



UNIVERSITI PUTRA MALAYSIA

***UTILISATION OF TREATED STRAW FROM NEWLY-DEVELOPED PadiU
PUTRA-1 RICE VARIETY AS FEED SUPPLEMENT FOR SUSTAINABLE
GOAT PRODUCTION***

AHMED MUIDEEN ADEWALE

IPTSM 2022 5



**UTILISATION OF TREATED STRAW FROM NEWLY-DEVELOPED PadiU
PUTRA-1 RICE VARIETY AS FEED SUPPLEMENT FOR SUSTAINABLE
GOAT PRODUCTION**

By

AHMED MUIDEEN ADEWALE

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in
Fulfillment of the Requirements for the Degree of Doctor of Philosophy**

February 2022

COPYRIGHT

All material contained within the thesis including, without limitation, text, logos, icons, photographs, and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express prior written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



DEDICATIONS

This Thesis Is Dedicated to My Father, Mother and My Siblings with Love, Humility
and Respect



© COPYRIGHT UPM

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Doctor of Philosophy

UTILISATION OF TREATED STRAW FROM NEWLY-DEVELOPED PadiU PUTRA-1 RICE VARIETY AS FEED SUPPLEMENT FOR A SUSTAINABLE GOAT PRODUCTION

By

AHMED MUIDEEN ADEWALE

February 2022

Chairman : Professor Awis Qurni bin Sazili, PhD
Institute : Tropical Agriculture and Food Security

The persistent challenges of meeting the global population's demand for animal protein in the ruminant industry, centred on feed to boost production. To bridge the niche areas for the realisation of food security, the availability of feed that is cheap, noncompetitive, qualitative, and sustainable is required. Agricultural waste, such as rice straw that is illicitly disposed of, causing health impairment and global environmental nuisance through burning could be improved by biological additive and used as ruminant feed. However, biological additives have been reported to improve straw quality through ensiling and possibly animal performance. As a result, the objective of the study refers to the use of enriched straw from a newly developed rice variety as a feed supplement for a sustainable goat production. This thesis comprised of three research chapters.

The aim of the first experiment was to examine the efficacy of biological additives on nutrient profiles, ensile quality, *in vitro* rumen fermentation in different varieties of treated rice straw; ML4, ML10, ML21, ML24, MR219 and PadiU Putra-1 were ensiled with water (Control), enzyme (E) and combination of bacteria and enzyme (BE) in a complete randomise design (CRD) experiment. The rice straw varieties, obtained from the paddy research farm were chopped to about 2-3 cm, treated according to the above treatments to attain 70% moisture content, compressed in a 1 L jar and ensiled for 30 days. Results showed that treatment with the combination of bacteria and enzyme had lower neutral detergent fiber (42.31%) and lower acid detergent fiber (21.08%) for PadiU Putra-1 than other treatments (enzyme and control) and varieties. For the ensiled extract, the lowest pH, NH₃, butyric acid and propionic acid were obtained in BE treatment in most of the rice straw varieties. In addition, straw treated with BE had significantly higher lactic acid content, *in vitro* gas production kinetics, digestibility, and rumen fermentation compared to the enzyme and control. This study revealed that a combination of bacterial and enzyme treatment effectively improved the quality of treated rice straw, with the PadiU Putra-1 variety being the most outstanding. Selection

of the appropriate biological treatment and variety of rice straw is important to improve straw quality. However, the treated rice straw needs further evaluation for its *in vivo* efficacy.

The second experiment investigated the effects of PadiU Putra-1 treated rice straw with biological additives on growth performance, nutrient digestibility, rumen fermentation, microbial population, and relative expressions of ruminal MCT1 and growth hormone genes in goats. Twenty-four male crossbred Boer goats were randomly assigned to the following diets: (i) Control (untreated straw); (ii) Enzyme treated straw; and (iii) BE, enzyme plus lactic acid bacteria treated straw. Each diet, in the form of total mixed ration consisted of 50% treated rice straw and 50% concentrate was fed at 3.5% DM body weight of the experimental animals for 14 weeks. The results showed that average daily gain (ADG) of goats was 13% and 26%, respectively, higher in enzyme and BE treatment than the control ($P<0.01$). Feed efficiency (gain:feed) was also enhanced by 8% and 23% in Enzyme and BE treatment diets relative to control. Goats fed treated straws had higher nutrient digestibility leading to higher digestible nutrient intake ($P<0.001$). Dietary treatments had no effect on rumen fermentation except propionate concentrations which were higher in the treated rice straw groups leading to lower acetate to propionate ratio ($P<0.01$). Goats fed treated rice straw had higher cellulolytic bacteria but lower protozoa and methanogens which resulted in lower methane concentration; greater expression of ruminal MCT1 and hepatic GHR, IGF-1 genes ($P<0.01$) indicating better rumen absorption, growth process and nutrient metabolism.

The third research chapter examines the influence of biologically treated rice straw on blood profiles (haematology and biochemistry), non-carcass, carcass characteristics and meat quality in goats. The blood samples from each animal were taken from the jugular vein using vacuum EDTA and serum tube for haematological and biochemical blood profile analysis ascertain the non-detrimental effect of the diet while the meat quality analysis was performed after the slaughtering and the ageing days. The blood profile results indicated dietary treatment had no detrimental effect on the haematological and biochemical profiles, but the days of sampling had significant effect. On the carcass trait, enzyme treatment increased slaughter weight, hot carcass, and cold carcass by 4%, 6%, and 6%, respectively, compared to the control, whereas BE treatment increased the above parameters by 9%, 13%, and 15%, respectively ($P<0.05$). In addition, the dietary treatment improved the chilling loss and dressing percentages, but had no effect on the non-carcass and primal cuts. Dietary treatment influenced the cooking loss percentage and shear force, whereas other physicochemical meat qualities were unaffected but had a post-mortem ageing effect.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**PENGUNAAN JERAMI YANG DIRAWAT DARIPADA PadiU PUTRA-1
YANG BAHARU DIHASILKAN SEBAGAI TAMBAHAN MAKANAN UNTUK
PENGELUARAN KAMBING LESTARI**

Oleh

AHMED MUIDEEN ADEWALE

Februari 2022

Pengerusi : Profesor Awis Qurni bin Sazili, PhD
Institut : Pertanian Tropika dan Sekuriti Makanan

Cabaran berterusan untuk memenuhi permintaan populasi global untuk protein hewani dalam industri ruminan, tertumpu kepada makanan untuk meningkatkan produksi. Untuk menghubungkan kawasan khusus untuk mewujudkan keselamatan makanan, ketersediaan makanan yang murah, tidak kompetitif, kualitatif, dan lestari diperlukan. Sisa pertanian, seperti jerami padi yang dibuang secara haram menyebabkan gangguan kesihatan dan gangguan persekitaran global melalui pembakaran dapat dielakkan dengan bahan tambahan biologi dan digunakan sebagai makanan ruminan. Walau bagaimanapun, bahan tambahan biologi telah dilaporkan dapat meningkatkan kualiti jerami melalui pengawetan hijau dan kemungkinan meningkatkan prestasi haiwan. Hasilnya, objektif kajian merujuk kepada penggunaan jerami yang diperkaya dari varietas padi yang baru dihasilkan sebagai makanan tambahan untuk pengeluaran kambing yang lestari. Tesis ini terdiri daripada tiga bab penyelidikan.

Tujuan bab penyelidikan pertama adalah untuk mengkaji keberkesanan bahan tambahan biologi terhadap profil nutrien, kualiti silase, fermentasi rumen secara in vitro dalam pelbagai jenis silase jerami padi; ML4, ML10, ML21, ML24, MR219 dan PadiU Putra-1 diawetkan dengan air (Kawalan), enzim (E) dan kombinasi bakteria dan enzim (BE) dalam eksperimen reka bentuk rawak (CRD) lengkap. Varieti jerami padi yang diperolehi dari ladang penyelidikan padi dipotong menjadi kira-kira 2-3 cm, dirawat mengikut rawatan seperti di atas untuk mencapai kandungan kelembapan 70%, dimampatkan dalam balang 1 L dan diawet selama 30 hari. Hasil kajian menunjukkan bahawa protein mentah dalam silase jerami padi yang dirawat lebih tinggi daripada kawalan. Rawatan dengan kombinasi bakteria dan enzim mempunyai serat pencuci neutral yang lebih rendah (42,31%) dan serat pencuci asid yang lebih rendah (21,08%) untuk PadiU Putra-1 daripada rawatan (enzim dan kawalan) dan varieti lain. Untuk ekstrak silase, pH terendah, NH₃, asid butir dan asid propionik diperolehi dalam rawatan BE pada kebanyakan varietas jerami padi. Jerami yang dirawat dengan BE memiliki kandungan asid laktat tertinggi, kinetik pengeluaran gas in vitro, pencernaan,

dan fermentasi rumen dibandingkan dengan enzim dan kawalan. Kajian ini mendedahkan bahawa gabungan rawatan bakteria dan enzim berkesan meningkatkan kualiti jerami padi yang dirawat, dengan varieti PadiU Putra-1 adalah yang paling menonjol. Pemilihan rawatan biologi dan varieti jerami padi adalah penting untuk meningkatkan kualiti jerami. Namun, jerami padi yang dirawat memerlukan penilaian lebih lanjut untuk keberkesanan in vivo.

Bab penyelidikan kedua menyelidiki kesan PadiU Putra-1 jerami padi yang dirawat dengan bahan tambahan biologi terhadap prestasi pertumbuhan, pencernaan nutrien, fermentasi rumen, populasi mikrob, dan ekspresi relatif MCT1 secara rumin dan gen hormon pertumbuhan pada kambing. Dua puluh empat ekor kambing Boer jantan dipilih secara rawak dan diberikan diet berikut: (i) Kawal (jerami yang tidak dirawat); (ii) Jerami yang dirawat dengan enzim; dan (iii) BE, enzim ditambah jerami yang dirawat bakteria asid laktik. Setiap diet dalam bentuk jumlah catuan campuran terdiri dari 50% silase jerami padi dan 50% diberi makan dengan berat badan DM sebanyak 3.5% untuk haiwan percubaan selama 14 minggu. Hasil kajian menunjukkan bahawa purata penambahan berat harian kambing adalah masing-masing 13% dan 26%, lebih tinggi enzim dan silage BE daripada kawalan ($P < 0.01$). Nisbah penukaran makanan juga ditingkatkan sebanyak 11% dan 19% dalam diet silase enzyme dan BE berbanding kawalan. Kambing yang diberi makan jerami yang diperlakukan mempunyai pencernaan nutrien yang lebih tinggi yang menyebabkan penyerapan nutrien yang dicerna lebih tinggi ($P < 0.001$). Rawatan makanan tidak dipengaruhi fermentasi rumen kecuali kepekatan propionat yang lebih tinggi pada kumpulan silase yang dirawat menyebabkan nisbah asetat ke propionat lebih rendah ($P < 0.01$). Kambing yang diberi makan silase mempunyai bakteria selulolitik yang lebih tinggi tetapi kandungan protozoa dan metanogen lebih rendah yang mengakibatkan kepekatan metana lebih rendah; ungkapan MCT1 rumin dan GHR hepatic, gen IGF-1 yang lebih besar ($P < 0.01$) menunjukkan penyerapan rumin yang lebih baik dan penggunaan nutrien untuk pengeluaran.

Bab penyelidikan ketiga meneliti pengaruh silase jerami padi yang dirawat secara biologi terhadap profil darah (hematologi dan biokimia), bahagian bukan bangkai, ciri bangkai dan kualiti daging pada kambing. Sampel darah dari setiap haiwan diambil dari urat jugular dengan menggunakan vakum EDTA dan tiub serum untuk analisis profil darah hematologi dan biokimia sementara analisis kualiti daging dilakukan setelah penyembelihan dan hari penuaan. Hasil profil darah menunjukkan rawatan diet tidak memberi kesan buruk pada profil hematologi dan biokimia, tetapi hari-hari pengambilan sampel mempunyai pengaruh yang ketara. Pada sifat bangkai, rawatan enzim meningkatkan berat penyembelihan, bangkai panas, dan bangkai sejuk masing-masing sebanyak 4%, 6%, dan 6%, berbanding dengan rawatan kawalan, sedangkan rawatan BE meningkatkan parameter di atas sebanyak 9%, 13%, dan 15%, masing-masing ($P < 0.05$). Di samping itu, rawatan diet meningkatkan peratusan kehilangan penyejukbekuan dan produksi bangkai, tetapi tidak berpengaruh pada bukan bangkai dan potongan primal. Rawatan makanan mempengaruhi peratusan kehilangan ketika memasak dan daya ricih, manakala kualiti fizikokimia daging yang lain tidak terjejas tetapi mempunyai kesan penuaan selepas kematian.

ACKNOWLEDGEMENTS

First of all, I want to thank Allah (S.W.T) to whom all adoration and praises is due for the successful completion of this work.

My utmost gratitude goes to my supervisor, chairman of my supervisory Committee, Prof. Dr. Awis Qurni Sazili for his patience, understanding, guidance and encouragement throughout the duration of this work in spite of his engagements. I wish to extend my special gratitude to my co-supervisor Dr. Liang Juan Boo, Prof. Dr. Mohd Raffi Yusop and Assoc. Prof. Dr. Nur Ain Izzati Mohd Zainudin for their magnanimous support, suggestion, good advice and germane criticism, may God reward you abundantly.

My special thanks to all members of my family for their love, motivation, encouragement and prayers. My surpassed parents, I remain deeply grateful for giving me the financial support and freedom to pursue my academic dream. I am exceedingly thankful for discomfoting yourself to offer me the best. May Allah in His Infinite Mercy bestow on you long life in good health and happiness to reap from the fruit of your labor (AMIN). You are indeed my all in all. Also my appreciation goes to my siblings (Haleemat, Morufat, Rashidat, my niece and my nephew) and Jaleelah for their persistence patience, prayers and encouragement. I am grateful for your awesome love, concern and encouragement throughout the study period.

I want to sincerely appreciate my colleagues and friends (Dr. Yusuf Oladosu, Dr. Atiq, Dr. Leela Lyana, Dr. Kifah, Dr. Idayu, Mrs. Batool, Dr. Anwar, Dr Suganya, Nadirah, Dr. Osama, Dr. Abubakar Ahmed, Renuh, Stephini, Abubakar, Liyana, Dr. Candy, Nizam, Zaihan, Mr. Muhammed, Wan Rasidah, Dr Pavan Kumar, and Dr. Ismaila Muhammad. Special thanks to the Department of Animal Science in Faculty of Agriculture, Universiti Putra Malaysia, specifically (*In vitro*, Meat Science and Nutrition Laboratory) where most of my research analysis were carried out. To all staff of Institute of Tropical Agriculture and Food Security, UPM thank you very much for making my life as a post graduate student a delightful experience. In addition, huge thanks to all the staff of Animal Research Centre (ITAFoS), Mr Eddy Shahriza, Mr Fazmen Lee, Mr Farhan and Mr Ahmad Zahidi and Mr Faizol from Animal Production Laboratory (ITAFoS) and Mr Khairul from Vaccine and Immunotherapeutics Laboratory (IBS) for their support during my farm work. To all those I couldn't mention who contributed to the success of this study in one way or the other, I say a big thanks to all.

