

## **UNIVERSITI PUTRA MALAYSIA**

DEVELOPMENT OF CHITOSAN-GRAPHENE OXIDE NANOCOMPOSITE FILMS FOR ACTIVE MARGARINE PACKAGING

**FOONG HAN LYN** 

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Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in the Fulfilment of the Requirements for the Degree of Doctor of Philosophy

October 2021

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Doctor of Philosophy

#### DEVELOPMENT OF CHITOSAN-GRAPHENE OXIDE NANOCOMPOSITE FILMS FOR ACTIVE MARGARINE PACKAGING

By

#### FOONG HAN LYN

October 2021

#### Chair : Nur Hanani Zainal Abedin, PhD Faculty : Food Science and Technology

Chitosan (CS) has gained significant attention as a food packaging material due to its film-forming ability, biocompatibility, and biodegradability. However, its applications have been limited by its weak mechanical properties and hydrophilicity. The aim of this study was to develop a chitosan-graphene oxide (CSGO) nanocomposite film with improved mechanical properties as well as water vapour, oxygen, and light barrier properties in comparison to pure CS film, for the antioxidant active packaging of palm olein-based margarine. In the first objective, GO samples with four different degrees of oxidation were synthesized by controlling the ratio of graphite to the oxidizing agent, potassium permanganate (KMnO<sub>4</sub>). The sample GO4, synthesized with a 1:8 w/w graphite:KMnO<sub>4</sub> ratio was embedded with abundant oxygen-containing groups, as supported by the Fourier-transform infrared (FTIR) and Raman spectra. The addition of GO4 into CS increased (p < 0.05) the mechanical strength and UV light barrier of the CS/GO4 composite. In the second objective, the effects of sonication time of GO4 (30, 60, and 120 min) and heating temperature of the films (30, 60, and 120 °C) on the structural and physical properties of the CSGO4 composites were investigated. After 120 min of sonication in a sonicator bath, graphene oxide nanosheets (GO120) of ~1 nm thick were obtained, as demonstrated using dynamic light scattering (DLS) technique and atomic force microscopy (AFM). The incorporation of GO120 decreased (p < 0.05) the light transmittance of CS films whereas heating the composites at 120 °C lowered (p <0.05) the water solubility and water vapour permeability (WVP). All of the films were completely decomposed within 28 days in a soil burial test. In the third objective, trisodium citrate (CIT) and sodium tripolyphosphate (TPP) solutions of different concentrations (0.5, 1.0, 2.0, and 3.0% w/v) were used as crosslinking agents for the films. Successful crosslinking was confirmed by FTIR spectroscopy. The hydrophilicity and light transmittance decreased (p < 0.05) with the increase in CIT and TPP. At 3.0% w/v, the elongation at break and tensile strength of the TPP-crosslinked CSGO films increased (p < 0.05) by 42 and 82%, respectively, outperforming CIT as a crosslinking agent. In the final objective, the effect of the concentrations of CS (1.5 and 2.0% w/v) and GO4 (0.5, 1.0, and 2.0% w/w CS) on the properties of nanocomposite films were

investigated. The WVP and oxygen permeability (OP) decreased (p < 0.05) by 43 and 54%, respectively. The antioxidant properties of the composite film increased (p < 0.05) with the concentration of GO4, as supported by the DPPH radical scavenging assay. The changes in the peroxide value (PV) and thiobarbituric acid reactive substances (TBARS) of the margarine samples were monitored for 30 d at 4 °C. For the margarine sample that was wrapped with the GOCS1.5 GO2.0 film (CS 1.5% w/v, GO 2.0% w/w CS), the PV and TBARS values were 36 and 79% lower (p < 0.05) in comparison to the low-density polyethylene films. The combination of these properties such as low WVP, OP, and light transmittance, as well as the radical scavenging activities suggests that the CS1.5 GO2.0 film could be a potential antioxidant active packaging for margarine.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Doktor Falsafah

#### PENBANGUNAN FILEM NANOKOMPOSIT KITOSAN-OKSIDA GRAFENA SEBAGAI PEMBUNGKUSAN AKTIF MAJERIN

Oleh

#### FOONG HAN LYN

#### Oktober 2021

# Pengerusi: Nur Hanani Zainal Abedin, PhDFakulti: Sains dan Teknologi Makanan

Kitosan (CS) telah mendapat perhatian sebagai bahan pembungkusan makanan kerana keupayaan pembentukan filem, keserasian bio, dan terbiodegradasinya. Walau bagaimanapun, aplikasi kitosan adalah terhad disebabkan oleh sifat mekanikal yang kurang memuaskan dan sifat hidrofiliknya. Projek ini bertujuan untuk membangunkan filem komposit kitosan-dioksida grafena (CSGO) dengan penambahbaikan sifat mekanikal serta pengurangan penembusan wap air, oksigen, dan cahaya, untuk dijadikan pembungkus aktif marjerin. Dalam objektif pertama, sampel-sampel GO yang mempunyai empat tahap pengoksidaan yang berbeza telah dihasilkan dengan penggunaan nisbah-nisbah yang berbeza untuk grafit dengan agen pengoksidaan, iaitu permanganat kalium (KMnO<sub>4</sub>). Spektroskopi inframerah transformasi Fourier (FTIR) dan spektroskopi Raman telah menunjukkan bahawa sampel GO4 yang dihasilkan dengan nisbah 1:8 w/w untuk grafit dan KMnO4 adalah kaya dengan kumpulankumpulan berfungsi yang mengandungi oksigen. Penambahan GO4 ke dalam CS telah meningkatkan (p < 0.05) kekuatan mekanikal dan mengurangkan (p < 0.05) kebolehan penembusan cahaya untuk filem CS. Untuk objektif kedua, kesan-kesan tempoh sonikasi (30, 60, dan 120 min) dan suhu pemanasan (30, 60, dan 120 °C) terhadap filem komposit telah dikaji. Ujian penyerakan cahaya dinamik (DLS) dan mikroskopi daya atom (AFM) menunjukkan bahawa kepingan nano oksida grafena dengan ketebalan ~1 nm (GO120) berjaya diperolehi selepas sonikasi selama 120 min dalam mandi ultrasonik. Penambahan GO120 mengurangkan (p < 0.05) kebolehan penembusan cahaya CS manakala pemanasan filem CSGO pada suhu 120 °C telah mengurangkan (p < 0.05) kelarutan dan kebolehtelapan wap air (WVP). Semua filem komposit terurai sepenuhnya di dalam tanah kompos dalam 28 hari. Dalam objektif ketiga, larutan trinatrium sitrat (CIT) dan natrium tripolifosfat (TPP) dengan kepekatan yang berbeza (0.5, 1.0. 2.0, dan 3.0% w/v) telah digunakan sebagai agen pautan silang untuk filem CS dan CSGO. Kejadian pautan silang telah disahkan melalui spektroskopi FTIR. Sifat hidrofilik dan kebolehan penembusan cahaya menurun (p < 0.05) dengan kenaikan kepekatan CIT dan TPP. Untuk filem CSGO yang dirawat dengan TPP, peratusan pemanjangan pada takat putus dan kekuatan mekanikal meningkat (p < 0.05) sebanyak 42 dan 82%, menunjukkan bahawa TPP adalah agen pautan silang yang lebih sesuai berbanding dengan CIT. Untuk

objektif yang terakhir, kesan-kesan kepekatan CS (1.5 dan 2.0% w/v) dan GO4 (0.5, 1.0, dan 2.0% w/w CS) telah dikaji. Untuk filem komposit, WVP dan kebolehtelapan oksigen (OP) menurun (p < 0.05) sebanyak 43 dan 54%. Aktiviti antioksida meningkat (p < 0.05) dengan kepekatan GO4 dan disokong oleh analisis aktiviti memerangkap radikal DPPH. Perubahan nilai peroksida (PV) dan uji asid tiobarbiturik (TBARS) untuk sampel marjerin dipantau selama 30 hari pada 4 °C. Sampel marjerin yang dibalut dengan filem CS1.5 GO2.0 menunjukkan penurunan (p < 0.05) PV dan TBARS sebanyak 36 dan 79% berbanding dengan filem polietilena berketumpatan rendah. Gabungan sifat-sifat filem CSGO seperti WVP, OP, dan penembusan cahaya yang rendah serta aktiviti memerangkap radikal DPPH membuktikan bahawa filem tersebut berpotensi untuk dijadikan pembungkusan aktif antioksida untuk marjerin.



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This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

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#### LIST OF ABBREVATIONS

ABTS	2,2'-azino-bis(3-ethylbenzothiazoline-6-sulfonic acid)
AOAC	Association of Analytical Communities
AOCS	American Oil Chemists' Society
APS	Average particle size
ASTM	American Society for Testing and Materials
СА	Crosslinking agent
CIE	International Commission on Illumination
CIT	Trisodium citrate
СМС	Carboxymethyl cellulose
CS	Chitosan 🚽 📕
CSGO	Chitosan-graphene oxide
DDA	Degree of deacetylation
DPPH	2,2-diphenyl-1-picrylhydrazyl
DSC	Differential scanning calorimeter
EAB	Elongation at break
EPA	Environmental Protection Agency
FAO	Food and Agriculture Organization
FDA	Food and Drug Administration
GBM	Graphene-based material
GNP	Graphene nanoplatelets
GO	Graphene oxide
GRAS	Generally Recognised as Safe
HDPE	High-density polyethylene
LBL	Layer-by-layer
LDPE	Low-density polyethylene
LLDPE	Linear low-density polyethylene
LOD	Limit of detection
MAP	Modified atmospheric packaging

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MDA	Malondialdehyde
MMT	Montmorillonite
OP	Oxygen permeability
PBAT	Polybutylene adipate terephthalate
PE	Polyethylene
PEC	Polyelectrolyte complexes
PEI	Polyethyleneimine
PET	Polyethylene terephthalate
PI	Polyimide
PLA	Polylactic acid
PMMA	Polymethyl methacrylate
PP	Polypropylene
PV	Peroxide value
PVA	Polyvinyl alcohol
rGO	Reduced graphene oxide
RH	Relative humidity
ROS	Reactive oxygen species
SEM	Scanning electron microscopy
SPC	Sustainable Packaging Coalition
TBA	Thiobarbituric acid
TBARS	Thiobarbituric acid reactive substances
TPP	Sodium tripolyphosphate
TS	Tensile strength
UV	Ultraviolet
WVP	Water vapour permeability
XPS	X-ray photoelectron spectroscopy
YM	Young's modulus

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#### **CHAPTER 1**

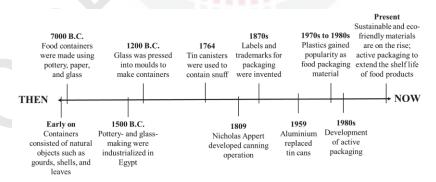
#### INTRODUCTION

#### 1.1 Background

Packaging is an essential part of the food system. It provides containment and protection to the contents against biological, chemical, and physical contaminations throughout the manufacturing and shipping processes, until they reach the final consumer (Robertson, 2010). Without packaging, the process of materials handling would be very inefficient, messy and costly. In addition, it would be almost impossible to convey information of a product to the respective consumers using modern marketing strategies.

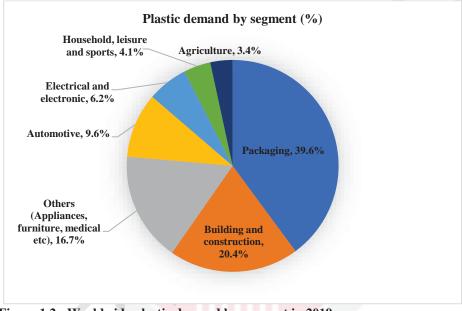
With the vast development of technologies, food packaging has evolved from simply a container to hold food to something that can play an active role in food quality (Figure 1.1). Active packaging improves the functionality of a package by incorporating active substance(s) into the packaging material or the package so that it could interact with the contents and environment to extend the shelf life while maintaining the organoleptic properties of food products (Brody, Bugusu, Han, Sand, & McHugh, 2008; Realini & Marcos, 2014). Active packaging can exist in the forms of antimicrobials, antioxidants, and oxygen scavengers, as well as modified atmosphere packaging (MAP), which is the modification of the internal gaseous composition of a package.

In addition to active packaging, sustainable and biodegradable packaging have garnered significant interest among consumers due to the increase in environmental awareness. This is due to the alarming growth of global petrochemical-based plastic demand and production in the recent decades (Piñeros-Hernandez, Medina-Jaramillo, López-Córdoba, & Goyanes, 2017).



**Figure 1.1 : A brief history of food packaging.** (Source: Berger & Welt, 2005; Frank et al., 2012; Robertson, 2019)

In fact, the worldwide production of petrochemical-based plastics in 2019 was 368 million tonnes, and packaging materials comprised the highest fraction of the total plastic demand, at approximately 40% (Figure 1.2) (*Plastics – the Facts 2020*, 2020). However, only 42% of the collected post-consumer packaging waste was recycled, according to a study conducted in Europe (*Plastics – the Facts 2020*, 2020). To make matters worse, a large fraction of the polymers used in the packaging industry are from non-renewable sources and therefore, exacerbating the environmental pollution. Biodegradable films produced from biopolymers such as polysaccharides, proteins, and waxes are promising alternatives to replace the plastic non-degradable films that presently plague the environment (dos Santos Caetano et al., 2018; Medina Jaramillo, Gutiérrez, Goyanes, Bernal, & Famá, 2016).



**Figure 1.2 : Worldwide plastic demand by segment in 2019.** (Source: *Plastics – the Facts 2020*, 2020)

Chitosan (CS) is a hydrophilic polysaccharide derived from the partial deacetylation of chitin, which is usually found in the exoskeleton of crustaceans (Islam, Shahruzzaman, Biswas, Sakib, & Rashid, 2020). Chitin is also present in some microorganisms such as bacteria, fungi, protozoan, and algae species, making it the second most abundant polysaccharide after cellulose (Lizardi-Mendoza, Argüelles Monal, & Goycoolea Valencia, 2016). Several chitin sources have been used for the production of CS, but the most common sources of chitin are the processing waste of shellfish (Gomes, Paschoalin, & Del Aguila, 2017). As a widely distributed and environmentally friendly biopolymer, CS has attracted significant attention as a packaging material due to its nontoxicity, biocompatibility, excellent film-forming ability, as well as antimicrobial and biodegradable properties (Ahmed, Mulla, Arfat, & Thai, 2017; Leceta, Guerrero, Ibarburu, Dueñas, & Caba, 2013; Zhong & Xia, 2008). In comparison to chitin, the amino and hydroxyl groups on CS facilitates the modification of CS by chemical reactions

(Lizardi-Mendoza et al., 2016), which enables researchers to tailor its properties for its usage.

The incorporation of nanoparticles into food packaging materials is known to improve packaging performances, such as mechanical properties as well as barrier properties in terms of light, water vapour, and gas (Souza & Fernando, 2016). One of the nanomaterials that is highlighted is graphene. Graphene is carbon in the form of a single-layered graphite. It possesses the thickness of one atomic layer and the carbon atoms are arranged in a honeycomb lattice structure in the sp<sup>2</sup> hybridization state. This two-dimensional structure was studied immensely by the theoreticians. The band structure of graphite was first calculated by Canadian physicist P. R. Wallace in 1947 (Wallace, 1947) and it was assumed not to exist freely until more than half a century later, when plane graphene was isolated by Andre Geim and Konstantin Novoselov from University of Manchester in 2004, which won them a Nobel Prize in 2010 (*The Nobel Prize in Physics 2010*, 2010).

