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Silicon-mediated herbivore defence in a pasture grass under reduced and Anthropocene levels of CO₂

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The uptake and accumulation of silicon (Si) in grass plants play a crucial role in alleviating both biotic and abiotic stresses. Si supplementation has been reported to increase activity of defence-related antioxidant enzyme, which helps to reduce oxidative stress caused by reactive oxygen species (ROS) following herbivore attack. Atmospheric CO_2 levels are known to affect Si accumulation in grasses; reduced CO₂ concentrations increase Si accumulation whereas elevated CO₂ concentrations often decrease Si accumulation. This can potentially affect antioxidant enzyme activity and subsequently insect herbivory, but this remains untested. We examined the effects of Si supplementation and herbivory by Helicoverpa armigera on antioxidant enzyme (catalase, CAT; superoxide dismutase, SOD; and ascorbate peroxidase, APX) activity in tall fescue grass (Festuca arundinacea) grown under CO_2 concentrations of 200, 410, and 640 ppm representing reduced, ambient, and elevated CO₂ levels, respectively. We also quantified foliar Si, carbon (C), and nitrogen (N) concentrations and determined how changes in enzymes and elemental chemistry affected H. armigera relative growth rates and plant consumption. Rising CO₂ concentrations increased plant mass and foliar C but decreased foliar N and Si. Si supplementation enhanced APX and SOD activity under the ranging CO₂ regimes. Si accumulation and antioxidant enzyme activity were at their highest level under reduced CO₂ conditions and their lowest level under future levels of CO₂. The latter corresponded with increased herbivore growth rates and plant consumption, suggesting that some grasses could become more susceptible to herbivory under projected CO₂ conditions.

KEYWORDS

antioxidant enzyme, carbon, carbon dioxide, herbivore, physical defences, plant defences, silicon

Introduction

Most grasses are silicon (Si) accumulators, which can account for up to 10% of their dry mass (Epstein, 1994). Si uptake and accumulation are a functional trait with multiple implications for plant biology and ecology (Epstein, 2009). Si is taken up as silicic acid [Si(OH)₄] via the roots and, after being transported into plant tissues, is deposited within and between plant cells, the cell wall, and silicified structures such as trichomes or other phytoliths (Perry et al., 1984; Kumar et al., 2017). Although the role of Si in protecting plants against multiple biotic (e.g., herbivores and pathogens) and abiotic (e.g., drought and salinity) stresses has been widely reported (Cooke and Leishman, 2011; Debona et al., 2017), an understanding of the exact mechanisms underpinning such protection remains incomplete (Coskun et al., 2019). However, the consensus is that Si supplementation enhances plant physical defences and integrates with the regulation of secondary metabolite defences (Reynolds et al., 2016; Alhousari and Greger, 2018; Hall et al., 2019; Waterman et al., 2020).

In terms of physical defences, it is well established that Si confers plant resistance and reduces plant damage caused by both vertebrate and invertebrate herbivores (Massey and Hartley, 2009; Hartley et al., 2015; Alhousari and Greger, 2018). Si deposition within and around plant cells makes plant tissues tougher and abrasive, causing wear on herbivore mouthparts, damages digestive organs, inhibits movement, and reduces the feeding efficiency of insect herbivores (Massey et al., 2006; Reynolds et al., 2009). Moreover, Si is known to alter grass physical defences such as macrohairs, silica cells, and prickle cells, which are linked to reduced feeding by insect herbivores (Hartley et al., 2015; Hall et al., 2020; Biru et al., 2021). Si uptake and accumulation have also been shown to be induced following herbivory (Massey et al., 2007; Islam et al., 2020; Biru et al., 2022).