Lastly, special thanks to all those who have contributed to the success of this research directly or indirect. May Almighty God bless you abundantly.

I certify that a Thesis Examination Committee has met on (date of viva voce) to conduct the final examination of Ahmed Muideen Adewale on his thesis entitled “Utilisation of enriched straw from newly developed rice variety as feed supplement for a sustainable goat production” in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U. (A) 106] 15 March 1998. The Committee recommends that the student be awarded the degree of Doctor of Philosophy.

Members of the Thesis Examination Committee were as follows:

Name of Chairperson, PhD

Title (e.g., Professor/Associate Professor/Ir; omit if irrelevant)

Name of Faculty

Universiti Putra Malaysia

(Chairman)

Name of Examiner 1, PhD

Title (e.g., Professor/Associate Professor/Ir; omit if irrelevant)

Name of Faculty

Universiti Putra Malaysia

(Internal Examiner)

Name of Examiner 2, PhD

Title (e.g., Professor/Associate Professor/Ir; omit if irrelevant)

Name of Faculty

Universiti Putra Malaysia

(Internal Examiner)

Name of External Examiner, PhD

Title (e.g., Professor/Associate Professor/Ir; omit if irrelevant)

Name of Department and/or Faculty

Name of Organisation (University/Institute)

Country

(External Examiner)

SITI SALWA ABD GANI, PhD

Associate Professor ChM. and Deputy Dean

School of Graduate Studies

Universiti Putra Malaysia

Date:

This thesis was submitted to the Senate of the Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Awis Qurni bin Sazili, PhD

Professor
Halal Products Research Institute
Universiti Putra Malaysia
(Chairman)

Liang Juan Boo, PhD

Senior Research Fellow
Institute of Tropical Agriculture and Food Security
Universiti Putra Malaysia
(Member)

Mohd Rafii bin Yusop, PhD

Professor
Institute of Tropical Agriculture and Food Security
Universiti Putra Malaysia
(Member)

Nur Ain Izzati binti Mohd Zainudin, PhD

Associate Professor
Faculty of Science
Universiti Putra Malaysia
(Member)

ZALILAH MOHD SHARIFF, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 10 November 2022

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) were adhered to.

Signature: _____
Name of
Chairman of
Supervisory
Committee: Professor Dr. Awis Qurni bin Sazili

Signature: _____
Name of
Member of
Supervisory
Committee: Dr. Liang Juan Boo

Signature: _____
Name of
Member of
Supervisory
Committee: Professor Dr. Mohd Rafii bin Yusop

Signature: _____
Name of
Member of
Supervisory
Committee: Associate Professor Dr. Nur Ain Izzati binti Mohd Zainudin

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xiv
LIST OF FIGURES	xvi
LIST OF ABBREVIATIONS	xvii
CHAPTER	
1 INTRODUCTION	1
2 LITERATURE REVIEW	4
2.1 Global rice and straw distribution	4
2.2 Malaysia rice (PadiU Putra 1) and straw distribution	4
2.3 Biological treatment of straw	7
2.4 Utilisation of biological treated straw as ruminant feed	8
2.5 Global distribution of goats	10
2.6 Malaysia goat population and distribution	11
2.6.1 Challenges faced by the Malaysian small ruminant sector to achieve sustainable production	13
2.6.2 Importance of goats	13
2.6.3 Feed and feeding of ruminant	14
2.7 Rumen ecosystem	14
2.7.1 Rumen bacteria	15
2.7.2 Protozoa	16
2.7.3 Methanogens	16
2.8 Meat quality and factors affecting it	17
2.8.1 pH	17
2.8.2 Colour	18
2.8.3 Water holding capacity	19
2.8.4 Drip loss	20
2.8.5 Cooking loss	20
2.8.6 Tenderness	21
2.9 Gene expression (MCT1) associated with absorption and transportation of short chain fatty acid in ruminant	22
2.10 Summary	23
3 EFFICACY OF BIOLOGICAL ADDITIVES ON NUTRIENT PROFILES, FERMENTATION QUALITIES, AND <i>In Vitro</i> GAS PRODUCTION KINETICS, DIGESTIBILITY AND RUMEN FERMENTATION IN DIFFERENT VARIETIES OF RICE STRAW	25
3.1 Introduction	25
3.2 Materials and Methods	26

3.2.1	Sample collection and preparation	26
3.2.2	Microbial culture and crude enzyme production	26
3.2.3	Nutrient composition and fermentation products determination	28
3.2.4	<i>In vitro</i> gas production	29
3.3	Statistical analyses	30
3.4	Results	31
3.4.1	Nutrient composition of different varieties of treated rice straw	31
3.4.2	Fermentation parameters of different varieties of biological treated rice straw	33
3.4.3	<i>In vitro</i> gas production and its kinetics of different varieties of treated rice straw	35
3.4.4	<i>In vitro</i> dry matter and organic matter digestibility of different varieties of treated rice straw	37
3.4.5	<i>In vitro</i> fermentation parameters of different varieties of treated rice straw	37
3.5	Discussion	40
3.5.1	Nutrient composition of different varieties of treated rice straw	40
3.5.2	Fermentation parameters of different varieties of biological treated rice straw	41
3.5.3	<i>In vitro</i> net gas production of different varieties of rice straw treatment	42
3.5.4	<i>In vitro</i> gas production kinetics of different varieties of treated rice straw	42
3.5.5	<i>In vitro</i> dry matter and organic matter digestibility of different varieties of treated rice straw	44
3.5.6	<i>In vitro</i> fermentation parameters of different varieties of treated rice straw	44
3.6	Conclusion	46
4	EFFECT OF BIOLOGICAL ADDITIVE TREATED RICE STRAW ON GROWTH PERFORMANCE, RUMEN FERMENTATION, MICROBIAL POPULATION, DIGESTIBLE NUTRIENT INTAKE AND GENE EXPRESSION IN GOATS	47
4.1	Introduction	47
4.2	Materials and Methods	48
4.2.1	Animal ethical note	48
4.2.2	Microbial culture, crude enzyme production and ensiling preparation	48
4.2.3	Experimental site, animals, housing and diet	49
4.2.4	Digestibility trial	50
4.2.5	Rumen pH, VFA and ammonia analysis	50
4.2.6	Rumen microbial population extraction and quantification	51
4.2.7	Gene expression analysis (hepatic GHR, IGF-1 and ruminal MCT-1 genes)	52
4.3	Statistical Analysis	53
4.4	Results	53

4.4.1	Intake and apparent nutrient digestibility	53
4.4.2	Growth performance profile	54
4.4.3	Rumen fermentation characteristics	55
4.4.4	Rumen microbial population in goats	55
4.4.5	Gene expression in rumen (MCT1) and hepatic (IGF1 and GHR)	56
4.5	Discussion	58
4.5.1	Intake and apparent digestibility	58
4.5.2	Growth performance	58
4.5.3	Rumen fermentation characteristics	59
4.5.4	Rumen microbial population	60
4.5.5	Gene expression in rumen (MCT1) and hepatic (IGF1 and GHR)	60
4.6	Conclusion	61
5	INFLUENCE OF BIOLOGICAL ADDITIVE TREATED RICE STRAW ON BLOOD PROFILES (HAEMATOLOGY AND BIOCHEMISTRY), CARCASS CHARACTERISTICS AND MEAT QUALITY IN GOATS	62
5.1	Introduction	62
5.2	Materials and Methods	63
5.2.1	Blood sampling	63
5.2.2	Tissue sampling and ageing of meat	63
5.2.3	Determination of muscle pH	64
5.2.4	Meat colour coordinate determination	65
5.2.5	Drip loss	65
5.2.6	Cooking loss	65
5.2.7	Determination of shear force values	66
5.3	Statistical Analysis	66
5.4	Results	66
5.4.1	Red blood cell haematological parameters in goats	66
5.4.2	White blood cell haematological parameters in goats	67
5.4.3	Serum alanine aminotransferase and aspartate aminotransferase in goats	68
5.4.4	Serum urea and total protein in goats	69
5.4.5	Serum glucose and cholesterol in goats	70
5.4.6	Weight of non-carcass parameters in goats	71
5.4.7	Weight of carcass parameters in goats	71
5.4.8	Percentage ratio, weight and tissue composition of primal cuts in goats	71
5.4.9	Physicochemical properties of different muscles from goats as influenced by biological treated rice straw and post mortem ageing	72
5.4.10	Colour coordinates of different muscles in goats	75
5.5	Discussion	77
5.5.1	Red blood cell haematological parameters in goats	77
5.5.2	White blood cell haematological parameters in goats	78
5.5.3	Serum alanine aminotransferase and aspartate aminotransferase in goats	79

5.5.4	Serum urea and total protein in goats	79
5.5.5	Serum glucose, cholesterol and triglyceride in goats	80
5.5.6	Weight of non-carcass parameters in goats	82
5.5.7	Weight of carcass parameters in goats	82
5.5.8	Percentage ratio, weight and tissue composition of primal cuts in goats	82
5.5.9	Physicochemical properties of different muscles from goats as influenced by biological treated rice straw and post mortem ageing in goats	83
5.5.10	Colour coordinate in goats	84
5.6	Conclusion	85
6	SUMMARY, CONCLUSION AND RECOMMENDATIONS FOR FUTURE RESEARCH	86
6.1	Summary	86
6.2	Conclusion	87
6.3	Recommendations for future research	87
	REFERENCES	88
	BIODATA OF STUDENT	117
	LIST OF PUBLICATIONS	118

LIST OF TABLES

Table		Page
2.1	Recent global rice and straw quantities	4
2.2	Total paddy production and productivity of the granary areas in 2017	5
2.3	Percentage changes in goat, cattle and sheep populations (2006–2016)	11
2.4	Top ten countries with the largest goat population in 2016	11
2.5	Population of livestock in Malaysia for 2016-2020	12
2.6	Registered goat farms in East Malaysia as of 2018	12
2.7	Malaysia goat per capital consumption and self-sufficiency ratio from 2016-2020	12
3.1	Effects of biological additive treatment and rice variety on the nutrient composition of treated rice straw	32
3.2	Effect of biological additive treatment and rice variety on the fermentation parameters of treated rice straw	34
3.3	Effect of biological additive treatment and rice variety on the <i>in vitro</i> net gas production, and <i>in vitro</i> gas production kinetic of treated rice straw.	36
3.4	Effect of biological additive treatment and rice variety on the <i>in vitro</i> dry matter digestibility and <i>in vitro</i> organic matter digestibility (IVDMD and IVOMD) of treated rice straw	37
3.5	Effect of biological additive treatment and rice variety on the <i>in vitro</i> rumen fermentation parameters of treated rice straw	39
4.1	Chemical composition of the feed ingredients; untreated (Control), enzyme (Enzyme) and LAB + enzyme (BE) treated rice straw, and the concentrate used for the experiment	49
4.2	Nutrient composition of the experimental diets containing 50% treated rice straw and 50% concentrate	50
4.3	Primers used for real time polymerase chain reaction for microbial quantification	51
4.4	List of primer sequences of target and reference genes	53

4.5	Nutrient intake and apparent digestibility of nutrients in goats fed biological treated rice straw over 90 days of feeding trial	54
4.6	Growth performance and feed efficiency of goats fed biological additive treated rice straw over 90 days of feeding trial	54
4.7	Rumen fermentation parameters in goats fed biological treated rice straw	55
5.1	Effect of biological treated rice straw on red blood cell haematological parameters in Boer crossbred goats	67
5.2	The effect of biological treated rice straw on white blood cell haematological parameters in goats	68
5.3	The effect of biological treated rice straw on ALT and AST biochemical parameters in goats	69
5.4	The effect of biological treated rice straw on biochemical parameters (serum renal function and total protein) in goats	69
5.5	The effect of biological treated rice straw on biochemical parameters (serum metabolites and ketone) in goats	70
5.6	Effect of biological treated rice straw on non-carcass weight components in goats	71
5.7	Effect of biological treated rice straw on carcass weight parameters in goats	71
5.8	Effect of biological treated straw on percentage ratio, weight and tissue composition of primal cuts effect in goats	72
5.9	Physicochemical properties of different muscles from goats as influenced by biological treated rice straw and post mortem ageing	74
5.10	Colour coordinate of different muscles from goats as influenced by biological treated rice straw and postmortem ageing	76

LIST OF FIGURES

Figure		Page
3.1	<i>In vitro</i> gas production equipment	30
4.1	Population of rumen microbes (Log 10 cell/ml) in goats fed biological treated rice straw	56
4.2	The relative expressions of ruminal MCT-1 in goat fed biological treated rice straw	57
4.3	The relative expressions of liver IGF-1 and GHR genes in goat fed biological treated rice straw	57
5.1	Primal cuts of goat carcass	64

LIST OF ABBREVIATIONS

ADF	Acid detergent fibre
ADL	Acid detergent lignin
NH ₃	Ammonia
ANOVA	Analysis of variance
CO ₂	Carbon dioxide
Cm	Centimeter
CTAB	Cetyltrimethylammonium bromide
Cfu	colony forming units
CF	Crude fiber
CP	Crude protein
D	Day
°C	degree Celsius
DNA	deoxyribonucleic acid
Na ₂ HPO ₄	disodium phosphate
DH ₂ O	distilled water
DM	Dry matter
EDTA	ethylene diamine tetraacetic acid
FAO	Food and Agricultural Organization
G	gram
H	hour
IVDMD	<i>In vitro</i> dry matter digestibility
Kg	kilogram
L	liter

MJ/Kg	megajoules per kilogram
ME	Metabolisable energy
CH ₄	Methane
μg	micro gram
μl	micro liter
μl	microliter
μm	micrometer
μM	micromole
μmol	Micromole
ml	milliliter
mM	Millimolar
Mmol	Millimole
Min	minute
M	Mole
KH ₂ PO ₄	monopotassium phosphate
Nm	Nanometer
NDF	Neutral detergent fiber
Ppm	Part per million
%	percent
PVPP	Polyvinyl polyrolidone
KCl	potassium chloride
Rpm	Revolutions per minute
S	second
NaCl	sodium chloride

SEM	Standard error of mean
H ₂ SO ₄	Tetraoxosulphate (vi) acid
TVFA	Total volatile fatty acid
pH _u	ultimate pH
USDA	United States Department of Agriculture
UPM	Universiti Putra Malaysia
v/v	volume per volume
CaCl ₂ •2H ₂ O	calcium chloride dehydrate
MgCl ₂ •6H ₂ O	magnesium chloride hexahydrate

CHAPTER 1

INTRODUCTION

The demand for animal protein products (meat and milk) has been estimated to skyrocket by 70% by 2050, solely because of the increase in the global population of more than 9 billion people with concurrent globalisation which influence the consumer taste as well as their perception (Ahmed *et al.*, 2018).

