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1	Myelination-Independent Functions of Oligodendrocyte Precursor Cells in
2	Health and Disease
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## 16 Abstract

17 Oligodendrocyte precursor cells (OPCs) are a population of tissue resident glial cells found 18 throughout the central nervous system (CNS), constituting approximately 5% of all CNS cells 19 and persisting from development to adulthood and ageing. The canonical role of OPCs is to 20 give rise to myelinating oligodendrocytes. However, additional functions of OPCs beyond 21 this traditional role as precursors have been suggested for a long time. In this Perspective, 22 we provide an overview of the multiple myelination-independent functions that have been 23 described for OPCs in the context of neuron development, angiogenesis, inflammatory 24 response, axon regeneration, and their recently discovered roles in neural circuit 25 remodelling.

26

#### 27 Main text

28 It is now increasingly appreciated that almost every aspect of central nervous system (CNS) 29 formation and function is critically regulated by glial cells <sup>1</sup>. Glia is used as an umbrella term 30 that encompasses astrocytes, microglia, and oligodendrocytes, each of which have 31 specialised functions in the CNS. Oligodendrocytes myelinate axons to facilitate conduction 32 and provide metabolic support to axons <sup>2</sup>. Myelination by oligodendrocytes is a dynamic 33 process that occurs over extended periods of life<sup>3</sup>, and it can change in response to 34 experience and neuronal activity; a process referred to as adaptive myelination <sup>4,5</sup>. New myelin is generally not formed by existing oligodendrocytes, but by differentiation of 35 specialised oligodendrocyte precursor cells (OPCs, Figure 1a)<sup>6</sup>. Interestingly, unlike other 36 37 undifferentiated stem and progenitor cells, OPCs neither reside in niches nor are they 38 transient cells. Instead, OPCs are evenly distributed throughout the CNS (comprising ~5% of 39 all cells in the adult mouse), thereby tiling the tissue with their process networks (Figure 1b, 40 c) 7-11. As OPCs reside within the tissue, they constantly integrate neural activity expressing 41 a broad range of neurotransmitter receptors and voltage-gated ion channels <sup>12</sup>. Furthermore, 42 OPCs are uniquely different from other glia in that they can form post-synaptic contacts with 43 neurons, allowing them to integrate neuronal activity with high spatial and temporal precision (Figure 1d) <sup>13,14</sup>. Together, these properties make OPCs a constant cell population that is 44 45 tightly integrated into neuronal networks, challenging the view that giving rise to myelinating 46 oligodendrocytes is their only function, and raising the question as to whether tissue resident 47 OPCs should be regarded a separate glial cell type (Box 1).

48

Despite the enduring question of what other roles OPCs have beyond the generation of
myelinating oligodendrocytes, definitive answers have remained sparse and somewhat
fragmented, especially in the healthy CNS. This is primarily owed to the circumstance that
experimental manipulation of OPCs frequently also affects myelin generation, which makes it

difficult to dissect primary OPC effects from secondary effects due to altered myelination.
However, a series of recent observations point to specific roles for OPCs in shaping the
development and function of neural circuits. In the light of these recent advances, 40 years
after the discovery of OPCs by Martin Raff and colleagues <sup>15</sup>, it is a prime moment to
summarise what is known about the myelination-independent functions of OPCs in the
healthy and diseased CNS, and to discuss their established properties in the context of their
recently reported roles neural circuit regulation.

60

61 Multiple functions of oligodendrocyte precursors in the healthy and damaged CNS 62 Guidance of migrating neurons. In the developing forebrain, OPCs are generated in 63 successive waves with the first OPCs arising during mid-embryogenesis in the ventral forebrain <sup>16</sup>. Intriguingly, although these ventrally specified OPCs expand and migrate to 64 65 populate the entire telencephalon by the time of birth, they are subsequently replaced by 66 OPCs that are born later in the dorsal cortex <sup>16</sup>. What is the function of these transient 67 precursors, given their limited contribution to the overall populations of OPCs and 68 myelinating oligodendrocytes in the postnatal brain? Recently, an elegant study by 69 Lepiemme and colleagues has shown that ventrally-born OPCs crucially regulate the migration of interneurons into the cortex <sup>17</sup>. Just like ventrally specified OPCs, cortical 70 71 interneurons arise from the medial ganglionic eminence, from where they concomitantly 72 migrate dorsally to the cortex. However, these two cell types do so using different migration 73 strategies. OPCs migrate along blood vessels <sup>18</sup>, whereas interneurons migrate through the 74 parenchyma following a long-range gradient of the chemokine Cxcl12 that is released from 75 the cortex <sup>19</sup>. However, endothelial cells throughout the CNS also release Cxcl12 and it had 76 remained unclear for a long time how interneurons manage to follow the cortical gradient 77 whilst ignoring the endothelial-derived Cxcl12. Lepiemme et al. showed that ventrally born 78 OPCs once associated with vessels, unidirectionally repel migrating interneurons. This

79 contact-driven repulsion allows interneurons to solely follow the Cxcl12 gradient coming from 80 the cortex <sup>17</sup>. It is a common biological phenomenon that the same signalling mechanisms 81 are repurposed in different contexts, but it creates the complication that it needs to be 82 ensured that the same signals coming from different sources do not get misinterpreted by 83 any one cell. The finding that ventrally born OPCs act as mediators to ensure migrating 84 interneurons ignore chemokines coming from the vasculature is a beautiful example of how intercellular crosstalk can achieve correct interpretation of such cues. This functionality also 85 86 demonstrates an important myelination-independent role for transient OPCs to regulate 87 circuit formation by ensuring proper guidance of interneurons.

