




# Triggers of Anaphylaxis in Mastocytosis Patients: Evidence of the Current Drug-Avoidance Recommendation

Tiago Azenha Rama, DMD, MD<sup>1,2,3,4\*</sup>,   
Mariana Castells, MD, PhD<sup>5</sup>

## Address

<sup>1</sup>Serviço de Imunoalergologia, Centro Hospitalar Universitário São João, Alameda Prof. Hernâni Monteiro, 4200-319 Porto, Portugal

<sup>2</sup>Serviço de Imunologia Básica e Clínica, Departamento de Patologia, Faculdade de Medicina, Universidade do Porto, Porto, Portugal

<sup>3</sup>EPIUnit - Institute of Public Health, University of Porto, Porto, Portugal

<sup>\*,4</sup>Laboratory for Integrative and Translational Research in Population Health (ITR), Porto, Portugal

Email: tarama@med.up.pt

<sup>5</sup>Division of Allergy and Clinical Immunology, Mastocytosis Center, Drug hypersensitivity and Desensitization Center, Brigham and Women's Hospital, Harvard Medical School, Boston, USA

© The Author(s) 2023

**Keywords** Mastocytosis · Mast cell activation · Anaphylaxis · Drug hypersensitivity

## Opinion statement

Mastocytosis is associated with a high risk of anaphylaxis, in part due to drug hypersensitivity reactions (DHR). Drugs associated with mast cell activation include nonsteroidal anti-inflammatory drugs (NSAIDs), drugs used in the perioperative setting, including general anesthetics, neuromuscular blocking agents (NMBAs) and opiates/opioids, radio-contrast media (RCM), vaccines, and antibiotics such as quinolones and vancomycin. To protect patients against DHR and anaphylaxis, general avoidance of potential drug triggers is common practice, which often deprives patients of important drugs at times of need and limits their options. We aimed to critically review current evidence on the indications to avoid drugs in children and adults with mastocytosis. Recent data shows that general avoidance of drugs with potential mast cell activation action is not indicated in all patients with mastocytosis, but guidelines are lacking. Drugs tolerated before and after the onset of mastocytosis should not be avoided and a personalized approach is recommended to

address drugs inducing mast cell activation. Pre-medication (RCM, local and general anesthetics, vaccines), use of safer alternatives (opioids, NBMA, NSAIDs in selected cases), and drug challenges (NSAIDs in most cases) are recommended to increase the safety of patients with mastocytosis when introduced to new drugs.

## Introduction

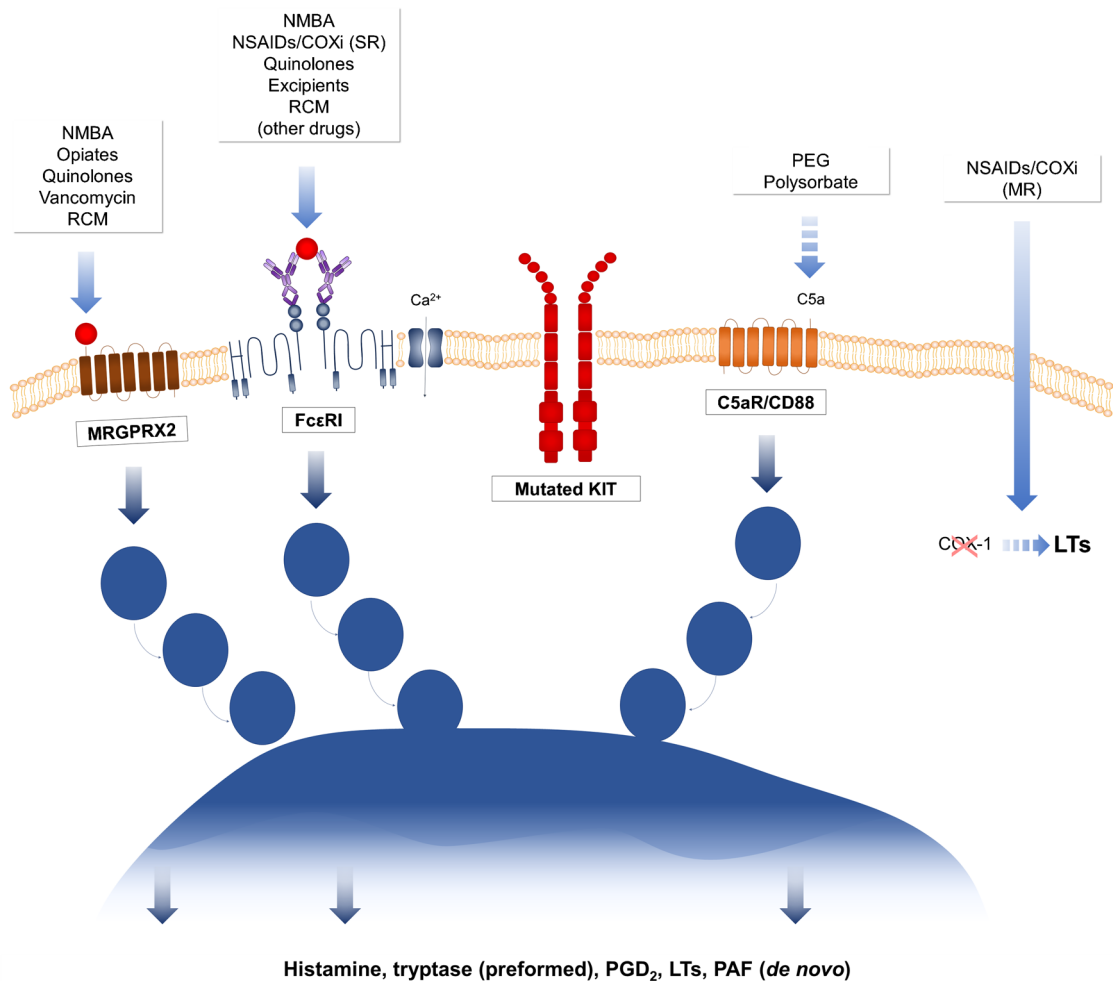
The term mastocytosis comprises a heterogeneous group of diseases characterized by clonal and phenotypically aberrant mast cell (MC) accumulation in different tissues and organs, including the skin, bone marrow, and gastrointestinal (GI) tract, with or without excessive MC activation [1].

MC express a myriad of activating receptors, including the stem cell factor receptor (KIT), immunoglobulin receptors (e.g., FcεRI and FcγRI), complement receptors (e.g., C3aR and C5aR/CD88), pattern recognition receptors (e.g., Toll-like receptors [TLR]) and the Mas-related G-protein coupled receptor member X2 (MRGPRX2) [2, 3]. When activated, MC may release a full array (i.e., degranulation) or a limited amount of mediators (i.e., piecemeal-degranulation or differential release of mediators) [4]. Immunoglobulin (Ig)E-independent (and notably MRGPRX2-dependent) degranulation has different kinetics and extent of mediators release as compared to IgE-mediated activation [5].

The majority of mastocytosis are caused by activating mutations in the gene that codifies for the KIT-receptor/CD117 (i.e., *KIT*), with the *KIT*<sup>D816V</sup> mutation being detected in over 90% of cases [6]. Such mutations are mostly sporadic and are restricted to MC and MC precursors in around two-thirds of cases [7]. Activating *KIT* mutations promote autophosphorylation and, or dimerization of the KIT receptor in a ligand-independent fashion, enhancing cell proliferation and survival [6], as well as IgE-mediated activation [8] and may lower the threshold for non-IgE-mediated MC activation [9]. Mastocytosis patients present mild to severe MC activation symptoms, which can be chronic and, or acute, including diarrhea, heartburn, urticaria, angioedema, flushing, hypotension, dizziness, brain fog, headaches, and life-threatening anaphylaxis [10].

The prevalence of anaphylaxis in mastocytosis patients is considerably higher than in the general population and may reach up to 50% [10••, 11], even though the prevalence of atopy is similar [12, 13]. According to the results found on two large case series, the percentage of allergy among mastocytosis patients ranges between 24 [14] and 28% [12, 13] in adults, and 17 [14] and 11% [15] in children. Interestingly, 67% of patients with anaphylaxis seem to be sensitized to known allergens [16]. Anaphylaxis is common among patients with systemic mastocytosis without skin lesions (typically adult males), in whom it may be present in up to 67% [12, 13]. In contrast, the frequency of anaphylaxis in mastocytosis with skin lesions (i.e., including cutaneous mastocytosis) is lower and found to be 16–19% [14, 16]. Mastocytosis patients at higher risk for anaphylaxis include those presenting without skin lesions, atopy, high total IgE, and lower serum baseline tryptase (sBT) [17], while recent data suggests a potential contribution of hereditary alpha-tryptasemia (HaT) genotypes [18–22].