Ruminant in general are very essential and they play a crucial role in the enhancement of livestock sustainability in every country due to their special feed utilization which differs from other animal groups as well as its general acceptance by all religion and races especially for goat (Wanapat *et al.*, 2008). In addition, they make use of agricultural by products which are not consumed by human into high quality resources for body metabolism through the conversion of plant protein mostly less quality protein as well as non-protein nitrogen into high-quality animal proteins as meat or milk (Xu *et al.*, 2019).

However, the quest for commercial animal production scale to meet up the challenges of demand for animal protein through proper feeding without impairment to the environment is of major interests among researchers to explore (Silva *et al.*, 2015). The utilisation of agricultural by products which are ubiquitous in a value-added mechanism through the use of biological additive to enhance the nutrient composition is an accepted innovation. The agricultural by product that is easy to get in proximity as well as cheap if not free in most places due to the fact that the farmers mostly dispose it off the farm by burning which causes environmental pollution or left on farm and flooded with water which triggers the increase in acidity present in the soil. However, there is no skepticism about forage quality been dependent on structural pattern, compositional ratio, age, plant part, soil fertility and these have made the nutritional composition to differ and unpredictable except post evaluation through *in vitro* techniques and *in vivo* trials (Reis *et al.*, 2016; Ahmed *et al.*, 2017).

Biological additive are mechanism of agricultural by product enhancement through microbes to improve the quality of the feed. Most frequently used microbes are lactic acid bacterial (improve silage fermentation and prevent spoilage) and fibrolytic enzyme from fungi. Individual microbes or combined has been employed for the treatment of straw for better acceptance and utilisation by the animals (Thomas *et al.*, 2013).

According to Kung and Charley (2010), biological treated rice straw has been demonstrated to improve the nutrient content relatively to the control through aerobic stability enhancement as well as inhibition of the aerobic pathogen which could have subjected the silage to deterioration. Elwakeel *et al.*, (2007) showed an increase in the *in vitro* dry matter digestibility of different four diary feedstuff when fibrolytic enzyme was applied. Thomas *et al.*, (2013) reported that differences among the varieties with reference to the treatment application was observed coupled with improvement in the

nutrient composition and treated rice straw fermentation parameters, increase in the true *in vitro* digestibility of the biological treatment relatively to the control and enhancement in the *in situ* disappearance was noted in the biological additive treatments in a findings where sorghum silage varieties (DBMR, PS 747, S700D and MMR) were treated by fibrolytic enzyme and inoculant.

Moreover, Elkholy *et al.*, (2009) demonstrated the impact of ensiled corn crop in sheep and the author discovered that the fiber crude fractions of the ensiled treatment were reduced, the ammonia as well as the pH of the ensiled treatment were lower compared with the control. In addition, the digestibility coefficient value of organic matter, crude protein and ether extract were all higher in sheep fed treated corn silage product relatively to the sheep in the control groups. The ruminal volatile fatty acid was enhanced couple with the rumen ammonia. Also, the blood parameters (total protein, blood urea nitrogen (due to high absorption of ammonia from the rumen to the blood), albumin and blood glucose) in sheep fed treated corn products were increased.

Furthermore, cattle fed grass treated with LAB, the influence of biological additives on silage quality manifested among the treatments with increase in voluntary intake as well as digestibility in the nutrient has evidenced by (Ando *et al.*, 2006). Most of the finding employed corn silage or whole corn silage and few on wheat silage for animal trial. In the few of the trial carried out on animal trial mostly in cattle, some positive result has been reported while some reported no difference in the performances (Zhang *et al.*, 2019).

The utilisation of ensiled straw has been tremendous especially in the area of ensile quality but more is yet to be executed on animal trial, even though the biological application cannot be exhausted on animal performance especially with the use of rice straw which are ubiquitous and causing environmental impairment due to burning disposal management. To the best of my knowledge, the treatment of rice straw (PadiU Putra-1) known for its blast resistance and high yielding properties has not been examined and this brings forth the objectives of the present study which is the utilisation of enriched straw from newly developed rice variety as feed supplement for a sustainable goat production.

Hypothesis statements:

The biological enrichment of PadiU Putra-1 rice straw will improve ensiled and nutritional qualities. It can also enhance the rumen fermentation profile, microbial population and performance of goat fed with biological treated rice straw.

Problem statement

1. The burning of different varieties of straw causes health and environmental impairment. Usually, the quality of straw variety is low, but information on its biological amelioration and how it impacts *in vitro* gas production techniques is yet to be understood.
2. In terms of animal performance, the understanding of PadiU Putra1 rice straw, which is known for blast resistance and high yield potential, remains unexplored.
3. The knowledge of how biological treated PadiU Putra 1 rice straw influences meat quality and blood profile is yet to be studied.

General objective

Utilisation of treated straw from newly developed padiu putra-1 rice variety as feed supplement for a sustainable goat production

Specific objectives

1. To evaluate the effects of biological additive treatments on nutrient profiles, fermentation qualities, and *in vitro* gas production kinetics, digestibility and rumen fermentation in different varieties of rice straw (ML4, ML10, ML21, ML24, MR219 and PadiU Putra-1).
2. To determine the influence of biological additive treated rice straw on growth performance, rumen fermentation, microbial population, digestible nutrient intake, and gene expression in Boer crossed bred goat.
3. To examine the impact of biological additive treated rice straw on blood profile (hematology and biochemistry), carcass characteristics and meat quality in goats.

REFERENCES

- Aaslyng, M. D., Bejerholm, C., Ertbjerg, P., Bertram, H. C., & Andersen, H. J. (2003). Cooking loss and juiciness of pork in relation to raw meat quality and cooking procedure. *Food Quality and Preference*, *14*(4), 277-288.
- Abd El-Galil, E. R. (2014). Using biological additives to manipulate rumen fermentation and improve Baladi goats performance. *Egyptian Journal of Nutrition and Feeds*, *17*(1), 29-42.
- Abd El-Rahman, H. H., Abedo, A. A., El-Nameary, Y. A. A., Abdel-Magid, S. S., & Mohamed, M. I. (2014). Effect of biological treatments of rice straw on growth performance, digestion and economic efficiency for growing calves. *Global Veterinaria*, *13*(1), 47-54.
- Abdelrahman, M. M., & Aljumaah, R. S. (2014). Dietary protein level and performance of growing Baladi kids. *Iranian Journal of Veterinary Research*, *15*(4), 353.
- Abdullah, F. A., Jx, A., & Noor, M. S. Z. (2020). The adoption of innovation in ruminant farming for food security in Malaysia: A narrative literature review. *Journal of Critical Reviews*.
- Abo-Donia, F. M., Abdel-Azim, S. N., Elghandour, M. M., Salem, A. Z., Buendía, G., & Soliman, N. A. M. (2014). Feed intake, nutrient digestibility and ruminal fermentation activities in sheep-fed peanut hulls treated with *Trichoderma viride* or urea. *Tropical Animal Health and Production*, *46*(1), 221-228.
- Acebron, L. B., & Dopico, D. C. (2000). The importance of intrinsic and extrinsic cues to expected and experienced quality: an empirical application for beef. *Food Quality and Preference*, *11*(3), 229-238.
- Acosta Aragón, Y., Jatkauskas, J., & Vrotniakiene, V. (2012). The effect of a silage inoculant on silage quality, aerobic stability, and meat production on farm scale. *International Scholarly Research Notices*, 1-6.
- Adams, N. R., & Briegel, J. R. (2005). Multiple effects of an additional growth hormone gene in adult sheep. *Journal of Animal Science*, *83*(8), 1868-1874.
- Addah, W., Baah, J., Okine, E. K., & McAllister, T. A. (2012). A third-generation esterase inoculant alters fermentation pattern and improves aerobic stability of barley silage and the efficiency of body weight gain of growing feedlot cattle. *Journal of Animal Science*, *90*(5), 1541-1552.
- Adesogan, A. T., Arriola, K. G., Jiang, Y., Oyebade, A., Paula, E. M., Pech-Cervantes, A. A., ... & Vyas, D. (2019). Symposium review: Technologies for improving fiber utilization. *Journal of Dairy Science*, *102*(6), 5726-5755.

- Adeyemi, K. D., Ahmed, M. A., Jotham, S., Roslan, N. A., Jahromi, M. F., Samsudin, A. A., & Sazili, A. Q. (2016). Rumen microbial community and nitrogen metabolism in goats fed blend of palm oil and canola oil. *Italian Journal of Animal Science*, *15*(4), 666-672.
- Adeyemi, K. D., Shittu, R. M., Sabow, A. B., Ebrahimi, M., & Sazili, A. Q. (2016). Influence of diet and post mortem ageing on oxidative stability of lipids, myoglobin and myofibrillar proteins and quality attributes of gluteus medius muscle in goats. *PLoS One*, *11*(5), e0154603.
- Agrawal, R., Satlewal, A., Kapoor, M., Mondal, S., & Basu, B. (2017). Investigating the enzyme-lignin binding with surfactants for improved saccharification of pilot scale pre-treated wheat straw. *Bioresource Technology*, *224*, 411-418.
- Ahmed, M. A., Adeyemi, K. D., Jahromi, M. F., Jusoh, S., Alimon, A. R., & Samsudin, A. A. (2017). Effects of dietary *Kleinhovia hospita* and *Leucaena leucocephala* leaves on rumen fermentation and microbial population in goats fed treated rice straw. *Tropical Animal Health and Production*, *49*(8), 1749-1756.
- Ahmed, M. A., Jusoh, S., Alimon, A. R., Ebrahimi, M., & Samsudin, A. A. (2018). Nutritive and anti-nutritive evaluation of *Kleinhovia hospita*, *Leucaena leucocephala* and *Gliricidia sepium* with respect to their effects on in vitro rumen fermentation and gas production. *Tropical Animal Science Journal*, *41*(2), 128-136.
- Aksu, T., Baytok, E., & Bolat, D. (2004). Effects of a bacterial silage inoculant on corn silage fermentation and nutrient digestibility. *Small Ruminant Research*, *55*(1-3), 249-252.
- Albers, S. V., Konings, W. N., & Driessen, A. J. (2007). Solute transport. *Archaea: Molecular and Cellular Biology*, 354-368.
- Al-Eissa, M. S., Alkahtani, S., Al-Farraj, S. A., Alarifi, S. A., Al-Dahmash, B., & Al-Yahya, H. (2012). Seasonal variation effects on the composition of blood in Nubian ibex (*Capra nubiana*) in Saudi Arabia. *African Journal of Biotechnology*, *11*(5), 1283-1286.
- Allen, M. S. (1997). Relationship between fermentation acid production in the rumen and the requirement for physically effective fiber. *Journal of Dairy Science*, *80*(7), 1447-1462.
- Alonso-Díaz, M. A., Torres-Acosta, J. F. J., Sandoval-Castro, C. A., & Hoste, H. (2010). Tannins in tropical tree fodders fed to small ruminants: a friendly foe? *Small Ruminant Research*, *89*(2-3), 164-173.
- Anaeto, M., Adeyeye, J. A., Chioma, G. O., Olarinmoye, A. O., & Tayo, G. O. (2010). Goat products: Meeting the challenges of human health and nutrition. *Agriculture and Biology Journal of North America*, *6*, 1231-1236.

- Anand, M. R., Kumar, H. S., Kommireddy, P., & Murthy, K. K. (2019). Secondary and micronutrient management practices in organic farming- an overview. *Current Agriculture Research Journal*, 7(1), 4.
- Anantasook, N., & Wanapat, M. (2012). Influence of rain tree pod meal supplementation on rice straw based diets using in vitro gas fermentation technique. *Asian-Australasian Journal of Animal Sciences*, 25(3), 325.
- Ando, S., Ishida, M., Oshio, S., & Tanaka, O. (2006). Effects of isolated and commercial lactic acid bacteria on the silage quality, digestibility, voluntary intake and ruminal fluid characteristics. *Asian-Australasian Journal of Animal Sciences*, 19(3), 386-389.
- AOAC. (2007). Official methods of analysis of the Association of Official Analytical Chemists (18th Ed). Association of Official Analytical Chemists, Washington D.C., USA.
- Ariff, O. M., Sharifah, N. Y., & Hafidz, A. W. (2015). Status of beef industry of Malaysia. *Malaysian Journal of Animal Science*, 18(2), 1-21.
- Arriola, K. G., Kim, S. C., Staples, C. R., & Adesogan, A. T. (2011). Effect of applying bacterial inoculants containing different types of bacteria to corn silage on the performance of dairy cattle. *Journal of Dairy Science*, 94(8), 3973-3979.
- Atti, N., Mahouachi, M., & Rouissi, H. (2006). The effect of spineless cactus (*Opuntia ficus-indica* f. *inermis*) supplementation on growth, carcass, meat quality and fatty acid composition of male goat kids. *Meat Science*, 73(2), 229-235.
- Aziz, M. A. (2010). Present status of the world goat populations and their productivity. *Lohman Information*, 45(2), 42-52.
- Babaeinasab, Y., Rouzbehan, Y., Fazaeli, H., & Rezaei, J. (2015). Chemical composition, silage fermentation characteristics, and in vitro ruminal fermentation parameters of potato-wheat straw silage treated with molasses and lactic acid bacteria and corn silage. *Journal of Animal Science*, 93(9), 4377-4386.
- Bach, A., Calsamiglia, S., & Stern, M. D. (2005). Nitrogen metabolism in the rumen. *Journal of Dairy Science*, 88, E9-E21.
- Bailey, M. J., Biely, P., & Poutanen, K. (1992). Interlaboratory testing of methods for assay of xylanase activity. *Journal of Biotechnology*, 23(3), 257-270.
- Baldwin, R. L., McLeod, K. R., Klotz, J. L., & Heitmann, R. N. (2004). Rumen development, intestinal growth and hepatic metabolism in the pre-and postweaning ruminant. *Journal of dairy science*, 87, E55-E65.