88

89 Angiogenesis. Crosstalk between OPCs and vasculature does not only regulate the 90 migration of newly formed neurons and OPCs to their target territories. In the developing 91 postnatal brain, OPCs themselves regulate local tissue vascularisation by acting as sensors 92 for hypoxia through activity of OPC-encoded hypoxia-inducible factor (HIF) <sup>20,21</sup>. On the one 93 hand, HIF activation stimulates OPCs proliferation and arrests their maturation to 94 myelinating oligodendrocytes. These autocrine effects are mediated through HIF-induced 95 secretion of Wnt7a/7b by OPCs <sup>20</sup>. At the same time, the same Wnt signalling mechanism 96 has paracrine effects and stimulates vessel growth into the hypoxic tissue where OPCs 97 directly contact sprouting vascular endothelial cells <sup>20,21</sup>. Disruption of hypoxia sensing by 98 OPCs in the forebrain leads to insufficient angiogenesis and axon degeneration, thus 99 highlighting the importance of these bi-directional interactions between OPCs and 100 vasculature to ensure healthy CNS development.

101

Mediator of tissue inflammation. Loss of myelin is a hallmark of demyelinating diseases
 such as Multiple Sclerosis (MS) and CNS injury. Remyelination of demyelinated axons is
 one of the few regenerative processes in the CNS that show relatively high efficiency due to

the lifelong abundance of OPCs which can, in principle, differentiate to become myelinating 105 oligodendrocytes at any time <sup>22</sup>. Upon demyelination, OPCs respond to inflammatory 106 107 cytokines and chemokines released by different immune cells, which affect the remyelination 108 process through regulation of OPC proliferation and differentiation <sup>23</sup>. However, several lines 109 of experimental evidence indicate that OPCs do not only respond to inflammation, but that 110 they actively contribute to driving inflammatory processes. OPCs themselves express 111 immune cues in response to demyelination such as CCL-2 and IL-1ß<sup>24</sup>. In fact, OPCs are 112 crucial for disease progression in experimental autoimmune encephalomyelintis (EAE). EAE 113 pathogenesis is markedly reduced when NFkb activator 1, a key signal transducer of interleukin 17 required for EAE induction, is selectively depleted from OPCs <sup>25</sup>, showing that 114 115 OPCs themselves actively participate in mediating inflammatory processes. More recently, 116 several reports provided further evidence for active roles of OPCs in inflammatory responses. Active MS lesions show aberrant clusters of perivascular OPCs <sup>26</sup>. These OPC 117 118 clusters interfere with the integrity of astrocyte endfeet and tight junctions in mouse models, 119 triggering breakdown of the blood-brain barrier and infiltration of inflammatory cells <sup>26</sup>. 120 Furthermore, single-cell transcriptomics revealed that demyelination primes oligodendrocytes to express immune genes <sup>27</sup>, and induces OPCs to phagocytose and 121 122 present myelin debris via major histocompatibility complex (MHC) class I and MHC-II <sup>28,29</sup>. 123 Together, these works show that tissue resident OPCs not only respond to inflammation, but 124 that they can act as mediators, perpetuators, and even inducers of inflammatory processes 125 in the CNS. 126

127 Glial scar formation and inhibition of axon regeneration. One of the main reasons why 128 severed CNS axons do not efficiently regenerate in mammals is the formation of a glial scar 129 around the lesion site, in addition to the presence of other inhibitors of axon regeneration like 130 myelin debris <sup>30,31</sup>. Following injury, OPCs, as well as astrocytes and microglia, increase their

proliferation rate and migrate to the site of damage where OPCs show upregulation of a 131 broad set of genes associated with inhibition of axonal growth beyond the scar <sup>32,33</sup>. 132 133 Chondroitin sulphate proteoglycans (CSPGs) are one major class of axon regeneration 134 inhibitory molecules. Many CSPGs are prominently expressed by OPCs, such as brevican, 135 neurocan, and phosphacan <sup>34</sup>. The commonly used OPC maker NG2 is also such a 136 proteoglycan (encoded by the *cspg4* gene) and has been known for over 20 years as one of 137 the key inhibitors of axonal regeneration within the glial scar<sup>35</sup>. Since then, a series of 138 studies has successfully devised strategies to enhance axon regeneration in spinal cord injury by either perturbating OPC accumulation within the lesion <sup>36</sup>, or by targeting NG2 139 140 using function-neutralising antibodies to promote regeneration and functional recovery in rat 141 models <sup>37,38</sup>. Besides the expression of inhibitory molecules that prevent axon growth, it has 142 also been suggested that OPCs might entrap dystrophic axons within lesions through the formation of synapse-like contacts <sup>39</sup>, which are known to exist between axons and OPCs in 143 144 the healthy CNS<sup>14</sup>. Even though compelling experimental evidence is lacking, it is tempting 145 to speculate that axon-OPC synapses have stabilizing functions on remodelling axons at the 146 site of transection <sup>39</sup>, similar to how neuronal synapses stabilise axon branch dynamics during development <sup>40,41</sup>. 147

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149 The effects on axon growth and sprouting are not necessarily an exclusive property of OPCs 150 in lesions, even though they may be most pronounced in scars characterised by increased 151 density of reactive OPCs. We should bear in mind that NG2-expressing OPCs equally exist 152 throughout the healthy CNS where they express the same growth inhibitory molecules, even 153 if at lower levels. It may therefore be that OPCs in glial scars formed in response to injury of 154 the CNS secrete additional factors to inhibit axonal regrowth. In addition, OPCs may also 155 serve important functions in providing guidance to axons during development and 156 remodelling of neural circuitry in the healthy CNS, as we discuss in the following sections.