Due to graphene's remarkable properties such as mechanical strength, flexibility, thermal and electrical conductivity, as well as high specific surface area, there have been high expectations for graphene to be incorporated into composite materials, as well as in biological applications (Mitura & Zarzycki, 2018). In addition, graphene also exhibits antimicrobial and antioxidant activities (Baali, Khecha, Bensouici, Speranza, & Hamdouni, 2019; Lalwani, Agati, Mahmud, & Sitharaman, 2016; Rajeswari & Prabu, 2018; Tayade, Borse, & Meshram, 2019), which makes it fitting as a component in active packaging. However, the bulk synthesis of defect free graphene sheets remains a challenge and is the greatest obstacle to commercialization (Merritt, Wan, Shollock, & Patole, 2018; Potts, Dreyer, Bielawski, & Ruoff, 2011). In addition, due to its highly hydrophobic properties, the direct incorporation of graphene into biopolymers in an aqueous solution can be difficult.

To overcome these problems, reduced graphene oxide (rGO), which is structurally similar to graphene, could be incorporated into the composite. The rGO can be extracted through Hummers' method, which involves the oxidation of graphite to graphene oxide (GO), followed by a reduction process into rGO (Figure 1.3). During the oxidation of graphite, the structure of GO is endowed with reactive oxygen-containing functional groups at the basal planes and edges of the GO sheets, which facilitates the exfoliation process into single layer GO. This also allows the interactions between GO and the functional groups on hydrophilic polymers through hydrogen bonds, covalent bonds, and electrostatic interactions, thus improving the dispersion of GO within the polymeric matrix (Kumar & Koh, 2014; Yan et al., 2017).

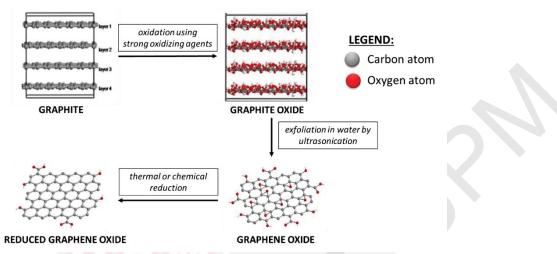


Figure 1.3 : Synthesis of graphene oxide (GO) and reduced graphene oxide (rGO) from graphite.

(Source: Naderi, Norouzi, Ganjali, & Gholipour-Ranjbar, 2016; Yuan et al., 2017)

The rGO can be incorporated into biopolymers via two approaches. The first approach involves the reduction of GO prior to incorporation, through methods such as microwave irradiation, chemical reductants, or thermal reduction. The second approach is in-situ reduction, where the reduction of GO occurs after the composite is cured. In this study, in-situ reduction was chosen due to the simplicity and safeness of this method as well as to avoid any possible aggregation of rGO in the polymeric matrix.

Besides the incorporation of nanofillers to improve the properties of biopolymeric composites, crosslinking is also an effective means to stabilize the structure of the polymeric network through the formation of a three-dimensional network of linked polymer chains. This usually results in improvements of the mechanical and barrier properties, as well as the stability of the material in water (Garavand et al., 2017; Mohamed, Mohd, Nurazzi, Siti Aisyah, & Mohd Fauzi, 2017; Nataraj, Sakkara, Meghwal, & Reddy, 2018). Depending on the properties of the crosslinking agent and the types of functional groups involved, crosslinking can be categorized into chemical (covalent), physical (ionic), and enzymatic crosslinking. Due to the ability of the amino groups in CS to be cationized in acidic media (Lizardi-Mendoza et al., 2016), physical crosslinking of CS can be effectively performed using negatively charged ions such as citrates (Li et al., 2018) as well as anionic molecules such as phosphate-bearing groups (Liang, Wang, & Chen, 2019).

In this study, the effectiveness of the composites in extending the shelf life of palm oilbased margarine was evaluated. Margarine was developed by a French chemist, Hippolyte Mège Mouriès, when Emperor Napoleon III demanded a cheaper butter alternative for the army. In the 1970s, scientists discovered that a diet high in saturated fat increased the level of low-density lipoprotein (LDL) cholesterol and decreased the level of high-density lipoprotein (HDL) cholesterol in the blood. This has caused many consumers to shift from butter to margarine, resulting in a spike in margarine's demand (Morris & Vaisey-Genser, 2003). In terms of composition, margarine is a water-in-oil emulsion which is made mainly of refined vegetable oils from sunflower, soybean, olive, or palm oil. Commercial margarine also contains additives such as emulsifier, salt, antioxidant(s), flavouring agent(s), and vitamins. In addition to enhancing the flavour and texture of food, margarine contributes nutritionally to our diet by being a source of fat-soluble vitamins such as vitamins A and D. With the advancement of research and technologies, more brands are now offering margarines that have a low trans fatty acids content as well as reduced fat and energy (Morris & Vaisey-Genser, 2003).

Concisely, CS is a valuable and potential material to be developed as a packaging material due to its excellent film-forming properties, oxygen barrier properties, as well as its abundance in nature. This can also reduce the waste generated from the shellfish processing industries. Various nanoparticles have been incorporated to enhance the properties of a CS film as food packaging material. Graphene is a promising nanofiller because of its remarkable mechanical strength, antimicrobial and antioxidant properties, as well as a high specific surface area. However, the lack of functional groups on graphene limits its interaction with CS. Therefore, rGO can be incorporated instead of pristine graphene. This involves the addition of the hydrophilic GO into CS, followed by an in-situ thermal reduction process. Besides the addition of nanofiller, the crosslinking of a CS has also been reported to improve its mechanical and barrier properties. However, a number of these studies focused on the synthesis of CS nanoparticles or beads (Babakhani & Sartaj, 2020; Hosseini, Soleimani, & Nikkhah, 2018; Jafari, Rad, Baharfar, Asghari, & Esfahani, 2020; Pan et al., 2020; Sang et al., 2020). On top of that, many studies which reported the crosslinking of CS films focused on the application on heavy metal removal from wastewater (Luna et al., 2019), as well as drug delivery and tissue engineering (Arteche Pujana, Pérez-Álvarez, Cesteros Iturbe, & Katime, 2013; Cho et al., 2016; Gierszewska & Ostrowska-Czubenko, 2016a). The effect of ionic crosslinking using sodium tripolyphosphate (TPP) and trisodium citrate (CIT) on the mechanical, physical, and antioxidant properties of chitosan-graphene oxide (CSGO) composite and its feasibility as a food packaging material, specifically for margarine, have yet to be investigated.

The development of a CSGO composite film material with improved mechanical strength, radical scavenging properties, as well as high light, water vapour, and oxygen barriers can be a promising packaging material for margarine to slow down the rancidification and therefore, extending its shelf life.

#### **1.2 Problem statement**

As a film packaging material, the strong hydrophilicity of CS can result in a compromised mechanical strength under humid environments (Elsabee & Abdou, 2013; Han, Yan, Chen, & Li, 2011), which often leads to compromised gas and water vapour barrier. The incorporation of graphene into CS is anticipated to significantly reinforce the material. However, pristine graphene is strongly hydrophobic which makes the incorporation into hydrophilic biopolymers such as CS, extremely challenging. GO is an inexpensive source to produce rGO, which shares comparable qualities with graphene (Gupta, Sharma, Singh, Arif, & Singh, 2017).

It is understood that the oxidation degree of GO can be controlled by the concentration of the oxidants, such as KMnO<sub>4</sub>, during the oxidation of graphite. Due to the abundance of oxygen-containing functional groups on its structure, a highly oxidized GO, which is synthesized using a graphite:KMnO<sub>4</sub> ratio of at least 1:6 (Marcano et al., 2010), is anticipated to incorporate homogeneously into a hydrophilic polymer such as CS, in comparison to GO with a lower oxidation degree. In addition, the epoxide groups in GO was shown to readily react with primary amine groups (Han et al., 2011). Due to the abundance of amino groups in CS, GO is expected to incorporate homogeneously into the polymeric network of CS. Nevertheless, the effect of GO oxidation degree on the incorporation into the CS polymeric matrix, as well as the effects on the structural and physical properties of the composite films as food packaging, are yet to be explored.

Previously, different sonication time have been used for the exfoliation of GO which ranged from 10 to 120 min (Liu et al., 2016; Meng, Ye, Coates, & Twigg, 2018; Pan, Wu, Bao, & Li, 2011; Wu, Wang, He, Zhang, & Zhang et al., 2017). A longer sonication time is expected to produce GO with a higher surface area-to-volume ratio, which will be more effective as a nanofiller (Cai et al., 2017; Tang, Ehlert, Lin, & Sodano, 2012). However, sonication can also damage and fragment the GO sheets (Dreyer, Park, Bielawski, & Ruoff, 2010; Parades, Villar-Rodil, Martínez-Alonso, & Tascón, 2008). In addition, the extremely high temperature resulting from the implosion of cavitation bubbles might thermally reduce GO, which could hinder the homogeneous incorporation into a hydrophilic polymer, such as CS.

The high hydrophilicity of GO is predicted to increase the water affinity of the composite (Yoo, Shin, Yoon, & Park, 2014). Therefore, the CSGO composite has to be treated in order to reduce its hydrophilicity and to improve its mechanical strength as well as oxygen and water vapour barrier properties under humid conditions. This could be achieved by the reduction of GO or the crosslinking process of the composite films to reduce the available functional groups.

Different temperatures have been applied for in-situ reduction of GO in previous studies, ranging from to 50 to 200 °C (Grande et al., 2017; Meng et al., 2016; Olowojoba et al., 2016; Toselli et al., 2015). However, different temperatures will cause a different saturation of oxygen-containing functional groups on the resulting GO nanosheets, which eventually influences the structural, mechanical, optical, and water resistant properties of the composites, as reported by Meng et al. (2016). Therefore, the optimum temperature for the in-situ thermal reduction of GO and its effects on the CSGO composites shall be determined.

Crosslinking has been used to effectively modify and improve the properties of CS. However, gelation occurs almost immediately once the crosslinking agents are introduced into the film-forming solution (Lin, Gu, & Cui, 2019; Nataraj et al., 2018; Yan et al., 2015), which makes it impossible to pour the solution onto the film-casting surface. However, this can be avoided by introducing the crosslinking agents after the films were cast, dried, and peeled.

The oxidative rancidity of margarine is often accelerated by heat, ultraviolet light, and oxygen (Riaz & Rokey, 2012). Therefore, the packaging material of the margarine can be improved in terms of its light and oxygen barrier. At present, many of the commercially available margarine in Malaysia adapt a polypropylene (PP) tub packaging (Figure 1.4). In fact, the demand for PP was one of the highest in the packaging sector along other resins such as low-density polyethylene (LDPE), high-density polyethylene (HDPE), and polyethylene terephthalate (PET) (*Plastics – the Facts 2020*, 2020). Unfortunately, PP is one of the least recycled post-consumer plastics which has a recycle ratio of only ~0.6% (Pavlík, Pavlíková, & Záleská, 2019). The huge volume of post-consumer waste has generated large amounts of urban solid residues (Vieira Ramos, Reis, Grafova, Grafov, & Monteiro, 2020). Therefore, a biodegradable active packaging with antioxidant properties as well as low light transmittance and oxygen permeability, can be a promising alternative to current petrochemical-based packaging for margarine.



Figure 1.4 : Examples of margarine brands sold in Malaysia that are packaged in polypropylene tubs.

#### **1.3** Objectives of the study

The aim of this study is to develop an antioxidant active packaging for margarine using chitosan (CS) with improved structural and physical properties, as well as water, light, and oxygen barrier through the incorporation of graphene oxide (GO) as well as heat and crosslinking treatments. There are four specific objectives in this study;

- 1. To prepare and evaluate the effect of oxidation degrees of GO on the structure and physical properties of CS films.
- 2. To evaluate the effect of sonication time and heating temperature on the structural and physical properties of chitosan-graphene oxide (CSGO) nanocomposite films.
- 3. To investigate the effect of trisodium citrate (CIT) and sodium tripolyphosphate (TPP) crosslinkers on the properties of CSGO nanocomposite films.
- 4. To evaluate the effect of CS and GO concentrations on the physicochemical properties of active packaging and their effects on storage stability of palm oil-based margarine.

The general research flow of this study, from Objective 1 to Objective 4, is summarized in Figure 1.5.

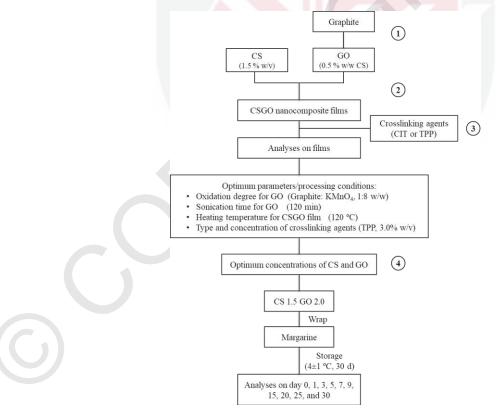


Figure 1.5 : Research flow of the study.

#### REFERENCES

- Abdollahi, M., Rezaei, M., & Farzi, G. (2012). A novel active bionanocomposite film incorporating rosemary essential oil and nanoclay into chitosan. *Journal of Food Engineering*, *111*, 343–350.
- Adilah, A. N., Jamilah, B., Noranizan, M. A., & Hanani, Z. A. N. (2018). Utilization of mango peel extracts on the biodegradable films for active packaging. *Food Packaging and Shelf Life*, 16, 1–7.
- Ahmad, M., Benjakul, S., Prodpran, T., & Agustini, T. W. (2012). Physico-mechanical and antimicrobial properties of gelatin film from the skin of unicorn leatherjacket incorporated with essential oils. *Food Hydrocolloids*, 28(1), 189–199.
- Ahmed, J., Mulla, M., Arfat, Y. A., & Thai, L. A. (2017). Mechanical, thermal, structural and barrier properties of crab shell chitosan/graphene oxide composite films. *Food Hydrocolloids*, 71, 141–148.
- Aider, M. (2010). Chitosan application for active bio–based films production and potential in the food industry: Review. *LWT Food Science and Technology*, *43*(6), 837–842.
- Akyuz, L., Kaya, M., Koc, B., Mujtaba, M., Ilk, S., Labidi, J., Salaberria, A. M., Cakmak, Y. S., & Yildiz, A. (2017). Diatomite as a novel composite ingredient for chitosan film with enhanced physicochemical properties. *International Journal of Biological Macromolecules*, 105, 1401–1411.
- Al-Fori, M., Dobretsov, S., Myint, M. T. Z., & Dutta, J. (2014). Antifouling properties of zinc oxide nanorod coatings. *Biofouling*, 30(7), 871–882.
- Al-Naamani, L., Dobretsov, S., & Dutta, J. (2016). Chitosan-zinc oxide nanoparticle composite coating for active food packaging applications. *Innovative Food Science* & *Emerging Technologies*, 38, 231–237.
- Alam, S. N., Sharma, N., & Kumar, L. (2017). Synthesis of graphene oxide (GO) by modified hummers method and its thermal reduction to obtain reduced graphene oxide (rGO). *Graphene*, 06(01), 1–18.
- Ali, A., Noh, N. M., & Mustafa, M. A. (2015). Antimicrobial activity of chitosan enriched with lemongrass oil against anthracnose of bell pepper. *Food Packaging* and Shelf Life, 3, 56–61.
- Alkarkhi, A. F. M., Muhammad, N. A. N., Alqaraghuli, W. A. A., Yusup, Y., Easa, A. M., & Huda, N. (2017). An investigation of food quality and oil stability indices of Muruku by cluster analysis and discriminant analysis. *International Journal on Advanced Science, Engineering and Information Technology*, 7(6), 2279–2285.
- Altiok, D., Altiok, E., & Tihminlioglu, F. (2010). Physical, antibacterial and antioxidant properties of chitosan films incorporated with thyme oil for potential wound healing applications. *Journal of Materials Science: Materials in Medicine*, 21(7), 2227–2236.
- Ang, P. K., Wang, S., Bao, Q., Thong, J. T. L., & Loh, K. P. (2009). High-throughput synthesis of graphene by intercalation–exfoliation of graphite oxide and study of ionic screening in graphene transistor. ACS Nano, 3(11), 3587–3594.