In addition to direct physical defences, Si potentially protects plants against herbivores by influencing production of plant biochemical defences (Reynolds et al., 2016; Yang et al., 2017), although there is much uncertainty about this since Si has limited chemical reactivity within the plant. Herbivore attack is associated with the induction of oxidative stress in plants, resulting from overproduction of reactive oxygen species (ROS) (Bi and Felton, 1995; Kerchev et al., 2012). For instance, insect herbivore attacks can induce various ROS such as hydrogen peroxide (H₂O₂), superoxide $(O_2 \bullet -)$, singlet oxygen $(^1O_2)$, or hydroxyl radicals (•OH) in cells (Sharma et al., 2012; Das and Roychoudhury, 2014). While ROS have signalling roles under physiological setup (Hasanuzzaman et al., 2020), (biotic) stress-induced ROS overproduction damages cell structures and functionality (Tripathy and Oelmüller, 2012; Das and Roychoudhury, 2014; Fichman and Mittler, 2020). In order to reduce excessive ROS content caused by the imbalance between free radical formation and the capability of cells to detoxify them (Pizzino et al., 2017), plants have developed efficient antioxidant enzymatic machinery to scavenge ROS (Tripathy and Oelmüller, 2012). The antioxidant defence system in the plant cell includes both enzyme constituents such as superoxide dismutase (SOD), catalase (CAT), ascorbate peroxidase (APX), glutathione reductase (GR), and non-enzyme constituents like cysteine (Cys), reduced glutathione (GSH), and ascorbic acid (AsA) (Farooq et al., 2013; Kim et al., 2017). Plants possess either constitutive or induced antioxidants (Sudhakar et al., 2001) and the increased activities of these enzymes in plant cells appear to better control oxidative stress (Das and Roychoudhury, 2014; Huang et al., 2019).

Exogenous Si application has been linked to enhanced activity of antioxidant enzyme defences such as CAT, APX, and SOD (Kim et al., 2016; Hasanuzzaman et al., 2018; Ahanger et al., 2020). However, the mode of action by which Si regulates antioxidant capacities is poorly understood. Previous work has shown that Si enhances SOD, CAT, APX, and peroxidase activities in rice (*Oryza sativa*) (Han et al., 2016), wheat (*Triticum aestivum* L.) (Gong et al., 2005), and maize (*Zea mays* L.) (Moussa, 2006). Si-mediated regulation of antioxidant defences, therefore, reduces the harmful effects of herbivore-induced oxidative stress (Ahanger et al., 2020; Acevedo et al., 2021).

Atmospheric concentrations of carbon dioxide (CO_2) have emerged as an important environmental driver of Si accumulation (Johnson and Hartley, 2018). In general, several studies report that elevated CO₂ concentrations (eCO₂) decrease Si accumulation (Ryalls et al., 2017; Johnson and Hartley, 2018; Johnson et al., 2023), but see Frew et al. (2017) and Fulweiler et al. (2014). In contrast, reduced levels of atmospheric CO₂ can lead to increased Si accumulation (Biru et al., 2020; Biru et al., 2021). These effects are likely due to carbon (C) being either more available under eCO₂ (Johnson and Hartley, 2018; Johnson et al., 2022) or less available under reduced CO₂ conditions (Biru et al., 2020). Si accumulation is often negatively correlated with C potentially due to stoichiometric dilution or Si substitution for C-based structural or defensive compounds (Raven, 1983; Hodson et al., 2005).

Given CO₂ is such an important driver of Si accumulation, which has been shown to influence enzymatic responses (e.g., Kim et al., 2016; Ahanger et al., 2020; Acevedo et al., 2021), CO2 may also affect production of plant biochemical defences (i.e., antioxidant enzyme), potentially via enhanced plant susceptibility to herbivore-induced oxidative stress. To our knowledge, no studies have yet investigated the effects of variable rates of Si accumulation on antioxidant enzyme activity and regulation of herbivore-induced oxidative stress under different CO₂ concentrations. Using tall fescue (Festuca arundinacea) and the generalist insect herbivore, cotton bollworm [Helicoverpa armigera (Hübner)], we investigated the effect of Si treatments on antioxidant enzyme activities and foliar chemistry of plants grown under three CO₂ concentrations (200, 410, and 640 ppm) and the consequences for insect herbivory. The objective of this study was to determine how Si treatments (+Si or -Si) under different CO2 concentrations affect antioxidant enzyme activity and foliar chemistry (e.g., C, N) in tall fescue and the consequences for insect herbivore growth rate and feeding efficiency. We hypothesised that (1) +Si and reduced CO₂ decrease shoot C concentrations but increase shoot N concentration, i.e., decreasing shoot C-to-N ratio, and (2) +Si together with reduced CO2 treatments increases antioxidant enzyme activity, whereas -Si together with elevated CO₂ would decrease antioxidant enzyme activity.