157

# Properties that enable oligodendrocyte precursors to participate in the regulation of neuronal circuitry in the healthy CNS

160 The proper formation of neural circuitry depends on accurate guidance and arborisation of 161 axonal and dendritic arbours, and the establishment of synaptic connections of appropriate 162 number and strength with the right neuronal partners <sup>42</sup>. These processes can be subject to 163 dynamic remodelling as forms of circuit refinement and plasticity. Both formation and 164 refinement of circuit connectivity can be fine-tuned by neuronal activity as one of the driving 165 forces to form, stabilise or enforce some synaptic connections, and to eliminate other synapses <sup>43</sup>. All major types of glia play important roles in regulating different aspects of 166 167 circuitry during development and plasticity. For instance, astrocytes regulate synaptogenesis 168 and neuronal homeostasis, microglia govern synapse pruning, and oligodendrocytes exert activity-dependent myelination <sup>5,44–46</sup>. Interestingly, OPCs are uniquely integrated into neural 169 170 circuits. OPCs express receptors for most neurotransmitters and voltage-gated ion channels, 171 and are accurate sensors for extracellular potassium <sup>12</sup>. Furthermore, OPCs are the only 172 glial cell type that forms synaptic contacts with glutamatergic and GABAergic neurons as 173 pre-synaptic partners <sup>47,48</sup>. The function of these axon:OPC synapses is still unclear, but they 174 allow OPCs to detect quantal release of neurotransmitter with high spatio-temporal 175 resolution to potentially discriminate the precise origins of activity over a broad range of firing 176 frequencies that wouldn't be possible to discriminate with ambient concentrations of 177 neurotransmitter surrounding the cell <sup>13</sup>. OPCs are not only listeners to neuronal networks, 178 they also have the capability to communicate to surrounding cells. Indeed, OPCs can use 179 exocytosis as a mechanism to externalise cargo <sup>49–51</sup>, and even release exosome vesicles <sup>52</sup>. 180 Myelinating oligodendrocytes exhibit exosome release in response to neurotransmitter 181 signalling <sup>53</sup>, which may equally be employed by OPCs to communicate with neurons. Together, these features equip OPCs to monitor neural activity like no other glial cell type, 182

raising the intriguing question of what OPCs do with the information that they receive from the network that they are integrated in, and how OPCs themselves may contribute to the form and function of networks in development and plasticity <sup>54</sup>.

186

187 The first direct evidence showing myelination-independent functions of OPCs in the context 188 of neuronal network function came from ex-vivo studies where it was shown that the NG2 189 proteoglycan is shed from the OPC cell surface in an neural activity-dependent manner, and 190 the inhibition of NG2 shedding impaired long-term potentiation of pyramidal neurons of the 191 somatosensory cortex <sup>55</sup>. This, and several later studies also reported that the manipulation 192 of OPCs and their function results in various behavioural deficits in mice: NG2-deficient mice 193 exhibit altered sensory-motor gating such as reduced pre-pulse inhibition <sup>55</sup>, genetic ablation of OPCs from the prefrontal cortex induces depressive-like behaviours <sup>56</sup>, depletion of 194 GABA<sub>B</sub> receptors from OPCs impairs social cognitive behaviour <sup>51</sup>, and the depletion of 195 196 Kir4.1 potassium channels from OPCs leads to improved spatial memory 57. Although all 197 these studies show that dysfunctional OPCs ultimately impair circuit function and animal 198 behaviour, they are difficult to interpret because it remains unclear which of these defects 199 result from myelination independent OPC functions, and which ones result from secondary 200 effects of impaired oligodendrogenesis and myelination by dysfunctional OPCs.

201

To test if OPCs directly regulate circuit form and function *in vivo*, it requires the design of studies where OPCs can be specifically manipulated without indirectly interfering with myelination. One possibility to achieve this specificity is to manipulate OPCs in CNS areas where they do not (or rarely) differentiate to myelinate axons during stages when functional circuits are present. Examples of such regions are the mouse barrel cortex where OPCs accumulate along the septa separating the barrels <sup>58</sup>, the molecular layer of the cerebellum <sup>59</sup>, as well as in the olfactory bulb where OPCs extend their processes into synaptic glomeruli <sup>60</sup>. However, the absence of reagents and assays to selectively interfere with
OPCs in these regions *in vivo* have thus far limited the investigation of region-specific OPC
functions in mammalian models.

212

## 213 Oligodendrocyte precursors directly sculpt circuit structure and function in the visual

214 system. In 2022, we published a study showing that OPCs regulate fine-tuning of neural circuitry in the visual system through regulation of axonal remodelling <sup>61</sup>. They used the optic 215 216 tectum of young zebrafish as model, as this brain region meets all criteria required to 217 specifically disentangle myelination-independent functions of OPCs. The optic tectum is the 218 primary visual processing centre of zebrafish (equivalent to the lateral geniculate nucleus in 219 mammals) and the site where retinal ganglion cell (RGC) axons form terminal arbours that 220 synapse to dendrites of tectal neurons. We showed that this synaptic region is interspersed 221 with OPCs that interact with surrounding axons and dendrites, but which do not differentiate 222 to myelinating oligodendrocytes, allowing them to study OPC functions without indirectly 223 interfering with myelination. Importantly, during these stages the zebrafish retinotectal 224 system is a functional circuit capable of processing complex sensory-motor transformations 225 <sup>62,63</sup>. We found using genetic global and specific local elimination of OPCs from the optic 226 tectum that RGC arbours showed exuberant sprouting and altered remodelling when OPCs 227 were conditionally eliminated during phases when visual processing is refined. Functionally, 228 these manipulations degraded the acuity of visual processing, meaning that the OPC-229 mediated effects on RGC arbour remodelling impaired synaptic connectivity in the retino-230 tectal system <sup>61</sup>. To our knowledge, this was the first study showing unambiguously that 231 OPCs have mature functions in regulating neural circuit function independently of their 232 canonical roles of generating myelinating oligodendrocytes, which opens a plethora of 233 questions. Do axon:OPC synapses and activity integration play a role in regulating axon 234 growth and remodelling, given that these processes can be controlled by neural activity <sup>64</sup>?

Are these constitutive effects of OPCs that can affect all axons regardless of their identity or is there specificity between subpopulations of axons and OPCs? It is known that specificity of circuit connectivity is governed by selective expression of matchmaking molecules between neurons with over 30 classes of retinal ganglion cell axons <sup>65,66</sup>, and that OPCs also express many of these matchmaking molecules <sup>67</sup>. Furthermore, what are the cellular morphogenic processes that govern OPC-mediated axon remodelling?