- Bandyk, C. A., Cochran, R. C., Wickersham, T. A., Titgemeyer, E. C., Farmer, C. G., & Higgins, J. J. (2001). Effect of ruminal vs post-ruminal administration of degradable protein on utilization of low-quality forage by beef steers. *Journal of Animal Science*, 79(1), 225-231.
- Barbera, S., & Tassone, S. (2006). Meat cooking shrinkage: Measurement of a new meat quality parameter. *Meat Science*, 73(3), 467-474.
- Bartoň, L., Bureš, D., & Kudrna, V. (2010). Meat quality and fatty acid profile of the musculus longissimus lumborum in Czech Fleckvieh, Charolais and Charolais× Czech Fleckvieh bulls fed different types of silages. *Czech Journal of Animal Science*, 55(11), 479-487.
- Basso, F. C., Adesogan, A. T., Lara, E. C., Rabelo, C. H. S., Berchielli, T. T., Teixeira, I. A. M. A., ... & Reis, R. A. (2014). Effects of feeding corn silage inoculated with microbial additives on the ruminal fermentation, microbial protein yield, and growth performance of lambs. *Journal of Animal Science*, 92(12), 5640-5650.
- Beauchemin, K. A., Colombatto, D., Morgavi, D. P., & Yang, W. Z. (2003). Use of exogenous fibrolytic enzymes to improve feed utilization by ruminants. *Journal of Animal Science*, 81(14_suppl_2), E37-E47.
- Becker, T. (2000). Consumer perception of fresh meat quality: a framework for analysis. *British Food Journal* 102(3), 158-176.
- Beharka, A. A., Nagaraja, T. G., Morrill, J. L., Kennedy, G. A., & Klemm, R. D. (1998). Effects of form of the diet on anatomical, microbial, and fermentative development of the rumen of neonatal calves. *Journal of Dairy Science*, 81(7), 1946-1955.
- Bekele, W., Melaku, S., & Mekasha, Y. (2013). Effect of substitution of concentrate mix with *Sesbania sesban* on feed intake, digestibility, body weight change, and carcass parameters of Arsi-Bale sheep fed a basal diet of native grass hay. *Tropical Animal Health and Production*, 45(8), 1677-1685.
- Berahir, Z., Omar, M. H., Zakaria, N. I., Ismail, M. R., Rosle, R., Roslin, N. A., & Che'Ya, N. N. (2021). Silicon improves yield performance by enhancement in physiological responses, crop imagery, and leaf and culm sheath morphology in new rice line, PadiU Putra. *BioMed Research International*, 2021.
- Bergman, E. N. (1990). Energy contributions of volatile fatty acids from the gastrointestinal tract in various species. *Physiological Reviews*, 70(2), 567-590.
- Bocian, M., Jankowiak, H., Reszka, P., & Banaszak, S. (2017). Pork quality with special emphasis on colour and its changes during storage. *Journal of Central European Agriculture*, 19(1), 102-113.
- Boeckert, C., Vlaeminck, B., Fievez, V., Maignien, L., Dijkstra, J., & Boon, N. (2008). Accumulation of trans C18: 1 fatty acids in the rumen after dietary algal

supplementation is associated with changes in the Butyrivibrio community. *Applied and Environmental Microbiology*, 74(22), 6923-6930.

- Bolumar, T., Enneking, M., Toepfl, S., & Heinz, V. (2013). New developments in shockwave technology intended for meat tenderization: Opportunities and challenges. A review. *Meat Science*, 95(4), 931-939.
- Bouton, P. E., Harris, P. V., & Shorthose, W. R. (1972). The effects of ultimate pH on ovine muscle: Water-holding capacity. *Journal of Food Science*, 37(3), 351-355.
- Bowker, B. C., Eastridge, J. S., Paroczay, E. W., Callahan, J. A., & Solomon, M. B. (2010). Aging/tenderization mechanisms. *Handbook of Meat Processing*. Ed. F. Toldrá. Blackwell Publishing, Ames, Iowa, 87-104.
- Broucek J. (2018). Options to methane production abatement in ruminants: A review. *Journal of Animal and Plant Sciences*, 28(2), 348-364.
- Bryant, M. P. (1970). Normal flora—rumen bacteria. *The American Journal of Clinical Nutrition*, 23(11), 1440-1450.
- Buckhaus, E. M., & Smith, Z. K. (2021). Effects of corn silage inclusion level and type of anabolic implant on animal growth performance, apparent total tract digestibility, beef production per hectare, and carcass characteristics of finishing steers. *Animals*, 11(2), 579.
- Campos, F. S., Carvalho, G. G. P., Santos, E. M., Araújo, G. G. L., Rebouças, R. A., Estrela-Lima, A., ... Magalhães, A. L. R. (2019). Metabolic profile and histopathology of kidneys and liver of lambs fed silages of forages adapted to a semi-arid environment. *South African Journal of Animal Science*, 49(3), 555-563.
- Cao, Y., Cai, Y., Takahashi, T., & Kongo, M. (2013). Ruminal digestibility and quality of silage conserved via fermentation by lactobacilli. *Lactic Acid Bacteria-R & D for Food, Health and Livestock Purposes*. InTech.
- Carlos, M. M. L., Leite, J. H. G. M., Chaves, D. F., Vale, A. M., Façanha, D. A. E., Melo, M. M., & Soto-Blanco, B. (2015). Blood parameters in the Morada Nova sheep: influence of age, sex and body condition score. *The Journal of Animal and Plant Sciences*, 25(4), 950-955.
- Castillo-González, A. R., Burrola-Barraza, M. E., Domínguez-Viveros, J., & Chávez-Martínez, A. (2014). Rumen microorganisms and fermentation. *Archivos de Medicina Veterinaria*, 46(3), 349-361.
- Chen, X. B., Chen, Y. K., Franklin, M. F., Ørskov, E. R., & Shand, W. J. (1992). The effect of feed intake and body weight on purine derivative excretion and microbial protein supply in sheep. *Journal of Animal Science*, 70(5), 1534-1542

- Choudhury, P. K., Salem, A. Z. M., Jena, R., Kumar, S., Singh, R., & Puniya, A. K. (2015). Rumen microbiology: An overview. In *Rumen Microbiology: from Evolution to Revolution* (pp. 3-16). Springer, New Delhi.
- Chukwu, S. C., Rafii, M. Y., Ramlee, S. I., Ismail, S. I., Oladosu, Y., Muhammad, I. I., ... & Yusuf, B. R. (2020). Recovery of recurrent parent genome in a marker-assisted backcrossing against rice blast and blight infections using functional markers and SSRs. *Plants*, 9(11), 1411.
- Coleman, G. S. (1975). The interrelationship between rumen ciliate protozoa and bacteria. I.W. McDonald, A.C.I. Warner (Eds.), *Digestion and Metabolism in the Ruminant*, University of New England, Armidale, N.S.W (1975), pp. 149-164
- Colombatto, D., Mould, F. L., Bhat, M. K., & Owen, E. (2007). Influence of exogenous fibrolytic enzyme level and incubation pH on the in vitro ruminal fermentation of alfalfa stems. *Animal Feed Science and Technology*, 137(1-2), 150-162.
- Colomer-Rocher, F., Morand-Fehr, P., & Kirton, A. H. (1987). Standard methods and procedures for goat carcass evaluation, jointing and tissue separation. *Livestock Production Science*, 17, 149-159.
- Connor, E. E., Li, R. W., Baldwin, R. L., & Li, C. (2010). Gene expression in the digestive tissues of ruminants and their relationships with feeding and digestive processes. *Animal*, 4(7), 993-1007.
- Contreras-Govea, F. E., Muck, R. E., Mertens, D. R., & Weimer, P. J. (2011). Microbial inoculant effects on silage and in vitro ruminal fermentation, and microbial biomass estimation for alfalfa, bmr corn, and corn silages. *Animal Feed Science and Technology*, 163(1), 2-10.
- Contreras-Zentella, M. L., & Hernández-Muñoz, R. (2016). Is liver enzyme release really associated with cell necrosis induced by oxidant stress? *Oxidative Medicine and Cellular Longevity*, 2016, 3529149. doi:10.1155/2016/3529149
- Cox, R. A., & Garcia-Palmieri, M. R. (1990). Cholesterol, Triglycerides, and Associated Lipoproteins In: Walker, HK; Hall, WD; Hurst, JW (ed) *Clinical Methods: The History, Physical, and Laboratory Examinations*. 3rd edition. Butterworths, Boston: pp. 153-160.
- Cuff, M. A., Lambert, D. W., & Shirazi-Beechey, S. P. (2002). Substrate-induced regulation of the human colonic monocarboxylate transporter, MCT1. *The Journal of Physiology*, 539(2), 361-371.
- da Silva TC, da Silva LD, Santos EM, Oliveira JS, Perazzo AF (2017) Importance of the Fermentation to Produce High-Quality Silage *Fermentation Processes*, 1-20.
- Daly, K., Cuff, M. A., Fung, F., & Shirazi-Beechey, S. P. (2005). The importance of colonic butyrate transport to the regulation of genes associated with colonic tissue homeostasis. *Biochemical Society Transactions*, 33, 733-735

- Daramola, J. O., Adeloye, A. A., Fatoba, T. A., & Soladoye, A. O. (2005). Haematological and biochemical parameters of West African Dwarf goats. *Livestock Research for Rural Development*, 17(8), 95.
- de Oliveira, J. S., Santos, E. M., & dos Santos, A. P. M. (2016). Intake and digestibility of silages. *Advances in Silage Production and Utilization*, 101.
- Deb, G. K., Nahar, T. N., Duran, P. G., & Presicce, G. A. (2016). Safe and sustainable traditional production: the water buffalo in Asia. *Frontiers in Environmental Science*, 4, 38.
- Dehority, B. A. (2003). *Rumen microbiology* (Vol. 372). Nottingham, UK: Nottingham University Press.
- Denman, S. E., & McSweeney, C. S. (2006). Development of a real-time PCR assay for monitoring anaerobic fungal and cellulolytic bacterial populations within the rumen. *FEMS Microbiology Ecology*, 58(3), 572-582.
- Department of Agriculture. (2019). Paddy statistics of Malaysia 2018.
- Department of Standards Malaysia (2009). MS1500: 2009. Malaysian Standards. Halal food— Production, preparation, handling and storage— General guideline. *Cyberjaya: Department of Standards Malaysia*, 1-13.
- DePeters, E. J., & Ferguson, J. D. (1992). Nonprotein nitrogen and protein distribution in the milk of cows. *Journal of Dairy Science*, 75(11), 3192-3209.
- Depreux, F. F. S., Grant, A. L., & Gerrard, D. E. (2002). Influence of halothane genotype and body-weight on myosin heavy chain composition in pig muscle as related to meat quality. *Livestock Production Science*, 73(2-3), 265-273.
- Direkvandi, E., Mohammadabadi, T., & Salem, A. Z. (2020). Effect of microbial feed additives on growth performance, microbial protein synthesis, and rumen microbial population in growing lambs. *Translational Animal Science*, 4(4), 1-10.
- Dong, Z., Wang, S., Zhao, J., Li, J., & Shao, T. (2020). Effects of additives on the fermentation quality, in vitro digestibility and aerobic stability of mulberry (*Morus alba* L.) leaves silage. *Asian-Australasian Journal of Animal Sciences*, 33(8), 1292.
- Drouin, P., Mari, L. J., & Schmidt, R. J. (2019). Lactic acid bacteria as microbial silage additives: current status and future outlook. *New Advances on Fermentation Processes*, 266.
- DVS. (2021). 2019/2020 Livestock Statistics. Department of Veterinary Services (DVS),
- DVS. (2018). Total Registration of Livestock Premises by Commodity in Peninsula Malaysia. Malaysia Open Data Portal, 19 September 2018.

- Elghandour, M. M. Y., Kholif, A. E., Salem, A. Z. M., Olafadehan, O. A., & Kholif, A. M. (2016). Sustainable anaerobic rumen methane and carbon dioxide productions from prickly pear cactus flour by organic acid salts addition. *Journal of Cleaner Production*, 139, 1362-1369.
- Elkholy, M. E. H., Hassanein, E. I., Soliman, M. H., Eleraky, W., Elgamel, M. F., & Ibraheim, D. (2009). Efficacy of feeding ensiled corn crop residues to sheep. *Pakistan Journal of Nutrition*, 8(12), 1858-1867.
- Elwakeel, E. A., Titgemeyer, E. C., Johnson, B. J., Armendariz, C. K., & Shirley, J. E. (2007). Fibrolytic enzymes to increase the nutritive value of dairy feedstuffs. *Journal of Dairy Science*, 90(11), 5226-5236.
- Enerson, B. E., & Drewes, L. R. (2003). Molecular features, regulation, and function of monocarboxylate transporters: implications for drug delivery. *Journal of Pharmaceutical Sciences*, 92(8), 1531-1544.
- Faiz, F., Queen, F., Amir, R. M., Ahmad, A., Ahmad, Z., Ameer, K., ... & Kabir, K. (2020). Physicochemical, microbiological and sensory characteristics of goats reared on organic rationing in Karakoram region. *Food Science and Technology*, 41, 381-387.
- Fang, J., Matsuzaki, M., Suzuki, H., Cai, Y., Horiguchi, K. I., & Takahashi, T. (2012). Effects of lactic acid bacteria and urea treatment on fermentation quality, digestibility and ruminal fermentation of roll bale rice straw silage in wethers. *Grassland science*, 58(2), 73-78.
- Fennema, O. R. (1985). Water and ice. Ch. 2. *Food Chemistry*. OR Fennema (Ed.), Marcel Dekker, Inc. New York, NY, 17-94.
- Fernando, S. C., Purvis, H. T., Najar, F. Z., Sukharnikov, L. O., Krehbiel, C. R., Nagaraja, T. G., ... & DeSilva, U. (2010). Rumen microbial population dynamics during adaptation to a high-grain diet. *Applied and Environmental Microbiology*, 76(22), 7482-7490.
- Ferry, J. G., & Kastead, K. A. (2007). Methanogenesis. In: Cavicchioli R (ed) *Archaea: Molecular and Cellular Biology*, ASM Press, Washington, DC, pp 288–314288-314.
- Firmenich, C. S., Schnepel, N., Hansen, K., Schmicke, M., & Muscher-Banse, A. S. (2020). Modulation of growth hormone receptor-insulin-like growth factor 1 axis by dietary protein in young ruminants. *British Journal of Nutrition*, 123(6), 652-663.
- Food and Agriculture Organization of the United Nations, Rice Market Monitor (RMM). FAO, Washington, DC (2017).
- Fouhse, J. M., Smiegielski, L., Tuplin, M., Guan, L. L., & Willing, B. P. (2017). Host immune selection of rumen bacteria through salivary secretory IgA. *Frontiers in Microbiology*, 8, 848.