- AOAC. (2000). Peroxide value of oils and fats 965.33.12. In *Official methods of analysis* of AOAC international (17th ed.). Maryland, USA: AOAC International.
- AOCS. (1998). 2–Thiobarbituric acid value. Direct method. In D. Firestone (Ed.), Official Methods and Recommended Practices of the American Oil Chemists' Society (5th ed.). Illinois, USA: AOCS Press.
- Apjok, R., Mihaly Cozmuta, A., Peter, A., Mihaly Cozmuta, L., Nicula, C., Baia, M., & Vulpoi, A. (2019). Active packaging based on cellulose–chitosan–Ag/TiO 2 nanocomposite for storage of clarified butter. *Cellulose*, 26(3), 1923–1946.
- Approved additives and E numbers. (2020, December 20) Retrieved from https://www.food.gov.uk/business-guidance/approved-additives-and-e-numbers
- Arfat, Y. A., Ahmed, J., Ejaz, M., & Mullah, M. (2018). Polylactide/graphene oxide nanosheets/clove essential oil composite films for potential food packaging applications. *International Journal of Biological Macromolecules*, 107, 194–203.
- Arteche Pujana, M., Pérez-Álvarez, L., Cesteros Iturbe, L. C., & Katime, I. (2013). Biodegradable chitosan nanogels crosslinked with genipin. *Carbohydrate Polymers*, 94(2), 836–842.
- Assis, R. Q., Lopes, S. M., Costa, T. M. H., Flôres, S. H., & Rios, A. de O. (2017). Active biodegradable cassava starch films incorporated lycopene nanocapsules. *Industrial Crops and Products*, *109*, 818–827.
- ASTM D 644–94: Standard test methods for moisture content of paper and paperboard by oven drying (1994). ASTM International: Pennsylvania, USA.
- ASTM D 882–02: Standard Test Method for Tensile Properties of Thin Plastic Sheeting (2002). ASTM International: Pennsylvania, USA.
- ASTM D 883–12: Standard Terminology Relating to Plastics (2012). ASTM International: Pennsylvania, USA.
- ASTM D 5338–98: Standard test method for determining aerobic biodegradation of plastic materials under controlled composting conditions (2003). ASTM International: Pennsylvania, USA.
- ASTM E 96–95: Standard test methods for water vapor transmission of materials (1995). ASTM International: Pennsylvania, USA.
- Atarés, L., & Chiralt, A. (2016). Essential oils as additives in biodegradable films and coatings for active food packaging. *Trends in Food Science and Technology*, 48, 51–62.
- Avelelas, F., Horta, A., Pinto, L. F. V., Marques, S. C., Nunes, P. M., Pedrosa, R., & Leandro, S. M. (2019). Antifungal and antioxidant properties of chitosan polymers obtained from nontraditional *Polybius henslowii* sources. *Marine Drugs*, 17(4), 1– 15.
- Avila–Sosa, R., Palou, E., Jiménez Munguía, M. T., Nevárez–Moorillón, G. V., Navarro Cruz, A. R., & López–Malo, A. (2012). Antifungal activity by vapor contact of essential oils added to amaranth, chitosan, or starch edible films. *International Journal of Food Microbiology*, 153(1–2), 66–72.

- Azeredo, H. M. C., & Waldron, K. W. (2016). Crosslinking in polysaccharide and protein films and coatings for food contact – A review. *Trends in Food Science and Technology*, 52, 109–122.
- Baali, N., Khecha, A., Bensouici, A., Speranza, G., & Hamdouni, N. (2019). Assessment of antioxidant activity of pure graphene oxide (GO) and ZnO–decorated reduced graphene oxide (rGO) using DPPH radical and H<sub>2</sub>O<sub>2</sub> scavenging assays. *Journal* of Carbon Research, 5(4), 75–83.
- Babakhani, A., & Sartaj, M. (2020). Removal of Cadmium (II) from aqueous solution using tripolyphosphate cross–linked chitosan. *Journal of Environmental Chemical Engineering*, 8(4), 103842.
- Bagaria, H. G., & Wong, M. S. (2011). Polyamine-salt aggregate assembly of capsules as responsive drug delivery vehicles. *Journal of Materials Chemistry*, 21(26), 9454–9466.
- Bai, H., Li, C., & Shi, G. (2011). Functional composite materials based on chemically converted graphene. Advanced Materials, 23, 1089–1115.
- Balaguer, M. P., Gómez-Estaca, J., Gavara, R., & Hernandez-Munoz, P. (2011). Functional properties of bioplastics made from wheat gliadins modified with cinnamaldehyde. *Journal of Agricultural and Food Chemistry*, 59(12), 6689–6695.
- Balaguer, M. P., Cerisuelo, J. P., Gavara, R., & Hernandez–Muñoz, P. (2013). Mass transport properties of gliadin films: Effect of cross-linking degree, relative humidity, and temperature. *Journal of Membrane Science*, 428, 380–392.
- Balandin, A. A., Ghosh, S., Bao, W., Calizo, I., Teweldebrhan, D., Miao, F., & Lau, C. N. (2008). Superior thermal conductivity of single–layer graphene. *Nano Letters*, 8(3), 902–907.
- Barbolina, I., Woods, C. R., Lozano, N., Kostarelos, K., Novoselov, K. S., & Roberts, I. S. (2016). Purity of graphene oxide determines its antibacterial activity. 2D *Materials*, 3(2), 025025.
- Barra, A., Ferreira, N. M., Martins, M. A., Lazar, O., Pantazi, A., Alexandru, A., Neumayer, S.M., Rodriguez, B. J., Enăchescu, M., Ferreira, P., & Nunes, C. (2019). Eco-friendly preparation of electrically conductive chitosan-reduced graphene oxide flexible bionanocomposites for food packaging and biological applications. *Composites Science and Technology*, 173, 53–60.
- Barra, A., Santos, J. D. C., Silva, M. R. F., Nunes, C., Ruiz–Hitzky, E., Gonçalves, I., Yildirim, S., Ferreira, P., & Marques, P. A. A. P. (2020). Graphene derivatives in biopolymer–based composites for food packaging applications. *Nanomaterials*, 10(10), 1–32.
- Bellich, B., D'Agostino, I., Semeraro, S., Gamini, A., & Cesàro, A. (2016). "The good, the bad and the ugly" of chitosans. *Marine Drugs*, 14, 1–31.
- Berger, J., Reist, M., Mayer, J. M., Felt, O., Peppas, N. A., & Gurny, R. (2004). Structure and interactions in covalently and ionically crosslinked chitosan hydrogels for biomedical applications. *European Journal of Pharmaceutics and Biopharmaceutics*, 57(1), 19–34.
- Berger, K. R., & Welt, B. (2005). *A Brief History of Packaging*. Florida, USA: Florida Cooperative Extension Service.

- Bo, Z., Shuai, X., Mao, S., Yang, H., Qian, J., Chen, J., Yan, J., & Cen, K. (2014). Green preparation of reduced graphene oxide for sensing and energy storage applications. *Scientific Reports*, *4*, 1–8.
- Briassoulis, D., Dejean, C., & Picuno, P. (2010). Critical review of norms and standards for biodegradable agricultural plastics part II: Composting. *Journal of Polymers* and the Environment, 18(3), 364–383.
- Brodie, B. C. (1859). On the atomic weight of graphite. *Royal Society of London*, 149, 423–429.
- Brody, A. L., Bugusu, B., Han, J. H., Sand, C. K., & McHugh, T. H. (2008). Innovative food packaging solutions. *Journal of Food Science*, *73*(8), 107–116.
- Budianto, E., Muthoharoh, S. P., & Nizardo, N. M. (2015). Effect of crosslinking agents, ph and temperature on swelling behavior of cross–linked chitosan hydrogel. *Asian Journal of Applied Sciences*, 03(05), 581–588.
- Bugnicourt, L., & Ladavière, C. (2016). Interests of chitosan nanoparticles ionically cross-linked with tripolyphosphate for biomedical applications. *Progress in Polymer Science*, 60, 1–17.
- Bumbudsanpharoke, N., & Ko, S. (2019). Nanoclays in food and beverage packaging. *Journal of Nanomaterials*, 2019, 8927167.
- Cai, C., Sang, N., Shen, Z., & Zhao, X. (2017). Facile and size-controllable preparation of graphene oxide nanosheets using high shear method and ultrasonic method. *Journal of Experimental Nanoscience*, 12(1), 1–16.
- Campos, E., Coimbra, P., & Gil, M. H. (2013). An improved method for preparing glutaraldehyde cross–linked chitosan–poly(vinyl alcohol) microparticles. *Polymer Bulletin*, 70(2), 549–561.
- Carr, R. A., & Vaisey–Genser, M. (2003). Margarine Methods of Manufacture. In *Encyclopedia of Food Sciences and Nutrition* (2nd ed., pp. 3709–3714). Massachusetts, USA: Academic Press.
- Carvalho, A. P. A. de, & Conte Junior, C. A. (2020). Green strategies for active food packagings: A systematic review on active properties of graphene-based nanomaterials and biodegradable polymers. *Trends in Food Science and Technology*, 103, 130–143.
- Catalán, V., Frühbeck, G., & Gómez-Ambrosi, J. (2018). Inflammatory and oxidative stress markers in skeletal muscle of obese subjects. In *Obesity: Oxidative Stress and Dietary Antioxidants* (pp. 163–189). Massachusetts, USA: Academic Press.
- Cerruti, P., Santagata, G., Gomez, G., Ambrogi, V., Carfagna, C., Malinconico, M., & Persico, P. (2011). Effect of a natural polyphenolic extract on the properties of a biodegradable starch–based polymer. *Polymer Degradation and Stability*, *96*, 839–846.
- Chaban, V. V., & Prezhdo, O. V. (2017). Microwave reduction of graphene oxide rationalized by reactive molecular dynamics. *Nanoscale*, 9(11), 4024–4033.
- Chang, S. Y., & Lai, H. M. (2016). Effect of trisodium citrate on swelling property and structure of cationic starch thin film. *Food Hydrocolloids*, *56*, 254–265.

- Chang, W., Liu, F., Sharif, H. R., Huang, Z., Goff, H. D., & Zhong, F. (2018). Preparation of chitosan films by neutralization for improving their preservation effects on chilled meat. *Food Hydrocolloids*, 90, 50–61.
- Chen, C., Hu, W., He, Y., Jiang, A., & Zhang, R. (2016). Effect of citric acid combined with UV-C on the quality of fresh-cut apples. *Postharvest Biology and Technology*, *111*, 126–131.
- Chen, H., Hu, X., Chen, E., Wu, S., McClements, D. J., Liu, S., Li, B., & Li, Y. (2016). Preparation, characterization, and properties of chitosan films with cinnamaldehyde nanoemulsions. *Food Hydrocolloids*, *61*, 662–671.
- Chen, J., Li, Y., Huang, L., Li, C., & Shi, G. (2014). High–yield preparation of graphene oxide from small graphite flakes via an improved Hummers method with a simple purification process. *Carbon*, 81, 826–834.
- Chen, J., Yao, B., Li, C., & Shi, G. (2013). An improved Hummers method for ecofriendly synthesis of graphene oxide. *Carbon*, 64(1), 225–229.
- Chen, L., & Liu, H. (2012). Effect of emulsifying salts on the physicochemical properties of processed cheese made from Mozzarella. *Journal of Dairy Science*, 95(9), 4823–4830.
- Chen, X., Wu, G., Cai, Z., Oyama, M., & Chen, X. (2014). Advances in enzyme-free electrochemical sensors for hydrogen peroxide, glucose, and uric acid. *Microchimica Acta*, 181, 689–705.
- Cho, I. S., Cho, M. O., Li, Z., Nurunnabi, M., Park, S. Y., Kang, S. W., & Huh, K. M. (2016). Synthesis and characterization of a new photo–crosslinkable glycol chitosan thermogel for biomedical applications. *Carbohydrate Polymers*, 144, 59– 67.
- Choe, G., Kim, S. W., Park, J., Park, J., Kim, S., Kim, Y. S., Ahn, Y., Jung, D.-W., Williams, D. R., & Lee, J. Y. (2019). Anti-oxidant activity reinforced reduced graphene oxide/alginate microgels: Mesenchymal stem cell encapsulation and regeneration of infarcted hearts. *Biomaterials*, 225, 119513.
- Chougui, N., Djerroud, N., Naraoui, F., Hadjal, S., Aliane, K., Zeroual, B., & Larbat, R. (2015). Physicochemical properties and storage stability of margarine containing *Opuntia ficus-indica* peel extract as antioxidant. *Food Chemistry*, 173, 382–390.
- Chowdhury, S. S., Pandey, P. R., Kumar, R., & Roy, S. (2017). Effect of shape of protrusions and roughness on the hydrophilicity of a surface. *Chemical Physics Letters*, 685, 34–39.
- Chunxiu, L., Renbi, B., & Li, N. (2004). Sodium tripolyphosphate (TPP) crosslinked chitosan membranes and application in humic acid removal. *AIChE Annual Meeting, Conference Proceedings*, 7147–7160.
- Cichello, S. A. (2015). Oxygen absorbers in food preservation: a review. *Journal of Food Science and Technology*, 52(4), 1889–1895.
- Clark, G. L., & Smith, A. F. (1936). X–Ray diffraction studies of chitin, chitosan, and derivatives. *The Journal of Physical Chemistry*, 40(7), 863–879.

- Cobos, M., Fernández, M. J., & Fernández, M. D. (2018). Graphene based poly(vinyl alcohol) nanocomposites prepared by in situ green reduction of graphene oxide by ascorbic acid: Influence of graphene content and glycerol plasticizer on properties. *Nanomaterials*, 8(12), 8–11.
- Cobos, M., González, B., Fernández, M. J., & Fernández, M. D. (2017). Chitosangraphene oxide nanocomposites: Effect of graphene oxide nanosheets and glycerol plasticizer on thermal and mechanical properties. *Journal of Applied Polymer Science*, 134(30), 1–14.
- Cobos, M., González, B., Fernández, M. J., & Fernández, M. D. (2018). Study on the effect of graphene and glycerol plasticizer on the properties of chitosan–graphene nanocomposites via in situ green chemical reduction of graphene oxide. *International Journal of Biological Macromolecules*, *114*, 599–613.
- Code of Federal Regulations Title 21, Section 182.1810 Sodium tripolyphosphate. (2019, December 9). Retrieved from https://www.accessdata.fda.gov/scripts/cdrh/cfdocs/cfcfr/CFRSearch.cfm?fr=182 .1810
- Code of Federal Regulations Title 21– Section 184.1751 Sodium Citrate. (2019, December 9) Retrieved from https://www.accessdata.fda.gov/scripts/cdrh/cfdocs/cfcfr/CFRSearch.cfm?fr=184 .1751
- Coles, R., & Kirwan, M. (2011). Introduction. In *Food and Beverage Packaging Technology* (2nd ed., pp. 1–29). Oxford, England: Blackwell Publishing Ltd.
- Compton, B. O. C., Kim, S., Pierre, C., & Torkelson, J. M. (2010). Crumpled Graphene Nanosheets as Highly Effective Barrier Property Enhancers. *Advanced Materials*, 22, 4759–4763.
- Costa, M. J., Marques, A. M., Pastrana, L. M., Teixeira, J. A., Sillankorva, S. M., & Cerqueira, M. A. (2018). Physicochemical properties of alginate-based films: Effect of ionic crosslinking and mannuronic and guluronic acid ratio. *Food Hydrocolloids*, 81, 442–448.
- Costa, S. S., Druzian, J. I., Machado, B. A. S., De Souza, C. O., & Guimaraes, A. G. (2014). Bi-functional biobased packing of the cassava starch, glycerol, licuri nanocellulose and red propolis. *PLoS ONE*, 9(11), 112554.
- D'Angelo, D., Bongiorno, C., Amato, M., Deretzis, I., La Magna, A., Fazio, E., & Scalese, S. (2017). Oxygen functionalities evolution in thermally treated graphene oxide featured by EELS and DFT calculations. *Journal of Physical Chemistry C*, *121*(9), 5408–5414.
- Dalla Rosa, M. (2019). Packaging Sustainability in the Meat Industry. In *Sustainable Meat Production and Processing* (pp. 161–179). Massachusetts, USA: Elsevier Inc.
- De Marchi, L., Pretti, C., Gabriel, B., Marques, P. A. A. P., Freitas, R., & Neto, V. (2018). An overview of graphene materials: Properties, applications and toxicity on aquatic environments. *Science of the Total Environment*, 1440–1456.
- de Moraes Crizel, T., de Oliveira Rios, A., D. Alves, V., Bandarra, N., Moldão–Martins, M., & Hickmann Flôres, S. (2018). Active food packaging prepared with chitosan and olive pomace. *Food Hydrocolloids*, 74, 139–150.