In mastocytosis, MC activation symptoms can be unprovoked and provoked by triggers [12]. Non-specific triggers include emotional stress/anxiety, dental eruption, fever, and the friction of mastocytosis skin lesions [23–25]. Specific triggers include drugs such as nonsteroidal anti-inflammatory drugs (NSAIDs), vaccines, opiates/opioids, neuromuscular blocking agents (NMBAs), radiocontrast media (RCM), quinolones, vancomycin, and *Hymenoptera* venoms [25, 26], which may induce MC activation through several mechanisms (Fig. 1). Due to the unpredictability of drug hypersensitivity reactions (DHR) and widespread perception of the risks associated with these drugs, mastocytosis patients are often told to empirically avoid all of these drugs. Avoidance of such drugs is



**Fig. 1** Mast cell receptors potentially involved in drug hypersensitivity reactions in mastocytosis patients. Activating *KIT* mutations lead to dimerization of *KIT* which lowers the threshold for mast cell (MC) activation due to the activation of the high-affinity immunoglobulin (Ig) E receptor (FcεRI) and the mas-related G-protein-coupled receptor X2 (MRGPRX2). Both receptors lead to MC degranulation when activated. IgE-mediated activation leads to a slow-onset and prolonged release of MC mediators dependent on the calcium influx, while MRGPRX2 induces a rapid-onset and brief (even if often severe) release. The C5a receptor (C5aR/CD88) may be overexpressed in patients with mastocytosis and also leads to MC activation due to the so-called complement activation-related pseudoallergy, in which IgM/IgG antibodies against drugs (e.g., excipients found in COVID-19 polyethyleneglycol [PEG] and polysorbate) activate the classical complement pathway culminating in the release of C5a. Following activation, MC granules merge into larger ones and migrate to the membrane with which they merge, releasing their content, which includes preformed mediators (i.e., histamine, tryptase). Later, de novo release of prostaglandin (PG) D<sub>2</sub>, cysteinyl-leukotrienes (LTs), and platelet-activating factor (PAF) occurs. Cyclooxygenase (COX) 1 inhibitors (e.g., nonsteroidal anti-inflammatory drugs and paracetamol) induce the release of leukotrienes (LTs) by decreasing the release of PGE<sub>2</sub> thereby promoting the action of leukotriene synthase.

frequently unnecessary as the associated risk of MC activation is often low and similar to the risk in the general population [10, 27, 28••, 29].

Here, we aimed to critically review current evidence on the indications to avoid drugs with mast cell activation potential in children and adults with mastocytosis.

## Nonsteroidal anti-inflammatory drugs and other cyclooxygenase inhibitors

NSAIDs are part of a broad group of cyclooxygenases (COX) inhibitors (COXi) that also includes analgesics and antipyretics such as salicylates (e.g., acetylsalicylic acid [ASA]), pyrazolones and paracetamol [30]. NSAIDs and ASA exert preferential COX-1 inhibition at the peripheral and central nervous systems, enacting anti-inflammatory, antipyretic, and pain control effects [31]. In contrast, paracetamol is a weak COX-1 inhibitor with analgesic and antipyretic effects caused by central nervous system COX-2 inhibition [32].

DHR to NSAIDs or other COXi may be caused by drugs within structurally related groups, with patients reacting to a single drug or structurally related drugs [33] through an IgE-mediated mechanism or by multiple structurally unrelated drugs [10••, 34] through a COX-1-inhibition-mediated depletion of PGE<sub>2</sub> mechanism [35], resulting in an over-production of cysteinyl-leukotrienes [36]. Reactions can range from urticaria, angioedema, and bronchospasm to anaphylaxis [37]. Patients reacting to multiple COX-1 inhibitors typically can tolerate weak COX-1 inhibitors (e.g., paracetamol) and COX-2 preferential or selective inhibitors (i.e., nimesulide/meloxicam or coxibs) [38]. Because there are no biomarkers of NSAID hypersensitivity, drug challenges with ASA (the strongest COX-1 inhibitor) are used to exclude/confirm reactivity to multiple NSAIDs [39, 40, 41]. Still, children under 16 must avoid ASA due to potential Reye's syndrome [42].

In mastocytosis, the reported prevalence of NSAID hypersensitivity is 14% in adults and 9% in pediatric patients [10••, 43, 44•], and NSAID-induced anaphylaxis is found in 2 to 11% of adult patients [10••, 14–16, 45] and 2% of children and adolescents [10••], and has led to frequent general avoidance [10••]. Regional differences exist, with ASA being the most frequent culprit in Spain and Italy [10••, 44•] and diclofenac in Sweden [44•]. Paracetamol, coxibs, and meloxicam are tolerated by most patients [10••].

A large single-center study conducted by the Spanish Network on Mastocytosis (*Red Española de Mastocitosis* [REMA]; with 469 patients, 382 adults and 87 children/adolescents) indicated that risk factors for NSAIDs hypersensitivity included female sex, with baseline symptoms of angioedema and pruritus, a previous history of anaphylaxis other than HVA, advanced disease, higher sBT levels, and multilineage *KIT*<sup>D816V</sup> mutation [10••]. A risk-predicting scoring model (Table 1) showed high sensitivity and requires validation in large multicenter studies.

Patients who tolerate NSAIDs before and after the onset of mastocytosis do not require testing, and avoidance is not indicated [10••, 44•, 46, 47]. For patients whose tolerance following the onset of the disease is unknown, a challenge with ibuprofen is recommended in children and adults with a negative score and with preferential/selective COX-2 inhibitors (e.g., meloxicam or coxibs) for adults with a positive score [10••]. Drug challenges need to be done by experienced providers after risk/benefit information to the patient in a shared decision process.

**Table 1. Scoring model proposed by the *Red Española de Mastocitosis* to screen for hypersensitivity to multiple NSAIDs or other COXi in adult patients with mastocytosis [10\*\*]**

Variable	Score
Gender	
Female	+6
Male	-2
Manifestations	
Flushing	+4
Absence of pruritus	-3
Skin lesions	-3
Anaphylaxis not caused by NSAIDs/COXi	+5
Anaphylaxis caused by <i>Hymenoptera</i> venoms	-4
Laboratory findings	
Multilineal D816V <i>KIT</i> mutation*	+4
sBT $\geq$ 48 ng/mL	+6

Patients scoring seven or above should undergo drug challenges with COX-2 preferential or selective inhibitors (respectively, meloxicam and coxibs), whereas those scoring below seven may undergo challenges with ibuprofen. \*If unknown, consider as negative

## Vaccines

Vaccines contain adjuvants which modulate both innate and adaptive immune responses [48] and excipients to stabilize and preserve formulations (e.g., ethylenediaminetetraacetic acid [EDTA]) and as antigen carriers (e.g., polyethylene glycol [PEG] nanolipid envelope). Microbial antigens, adjuvants, and excipients may act as allergens or as pathogen-associated molecular patterns (PAMP), leading to IgE- and non-IgE-mediated activation of MC. IgE-mediated reactions have been described against excipients (e.g., gelatin, milk and egg proteins, latex, tromethamine) and microbial antigens (e.g., protein conjugates within the diphtheria and *Haemophilus influenzae* type b vaccines, and tetanus toxoid) [49–51]. SARS-Cov-2 mRNA vaccines' PEGylated nanolipid envelopes have been implicated in IgE-mediated anaphylactic reactions, which have been more frequent in females with previous allergic reactions, including anaphylaxis to drugs and foods [52–55]. Non-IgE-mediated mechanisms, such as dose-dependent complement-mediated MC activation (complement activation-related pseudoallergy [CARPA]) [52, 56–58], have been suggested as a potential cause of DHR to COVID-19 vaccines in mastocytosis patients, as MC from patients with systemic mastocytosis overexpress the complement receptor for C5 (C5aR/CD88) [59].

