Patient characteristics – cancer

	AII	Possible	Probable	Definite	P	P (Pos vs Pro)	P (Pos vs Def)	P (Pro vs Def)
Primary Cancer Type ^a								
Non-small cell lung cancer	5 (19)	1 (17)	1 (11)	3 (27)		0.999	0.999	0.591
Renal cell carcinoma	5 (19)	2 (33)	3 (33)	0		0.999	0.110	0.074
Melanoma	6 (23)	2 (33)	0	4 (36)		0.143	0.999	0.094
Other cancer	10 (38)	1 (17)	5 (56)	4 (36)		0.287	0.600	0.653
Prior Radiation ^a								
Chest radiation	4 (15)	0	2 (40)	2 (18)		0.486	0.515	0.999
Other radiation	11 (42)	3 (100)	3 (60)	5 (45)		0.622	0.999	0.670
Prior Treatment ^a								
Anthracyclines	2 (7)	0	2 (22)	0		0.486	0.999	0.190
VEGF inhibitors	7 (27)	4 (67)	3 (33)	0		0.315	0.0063	0.074
Other cancer treatment	14 (54)	2 (33)	4 (45)	8 (73)		0.999	0.162	0.362
ICI Therapy ^a								
Nivolumab (anti-PD-1)	8 (31)	3 (50)	2 (22)	3 (27)		0.329	0.600	0.999
Ipilimumab (anti-CTLA-4)	1 (4)	0	1 (11)	0		0.999	0.999	0.450
Pembrolizumab (anti-PD-1)	8 (31)	3 (50)	2 (22)	3 (27)		0.329	0.600	0.999
Atezolizumab (anti-PD-L1)	2 (7)	0	1 (11)	1 (9)		0.999	0.999	0.999
Combination therapy (Nivolumab + Ipilimumab)	7 (27)	0	3 (33)	4 (36)		0.229	0.237	0.999
Total ICI Cycles ^a	9±12	23 ± 17	4 ± 3	5 ± 4	0.0011	0.0223	0.0222	0.999
Cycle 1 to clinical diagnosis (mo) ^a	9±13	22 ± 14	8 ± 13	3 ± 3	0.0085	0.069	0.0071	0.997
Cycle 1 to death (mo) ^b	16 ± 17	32 ± 20	6 ± 4	12 ± 12	0.0078	0.0085	0.0392	0.999
Clinical diagnosis to death (mo) ^b	7 ± 10	10 ± 8	3 ± 3	9±12	0.325			
Deceased ^a	22 (85)	6 (100)	7 (78)	9 (82)		0.486	0.515	0.999

Values are expressed as mean \pm standard deviation or n (%). Fisher's exact test was used when comparing categorical variables and Student's t-test or analysis of variance when comparing continuous variables. P values < 0.05 were considered significant. The n are defined as the following for All, P ossible, P robable, and P Definite, respectively: P = 26, 6, 9, 11; P = 22, 6, 7, 9. P ossible, P robable, and P of the proposed reclassified myocarditis patient cohorts

CTLA-4 Cytotoxic T lymphocyte-associated protein 4, ICI Immune checkpoint inhibitors, mo months, PD-1 Programmed cell death protein 1, PD-L1 Programmed cell death ligand-1, VEGF Vascular endothelial growth factor

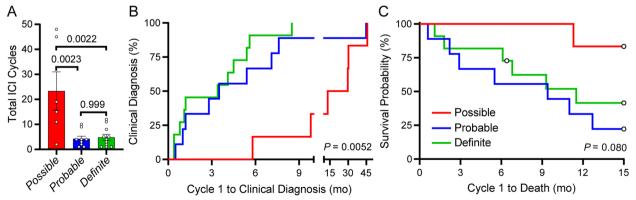


Fig. 4 Total immune checkpoint inhibitor (ICI) cycles, time interval, and survival probability between first exposure to clinical diagnosis and death in reclassified patients. **A** Comparison of total ICI cycles. **B** Percent of patients with clinical diagnosis for myocarditis following cycle 1 of ICI treatment. **C** Percent survival probability of patients at 15 months following cycle 1 of ICI treatment. The n are defined as Possible (n = 6), Probable (n = 9), and Possible (n = 11) using the reclassified myocarditis patient cohorts. Open circle symbols represent censored patients in Possible (n = 5), Probable (n = 2), and Possible (n = 5) cohorts. Data were compared using the Mantel-Cox log-rank test with P < 0.05 being considered statistically significant