- Franzolin, R., Rosales, F. P., & Soares, W. V. B. (2010). Effects of dietary energy and nitrogen supplements on rumen fermentation and protozoa population in buffalo and zebu cattle. *Revista Brasileira de Zootecnia*, 39, 549-555.
- Freise, K., Brewer, S., & Novakofski, J. (2005). Duplication of the pale, soft, and exudative condition starting with normal post mortem pork. *Journal of Animal Science*, 83(12), 2843-2852.
- Frizzo, L. S., Soto, L. P., Zbrun, M. V., Bertozzi, E., Sequeira, G., Armesto, R. R., & Rosmini, M. R. (2010). Lactic acid bacteria to improve growth performance in young calves fed milk replacer and spray-dried whey powder. *Animal Feed Science and Technology*, 157(3-4), 159-167.
- Fugita, C. A., Prado, I. N. D., Jobim, C. C., Zawadzki, F., Valero, M. V., Pires, M. C. D. O., ... & Françoze, M. C. (2012). Corn silage with and without enzyme-bacteria inoculants on performance, carcass characteristics and meat quality in feedlot finished crossbred bulls. *Revista Brasileira de Zootecnia*, 41(1), 154-163.
- Fuller, R. (1989). Probiotics in man and animals. *Journal of Applied Bacteriology*, 66(5), 365-378.
- Gäbel, G., & Aschenbach, J. R. (2006). Ruminal SCFA absorption: channelling acids without harm. *Ruminant physiology: digestion, metabolism and impact of nutrition on gene expression, immunology and stress* (ed. K Sejrsen, T Hvelplund and MO Nielsen), 173-195.
- Gaili, E. S., & Ali, A. E. (1985). Meat from Sudan desert sheep and goats: Part 2—composition of the muscular and fatty tissues. *Meat Science*, 13(4), 229-236.
- Gandra, J. R., De Oliveira, E. R., de Goes, R. H. T. B., De Oliveira, K. M. P., Takiya, C. S., Del Valle, T. A., ... & Pause, A. D. S. (2017). Microbial inoculant and an extract of *Trichoderma longibrachiatum* with xylanase activity effect on chemical composition, fermentative profile and aerobic stability of guinea grass (*Panicum maximum* Jacq.) silage. *Journal of Animal and Feed Sciences*, 26(4), 339-347.
- García-Segovia, P., Andrés-Bello, A., & Martínez-Monzó, J. (2007). Effect of cooking method on mechanical properties, color and structure of beef muscle (M. pectoralis). *Journal of Food Engineering*, 80(3), 813-821.
- Getachew, G., DePeters, E., & Robinson, P. (2004). In vitro gas production provides effective method for assessing ruminant feeds. *California Agriculture*, 58(1), 54-58.
- Ghose, T. K. (1987). Measurement of cellulase activities. *Pure and applied Chemistry*, 59(2), 257-268.
- Glitsch, K. (2000). Consumer perceptions of fresh meat quality: cross-national comparison. *British Food Journal*, 102, 177-194

- Gofur, M. R., Hossain, K. M. M., Khaton, R., & Hasan, M. R. (2014). Effect of testosterone on physio-biochemical parameters and male accessory sex glands of Black Bengal goat. *International Journal of Emerging Technology and Advanced Engineering*, 9, 456-465.
- Graham, C., Gatherar, I., Haslam, I., Glanville, M., & Simmons, N. L. (2007). Expression and localization of monocarboxylate transporters and sodium/proton exchangers in bovine rumen epithelium. *American Journal of Physiology-Regulatory, Integrative and Comparative Physiology*, 292(2), R997-R1007.
- Guerrero, A., Velandia Valero, M., Campo, M. M., & Sañudo, C. (2013). Some factors that affect ruminant meat quality: from the farm to the fork. Review. *Acta Scientiarum. Animal Sciences*, 35(4), 335-347.
- Gunun, P., Wanapat, M., & Anantasook, N. (2013). Effects of physical form and urea treatment of rice straw on rumen fermentation, microbial protein synthesis and nutrient digestibility in dairy steers. *Asian-Australasian journal of Animal Sciences*, 26(12), 1689.
- Hadjiagapiou, C., Schmidt, L., Dudeja, P. K., Layden, T. J., & Ramaswamy, K. (2000). Mechanism (s) of butyrate transport in Caco-2 cells: role of monocarboxylate transporter 1. *American Journal of Physiology-Gastrointestinal and Liver Physiology*, 279(4), G775-G780.
- Halestrap, A. P., & Meredith, D. (2004). The SLC16 gene family—from monocarboxylate transporters (MCTs) to aromatic amino acid transporters and beyond. *Pflügers Archiv*, 447(5), 619-628.
- Halestrap, A. P., & Price, N. T. (1999). The proton-linked monocarboxylate transporter (MCT) family: structure, function and regulation. *Biochemical Journal*, 343(2), 281-299.
- Hashim, F. A. H. (2015). Strategies to strengthen livestock industry in Malaysia. Malaysian Agricultural Research and Development Institute (MARDI), 1-6.
- He, L., Wu, H., Meng, Q., & Zhou, Z. (2018). Growth performance, carcass traits, blood parameters, rumen enzymes, and fattening earnings of cattle fed corn silage/corn stalk silage-based finishing diets. *Czech Journal of Animal Science*, 63(12), 483-491.
- He, L., Zhou, W., Wang, Y., Wang, C., Chen, X., & Zhang, Q. (2018). Effect of applying lactic acid bacteria and cellulase on the fermentation quality, nutritive value, tannins profile and in vitro digestibility of *Neolamarckia cadamba* leaves silage. *Journal of Animal Physiology and Animal Nutrition*, 102(6), 1429-1436.
- Heinze, P. H., Smit, M. C., Naude, R. T., & Boccard, R. L. (1986). Influence of breed and age on collagen content and solubility of some ovine and goat muscles. *Proc. 32nd Eur. Meet. Meat Research Workers, INRA, Theix, France*, 169-173.

- Henderson, G., Cox, F., Ganesh, S., Jonker, A., Young, W., Abecia, L., ... & Attwood, G. T. (2015). Rumen microbial community composition varies with diet and host, but a core microbiome is found across a wide geographical range. *Scientific Reports*, 5, 14567.
- Hetta, M., Cone, J. W., Gustavsson, A. M., & Martinsson, K. (2003). The effect of additives in silages of pure timothy and timothy mixed with red clover on chemical composition and in vitro rumen fermentation characteristics. *Grass and Forage Science*, 58(3), 249-257.
- Honikel, K.O. (1998). Reference methods for the assessment of physical characteristics of meat. *Meat Science* 49(4), 447-457.
- Hook, S. E., Wright, A. D. G., & McBride, B. W. (2010). Methanogens: methane producers of the rumen and mitigation strategies. *Archaea*, 1-11.
- Huff-Lonergan, E. (2010). Chemistry and biochemistry of meat. *Handbook of meat Processing*, 1 5-24
- Huhtanen, P., Khalili, H., Nousiainen, J. I., Rinne, M., Jaakkola, S., Heikkilä, T., & Nousiainen, J. (2002). Prediction of the relative intake potential of grass silage by dairy cows. *Livestock Production Science*, 73(2-3), 111-130.
- Hungate, R. E. (1966). *The Rumen and its Microbes* Academic Press NY.
- Hunt, M.C. (1980). Meat color measurements. *Proceedings of the Meat Conference of the American Meat Science Association. Purdue University, Lafayette, Indiana*. Pp. 41-46.
- Huntington, G. B., Prior, R. L., & Britton, R. A. (1981). Glucose and lactate absorption and metabolic interrelationships in steers changed from low to high concentrate diets. *The Journal of Nutrition*, 111(7), 1164-1172.
- Hussein, H. A., Thurmann, J. P., & Staufienbiel, R. (2020). 24-h variations of blood serum metabolites in high yielding dairy cows and calves. *BMC Veterinary Research*, 16(1), 1-11.
- Huws, S. A., Creevey, C. J., Oyama, L. B., Mizrahi, I., Denman, S. E., Popova, M., ... & Morgavi, D. P. (2018). Addressing global ruminant agricultural challenges through understanding the rumen microbiome: past, present, and future. *Frontiers in Microbiology*, 9, 2161.
- Huws, S. A., Lee, M. R., Muetzel, S. M., Scott, M. B., Wallace, R. J., & Scollan, N. D. (2010). Forage type and fish oil cause shifts in rumen bacterial diversity. *FEMS Microbiology Ecology*, 73(2), 396-702.
- Huyen, N. T., Tuan, B. Q., Ngo, X. N., Nguyen, T. B. T., & Le, N. T. T. (2019). Effect of using fungal treated rice straw in sheep diet on nutrients digestibility and microbial protein synthesis. *Asian Journal of Animal Sciences*, 13(1), 1-7.

- Ibrahim, N. A., Mohamed, W. N. W., Noh, A. M., & Saminathan, M. (2021). Growth performance and carcass traits of goats fed with oil palm by-products based feed pellet. *Journal of Oil Palm Research*, 33(2), 327-334.
- Ishler, V. A., Heinrichs, A. J., & Varga, G. B. (1996). *From feed to milk: understanding rumen function* (Vol. 422). Pennsylvania State University.
- James, C., & James, S. J. (2010). Freezing/thawing. *Handbook of Meat Processing*, Wiley-Blackwell, USA (2010), pp. 105-125.
- Jankowiak, H., Cebulska, A., & Bocian, M. (2021). The relationship between acidification (pH) and meat quality traits of polish white breed pigs. *European Food Research and Technology*, 247(11), 2813-2820.
- Janssen, P. H., & Kirs, M. (2008). Structure of the archaeal community of the rumen. *Applied and Environmental Microbiology*, 74(12), 3619-3625.
- Jawasreh, K., Awawdeh, F., Ismail, Z. B., Al-Rawashdeh, O., & Al-Majali, A. (2010). Normal hematology and selected serum biochemical values in different genetic lines of Awassi ewes in Jordan. *Internet Journal of Veterinary Medicine*, 7(2), 1-5.
- Jing, X. P., Peng, Q. H., Hu, R., Zou, H. W., Wang, H. Z., Yu, X. Q., ... & Wang, Z. S. (2018). Dietary supplements during the cold season increase rumen microbial abundance and improve rumen epithelium development in Tibetan sheep. *Journal of Animal Science*, 96(1), 293-305.
- Johnson, K. A., & Johnson, D. E. (1995). Methane emissions from cattle. *Journal of Animal Science*, 73(8), 2483-2492.
- Jones, L. (2012). Inoculants: Insurance or strategy.
- Joo, S. T., Kauffman, R. G., Kim, B. C., & Park, G. B. (1999). The relationship of sarcoplasmic and myofibrillar protein solubility to colour and water-holding capacity in porcine longissimus muscle. *Meat Science*, 52(3), 291-297.
- Joo, S. T., Kim, G. D., Hwang, Y. H., & Ryu, Y. C. (2013). Control of fresh meat quality through manipulation of muscle fiber characteristics. *Meat Science*, 95(4), 828-836.
- Jurkovich, V., Kutasi, J., Fébel, H., Reiczigel, J., Brydl, E., Könyves, L., & Rafai, P. (2006). Rumen fermentation response to a direct-fed xylanase enzyme preparation from *Thermomyces lanuginosus* in sheep. *Acta Veterinaria Hungarica*, 54(3), 333-342.
- Kadim, I. T., & Mahgoub, O. (2012). Nutritive value and quality characteristics of goat meat. *Goat meat production and quality*, Wallingford: CABI Publishing; p. 292-323.

- Kadim, I. T., Mahgoub, O., Al-Ajmi, D. S., Al-Maqbaly, R. S., Al-Saqri, N. M., & Ritchie, A. (2004). An evaluation of the growth, carcass and meat quality characteristics of Omani goat breeds. *Meat Science*, 66(1), 203-210.
- Kadim, I. T., Mahgoub, O., AL-MARZOOQI, W., Khalaf, S., AL-SINAWI, S. S., & AL-AMRI, I. (2010). Effects of transportation during the hot season, breed and electrical stimulation on histochemical and meat quality characteristics of goat longissimus muscle. *Animal Science Journal*, 81(3), 352-361.
- Kadim, T., & Mahgoub, O. (2012). 13 Nutritive Value and Quality. *Goat Meat Production and Quality*, 292.
- Kafilzadeh, F., & Heidary, N. (2013). Chemical composition, in vitro digestibility and kinetics of fermentation of whole-crop forage from 18 different varieties of oat (*Avena sativa* L.). *Journal of Applied Animal Research*, 41(1), 61-68.
- Kala, A., Kamra, D. N., Kumar, A., Agarwal, N., Chaudhary, L. C., & Joshi, C. G. (2017). Impact of levels of total digestible nutrients on microbiome, enzyme profile and degradation of feeds in buffalo rumen. *PloS One*, 12(2), e0172051.
- Kaliber, M.; Koluman, N.; & Silanikove, N. (2016). Physiological and behavioural basis for the successful adaptation of goats to severe water restriction under hot environmental conditions. *Animal*, 10, 82–88.
- Kannan, G., Kouakou, B., & Gelaye, S. (2001). Color changes reflecting myoglobin and lipid oxidation in chevon cuts during refrigerated display. *Small Ruminant Research*, 42(1), 67-74.
- Kanyinji, F., Kumagai, H., Maeda, T., Kaneshima, S., & Yokoi, D. (2009). Effects of supplementary inosine on nutrient digestibility, ruminal fermentation and nitrogen balance in goats fed high amount of concentrate. *Animal Feed Science and Technology*, 152(1-2), 12-20.
- Karaşahin, T., Aksoy, N. H., Haydardedeoglu, A. E., Dursun, Ş., Bulut, G., Çamkerten, G., ... & İlğün, R. (2019). Serum cholesterol levels in Hair goats of Aksaray Region. *Indian Journal of Animal Research*, 53(1), 63-66.
- Keady, T. W. J., & Steen, R. W. J. (1995). The effects of treating low dry-matter, low digestibility grass with a bacterial inoculant on the intake and performance of beef cattle, and studies on its mode of action. *Grass and Forage Science*, 50(3), 217-226.
- Keady, T. W. J., & Steen, R. W. J. (1996). Effects of applying a bacterial inoculant to silage immediately before feeding on silage intake, digestibility, degradability and rumen volatile fatty acid concentrations in growing beef cattle. *Grass and Forage Science*, 51(2), 155-162.
- Khan, A., Rehman, S., Imran, R., & Pitafi, K. D. (2013). Analysis of serum cholesterol level in goats breeds in Gilgit-Baltistan area of Pakistan. *Journal of Agricultural Science and Technology. A*, 3(4A), 302.