Anaphylaxis to vaccines is rare in the general population, amounting to an overall rate of 1.31 per million vaccine doses [60], and is increased ten-fold for SARS-COV-2 vaccines [49]. In mastocytosis, MC activation episodes related to vaccination have been reported in children and in adults [61–63]. The frequency of reactions ranges from 4 to 13% in pediatric mastocytosis

patients [61, 64, 65], with diffuse cutaneous mastocytosis (DCM) having been deemed a risk factor in a Spanish study population [63]. Although there is no correlation between lesion burden and vaccination-associated DHR, the administration of single vaccinations instead of multiple simultaneous vaccinations with pre-medications has been recommended [61], especially in patients with DCM [63, 66, 67].

Five studies with a total of 372 patients have described the safety of COVID-19 vaccination in patients with mastocytosis and other clonal and non-clonal MC activation disorders [27, 68–71]. In two studies, none out of 73 [68] and nine out of 30 [27] had a prior history of anaphylaxis. Overall, 12 hypersensitivity reactions were reported including anaphylaxis in non-pre-medicated (10%) and pre-medicated (0.6%) patients.

The data support the safe vaccination of all pediatric and adult mastocytosis patients as recommended by worldwide agencies. Pre-medication is recommended with H1 and H2 antihistamines and leukotriene receptor antagonists, but not steroids, and vaccination should be followed by a prolonged observation period of 30 min.

## Drugs used in the perioperative setting and opiates/opioids

There is a perception by patients and providers that general anesthesia is a high-risk procedure for mastocytosis patients [46, 47, 72–75]. However, in a large retrospective study of patients with mastocytosis ( $n = 501$  patients, 459 adults, and 42 children), the risk of DHR during the perioperative period was shown to be low (DHR in 2% of adults and 4% of children, and anaphylaxis in 0.4% of adults and 2% of children) [75]. Risk factors identified in the study included a prior history of anaphylaxis, major surgery, and lack of pre-medication [75]. A limited study of a pediatric population ( $n = 22$  patients) showed that 9% of children who underwent general anesthesia presented with flushing without hemodynamic compromise [76].

Neuromuscular blocking agents are the most frequent elicitors of perioperative anaphylaxis in the general population [77, 78], which may be caused by IgE-mediated and MRGPRX2-related MC activation (for drugs containing the tetrahydroisoquinoline [THIQ] motif) [79, 80]. Non-depolarizing benzylisoquinoline NMBAs atracurium and mivacurium have been identified as having the highest potential for MC activation [79, 81] due to MRGPRX2-related activation [82]. Atracurium cause severe reactions in mastocytosis patients and its use is not recommended due to potential fatalities [73]. Succinylcholine and isoquinoline-type NMBAs, such as cisatracurium, have the lowest potential for MC activation [79, 83, 84] and are considered safe for mastocytosis patients. Succinylcholine is rarely associated with IgE-mediated reactions [26, 85]. Aminosteroid NMBAs include rocuronium which can induce IgE-mediated reactions and is not recommended in mastocytosis patients, while vecuronium, pancuronium, and rapacuronium rarely induce MC activation [79], and their use is recommended in mastocytosis patients [26, 73].

Opiates/opioids, including morphine, pethidine/meperidine, codeine, and others, are avoided by mastocytosis patients based on in vivo and in vitro

data showing direct MC activation [86, 87]. Codeine, an opiate frequently used in over-the-counter cough syrups, seems to display the strongest MC degranulation potential, followed by morphine and meperidine [88]. These effects are dose-dependent [89, 90] and thought to relate to the activation of MC  $\delta$ -receptors [89], the release of histamine by free radicals [91], and MRGPRX2-mediated activation [92]. In vitro studies show that connective tissue mast cells termed MC<sub>TC</sub> undergo degranulation following morphine exposure [93], whereas pulmonary, intestinal, and cardiac mucosal mast cells (i.e., MC<sub>T</sub>) do not degranulate [94]. Buprenorphine seems to only induce degranulation of pulmonary MCs [90]. Fentanyl and other piperidine-derived synthetic opioids have been shown to be safe for mastocytosis patients, as these induce little MC activation [88, 89]. Opioids used in analgesia, including oxycodone, hydromorphone, oxymorphone, tapentadol, and tramadol (a partial opioid-receptor agonist), have been shown not to induce in vivo histamine release [92]. Even though in vivo responsiveness to morphine in mastocytosis patients (assessed by wheal size following skin prick testing) is not increased in comparison with healthy individuals and asthma patients [95], its use is not recommended, and the use of tramadol, and piperidine derivatives has been recommended instead [26, 96].

IgE-mediated DHR to hypnotics, propofol, thiopental, and ketamine are less common than reactions to NMBAs, and opiates/opioids and are not contraindicated in mastocytosis patients [97]. Propofol, thiopental, and ketamine induce the release of histamine by cutaneous and pulmonary MCs [98], whereas propofol may inhibit intestinal and cardiac MC activation [99–102]. Benzodiazepine (BZPs) IgE-mediated DHR are exceedingly rare in the general population [103], and MCs have been shown to have high affinity binding sites for BZPs [104, 105], which may inhibit MC proliferation and activation [106]. BZPs are considered safe in mastocytosis, and their use as pre-medication is recommended to reduce procedural anxiety [107]. Currently used inhaled general anesthetics seem to be devoid of MC activation potential, IgE-mediated reactions have not been described, and their use should not be avoided in mastocytosis patients [108, 109].

The prevalence of anaphylaxis in the perioperative period is higher for mastocytosis (1:250 in adults and 2:100 in children) [75] as compared to the general population (1:5000 to 1:13000) [110]. Specific and non-specific triggers include the drugs described above, procedural anxiety, temperature changes, pressure and lesional friction, and gastrointestinal organ manipulations. The role of pre-medications has not been validated in controlled studies, and the current recommendations include glucocorticoids, H1 and H2 antihistamines, leukotriene receptor antagonists and BZP [26]. Omalizumab has been shown to protect mastocytosis patients against anaphylaxis [111], and its protective effect in the perioperative period also warrants further studies.

Local anesthetics (LAs) are divided into ester-type LAs (e.g., benzocaine, procaine), which are associated with immediate DHR [112], and amide LAs which rarely cause immediate DHR or anaphylaxis [113, 114]. Preservatives and antioxidants contained in LAs formulations, or LAs/vasoconstrictor association formulations, including parabens, metabisulfite, and latex [112, 115–117], have rarely been identified as reaction culprits. Skin tests' predictive values are not well defined since false positive tests followed by negative

subcutaneous challenges have been reported [118]. Non-immune-mediated reactions such as syncope resulting from parasympathetic hyperstimulation or hyperexcitability of peripheral nerves have been described [119]. The prevalence of DHR to LAs has been described as 0.8% in mastocytosis adults [75], and ester-type LAs are not recommended [26, 46, 96]. Pre-medication may be recommended prior to procedures requiring local anesthesia—usually, a non-sedative H1 antihistamine 1 h before the procedure.

## Radiocontrast media

RCM are an uncommon cause of DHR and anaphylaxis in the general population [120, 121] and in mastocytosis patients [14, 16, 46, 122]. In the general population, hyperosmolar/ionic RCM have been associated with a higher frequency of severe DHR [120]. The prevalence of DHR to non-ionic/hyposmolar RCM is much lower at 0.1% in a large cohort, including 299,413 patients who underwent contrast tomography [121]. Still, RCM osmolarity does not seem to correlate with histamine release [123]. In vitro studies indicate that hyposmolar non-ionic RCM (e.g., iohexol) do not induce direct MC activation [124]. Reactions to RCM are thought to be IgE- [125] or MRGPRX2-mediated [126, 127], and the role of excipients, notably tromethamine, has not been defined.

Literature reports on DHR associated with hyperosmolar or hyposmolar RCM in mastocytosis patients are scarce, and, based on in vitro data indicating MC activation with hyperosmolar RCM [128], the use of hyperosmolar/ionic RCM has been contraindicated [129]. DHR reports in mastocytosis patients include iohexol [130], non-disclosed angiography contrast media [131], non-disclosed non-ionic ICM [122], and otherwise unspecified contrast media [15, 122, 132–134]. Some of the reports indicate that patients were able to tolerate RCM with pre-medication [122, 130]. Pre-medications prior to RCM are recommended, but no controlled studies exist to provide outcomes [26, 46, 47, 122, 134]. Evaluation of mastocytosis patients with suspected DHR to RCM may include skin testing and drug challenge testing [46]. In the unusual event of DHR to multiple RCM, tromethamine hypersensitivity should be investigated. Paramagnetic contrasts (e.g., gadolinium derivatives) may be used as alternatives to RCM, but gadobutrol should be avoided for those allergic to tromethamine [50].