Jimenez et al. Cardio-Oncology (2023) 9:14 Page 8 of 11

 Table 4
 Steroid treatment before and after potential reclassification

	Initial Clinical Diagno	osis	Potential Reclassified Diagnosis			
Steroid	Possible $(n=8)$	<i>Probable</i> (<i>n</i> = 13)	Definite (n = 5)	Possible (n = 6)	<i>Probable</i> (<i>n</i> = 9)	Definite (n = 11)
High	3 (38)	8 (62)	3 (60)	2 (33)	5 (56)	7 (64)
Other	1 (12)	3 (23)	1 (20)	1 (17)	3 (33)	1 (9)
None	4 (50)	2 (15)	1 (20)	3 (50)	1 (11)	3 (27)

Values are expressed as *n* (%). High dose steroids are defined as the equivalent of intravenous methylprednisolone of 1 gm or higher and Other steroids are defined as less than 1 gm

Discussion

While the Dallas criteria can identify a subset of myocarditis patients, this study demonstrates that quantification of macrophage abundance adds additional information that correlates with clinical presentation and assists in establishing the diagnosis of the ICI-associated myocarditis. In this study, we present 26 cases of suspected ICI-associated myocarditis with detailed clinical evaluation and outcomes. After applying standard accepted tools for the diagnosis and classification of ICI-associated myocarditis [14, 15], only 5/26 patients met criteria for Definite myocarditis. Of the 17 patients that underwent EMB, only 2 patients satisfied the Dallas criteria for myocarditis. When supplemental IHC staining in EMBs was performed, 7 additional patients could meet criteria for pathologic cardiac inflammation given positive CD68-IHC staining (>50 CD68+ cells/HPF) and may be reclassified as having a Definitive diagnosis of ICI-associated myocarditis. The remaining 8 patients would maintain their original classification (Fig. 5A, B). These data highlight that deeper immunophenotyping of myocardial inflammation may increase the diagnostic yield of EMB.

Pathologic presentations of ICI-associated myocarditis may vary based on severity of disease, sampling bias, prior immunosuppressive treatment, cardiovascular comorbidities, and concomitant cardiotoxic cancer therapies [21, 22, 27, 28]. The utility of EMB for the diagnosis of ICI-associated myocarditis has many potential limitations which are only partially explained by sampling bias and inter-observer variability. There are established limitations of the Dallas criteria [16], and likely contribute to the underdiagnosis of ICI-associated myocarditis. Other important clinical observations limiting EMB as a crucial diagnostic test in ICI-associated myocarditis include the perceived additional risks related to the procedure, availability of procedural expertise, and availability of pathology expertise to perform detailed IHC testing. Despite these limitations, major professional groups advising about cardiac safety during ICI therapy have recommended EMB as part of the diagnostic paradigm [9, 10, 14].

Although it has been shown that the diagnostic utility of the Dallas criteria can be improved by the use of IHC [17], this has not been well established for ICI-associated myocarditis. As a means to screen EMB for ICI-associated myocarditis, recent work by Palaskas et al. [28] proposes the use of light and electron microscopy in addition to IHC staining while Champion et al. [21] propose the use of the ISHLT (International Society of Heart and Lung Transplantation) criteria that is used for acute cellular rejection. Given the significant variability, it appears that ICI-associated myocarditis presents histologically along a spectrum. Hence, a gap in the literature is the lack of a "gold standard" for diagnosis, which presents uncertainty as to which classification scheme, H&E alone or H&E and IHC, is clinically superior.