- Dean, K., Sangwan, P., Way, C., Zhang, X., Martino, V. P., Xie, F., Halley, P. J., Pollet, E., & Avérous, L. (2013). Glycerol plasticised chitosan: A study of biodegradation via carbon dioxide evolution and nuclear magnetic resonance. *Polymer Degradation and Stability*, 98(6), 1236–1246.
- Debandi, M. V, Bernal, C., & Francois, N. J. (2016). Development of biodegradable films based on chitosan/glycerol blends suitable for biomedical applications. *Journal of Tissue Science & Engineering*, 07(03), 1000187.
- Decker, J. J., Meyers, K. P., Paul, D. R., Schiraldi, D. A., Hiltner, A., & Nazarenko, S. (2015). Polyethylene-based nanocomposites containing organoclay: A new approach to enhance gas barrier via multilayer coextrusion and interdiffusion. *Polymer*, 61, 42–54.
- Dey, A., & Neogi, S. (2019). Oxygen scavengers for food packaging applications: A review. Trends in Food Science and Technology, 90, 26–34.
- Domínguez, R., Barba, F. J., Gómez, B., Putnik, P., Bursać Kovačević, D., Pateiro, M., Santos, E. M., & Lorenzo, J. M. (2018). Active packaging films with natural antioxidants to be used in meat industry: A review. *Food Research International*, 113, 93–101.
- dos Santos Caetano, K., Almeida Lopes, N., Haas Costa, T. M., Brandelli, A., Rodrigues, E., Hickmann Flôres, S., & Cladera–Olivera, F. (2018). Characterization of active biodegradable films based on cassava starch and natural compounds. *Food Packaging and Shelf Life*, 16, 138–147.
- Dreyer, D. R., Park, S., Bielawski, C. W., & Ruoff, R. S. (2010). The chemistry of graphene oxide. *Chemical Society Reviews*, 39(1), 228–240.
- Ebrahimi, H., Abedi, B., Bodaghi, H., Davarynejad, G., Haratizadeh, H., & Conte, A. (2018). Investigation of developed clay-nanocomposite packaging film on quality of peach fruit (*Prunus persica* Cv. Alberta) during cold storage. *Journal of Food Processing and Preservation*, 42(2), 1–9.
- Echegoyen, Y., Rodríguez, S., & Nerín, C. (2016). Nanoclay migration from food packaging materials. Food Additives and Contaminants – Part A Chemistry, Analysis, Control, Exposure and Risk Assessment, 33(3), 530–539.
- Edwards, R. S., & Coleman, K. S. (2013). Graphene synthesis: relationship to applications. *Nanoscale*, 5(1), 38–51.
- Elsabee, M. Z., Morsi, R. E., & Fathy, M. (2016). Chitosan-Oregano Essential Oil Blends
   Use as Antimicrobial Packaging Material. In *Antimicrobial Food Packaging* (pp. 539–551). Massachusetts, USA: Elsevier Inc.
- Elsabee, M. Z., & Abdou, E. S. (2013). Chitosan based edible films and coatings: A review. *Materials Science and Engineering C*, *33*(4), 1819–1841.
- Fan, H., Wang, L., Zhao, K., Li, N., Shi, Z., Ge, Z., & Jin, Z. (2010). Fabrication, mechanical properties, and biocompatibility of graphene-reinforced chitosan composites. *Biomacromolecules*, 11(9), 2345–2351.
- Fang, C., Zhang, J., Chen, X., & Weng, G. J. (2020). Calculating the electrical conductivity of graphene nanoplatelet polymer composites by a Monte Carlo method. *Nanomaterials*, 10(6), 1–15.

- FAO- Codex Standards for Fats and Oils from Vegetable Sources. (2019, December 10) Retrieved from http://www.fao.org/3/y2774e/y2774e04.htm/
- Farhanini, Y., Khing, N. T., Hao, C. C., Sang, L. P., Muhamad, N. B., & Md Saleh, N. (2018). The electrochemical behavior of zinc oxide/reduced graphene oxide composite electrode in dopamine. *Malaysian Journal of Analytical Sciences*, 22(2), 227–237.
- Fasihnia, S. H., Peighambardoust, S. H., & Peighambardoust, S. J. (2018). Nanocomposite films containing organoclay nanoparticles as an antimicrobial (active) packaging for potential food application. *Journal of Food Processing and Preservation*, 42(2).
- Fideles, T. B., Santos, J. L., Tomás, H., Furtado, G. T. F. S., Lima, D. B., Borges, S. M. P., & Fook, M. V. L. (2018). Characterization of chitosan membranes crosslinked by sulfuric acid. *Open Access Library Journal*, 5, 1–13.
- Filip, V., Hrádková, I., & Šmidrkal, J. (2009). Antioxidants in margarine emulsions. *Czech Journal of Food Sciences*, 27, 25–28.
- Food Act 1983 (Act 281) & Regulations, Malaysia. (2016). Selangor, Malaysia: International Law Book Services.
- Frank, W. B. ., Haupin, W. E. ., Vogt, H., Bruno, M., Thonstad, J., Dawless, R. K., Kvande, H., & Taiwo, O. A. (2012). Aluminum. In Ullmann's Encyclopedia of Industrial Chemistry (Vol. 2, pp. 483–519). Weinheim, Germany: Wiley–VCH Verlag GmbH & Co. KGaA.
- Galus, S., & Kadzińska, J. (2015). Whey protein edible films modified with almond and walnut oils. *Food Hydrocolloids*, 52, 78–86.
- Garavand, F., Rouhi, M., Razavi, S. H., Cacciotti, I., & Mohammadi, R. (2017). Improving the integrity of natural biopolymer films used in food packaging by crosslinking approach: A review. *International Journal of Biological Macromolecules*, 104, 687–707.
- Gascho, J. L. S., Costa, S. F., Recco, A. A. C., & Pezzin, S. H. (2019). Graphene oxide films obtained by vacuum filtration: X-ray diffraction evidence of crystalline reorganization. *Journal of Nanomaterials*, 2019, 12–16.
- Geim, A. K., & Novoselov, K. S. (2007). The rise of graphene. *Nature Materials*, 6(3), 183–191.
- George, G., Simon, S. M., Prakashan, V., Sajna, M., Faisal, M., Wilson, R., Chandran, A., Bijhu, P. R., Joseph, C., & Unnikrishnan, N. V. (2018). Green and facile approach to prepare polypropylene: In situ reduced graphene oxide nanocomposites with excellent electromagnetic interference shielding properties. *RSC Advances*, 8(53), 30412–30428.
- Gierszewska–Drużyńska, M., & Ostrowska–Czubenko, J. (2011). Influence of crosslinking process conditions on molecular and supermolecular structure of chitosan hydrogel membrane. *Progress on Chemistry and Application of Chitin and Its Derivatives*, *16*, 15–22.
- Gierszewska, M., & Ostrowska–Czubenko, J. (2016a). Chitosan–based membranes with different ionic crosslinking density for pharmaceutical and industrial applications. *Carbohydrate Polymers*, *153*, 501–511.

- Gierszewska, M., & Ostrowska–Czubenko, J. (2016b). Equilibrium swelling study of crosslinked chitosan membranes in water, buffer and salt solutions. *Progress on Chemistry and Application of Chitin and Its Derivatives*, 21, 55–62.
- Glossary of Packaging Terms (6th ed.) (1988). Connecticut, USA: The Packaging Institute International.
- Glover, A. J., Cai, M., Overdeep, K. R., Kranbuehl, D. E., & Schniepp, H. C. (2011). In situ reduction of graphene oxide in polymers. *Macromolecules*, 44, 9821–9829.
- Goh, K., Heising, J. K., Yuan, Y., Karahan, H. E., Wei, L., Zhai, S., Koh, J., Htin, N. M., Zhang, F., Wang, R., Fane, A., G., Dekker, M., Dehghani, F., & Chen, Y. (2016). Sandwich–architectured poly(lactic acid)–graphene composite food packaging films. ACS Applied Materials and Interfaces, 8(15), 9994–10004.
- Gomes, L. P., Paschoalin, V. M. F., & Del Aguila, E. M. (2017). Chitosan nanoparticles: Production, physicochemical characteristics and nutraceutical applications. *Revista Virtual de Quimica*, 9(1), 387–409.
- Gómez–Estaca, J., López–de–Dicastillo, C., Hernández–Muñoz, P., Catalá, R., & Gavara, R. (2014). Advances in antioxidant active food packaging. *Trends in Food Science and Technology*, 35(1), 42–51.
- Gomez-Navarro, C., Weitz, R. T., Bittner, A. M., Scolari, M., Mews, A., Burghard, M., & Kern, K. (2007). Electronic transport properties of individual chemically reduced graphene oxide sheets. *Nano Letters*, 7(11), 3499–3503.
- Gong, H. P., Hua, W. M., Yue, Y. H., & Gao, Z. (2017). Graphene oxide for acid catalyzed-reactions: Effect of drying process. *Applied Surface Science*, 397, 44–48.
- Gouvêa, R. F., Del Aguila, E. M., Paschoalin, V. M. F., & Andrade, C. T. (2018). Extruded hybrids based on poly(3–hydroxybutyrate–co–3–hydroxyvalerate) and reduced graphene oxide composite for active food packaging. *Food Packaging and Shelf Life*, *16*(December 2017), 77–85.
- Graf, D., Molitor, F., Ensslin, K., Stampfer, C., Jungen, A., Hierold, C., & Wirtz, L. (2007). Spatially resolved raman spectroscopy of single- and few-layer graphene. *Nano Letters*, 7(2), 238–242.
- Grande, C. D., Mangadlao, J., Fan, J., De Leon, A., Delgado–Ospina, J., Rojas, J. G., Rodrigues, D. F., & Advincula, R. (2017). Chitosan cross–linked graphene oxide nanocomposite films with antimicrobial activity for application in food industry. *Macromolecular Symposia*, 374(1), 1–8.
- Guan, G., Abul Kalam Azad, M., Lin, Y., Kim, S. W., Tian, Y., Liu, G., & Wang, H. (2019). Biological effects and applications of chitosan and chito–oligosaccharides. *Frontiers in Physiology*, *10*, 1–10.
- Gudarzi, M. M. (2016). Colloidal stability of graphene oxide: Aggregation in two dimensions. *Langmuir*, *32*(20), 5058–5068.
- Guerrero, P., Muxika, A., Zarandona, I., & de la Caba, K. (2018). Crosslinking of chitosan films processed by compression molding. *Carbohydrate Polymers*, *206*, 820–826.

- Guerrero, P., Nur Hanani, Z. A., Kerry, J. P., & de la Caba, K. (2011). Characterization of soy protein–based films prepared with acids and oils by compression. *Journal of Food Engineering*, *107*(1), 41–49.
- Guo, H., Wang, X., Qian, Q., Wang, F., & Xia, X. (2009). A green approach to the synthesis of graphene nanosheets. ACS Nano, 3(9), 2653–2659.
- Guo, J., Ge, L., Li, X., Mu, C., & Li, D. (2014). Periodate oxidation of xanthan gum and its crosslinking effects on gelatin–based edible films. *Food Hydrocolloids*, *39*, 243–250.
- Gupta, V., Sharma, N., Singh, U., Arif, M., & Singh, A. (2017). Higher oxidation level in graphene oxide. *Optik*, *143*, 115–124.
- Habte, A. T., Ayele, D. W., & Hu, M. (2019). Synthesis and characterization of reduced graphene oxide (rGO) started from graphene oxide (GO) using the Tour method with different parameters. *Advances in Materials Science and Engineering*, 2019, 1–9.
- Hajji, S., Salem, R. B. S. Ben, Hamdi, M., Jellouli, K., Ayadi, W., Nasri, M., & Boufi, S. (2017). Nanocomposite films based on chitosan–poly(vinyl alcohol) and silver nanoparticles with high antibacterial and antioxidant activities. *Process Safety and Environmental Protection*, 111, 112–121.
- Ham, H., Van Khai, T., Park, N. H., So, D. S., Lee, J.W., Han, G. N., Kwon, Y. J., Cho, H. Y., & Kim, W. H. (2014). Freeze–drying–induced changes in the properties of graphene oxides. *Nanotechnology*, 25(23), 235601.
- Hamedi, H., Moradi, S., Hudson, S. M., & Tonelli, A. E. (2018). Chitosan based hydrogels and their applications for drug delivery in wound dressings: A review. *Carbohydrate Polymers*, *199*, 445–460.
- Hamman, J. H. (2010). Chitosan based polyelectrolyte complexes as potential carrier materials in drug delivery systems. *Marine Drugs*, 8(4), 1305–1322.
- Han, D., Wang, X., Zhao, Y., Chen, Y., Tang, M., & Zhao, Z. (2017). High–quality graphene synthesis on amorphous SiC through a rapid thermal treatment. *Carbon*, *124*, 105–110.
- Han, D., Yan, L., Chen, W., & Li, W. (2011). Preparation of chitosan/graphene oxide composite film with enhanced mechanical strength in the wet state. *Carbohydrate Polymers*, 83(2), 653–658.
- Harish Prashanth, K. V., & Tharanathan, R. N. (2006). Crosslinked chitosan Preparation and characterization. *Carbohydrate Research*, 341(1), 169–173.
- He, H., Klinowski, J., Forster, M., & Lerf, A. (1998). A new structural model for graphite oxide. *Chemical Physics Letters*, 287, 53–56.
- Hejjaji, E. M. A., Smith, A. M., & Morris, G. A. (2017). Designing chitosan– tripolyphosphate microparticles with desired size for specific pharmaceutical or forensic applications. *International Journal of Biological Macromolecules*, 95, 564–573.

- Higueras, L., López–Carballo, G., Gavara, R., & Hernández–Muñoz, P. (2014). Reversible covalent immobilization of cinnamaldehyde on chitosan films via schiff base formation and their application in active food packaging. *Food and Bioprocess Technology*, 8(3), 526–538.
- Homez–Jara, A., Daza, L. D., Aguirre, D. M., Muñoz, J. A., Solanilla, J. F., & Váquiro, H. A. (2018). Characterization of chitosan edible films obtained with various polymer concentrations and drying temperatures. *International Journal of Biological Macromolecules*, 113, 1233–1240.
- Hosseini, S. F., Soleimani, M. R., & Nikkhah, M. (2018). Chitosan/sodium tripolyphosphate nanoparticles as efficient vehicles for antioxidant peptidic fraction from common kilka. *International Journal of Biological Macromolecules*, 111, 730–737.
- Hu, W., Wang, Z., Xiao, Y., Zhang, S., & Wang, J. (2019). Advances in crosslinking strategies of biomedical hydrogels. *Biomaterials Science*, 7(3), 843–855.
- Hu, W., Peng, C., Luo, W., Lv, M., Li, X., Li, D., Huang, Q., & Fan, C. (2010). Graphene-based antibacterial paper, 4(7), 4317–4323.
- Huang, H. D., Ren, P. G., Chen, J., Zhang, W. Q., Ji, X., & Li, Z. M. (2012). High barrier graphene oxide nanosheet/poly(vinyl alcohol) nanocomposite films. *Journal of Membrane Science*, 409–410, 156–163.
- Huang, Y., Mei, L., Chen, X., & Wang, Q. (2018). Recent developments in food packaging based on nanomaterials. *Nanomaterials*, *8*, 1–29.
- Hummers, W. S., & Offeman, R. E. (1958). Preparation of graphitic oxide. *Journal of the American Chemical Society*, 80(6), 1339.
- Ickecan, D., Zan, R., & Nezir, S. (2017). Eco-friendly synthesis and characterization of reduced graphene oxide. *Journal of Physics: Conference Series*, 902(1), 6426–6432.
- Islam, M. M., Shahruzzaman, M., Biswas, S., Nurus Sakib, M., & Rashid, T. U. (2020). Chitosan based bioactive materials in tissue engineering applications– A review. *Bioactive Materials*, 5(1), 164–183.
- Islam, N., Dmour, I., & Taha, M. O. (2019). Degradability of chitosan micro/nanoparticles for pulmonary drug delivery. *Heliyon*, 5(5), 1–22.
- Izzreen, I., & Noriham, A. (2011). Evaluation of the antioxidant potential of some Malaysian herbal aqueous extracts as compared with synthetic antioxidants and ascorbic acid in cakes. *International Food Research Journal*, 18(2), 583–587.
- Jacobsen, C. (2016). Oxidative Stability and Shelf Life of Food Emulsions. In *Oxidative Stability and Shelf Life of Foods Containing Oils and Fats* (pp. 287–312). Massachusetts, USA: Elsevier Inc.
- Jafari, Z., Rad, A. S., Baharfar, R., Asghari, S., & Esfahani, M. R. (2020). Synthesis and application of chitosan/tripolyphosphate/graphene oxide hydrogel as a new drug delivery system for Sumatriptan Succinate. *Journal of Molecular Liquids*, *315*, 113835.