## Antibacterials

Quinolones (e.g., ciprofloxacin, levofloxacin, moxifloxacin) are broad-spectrum antibiotics involved in immediate DHR both through IgE-dependent and MRGPRX2 activation [135]. Quinolones' safety [136] and resistance [137] profiles have made their use mostly limited to multiresistant bacteria (e.g., *Pseudomonas aeruginosa*). The prevalence of immediate DHR to



fluoroquinolones is as high as 1 out of 1100 treatments [138]. In mastocytosis, only one case of DHR has been reported to date [139]—curiously, in a patient with bone marrow mastocytosis with Hymenoptera venom anaphylaxis, a disease subvariant that seems less prone to DHR [29]. Recommendations to avoid quinolones in patients with mastocytosis have been based on the high rate and severity of the reactions [140].

Vancomycin is a glycopeptide used to treat bacterial gram<sup>+</sup> infections (e.g., methicillin-resistant *Staphylococcus aureus*, *Enterococci*) and a weak MRGPRX2 agonist [141]. Vancomycin infusions can induce histamine release in healthy volunteers related to the infusion rate (formerly, red man syndrome) [142]. There is no evidence that slow-rate infusions may be associated with MC activation in patients with mastocytosis, but vancomycin should be avoided in these patients based on its MC activation properties.

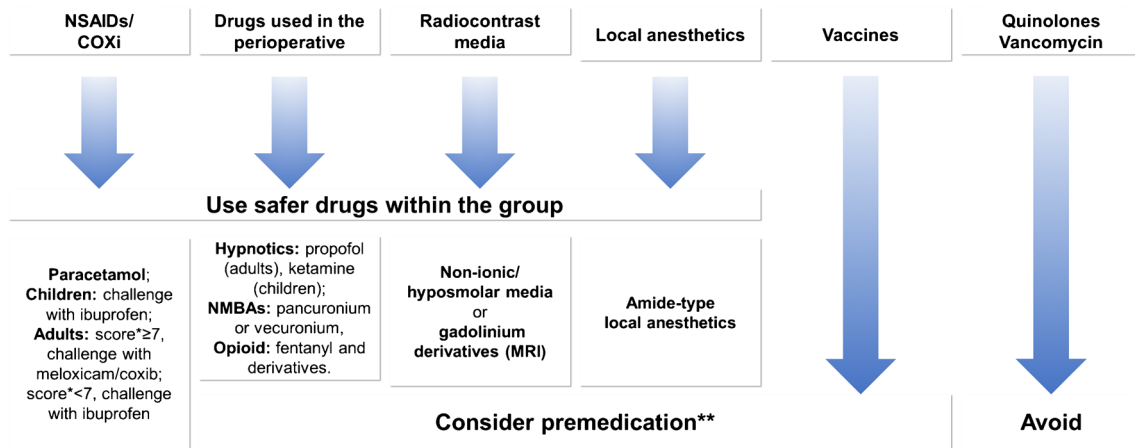
## Conclusion

Mastocytosis has been associated with an increased risk of DHR to a wide array of drugs, including NSAIDs, drugs used in the perioperative setting, opiates and opioids, vaccines, RCM, antibiotics such as quinolones, and vancomycin and identifying patients at risk is paramount to prevent unnecessary avoidance of critical drugs. The proportion of patients with DHR depends on the drug and the disease with adult patients presenting higher incidence than children (Table 2). Avoidance should be restricted to specific drugs to which there is a history of reactions. Drugs with the best safety

**Table 2. Frequencies for drug hypersensitivity reactions of drugs frequently avoided in mastocytosis patients**

Drug group	Frequency of hypersensitivity in mastocytosis	References
NSAIDs/COXi	Children: up to 9% Adults: up to 14%	[10 <sup>••</sup> , 43, 44 <sup>•</sup> ]
RCM	Children: unknown (rare) Adults: rare	[122]
Drugs used in the perioperative		
General anesthesia	Children: 2–18% Adults: 0.4%	[75, 76]
Local anesthesia	Children: unknown (rare) Adults: 0.8%	[75]
Vaccines	Children: 4–13% Adults: 3% (COVID-19 vaccines)	[61, 65] [27, 68, 69, 70, 71]
Quinolones	Unknown	-
Vancomycin	Unknown	-

COXi cyclooxygenase inhibitors, NSAIDs nonsteroidal anti-inflammatory drugs, RCM radiocontrast media



**Fig. 2** Drugs that are frequently avoided in patients with mastocytosis and suggested approach. COXi, cyclooxygenase inhibitors; MRI, magnetic resonance imaging; NMBA, neuromuscular blocking agents; NSAIDs, nonsteroidal anti-inflammatory drugs; RCM, radiocontrast media. \* see Table 1. Asterisk (\*\*) indicates prednisolone 0.5 to 1 mg/Kg (or equivalent) 12 and 1 h before, and cetirizine 10 mg (or other non-sedating H1 antihistamine on equivalent dose), famotidine 20 mg and montelukast 10 mg 1 h before major surgery and procedures in patients with previous reactions/same dosing of glucocorticoids 13, 7 and 1 h before RCM infusion. Cetirizine 10 mg (or other non-sedating H1 antihistamine on equivalent dose) and famotidine 20 mg for vaccination and minor procedures. Omalizumab may be used in patients with multiple reactions to multiple drugs who have failed other pre-medications.

profile should be preferred over those known to elicit MC activation and drug challenges for less preferred drugs should be done at times of need by experienced providers. Patients should not be labeled as allergic to drugs to which they have not been exposed before. Delabeling patients with allergy labels not sustained by objective data should be done to improve the quality of life of mastocytosis patients. Pre-medications should be used judiciously before tests and procedures known to induce reactions (Fig. 2). Pre-emptive and empirical avoidance of drugs is not recommended in children with mastocytosis who have not presented previous reactions or have not been exposed to the drug. Further clinical and basic research and the generation of data bases will help understand DHR in mastocytosis and guide treatments and procedures, improving safety.

## Funding

Open access funding provided by FCT|FCCN (b-on).

## Compliance with Ethical Standards

---

### Conflict of Interest

The authors declare no competing interests.

### Human and Animal Rights and Informed Consent

This article does not contain any studies with human or animal subjects performed by any of the authors.

### Open Access

This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

## References and Recommended Reading

---

Papers of particular interest, published recently, have been highlighted as:

- Of importance
- Of major importance

1. Valent P, Akin C, Hartmann K, Alvarez-Twose I, Brockow K, Hermine O, et al. Updated diagnostic criteria and classification of mast cell disorders: a consensus proposal. *Hemasphere*. 2021;5(11):e646.
2. Phong BL, D'Souza SJ, Baudier RL, Wu E, Immethun VE, Bauer DL, et al. IgE-activated mast cells enhance TLR4-mediated antigen-specific CD4+ T cell responses. *Sci Rep*. 2021;11(1):9686.
3. Tatemoto K, Nozaki Y, Tsuda R, Konno S, Tomura K, Furuno M, et al. Immunoglobulin E-independent activation of mast cell is mediated by Mrg receptors. *Biochem Biophys Res Commun*. 2006;349(4):1322–8.
4. Galli SJ, Gaudenzio N, Tsai M. Mast cells in inflammation and disease: recent progress and ongoing concerns. *Annu Rev Immunol*. 2020;38(1):49–77.
5. Gaudenzio N, Sibilano R, Marichal T, Starkl P, Reber LL, Cenac N, et al. Different activation signals induce distinct mast cell degranulation strategies. *J Clin Invest*. 2016;126(10):3981–98.
6. Akin C, Metcalfe DD. The biology of Kit in disease and the application of pharmacogenetics. *J Allergy Clin Immunol*. 2004;114(1):13–9.
7. Garcia-Montero AC, Jara-Acevedo M, Teodosio C, Sanchez ML, Nunez R, Prados A, et al. KIT mutation in mast cells and other bone marrow hematopoietic cell lineages in systemic mast cell disorders: a prospective study of the Spanish Network on Mastocytosis (REMA) in a series of 113 patients. *Blood*. 2006;108(7):2366–72.
8. Iwaki S, Spicka J, Tkaczyk C, Jensen BM, Furumoto Y, Charles N, et al. Kit- and Fc epsilonRI-induced differential phosphorylation of the transmembrane adaptor molecule NTAL/LAB/LAT2 allows flexibility in its scaffolding function in mast cells. *Cell Signal*. 2008;20(1):195–205.
9. Kitayama H, Kanakura Y, Furitsu T, Tsujimura T, Oritani K, Ikeda H, et al. Constitutively activating mutations of c-kit receptor tyrosine kinase confer factor-independent growth and tumorigenicity of factor-dependent hematopoietic cell lines. *Blood*. 1995;85(3):790–8.
10. •• Rama TA, Morgado JM, Henriques A, Escribano L, Alvarez-Twose I, Sanchez-Munoz L, et al. Mastocytosis presenting with mast cell-mediator release-associated symptoms elicited by cyclo oxygenase inhibitors: prevalence, clinical, and laboratory features. *Clin Transl Allergy*. 2022;12(3):e12132. Findings from this study