241

242 Do OPCs guide or prune axons to regulate circuit connectivity? Overshooting axonal 243 sprouting and faulty synapse formation in the absence of OPCs can occur by two 244 mechanisms which do not need to be mutually exclusive. Lack of inhibition/stabilisation of 245 axon arbours resulting in exuberant sprouting, and/or lack of active removal of surplus 246 connections through phagocytosis. Two recent studies have reported the presence of pre-247 synaptic axonal material within OPCs, suggesting that OPCs actively prune synapses 248 through phagocytosis <sup>68,69</sup>. Three-dimensional electron microscopy reconstructions in the 249 developing mouse visual cortex found that OPCs frequently surround fine axonal filopodia, 250 and that OPCs themselves contain a high number of phagolysosomes and fragments of 251 axons, indicating that they prune synaptic components <sup>69</sup>. Similarly, a light-microscopic study 252 revealed pre-synapses of thalamocortical axons within OPCs of the visual cortex, and the 253 amount synaptic material within OPCs was dependent on the degree of synaptic remodelling 254 during eye opening 68. Thus, OPCs have the capacity to ingest axons, which may mean that 255 they actively prune neuronal circuits. In this case, it will be important to elucidate the 256 difference between synapse pruning by OPCs and by microglia, which are thought to be the 257 main phagocytic cells in circuit plasticity <sup>45</sup>. Another important consideration to make here is 258 that each phagocytosed pre-synapse may have been the pre-synaptic partner of an 259 axon:OPC synapse. Such a scenario would also be consistent with the data available to this 260 date, but substantially change the interpretation of the role of the ingested presynaptic

material. In the latter case, the pre-synapses ingested by OPCs would not necessarily affect
number of neuronal synapses and thus circuit connectivity, but rather alter how OPCs are
connected to neurons through their synapse-like connections, whose roles are still entirely
unclear.

265

266 Whether or not OPCs prune circuits though phagocytosis of neuronal synapses, our own 267 work suggests that guidance is at least one mechanism that is in place because they 268 observed that axonal filopodia frequently retract upon contact with OPC processes <sup>61</sup>, similar 269 to their repulsive properties within glial scars and their role in guiding migrating interneurons 270 <sup>17,35</sup>. Indeed, OPCs express a range of molecules beyond proteoglycans that can guide 271 axons. One such example is semaphorin 5a, which is highly enriched in OPCs and which inhibits growth of retinal ganglion cell axons *in vitro*<sup>70</sup>, and synaptogenesis in the 272 hippocampal dentate gyrus *in vivo*<sup>71</sup>. Interestingly, mutations in semaphorin 5a are linked to 273 274 autism in humans 72, which encompasses a range of complex disorders with defects in 275 neural circuitry. It is tempting to speculate if these defects in synapse formation result from 276 altered regulation of axon remodelling by dysfunctional OPCs, which remains to be 277 investigated in future studies.

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## 279 Conclusions and outlook

Although suggested for a long time, it is now clear that tissue resident OPCs have a number of functions in the healthy and diseased CNS that are independent of their canonical roles in developmental, adaptive, and regenerative myelination (Figure 2). Should we stop using the term OPC in order to reflect the circumstance that these cells are more than just a precursor? We think that it is neither necessary nor helpful to completely overhaul the terminology. From the literature to date, all OPCs, NG2 cells, or how else they are known (Box 1), are members of the same overall cell type that expresses the same set of core markers, and they all can in principle give rise to myelinating oligodendrocytes. Intrinsic and
extrinsic factors allow segregation of OPCs into different states and even functionally
different groups that restrict their roles and propensities to differentiate <sup>11,73,74</sup>. However,
despite these potential groupings and the non-canonical roles of this type of glia in the
healthy and diseased CNS, there is no evidence that any of these cells isn't also a precursor
with the potential to give rise to a myelinating oligodendrocyte.

293

294 Irrespective of this opinion on the nomenclature of tissue resident OPCs, the collective 295 findings on how OPCs directly participate in different processes in the healthy and diseased 296 nervous system raise several questions. Are there unifying attributes that collectively 297 describe the non-canonical roles of OPCs independent of their context? We propose that 298 one possibility would be to regard tissue-resident OPCs as sensors of physiological signals, 299 which they integrate to subsequently act as mediators to other cells that locally surround 300 them. Examples are their described roles in sensing hypoxia to stimulate angiogenesis <sup>20</sup>, in acting as antigen presenting cells upon demyelination <sup>28,29</sup>, and in transducing IL-17 301 302 mediated EAE <sup>25</sup>. Along those lines, a very recent study revealed exacerbated microglial reactions following prion infection in the absence of OPCs <sup>75</sup>, further corroborating the idea 303 304 that OPCs act as mediators to surrounding cells.

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In addition to their roles as signal integrators and mediators to non-neuronal cells, OPCs also directly talk back to the axons they interact with, whether through regulation of their remodelling in circuit maturation <sup>61</sup>, ingestion of pre-synapses <sup>68,69</sup>, or direct bi-directional cross talk <sup>51</sup>. It is an imminent open topic to address the role of axon:OPC synapses in this context. Is the ability of OPCs to integrate synaptic and non-synaptic input used to directly shape circuit development and plasticity? One simple possibility is that axon:OPC synapses could act as temporary guideposts or placeholders to stabilise axon terminals in target areas 313 when they are not yet connected to neuronal postsynapses. It is known that synapse formation guides and stabilises growing axons <sup>40,41</sup>. Synaptic contacts to OPCs could 314 315 facilitate these processes just like they have been proposed to entrap dystrophic axons in 316 lesions <sup>39</sup>. Albeit speculative, OPCs could even employ such contacts to help establish 317 functional boundaries within the CNS. Synaptic specificity and matchmaking between 318 individual neurons are regulated by complex combinations of adhesion molecules that each 319 cell expresses 65. It will be interesting to investigate if OPCs express these molecules in a 320 similarly variegated manner as neurons do. This would enable OPCs to participate in these 321 matchmaking processes, which would add a new level of complexity into the diversity of 322 oligodendrocytes and their interactions with surrounding axons. Regardless of the precise 323 mechanisms by which OPCs integrate into and shape circuits, the overall role of non-324 canonical OPC functions in the context of CNS circuit disorders will be an exciting area for 325 future research. For example, it was shown that fifty percent of dysregulated genes in 326 patients who suffered from major depressive disorders were in fact encoded by OPCs 76. 327 How do dysfunctional OPCs contribute to these and other disorders where circuit 328 connectivity and function is disrupted? They may likely lead to faulty myelination especially 329 in areas that show variable degrees of myelination. However, it may also be direct effects of 330 dysfunctional OPCs as they sculpt the nervous system. The continued development of more refined reagents and assays will allow to dissect to these and other questions on the role of 331 332 neuron:OPC crosstalk in the time to come.

