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Expression of intermediate filaments at muscle insertions in human fetuses

Shinichi Abe^{1,2*}, Sun-ki Rhee², Makoto Osonoi¹, Takuo Nakamura³, Baik Hwan Cho⁴, Gen Murakami⁵, Yoshinobu Ide²

¹Oral Health Science Center hrc7, Tokyo Dental College, Chiba, Japan

²Department of Anatomy, Tokyo Dental College, Chiba, Japan

³Department of Physical Therapy, Sapporo Medical University School of Health

Sciences, Sapporo, Japan

⁴Department of Surgery, Chonbuk National University College of Medicine, Jeonju,

Korea

⁵Division of Internal Medicine, Iwamizawa Kojin-kai Hospital, Iwamizawa, Japan

*Corresponding author.

Dr. Shinichi Abe

Department of Anatomy, Tokyo Dental College

1-2-2 Masago, Mihama-ku, Chiba-shi, Chiba 261-8502, Japan

Tel: +81-43-270-3571, Fax: +81-43-277-4010, E-mail: abesh@tdc.ac.jp

Abstract

Desmin and vimentin are intermediate filaments that play crucial roles in maturation,

maintenance and recovery of muscle fibers and mesenchymal cells. The expression of

these proteins has not been investigated extensively in human fetuses. In the present

study, we examined the immunohistochemical expression of intermediate filaments in

skeletal muscles of the head, neck and thorax in 12 mid-term human fetuses at 9-18

weeks of gestation. We also used immunohistochemistry to localise expression of the

myosin heavy chain, and silver-impregnation to identify the fetal endomysium.

Expression of desmin and vimentin was already detectable in intercostal muscle at 9

weeks, especially at sites of muscle attachment to the perichondrium. At this stage,

myosin heavy chain was expressed throughout the muscle fibers, and the endomysium

had already developed. Beginning with punctate expression, the positive areas became

diffusely distributed in the muscle fibers. At 15-18 weeks, intermediate filament proteins

were extensively expressed in all of the muscles examined. Expression at the

bone-muscle interface was continuous with expression along the intramuscular tendon

fibres. These results suggest that the development of intermediate filaments begins in

areas of mechanical stress due to early muscle contraction. Their initially punctate

distribution, as observed here, probably corresponds to the earliest stage of fetal

enthesis formation.

Key words: desmin, vimentin, intercostal muscle, enthesis, human fetus.

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Introduction

Desmin, a major intermediate filament component, plays a crucial role in maturation, maintenance and recovery of skeletal and cardiac muscle fibers because it forms an interlinking scaffold around myofibrils with connections to the sarcolemma and nuclear membrane (Goldfarb et al., 2008; Tidball, 1992). In normal adult skeletal muscle, desmin is abundant at myotendinous and neuromuscular junctions (Carlsson et al., 1999). Vimentin, another type of intermediate filament, coexists with desmin in human fetal skeletal muscle fibers and plays an important role in maintaining the stability of mesenchymal cells, and also in signal transduction (Yang and Makita, 1996). Vimentin is also known to be a marker of osteoprogenitor cells (Shapiro et al., 1995; Watanabe et al., 1993; Xian et al., 2004) and is expressed in the cell body and processes of osteoblasts and osteocytes (Shapiro et al., 1995).

Although development of these intermediate filaments is probably closely related to muscle fiber maturation. However, their expression has not been fully examined in human fetal muscles according to our knowledge,. In particular, there is no information to indicate whether the expression of desmin and vimentin exhibits heterogeneity at a specific site in a muscle (e.g., the center or periphery of a muscle fiber), a region of the body (e.g., the head, thorax or upper extremity) or a specific developmental stage.

Accordingly, to investigate the expression and distribution of these intermediate filaments in human fetal skeletal muscles, the present immunohistochemical study was conducted. For easy understanding of the intramuscular distribution of positive reactivity, we used relatively large sections showing the topographical anatomy of the muscles examined.

Materials and Methods

The study was performed in accordance with the provisions of the Declaration of Helsinki 1995 (as revised in Edinburgh 2000). We examined the skeletal muscle of the head, neck and thorax in 12 mid-term human fetuses at 9-18 weeks of gestation (three fetuses in each group). With the agreement of the families concerned, these specimens were donated to the Department of Anatomy, Chonbuk National University in Korea. Moreover, their use for research was approved by the university ethics committee, and this study was performed without breaking any rules of the facility to which the lead author belongs. The fetuses had been obtained by induced abortions. After abortion, each of the mothers was personally informed by an obstetrician about the possibility of donating the fetus for research: no attempt was made to encourage donation. The donated fetuses were fixed with 10% w/w formalin solution for more than 3 months. Because of randomization of the specimen numbering, it was not possible to trace any of the families concerned.