Materials and methods

Plant material and growth conditions

Tall fescue (Festuca arundinacea) is a common pasture grass and a high Si accumulator (Hodson et al., 2005). Seeds of tall fescue (accession T 9627), obtained from Margot Forde Germplasm Centre (Palmerston North, New Zealand), were sterilised in a solution of 0.9% sodium hypochlorite and 0.1% Triton X-100 for 30 min, followed by washing several times with water before being inserted into perlite irrigated with water. Seeds were stratified at 4°C for 3 days, and plants were grown in the glasshouse for 2 weeks using a rectangular plastic tray to achieve uniform seedling growth. Two weeks after germination, individual plants were transferred to hydroponics cups. The hydroponics setup consisted of two nested disposable plastic cups as per Hall et al. (2019). Each cup was filled with approximately 350 mL of full-strength standard hydroponic solutions following the protocol of Jung et al. (2015). Seedlings were grown in three plant growth chambers (TPG-1260TH, Thermoline Scientific, NSW, Australia), maintained at a reduced CO₂ of 200 ppm, an ambient CO₂ of 410 ppm, and an elevated CO₂ of 640 ppm CO2 concentrations; the latter CO2 concentration predicted for 2100 under the RCP6.0 scenario outlined by the IPCC (2014). Chambers were illuminated with five 400-W Sunmaster Dual Spectrum High-Pressure Sodium globes at 350 µmol m⁻² s⁻¹ at the plant canopy level. Daytime air temperature was regulated at 26° C and fell to 18°C at night on a 15L:9D photoperiod cycle. Humidity was controlled at 50% (\pm 6%). Carbon dioxide within the chambers was monitored by a Li-Cor LI-820 CO2 gas analyser, with CO2 (food grade, Air Liquide, NSW, Australia) injected from pressurised cylinders through solenoid valves. For reduced CO₂ treatment in the chamber, the computer controller constantly monitors CO₂ and powers fans to direct chamber air through the scrubbers filled with Sodasorb[®] (W.R. Grace & Co, Chicago, USA).

Experimental design

The experimental design consisted of 252 hydroponically grown tall fescue plants. The experiment comprised a factorial combination of CO₂ concentrations (200, 410, or 640 ppm), Si (+Si or -Si), and herbivore (herbivory, +H; no herbivory, -H) treatments (see Figure 1). Si treatments (+Si) used liquid potassium silicate (K₂SiO₃) (Agsil32, PQ Australia, SA, Australia) at a concentration equivalent to 2 mM SiO₂. Chemically, silicic acid polymerises to form silica gel when the concentration of silicic acid exceeds 2 mM (Ma and Yamaji, 2006). Potassium chloride was added to the control (-Si) cups to balance additional K⁺ ions in +Si treatments. The pH of both +Si and -Si solutions was adjusted to 5.6 using hydrochloric acid to reduce silicate polymerisation (Ma and Yamaji, 2006). Solutions were replaced weekly for the first 2 weeks and then three times a week afterwards. Cups were rotated and chambers were swapped weekly to minimise chamber effects and pseudo-replication as previously described by Johnson et al. (2018). Plants were grown hydroponically for a further 6 weeks (42 days, Figure 1) before insect inoculation.

Herbivore performance

To assess the impacts of different CO₂ concentrations and Si supplementation on the growth of H. armigera larvae, a feeding performance assay was conducted. Helicoverpa armigera third instar larvae supplied by CSIRO Agriculture & Food, Narrabri, Australia, reared on an artificial diet (Teakle and Jensen, 1985), were used for feeding assays. Initially, larvae were starved for 24 h and weighed, and a single larva was either applied for each plant shoot [herbivory (+H)]) or kept control [no herbivory (-H)] (see Figure 1 for details). Each plant was then caged with transparent Perspex sheaths and herbivores were allowed to feed on shoot parts of the plants for 6 days, after which they were removed and starved for a further 24 h to allow the frass to pass, before being reweighed. All frass were collected, dried, and weighed. Frass production was used as a surrogate for plant consumption. RGR was calculated according to Massey and Hartley (2009). RGR estimates the change in larval fresh mass relative to initial mass and was calculated as mass gained (mg)/initial mass (mg)/time (days).

Antioxidant enzyme activity assays

Immediately after herbivore removal, plants from all treatment groups were harvested into liquid nitrogen and stored at -80° C until analysis. For the measurement of enzymatic activities, ca. 0.05 g of leaf tissue was ground in liquid N and homogenised in 3 mL of ice-cold 100 mM K-phosphate buffer (pH 6.8) containing 0.1 mM EDTA. The homogenate was centrifuged at 16,000 g for 15 min and the supernatant was used as the source of crude extracts. The supernatant was utilised to measure the activity of antioxidant enzyme such as CAT, APX, and SOD.