- Khazaal, K., Dentinho, M. T., Ribeiro, J. M., & Ørskov, E. R. (1995). Prediction of apparent digestibility and voluntary intake of hays fed to sheep: comparison between using fibre components, in vitro digestibility or characteristics of gas production or nylon bag degradation. *Animal Science*, 61(3), 527-538.
- Khuntia, A., & Chaudhary, L. C. (2002). Performance of male crossbred calves as influenced by substitution of grain by wheat bran and the addition of lactic acid bacteria to diet. *Asian-Australasian Journal of Animal Sciences*, 15(2), 188-194.
- Kim, J. G., Ham, J. S., Li, Y. W., Park, H. S., Huh, C. S., & Park, B. C. (2017). Development of a new lactic acid bacterial inoculant for fresh rice straw silage. *Asian-Australasian Journal of Animal Sciences*, 30(7), 950.
- Kinley, R. D., de Nys, R., Vucko, M. J., Machado, L., & Tomkins, N. W. (2016). The red macroalgae *Asparagopsis taxiformis* is a potent natural antimethanogenic that reduces methane production during in vitro fermentation with rumen fluid. *Animal Production Science*, 56(3), 282-289.
- Kiran, S., Bhutta, A. M., Khan, B. A., Durrani, S., Ali, M., & Iqbal, F. (2012). Effect of age and gender on some blood biochemical parameters of apparently healthy small ruminants from Southern Punjab in Pakistan. *Asian Pacific Journal of Tropical Biomedicine*, 2(4), 304-306.
- Kirat, D., Inoue, H., Iwano, H., Hirayama, K., Yokota, H., Taniyama, H., & Kato, S. (2005). Expression and distribution of monocarboxylate transporter 1 (MCT1) in the gastrointestinal tract of calves. *Research in Veterinary Science*, 79(1), 45-50.
- Kirat, D., Masuoka, J., Hayashi, H., Iwano, H., Yokota, H., Taniyama, H., & Kato, S. (2006). Monocarboxylate transporter 1 (MCT1) plays a direct role in short-chain fatty acids absorption in caprine rumen. *The Journal of Physiology*, 576(2), 635-647.
- Koho, N. M., Taponen, J., Tiihonen, H., Manninen, M., & Pösö, A. R. (2011). Effects of age and concentrate feeding on the expression of MCT 1 and CD147 in the gastrointestinal tract of goats and Hereford finishing beef bulls. *Research in Veterinary Science*, 90(2), 301-305.
- Koike, S., & Kobayashi, Y. (2009). Fibrolytic rumen bacteria: their ecology and functions. *Asian-Australasian Journal of Animal Sciences*, 22(1), 131-138.
- Koike, S., Yoshitani, S., Kobayashi, Y., & Tanaka, K. (2003). Phylogenetic analysis of fiber-associated rumen bacterial community and PCR detection of uncultured bacteria. *FEMS Microbiology Letters*, 229(1), 23-30.
- Koike, S.; Kobayashi, Y. (2001). Development and use of competitive PCR assays for the rumen cellulolytic bacteria: *Fibrobacter succinogenes*, *Ruminococcus albus* and *Ruminococcus flavefaciens*. *FEMS Microbiology Letters*, 204, 361-366.

- Komala, T. S., Mahadi, Y., Khairunnisak, M., & Ramlan, M. (2011). Studies on nutritionally-related blood metabolites: total protein and glucose levels in goats of Kinta and Hilir Perak district. *Malaysian Journal of Veterinary Research* 2(2):9–16.
- Koohmaraie, M., Kent, M. P., Shackelford, S. D., Veiseth, E., & Wheeler, T. L. (2002). Meat tenderness and muscle growth: is there any relationship? *Meat Science*, 62(3), 345-352.
- Krause, D. O., Denman, S. E., Mackie, R. I., Morrison, M., Rae, A. L., Attwood, G. T., & McSweeney, C. S. (2003). Opportunities to improve fiber degradation in the rumen: microbiology, ecology, and genomics. *FEMS Microbiology Reviews*, 27(5), 663-693.
- Krehbiel, C. R., Rust, S. R., Zhang, G., & Gilliland, S. E. (2003). Bacterial direct-fed microbials in ruminant diets: Performance response and mode of action. *Journal of Animal Science*, 81, 120-132.
- Kristensen, N. B., Sloth, K. H., Højberg, O., Spliid, N. H., Jensen, C., & Thøgersen, R. (2010). Effects of microbial inoculants on corn silage fermentation, microbial contents, aerobic stability, and milk production under field conditions. *Journal of Dairy Science*, 93(8), 3764-3774.
- Kung Jr, L., Carmean, B. R., & Tung, R. S. (1990). Microbial inoculation or cellulase enzyme treatment of barley and vetch silage harvested at three maturities. *Journal of Dairy Science*, 73(5), 1304-1311.
- Kung Jr, L., Chen, J. H., Kreck, E. M., & Knutsen, K. (1993). Effect of microbial inoculants on the nutritive value of corn silage for lactating dairy cows. *Journal of Dairy Science*, 76(12), 3763-3770.
- Kung Jr, L., Shaver, R. D., Grant, R. J., & Schmidt, R. J. (2018). Silage review: Interpretation of chemical, microbial, and organoleptic components of silages. *Journal of Dairy Science*, 101(5), 4020-4033.
- Kung Jr, L., Taylor, C. C., Lynch, M. P., & Neylon, J. M. (2003). The effect of treating alfalfa with *Lactobacillus buchneri* 40788 on silage fermentation, aerobic stability, and nutritive value for lactating dairy cows. *Journal of Dairy Science*, 86(1), 336-343.
- Kung, L., & Charley, R. (2010). Forage inoculants: Key silage management topics.
- Kuzinski, J., Zitnan, R., Viergutz, T., Legath, J., & Schweigel, M. (2011). Altered Na⁺/K⁺-ATPase expression plays a role in rumen epithelium adaptation in sheep fed hay ad libitum or a mixed hay/concentrate diet. *Veterinarni Medicina*, 56(1), 35-47.
- Lane, D. J. (1991). 16S/23S rRNA sequencing. *Nucleic acid techniques in bacterial systematics*, 115-175. Edited by Stackebrandt E., Goodfellow M. Chichester: Wiley

- Lane, M. A., Baldwin IV, R. L., & Jesse, B. W. (2002). Developmental changes in ketogenic enzyme gene expression during sheep rumen development. *Journal of Animal Science*, 80(6), 1538-1544.
- Lange, M., Westermann, P., & Ahring, B. K. (2005). Archaea in protozoa and metazoa. *Applied Microbiology and Biotechnology*, 66(5), 465-474.
- Larue, R., Yu, Z., Parisi, V. A., Egan, A. R., & Morrison, M. (2005). Novel microbial diversity adherent to plant biomass in the herbivore gastrointestinal tract, as revealed by ribosomal intergenic spacer analysis and rrs gene sequencing. *Environmental Microbiology*, 7(4), 530-543.
- Lawrie, R., & Ledward, D. (2006). *Lawrie's Meat Science*. (7th ed.), Woodhead Publishing, Cambridge
- Ledward, D. A. (1985). Post-slaughter influences on the formation of metmyoglobin in beef muscles. *Meat Science*, 15(3), 149-171.
- Li M, Zi X, Zhou H, Lv R, Tang J, Cai Y (2019) Silage fermentation and ruminal degradation of cassava foliage prepared with microbial additive. *AMB Express* 9, 180.
- Li, F., Xie, G., Huang, J., Zhang, R., Li, Y., Zhang, M., ... & Qu, C. (2017). Os CESA 9 conserved-site mutation leads to largely enhanced plant lodging resistance and biomass enzymatic saccharification by reducing cellulose DP and crystallinity in rice. *Plant Biotechnology Journal*, 15(9), 1093-1104.
- Li, M., Zhou, H., Zi, X., & Cai, Y. (2017). Silage fermentation and ruminal degradation of stylo prepared with lactic acid bacteria and cellulase. *Animal Science Journal*, 88(10), 1531-1537.
- Li, P., Wang, T., Mao, Y., Zhang, Y., Niu, L., Liang, R., ... & Luo, X. (2014). Effect of ultimate pH on post mortem myofibrillar protein degradation and meat quality characteristics of Chinese yellow crossbreed cattle. *The Scientific World Journal*, 2014. 1-8.
- Lindahl, G., Lundström, K., & Tornberg, E. (2001). Contribution of pigment content, myoglobin forms and internal reflectance to the colour of pork loin and ham from pure breed pigs. *Meat Science*, 59(2), 141-151.
- Livak, K. J., & Schmittgen, T. D. (2001). Analysis of relative gene expression data using real-time quantitative PCR and the $2^{-\Delta\Delta CT}$ method. *Methods*, 25(4), 402-408.
- Loor, J. J., Elolimy, A. A., & McCann, J. C. (2016). Dietary impacts on rumen microbiota in beef and dairy production. *Animal Frontiers*, 6(3), 22-29.
- MacDougall, D. B. (1982). Changes in the colour and opacity of meat. *Food Chemistry*, 9(1-2), 75-88.

- Madan, J., Sindhu, S., Gupta, M., & Kumar, S. (2016). Hematobiochemical profile and mineral status in growing beetal goat kids. *Journal of Cell and Tissue Research*, 16(1).
- Mahesh, M. S., & Mohini, M. (2013). Biological treatment of crop residues for ruminant feeding: A review. *African Journal of Biotechnology*, 12(27), 4221-4231
- Mahgoub, O., Kadim, I. T., Tageldin, M. H., Al-Marzooqi, W. S., Khalaf, S. Q., & Ali, A. A. (2008). Clinical profile of sheep fed non-conventional feeds containing phenols and condensed tannins. *Small Ruminant Research*, 78(1-3), 115-122.
- Malecky, M., Ghadbeigi, M., Aliarabi, H., Bahari, A. A., & Zaboli, K. (2017). Effect of replacing alfalfa with processed potato vines on growth performance, ruminal and total tract digestibility and blood metabolites in fattening lambs. *Small Ruminant Research*, 146, 13-22.
- Mamuad, L. L., Kim, S. H., Biswas, A. A., Yu, Z., Cho, K. K., Kim, S. B., ... & Lee, S. S. (2019). Rumen fermentation and microbial community composition influenced by live *Enterococcus faecium* supplementation. *Amb Express*, 9(1), 1-12.
- Marichal, A., Castro, N., Capote, J., Zamorano, M. J., & Argüello, A. (2003). Effects of live weight at slaughter (6, 10 and 25 kg) on kid carcass and meat quality. *Livestock Production Science*, 83(2-3), 247-256.
- Marinova, P., Banskalieva, V., Alexandrov, S., Tzvetkova, V., & Stanchev, H. (2001). Carcass composition and meat quality of kids fed sunflower oil supplemented diet. *Small Ruminant Research*, 42(3), 217-225.
- Matthews, C., Crispie, F., Lewis, E., Reid, M., O'Toole, P. W., & Cotter, P. D. (2019). The rumen microbiome: a crucial consideration when optimising milk and meat production and nitrogen utilisation efficiency. *Gut Microbes*, 10(2), 115-132.
- Mazhangara, I. R., Chivandi, E., Mupangwa, J. F., & Muchenje, V. (2019). The potential of goat meat in the red meat industry. *Sustainability*, 11(13), 3671.
- Mbassa, G. K., & Poulsen, J. S. D. (1993). Reference ranges for clinical chemical values in Landrace goats. *Small Ruminant Research*, 10(2), 133-142.
- McGill, M. R. (2016). The past and present of serum aminotransferases and the future of liver injury biomarkers. *EXCLI Journal*, 15, 817-828.
- McSweeney, C., & Mackie, R. (2012). Commission on genetic resources for food and agriculture. *Micro-organisms and ruminant digestion: State of knowledge, trends and future prospects. Background Study Paper (FAO)*, 61, 1-62.
- Meat and Livestock Australia (2018). The Malaysian market.

- Menke, K. H. (1988). Estimation of the energetic feed value obtained from chemical analysis and in vitro gas production using rumen fluid. *Animal Research and Development*, 28, 7-55.
- Merck Veterinary Manual (2016).
- Milletich, S. (2002). *Circadiane Rhythmen hämatologischer und chemischer Blutparameter beim Hengst*. Dissertation Veterinary Medicine. Wien: Universität Wien
- Mirimo, D., & Shamsudin, M. N. (2018). Price relations between Malaysia rice sector and selected ASEAN countries. *International Journal of Community Development and Management Studies*, 2, 131-144.
- Mohamed-Nour, I. A., & Abu-Goukh, A. A. (2010). Effect of Ethrel in aqueous solution and ethylene released from Ethrel on guava fruit ripening. *Agriculture and Biology Journal of North America*, 1(3), 232-237.
- Mohammed, S. A., Razzaque, M. A., Omar, A. E., Albert, S., & Al-Gallaf, W. M. (2016). Biochemical and hematological profile of different breeds of goat maintained under intensive production system. *African Journal of Biotechnology*, 15(24), 1253-1257.
- Morand-Fehr, P., Boutonnet, J. P., Devendra, C., Dubeuf, J. P., Haenlein, G. F. W., Holst, P., ... & Capote, J. (2004). Strategy for goat farming in the 21st century. *Small Ruminant Research*, 51(2), 175-183.
- Morgavi, D. P., Forano, E., Martin, C., & Newbold, C. J. (2010). Microbial ecosystem and methanogenesis in ruminants. *Animal*, 4(7), 1024-1036.
- Moss, A. R., Jouany, J. P., & Newbold, J. (2000, May). Methane production by ruminants: its contribution to global warming. In *Annales de zootechnie* 49(3), 231-253).
- Moyo, M., & Nsahlai, I. (2018). Rate of passage of digesta in ruminants; Are goats different. *Goat Science; Kukovics, S., Ed.; IntechOpen: London, UK*, 39-74.
- Muchenje, V., Dzama, K., Chimonyo, M., Raats, J.G., and Strydom, P. E. (2008). Meat quality of Nguni, Bonsmara and Aberdeen Angus steers raised on natural pasture in the Eastern Cape, South Africa. *Meat Science*, 79, 20–28.
- Muck, R. (2013). Recent advances in silage microbiology. *Agricultural and Food Science*, 22(1), 3-15
- Muck, R. E. (2010, February). Silage additives and management issues. In *Proceedings of Idaho Alfalfa Forage Conference, Best Western Burley, Idaho, USA* (pp. 49-55).