Immunohistochemical procedures

In order to analyze the expression of protein in the samples, their immunolocalization of desmin, vimentin and myosin was investigated. The primary antibodies used were: 1) mouse monoclonal anti-human striated muscle myosin heavy chain (dilution, 1:100; Dako, Glostrup, Denmark), 2) mouse monoclonal anti-human vimentin (dilution, 1:10; Dako) and, 3) mouse monoclonal anti-human desmin (dilution, 1:50; Dako). Pretreatment with an autoclave was not conducted because of the fragile nature of the fetal tissues. The secondary antibody (Dako Chem Mate Envison Kit; Dako) was labeled with horseradish

peroxidase (HRP), and antigen-antibody reactions were detected via an HRP-catalyzed reaction with diaminobenzidine. Counterstaining with hematoxylin was performed on the same samples.

In addition, silver impregnation staining (Lillie et al., 1980) was performed to show the endomysium, in which argyrophilic reticular fibers are stained black, and collagen fibers wine red.

Results

Immunohistochemical examination

Expression of desmin

High immunoreactivity for desmin was seen mostly in the head, neck, and thoracic and shoulder girdle muscles at 9-12 weeks. In the intercostal muscles, a site of desmin expression was clearly localized at the rib insertion, with a weak or negative reaction in the central part of the muscles (Fig. 1). The specimens did not show evidence of the diaphragm at 9 weeks. However, at 12 weeks, positive innmunoreaction of desmin was diffusely expressed in the diaphragm. At 12 weeks, desmin immunolocalization was identifiable in the masticatory, tongue, and shoulder girdle and back muscles (Fig. 2). The spots of immunoreactivity were distributed along intramuscular aponeurotic tissues (or intramuscular tendons) as well as at bone or cartilage insertions. The intermediate tendon of the omohyoideus also showed sites of desmin expression. In the tongue, sites of desmin expression appeared near the sagittal septum (Fig. 2A). Among the complex muscles of facial expression, sites of desmin expression were found at the insertion to the base of the nasal cartilages.

At 15 weeks, because the entire length of muscle fibers had become diffusely

positive for desmin, sites of desmin expression became indistinct in some muscles, including the intercostal muscles (Figs. 2G, 3 and 4). Sites of desmin expression then became evident in muscles containing thick intramuscular tendons (e.g., the muscles around the shoulder joint: Fig. 3). Notably, sites of desmin expression were not clearly identified near the sagittal septum of the tongue due to the diffuse nature of the desmin expression, and instead became distinctive around the parasagittal septum (Fig. 4A). At 18 weeks, in most muscles including the intercostals, the punctate expression of desmin changed into diffuse one throughout the entire muscles. In back muscles extending between the vertebral transverse processes at 15-18 weeks, the appearance of sites of desmin expression was delayed, even though the muscle configuration was similar to that of the intercostals. Moreover, in some muscles such as the serratus anterior and multifidus, desmin localization was evident not only at the muscle insertion but also in the central part of the muscle fibers.

Expression of vimentin

Sites of desmin expression, localization within a muscle, and the chronological sequence of expression were similar to those observed for desmin immunoreactivity. At 9-12 weeks, sites of vimentin expression were evident at the rib insertions of intercostal muscles (Fig. 1B), but the expression became diffusely positive at 15 weeks in these muscles. Notably, at 15 weeks, sites of vimentin expression were extensively distributed along intramuscular tendons and at the muscle-tendon interface (Figs. 2B,D, F and 3C), being apparently more frequent than desmin expression. At 18 weeks, vimentin was diffusely expressed throughout the muscle fibers. In addition, osteoprogenitor cells strongly expressed vimentin along the margins of ossification sites. The dura mater and membranous

inner ear were also positive for vimentin.

Expression of the myosin heavy chain

The myosin heavy chain is abundantly present in fetal skeletal muscle.

Accordingly, all samples showed diffuse expression in the muscle fibers at 9 weeks (Fig. 1C), and we were unable to determine the developmental stage at which myosin expression was initiated.

Silver staining

The spots were located diffusely within the end of the muscle fibers according to silver staining and the nuclei were likely to overlap with the reactive zone (Fig. 1E). The endomysium was stained black by silver impregnation (Figs. 1E and 3D) and collagen bundles in the perichondrium and tendon were stained wine red. In the intercostal muscles, the endomysium had already developed by 9 weeks. Likewise, the endomysium was evident when desmin displayed spotty expression at the sites of muscle fiber attachment.