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- 342 The authors declare no competing interests.
- 343

## 344 References

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- 346 1. Allen, N. J. & Lyons, D. A. Glia as architects of central nervous system formation and
   347 function. *Science* 362, 181–185 (2018).
- 348 2. Simons, M. & Nave, K.-A. Oligodendrocytes: Myelination and Axonal Support. *Cold Spring*349 *Harbor perspectives in biology* 8, a020479 (2015).
- 350 3. Miller, D. J. *et al.* Prolonged myelination in human neocortical evolution. *PNAS* 109, 16480–16485 (2012).
- 4. Mount, C. W. & Monje, M. Wrapped to Adapt: Experience-Dependent Myelination. *Neuron* **95**, 743–756 (2017).
- 354 5. Xin, W. & Chan, J. R. Myelin plasticity: sculpting circuits in learning and memory. *Nat Rev* 355 *Neurosci* 21, 682–694 (2020).

6. Czopka, T., ffrench-Constant, C. & Lyons, D. A. Individual oligodendrocytes have only a
few hours in which to generate new myelin sheaths in vivo. *Developmental Cell* 25, 599–609
(2013).

- 7. Dawson, M. R. L., Polito, A., Levine, J. M. & Reynolds, R. NG2-expressing glial progenitor
  cells: an abundant and widespread population of cycling cells in the adult rat CNS. *Mol Cell Neurosci* 24, 476–488 (2003).
- 8. Kirby, B. B. *et al.* In vivo time-lapse imaging shows dynamic oligodendrocyte progenitor
  behavior during zebrafish development. *Nature Neuroscience* 9, 1506–1511 (2006).

9. Hughes, E. G., Kang, S. H., Fukaya, M. & Bergles, D. E. Oligodendrocyte progenitors
balance growth with self-repulsion to achieve homeostasis in the adult brain. *Nature Neuroscience* 16, 668–676 (2013).

- 367 10. Bergles, D. E. & Richardson, W. D. Oligodendrocyte Development and Plasticity. *Csh* 368 *Perspect Biol* 8, a020453 (2015).
- 11. Marisca, R. *et al.* Functionally distinct subgroups of oligodendrocyte precursor cells
  integrate neural activity and execute myelin formation. *Nature Neuroscience* 23, 363–374
  (2020).
- 12. Maldonado, P. P. & Angulo, M. C. Multiple Modes of Communication between Neurons
  and Oligodendrocyte Precursor Cells. *Neurosci* 21, 266–276 (2015).
- 13. Gallo, V., Mangin, J.-M., Kukley, M. & Dietrich, D. Synapses on NG2-expressing
  progenitors in the brain: multiple functions? *The Journal of Physiology* 586, 3767–3781
  (2008).
- 14. Bergles, D. E., Jabs, R. & Steinhäuser, C. Neuron-glia synapses in the brain. *Brain Res Rev* 63, 130–137 (2010).

- 15. Raff, M. C., Miller, R. H. & Noble, M. A glial progenitor cell that develops in vitro into an
  astrocyte or an oligodendrocyte depending on culture medium. *Nature* **303**, 390–396 (1983).
- 381 The discovery of what is now called OPC. Must know!
- 16. Kessaris, N. *et al.* Competing waves of oligodendrocytes in the forebrain and postnatal
   elimination of an embryonic lineage. *Nature Neuroscience* 9, 173–179 (2006).
- 17. Lepiemme, F. *et al.* Oligodendrocyte precursors guide interneuron migration by
- unidirectional contact repulsion. *Science* **376**, eabn6204 (2022).
- 386 **A very nice study study demonstrating a role for transient embryonic OPCs in** 387 **regulating interneuron migration.**
- 18. Tsai, H.-H. *et al.* Oligodendrocyte precursors migrate along vasculature in the developing
   nervous system. *Science* 351, 379–384 (2016).
- 19. Marín, O. Cellular and molecular mechanisms controlling the migration of neocortical
   interneurons. *Eur J Neurosci* 38, 2019–2029 (2013).
- 392 20. Yuen, T. J. *et al.* Oligodendrocyte-encoded HIF function couples postnatal myelination
- and white matter angiogenesis. *Cell* **158**, 383–396 (2014).
- 394 This is the first documentation of OPC-mediated regulation of angiogenesis.
- 21. Chavali, M. *et al.* Wnt-Dependent Oligodendroglial-Endothelial Interactions Regulate
  White Matter Vascularization and Attenuate Injury. *Neuron* **108**, 1130-1145.e5 (2020).
- 22. Franklin, R. J. M. & ffrench-Constant, C. Regenerating CNS myelin from mechanisms
  to experimental medicines. *Nature Reviews Neuroscience* 18, 753–769 (2017).
- 23. Lloyd, A. F. & Miron, V. E. The pro-remyelination properties of microglia in the central
  nervous system. *Nat Rev Neurol* 15, 447–458 (2019).
- 401 24. Moyon, S. *et al.* Demyelination causes adult CNS progenitors to revert to an immature
  402 state and express immune cues that support their migration. *Journal of Neuroscience* 35, 4–
  403 20 (2015).
- 404 25. Kang, Z. *et al.* Act1 mediates IL-17-induced EAE pathogenesis selectively in NG2+ glial 405 cells. *Nature Neuroscience* **16**, 1401–1408 (2013).
- 406 This publication strikingly documents that OPCs are crucial mediators of EAE
   407 progression.
- 408 26. Niu, J. *et al.* Aberrant oligodendroglial-vascular interactions disrupt the blood-brain
  409 barrier, triggering CNS inflammation. *Nature Neuroscience* 22, 709–718 (2019).
- 410 27. Meijer, M. *et al.* Epigenomic priming of immune genes implicates oligodendroglia in
  411 multiple sclerosis susceptibility. *Neuron* **110**, 1193-1210.e13 (2022).
- 412 28. Falcão, A. M. *et al.* Disease-specific oligodendrocyte lineage cells arise in multiple
  413 sclerosis. *Nature Medicine* 24, 1837–1844 (2018).
- 414 29. Kirby, L. *et al.* Oligodendrocyte precursor cells present antigen and are cytotoxic targets 415 in inflammatory demyelination. *Nature communications* **10**, 3887–20 (2019).