Identifying the optimal IHC panel represents an important next step. However, there is no consensus for which IHC stains should be performed in cases of suspected ICI-associated myocarditis and several studies have reported a wide array of IHC staining including CD4, CD8, CD3, CD68, CD20, PD-1, and PD-L1 [21, 22, 27, 28]. While the heart is interspersed with a variety of immune cells in the steady state, the bulk of immune cells within the heart are comprised of tissue resident macrophages [18, 29]. Macrophage numbers in the heart are dynamic, increasing with aging, hemodynamic stress, and myocardial injury [30]. Myocardial infiltration of CD68⁺ macrophages has previously been described in ICI-associated myocarditis with rapid and late onset [27]. Notably, the proposed reclassified *Definite* patient cohort is associated with an earlier onset of myocarditis following cycle 1 of treatment compared to the Possible group. In addition, survival outcomes were close to achieving statistical significance with the Definite group trending towards lower survival probability compared to the Possible cohort. These findings serve to highlight that patients with Definite ICI-associated myocarditis may be at risk for earlier onset myocarditis and decreased survival probability. The seriousness of the potential clinical diagnosis reaffirms that early aggressive treatment should be instituted for patients with Definite ICI-associated myocarditis.

Jimenez et al. Cardio-Oncology (2023) 9:14 Page 9 of 11

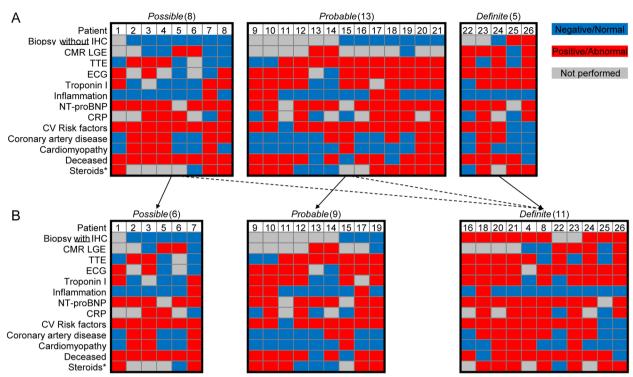


Fig. 5 Myocarditis classification with and without supplemental immunohistochemistry (IHC) staining. Heatmaps of standard myocarditis classification criteria and selected variables from patients with immune checkpoint inhibitor-associated myocarditis. **A** Myocarditis classification without IHC and **B** potential reclassification with CD68 positive IHC. Each column represents characteristics from one patient (n = 26). Blue boxes represent negative/normal data, red boxes represent positive/abnormal data, and gray boxes represent studies not performed. *For steroids, red boxes represent high dose steroids that are defined as the equivalent of intravenous methylprednisolone of 1 gm or higher, blue boxes represent other steroids which are defined as a less than 1 gm, and gray boxes represent no steroids given. CMR = cardiac magnetic resonance imaging; CRP = C-reactive protein; CV = cardiovascular; ECG = electrocardiogram; LGE = late gadolinium enhancement; NT-proBNP = N-terminal pro-brain natriuretic peptide

Study limitations

This study requires additional validation of CD68 IHC staining in EMBs from patients with suspected ICI-associated myocarditis. Increased macrophage infiltration can be seen in other forms of cardiac injury such as acute myocardial infarction and heart failure. However, we anticipate that CD68 IHC staining will improve the sensitivity of diagnosing ICI-associated myocarditis in the appropriate clinical context. This limitation is not unique to macrophages since increased T-cell (CD3) and PD-L1 staining is also observed with non-myocarditis pathology [31]. Our study was limited by the small number of patients that underwent an EMB, although this is within clinical expectations as patients may already be presumed to have ICI-associated myocarditis or be too sick to undergo an EMB. We recognize that this is a retrospective study and clinical information is limited to what is recorded in the medial record and not all CMR studies had T1 and T2 mapping. Finally, we recognize inconsistencies between CMR and EMB biopsy findings that relate to sampling bias and inherent limitation of magnetic imaging without the use of T1 mapping and application of the modified Lake Louise Criteria [32, 33]. Our initial findings support the rationale of designing a prospective multi-center cohort study to address the limitations we have highlighted.