- Muck, R. E., Nadeau, E. M. G., McAllister, T. A., Contreras-Govea, F. E., Santos, M. C., & Kung Jr, L. (2018). Silage review: Recent advances and future uses of silage additives. *Journal of Dairy Science*, *101*(5), 3980-4000.
- Muller, F., Huber, K., Pfannkuche, H., Aschenbach, J. R., Breves, G., & Gabel, G. (2002). Transport of ketone bodies and lactate in the sheep ruminal epithelium by monocarboxylate transporter 1. *American Journal of Physiology-Gastrointestinal and Liver Physiology*, *283*(5), G1139-G1146.
- Mupangwa, J. F., Ngongoni, N. T., & Hamudikuwanda, H. (2006). The effect of stage of growth and method of drying fresh herbage on chemical composition of three tropical herbaceous forage legumes. *Tropical and Subtropical Agroecosystems*, *6*(1), 23-30.
- Murillo, M., Herrera, E., Carrete, F. O., Ruiz, O., & Serrato, J. S. (2012). Chemical composition, in vitro gas production, ruminal fermentation and degradation patterns of diets by grazing steers in native range of North Mexico. *Asian-Australasian Journal of Animal Sciences*, *25*(10), 1395.
- Mushi, D. E., Safari, J., Mtenga, L. A., Kifaro, G. C., & Eik, L. O. (2009). Effects of concentrate levels on fattening performance, carcass and meat quality attributes of Small East African× Norwegian crossbred goats fed low quality grass hay. *Livestock Science*, *124*(1-3), 148-155.
- Nadeau, E. M. G., Buxton, D. R., Russell, J. R., Allison, M. J., & Young, J. W. (2000). Enzyme, bacterial inoculant, and formic acid effects on silage composition of orchardgrass and alfalfa. *Journal of Dairy Science*, *83*(7), 1487-1502.
- Nazifi, S., Saeb, M., & Abedi, M. (2003). Serum lipid profiles and their correlation with thyroid hormones in clinically healthy Turkoman horses. *Comparative Clinical Pathology*, *12*(1), 49-52.
- Ni, K., Wang, Y., Pang, H., & Cai, Y. (2014). Effect of cellulase and lactic acid bacteria on fermentation quality and chemical composition of wheat straw silage. *American Journal of Plant Sciences*, *5*, 1877-1884.
- Nkosi, B. D., Meeske, R., Langa, T., & Thomas, R. S. (2011). Effects of bacterial silage inoculants on whole-crop maize silage fermentation and silage digestibility in rams. *South African Journal of Animal Science*, *41*(4), 350-359.
- Nkosi, B. D., Vadlani, P. V., Brijwani, K., Nanjunda, A., & Meeske, R. (2012). Effects of bacterial inoculants and an enzyme on the fermentation quality and aerobic stability of ensiled whole-crop sweet sorghum. *South African Journal of Animal Science*, *42*(3), 232-240.
- Nori, H., Halim, R. A., & Ramlan, M. F. (2008). Effects of nitrogen fertilization management practice on the yield and straw nutritional quality of commercial rice varieties. *Malaysian Journal of Mathematical Sciences*, *2*(2), 61-71.

- Nsereko, V. L., Smiley, B. K., Rutherford, W. M., Spielbauer, A., Forrester, K. J., Hettinger, G. H., ... & Harman, B. R. (2008). Influence of inoculating forage with lactic acid bacterial strains that produce ferulate esterase on ensilage and ruminal degradation of fiber. *Animal Feed Science and Technology*, 145(1-4), 122-135.
- Nurjana, D. J., & Suharti, S. (2016). Improvement of napier grass silage nutritive value by using inoculant and crude enzymes from *Trichoderma reesei* and its effect on in vitro rumen fermentation. *Media Peternakan*, 39(1), 46-52.
- Offer, G. (1988a). The structural basis of water-holding capacity in meat. Part 2. Drip losses. R. Lawrie (Ed.), *Developments in Meat Science*, Vol. 4, Elsevier Science Publications, London, pp. 61-171.
- Offer, G., & Cousins, T. (1992). The mechanism of drip production: formation of two compartments of extracellular space in muscle post mortem. *Journal of the Science of Food and Agriculture*, 58(1), 107-116.
- Offer, G., & Knight, P. (1988b). Structural basis of water-holding in meat. 2. Drip losses. R. Lawrie (Ed.), *Developments in Meat Science*, Vol. 4, Elsevier Science Publications, London, pp. 173
- Okonkwo, J. C., Omeje, I. S., Okonkwo, I. F., & Umeghalu, I. C. E. (2010). Effects of breed, sex and source within breed on the blood bilirubin, cholesterol and glucose concentrations of Nigerian goats. *Pakistan Journal of Nutrition*, 9(2), 120-124.
- Okpanachi, U., Okpanachi, G. A. C., Kaye, J., Agu, C. I., & Odah, E. O. (2019). Haematological profile and serum biochemistry of West African Dwarf goats fed sun-dried yellow cashew pulp based diets. *Journal of Applied Sciences*, 19(4), 319-324.
- Olafadehan, O. A. (2011). Changes in haematological and biochemical diagnostic parameters of Red Sokoto goats fed tannin-rich *Pterocarpus erinaceus* forage diets. *Veterinarski arhiv*, 81(4), 471-483.
- Omer, H. A. A., Ali, F. A. F., & Gad, S. M. (2012). Replacement of clover hay by biologically treated corn stalks in growing sheep rations. *Journal of Agricultural Science (Toronto)*, 4(2), 257-268.
- Ørskov, E. R., & McDonald, Y. (1979). The estimation of protein degradability in the rumen from determining the digestibility of feeds in the rumen. *The Journal of Agricultural Science*, 92, 499-503.
- Oskoueian, E., Jafari, S., Noura, R., Jahromi, M. F., Meng, G. Y., & Ebrahimi, M. (2019). Application of different types of lactic acid bacteria inoculant on ensiled rice straw; effects on silage quality, rumen fermentation, methane production and microbial population. *BioRxiv*, 612556.

- Ozduven ML, Koc F, Akay V (2017) Effects of bacterial inoculants and enzymes on the fermentation, aerobic stability and in vitro organic matter digestibility characteristics of sunflower silages. *Pakistan Journal of Nutrition* 16, 22-27.
- Panadi, M., Mohamed, W. Z., Rusli, N. D., & Mat, K. (2018). Effects of medicated and non-medicated multi-nutrient block supplementation on gastrointestinal parasite infestation and blood hematological parameters of lactating Saanen goats. *Sains Malaysiana*, 47(7), 1447-1453.
- Paraskevopoulou, C., Theodoridis, A., Johnson, M., Ragkos, A., Arguile, L., Smith, L., ... & Arsenos, G. (2020). Sustainability assessment of goat and sheep farms: a comparison between European countries. *Sustainability*, 12(8), 3099.
- Pathare, P. B., & Roskilly, A. P. (2016). Quality and energy evaluation in meat cooking. *Food Engineering Reviews*, 8(4), 435-447.
- Patra, A., Park, T., Kim, M., & Yu, Z. (2017). Rumen methanogens and mitigation of methane emission by anti-methanogenic compounds and substances. *Journal of Animal Science and Biotechnology*, 8(1), 13.
- Pearson, A. M., & Gillett, T. A. (1996). *Processed Meats*. 3rd edn. Chapman & Hall, New York.
- Pérez, J. M., González, F. J., Granados, J. E., Pérez, M. C., Fandos, P., Soriguer, R. C., & Serrano, E. (2003). Hematologic and biochemical reference intervals for Spanish ibex. *Journal of Wildlife Diseases*, 39(1), 209-215.
- Peripolli, V., Barcellos, J. O. J., Prates, Ê. R., McManus, C., Stella, L. A., Camargo, C. M., ... & Bayer, C. (2017). Additives on in vitro ruminal fermentation characteristics of rice straw. *Revista Brasileira de Zootecnia*, 46, 240-250.
- Pieniak-Lendzion, K., Niedziolka, R., Horoszewicz, E., & Brokowska, T. (2008). Evaluation of slaughter value and physicochemical attributes of goat meat. *Electronic Journal of Polish Agriculture University*, 11, 11-116.
- Pino, F., Mitchell, L. K., Jones, C. M., & Heinrichs, A. J. (2018). Comparison of diet digestibility, rumen fermentation, rumen rate of passage, and feed efficiency in dairy heifers fed ad-libitum versus precision diets with low and high quality forages. *Journal of Applied Animal Research*, 46(1), 1296-1306.
- Polat, U., Gencoglu, H., & Turkmen, I. I. (2009). The effects of partial replacement of corn silage on biochemical blood parameters in lactating primiparous dairy cows. *Veterinárni Medicína*, 54(9), 407-411.
- Ponsuksili, S., Jonas, E., Murani, E., Phatsara, C., Srikanchai, T., Walz, C., ... & Wimmers, K. (2008). Trait correlated expression combined with expression QTL analysis reveals biological pathways and candidate genes affecting water holding capacity of muscle. *BMC Genomics*, 9(1), 1-14.

- Pratiwi, N. W., Murray, P. J., & Taylor, D. G. (2007). Feral goats in Australia: A study on the quality and nutritive value of their meat. *Meat Science*, 75(1), 168-177.
- Pratt DS. Liver chemistry and function test. In: Feldman M, Friedman LS, Brandt LJ. (eds.) Sleisenger and Fordran's gastrointestinal and liver diseases. Philadelphia: Elsevier Publishers; 2010, p. 118-124.
- Purslow, P. P. (2005). Intramuscular connective tissue and its role in meat quality. *Meat Science*, 70(3), 435-447.
- Queiroz, O. C. M., Adesogan, A. T., Arriola, K. G., & Queiroz, M. F. S. (2012). Effect of a dual-purpose inoculant on the quality and nutrient losses from corn silage produced in farm-scale silos. *Journal of Dairy Science*, 95(6), 3354-3362.
- Queiroz, O. C. M., Kim, S. C., & Adesogan, A. T. (2012). Effect of treatment with a mixture of bacteria and fibrolytic enzymes on the quality and safety of corn silage infested with different levels of rust. *Journal of Dairy Science*, 95(9), 5285-5291.
- Rabelo, C. H. S., Basso, F. C., Lara, E. C., Jorge, L. G. O., Härter, C. J., Mari, L. J., & Reis, R. A. (2017). Effects of *Lactobacillus buchneri* as a silage inoculant or probiotic on in vitro organic matter digestibility, gas production and volatile fatty acids of low dry-matter whole-crop maize silage. *Grass and Forage Science*, 72(3), 534-544.
- Ramprabhu, R., Chellapandian, M., Balachandran, S., & Rajeswar, J. J. (2010). Influence of age and sex on blood parameters of Kanni goats in Tamil Nadu. *The Indian Journal of Small Ruminants*, 16(2), 249-251.
- Raza, A., Razzaq, A., Mehmood, S. S., Zou, X., Zhang, X., Lv, Y., & Xu, J. (2019). Impact of climate change on crops adaptation and strategies to tackle its outcome: A review. *Plants*, 8(2), 34.
- Reis, S. T. D., Lima, M. V. G., Sales, E. C. J. D., Monção, F. P., Rigueira, J. P. S., & Santos, L. D. T. (2016). Fermentation kinetics and in vitro degradation rates of grasses of the genus *Cynodon*. *Acta Scientiarum. Animal Sciences*, 38(3), 249-254.
- Rentfrow, G., Brewer, M. S., Carr, T. R., Berger, L. A., & McKeith, F. K. (2004). The effects of feeding elevated levels of vitamins D3 and E on beef quality. *Journal of Muscle Foods*, 15(3), 205-223.
- Rinne, M., Nousiainen, J., & Huhtanen, P. (2009). Effects of silage protein degradability and fermentation acids on metabolizable protein concentration: A meta-analysis of dairy cow production experiments. *Journal of Dairy Science*, 92(4), 1633-1642.
- Roeder, B., Ramsey, W. S., Hafley, B. S., Miller, R. K., Davis, E. E., & Branson, R. (1999). Consumer acceptance and quality profile of goat meat. Final Report to Texas Department of Agriculture, Austin, TX

- Rowghani E & Zamiri M (2009) The effects of a microbial inoculant and formic acid as silage additives on chemical composition, ruminal degradability and nutrient digestibility of corn silage in sheep. *Iranian Journal of Veterinary*. 10(2), 110-118.
- Rowghani, E., Zamiri, M. J., Khorvash, M., & Abdollahipanah, A. (2008). The effects of *Lactobacillus plantarum* and *Propionibacterium acidipropionici* on corn silage fermentation, ruminal degradability and nutrient digestibility in sheep. *Iranian Journal of Veterinary Research*, 9(4), 308-315.
- Sahoo, A. (2018). Silage for Climate Resilient Small Ruminant Production. *Ruminants: The Husbandry, Economic and Health Aspects*, 11.
- Saleem, A. M., Zanouny, A. I., & Singer, A. M. (2017). Growth performance, nutrients digestibility, and blood metabolites of lambs fed diets supplemented with probiotics during pre-and post-weaning period. *Asian-Australasian Journal of Animal Sciences*, 30(4), 523.
- SAS (2012). Statistical Analysis System package (SAS) Version 9.4 software. SAS Institute Inc., Cary, NC, USA.
- Satter, L. D., & Slyter, L. L. (1974). Effect of ammonia concentration on rumen microbial protein production in vitro. *British Journal of Nutrition*, 32(2), 199-208.
- Saxena, M., Saxena, J., Nema, R., Singh, D., & Gupta, A. (2013). Phytochemistry of medicinal plants. *Journal of Pharmacognosy and Phytochemistry*, 1(6), 168-182.
- Sazili, A.Q., Parr, T., Sensky, P.L., Jones, S.W., Bardsley, R.G. and Buttery, P.J. (2005). The relationship between slow and fast myosin heavy chain content, calpastatin and meat tenderness in different ovine skeletal muscles. *Meat Science* 69(1): 17-25.
- Schmidt RJ, Hu W, Mills JA, Kung JrL (2009) The development of lactic acid bacteria and *Lactobacillus buchneri* and their effects on the fermentation of alfalfa silage. *Journal of Dairy Science* 92, 5005-5010.
- Sebsibe, A. (2008). Sheep and goat meat characteristics and quality. *Sheep and Goat Production Handbook for Ethiopia. Ethiopian Sheep and Goats Productivity Improvement Program (ESGPIP), Addis Ababa, Ethiopia*. pp323-328.
- Sehested, J., Diernæs, L., Møller, P. D., & Skadhauge, E. (1999). Ruminal transport and metabolism of short-chain fatty acids (SCFA) in vitro: effect of SCFA chain length and pH. *Comparative Biochemistry and Physiology Part A: Molecular & Integrative Physiology*, 123(4), 359-368.
- Sen, A. R., Santra, A., & Karim, S. A. (2004). Carcass yield, composition and meat quality attributes of sheep and goat under semiarid conditions. *Meat Science*, 66(4), 757-763.