- 416 30. Schwab, M. & Caroni, P. Oligodendrocytes and CNS myelin are nonpermissive
- substrates for neurite growth and fibroblast spreading in vitro. *J. Neurosci.* 8, 2381–2393
  (1988).
- 419 31. Zheng, B. & Tuszynski, M. H. Regulation of axonal regeneration after mammalian spinal 420 cord injury. *Nat. Rev. Mol. Cell Biol.* **24**, 396–413 (2023).
- 32. Silver, J. & Miller, J. H. Regeneration beyond the glial scar. *Nature Reviews Neuroscience* 5, 146–156 (2004).
- 33. Adams, K. L. & Gallo, V. The diversity and disparity of the glial scar. *Nature Neuroscience* 21, 9–15 (2018).
- 425 34. Morgenstern, D. A., Asher, R. A. & Fawcett, J. W. Chondroitin sulphate proteoglycans in
  426 the CNS injury response. *Prog Brain Res* 137, 313–332 (2002).
- 35. Tan, A. M., Zhang, W. & Levine, J. M. NG2: a component of the glial scar that inhibits
  axon growth. *J Anat* 207, 717–725 (2005).
- 429 36. Rodriguez, J. P. *et al.* Abrogation of β-catenin signaling in oligodendrocyte precursor 430 cells reduces glial scarring and promotes axon regeneration after CNS injury. *Journal of*
- 431 *Neuroscience* **34**, 10285–10297 (2014).
- 37. Tan, A. M., Colletti, M., Rorai, A. T., Skene, J. H. P. & Levine, J. M. Antibodies against
  the NG2 proteoglycan promote the regeneration of sensory axons within the dorsal columns
  of the spinal cord. *Journal of Neuroscience* 26, 4729–4739 (2006).
- 38. Petrosyan, H. A. *et al.* Neutralization of inhibitory molecule NG2 improves synaptic
  transmission, retrograde transport, and locomotor function after spinal cord injury in adult
  rats. *Journal of Neuroscience* **33**, 4032–4043 (2013).
- 438 39. Filous, A. R. *et al.* Entrapment via Synaptic-Like Connections between NG2
- 439 Proteoglycan+ Cells and Dystrophic Axons in the Lesion Plays a Role in Regeneration
  440 Failure after Spinal Cord Injury. *J Neurosci* 34, 16369–16384 (2014).
- 40. Ruthazer, E. S., Li, J. & Cline, H. T. Stabilization of axon branch dynamics by synaptic 42 maturation. *Journal of Neuroscience* **26**, 3594–3603 (2006).
- 443 41. Meyer, M. P. & Smith, S. J. Evidence from in vivo imaging that synaptogenesis guides
  444 the growth and branching of axonal arbors by two distinct mechanisms. *Journal of*445 *Neuroscience* 26, 3604–3614 (2006).
- 446 42. Weiner, J. A., Jontes, J. D. & Burgess, R. W. Introduction to mechanisms of neural circuit 447 formation. *Front Mol Neurosci* **6**, 12 (2013).
- 448 43. Katz, L. C. & Shatz, C. J. Synaptic activity and the construction of cortical circuits.
  449 *Science* 274, 1133–1138 (1996).
- 44. Chung, W.-S., Allen, N. J. & Eroglu, C. Astrocytes Control Synapse Formation, Function,
  and Elimination. *Csh Perspect Biol* 7, a020370 (2015).

- 452 45. Schafer, D. P. & Stevens, B. Microglia Function in Central Nervous System Development 453 and Plasticity. *Csh Perspect Biol* **7**, a020545 (2015).
- 46. Pan, Y. & Monje, M. Activity Shapes Neural Circuit Form and Function: A Historical
  Perspective. *J Neurosci* 40, 944–954 (2020).

47. Bergles, D. E., Roberts, J. D. B., Somogyl, P. & Jahr, C. E. Glutamatergic synapses on
oligodendrocyte precursor cells in the hippocampus. *Nature* 405, 187–191 (2000). *This seminal study revealed that chemical synapses do not only exist between neurons but also between axons and OPCs.*

- 460 48. Lin, S. & Bergles, D. E. Synaptic signaling between GABAergic interneurons and
  461 oligodendrocyte precursor cells in the hippocampus. *Nature Neuroscience* 7, 24–32 (2004).
- 462 49. Lam, M. *et al.* CNS myelination requires VAMP2/3-mediated membrane expansion in 463 oligodendrocytes. *Nat Commun* **13**, 5583 (2022).
- 464 50. Pan, L. *et al.* Oligodendrocyte-lineage cell exocytosis and L-type prostaglandin D
  465 synthase promote oligodendrocyte development and myelination. *Elife* **12**, e77441 (2023).
- 466 51. Fang, L.-P. *et al.* Impaired bidirectional communication between interneurons and
  467 oligodendrocyte precursor cells affects social cognitive behavior. *Nat Commun* **13**, 1394
  468 (2022).
- 469 52. Goncalves, M. B. *et al.* Regulation of Myelination by Exosome Associated Retinoic Acid
  470 Release from NG2-Positive Cells. *J Neurosci* **39**, 3013–3027 (2019).
- 471 53. Frühbeis, C. *et al.* Neurotransmitter-Triggered Transfer of Exosomes Mediates
  472 Oligodendrocyte–Neuron Communication. *Plos Biol* **11**, e1001604 (2013).
- 473 54. Mangin, J.-M. & Gallo, V. The curious case of NG2 cells: transient trend or game
- 474 changer? *ASN neuro* **3**, e00052 (2011).