CAT activity was measured spectrophotometrically following the method of Fimognari et al. (2020) and Maksimović and Živanović (2012). Reaction mixture consists of 50 mM potassium phosphate buffer, pH 7.0, 20 mM H₂O₂, and crude extract. Absorbance at 240 nm was recorded for 130s using CLARIOstar[®] plate reader (BMG Labtech, Ortenberg, Germany) in 96-well plates (96-well Flat Bottom Plate, Greiner Bio-one, Australia). For control reactions, H₂O₂ was omitted. Enzyme activity was calculated using the molar extinction coefficient 36×10^3 mM⁻¹ m⁻¹ and expressed as µmol H₂O₂ oxidised g⁻¹ FW min⁻¹.

APX activity was assayed according to Hartmann and Asch (2019). The reaction mixture consists of 50 mM potassium phosphate buffer, pH 7.0, 0.2 mM ascorbate, 0.2 mM H_2O_2 and crude extract. Absorbance at 290 nm was recorded for 130 s using CLARIOstar[®] in 96-well plates. APX activity was calculated according to Hartmann and Asch (2019); one unit of APX is defined as the amount of enzyme required to oxidise 1 µmol of ascorbic acid per minute.

SOD activity was estimated following the inhibition of photochemical reduction of nitroblue tetrazolium (NBT) by the enzyme according to Hartmann and Asch (2019). The reaction mixture contained 0.05 M sodium carbonate, 13.3 mM methionine, 1.3 μ M riboflavin, 21 μ M NBT, and plant extract (Hartmann and Asch, 2019). The reaction took place in a chamber under



illumination of a 30-W fluorescent lamp at 25°C. The reaction was started by turning the fluorescent lamp on and stopped 5 min later by turning it off. The blue formazan produced by NBT photoreduction was measured as increase in absorbance at 560 nm. The blank solution had the same complete reaction mixture but was kept in the dark. One SOD unit was defined as the amount of enzyme required to inhibit 50% of the NBT photoreduction in comparison with wells lacking the plant extract and expressed as units of enzyme activity g^{-1} FW min⁻¹ (Cavalcanti et al., 2004).

Analyses of foliar chemistry

Harvested sample leaves were oven-dried for 3 days at 60°C and ball milled for further analysis (see sample size from Figure 1). For foliar Si analysis, roughly 80 mg of ground leaf material was analysed using x-ray fluorescence spectrometry (Epsilon 3^x, PANalytical, EA Almelo, The Netherlands) as per Reidinger et al. (2012). Si measurements were calibrated against a certified plant reference material of known Si concentrations (Hiltpold et al., 2017). For foliar C and N concentration, approximately 7 mg of ground leaf material was analysed using Elementar Vario EL Cube, CHNOS elemental analyser (Elementar Analysensysteme GmbH, Hanau, Germany), at a combustion temperature of 950°C.

Statistical analysis

All data were analysed using SPSS (version 27) statistical software. Before analysis, all data were checked for assumptions of normality for residuals according to inspection of quantile– quantile plots. Plant dry mass was analysed on square-root transformed data whereas CAT, C:N ratio, and RGR were analysed on log10 transformed data, as they did not meet the assumptions of normality. Foliar Si was analysed using two-way analysis of variance (ANOVA) type = II, comparing CO_2 , and herbivory (larval fed vs. undamaged controls) as treatments and their interaction. For foliar Si analyses, control (–Si) plants were omitted since –Si plants had Si concentrations lower than the machine detection limits (Reidinger et al., 2012). Plant dry mass, antioxidant enzyme activities (CAT, APX, and SOD), foliar C, N, and C-to-N ratio concentrations were all analysed using three-way ANOVA type = II, comparing CO_2 , Si (Si-supplemented vs. non-Si-supplemented plants) and herbivory as treatments and their interaction. Additionally, we tested the independent effects of CO_2 on antioxidant enzyme using a one-way ANCOVA, with CO_2 levels as a fixed factor and foliar Si concentration fitted as a covariate. For herbivore RGR and frass produced, three insects escaped, so data were analysed using type = III ANOVAs due to the unbalanced design. Bonferroni *post hoc* test (Aslam and Albassam, 2020) was applied for pairwise multiple comparisons when interaction terms were statistically significant. Potential relationships between foliar Si and herbivore RGR, frass produced, CAT, APX, and SOD enzymes activity were investigated using Spearman's rank correlation test.