- show that NSAIDs and other COX<sub>i</sub> are tolerated by around 9/10 of patients with mastocytosis.
11. Butterfield JH. Systemic mastocytosis: clinical manifestations and differential diagnosis. *Immunol Allergy Clin North Am.* 2006;26(3):487–513.
  12. Matito A, Alvarez-Twose I, Morgado JM, Sanchez-Munoz L, Orfao A, Escribano L. Anaphylaxis as a clinical manifestation of clonal mast cell disorders. *Curr Allergy Asthma Rep.* 2014;14(8):450.
  13. Escribano L, Orfao A. Anaphylaxis in mastocytosis. In: Castells MC, editor. *Anaphylaxis and hypersensitivity reactions.* New York: Springer; 2011.
  14. Gonzalez de Olano D, de la Hoz CB, Nunez Lopez R, Sanchez Munoz L, Cuevas Agustin M, Dieguez MC, et al. Prevalence of allergy and anaphylactic symptoms in 210 adult and pediatric patients with mastocytosis in Spain: a study of the Spanish network on mastocytosis (REMA). *Clin Exp Allergy.* 2007;37(10):1547–55.
  15. Brockow K, Jofer C, Behrendt H, Ring J. Anaphylaxis in patients with mastocytosis: a study on history, clinical features and risk factors in 120 patients. *Allergy.* 2008;63(2):226–32.
  16. Gulen T, Hagglund H, Dahlen B, Nilsson G. High prevalence of anaphylaxis in patients with systemic mastocytosis - a single-centre experience. *Clin Exp Allergy.* 2014;44(1):121–9.
  17. Gulen T, Ljung C, Nilsson G, Akin C. Risk factor analysis of anaphylactic reactions in patients with systemic mastocytosis. *J Allergy Clin Immunol Pract.* 2017;5(5):1248–55.
  18. Greiner G, Sprinzel B, Gorska A, Ratzinger F, Gurbisz M, Witzeneder N, et al. Hereditary alpha tryptasemia is a valid genetic biomarker for severe mediator-related symptoms in mastocytosis. *Blood.* 2021;137(2):238–47.
  19. Sordi B, Vanderwert F, Crupi F, Gesullo F, Zanotti R, Bonadonna P, et al. Disease correlates and clinical relevance of hereditary alpha-tryptasemia in patients with systemic mastocytosis. *J Allergy Clin Immunol.* 2023;151(2):485–93 e11.
  20. Chollet MB, Akin C. Hereditary alpha tryptasemia is not associated with specific clinical phenotypes. *J Allergy Clin Immunol.* 2022;149(2):728–35 e2.
  21. Couto ML, Silva M, Barbosa MJ, Ferreira F, Fragoso AS, Azenha RT. Defining hereditary alpha-tryptasemia as a risk/modifying factor for anaphylaxis: are we there yet? *Eur Ann Allergy Clin Immunol.* 2023;55(4):152–60.
  22. Rama TA, Torrado I, Henriques AF, Sanchez-Munoz L, Jara-Acevedo M, Navarro-Navarro P, et al. Mast cell activation syndromes: comparison between two scoring models to predict for mast cell clonality. *J Allergy Clin Immunol Pract.* 2023;11(3):908–19 e4.
  23. Hartmann K, Escribano L, Grattan C, Brockow K, Carter MC, Alvarez-Twose I, et al. Cutaneous manifestations in patients with mastocytosis: consensus report of the European Competence Network on Mastocytosis; the American Academy of Allergy, Asthma & Immunology; and the European Academy of Allergology and Clinical Immunology. *J Allergy Clin Immunol.* 2016;137(1):35–45.
  24. Escribano L, Akin C, Castells M, Schwartz LB. Current options in the treatment of mast cell mediator-related symptoms in mastocytosis. *Inflamm Allergy Drug Targets.* 2006;5(1):61–77.
  25. Alvarez-Twose I, Carter M. Pediatric mastocytosis. In: Akin C, editor. *Mastocytosis.* Cham: Springer; 2020. p. 93–114.
  26. Castells M. Drug allergy and perioperative management of mastocytosis. In: Akin C, editor. *Mastocytosis.* Cham: Springer International Publishing; 2020. p. 175–86.
  27. Rama TA, Miranda J, Silva D, Amaral L, Castro E, Coimbra A, et al. COVID-19 vaccination is safe among mast cell disorder patients, under adequate premedication. *Vaccines.* 2022;10(5):718.
  - 28.●● Giannetti MP, Olivieri F, Godwin G, Weller E, Nicoloso-SantaBarbara J, Bonadonna P, et al. Outcomes of COVID-19 vaccination in 323 patients with clonal and non-clonal mast cell activation disorders. *Allergy.* 2023;78(1):301–4. Findings from this large multicentric study show that COVID-19 vaccination is safe in patients with mastocytosis.
  29. Rama TA, Torrado I, Henriques AF, Sanchez-Munoz L, Matito A, Alvarez-Twose I. Drug hypersensitivity in indolent systemic mastocytosis, without skin lesions, presenting with anaphylaxis to hymenoptera venoms. *Allergy.* 2021;76(Suppl. 1):388.
  30. WHO Collaborating Centre for Drug Statistics Methodology (2018) Guidelines for ATC classification and DDD assignment, 2019. Oslo. [https://www.whocc.no/atc\\_ddd\\_index\\_and\\_guidelines/guidelines/](https://www.whocc.no/atc_ddd_index_and_guidelines/guidelines/).
  31. Toth L, Muszbek L, Komaromi I. Mechanism of the irreversible inhibition of human cyclooxygenase-1 by aspirin as predicted by QM/MM calculations. *J Mol Graph Model.* 2013;40:99–109.
  32. Graham GG, Davies MJ, Day RO, Mohamudally A, Scott KF. The modern pharmacology of paracetamol: therapeutic actions, mechanism of action, metabolism, toxicity and recent pharmacological findings. *Inflammopharmacology.* 2013;21(3):201–32.
  33. Blanca-Lopez N, Perez-Sanchez N, Agundez JA, Garcia-Martin E, Torres MJ, Cornejo-Garcia JA, et al. Allergic reactions to metamizole: immediate and delayed responses. *Int Arch Allergy Immunol.* 2016;169(4):223–30.
  34. Kowalski ML, Asero R, Bavbek S, Blanca M, Blanca-Lopez N, Bochenek G, et al. Classification and practical approach to the diagnosis and management of hypersensitivity to nonsteroidal anti-inflammatory drugs. *Allergy.* 2013;68(10):1219–32.
  35. Mastalerz L, Sanak M, Gawlewicz-Mroccka A, Gielicz A, Cmiel A, Szczeklik A. Prostaglandin E<sub>2</sub> systemic production in patients with asthma with and without aspirin hypersensitivity. *Thorax.* 2008;63(1):27–34.
  36. Steinke JW, Negri J, Liu L, Payne SC, Borish L. Aspirin activation of eosinophils and mast cells: implications in the pathogenesis of aspirin-exacerbated respiratory disease. *J Immunol.* 2014;193(1):41–7.