- Jahit, I. S., Nazmi, N. N. M., Isa, M. I. N., & Sarbon, N. M. (2016). Preparation and physical properties of gelatin/CMC/chitosan composite films as affected by drying temperature. *International Food Research Journal*, 23(3), 1068–1074.
- Jamróz, E., Kopel, P., Tkaczewska, J., Dordevic, D., Jancikova, S., Kulawik, P., Milosavljevic, V., Dolezelikova, K., Smerkova, K., & Adam, V. (2019). Nanocomposite furcellaran films-the influence of nanofillers on functional properties of furcellaran films and effect on linseed oil preservation. *Polymers*, 11(12), 2046.
- Janero, D. R. (1990). Malondialdehyde and thiobarbituric acid-reactivity as diagnostic indices of lipid peroxidation and peroxidative tissue injury. *Free Radical Biology and Medicine*, 9(6), 515–540.
- Jiang, J. W., Wang, J. S., & Li, B. (2009). Young's modulus of graphene: A molecular dynamics study. *Physical Review B – Condensed Matter and Materials Physics*, 80(11), 15–18.
- Jiang, T., Feng, L., & Zheng, X. (2012). Effect of chitosan coating enriched with thyme oil on postharvest quality and shelf life of shiitake mushroom (*Lentinus edodes*). *Journal of Agricultural and Food Chemistry*, 60(1), 188–196.
- Jin, R., Moreira Teixeira, L. S., Dijkstra, P. J., Karperien, M., van Blitterswijk, C. A., Zhong, Z. Y., & Feijen, J. (2009). Injectable chitosan-based hydrogels for cartilage tissue engineering. *Biomaterials*, 30(13), 2544–2551.
- Jing, X., Mi, H. Y., Napiwocki, B. N., Peng, X. F., & Turng, L. S. (2017). Musselinspired electroactive chitosan/graphene oxide composite hydrogel with rapid self-healing and recovery behavior for tissue engineering. *Carbon*, 125, 557–570.
- Jóźwiak, T., Filipkowska, U., Szymczyk, P., Rodziewicz, J., & Mielcarek, A. (2017). Effect of ionic and covalent crosslinking agents on properties of chitosan beads and sorption effectiveness of Reactive Black 5 dye. *Reactive and Functional Polymers*, 114, 58–74.
- Justin, R., & Chen, B. (2014). Strong and conductive chitosan–reduced graphene oxide nanocomposites for transdermal drug delivery. *Journal of Materials Chemistry B*, 2, 3759–3770.
- Kaczmarek, M. B., Struszczyk–Swita, K., Li, X., Szczęsna–Antczak, M., & Daroch, M. (2019). Enzymatic modifications of chitin, chitosan, and chitooligosaccharides. *Frontiers in Bioengineering and Biotechnology*, 7, 243.
- Kammoun, M., Haddar, M., Kossentini, T., Dammak, M., & Sayari, A. (2013). Biological properties and biodegradation studies of chitosan biofilms plasticized with PEG and glycerol. *International Journal of Biological Macromolecules*, 62, 433–438.
- Kang, S., Herzberg, M., Rodrigues, D. F., & Elimelech, M. (2008). Antibacterial effects of carbon nanotubes: Size does matter! *Langmuir*, 24(13), 6409–6413.
- Kazi, S. N., Badarudin, A., Zubir, M. N. M., Ming, H. N., Misran, M., Sadeghinezhad, E., Mehrali, M., & Syuhada, N. I. (2015). Investigation on the use of graphene oxide as novel surfactant to stabilize weakly charged graphene nanoplatelets. *Nanoscale Research Letters*, 10(1), 16–18.

- Khan, Y. H., Islam, A., Sarwar, A., Gull, N., Khan, S. M., Munawar, M. A., Zia, S., Sabir, A., Shafiq, M., & Jamil, T. (2016). Novel green nano composites films fabricated by indigenously synthesized graphene oxide and chitosan. *Carbohydrate Polymers*, 146, 131–138.
- Khanum, R., & Thevanayagam, H. (2017). Lipid peroxidation: Its effects on the formulation and use of pharmaceutical emulsions. Asian Journal of Pharmaceutical Sciences, 12(5), 401–411.
- Kim, H., Lee, S., & Lee, K. (2018). Scalable production of large single–layered graphenes by microwave exfoliation 'in deionized water'. *Carbon*, 134, 431–438.
- Kolanthai, E., Sindu, P. A., Khajuria, D. K., Veerla, S. C., Kuppuswamy, D., Catalani, L. H., & Mahapatra, D. R. (2018). Graphene oxide – A Tool for the preparation of chemically crosslinking free alginate–chitosan–collagen scaffolds for bone tissue engineering. ACS Applied Materials and Interfaces, 10(15), 12441–12452.
- Kong, F., & Singh, R. P. (2011). Advances in instrumental methods to determine food quality deterioration. In *Food and Beverage Stability and Shelf Life* (pp. 381–404). Cambridge, UK: Woodhead Publishing Limited.
- Kowalczyk, D., & Biendl, M. (2016). Physicochemical and antioxidant properties of biopolymer/candelilla wax emulsion films containing hop extract – A comparative study. *Food Hydrocolloids*, 60, 384–392.
- Krishnamoorthy, K., Veerapandian, M., Yun, K., & Kim, S. J. (2013). The chemical and structural analysis of graphene oxide with different degrees of oxidation. *Carbon*, 53, 38–49.
- Kumar, G., Bristow, J. F., Smith, P. J., & Payne, G. F. (2000). Enzymatic gelation of the natural polymer chitosan. *Polymer*, 41(6), 2157–2168.
- Kumar, S., & Koh, J. (2014). Physiochemical and optical properties of chitosan based graphene oxide bionanocomposite. *International Journal of Biological Macromolecules*, 70, 559–564.
- Lalwani, G., Agati, M. D., Mahmud, A., & Sitharaman, B. (2016). Toxicology of graphene-based nanomaterials. Advanced Drug Delivery Reviews, 105, 109–144.
- Landi, G., Sorrentino, A., Iannace, S., & Neitzert, H. C. (2017). Differences between graphene and graphene oxide in gelatin based systems for transient biodegradable energy storage applications. *Nanotechnology*, 28(5), 1–12.
- Laycock, B., Nikolić, M., Colwell, J. M., Gauthier, E., Halley, P., Bottle, S., & George, G. (2017). Lifetime prediction of biodegradable polymers. *Progress in Polymer Science*, 71, 144–189.
- Layek, R. K., Samanta, S., & Nandi, A. K. (2012). Graphene sulphonic acid/chitosan nano biocomposites with tunable mechanical and conductivity properties. *Polymer*, *53*(11), 2265–2273.
- Le, G. T. T., Manyam, J., Opaprakasit, P., Chanlek, N., Grisdanurak, N., & Sreearunothai, P. (2018). Divergent mechanisms for thermal reduction of graphene oxide and their highly different ion affinities. *Diamond and Related Materials*, 89, 246–256.

- Leceta, I., Guerrero, P., Ibarburu, I., Dueñas, M. T., & Caba, K. De. (2013). Characterization and antimicrobial analysis of chitosan-based films. *Journal of Food Engineering*, 116(4), 889–899.
- Leceta, I., Molinaro, S., Guerrero, P., Kerry, J. P., & de la Caba, K. (2015). Quality attributes of map packaged ready-to-eat baby carrots by using chitosan-based coatings. *Postharvest Biology and Technology*, 100, 142–150.
- Lee, C., Wei, X., Kysar, J. W., & Hone, J. (2008). Measurement of the elastic properties and intrinsic strength of monolayer graphene. *Science*, *321*, 385–389.
- Lee, E., Park, S. J., Lee, J. H., Kim, M. S., & Kim, C. H. (2016). Preparation of chitosan– TPP nanoparticles and their physical and biological properties. *Asian Journal of Pharmaceutical Sciences*, 11(1), 166–167.
- Lee, Y. M., Yoon, Y., Yoon, H., Song, S., Park, H. M., Lee, Y. Y., Shin, H., Hwang, S. W., & Yeum, K. J. (2018). Enhanced antioxidant activity of bioactives in colored grains by nano-carriers in human lens epithelial cells. *Molecules*, 23(6), 1327.
- Lerf, A., He, H., Forster, M., & Klinowski, J. (1998). Structure of graphite oxide revisited. *The Journal of Physical Chemistry B*, 102(23), 4477–4482.
- Leszczynska, A., & Pielichowski, K. (2008). Application of thermal analysis methods for characterization of polymer/montmorillonite nanocomposites. *Journal of Thermal Analysis and Calorimetry*, 93, 677–687.
- Li, B., Yang, J., Huang, Q., Zhang, Y., Peng, C., Zhang, Y., He, Y., Shi, J., Li, W., Hu, J., & Fan, C. (2013). Biodistribution and pulmonary toxicity of intratracheally instilled graphene oxide in mice. *NPG Asia Materials*, 5(4), 1–8.
- Li, D., Müller, M. B., Gilje, S., Kaner, R. B., & Wallace, G. G. (2008). Processable aqueous dispersions of graphene nanosheets. *Nature Nanotechnology*, 3(2), 101–105.
- Li, F., Yu, H. Y., Wang, Y. Y., Zhou, Y., Zhang, H., Yao, J. M., Abdalkarim, S. Y. H., & Tam, K. C. (2019). Natural biodegradable poly(3-hydroxybutyrate-co-3hydroxyvalerate) nanocomposites with multifunctional cellulose nanocrystals/graphene oxide hybrids for high-performance food packaging. *Journal of Agricultural and Food Chemistry*, 67(39), 10954–10967.
- Li, H., Yang, C., Chen, C., Ren, F., Li, Y., Mu, Z., & Wang, P. (2018). The use of trisodium citrate to improve the textural properties of acid–induced, transglutaminase–treated micellar casein gels. *Molecules*, 23(7).
- Li, N., Zhang, X., Song, Q., Su, R., Zhang, Q., Kong, T., Liu, L., Jin, G., Tang, M., & Cheng, G. (2011). The promotion of neurite sprouting and outgrowth of mouse hippocampal cells in culture by graphene substrates. *Biomaterials*, *32*(35), 9374–9382.
- Li, T., Song, X., Weng, C., Wang, X., Sun, L., Gong, X., Yang, L., & Chen, C. (2018). Self–crosslinking and injectable chondroitin sulfate/pullulan hydrogel for cartilage tissue engineering. *Applied Materials Today*, 10, 173–183.
- Li, Y. Q., Yu, T., Yang, T. Y., Zheng, L. X., & Liao, K. (2012). Bio–Inspired nacre–like composite films based on graphene with superior mechanical, electrical, and biocompatible properties. *Advanced Materials*, 24(25), 3426–3431.

- Lian, H., Shi, J., Zhang, X., & Peng, Y. (2020). Effect of the added polysaccharide on the release of thyme essential oil and structure properties of chitosan based film. *Food Packaging and Shelf Life*, 23(61), 100467.
- Liang, J., Wang, R., & Chen, R. (2019). The impact of cross-linking mode on the physical and antimicrobial properties of a chitosan/bacterial cellulose composite. *Polymers*, *11*(3), 491.
- Liang, S., Yi, M., Shen, Z., Liu, L., Zhang, X., & Ma, S. (2014). One-step green synthesis of graphene nanomesh by fluid-based method. *RSC Advances*, 4(31), 16127–16131.
- Lim, H. N., Huang, N. M., & Loo, C. H. (2012). Facile preparation of graphene–based chitosan films: Enhanced thermal, mechanical and antibacterial properties. *Journal* of Non–Crystalline Solids, 358(3), 525–530.
- Lim, L. Y., Khor, E., & Ling, C. E. (1999). Effects of dry heat and saturated steam on the physical properties of chitosan. *Journal of Biomedical Materials Research*, 48(2), 111–116.
- Lin, L., Gu, Y., & Cui, H. (2019). Moringa oil/chitosan nanoparticles embedded gelatin nanofibers for food packaging against *Listeria monocytogenes* and *Staphylococcus aureus* on cheese. *Food Packaging and Shelf Life*, 19, 86–93.
- Liu, D., Bian, Q., Li, Y., Wang, Y., Xiang, A., & Tian, H. (2016). Effect of oxidation degrees of graphene oxide on the structure and properties of poly(vinyl alcohol) composite films. *Composites Science and Technology*, 129, 146–152.
- Liu, F., Chang, W., Chen, M., Xu, F., Ma, J., & Zhong, F. (2019). Tailoring physicochemical properties of chitosan films and their protective effects on meat by varying drying temperature. *Carbohydrate Polymers*, 212, 150–159.
- Liu, L., Li, C., Bao, C., Jia, Q., Xiao, P., Liu, X., & Zhang, Q. (2012). Preparation and characterization of chitosan/graphene oxide composites for the adsorption of Au(III) and Pd(II). *Talanta*, *93*, 350–357.
- Liu, M., Zhang, X., Wu, W., Liu, T., Liu, Y., Guo, B., & Zhang, R. (2019). One-step chemical exfoliation of graphite to ~ 100 % few-layer graphene with high quality and large size at ambient temperature. *Chemical Engineering Journal*, 355, 181–185.
- Liu, S., Zeng, T. H., Hofmann, M., Burcombe, E., Wei, J., Jiang, R., Kong, J., & Chen, Y. (2011). Antibacterial activity of graphite, graphite oxide, graphene oxide, and reduced graphene oxide: Membrane and oxidative stress. *ACS Nano*, 5(9), 6971–6980.
- Lizardi-Mendoza, J., Argüelles Monal, W. M., & Goycoolea Valencia, F. M. (2016). Chemical Characteristics and Functional Properties of Chitosan. In *Chitosan in the Preservation of Agricultural Commodities* (pp. 3–31). Elsevier Inc.
- Lockhart, H. E. (1997). A paradigm for packaging. *Packaging Technology and Science*, 10(5), 237–252.
- Lopes, C. de O., Barcelos, M. de F. P., Dias, N. A. A., Carneiro, J. de D. S., & de Abreu, W. C. (2014). Effect of the addition of spices on reducing the sodium content and increasing the antioxidant activity of margarine. *LWT – Food Science and Technology*, 58(1), 63–70.