Results

Plant dry mass and foliar chemistry

Averaged across CO_2 treatments, Si supply increased plant dry mass by 160% relative to those grown without Si supply, whereas elevated CO_2 increased plant dry mass by twofold and threefold compared to reduced CO_2 and ambient CO_2 , respectively (Figure 2A; Table 1). Furthermore, herbivory decreased plant dry mass in Si-free (control) plants by 1.5-, 2-, and 2-fold under reduced, ambient, and elevated CO_2 , respectively, compared to Si-supplemented plants (Figure 2A). Si supplementation decreased foliar C concentrations under all CO_2 regimes. This effect was reversed when herbivores were present and foliar C concentrations increased to levels observed in Si-free plants (Figure 2B; Table 1). Si supply decreased foliar C by 149%, 177%, and 107% under reduced, ambient, and elevated CO₂, respectively, regardless of herbivore treatments (Figure 2B). Reduced CO₂ significantly decreased foliar C (Figure 2B; Table 1). Si supply decreased foliar N under all CO₂ regimes. However, there was also an effect of CO₂ whereby reduced CO₂ increased foliar N by twofold and threefold compared to ambient and elevated CO₂, respectively, irrespective of herbivore treatments (Figure 2C; Table 1). In addition to variations in C and N concentrations, there was also an effect on their ratio. Si supply increased foliar C:N ratio especially when plants were damaged by herbivores or in herbivore-free plants only under elevated CO₂ (Figure 2D; Table 1). While herbivory increased foliar C:N ratio regardless of CO₂ levels, reduced CO₂ decreased foliar C:N ratio relative to elevated CO₂ (Figure 2D; Table 1). Foliar Si accumulation [% dry weight (DW)] was significantly higher under reduced CO₂ relative to ambient and elevated CO₂ (Figure 3; Table 1).

Antioxidant enzyme activity was enhanced by Si uptake and reduced CO₂

CAT activity increased in response to Si supply and herbivore damage overall, although this was only apparent under reduced CO_2 concentrations (Table 1; Figure 4A). The significant interaction between Si and CO_2 reflects that Si impacts were only apparent under reduced CO_2 concentrations (Table 1). In contrast, Si supply increased APX enzymes activity under all CO_2 regimes and SOD enzyme activity under reduced and ambient CO_2 (Table 1; Figures 4B, C). Herbivory caused higher APX activity specifically in Si-supplemented plants (Table 1; Figure 4B); however, it had no significant effect on SOD enzyme activity (Table 1; Figure 4C). Overall, antioxidant enzyme activity (CAT, APX, and SOD) declined with increasing CO_2 concentrations (Table 1;



FIGURE 2

Effects of Si supply (+Si or –Si) and herbivory treatments (+H or –H) on (A) plant dry mass, (B) foliar C, (C) foliar N, and (D) foliar C:N of tall fescue grass grown under reduced, ambient, and elevated CO_2 concentration. Means \pm SE shown. N = 6-8. Uppercase letters represent differences between CO_2 concentrations whereas lowercase letters indicate differences between Si treatments (Panel 1A) and herbivore treatments (Panel 1C). Statistically significant effects are indicated *p < 0.05, and ***p < 0.001.

TABLE 1 Results of ANOVA for plant biomass, foliar chemistry, and antioxidant enzyme as affected by CO₂ levels, Si treatment, and herbivore presence and their interactive effects.