## This is an excellent review which should not be overlooked when thinking about the myelination-independent roles of OPCs.

55. Sakry, D. *et al.* Oligodendrocyte precursor cells modulate the neuronal network by

478 activity-dependent ectodomain cleavage of glial NG2. *PLoS Biology* **12**, e1001993 (2014).

- 479 One of the first publications showing that OPCs can directly regulate neuronal
   480 function.
- 481 56. Birey, F. *et al.* Genetic and Stress-Induced Loss of NG2 Glia Triggers Emergence of
  482 Depressive-like Behaviors through Reduced Secretion of FGF2. *Neuron* 88, 941–956
  483 (2015).
- 484 57. Timmermann, A. *et al.* Dysfunction of NG2 glial cells affects neuronal plasticity and 485 behavior. *Glia* (2023) doi:10.1002/glia.24352.
- 58. Mangin, J.-M., Li, P., Scafidi, J. & Gallo, V. Experience-dependent regulation of NG2
  progenitors in the developing barrel cortex. *Nature Neuroscience* 15, 1192–1194 (2012).
- 488 59. Goebbels, S. *et al.* A neuronal PI(3,4,5)P3-dependent program of oligodendrocyte
  489 precursor recruitment and myelination. *Nature Neuroscience* 20, 10–15 (2017).

- 490 60. Rungta, R. L., Chaigneau, E., Osmanski, B.-F. & Charpak, S. Vascular
- 491 Compartmentalization of Functional Hyperemia from the Synapse to the Pia. *Neuron* **99**,
- 492 362-375.e4 (2018).
- 493 61. Xiao, Y., Petrucco, L., Hoodless, L. J., Portugues, R. & Czopka, T. Oligodendrocyte
  494 precursor cells sculpt the visual system by regulating axonal remodeling. *Nat Neurosci* 25,
  495 280–284 (2022).

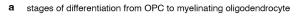
## The first publication that unambiguously shows that OPCs fine-tune assembly and

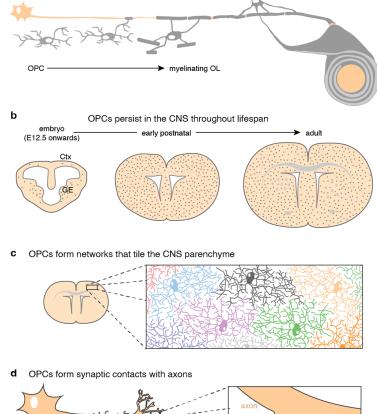
- 497 *function of neural circuits.*
- 498 62. Bollmann, J. H. The Zebrafish Visual System: From Circuits to Behavior. *Annual review*499 *of vision science* 5, 269–293 (2019).
- 500 63. Baier, H. & Wullimann, M. F. Anatomy and function of retinorecipient arborization fields 501 in zebrafish. *J Comp Neurol* **529**, 3454–3476 (2021).
- 64. Hua, J. Y. & Smith, S. J. Neural activity and the dynamics of central nervous system
  development. *Nature Neuroscience* 7, 327–332 (2004).
- 504 65. Sanes, J. R. & Zipursky, S. L. Synaptic Specificity, Recognition Molecules, and 505 Assembly of Neural Circuits. *Cell* **181**, 536–556 (2020).
- 506 66. Kölsch, Y. *et al.* Molecular classification of zebrafish retinal ganglion cells links genes to cell types to behavior. *Neuron* **109**, 645-662.e9 (2020).
- 508 67. Almeida, R. G. & Lyons, D. A. On the resemblance of synapse formation and CNS myelination. *Neuroscience* **276**, 98–108 (2014).
- 510 68. Auguste, Y. S. S. *et al.* Oligodendrocyte precursor cells engulf synapses during circuit 511 remodeling in mice. *Nat Neurosci* **25**, 1273–1278 (2022).
- 512 This article shows that OPCs can phagocytose axonal pre-synapses, and that the
- 513 degree of engulfment by OPCs is altered by sensory experience.
- 514 69. Buchanan, J. *et al.* Oligodendrocyte precursor cells ingest axons in the mouse
- 515 neocortex. *Proc National Acad Sci* **119**, e2202580119 (2022).
- 516 This article presents careful 3D reconstructions of electron microscopic images that
- 517 reveal the presence of axonal material inside OPCs. Together with reference #68 the
- 518 *first documentation that OPCs also have these roles.*
- 519 70. Goldberg, J. L. *et al.* An oligodendrocyte lineage-specific semaphorin, Sema5A, inhibits
  520 axon growth by retinal ganglion cells. *Journal of Neuroscience* 24, 4989–4999 (2004).
- 521 71. Duan, Y. *et al.* Semaphorin 5A inhibits synaptogenesis in early postnatal- and adult-born
  522 hippocampal dentate granule cells. *eLife* 3, (2014).
- 523 72. Weiss, L. A., Arking, D. E., Consortium, G. D. P. of J. H. & the A., Daly, M. J. &
- 524 Chakravarti, A. A genome-wide linkage and association scan reveals novel loci for autism. 525 *Nature* **461**, 802–808 (2009).
- 526 73. Viganò, F., Möbius, W., Götz, M. & Dimou, L. Transplantation reveals regional
- 527 differences in oligodendrocyte differentiation in the adult brain. *Nature Neuroscience* **16**,
- 528 1370–1372 (2013).