- Luna, M. S. De, Ascione, C., Santillo, C., Verdolotti, L., Lavorgna, M., Buonocore, G. G., Castaldo, R., Filippone, G., Xia, H., & Ambrosio, L. (2019). Optimization of dye adsorption capacity and mechanical strength of chitosan aerogels through crosslinking strategy and graphene oxide addition. *Carbohydrate Polymers*, 211, 195–203.
- Lv, C., Hue, Q., Xia, D., Ma, M., Xie, J., & Chwn, H. (2010). Effect of chemisorption on the interfacial bonding characteristics of graphene–polymer composites. *Journal of Physical Chemistry C*, 114, 6588–6594.
- Lv, Y. N., Wang, J. F., Long, Y., Tao, C. A., Xia, L., & Zhu, H. (2012). How graphene layers depend on drying methods of graphene oxide. *Advanced Materials Research*, 554, 597–600.
- Mahendran, R., Sridharan, D., Santhakumar, K., Selvakumar, T. A., Rajasekar, P., & Jang, J.-H. (2016). Graphene oxide reinforced polycarbonate nanocomposite films with antibacterial properties. *Indian Journal of Materials Science*, 2016, 1–10.
- Maitra, J., & Shukla, V. K. (2014). Cross–linking in Hydrogels A Review. American Journal of Polymer Science, 4(2), 25–31.
- Marcano, D. C., Kosynkin, D. V., Berlin, J. M., Sinitskii, A., Sun, Z., Slesarev, A., Alemany, L. B., Lu, W., & Tour, J. M. (2010). Improved synthesis of graphene oxide. ACS Nano, 4(8), 4806–4814.
- Marcus, J. B. (2013). Food Science Basics: Healthy Cooking and Baking Demystified. In *Culinary Nutrition* (pp. 51–97). Massachusetts, USA: Elsevier Inc.
- Marin, E., Briceño, M. I., & Caballero–George, C. (2013). Critical evaluation of biodegradable polymers used in nanodrugs. *International Journal of Nanomedicine*, 8, 3071–3091.
- Maryam Adilah, Z. A., Jamilah, B., & Nur Hanani, Z. A. (2018). Functional and antioxidant properties of protein–based films incorporated with mango kernel extract for active packaging. *Food Hydrocolloids*, 74, 207–218.
- McAllister, M. J., Li, J.-L., Adamson, D. H., Schniepp, H. C., Abdala, A. A., Liu, J., Herrera–Alonso, M., Milius, D. L. Car, R., Prud'homme, R. K., & Aksay, I. A. (2007). Single sheet functionalized graphene by oxidation and thermal expansion of graphite. *Chemistry of Materials*, 19(18), 4396–4404.
- Medina–Jaramillo, C., Ochoa–Yepes, O., Bernal, C., & Famá, L. (2017). Active and smart biodegradable packaging based on starch and natural extracts. *Carbohydrate Polymers*, 176, 187–194.
- Medina Jaramillo, C., Gutiérrez, T. J., Goyanes, S., Bernal, C., & Famá, L. (2016). Biodegradability and plasticizing effect of yerba mate extract on cassava starch edible films. *Carbohydrate Polymers*, *151*, 150–159.
- Mehta, B. M., Darji, V. B., & Aparnathi, K. D. (2015). Comparison of five analytical methods for the determination of peroxide value in oxidized ghee. *Food Chemistry*, 185, 449–453.
- Mellado, C., Figueroa, T., Baez, R., Meléndrez, M., & Fernández, K. (2019). Effects of probe and bath ultrasonic treatments on graphene oxide structure. *Materials Today Chemistry*, 13, 1–7.

- Meng, N., Priestley, R. C. E., Zhang, Y., Wang, H., & Zhang, X. (2016). The effect of reduction degree of GO nanosheets on microstructure and performance of PVDF/GO hybrid membranes. *Journal of Membrane Science*, 501, 169–178.
- Meng, Y., Ye, L., Coates, P., & Twigg, P. (2018). In situ cross–linking of poly(vinyl alcohol)/graphene oxide–polyethylene glycol nanocomposite hydrogels as artificial cartilage replacement: Intercalation structure, unconfined compressive behavior, and biotribological behaviors. *Journal of Physical Chemistry C*, 122(5), 3157–3167.
- Merritt, S., Wan, C., Shollock, B., & Patole, S. (2018). Polymer/Graphene Nanocomposites for Food Packaging. In G. Cirillo, M. A. Kozlowski, & U. G. Spizzirri (Eds.), *Composites Materials for Food Packaging* (pp. 251–268). Beverly, MA: Scrivener Publishing LLC.
- Migneault, I., Dartiguenave, C., Betrand, M. J., & Waldron, K. C. (2004). Threedimensional carbon-based architectures for oil remediation: From synthesis and modification to functionalization. *Biotechniques*, 37, 790–802.
- Mitura, K. A., & Zarzycki, P. K. (2018). Forms of Carbon in Food Packaging. In *Role of Materials Science in Food Bioengineering* (pp. 73–107). Amsterdam, Netherlands: Elsevier Inc.
- Miwa, M., Nakajima, A., Fujishima, A., Hashimoto, K., & Watanabe, T. (2000). Effects of the surface roughness on sliding angles of water droplets on superhydrophobic surfaces. *Langmuir*, *16*(13), 5754–5760.
- Moeini, A., Pedram, P., Makvandi, P., Malinconico, M., & Gomez d'Ayala, G. (2020). Wound healing and antimicrobial effect of active secondary metabolites in chitosan-based wound dressings: A review. *Carbohydrate Polymers*, 233, 115839.
- Mohamed, R., Mohd, N., Nurazzi, N., Siti Aisyah, M. I., & Mohd Fauzi, F. (2017). Swelling and tensile properties of starch glycerol system with various crosslinking agents. *IOP Conference Series: Materials Science and Engineering*, 223(1), 012059.
- Moon, I. K., Lee, J., Ruoff, R. S., & Lee, H. (2010). Reduced graphene oxide by chemical graphitization. *Nature Communications*, *1*, 73.
- Morimune, S., Nishino, T., & Goto, T. (2012). Poly(vinyl alcohol)/graphene oxide nanocomposites prepared by a simple eco-process. *Polymer Journal*, 44(10), 1056–1063.
- Morris, D. H., & Vaisey–Genser, M. (2003). Margarine Dietary Importance. In Encyclopedia of Food Sciences and Nutrition (2nd ed., pp. 3968–3974). Massachusetts, USA: Academic Press.
- Mu, C., Guo, J., Li, X., Lin, W., & Li, D. (2012). Preparation and properties of dialdehyde carboxymethyl cellulose crosslinked gelatin edible films. *Food Hydrocolloids*, 27(1), 22–29.
- Mujtaba, M., Morsi, R. E., Kerch, G., Elsabee, M. Z., Kaya, M., Labidi, J., & Mahmood, K. (2019). Current advancements in chitosan–based film production for food technology; A review. *International Journal of Biological Macromolecules*, 121, 889–904.

- Nadeem, M., Imran, M., Taj, I., Ajmal, M., & Junaid, M. (2017). Omega–3 fatty acids, phenolic compounds and antioxidant characteristics of chia oil supplemented margarine. *Lipids in Health and Disease*, *16*(1), 1–12.
- Naderi, H. R., Norouzi, P., Ganjali, M. R., & Gholipour–Ranjbar, H. (2016). Synthesis of a novel magnetite/nitrogen–doped reduced graphene oxide nanocomposite as high performance supercapacitor. *Powder Technology*, 302, 298–308.
- Nasti, A., Zaki, N. M., De Leonardis, P., Ungphaiboon, S., Sansongsak, P., Rimoli, M. G., & Tirelli, N. (2009). Chitosan/TPP and chitosan/TPP-hyaluronic acid nanoparticles: Systematic optimisation of the preparative process and preliminary biological evaluation. *Pharmaceutical Research*, 26(8), 1918–1930.
- Nataraj, D., Sakkara, S., Meghwal, M., & Reddy, N. (2018). Crosslinked chitosan films with controllable properties for commercial applications. *International Journal of Biological Macromolecules*, 120, 1256–1264.
- Nazir, N., Diana, A., & Sayuti, K. (2017). Physicochemical and fatty acid profile of fish oil from head of tuna (*Thunnus albacares*) extracted from various extraction method. *International Journal on Advanced Science, Engineering and Information Technology*, 7(2), 709–715.
- Nethravathi, C., & Rajamathi, M. (2008). Chemically modified graphene sheets produced by the solvothermal reduction of colloidal dispersions of graphite oxide. *Carbon*, 46(14), 1994–1998.
- Ngo, D. H., & Kim, S. K. (2014). Antioxidant effects of chitin, chitosan, and their derivatives. *Advances in Food and Nutrition Research*, 73, 15–31.
- Nor Adilah, A., Noranizan, M. A., Jamilah, B., & Nur Hanani, Z. A. (2020). Development of polyethylene films coated with gelatin and mango peel extract and the effect on the quality of margarine. *Food Packaging and Shelf Life*, 26, 100577.
- Novoselov, K. S., Geim, A. K., Morozov, S. V, Jiang, D., Zhang, Y., Dubonos, S. V, Grigorieva, I. V., & Firsov, A. A. (2004). Electric field effect in atomically thin carbon films. *Science*, *306*, 666–669.
- Nur Fatin Nazurah, R., & Nur Hanani, Z. A. (2017). Physicochemical characterization of kappa–carrageenan (*Euchema cottoni*) based films incorporated with various plant oils. *Carbohydrate Polymers*, 157, 1479–1487.
- Nurul Syahida, S., Ainun, Z. M. A., Ismail–Fitry, M. R., & Nur Hanani, Z. A. (2020). Development and characterisation of gelatine/palm wax/lemongrass essential oil (GPL)–coated paper for active food packaging. *Packaging Technology and Science*, 1–15.
- O'Sullivan, M. G. (2017). Shelf Life and Sensory Quality of Foods and Beverages. In A Handbook for Sensory and Consumer–Driven New Product Development (pp. 103–123). Sawston, United Kingdom: Woodhead Publishing Limited.
- Ojeda, G. A., Sgroppo, S. C., Martín-Belloso, O., & Soliva-Fortuny, R. (2019). Chitosan/tripolyphosphate nanoaggregates enhance the antibrowning effect of ascorbic acid on mushroom slices. *Postharvest Biology and Technology*, *156*, 110934.

- Olowojoba, G. B., Eslava, S., Gutierrez, E. S., Kinloch, A. J., Mattevi, C., Rocha, V. G., & Taylor, A. C. (2016). In situ thermally reduced graphene oxide/epoxy composites: thermal and mechanical properties. *Applied Nanoscience*, 6(7), 1015– 1022.
- Ou, L., Song, B., Liang, H., Liu, J., Feng, X., Deng, B., Sun, T., & Shao, L. (2016). Toxicity of graphene–family nanoparticles: A general review of the origins and mechanisms. *Particle and Fibre Toxicology*, 13(1), 1–24.
- Pal, N., Banerjee, S., Roy, P., & Pal, K. (2019). Reduced graphene oxide and PEG– grafted TEMPO–oxidized cellulose nanocrystal reinforced poly–lactic acid nanocomposite film for biomedical application. *Materials Science and Engineering C*, 104, 109956.
- Pan, C., Qian, J., Zhao, C., Yang, H., Zhao, X., & Guo, H. (2020). Study on the relationship between crosslinking degree and properties of TPP crosslinked chitosan nanoparticles. *Carbohydrate Polymers*, 241, 116349.
- Pan, S., & Aksay, I. A. (2011). Factors controlling the size of graphene oxide sheets produced via the graphite oxide route. ACS Nano, 5(5), 4073–4083.
- Pan, Y., Wu, T., Bao, H., & Li, L. (2011). Green fabrication of chitosan films reinforced with parallel aligned graphene oxide. *Carbohydrate Polymers*, 83(4), 1908–1915.
- Parades, J. I., Villar-Rodil, S., Martínez-Alonso, A., & Tascón, J. M. D. (2008). Graphene oxide dispersions in organic solvents. *Langmuir*, 24(19), 10560–10564.
- Park, S. Il, Marsh, K. S., & Dawson, P. (2010). Application of chitosan-incorporated LDPE film to sliced fresh red meats for shelf life extension. *Meat Science*, 85(3), 493–499.
- Parsa, A., & Salout, S. A. (2016). Investigation of the antioxidant activity of electrosynthesized polyaniline/reduced graphene oxide nanocomposite in a binary electrolyte system on ABTS and DPPH free radicals. *Journal of Electroanalytical Chemistry*, 760, 113–118.
- Paulchamy, B., Arthi, G., & Lignesh, B. (2015). A simple approach to stepwise synthesis of graphene oxide nanomaterial. *Journal of Nanomedicine & Nanotechnology*, 6(1), 2–5.
- Pavlík, Z., Pavlíková, M., & Záleská, M. (2019). Properties of concrete with plastic polypropylene aggregates. In Use of Recycled Plastics in Eco-efficient Concrete (pp. 189–213). Massachusetts, USA: Elsevier Inc.
- Pei, S., & Cheng, H. M. (2012). The reduction of graphene oxide. *Carbon*, 50(9), 3210–3228.
- Pelissari, F. M., Grossmann, M. V. E., Yamashita, F., & Pined, E. A. G. (2009). Antimicrobial, mechanical, and barrier properties of cassava starch-chitosan films incorporated with oregano essential oil. *Journal of Agricultural and Food Chemistry*, 57(16), 7499–7504.
- Perrozzi, F., Prezioso, S., & Ottaviano, L. (2015). Graphene oxide: From fundamentals to applications. *Journal of Physics Condensed Matter*, 27(1), 13002.

- Pieróg, M., Gierszewska–Drużyńska, M., & Ostrowska–Czubenko, J. (2009). Effect of ionic crosslinking agents on swelling behaviour of chitosan hydrogel membranes. *Progress on Chemistry and Application of Chitin and Its Derivatives*, 14, 75–82.
- Piñeros–Hernandez, D., Medina–Jaramillo, C., López–Córdoba, A., & Goyanes, S. (2017). Edible cassava starch films carrying rosemary antioxidant extracts for potential use as active food packaging. *Food Hydrocolloids*, 63, 488–495.
- Pinto, A. M., Cabral, J., Tanaka, D. A. P., Mendes, A. M., & Magalhães, F. D. (2013). Effect of incorporation of graphene oxide and graphene nanoplatelets on mechanical and gas permeability properties of poly(lactic acid) films. *Polymer International*, 62(1), 33–40.
- Pinto, A. M., Gonçalves, I. C., & Magalhães, F. D. (2013). Graphene–based materials biocompatibility: A review. *Colloids and Surfaces B: Biointerfaces*, 111, 188–202.
- Pirsa, S., & Asadi, S. (2021). Innovative smart and biodegradable packaging for margarine based on a nano composite polylactic acid/lycopene film. Food Additives and Contaminants – Part A, 38(5), 856–869.
- Plastics the Facts 2020. (2020, December 9). Retrieved from https://plasticseurope.org/wp-content/uploads/2021/09/Plastics\_the\_facts-WEB-2020\_versionJun21\_final.pdf
- Pokharkar, D. R., & Bhumkar, V. (2006). Studies on effect of pH on cross-linking of chitosan with sodium tripolyphosphate: A technical note. AAPS PharmSciTech, 7(2), 138–143.
- Ponsubha, S., & Jaiswal, A. K. (2019). Effect of interpolymer complex fomation between chondroitin sulfate and chitosan–gelatin hydrogel on physocochemical and rheological properties. *BBA Biomembranes*, 183135.
- Poon, L., Wilson, L. D., & Headley, J. V. (2014). Chitosan-glutaraldehyde copolymers and their sorption properties. *Carbohydrate Polymers*, 109, 92–101.
- Potts, J. R., Dreyer, D. R., Bielawski, C. W., & Ruoff, R. S. (2011). Graphene-based polymer nanocomposites. *Polymer*, 52(1), 5–25.
- Prateepchanachai, S., Thakhiew, W., Devahastin, S., & Soponronnarit, S. (2019). Improvement of mechanical and heat sealing properties of chitosan films via the use of glycerol and gelatin blends in film–forming solution. *International Journal* of Biological Macromolecules, 131, 589–600.
- Priyadarshi, R., Kumar, B., Deeba, F., & Kulshreshtha, A. (2018). Chitosan films incorporated with apricot (*Prunus armeniaca*) kernel essential oil as active food packaging material. *Food Hydrocolloids*, 85, 158–166.
- Qi, X., Zhou, T., & Deng, S. (2014). Size–specified graphene oxide sheets: ultrasonication assisted preparation and characterization. *Journal of Materials Science*, 49, 1785–1793.
- Qiu, Y., Wang, Z., Owens, A. C. E., Kulaots, I., Chen, Y., Kane, A. B., & Hurt, R. H. (2014). Antioxidant chemistry of graphene–based materials and its role in oxidation protection technology. *Nanoscale*, 6, 11744–11755.