| Response | Figures | Factors | | | | | | |
|------------------------------|---------|---------------------------------|---------------------------------|---------------------------------|---------------------------------|---------------------------------|---------------------------------|-------------------------------------|
| variables | | CO ₂ | Si | Herbivory | CO ₂ × Si | CO ₂ × Herbivory | Si × Herbivory | CO ₂ × Si × Herbivory |
| Biomass and foliar chemistry | | | | | | | | |
| Biomass ^z | 1A | $F_{2,60} = 153.1$ p < 0.001 | $F_{1,60} = 44.61$ p < 0.001 | $F_{1,60} = 14.23$ p < 0.001 | $F_{2,60} = 5.761$ p = 0.005 | $F_{2,54} = 0.545$ p = 0.582 | $F_{1,60} = 0.489$ p = 0.487 | $F_{2,60} = 1.043$ p = 0.359 |
| C ^x | 1B | $F_{2,84} = 41.34$ p < 0.001 | $F_{1,84} = 10.54$ p = 0.002 | $F_{1,84} = 145.5$ p < 0.001 | $F_{2,84} = 0.211$ p = 0.801 | $F_{2,84} = 0.515$ p = 0.599 | $F_{1,84} = 0.062$ p = 0.804 | $F_{2,84} = 0.409$ p = 0.666 |
| N ^x | 1C | $F_{2,84} = 54.73$ p < 0.001 | $F_{1,84} = 7.973$ p = 0.006 | $F_{1,84} = 123.1$ p < 0.001 | $F_{2,84} = 0.072$ p = 0.930 | $F_{2,84} = 4.151$ p = 0.019 | $F_{1,84} = 0.645$ p = 0.424 | $F_{2,84} = 1.856$ p = 0.163 |
| C:N ^z | 1D | $F_{2,84} = 62.33$ p < 0.001 | $F_{1,84} = 7.512$ p = 0.007 | $F_{1,84} = 154.7$ p < 0.001 | $F_{2,84} = 0.102$ p = 0.903 | $F_{2,84} = 0.146$ p = 0.864 | $F_{1,84} = 1.818$ p = 0.181 | $F_{2,84} = 1.462$ p = 0.237 |
| Si ^a | 2 | $F_{2,54} = 19.31$ p < 0.001 | | $F_{1,54} = 4.267$ p = 0.044 | | $F_{2,54} = 0.949$ p = 0.393 | | |
| Antioxidant enzyme activity | | | | | | | | |
| CAT ^y | 3A | $F_{2,45} = 19.19$ p < 0.001 | $F_{1,45} = 5.672$ p = 0.022 | $F_{1,45} = 4.290$ p = 0.044 | $F_{2,45} = 3.843$ p = 0.029 | $F_{2,45} = 0.649$ p = 0.528 | $F_{1,45} = 0.000$ p = 0.989 | $F_{2,45} = 0.007$ p = 0.993 |
| APX ^x | 3B | $F_{2,34} = 13.69$ p < 0.001 | $F_{1,34} = 33.36$ p < 0.001 | $F_{1,34} = 20.57$ p < 0.001 | $F_{2,34} = 1.218$ p = 0.308 | $F_{2,34} = 0.199$ p = 0.820 | $F_{1,34} = 0.054$ p = 0.818 | $F_{2,34} = 0.066$ p = 0.936 |
| SOD ^x | 3C | $F_{2,45} = 7.489$ p = 0.002 | $F_{1,45} = 8.781$ p = 0.005 | $F_{1,45} = 2.047$ p = 0.159 | $F_{2,45} = 0.061$ p = 0.941 | $F_{2,45} = 0.018$ p = 0.983 | $F_{1,45} = 0.109$ p = 0.743 | $F_{2,45} = 0.094$ p = 0.910 |
| Herbivore performance | | | | | | | | |
| RGR ^a | 5A | $F_{2,39} = 6.067$ p = 0.005 | $F_{1,39} = 10.03$ p = 0.003 | | $F_{2,39} = 0.224$ p = 0.801 | | | |
| Frass ^a | 5B | $F_{2,39} = 2.510$ p = 0.094 | $F_{1,39} = 4.825$ p = 0.034 | | $F_{2,39} = 0.635$ p = 0.536 | | | |

^aAnalysed using a two-way ANOVA.

^xAnalysed using a three-way ANOVA.

^yAnalysed using a three-way ANOVA on square-root transformed data.

^zAnalysed using a three-way ANOVA on log10 transformed data.

Statistically significant factors at p-values <0.05 are indicated in bold.

Figures 3A–C). Including foliar Si as a covariate in ANCOVA indicated that the changes in antioxidant enzyme activity were linked to CO₂ levels, which fully explained the observed changes in CAT ($F_{1,53} = 11.67$, p = 0.001) and APX ($F_{1,42} = 5.79$, p = 0.021) but not in SOD ($F_{1,53} = 1.130$, p = 0.258). There was a positive correlation between foliar Si concentrations and concentration of CAT under reduced CO₂, and concentration of APX under elevated CO₂ (Figures 5A, B). Interestingly, frass produced was negatively correlated with SOD (r = -0.310, p = 0.038), but had marginally non-significant negative correlation with APX (r = -0.286, p = 0.057). However, there was no such relationship observed between CAT and frass produced (r = -0.191, p = 0.208) (data not shown).