Discussion

The present study appears to be the first to have demonstrated the chronological sequence and heterogeneous distribution of desmin expression in human fetuses. Desmin expression was apparently positive in the intercostal muscles, and thereafter seemed to spread to other muscles. The early appearance of desmin expression in the intercostal muscles was probably attributable to the short muscle fibers there, as tension is conducted easily along the entire length of such short fibers in comparison with long ones. Even in comparison with small muscles of adults, such as the tensor tympani and subclavius, the muscle fibers in fetuses were seemed to be

larger than in intercostal muscles. Although the intercostal and intervertebral muscles are morphologically similar, the difference in the developmental stage at which desmin expression first appeared in them might be explained by a difference in the stage at which they begin contraction.

Desmin expression in muscle attachments to the pericondrium (or at future sites of bone insertion) was followed by expression along the intramuscular tendons. In lingual muscles, desmin expression was initiated from the midsagittal septum, and then the site of strongest expression moved to the parasagittal septum. The expression of vimentin also demonstrated a similar chronological sequence. The results from week 15 would suggest that the expression occurs in muscle cells (Hesselmans et al., 1993). Such heterogeneity in the site and stage of expression suggested progressive maturation of the muscle fiber attachments. Expression of the myosin heavy chain (Abe et al., 2007; Gojo et al., 2002; Maejima et al., 2005; Okubo et al., 2006; Shida et al., 2005; Suzuki et al., 2007; Usami et al., 2003; Yanagisawa et al., 2006; Yoshii et al., 2008) appeared to have been completed before that of desmin was initiated. Likewise, silver impregnation showed that the endomysium of each muscle fiber was apparently in tight contact with the perichondrium, insertion tendon or intramuscular tendon until the expression of desmin appeared. Therefore, it was suggested that desmin expression coincided with the initiation of mechanical stress between the endomysium and mature muscle fiber accompanying early contraction.

Vimentin, another intermediate filament, is a well known marker of osteoprogenitor cells (Shapiro et al., 1995; Watanabe et al., 1993; Xian et al., 2004) and plays an important role in maintaining the stability of mesenchymal cells (Yang and Makita, 1996). Thus, the concomitant expression of both vimentin and desmin observed in this study

appeared to suggest maturation of the developing muscle-tendon and muscle-bone interfaces. During the development of rabbit fetal limb muscles, insertion regions are characterized by the presence of coarse-fibered periosteal bone and chondroid bone, both being morphologically intermediate between fibrocartilage and lamellar bone (Hurov, 1986). Fibrocartilage provides the bone-tendon interface, or enthesis: the fibrocartilage cells are commonly packed with intermediate filaments, which could be involved in the transduction of mechanical load (Benjamin et al., 1998). Although previous studies did not include any data for early stages of development, the punctate expression of desmin and vimentin observed in the present study probably corresponded to the morphology present at the earliest stage of enthesis development.

The present findings were consistent with those of a previous study of human fetal cardiac muscles, in which desmin expression increased progressively with fetal age (Kim, 1996). Notably, that study also demonstrated that, in contrast to desmin, vimentin-positive areas diminished progressively in cardiac muscle, despite the fact that vimentin is present in not only muscle cells but also connective tissue cells. A similar dynamic relationship between desmin and vimentin has been reported in human skeletal muscle fibers (Yang and Makita, 1996; Barbet et al., 1991). In the present study, vimentin immunoreactivity extended diffusely throughout muscle fibers by 18 weeks. However, this observation did not indicate a decrease of expression. Unfortunately, we were unable to obtain any data for stages later than 18 weeks because of the limited materials available.

It is well known that the periosteum plays an essential role in tendon and ligament migration during the growth of long bones (Muhl and Gedak, 1986). On the other hand, osteoblasts are likely to control the activity of osteoclasts, as well as the distribution of

differentiated osteoclasts, via the RANK-RANKL molecular system (Takahashi et al., 1999; Yamamoto et al., 2006). Further studies may reveal a type of osteoblast differentiation in the periosteum that is regulated by a molecular cascade during muscle fiber development, maturation and degeneration.

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Figure legends

Figure 1. Immunohistochemistry of fuman fetal intercostal muscles at 9 weeks.

A, B) Immunoreactivity for desmin and vimentin is restricted to the muscle insertions. C) Whereas, MyH is distributed diffusely throughout the muscles. D) Higher-magnification view of the square in figure A. E) Silver impregnation demonstrates the endomysium (stained black) connecting to the perichondrium of the rib (stained wine red). EI, external intercostal muscle; II, internal intercostal muscle; ICN, intercostal nerve; P, pectoralis major (caudal end); PL, pleural cavity; rib 1 and rib 2, first and second ribs.

Figure 2. Condensations of desmin and vimentin at 12-week human fetus.