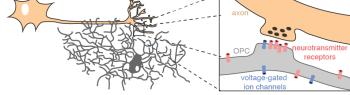
- 529 74. Spitzer, S. O. *et al.* Oligodendrocyte Progenitor Cells Become Regionally Diverse and
  530 Heterogeneous with Age. *Neuron* **101**, 1–13 (2019).
- 531 75. Liu, Y. *et al.* NG2 glia protect against prion neurotoxicity by inhibiting prostaglandin E2 532 signaling. (2023) doi:10.1101/2023.04.04.535590.
- 533 76. Nagy, C. *et al.* Single-nucleus transcriptomics of the prefrontal cortex in major
- 534 depressive disorder implicates oligodendrocyte precursor cells and excitatory neurons. *Nat* 535 *Neurosci* **23**, 771–781 (2020).
- 536 77. Butt, A. M., Hamilton, N., Hubbard, P., Pugh, M. & Ibrahim, M. Synantocytes: the fifth 537 element. *J Anat* **207**, 695–706 (2005).
- 538 78. Nishiyama, A., Komitova, M., Suzuki, R. & Zhu, X. Polydendrocytes (NG2 cells):
  539 multifunctional cells with lineage plasticity. *Nature Reviews Neuroscience* **10**, 9–22 (2009).
- 540 79. Dimou, L. & Gallo, V. NG2-glia and their functions in the central nervous system. *Glia* 63,
  541 1429–1451 (2015).
- 80. Richardson, W. D., Young, K. M., Tripathi, R. B. & McKenzie, I. NG2-glia as Multipotent
  Neural Stem Cells: Fact or Fantasy? *Neuron* 70, 661–673 (2011).
- 81. Nishiyama, A., Boshans, L., Goncalves, C. M., Wegrzyn, J. & Patel, K. D. Lineage, fate,
  and fate potential of NG2-glia. *Brain Res* 1638, 116–128 (2016).
- 82. Zawadzka, M. *et al.* CNS-resident glial progenitor/stem cells produce Schwann cells as
  well as oligodendrocytes during repair of CNS demyelination. *Cell Stem Cell* 6, 578–590
  (2010).
- 83. Emery, B. & Lu, Q. R. Transcriptional and Epigenetic Regulation of Oligodendrocyte
  Development and Myelination in the Central Nervous System. *Cold Spring Harbor perspectives in biology* 7, a020461 (2015).
- 84. Sock, E. & Wegner, M. Using the lineage determinants Olig2 and Sox10 to explore
  transcriptional regulation of oligodendrocyte development. *Dev. Neurobiol.* 81, 892–901
  (2021).
- 555

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## 557 Figures Legends:



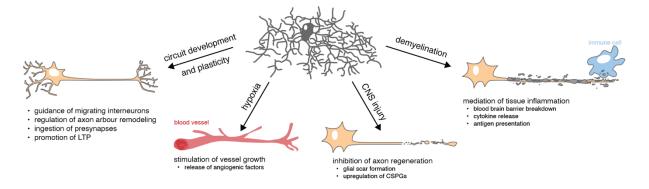




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## 559 Figure 1: Characteristics of oligodendrocyte precursor cells (OPCs)

- **a)** The canonical role for OPCs is to give rise to myelinating oligodendrocytes in health and
- disease. OPCs have highly branched processes which they iteratively wrap around axons toform myelin sheaths during their differentiation.
- 563 b) OPCs are specified in discrete CNS regions (in the mouse telencephalon starting from the
- 564 ganglionic eminence (GE) between embryonic day 12.5-15, and from the dorsal cortex (Ctx)
- 565 from around birth), from where they disperse to evenly distribute throughout the CNS and 566 where they persist into adulthood.
- **c)** With the parenchyma, each OPC forms a highly branched process network, with each cell occupying its own territory. OPCs sense their territory and expand into regions not occupied
- by an OPC through migration, growth, or proliferation, resulting in a tiling of the CNS.
- **d)** OPCs closely interact with and receive information from axons throughout development
- and adulthood. They integrate neural activity via neurotransmitter receptors and voltage-
- 572 gated ion channels and, once settled in the parenchyma, can form synaptic contacts with
- 573 neuronal pre-synapses.



575

## **Figure 2: Multifunctional OPCs in the healthy and damaged CNS.** Several non-canonical functions have been attributed to OPCs in different contexts as discussed in this

- Perspective.

#### 580 Box 1: Identity and terminology of oligodendrocyte precursor cells

581 Tissue-resident oligodendrocyte precursor cells (OPCs) represent about 5% of all CNS cells 582 which tile the tissue with their complex process networks 7. They can differentiate to 583 myelinating oligodendrocytes throughout life as part of developmental, adaptive, and 584 regenerative myelination, but OPC numbers are maintained in constant homeostasis 4,5,10,22. 585 Due to the permanent presence of OPCs throughout the CNS it has been speculated for a 586 long time that OPCs are likely more than just a precursor. Hence, over the years OPCs have 587 been given different names to reflect this circumstance, such as synantocytes, polydendrocytes, or simply NG2 glia because all OPCs express the NG2 antigen 77-79. In 588 589 fact, at the time of their first discovery in the early 1980s, OPCs were named O-2A 590 progenitors because they can give rise to oligodendrocytes and type 2 astrocytes in culture 591 <sup>15</sup>. Some refer to OPCs as oligodendrocyte progenitor cells because it has been reported 592 that they can give rise to multiple neural lineages <sup>80,81</sup> (even Schwann cells in disease 593 contexts <sup>82</sup>). However, no matter how these cells are referred to, they all are the same cell 594 type that shares a set of key transcription factors and markers, most importantly the two 595 oligodendrocyte lineage determinants Olig2 and Sox10<sup>83,84</sup>. This gives them the principal 596 ability to give rise to myelinating OLs, even though additional markers may be discovered in 597 the future that distinguish between OPCs that can generate OLs and those that do not, and 598 even though OPCs fulfil a series of additional functions in the healthy and diseased CNS as 599 presented in this article.