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Clinical Performance of Reverse Transcription Loop-mediated Isothermal Amplification COVID-19 Assay on Gold-nanoparticle-modified Screen-printed Carbon Electrode Using Differential Pulse Voltammetry
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Abstract

The World Health Organization (WHO) has recommended real-time reverse transcription polymerase chain reaction (RT-PCR) as the gold standard for coronavirus disease detection. In this study, we aim to validate the clinical performance of reverse transcription loop-mediated isothermal amplification (RT-LAMP) assay on a gold-nanoparticle-modified screen-printed carbon electrode (AuNP/SPCE) using differential pulse voltammetry (DPV) and to compare it with real-time RT-PCR. The electrodeposited AuNP on SPCE was quasi-spherical with a size of ± 500 nm. The developed RT-LAMP primer was designed from the GenBank database using the NCBI Multiple Alignment tools and Jalview software. Nasopharyngeal clinical samples were obtained from suspected COVID-19 patients ($n = 148$). The RT-LAMP products were dropped on the modified AuNP/SPCE under DPV setting, which resulted in current change (ΔI) responses. The positive and negative samples produced significantly different ΔI signals with a p-value < 0.0001 at a 95% confidence interval using Student's t-test. The RT-LAMP assay using Au/SPCE exhibited a 30 s response time per analysis. The clinical sensitivity and specificity obtained were 79.7 and 85.1%, respectively, with a detection limit of 0.4 copies μl^{-1} . Hence, this proposed method is suitable for COVID-19 RNA detection in resource-limited settings. © 2023 M Y U Scientific Publishing Division. All rights reserved.

Author Keywords

AuNP/SPCE; COVID-19; differential pulse voltammetry; electrochemical sensor; RT-LAMP; RT-PCR

Index Keywords

Carbon, Electrochemical electrodes, Electrochemical sensors, Gold nanoparticles, Isotherms, Metal nanoparticles, Polymerase chain reaction, Voltammetry; AuNP/SPCE, Clinical performance, Differential pulse voltammetry, Gold nanoparticle, Gold Nanoparticles, Loop mediated isothermal amplifications, Loop-mediated isothermal amplifications, Reverse transcription, Reverse transcription loop-mediated isothermal amplification, Reverse transcription-polymerase chain reaction; COVID-19

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