A, C, E and G show desmin immunohistochemistry, and B, D and F show vimentin expression. A, B) In the tongue, condensations (spots) of desmin or vimentin expression

are evident near the sagittal septum (asterisk). C, D) In the biceps brachii muscle, the spots are arranged in line along the distal tendon. E-G) In the trapezius muscle, the spots are located along the insertion into the clavicle (E and F). G) The trapezius muscle fibers exhibit desmin spots in spite of their diffuse desmin expression at 15 weeks.

Figure 3. Subscapularis muscle in a human fetus at 15 weeks.

A and B show immunohistochemistry for desmin (B, higher magnification), C shows that for vimentin in an adjacent section, and D demonstrates silver impregnation of the section adjacent to that in figure A. Orientations of B-D are shown as squares and circles in A. The desmin or vimentin spots (B and C) are evident in and near the intramuscular tendon (asterisk). The black-stained endomysium connects to the wine red-stained intramuscular tendon (D). SA, serratus anterior.

Figure 4. Sites of desmin expression in muscles of a human fetus at 15 weeks.

A specimen differing from that shown in Fig. 3. A) Sites of desmin expression are not evident along the sagittal septum of the tongue (asterisk), but around the parasagittal septum (star). GH: geniohyoideus. B) The tendon of the tensor tympani with Sites of desmin expression. C) The lateral pterygoid muscle inserts to the mandibular head. D) The laryngeal muscle inserting to the posterolateral part of the arytenoid cartilage. E) The origin of the supraspinatus muscle from the scapular plate. All figures were prepared at the same magnification.

Fig.1

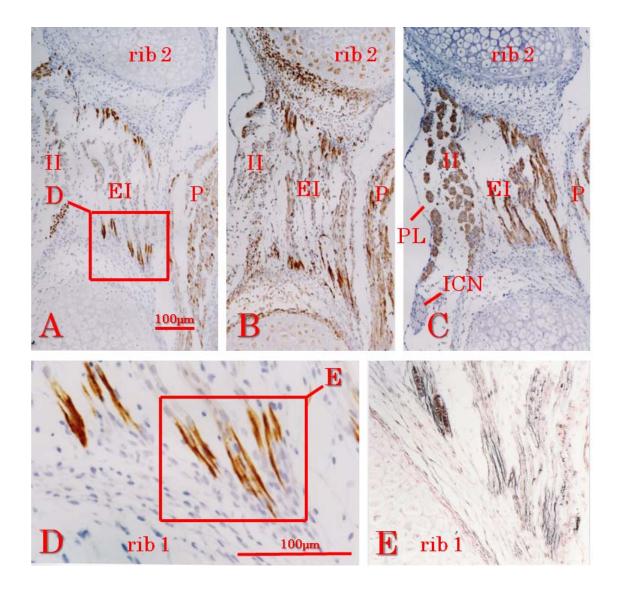


Fig.2

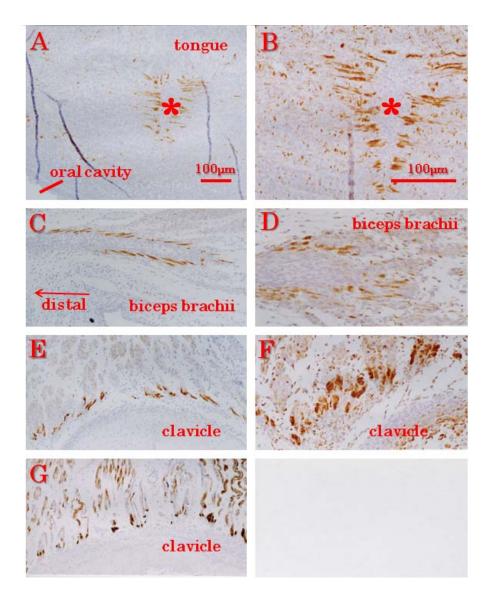


Fig.3

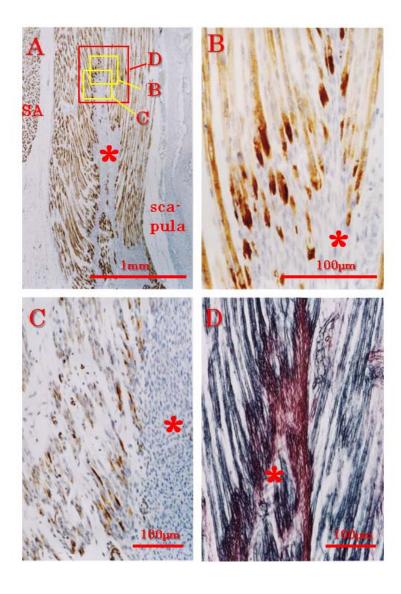


Fig.4