Conclusions

Collectively, our findings highlight the possible utility of supplemental CD68 IHC staining when performing EMB to aid in the diagnosis of ICI-associated myocarditis. Given that ICI-associated myocarditis carries significant morbidity and mortality, early and accurate diagnosis is of paramount importance as it will improve the implementation of appropriate immunosuppressive therapy.

Perspectives

Competency in medical knowledge

Immune checkpoint inhibitor (ICI) myocarditis is associated with significant morbidity and mortality. Endomyocardial biopsy (EMB) is considered one of the gold standards for diagnosis; however, the sensitivity of

Jimenez et al. Cardio-Oncology (2023) 9:14 Page 10 of 11

biopsies and the pathological features that correlate with the clinical diagnosis of myocarditis remain incompletely defined. While the Dallas criteria can identify a subset of myocarditis patients, quantification of macrophage abundance adds additional information that correlates with clinical presentation and assists in establishing the diagnosis of the ICI-associated myocarditis.

Translational outlook

Limitations exist with solely using Dallas criteria to diagnose ICI-associated myocarditis from EMB and these findings suggest that deeper immunophenotyping of myocardial inflammation may increase the diagnostic yield of EMB.

Abbreviations

ASCO American Society of Clinical Oncology CD68 Cluster of differentiation 68 CMR Cardiac magnetic resonance imaging

CRP C-reactive protein

CTLA-4 Cytotoxic T lymphocyte-associated protein 4

Dx Diagnosis
ECG Electrocardiogram
EMB Endomyocardial biopsy
HPF High powered field

ICI Immune checkpoint inhibitors

 ID
 Identification

 IHC
 Immunohistochemistry

 LVEF
 Left ventricle ejection fraction

 MRI
 Magnetic resonance imaging

NCCN National Comprehensive Cancer Network
NT-proBNP N-terminal pro-brain natriuretic peptide
PD-1 Programmed cell death protein 1
PD-L1 Programmed cell death ligand-1
RAAS Renin-angiotensin-aldosterone system
VEGF Vascular endothelial growth factor

Authors' contributions

JJ, NK, DJL, CYL and KJL were responsible for conceptualization of the study. KJL, DJL, JJ, JDM, NK, KWZ, and CYL contributed to experimental design, acquired data, and performed data analysis. NK and CYL performed histopathological analysis. DJL, JJ, NK, KWZ, CYL, KJL extracted electronic medical patient information. JJ and KJL performed statistical analysis. JJ and NK wrote the original draft of the manuscript, and all authors approved the final version.

Funding

JJ is supported by funding provided by the National Institutes of Health (R25 HL105400) and the Paul and Patti Eisenberg Scholar Award of Washington University in the Division of Cardiology. KJL is supported by funding provided by the Children's Discovery Institute of Washington University and St. Louis Children's Hospital, the Foundation of Barnes-Jewish Hospital, the Burroughs Foundation Welcome Fund, the Leducq Foundation, and the National Institutes of Health (R35 HL161185, RO1 HL151078, R21 Al148877).

Availability of data and materials

All data generated or analyzed during this study are included in this published article.

Declarations

Ethics approval and consent to participate

The study was approved by the Washington University in St. Louis institutional review board (ID #201909079) and the requirement for written informed consent was waived.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

Author detail:

¹Center for Cardiovascular Research, Department of Medicine, Cardiovascular Division, Washington University School of Medicine, 660 South Euclid Campus, Box 8086, St. Louis, MO 63110, USA. ²Cardio-Oncology Center of Excellence, Department of Medicine, Cardiovascular Division, Washington University School of Medicine, 660 South Euclid Campus, Box 8086, St. Louis, MO 63110, USA. ³Department of Pathology and Immunology, Washington University School of Medicine, St. Louis, MO, USA. ⁴Department of Developmental Biology, Washington University School of Medicine, St. Louis, MO, USA.