37. Mastalerz L, Setkowicz M, Sanak M, Szczeklik A. Hypersensitivity to aspirin: common eicosanoid alterations in urticaria and asthma. *J Allergy Clin Immunol*. 2004;113(4):771–5.
38. Daham K, James A, Balgoma D, Kupczyk M, Billing B, Lindeberg A, et al. Effects of selective COX-2 inhibition on allergen-induced bronchoconstriction and airway inflammation in asthma. *J Allergy Clin Immunol*. 2014;134(2):306–13.
39. Kowalski ML, Makowska JS, Blanca M, Bavbek S, Bochenek G, Bousquet J, et al. Hypersensitivity to nonsteroidal anti-inflammatory drugs (NSAIDs) - classification, diagnosis and management: review of the EAACI/ENDA(#) and GA2LEN/HANNA\*. *Allergy*. 2011;66(7):818–29.
40. Kidon M, Blanca-Lopez N, Gomes E, Terreehorst I, Tanno L, Ponvert C, et al. EAACI/ENDA Position Paper: diagnosis and management of hypersensitivity reactions to non-steroidal anti-inflammatory drugs (NSAIDs) in children and adolescents. *Pediatr Allergy Immunol*. 2018;29(5):469–80.
41. Woessner KM, Castells M. NSAID single-drug-induced reactions. *Immunol Allergy Clin North Am*. 2013;33(2):237–49.
42. Belay ED, Bresee JS, Holman RC, Khan AS, Shahriari A, Schonberger LB. Reye's syndrome in the United States from 1981 through 1997. *N Engl J Med*. 1999;340(18):1377–82.
43. Sanchez-Matas I, Matito-Bernechea A, Gonzalez de Olano D, Alvarez-Twose I, Sanchez-Munoz L, de la Hoz CB, et al. Prevalence of hypersensitivity reactions to nonsteroidal anti-inflammatory drugs in 212 patients with mastocytosis in Spain. *Allergy*. 2009;64:574–5.
44. • Bonadonna P, Olivieri F, Jarkvist J, Nalin F, Zanotti R, Maclachlan L, et al. Non-steroidal anti-inflammatory drug-induced anaphylaxis infrequent in 388 patients with mastocytosis: a two-center retrospective cohort study. *Front Allergy*. 2022;3:1071807. Findings from this study performed in two independent centers confirm that around 9/10 mastocytosis patients tolerate NSAIDs.
45. Hermans MAW, van der Vet SQA, van Hagen PM, van Wijk RG, van Daele PLA. Low frequency of acetyl salicylic acid hypersensitivity in mastocytosis: the results of a double-blind, placebo-controlled challenge study. *Allergy*. 2018;73(10):2055–62.
46. Carter MC, Metcalfe DD, Matito A, Escribano L, Butterfield JH, Schwartz LB, et al. Adverse reactions to drugs and biologics in patients with clonal mast cell disorders: a Work Group Report of the Mast Cells Disorder Committee, American Academy of Allergy, Asthma & Immunology. *J Allergy Clin Immunol*. 2019;143(3):880–93.
47. Bonadonna P, Pagani M, Aberer W, Bilo MB, Brockow K, Oude Elberink H, et al. Drug hypersensitivity in clonal mast cell disorders: ENDA/EAACI position paper. *Allergy*. 2015;70(7):755–63.
48. Reed SG, Orr MT, Fox CB. Key roles of adjuvants in modern vaccines. *Nat Med*. 2013;19(12):1597–608.
49. McNeil MM, Destefano F. Vaccine-associated hypersensitivity. *J Allergy Clin Immunol*. 2018;141(2):463–72.
50. Azenha Rama T, Moco Coutinho R, Mota D, Moreira A, Cernadas J. Hypersensitivity to the Moderna COVID-19 vaccine caused by tromethamine: PEG is not always the culprit excipient. *J Investig Allergol Clin Immunol*. 2022;32(5):414–5.
51. Mark A, Bjorksten B, Granstrom M. Immunoglobulin E responses to diphtheria and tetanus toxoids after booster with aluminium-adsorbed and fluid DT-vaccines. *Vaccine*. 1995;13(7):669–73.
52. Klimek L, Novak N, Cabanillas B, Jutel M, Bousquet J, Akdis CA. Allergenic components of the mRNA-1273 vaccine for COVID-19: possible involvement of polyethylene glycol and IgG-mediated complement activation. *Allergy*. 2021;76(11):3307–13.
53. Li DH, Lee E, Song C. Successful mRNA COVID-19 vaccination in a patient with a history of severe polyethylene glycol anaphylaxis. *Research Square*. 2022;
54. Maltezou HC, Anastassopoulou C, Hatziantoniou S, Poland GA, Tsakris A. Anaphylaxis rates associated with COVID-19 vaccines are comparable to those of other vaccines. *Vaccine*. 2022;40(2):183–6.
55. Sobczak M, Pawliczak R. The risk of anaphylaxis behind authorized COVID-19 vaccines: a meta-analysis. *Clin Mol Allergy*. 2022;20(1):1.
56. Stone CA Jr, Liu Y, Relling MV, Krantz MS, Pratt AL, Abreo A, et al. Immediate hypersensitivity to polyethylene glycols and polysorbates: more common than we have recognized. *J Allergy Clin Immunol Pract*. 2019;7(5):1533–40 e8.
57. Wenande E, Garvey LH. Immediate-type hypersensitivity to polyethylene glycols: a review. *Clin Exp Allergy*. 2016;46(7):907–22.
58. Schreiner M, Zobel C, Baumgarten U, Uhlmann T, Vanderssee S. Anaphylactic reactions to polyethylene glycol-containing bowel cleansing preparations after Moderna COVID-19 vaccination. *Endoscopy*. 2022;54(5):517–8.
59. Morgado JM, Sanchez-Munoz L, Teodosio C, Mora LME. Identification and immunophenotypic characterization of normal and pathological mast cells. *Methods Mol Biol*. 2020;2163:331–53.
60. McNeil MM, Weintraub ES, Duffy J, Sukumaran L, Jacobsen SJ, Klein NP, et al. Risk of anaphylaxis after vaccination in children and adults. *J Allergy Clin Immunol*. 2016;137(3):868–78.
61. Zanoni G, Zanotti R, Schena D, Sabbadini C, Opri R, Bonadonna P. Vaccination management in children and adults with mastocytosis. *Clin Exp Allergy*. 2017;47(4):593–6.
62. Parente R, Pucino V, Magliacane D, Petraroli A, Loffredo S, Marone G, et al. Evaluation of vaccination safety in children with mastocytosis. *Pediatr Allergy Immunol*. 2017;28(1):93–5.
63. Alvarez-Twose I, Vano-Galvan S, Sanchez-Munoz L, Morgado JM, Matito A, Torrelo A, et al. Increased serum baseline tryptase levels and extensive skin involvement