- Senger, C. C. D., Mühlbach, P. R. F., Sánchez, L. M. B., Netto, D. P., & Lima, L. D. D. (2005). Chemical composition and *in vitro* digestibility of maize silages with different maturities and packing densities. *Ciência Rural*, 35(6), 1393-1399.
- Sentandreu, M. A., Coulis, G., & Ouali, A. (2002). Role of muscle endopeptidases and their inhibitors in meat tenderness. *Trends in Food Science & Technology*, 13(12), 400-421.
- Shackelford, S. D., Morgan, J. B., Cross, H. R., & Savell, J. W. (1991). Identification of threshold levels for Warner-Bratzler shear force in beef top loin steaks. *Journal of Muscle Foods*, 2(4), 289-296.
- Sharma, A., Kumar, V., Shahzad, B., Tanveer, M., Sidhu, G. P. S., Handa, N., ... & Thukral, A. K. (2019). Worldwide pesticide usage and its impacts on ecosystem. *SN Applied Sciences*, 1(11), 1-16.
- Shen Z, Seyfert HM, Löhrike B, Schneider F, Zitnan R, Chudy A, Kuhla S, Hammon H, Blum JW, Martens H, Hagemeister H, Voigt J (2004) An energy-rich diet causes rumen papillae proliferation associated with more IGF type 1 receptors and increased plasma IGF-1 concentrations in young goats. *The Journal of Nutrition*, 134(1), 11-17
- Sheperd, A. C., & Kung Jr, L. (1996). Effects of an enzyme additive on composition of corn silage ensiled at various stages of maturity. *Journal of Dairy Science*, 79(10), 1767-1773.
- Shrivastava, B., Nandal, P., Sharma, A., Jain, K. K., Khasa, Y. P., Das, T. K., ... & Kuhad, R. C. (2012). Solid state bioconversion of wheat straw into digestible and nutritive ruminant feed by *Ganoderma* sp. rckk02. *Bioresource Technology*, 107, 347-351.
- Silva, J. D. L., Guim, A., Carvalho, F. F. R. D., Mattos, C. W., Garcia, D. A., Silva, E. R. R. D., ... & Vasconcelos, J. L. D. A. (2015). Metabolic profile of lactating goats fed integral mango meal. *Revista Brasileira de Saúde e Produção Animal*, 16(4), 885-892.
- Simela, L., & Merkel, R. (2008). The contribution of chevon from Africa to global meat production. *Meat Science*, 80(1), 101-109.
- Singh, P. K., Shrivastava, N., & Ojha, B. K. (2019). "Enzymes in the Meat Industry". In *Enzymes in Food Biotechnology*, Elsevier Inc (pp. 111-128).
- Sirois, M. (1995). Veterinary clinical laboratory procedure. Mosby year book. *Inc. St Louis, Missouri, USA*.
- Solaiman, S., Thomas, J., Dupre, Y., Min, B. R., Gurung, N., Terrill, T. H., & Haenlein, G. F. W. (2010). Effect of feeding sericea lespedeza (*Lespedeza cuneata*) on growth performance, blood metabolites, and carcass characteristics of Kiko crossbred male kids. *Small Ruminant Research*, 93(2-3), 149-156.

- Solorzano, L. (1969). Determination of ammonia in natural waters by the phenylhypochlorite method 1 1 This research was fully supported by US Atomic Energy Commission Contract No. ATS (11-1) GEN 10, PA 20. *Limnology and Oceanography*, 14(5), 799-801.
- Soul, W., Mupangwa, J., Muchenje, V., & Mpendulo, T. C. (2019). Biochemical indices and hematological parameters of goats fed lablab purpureus and vigna unguiculata as supplements to a chloris gayana basal diet. *Veterinary and Animal Science*, 8, 100073.
- Stevens, C. E., & Stettler, B. K. (1966). Factors affecting the transport of volatile fatty acids across rumen epithelium. *American Journal of Physiology-Legacy Content*, 210(2), 365-372.
- Strzyżewski, T., Bilka, A., & Krystofiak, K. (2008). Correlation between pH value of meat and its colour. *Nauka Przyr Technol*, 2, 12.
- Sucu, E., & Filya, I. (2006) Effects of homo fermentative lactic acid bacterial inoculants on the fermentation and aerobic stability characteristics of low dry matter corn silages. *Turkish Journal of Veterinary and Animal Sciences*, 30, 83-88.
- Sutton, J. D., McGilliard, A. D., & Jacobson, N. L. (1963). Functional development of rumen mucosa. Absorptive ability. *Journal of Dairy Science*, 46(5), 426-436.
- Sutton, J. D. (1985). Digestion and absorption of energy substrates in the lactating cow. *Journal of Dairy Science*, 68(12), 3376-3393.
- Swatland, H. J. (1982). The challenges of improving meat quality. *Canadian Journal of Animal Science*, 62(1), 15-24.
- Sylvester, J. T., Karnati, S. K. R., Yu, Z., Newbold, C. J., & Firkins, J. L. (2005). Evaluation of a real-time PCR assay quantifying the ruminal pool size and duodenal flow of protozoal nitrogen. *Journal of Dairy Science*, 88(6), 2083-2095.
- Tahuk, P. K., & Bira, G. F. (2020). Carcass and meat characteristics of male Kacang goat fattened by complete silage. *Veterinary World*, 13(4), 706.
- Tajik, J., & Nazifi, S. (2011). Serum concentrations of lipids and lipoproteins and their correlations together and with thyroid hormones in Iranian water buffalo (*Bulbalus bulbalis*). *Asian Journal of Animal Sciences*, 5(3), 196-201.
- Tamai, I., Sai, Y., Ono, A., Kido, Y., Yabuuchi, H., Takanaga, H., ... & Tsuji, A. (1999). Immunohistochemical and functional characterization of pH-dependent intestinal absorption of weak organic acids by the monocarboxylic acid transporter MCT1. *Journal of Pharmacy and Pharmacology*, 51(10), 1113-1121.

- Tanweer, F. A., Rafii, M. Y., Sijam, K., Rahim, H. A., Ahmed, F., Ashkani, S., & Latif, M. A. (2015). Introgression of blast resistance genes (putative Pi-b and Pi-kh) into elite rice cultivar MR219 through marker-assisted selection. *Frontiers in plant science*, 6, 1002.
- Tekliye, L., Mekuriaw, Y., Asmare, B., & Mehret, F. (2018). Nutrient intake, digestibility, growth performance and carcass characteristics of Farta sheep fed urea-treated rice straw supplemented with graded levels of dried Sesbania sesban leaves. *Agriculture and Food Security*, 7(1), 1-10.
- Thomas, M. E., Foster, J. L., McCuiston, K. C., Redmon, L. A., & Jessup, R. W. (2013). Nutritive value, fermentation characteristics, and in situ disappearance kinetics of sorghum silage treated with inoculants. *Journal of Dairy Science*, 96(11), 7120-7131.
- Thompson, J. (2002). Managing meat tenderness. *Meat Science*, 62(3), 295-308.
- Thornton, P. K. (2010). Livestock production: recent trends, future prospects. *Philosophical Transactions of the Royal Society B: Biological Sciences*, 365(1554), 2853-2867.
- Thorsteinsson, M., Martin, H. L., Larsen, T., Sehested, J., & Vestergaard, M. (2020). The effects of supplementation of yeast (*Saccharomyces cerevisiae*) and postbiotic from *Lactobacillus acidophilus* on the health and growth performance of young Jersey heifer calves. *Journal of Animal and Feed Sciences*, 29(3), 224-233.
- Tian, J., Yu, Y., Yu, Z., Shao, T., Na, R., & Zhao, M. (2014). Effects of lactic acid bacteria inoculants and cellulase on fermentation quality and in vitro digestibility of *Leymus chinensis* silage. *Grassland Science*, 60(4), 199-205
- Troy, D. J., & Kerry, J. P. (2010). Consumer perception and the role of science in the meat industry. *Meat Science*, 86(1), 214-226
- Van Soest, P. J. (1994). *Nutritional ecology of the ruminant*. Cornell University Press. USA.
- Van Soest, P. V., Robertson, J. B., & Lewis, B. (1991). Methods for dietary fiber, neutral detergent fiber, and nonstarch polysaccharides in relation to animal nutrition. *Journal of Dairy Science*, 74(10), 3583-3597.
- Van Vuuren, A. M., Bergsma, K., Frol-Kramer, F., & Van Beers, J. A. C. (1989). Effects of addition of cell wall degrading enzymes on the chemical composition and the *in sacco* degradation of grass silage. *Grass and Forage Science*, 44(2), 223-230.
- Vasanthi, C., Venkataramanujam, V., & Dushyanthan, K. (2007). Effect of cooking temperature and time on the physico-chemical, histological and sensory properties of female carabeef (buffalo) meat. *Meat Science*, 76(2), 274-280.

- Wallace, R. J., Rooke, J. A., Duthie, C. A., Hyslop, J. J., Ross, D. W., McKain, N., ... & Roehe, R. (2014). Archaeal abundance in post-mortem ruminal digesta may help predict methane emissions from beef cattle. *Scientific Reports*, 4, 5892.
- Wanapat, M., Cherdthong, A., Pakdee, P., and Wanapat, S. (2008). Manipulation of rumen ecology by dietary lemongrass (*Cymbopogon citratus* Stapf.) powder supplementation. *Journal of Animal Science*, 86(12), 3497–3503.
- Wang Y, He L, Xing Y, Zhou W, Pian R, Yang F, Zhang Q (2019) Bacterial diversity and fermentation quality of *Moringa oleifera* leaves silage prepared with lactic acid bacteria inoculants and stored at different temperatures. *Bioresource Technology* 284, 349-358.
- Wang, S., Guo, G., Li, J., Chen, L., Dong, Z., & Shao, T. (2018). Improvement of fermentation profile and structural carbohydrate compositions in mixed silages ensiled with fibrolytic enzymes, molasses and *Lactobacillus plantarum* MTD-1. *Italian Journal of Animal Science*, 18, 328-335.
- Wang, W., Wu, X., Chen, A., Xie, X., Wang, Y., & Yin, C. (2016). Mitigating effects of ex situ application of rice straw on CH₄ and N₂O emissions from paddy-upland coexisting system. *Scientific Reports*, 6, 37402.
- Warner, A. C. I. (1956). Proteolysis by rumen micro-organisms. *Microbiology*, 14(3), 749-762.
- Warner, R. D., Greenwood, P. L., Pethick, D. W., & Ferguson, D. M. (2010). Genetic and environmental effects on meat quality. *Meat Science*, 86(1), 171-183.
- Warriss, P. D. (2010). *Meat Science: An Introductory Text*. Cambridge: CABI publishing
- Webb, E., Casey, N., & Simela, L. (2005). Goat meat quality. *Small Ruminant Research*, 60, 153-166.
- Weinberg, Z. G., Muck, R. E., & Weimer, P. J. (2003). The survival of silage inoculant lactic acid bacteria in rumen fluid. *Journal of Applied Microbiology*, 94(6), 1066-1071.
- Weinberg, Z. G., Shatz, O., Chen, Y., Yosef, E., Nikbahat, M., Ben-Ghedalia, D., & Miron, J. (2007). Effect of lactic acid bacteria inoculants on in vitro digestibility of wheat and corn silages. *Journal of Dairy Science*, 90(10), 4754-4762.
- Weir, C. E. (1960). The science of meat and meat products. *American Meat Institute* Pag, 212.
- Whitley, N. C., Cazac, D., Rude, B. J., Jackson-O'Brien, D., & Parveen, S. (2009). Use of a commercial probiotic supplement in meat goats. *Journal of animal science*, 87(2), 723-728.

- Williams, A. G., & Coleman, G. S. (1985). Hemicellulose-degrading enzymes in rumen ciliate protozoa. *Current Microbiology*, 12(2), 85-90.
- Williams, A. G., & Coleman, G. S. (1997). The rumen protozoa. In *The Rumen Microbial Ecosystem* (pp. 73-139). Springer, Dordrecht.
- Williams, S. D., & Shinnars, K. J. (2012). Farm-scale anaerobic storage and aerobic stability of high dry matter sorghum as a biomass feedstock. *Biomass and Bioenergy*, 46, 309-316.
- Wolin, M. J., & Miller, T. L. (1988). In: Hobson PN, ed. Microbe–Microbe Interactions. Rumen Microbial Ecosystem. Essex, UK: Elsevier Science. Pp. 343-359.
- Wright, L. I., Scanga, J. A., Belk, K. E., Engle, T. E., Tatum, J. D., and Person, R. C. (2005). Benchmarking value in the pork supply chain: Characterization of US pork in the retail marketplace. *Meat Science*, 71(3), 451–463.
- Xing, T., Gao, F., Tume, R. K., Zhou, G., & Xu, X. (2019). Stress effects on meat quality: a mechanistic perspective. *Comprehensive reviews in food science and food safety*, 18(2), 380-401.
- Xu, Y., Li, Z., Moraes, L. E., Shen, J., Yu, Z., & Zhu, W. (2019). Effects of incremental urea supplementation on rumen fermentation, nutrient digestion, plasma metabolites, and growth performance in fattening lambs. *Animals*, 9(9), 652.
- Yan, L., Zhang, B., & Shen, Z. (2014). Dietary modulation of the expression of genes involved in short-chain fatty acid absorption in the rumen epithelium is related to short-chain fatty acid concentration and pH in the rumen of goats. *Journal of Dairy Science*, 97(9), 5668-5675.
- Yang, C., Zhang, J., Ahmad, A. A., Bao, P., Guo, X., Long, R., ... & Yan, P. (2019). Dietary energy levels affect growth performance through growth hormone and insulin-like growth factor 1 in Yak (*Bos grunniens*). *Animals*, 9(2), 39.
- Yitbarek, M. B., & Tamir, B. (2014). Silage additives. *Open Journal of Applied Sciences*, 2014. 4, 258-274.
- Zayadi, R. A. (2021). Current outlook of livestock industry in Malaysia and ways towards sustainability. *Journal of Sustainable Natural Resources*, 2(2), 1-11.
- Zhang, Y. G., Xin, H. S., & Hua, J. L. (2010). Effects of treating whole-plant or chopped rice straw silage with different levels of lactic acid bacteria on silage fermentation and nutritive value for lactating Holsteins. *Asian-Australasian Journal of Animal Sciences*, 23(12), 1601-1607.

- Zhang, Y., Zhao, X., Chen, W., Zhou, Z., Meng, Q., & Wu, H. (2019). Effects of adding various silage additives to whole corn crops at ensiling on performance, rumen fermentation, and serum physiological characteristics of growing-finishing cattle. *Animals*, 9(9), 695.
- Zhao, J., Dong, Z., Li, J., Chen, L., Bai, Y., Jia, Y., & Shao, T. (2019). Effects of lactic acid bacteria and molasses on fermentation dynamics, structural and nonstructural carbohydrate composition and in vitro ruminal fermentation of rice straw silage. *Asian-Australasian Journal of Animal Sciences*, 32(6), 783.
- Zhu, Y., Nishino, N., & Xusheng, G. (2010). Chemical changes during ensilage and in sacco degradation of two tropical grasses: Rhodes grass and guinea grass treated with cell wall-degrading enzymes. *Asian-Australasian Journal of Animal Sciences*, 24(2), 214-221.
- Zopollatto, M., Daniel, J. L. P., & Nussio, L. G. (2009). Microbial silage additives in Brazil: review of aspects of ensilage and animal performance. *Revista Brasileira de Zootecnia*, 38(Supplement), 170-189.