Received: 16 November 2022 Accepted: 10 March 2023 Published online: 13 March 2023

References

- Akinleye A, Rasool Z. Immune checkpoint inhibitors of PD-L1 as cancer therapeutics. J Hematol Oncol. 2019;12:92.
- Nishimura H, Nose M, Hiai H, Minato N, Honjo T. Development of lupuslike autoimmune diseases by disruption of the PD-1 gene encoding an ITIM motif-carrying immunoreceptor. Immunity. 1999;11:141–51.
- 3. Dong H, Zhu G, Tamada K, Chen L. B7–H1, a third member of the B7 family, co-stimulates T-cell proliferation and interleukin-10 secretion. Nat Med. 1999;5:1365–9.
- 4. Leach DR, Krummel MF, Allison JP. Enhancement of antitumor immunity by CTLA-4 blockade. Science. 1996;271:1734–6.
- Seidel JA, Otsuka A, Kabashima K. Anti-PD-1 and Anti-CTLA-4 Therapies in Cancer: Mechanisms of Action, Efficacy, and Limitations. Front Oncol. 2018;8:86
- Puzanov I, Diab A, Abdallah K, et al. Managing toxicities associated with immune checkpoint inhibitors: consensus recommendations from the Society for Immunotherapy of Cancer (SITC) Toxicity Management Working Group. J Immunother Cancer. 2017;5:95.
- Salem JE, Manouchehri A, Moey M, et al. Cardiovascular toxicities associated with immune checkpoint inhibitors: an observational, retrospective, pharmacovigilance study. Lancet Oncol. 2018;19:1579–89.
- Wang DY, Salem JE, Cohen JV, et al. Fatal Toxic Effects Associated With Immune Checkpoint Inhibitors: A Systematic Review and Meta-analysis. JAMA Oncol. 2018;4:1721–8.
- Brahmer JR, Lacchetti C, Schneider BJ, et al. Management of Immune-Related Adverse Events in Patients Treated With Immune Checkpoint Inhibitor Therapy: American Society of Clinical Oncology Clinical Practice Guideline. J Clin Oncol. 2018;36:1714–68.
- Thompson JA, Schneider BJ, Brahmer J, et al. Management of Immunotherapy-Related Toxicities, Version 1.2022, NCCN Clinical Practice Guidelines in Oncology. J Natl Compr Canc Netw. 2022;20:387-405.
- Caforio AL, Pankuweit S, Arbustini E, et al. Current state of knowledge on aetiology, diagnosis, management, and therapy of myocarditis: a position statement of the European Society of Cardiology Working Group on Myocardial and Pericardial Diseases. Eur Heart J. 2013;34(2636–48):2648a–2648d.
- Ammirati E, Frigerio M, Adler ED, et al. Management of Acute Myocarditis and Chronic Inflammatory Cardiomyopathy: An Expert Consensus Document. Circ Heart Fail. 2020;13:e007405.
- Ball S, Ghosh RK, Wongsaengsak S, et al. Cardiovascular Toxicities of Immune Checkpoint Inhibitors: JACC Review Topic of the Week. J Am Coll Cardiol. 2019;74:1714–27.
- Bonaca MP, Olenchock BA, Salem JE, et al. Myocarditis in the Setting of Cancer Therapeutics: Proposed Case Definitions for Emerging Clinical Syndromes in Cardio-Oncology. Circulation. 2019;140:80–91.
- Schiffer WB, Deych E, Lenihan DJ, Zhang KW. Coronary and aortic calcification are associated with cardiovascular events on immune checkpoint inhibitor therapy. Int J Cardiol. 2021;322:177–82.
- Baughman KL. Diagnosis of myocarditis: death of Dallas criteria. Circulation. 2006;113:593–5.