- are predictors for the severity of mast cell activation episodes in children with mastocytosis. *Allergy*. 2012;67(6):813–21.
64. Abuhay H, Clark AS, Carter MC. Occurrence of unexpected adverse reactions to vaccines in children with mastocytosis. *J Pediatr Res*. 2020;7(1):81–6.
  65. Johansen ML, Lawley LP. Assessing vaccination reactions in pediatric patients with maculopapular cutaneous mastocytosis. *Pediatr Dermatol*. 2021;38(2):502–3.
  66. Bankova LG, Walter JE, Iyengar SR, Lorenzo ME, Hornick JL, Castells MC. Generalized bullous eruption after routine vaccination in a child with diffuse cutaneous mastocytosis. *J Allergy Clin Immunol Pract*. 2013;1(1):94–6.
  67. Gupta M, Akin C, Sanders GM, Chan MP, Ross CW, Castells MC. Blisters, Vaccines, and mast cells: a difficult case of diffuse cutaneous mastocytosis. *J Allergy Clin Immunol Pract*. 2019;7(4):1370–2.
  68. Lazarinis N, Bossios A, Gulen T. COVID-19 vaccination in the setting of mastocytosis-Pfizer-BioNTech mRNA vaccine is safe and well tolerated. *J Allergy Clin Immunol Pract*. 2022;10(5):1377–9.
  69. Sriskandarajah P, Hobart J, Radia DH, Whyte AF. A UK survey examining the experience of adults with mastocytosis receiving COVID-19 vaccination. *Hemasphere*. 2021;5(11):e650.
  70. Ruano-Zaragoza M, Carpio-Escalona LV, Diaz-Beya M, Piris-Villarespa M, Castano-Diez S, Munoz-Cano R, et al. Safety of COVID-19 vaccination in patients with clonal mast cell disorders. *J Allergy Clin Immunol Pract*. 2022;10(5):1374–6 e3.
  71. Kaakati R, Khokhar D, Akin C. Safety of COVID-19 vaccination in patients with mastocytosis and monoclonal mast cell activation syndrome. *J Allergy Clin Immunol Pract*. 2021;9(8):3198–9.
  72. Escribano L, Akin C, Castells M, Orfao A, Metcalfe DD. Mastocytosis: current concepts in diagnosis and treatment. *Ann Hematol*. 2002;81(12):677–90.
  73. Hermans MAW, Arends NJT, Gerth van Wijk R, van Hagen PM, Kluin-Nelemans HC, Oude Elberink HNG, et al. Management around invasive procedures in mastocytosis: an update. *Ann Allergy Asthma Immunol*. 2017;119(4):304–9.
  74. Dewachter P, Castells MC, Hepner DL, Mouton-Faivre C. Perioperative management of patients with mastocytosis. *Anesthesiology*. 2014;120(3):753–9.
  75. Matito A, Morgado JM, Sanchez-Lopez P, Alvarez-Twose I, Sanchez-Munoz L, Orfao A, et al. Management of anesthesia in adult and pediatric mastocytosis: a study of the Spanish Network on Mastocytosis (REMA) based on 726 anesthetic procedures. *Int Arch Allergy Immunol*. 2015;167(1):47–56.
  76. Carter MC, Uzzaman A, Scott LM, Metcalfe DD, Quezado Z. Pediatric mastocytosis: routine anesthetic management for a complex disease. *Anesth Analg*. 2008;107(2):422–7.
  77. Mertes PM, Laxenaire MC, Alla F. Groupe d'Etudes des Reactions Anaphylactoides P. Anaphylactic and anaphylactoid reactions occurring during anesthesia in France in 1999–2000. *Anesthesiology*. 2003;99(3):536–45.
  78. Harboe T, Guttormsen AB, Irgens A, Dybendal T, Florvaag E. Anaphylaxis during anesthesia in Norway: a 6-year single-center follow-up study. *Anesthesiology*. 2005;102(5):897–903.
  79. Koppert W, Blunk JA, Petersen LJ, Skov P, Rentsch K, Schmelz M. Different patterns of mast cell activation by muscle relaxants in human skin. *Anesthesiology*. 2001;95(3):659–67.
  80. Navines-Ferrer A, Serrano-Candelas E, Lafuente A, Munoz-Cano R, Martin M, Gastaminza G. MRGPRX2-mediated mast cell response to drugs used in perioperative procedures and anaesthesia. *Sci Rep*. 2018;8(1):11628.
  81. Renauld V, Goudet V, Mouton-Faivre C, Debaene B, Dewachter P. Case report: perioperative immediate hypersensitivity involves not only allergy but also mastocytosis. *Can J Anaesth*. 2011;58(5):456–9.
  82. McNeil BD, Pundir P, Meeker S, Han L, Udem BJ, Kulka M, et al. Identification of a mast-cell-specific receptor crucial for pseudo-allergic drug reactions. *Nature*. 2015;519(7542):237–41.
  83. Stellato C, de Paulis A, Cirillo R, Mastronardi P, Mazarella B, Marone G. Heterogeneity of human mast cells and basophils in response to muscle relaxants. *Anesthesiology*. 1991;74(6):1078–86.
  84. Doenicke A, Soukup J, Hoerneck R, Moss J. The lack of histamine release with cisatracurium: a double-blind comparison with vecuronium. *Anesth Analg*. 1997;84(3):623–8.
  85. Reddy JI, Cooke PJ, van Schalkwyk JM, Hannam JA, Fitzharris P, Mitchell SJ. Anaphylaxis is more common with rocuronium and succinylcholine than with atracurium. *Anesthesiology*. 2015;122(1):39–45.
  86. Baldo BA, Pham NH. Histamine-releasing and allergic properties of opioid analgesic drugs: resolving the two. *Anaesth Intensive Care*. 2012;40(2):216–35.
  87. Edston E, van Hage-Hamsten M. Anaphylactoid shock—a common cause of death in heroin addicts? *Allergy*. 1997;52(9):950–4.
  88. Blunk JA, Schmelz M, Zeck S, Skov P, Likar R, Koppert W. Opioid-induced mast cell activation and vascular responses is not mediated by mu-opioid receptors: an in vivo microdialysis study in human skin. *Anesth Analg*. 2004;98(2):364–70.
  89. Sheen CH, Schleimer RP, Kulka M. Codeine induces human mast cell chemokine and cytokine production: involvement of G-protein activation. *Allergy*. 2007;62(5):532–8.
  90. Stellato C, Cirillo R, de Paulis A, Casolaro V, Patella V, Mastronardi P, et al. Human basophil/mast cell releasability. IX. Heterogeneity of the effects of opioids on mediator release. *Anesthesiology*. 1992;77(5):932–40.
  91. Di Bello MG, Masini E, Ioannides C, Ndisang JF, Raspanti S, Bani Sacchi T, et al. Histamine release from rat mast cells induced by the metabolic activation