Si supply and low CO₂ environment suppressed herbivore RGR and feeding efficiency

Si supplementation decreased RGR under all CO₂ regimes; RGR was significantly lower under reduced CO₂ compared to elevated

 CO_2 (Table 1; Figure 6A). Si supply decreased the amount of frass produced by caterpillars (indicative of feeding efficiency) under all CO_2 levels; CO_2 had no significant effect on frass production, although there was a large increase in production in Si-free plants grown under elevated CO_2 (Table 1; Figure 6B). While herbivore RGR and frass produced were negatively correlated with foliar Si under ambient CO_2 , there was no such relationship observed under the other two CO_2 regimes (Figures 6C, D). Here, the correlation between rate of herbivore feeding on foliar tissues of tall fescue grown under different CO_2 concentrations potentially indicates a new perspective towards mitigating challenges of CO_2 enriched environment on plant defences.

Discussion

We demonstrated that reduced levels of atmospheric CO_2 caused plants to accumulate more Si and produce higher levels of antioxidant enzyme relative to future levels of atmospheric CO_2 . These increased levels of Si and antioxidant enzyme concentrations



under reduced levels of CO2 were associated with reduced insect herbivore performance. In contrast, herbivore performance and plant consumption (frass production as proxy) were highest under elevated atmospheric CO2 conditions, which typically had the lowest levels of Si and antioxidant enzyme. To our knowledge, this is the first study to address the relationship between Si defences and antioxidant enzyme production in the context of variable atmospheric CO₂ conditions, which we summarise in Figure 7.

Direct and Si-mediated impacts of CO₂ on the activity of antioxidant enzyme

The effects of CO₂ in this study on Si concentrations are broadly similar to the few studies exploring this, whereby elevated CO₂ leads to decreased Si accumulation (Ryalls et al., 2017; Johnson and Hartley, 2018; Biru et al., 2022; Johnson et al., 2022; Johnson et al., 2023), whereas reduced CO₂ leads to increased Si accumulation. To



whereas lowercase letters indicate differences between Si treatments (Panel 1A). Statistically significant effects are indicated *p < 0.05, and ***p < 0.001



the best of our knowledge, the current study is only the third study to investigate the latter (Biru et al., 2020; Biru et al., 2021). Si has been hypothesised to act as a structural substitute for C at a lower metabolic cost, particularly when CO_2 concentrations were lower in the Miocene (Craine, 2009).

Exogenous application of Si has been shown to increase antioxidant enzyme including CAT, APX, and SOD (Kim et al., 2016; Hasanuzzaman et al., 2018; Ahanger et al., 2020). It seems likely that changes in Si in response to CO_2 growing conditions influenced activity of antioxidant enzyme in the current study. Additionally, the ANCOVA results indicated that CO_2 affects antioxidant enzyme activity (e.g., CAT and APX), which were mostly explained by the direct impacts of CO_2 levels on antioxidant enzyme. However, the observed positive correlation of leaf Si with CAT and APX under reduced CO_2 and elevated CO_2 , respectively, suggests that CAT defence response may be linked to higher levels of Si under reduced CO_2 , whereas APX defence response may be associated with increased induction of defence responses following herbivore attack under elevated CO₂. Overall, our results demonstrated significant augmentation of antioxidant enzyme responses via increased Si uptake under reduced CO2 as well as by the direct effect of CO₂ levels (see Figure 7). In contrast, previous studies have reported that elevated CO2 increases activity of antioxidant enzyme in different plants (Wang et al., 2003; Moghimifam et al., 2020). For example, Moghimifam et al. (2020) found that elevated CO₂ enhances CAT, polyphenol oxidase (PPO), SOD, and proline activity in algae (Dunaliella sp.). Our result may reflect that reduced CO₂ often increases photorespiration (Moroney et al., 2013; Voss et al., 2013) and since photorespiration is the key source for ROS production (Voss et al., 2013), reduced CO₂ may cause increased antioxidant enzyme activity in order to scavenge excessive ROS produced. However, these studies did not address whether Si played a role in these changes, while our findings suggest Si as an important influencer of the activity of these enzymes.



Effects of Si supply (+Si or -Si) and CO₂ level on (A) relative growth rate (RGR) of *Helicoverpa armigera* fed on tall fescue grass, (B) frass produced, and the relationship between foliar Si concentrations and (C) RGR and (D) frass produced. Means \pm SE shown. N = 7-8. For (C) and (D), the solid line represents linear regression through all data points and dashed lines indicate that no significant relationship was observed. Statistically significant effects are indicated *p < 0.05.