- Rajeswari, R., & Prabu, H. G. (2018). Synthesis characterization, antimicrobial, antioxidant, and cytotoxic activities of ZnO nanorods on reduced graphene oxide. *Journal of Inorganic and Organometallic Polymers and Materials*, 28(3), 679– 693.
- Ramezani, Z., Zarei, M., & Raminnejad, N. (2015). Comparing the effectiveness of chitosan and nanochitosan coatings on the quality of refrigerated silver carp fillets. *Food Control*, 51, 43–48.
- Ramos, M., Valdés, A., Beltrán, A., & Garrigós, M. (2016). Gelatin–based films and coatings for food packaging applications. *Coatings*, 6(4), 41.
- Rana, V., Babita, K., Goyal, D., & Tiwary, A. K. (2005). Sodium citrate cross–linked chitosan films: Optimization as substitute for human/rat/rabbit epidermal sheets. *Journal of Pharmacy and Pharmaceutical Sciences*, 8(1), 10–17.
- Rattana, T., Chaiyakun, S., Witit–Anun, N., Nuntawong, N., Chindaudom, P., Oaew, S., Kedkeaw, C., & Limsuwan, P. (2012). Preparation and characterization of graphene oxide nanosheets. *Procedia Engineering*, 32, 759–764.
- Ray Chowdhuri, A., Tripathy, S., Chandra, S., Roy, S., & Sahu, S. K. (2015). A ZnO decorated chitosan–graphene oxide nanocomposite shows significantly enhanced antimicrobial activity with ROS generation. *RSC Advances*, 5(61), 49420–49428.
- Realini, C. E., & Marcos, B. (2014). Active and intelligent packaging systems for a modern society. *Meat Science*, 98(3), 404–419.
- Reddy, N., & Yang, Y. (2010). Citric acid cross-linking of starch films. *Food Chemistry*, *118*(3), 702–711.
- Rhim, J.–W. (2013). Effect of PLA lamination on performance characteristics of agar/κ– carrageenan/clay bio–nanocomposite film. *Food Research International*, 51, 714– 722.
- Riaz, M. N., & Rokey, G. J. (2012). Impact of particle size and other ingredients on extruded foods and feeds. In *Extrusion Problems Solved* (pp. 55–63). Sawston, United Kingdom: Woodhead Publishing Limited.
- Risch, S. J. (2009). Food packaging history and innovations. *Journal of Agricultural and Food Chemistry*, 57(18), 8089–8092.
- Robertson, G. (2010). *Food Packaging and Shelf Life: A Practical Guide*. Florida, USA: CRC Press.
- Robertson, G. L. (2012). *Food Packaging Principles and Practice* (3rd ed.). Florida, USA: CRC Press.
- Robertson, G. L. (2019). *History of Food Packaging*. *Reference Module in Food Science*. Massachusetts, USA: Elsevier.
- Rouhi, J., Mahmud, S., Naderi, N., Raymond Ooi, C. H., & Mahmood, M. R. (2013). Physical properties of fish gelatin–based bio–nanocomposite films incorporated with ZnO nanorods. *Nanoscale Research Letters*, 8(1), 1–6.
- Roy, J. C., Salaün, F., Giraud, S., Ferri, A., Chen, G., & Guan, J. (2017). Solubility of Chitin: Solvents, Solution Behaviors and Their Related Mechanisms. In *Solubility* of *Polysaccharides* (pp. 109–127). London, United Kingdom: IntechOpen Limited.

- Rozada, R., Paredes, J. I., López, M. J., Villar–Rodil, S., Cabria, I., Alonso, J. A., Martínez–Alonso, A., & Tascón, J. M. D. (2015). From graphene oxide to pristine graphene: Revealing the inner workings of the full structural restoration. *Nanoscale*, 7(6), 2374–2390.
- Rubentheren, V., Ward, T. A., Chee, C. Y., Nair, P., Salami, E., & Fearday, C. (2016). Effects of heat treatment on chitosan nanocomposite film reinforced with nanocrystalline cellulose and tannic acid. *Carbohydrate Polymers*, 140, 202–208.
- Ryan, P. G., Moore, C. J., Van Franeker, J. A., & Moloney, C. L. (2009). Monitoring the abundance of plastic debris in the marine environment. *Philosophical Transactions of the Royal Society B: Biological Sciences*, *364*, 1999–2012.
- Sanchez, V. C., Jachak, A., Hurt, R. H., & Kane, A. B. (2012). Biological interactions of graphene–family nanomaterials: An interdisciplinary review. *Chemical Research in Toxicology*, 25(1), 15–34.
- Sang, Z., Qian, J., Han, J., Deng, X., Shen, J., Li, G., & Xie, Y. (2020). Comparison of three water-soluble polyphosphate tripolyphosphate, phytic acid, and sodium hexametaphosphate as crosslinking agents in chitosan nanoparticle formulation. *Carbohydrate Polymers*, 230, 115577.
- Santonicola, S., Ibarra, V. G., Sendón, R., Mercogliano, R., & de Quirós, A. (2017). Antimicrobial films based on chitosan and methylcellulose containing natamycin for active packaging applications. *Coatings*, 7(177), 1–10.
- Sanuja, S., Agalya, A., & Umapathy, M. J. (2015). Synthesis and characterization of zinc oxide-neem oil-chitosan bionanocomposite for food packaging application. *International Journal of Biological Macromolecules*, 74, 76–84.
- Sasikala, S. P., Poulin, P., & Aymonier, C. (2017). Advances in subcritical hydro-/solvothermal processing of graphene materials. *Advanced Materials*, 29(22), 1605473.
- Schreiber, S. B., Bozell, J. J., Hayes, D. G., & Zivanovic, S. (2013). Introduction of primary antioxidant activity to chitosan for application as a multifunctional food packaging material. *Food Hydrocolloids*, 33(2), 207–214.
- Schrödter, K., Bettermann, G., Staffel, T., Wahl, F., Klein, T., & Hofmann, T. (2012). Phosphoric Acid and Phosphates. In *Ullman's encyclopedia of industrial chemistry* (pp. 679–724). Weinheim, Germany: Wiley–VCH Verlag GmbH & Co. KGaA.
- Shankar, S., Oun, A. A., & Rhim, J. W. (2018). Preparation of antimicrobial hybrid nano-materials using regenerated cellulose and metallic nanoparticles. *International Journal of Biological Macromolecules*, *107*, 17–27.
- Shao, L., Chang, X., Zhang, Y., Huang, Y., Yao, Y., & Guo, Z. (2013). Graphene oxide cross–linked chitosan nanocomposite membrane. *Applied Surface Science*, 280, 989–992.
- Sharma, C., Dhiman, R., Rokana, N., & Panwar, H. (2017). Nanotechnology: An untapped resource for food packaging. *Frontiers in Microbiology*, 8, 1–22.
- Sharma, L., Sharma, H. K., & Saini, C. S. (2018). Edible films developed from carboxylic acid cross–linked sesame protein isolate: barrier, mechanical, thermal, crystalline and morphological properties. *Journal of Food Science and Technology*, *55*(2), 532–539.

- Sharma, S., Swetha, K. L., & Roy, A. (2019). Chitosan-chondroitin sulfate based polyelectrolyte complex for effective management of chronic wounds. *International Journal of Biological Macromolecules*, 132, 97–108.
- Shen, L., Wang, D., Jin, Z., Che, L., Cai, N., Wang, Y., & Lu, Y. (2019). The effect of drying modes on aqueous dispersion of graphene oxide solids. *Functional Materials Letters*, 12(4), 1950043.
- Sheng, X., Xie, D., Cai, W., Zhang, X., Zhong, L., & Zhang, H. (2014). In-situ thermal reduction of graphene nanosheets based polymethyl methacrylate nanocomposites with effective reinforcements. *Industrial & Engineering Chemistry Research*, 54(2), 649–658.
- Shi, R., Bi, J., Zhang, Z., Zhu, A., Chen, D., Zhou, X., Zhang, L., & Tian, W. (2008). The effect of citric acid on the structural properties and cytotoxicity of the polyvinyl alcohol/starch films when molding at high temperature. *Carbohydrate Polymers*, 74(4), 763–770.
- Shih, C.-J., Lin, S., Sharma, R., Strano, M. S., & Blankschtein, D. (2011). Understanding the pH–dependent behavior of graphene oxide aqueous solutions: A comparative experimental and molecular dynamics simulation study. *Langmuir*, 28(1), 235–241.
- Shojaee–Aliabadi, S., Hosseini, H., Mohammadifar, M. A., Mohammadi, A., Ghasemlou, M., Ojagh, S. M., Hosseini, S. M., & Khaksar, R. (2013). Characterization of antioxidant–antimicrobial κ–carrageenan films containing *Satureja hortensis* essential oil. *International Journal of Biological Macromolecules*, 52(1), 116–124.
- Shu, X. Z., Zhu, K. J., & Song, W. (2001). Novel pH–sensitive citrate cross–linked chitosan film for drug controlled release. *International Journal of Pharmaceutics*, 212, 19–28.
- Siracusa, V., Romani, S., Gigli, M., Mannozzi, C., Cecchini, J. P., Tylewicz, U., & Lotti, N. (2018). Characterization of active edible films based on citral essential oil, alginate and pectin. *Materials*, 11(10), 1980.
- Sirelkhatim, A., Mahmud, S., Seeni, A., Kaus, N. H. M., Ann, L. C., Bakhori, S. K. M., Hassan, H., & Mohamad, D. (2015). Review on zinc oxide nanoparticles: Antibacterial activity and toxicity mechanism. *Nano–Micro Letters*, 7(3), 219–242.
- Siripatrawan, U., & Kaewklin, P. (2018). Fabrication and characterization of chitosantitanium dioxide nanocomposite film as ethylene scavenging and antimicrobial active food packaging. *Food Hydrocolloids*, 84, 125–134.
- Sivaselvi, K., & Ghosh, P. (2017). Characterization of modified chitosan thin film. Materials Today: Proceedings, 4(2), 442–451.
- Solodovnik, T., Stolyarenko, H., Slis, A., & Kultenko, V. (2017). Study of heat treatment effect on structure and solubility of chitosan films. *Chemistry & Chemical Technology*, *11*(2), 175–179.
- Soltani, T., & Lee, B. K. (2017). Low intensity–ultrasonic irradiation for highly efficient, eco–friendly and fast synthesis of graphene oxide. *Ultrasonics Sonochemistry*, 38(2), 693–703.
- Song, X., Zuo, G., & Chen, F. (2018). Effect of essential oil and surfactant on the physical and antimicrobial properties of corn and wheat starch films. *International Journal of Biological Macromolecules*, 107, 1302–1309.

- Sothornvit, R. (2019). Nanostructured materials for food packaging systems: new functional properties. *Current Opinion in Food Science*, 25, 82–87.
- Souza, J. M., Caldas, A. L., Tohidi, S. D., Molina, J., Souto, A. P., Fangueiro, R., & Zille, A. (2014). Properties and controlled release of chitosan microencapsulated limonene oil. *Brazilian Journal of Pharmacognosy*, 24(6), 691–698.
- Souza, V. G. L., & Fernando, A. L. (2016). Nanoparticles in food packaging: Biodegradability and potential migration to food– A review. *Food Packaging and Shelf Life*, 8, 63–70.
- Staudenmaier, L. (1898). Method for the preparation of the graphite acid. *European Journal of Inorganic Chemistry*, 31(2), 1481–1487.
- Štular, D., Šobak, M., Mihelčič, M., Šest, E., German Ilić, I., Jerman, I., Simončič, B., & Tomšič, B. (2019). Proactive release of antimicrobial essential oil from a "smart" cotton fabric. *Coatings*, 9(4), 242.
- Su, C. Y., Xu, Y., Zhang, W., Zhao, J., Tang, X., Tsai, C. H., & Li, L. J. (2009). Electrical and spectroscopic characterizations of ultra–large reduced graphene oxide monolayers. *Chemistry of Materials*, 21(23), 5674–5680.
- Su, Y., Kravets, V. G., Wong, S. L., Waters, J., Geim, A. K., & Nair, R. R. (2014). Impermeable barrier films and protective coatings based on reduced graphene oxide. *Nature Communications*, 5, 1–5.
- Sun, T., Wu, C. ling, Hao, H., Dai, Y., & Li, J. rong. (2016). Preparation and preservation properties of the chitosan coatings modified with the in situ synthesized nano SiO<sub>x</sub>. *Food Hydrocolloids*, 54, 130–138.
- Sundramoorthy, A. K., Vignesh Kumar, T. H., & Gunasekaran, S. (2018). Graphene– Based Nanosensors and Smart Food Packaging Systems for Food Safety and Quality Monitoring. In *Graphene Bioelectronics* (pp. 267–306). Amsterdam, Netherlands: Elsevier Inc.
- Suresh, D., Udayabhanu, Pavan Kumar, M. A., Nagabhushana, H., & Sharma, S. C. (2015). Cinnamon supported facile green reduction of graphene oxide, its dye elimination and antioxidant activities. *Materials Letters*, 151, 93–95.
- Sustainable Packaging Criteria. (2020, August 16). Retrieved from https://sustainablepackaging.org/wp-content/uploads/2017/09/Definition-of-Sustainable-Packaging.pdf/
- Syed, A. (2016). Oxidative Stability and Shelf Life of Vegetable Oils. In *Oxidative Stability and Shelf Life of Foods Containing Oils and Fats* (pp. 187–207). Massachusetts, USA: Elsevier Inc.
- Szabó, T., Berkesi, O., Forgó, P., Josepovits, K., Sanakis, Y., Petridis, D., & Dékány, I. (2006). Evolution of surface functional groups in a series of progressively oxidized graphite oxides. *Chemistry of Materials*, 18(11), 2740–2749.
- Szymańska, E., & Winnicka, K. (2015). Stability of chitosan A challenge for pharmaceutical and biomedical applications. *Marine Drugs*, *13*(4), 1819–1846.
- Talbot, G. (2016). The Stability and Shelf Life of Fats and Oils. In *The Stability and Shelf Life of Food* (2nd ed., pp. 461–503). Sawston, United Kingdom: Woodhead Publishing Limited.