Jimenez et al. Cardio-Oncology (2023) 9:14 Page 11 of 11

- 17. Kindermann I, Barth C, Mahfoud F, et al. Update on myocarditis. J Am Coll Cardiol. 2012;59:779–92.
- 18. Tucker NR, Chaffin M, Fleming SJ, et al. Transcriptional and Cellular Diversity of the Human Heart. Circulation. 2020;142:466–82.
- Azzawi M, Hasleton PS, Kan SW, Hillier VF, Quigley A, Hutchinson IV. Distribution of myocardial macrophages in the normal human heart. J Anat. 1997;191(Pt 3):417–23.
- 20. van der Laan AM, Ter Horst EN, Delewi R, et al. Monocyte subset accumulation in the human heart following acute myocardial infarction and the role of the spleen as monocyte reservoir. Eur Heart J. 2014;35:376–85.
- Champion SN, Stone JR. Immune checkpoint inhibitor associated myocarditis occurs in both high-grade and low-grade forms. Mod Pathol. 2020;33:99–108.
- Johnson DB, Balko JM, Compton ML, et al. Fulminant Myocarditis with Combination Immune Checkpoint Blockade. N Engl J Med. 2016;375:1749–55.
- Mahmood SS, Fradley MG, Cohen JV, et al. Myocarditis in Patients Treated With Immune Checkpoint Inhibitors. J Am Coll Cardiol. 2018;71:1755–64.
- Zhang L, Zlotoff DA, Awadalla M, et al. Major Adverse Cardiovascular Events and the Timing and Dose of Corticosteroids in Immune Checkpoint Inhibitor-Associated Myocarditis. Circulation. 2020;141:2031–4.
- Ferreira VM, Schulz-Menger J, Holmvang G, et al. Cardiovascular Magnetic Resonance in Nonischemic Myocardial Inflammation: Expert Recommendations. J Am Coll Cardiol. 2018;72:3158–76.
- Lang RM, Badano LP, Mor-Avi V, et al. Recommendations for cardiac chamber quantification by echocardiography in adults: an update from the American Society of Echocardiography and the European Association of Cardiovascular Imaging. J Am Soc Echocardiogr. 2015;28(1–39): e14.
- Hardy T, Yin M, Chavez JA, et al. Acute fatal myocarditis after a single dose
 of anti-PD-1 immunotherapy, autopsy findings: a case report. Cardiovasc
 Pathol. 2020;46: 107202.
- Palaskas NL, Segura A, Lelenwa L, et al. Immune checkpoint inhibitor myocarditis: elucidating the spectrum of disease through endomyocardial biopsy. Eur J Heart Fail. 2021;23:1725–35.
- Koenig AL, Shchukina I, Amrute J, et al. Single-cell transcriptomics reveals cell-type-specific diversification in human heart failure. Nat Cardiovasc Res. 2022;1:263–80.
- 30. Jimenez J, Lavine KJ. The Dynamic Role of Cardiac Macrophages in Aging and Disease. Curr Cardiol Rep. 2022;24:925–33.
- Kushnareva E, Kushnarev V, Artemyeva A, Mitrofanova L, Moiseeva O. Myocardial PD-L1 Expression in Patients With Ischemic and Non-ischemic Heart Failure. Front Cardiovasc Med. 2021;8: 759972.
- Wei S, Fu J, Chen L, Yu S. Performance of cardiac magnetic resonance imaging for diagnosis of myocarditis compared with endomyocardial biopsy: a meta-analysis. Med Sci Monit. 2017;23:3687–96.
- Thavendiranathan P, Zhang L, Zafar A, et al. Myocardial T1 and T2 mapping by magnetic resonance in patients with immune checkpoint inhibitor-associated myocarditis. J Am Coll Cardiol. 2021;77:1503–16.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- $\bullet\,$ thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