(A) Current knowledge of the interaction effect between Si and antioxidant enzyme (AOE) activity on insect herbivory. Our key results from this study are indicated in panels (**B**, **C**). (**B**) Reduced CO₂ enhances AOE defences by (1) directly enhancing AOE activity and (2) indirectly increasing Si uptake, which leads to reduced herbivore performance. (**C**) Elevated CO₂ reduces AOE defences by (3) directly reducing AOE activity and (4) indirectly decreasing Si uptake, which leads to increased herbivore performance. Positive and negative effects for both plants and insects are indicated by blue arrows and red lines, respectively.

Herbivory and Si enhanced antioxidant enzyme activity

In addition to CO_2 having direct and Si-mediated impacts on antioxidant enzyme activity, it is also possible that the amount of herbivore damage played a role in antioxidant enzyme activity. Herbivores induced higher activity of CAT and APX, so these enzymes activity is at least associated with the levels of damage done to the plant, which has been similarly reported in previous studies (Leitner et al., 2005; Yang et al., 2017). Increased levels of antioxidant enzyme under reduced CO_2 may reflect the fact that insects were doing less damage to the plant under these conditions and, therefore, there was less ROS oxidative stress that may have persisted in plant tissues to react with higher levels of antioxidant enzyme. Results from the ANCOVA and correlation tests also revealed that CO_2 directly changed antioxidant enzyme activity and altered feeding efficiency (leaf consumption), which eventually reduced the growth rate of herbivores.

Diminishing Si defence with rising CO₂

The recent finding by Biru et al. (2021) showed that *H. armigera* RGR was lowest when fed on the model grass *Brachypodium distachyon* grown under reduced CO_2 due to these plants having higher levels of Si defences compared to plants grown under ambient and elevated CO_2 concentrations. In the current study, we observed that tall fescue had the highest concentrations of foliar N when grown under reduced CO_2 , which, in theory, could have promoted herbivore RGR because N is frequently the limiting factor in insect herbivore diets (Mattson, 1980; Huberty and Denno, 2006). The lower production of frass under reduced CO_2 , which we used as a proxy for plant consumption, suggests that herbivores were deterred from feeding; thus, they would have not been able to access these N resources.

Understanding the diminishing levels of Si-based plant defences against herbivores under future elevated atmospheric CO₂ concentrations has received limited attention. Previous studies have shown that elevated CO2 concentrations decrease Si accumulation in different Poaceae genera (e.g., grass species and wheat) (Johnson and Hartley, 2018; Biru et al., 2022; Johnson et al., 2022), and this was associated with reduced Si defences while increasing herbivore performance; however, this effect is not always reproducible (Frew et al., 2017). The impact of elevated CO2 on Si defences reflects that plants switch from Si defences to Cbased defences due to higher C availability under this scenario (Johnson and Hartley, 2018; Johnson et al., 2022). Although not examined in the context of Si defences, previous studies have demonstrated that elevated atmospheric CO₂ increased consumption and growth rate of the generalist (Pseudaletia unipuncta) and specialist (Spodoptera frugiperda) insect herbivores when fed on C₃ grass relative to lower atmospheric CO₂ conditions (Barbehenn et al., 2004). Johnson et al. (2020) also reported that H. armigera RGR increased under elevated CO2 as a result of lower plant defence signalling and minimal reductions in the nutritional quality of lucerne (Medicago sativa).

Conclusions

The present study provides further evidence that CO_2 concentrations are strong drivers of Si accumulation in an important plant species, not previously reported on. We found strong evidence that reduced CO_2 increased foliar Si concentration and antioxidant enzyme levels, which potentially linked to suppressed insect herbivore performance. This suggests that the negative effects of silicification, whether via physical or biochemical mechanisms, are stronger under reduced CO_2 . We showed a strong linkage between Si supplementation and activity of antioxidant enzyme, which may help in alleviating the harmful effects of

herbivore-induced oxidative stress on plant defence responses. Although Si defences are minimal under elevated atmospheric CO_2 conditions, many agricultural soils can become deficient in bioavailable Si (Haynes, 2017), which points to the importance of maintaining Si levels in soils under future projected atmospheric CO_2 conditions.

Data availability statement

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation.

Author contributions

FB: Conceptualization, Data curation, Formal Analysis, Methodology, Validation, Writing – original draft. CC: Supervision, Writing – review & editing. RE: Supervision, Writing – review & editing. SJ: Conceptualization, Supervision, Writing – review & editing.

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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