- of drugs of abuse into free radicals. *Inflamm Res*. 1998;47(3):122–30.
92. Baldo BA, Pham NH. Opioid toxicity: histamine, hypersensitivity, and MRGPRX2. *Arch Toxicol*. 2023;97(2):359–75.
  93. Church MK, Pao GJ, Holgate ST. Characterization of histamine secretion from mechanically dispersed human lung mast cells: effects of anti-IgE, calcium ionophore A23187, compound 48/80, and basic polypeptides. *J Immunol*. 1982;129(5):2116–21.
  94. Tharp MD, Kagey-Sobotka A, Fox CC, Marone G, Lichtenstein LM, Sullivan TJ. Functional heterogeneity of human mast cells from different anatomic sites: in vitro responses to morphine sulfate. *J Allergy Clin Immunol*. 1987;79(4):646–53.
  95. Gulen T, Moller Westerberg C, Lyberg K, Ekoff M, Kolmert J, Bood J, et al. Assessment of in vivo mast cell reactivity in patients with systemic mastocytosis. *Clin Exp Allergy*. 2017;47(7):909–17.
  96. Bonadonna P, Lombardo C. Drug allergy in mastocytosis. *Immunol Allergy Clin North Am*. 2014;34(2):397–405.
  97. Mertes PM, Ebo DG, Garcez T, Rose M, Sabato V, Takazawa T, et al. Comparative epidemiology of suspected perioperative hypersensitivity reactions. *Br J Anaesth*. 2019;123(1):e16–28.
  98. Stellato C, Casolaro V, Ciccarelli A, Mastronardi P, Mazzarella B, Marone G. General anaesthetics induce only histamine release selectively from human mast cells. *Br J Anaesth*. 1991;67(6):751–8.
  99. Fujimoto T, Nishiyama T, Hanaoka K. Inhibitory effects of intravenous anesthetics on mast cell function. *Anesth Analg*. 2005;101(4):1054–9.
  100. Yu X, Sun X, Zhao M, Hou Y, Li J, Yu J, et al. Propofol attenuates myocardial ischemia reperfusion injury partly through inhibition of resident cardiac mast cell activation. *Int Immunopharmacol*. 2018;54:267–74.
  101. Li Y, Sun X, Juan Z, Guan X, Wang M, Meng Y, et al. Propofol pretreatment alleviates mast cell degranulation by inhibiting SOC to protect the myocardium from ischemia-reperfusion injury. *Biomed Pharmacother*. 2022;150:113014.
  102. Gan X, Xing D, Su G, Li S, Luo C, Irwin MG, et al. Propofol attenuates small intestinal ischemia reperfusion injury through inhibiting NADPH oxidase mediated mast cell activation. *Oxid Med Cell Longev*. 2015;2015:167014.
  103. Haybarger E, Young AS, Giovannitti JA Jr. Benzodiazepine allergy with anesthesia administration: a review of current literature. *Anesth Prog*. 2016;63(3):160–7.
  104. Suzuki-Nishimura T, Sano T, Uchida MK. Effects of benzodiazepines on serotonin release from rat mast cells. *Eur J Pharmacol*. 1989;167(1):75–85.
  105. Miller LG, Lee-Paritz A, Greenblatt DJ, Theoharides TC. High-affinity benzodiazepine binding sites on rat peritoneal mast cells and RBL-1 cells: binding characteristics and effects on granule secretion. *Pharmacology*. 1988;36(1):52–60.
  106. Bidri M, Royer B, Averlant G, Bismuth G, Guillosson JJ, Arock M. Inhibition of mouse mast cell proliferation and proinflammatory mediator release by benzodiazepines. *Immunopharmacology*. 1999;43(1):75–86.
  107. Rama TA, Corte-Real I, Gomes PS, Escrivano L, Fernandes MH. Mastocytosis: oral implications of a rare disease. *J Oral Pathol Med*. 2010;40(6):441–50.
  108. Hepner DL, Castells MC. Anaphylaxis during the perioperative period. *Anesth Analg*. 2003;97(5):1381–95.
  109. Ahmad N, Evans P, Lloyd-Thomas AR. Anesthesia in children with mastocytosis--a case based review. *Paediatr Anaesth*. 2009;19(2):97–107.
  110. Mertes PM, Tajima K, Regnier-Kimmoun MA, Lambert M, Iohom G, Gueant-Rodriguez RM, et al. Perioperative anaphylaxis. *Med Clin North Am*. 2010;94(4):761–89.
  111. Broesby-Olsen S, Vestergaard H, Mortz CG, Jensen B, Havelund T, Hermann AP, et al. Omalizumab prevents anaphylaxis and improves symptoms in systemic mastocytosis: efficacy and safety observations. *Allergy*. 2018;73(1):230–8.
  112. Phillips JF, Yates AB, Deshazo RD. Approach to patients with suspected hypersensitivity to local anesthetics. *Am J Med Sci*. 2007;334(3):190–6.
  113. Malinovsky JM, Chiriac AM, Tacquard C, Mertes PM, Demoly P. Allergy to local anesthetics: reality or myth? *Presse Med*. 2016;45(9):753–7.
  114. Kvisselgaard AD, Mosbech HF, Fransson S, Garvey LH. Risk of immediate-type allergy to local anesthetics is overestimated--results from 5 years of provocation testing in a Danish Allergy Clinic. *J Allergy Clin Immunol Pract*. 2018;6(4):1217–23.
  115. Campbell JR, Maestrello CL, Campbell RL. Allergic response to metabisulfite in lidocaine anesthetic solution. *Anesth Prog*. 2001;48(1):21–6.
  116. Becker DE, Reed KL. Essentials of local anesthetic pharmacology. *Anesth Prog* 2006;53(3):98–108.
  117. Specia SJ, Boynes SC, Cuddy MA. Allergic reactions to local anesthetic formulations. *Dent Clin N Am*. 2010;54(4):655–64.
  118. Kvisselgaard AD, Kroigaard M, Mosbech HF, Garvey LH. No cases of perioperative allergy to local anaesthetics in the Danish Anaesthesia Allergy Centre. *Acta Anaesthesiol Scand*. 2017;61(2):149–55.
  119. Ring J, Franz R, Brockow K. Anaphylactic reactions to local anesthetics. *Chem Immunol Allergy*. 2010;95:190–200.
  120. Macy E. Practical management of patients with a history of immediate hypersensitivity to common non-beta-lactam drugs. *Curr Allergy Asthma Rep*. 2016;16(1):4.
  121. Kolbe AB, Hartman RP, Hoskin TL, Carter RE, Maddox DE, Hunt CH, et al. Premedication of patients for prior urticarial reaction to iodinated contrast medium. *Abdom Imaging*. 2014;39(2):432–7.
  122. Schwaab J, Brockow K, Riffel P, Lubke J, Naumann N, Jawhar M, et al. Low risk of contrast media-induced hypersensitivity reactions in all subtypes of systemic mastocytosis. *Ann Allergy Asthma Immunol*. 2022;128(3):314–8.
  123. Stellato C, Crescenzo GD, Patella V, Mastronardi P, Mazzarella B, Marone G. Human basophil/mast cell

- releasability. XI. Heterogeneity of the effects of contrast media on mediator release. *J Allergy Clin Immunol.* 1996;97(3):838–50.
124. Kulka M, Sheen CH, Tancowny BP, Grammer LC, Schleimer RP. Neuropeptides activate human mast cell degranulation and chemokine production. *Immunology.* 2008;123(3):398–410.
  125. Yoon SH, Lee SY, Kang HR, Kim JY, Hahn S, Park CM, et al. Skin tests in patients with hypersensitivity reaction to iodinated contrast media: a meta-analysis. *Allergy.* 2015;70(6):625–37.
  126. Trautmann A, Brockow K, Behle V, Stoevesandt J. Radiocontrast media hypersensitivity: skin testing differentiates allergy from nonallergic reactions and identifies a safe alternative as proven by intravenous provocation. *J Allergy Clin Immunol Pract.* 2019;7(7):2218–24.
  127. Yuan F, Zhang C, Sun M, Wu D, Cheng L, Pan B, et al. MRGPRX2 mediates immediate-type pseudo-allergic reactions induced by iodine-containing iohexol. *Biomed Pharmacother.* 2021;137:111323.
  128. Lyberg K, Ekoff M, Westerberg CM, Engblom C, Dahlen B, Gulen T, et al. Mast cells derived from systemic mastocytosis exhibit an increased responsiveness to hyperosmolarity. *Allergy.* 2022;77(6):1909–11.
  129. De la Hoz B, Gonzalez de Olano D, Alvarez I, Sanchez L, Nunez R, Sanchez I, et al. Guidelines for the diagnosis, treatment and management of mastocytosis. *An Sist Sanit Navar.* 2008;31(1):11–32.
  130. Weingarten TN, Volcheck GW, Sprung J. Anaphylactoid reaction to intravenous contrast in patient with systemic mastocytosis. *Anaesth Intensive Care.* 2009;37(4):646–9.
  131. Valabhji J, Robinson S, Johnston D, Bellamy M, Davies W, Bain BJ. Unexplained loss of consciousness: systemic mastocytosis. *J R Soc Med.* 2000;93(3):141–2.
  132. Kors JW, van Doormaal JJ, JGR DM. Anaphylactoid shock following Hymenoptera sting as a presenting symptom of systemic mastocytosis. *J Intern Med.* 1993;233(3):255–8.
  133. Bonadonna P, Zanotti R, Pagani M, Caruso B, Perbellini O, Colarossi S, et al. How much specific is the association between hymenoptera venom allergy and mastocytosis? *Allergy.* 2009;64(9):1379–82.
  134. Alvarez-Twose I, Gonzalez de Olano D, Sanchez-Munoz L, Matito A, Esteban-Lopez MI, Vega A, et al. Clinical, biological, and molecular characteristics of clonal mast cell disorders presenting with systemic mast cell activation symptoms. *J Allergy Clin Immunol.* 2010;125(6):1269–78 e2.
  135. Manfredi M, Severino M, Testi S, Macchia D, Ermini G, Pichler WJ, et al. Detection of specific IgE to quinolones. *J Allergy Clin Immunol.* 2004;113(1):155–60.
  136. Yarrington ME, Anderson DJ, Dodds Ashley E, Jones T, Davis A, Johnson M, et al. Impact of FDA black box warning on fluoroquinolone and alternative antibiotic use in southeastern US hospitals. *Infect Control Hosp Epidemiol.* 2019;40(11):1297–300.
  137. Aldred KJ, Kerns RJ, Osheroff N. Mechanism of quinolone action and resistance. *Biochemistry.* 2014;53(10):1565–74.
  138. Burke P, Burne SR. Allergy associated with ciprofloxacin. *BMJ.* 2000;320(7236):679.
  139. Giavina-Bianchi P, Gonçalves DG, Zanandrea A, Borges De Castro R, Garro LS, Kalil J, et al. Anaphylaxis to quinolones in mastocytosis: hypothesis on the mechanism. *The Journal of Allergy and Clinical Immunology. In Pract.* 2019;7(6):2089–90.
  140. Weiler CR. Mastocytosis, quinolones, MRGPRX2, and anaphylaxis. *The Journal of Allergy and Clinical Immunology. In Pract.* 2019;7(6):2091–2.
  141. McNeil BD. MRGPRX2 and adverse drug reactions. *Front Immunol.* 2021;12:676354.
  142. Alvarez-Arango S, Ogunwole SM, Sequist TD, Burk CM, Blumenthal KG. Vancomycin infusion reaction - moving beyond "Red Man Syndrome". *N Engl J Med.* 2021;384(14):1283–6.

## Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.