- Tang, H., Ehlert, G. J., Lin, Y., & Sodano, H. A. (2012). Highly efficient synthesis of graphene nanocomposites. *Nano Letters*, 12(1), 84–90.
- Tang, S., Jin, S., Zhang, R., Liu, Y., Wang, J., Hu, Z., Lu, W., Yang, S., Qiao, W., Ling, L., & Jin, M. (2019). Effective reduction of graphene oxide via a hybrid microwave heating method by using mildly reduced graphene oxide as a susceptor. *Applied Surface Science*, 473, 222–229.
- Tang, Z., Wu, X., Guo, B., Zhang, L., & Jia, D. (2012). Preparation of butadiene– styrene–vinyl pyridine rubber–graphene oxide hybrids through co–coagulation process and in situ interface tailoring. *Journal of Materials Chemistry*, 22(15), 7492–7501.
- Tanuma, H., Kiuchi, H., Kai, W., Yazawa, K., & Inoue, Y. (2009). Characterization and enzymatic degradation of peg–cross–linked chitosan hydrogel films. *Journal of Applied Polymer Science*, 114, 1902–1907.
- Tayade, U. S., Borse, A. U., & Meshram, J. S. (2019). Green reduction of graphene oxide and its applications in band gap calculation and antioxidant activity. *Green Materials*, 7(3), 143–155.
- Tegou, E., Pseiropoulos, G., Filippidou, M. K., & Chatzandroulis, S. (2016). Lowtemperature thermal reduction of graphene oxide films in ambient atmosphere: Infra-red spectroscopic studies and gas sensing applications. *Microelectronic Engineering*, 159, 146–150.
- *The Nobel Prize in Physics 2010.* (2010, October 5). Retrieved from https://www.nobelprize.org/prizes/physics/2010/press-release/
- Tian, Z., Liu, W., & Li, G. (2016). The microstructure and stability of collagen hydrogel cross-linked by glutaraldehyde. *Polymer Degradation and Stability*, 130, 264–270.
- Tokur, B., Korkmaz, K., & Ayas, D. (2006). Comparison of two thiobarbituric acid (TBA) method for monitoring lipid oxidation in fish. *Ege Journal of Fisheries & Aquatic Sciences*, 23, 331–334.
- Tonelli, F. M., Goulart, V. A., Gomes, K. N., Ladeira, M. S., Santos, A. K., Lorençon, E., Ladeira, M. S., Santos, A. K., Lorençon, E., Ladeira, L. O., & Resende, R. R. (2015). Graphene–based nanomaterials: Biological and medical applications and toxicity. *Nanomedicine*, 10(15), 2423–2450.
- Toselli, M., Fabiani, D., Mancinelli, P., Fréchette, M., Heid, T., David, E., & Saccani, A. (2015). In situ thermal reduction of graphene oxide forming epoxy nanocomposites and their dielectric properties. *Polymer Composites*, *36*(2), 294–301.
- Tripathi, S., Mehrotra, G. K., & Dutta, P. K. (2009). Physicochemical and bioactivity of cross–linked chitosan–PVA film for food packaging applications. *International Journal of Biological Macromolecules*, 45(4), 372–376.
- Tschoppe, K., Beckert, F., Beckert, M., & Mülhaupt, R. (2015). Thermally reduced graphite oxide and mechanochemically functionalized graphene as functional fillers for epoxy nanocomposites. *Macromolecular Materials and Engineering*, 300(2), 140–152.
- Tseng, I. H., Liao, Y. F., Chiang, J. C., & Tsai, M. H. (2012). Transparent polyimide/graphene oxide nanocomposite with improved moisture barrier property. *Materials Chemistry and Physics*, 136(1), 247–253.

- Tummala, N. R., & Striolo, A. (2008). Role of counterion condensation in the self– assembly of SDS surfactants at the water–graphite interface. *Journal of Physical Chemistry B*, 112(7), 1987–2000.
- Vecitis, C. D., Zodrow, K. R., Kang, S., & Elimelech, M. (2010). Electronic-structuredependent bacterial cytotoxicity of single-walled carbon nanotubes. ACS Nano, 4(9), 5471–5479.
- Velickova, E., Winkelhausen, E., Kuzmanova, S., Moldão–Martins, M., & Alves, V. D. (2015). Characterization of multilayered and composite edible films from chitosan and beeswax. *Food Science and Technology International*, 21(2), 83–93.
- Vieira Ramos, F. J. H. T., Reis, R. H. M., Grafova, I., Grafov, A., & Monteiro, S. N. (2020). Eco-friendly recycled polypropylene matrix composites incorporated with geopolymer concrete waste particles. *Journal of Materials Research and Technology*, 9(3), 3084–3090.
- Voiry, D., Yang, J., Kupferberg, J., Fullon, R., Lee, C., Jeong, H. Y., Shin, H. S., & Chhowalla, M. (2016). High-quality graphene via microwave reduction of solution-exfoliated graphene oxide. *Science*, 353(6306), 1413–1416.
- Wallace, P. R. (1947). The band theory of graphite. *Physical Review*, 71(9), 622–634.
- Wang, G., Shen, X., Wang, B., Yao, J., & Park, J. (2009). Synthesis and characterisation of hydrophilic and organophilic graphene nanosheets. *Carbon*, 47(5), 1359–1364.
- Wang, J., Salihi, E. C., & Šiller, L. (2017). Green reduction of graphene oxide using alanine. *Materials Science and Engineering C*, 72, 1–6.
- Wang, J., Wang, X., Xu, C., Zhang, M., & Shang, X. (2011). Preparation of graphene/poly(vinyl alcohol) nanocomposites with enhanced mechanical properties and water resistance. *Polymer International*, 60(5), 816–822.
- Wang, W., Wang, Z., Liu, Y., Li, N., Wang, W., & Gao, J. (2012). Preparation of reduced graphene oxide/gelatin composite films with reinforced mechanical strength. *Materials Research Bulletin*, 47(9), 2245–2251.
- Wang, Y., Shi, Z. X., & Yin, J. (2011). Facile synthesis of soluble graphene via a green reduction of graphene oxide in tea solution and its biocomposites. ACS Applied Materials and Interfaces, 3(4), 1127–1133.
- Waraho, T., Mcclements, D. J., & Decker, E. A. (2011). Mechanisms of lipid oxidation in food dispersions. *Trends in Food Science and Technology*, 22(1), 3–13.
- Wexler, A. (1976). Vapor pressure equation for water in range 3 to 100 degrees C. Journal of Research of the National Bureau of Standards – A, 80A(3), 775–785.
- Wu, L.-L., Wang, J., He, X., Zhang, T., & Sun, H. (2014). Using graphene oxide to enhance the barrier properties of poly(lactic acid) film. *Packaging Technology and Science*, 27, 693–700.
- Wu, Z. S., Ren, W., Gao, L., Liu, B., Jiang, C., & Cheng, H. M. (2009). Synthesis of high-quality graphene with a pre-determined number of layers. *Carbon*, 47(2), 493–499.

- Wu, Z. S., Ren, W., Gao, L., Zhao, J., Chen, Z., Liu, B., Tang, D., Yu, B., Jiang, C., & Cheng, H. M. (2009). Synthesis of graphene sheets with high electrical conductivity and good thermal stability by hydrogen arc discharge exfoliation. *ACS Nano*, 3(2), 411–417.
- Xie, C., Wu, X., Guo, X., Long, C., Li, S., Hu, C. A. A., & Yin, Y. (2016). Maternal chitosan oligosaccharide supplementation affecting expression of circadian clock genes, and possible association with hepatic cholesterol accumulation in suckling piglets. *Biological Rhythm Research*, 47(2), 253–265.
- Yamoneka, J., Malumba, P., Lognay, G., Blecker, C., & Danthine, S. (2019). *Irvingia gabonensis* seed fat as hard stock to formulate blends for trans free margarines. *LWT – Food Science and Technology*, 101, 747–756.
- Yan, H., Jiang, L., Xu, X., Li, Y., Shen, Y., & Zhu, S. (2017). Ultrastrong composite film of chitosan and silica-coated graphene oxide sheets. *International Journal of Biological Macromolecules*, 104, 936–943.
- Yan, J., Hu, C., Chen, K., & Lin, Q. (2019). Release of graphene from graphene– polyethylene composite films into food simulants. *Food Packaging and Shelf Life*, 20, 100310.
- Yan, L., Wang, Y., Xu, X., Zeng, C., Hou, J., Lin, M., Xu, J., Sun, F., Huang, X., Dai, L., Lu, F., & Liu, Y. (2012). Can graphene oxide cause damage to eyesight? *Chemical Research in Toxicology*, 25(6), 1265–1270.
- Yan, N., Capezzuto, F., Buonocore, G. G., Tescione, F., Lavorgna, M., Xia, H., & Ambrosio, L. (2015). Borate cross-linking chitosan/graphene oxide films: Toward the simultaneous enhancement of gases barrier and mechanical properties. *AIP Conference Proceedings*, 1695(1), 020018.
- Yang, B., Dong, N., & Wang, S. (2018). Qualitative analysis of reduction degree in reduced graphene oxide solution by femtosecond laser-induced breakdown spectroscopy. *IOP Conference Series: Materials Science and Engineering*, 382(2), 022020.
- Yang, D., Velamakanni, A., Bozoklu, G., Park, S., Stoller, M., Piner, R. D., Stankovich, S., Jung, I., Field, D. A., Ventrice Jr., C. A., & Ruoff, R. S. (2009). Chemical analysis of graphene oxide films after heat and chemical treatments by X–ray photoelectron and Micro–Raman spectroscopy. *Carbon*, 47(1), 145–152.
- Yang, K., Wan, J., Zhang, S., Zhang, Y., Lee, S.-T., & Liu, Z. (2010). In vivo pharmacokinetics, long-term biodistribution and toxicology of PEGylated graphene in mice. ACS Nano, 5(1), 516–522.
- Yang, K., Zhang, S., Zhang, G., Sun, X., Lee, S. T., & Liu, Z. (2010). Graphene in mice: Ultrahigh in vivo tumor uptake and efficient photothermal therapy. *Nano Letters*, 10(9), 3318–3323.
- Yang, X., Tu, Y., Li, L., Shang, S., & Tao, X. (2010). Well–dispersed chitosan/graphene oxide nanocomposites. ACS Applied Materials and Interfaces, 2(6), 1707–1713.
- Yang, Y.-H., Bolling, L., Haile, M., & Grunlan, J. C. (2012). Improving oxygen barrier and reducing moisture sensitivity of weak polyelectrolyte multilayer thin films with crosslinking. *RSC Advances*, 2(32), 12355–12363.

- Yang, Y.-M., Zhao, Y.-H., Liu, X.-H., Ding, F., & Gu, X.-S. (2007). The effect of different sterilization procedures on chitosan dried powder. *Journal of Applied Polymer Science*, 104, 1968–1972.
- Yang, Z. Z., Zheng, Q. Bin, Qiu, H. X., Li, J., & Yang, J. H. (2015). A simple method for the reduction of graphene oxide by sodium borohydride with CaCl<sub>2</sub> as a catalyst. *Xinxing Tan Cailiao/New Carbon Materials*, 30(1), 41–47.
- Ye, S., & Feng, J. (2013). A new insight into the in situ thermal reduction of graphene oxide dispersed in a polymer matrix. *Polymer Chemistry*, 4(6), 1765–1768.
- Yeng, C. M., Husseinsyah, S., & Ting, S. S. (2013). Chitosan/corn cob biocomposite films by cross-linking with glutaraldehyde. *BioResources*, 8(2), 2910–2923.
- Yesildag, N., Hopmann, C., Adamy, M., & Windeck, C. (2017). Properties of polyamide 6-graphene-composites produced and processed on industrial scale. AIP Conference Proceedings, 1914, 150001.
- Yildirim, S., & Röcker, B. (2018). Active packaging. In Nanomaterials for Food Packaging: Materials, Processing Technologies, and Safety Issues (pp. 173–202). Amsterdam, Netherlands: Elsevier Inc.
- Yoksan, R., & Chirachanchai, S. (2010). Silver nanoparticle-loaded chitosan-starch based films: Fabrication and evaluation of tensile, barrier and antimicrobial properties. *Materials Science & Engineering C*, 30(6), 891–897.
- Yoo, B. M., Shin, H. J., Yoon, H. W., & Park, H. B. (2014). Graphene and graphene oxide and their uses in barrier polymers. *Journal of Applied Polymer Science*, 131(1), 39628.
- Young, R. J., Kinloch, I. A., Gong, L., & Novoselov, K. S. (2012). The mechanics of graphene nanocomposites: A review. *Composites Science and Technology*, 72(12), 1459–1476.
- Yousefi, M., Dadashpour, M., Hejazi, M., Hasanzadeh, M., Behnam, B., de la Guardia, M., Shadjou, N., & Mokhtarzadeh, A. (2017). Anti-bacterial activity of graphene oxide as a new weapon nanomaterial to combat multidrug-resistance bacteria. *Materials Science and Engineering: C*, 74, 568–581.
- Youssef, A. M., & El–Sayed, S. M. (2018). Bionanocomposites materials for food packaging applications: Concepts and future outlook. *Carbohydrate Polymers*, 193, 19–27.
- Youssef, A. M., El–Sayed, S. M., El–Sayed, H. S., Salama, H. H., & Dufresne, A. (2016). Enhancement of Egyptian soft white cheese shelf life using a novel chitosan/carboxymethyl cellulose/zinc oxide bionanocomposite film. *Carbohydrate Polymers*, 151, 9–19.
- Yu, L., Lim, Y. S., Han, J. H., Kim, K., Kim, J. Y., Choi, S. Y., & Shin, K. (2012). A graphene oxide oxygen barrier film deposited via a self–assembly coating method. *Synthetic Metals*, *162*(7–8), 710–714.
- Yuan, G., Chen, X., & Li, D. (2016). Chitosan films and coatings containing essential oils: The antioxidant and antimicrobial activity, and application in food systems. *Food Research International*, 89, 117–128.

- Yuan, J., Gao, H., Sui, J., Duan, H., Chen, W. N., & Ching, C. B. (2012). Cytotoxicity evaluation of oxidized single-walled carbon nanotubes and graphene oxide on human hepatoma HepG2 cells: An iTRAQ-coupled 2D LC-MS/MS proteome analysis. *Toxicological Sciences*, 126(1), 149–161.
- Yuan, R., Yuan, J., Wu, Y., Chen, L., Zhou, H., & Chen, J. (2017). Efficient synthesis of graphene oxide and the mechanisms of oxidation and exfoliation. *Applied Surface Science*, 416, 868–877.
- Yuan, X., Zheng, J., Jiao, S., Cheng, G., Feng, C., Du, Y., & Liu, H. (2019). A review on the preparation of chitosan oligosaccharides and application to human health, animal husbandry and agricultural production. *Carbohydrate Polymers*, 220, 60– 70.
- Zakhem, E., & Bitar, K. (2015). Development of chitosan scaffolds with enhanced mechanical properties for intestinal tissue engineering applications. *Journal of Functional Biomaterials*, 6(4), 999–1011.
- Zeid, A., Karabagias, I. K., Nassif, M., & Kontominas, M. G. (2019). Preparation and evaluation of antioxidant packaging films made of polylactic acid containing thyme, rosemary, and oregano essential oils. *Journal of Food Processing and Preservation*, 43(10), 1–11.
- Zhang, D., Yang, S., Chen, Y., Liu, S., Zhao, H., & Gu, J. (2018). 60Co γ–ray irradiation crosslinking of chitosan/graphene oxide composite film: Swelling, thermal stability, mechanical, and antibacterial properties. *Polymers*, *10*(3), 294.
- Zhang, J., Yang, H., Shen, G., Cheng, P., Zhang, J., & Guo, S. (2010). Reduction of graphene oxide vial-ascorbic acid. *Chemical Communications*, 46(7), 1112–1114.
- Zhang, J., Ding, E., Xu, S., Li, Z., Fakhri, A., & Gupta, V. K. (2020). Production of metal oxides nanoparticles based on poly–alanine/chitosan/reduced graphene oxide for photocatalysis degradation, anti-pathogenic bacterial and antioxidant studies. *International Journal of Biological Macromolecules*, 164, 1584–1591.
- Zhang, L., Liang, J., Huang, Y., Ma, Y., Wang, Y., & Chen, Y. (2009). Size-controlled synthesis of graphene oxide sheets on a large scale using chemical exfoliation. *Carbon*, 47(14), 3365–3368.
- Zhang, Q., Qian, X., Thebo, K. H., Cheng, H. M., & Ren, W. (2018). Controlling reduction degree of graphene oxide membranes for improved water permeance. *Science Bulletin*, 63(12), 788–794.
- Zhang, W., & Jiang, W. (2020). Antioxidant and antibacterial chitosan film with tea polyphenols-mediated green synthesis silver nanoparticle via a novel one-pot method. *International Journal of Biological Macromolecules*, 155, 1252–1261.
- Zhang, X., Yin, J., Peng, C., Hu, W., Zhu, Z., Li, W., Fan, C., & Huang, Q. (2011). Distribution and biocompatibility studies of graphene oxide in mice after intravenous administration. *Carbon*, 49(3), 986–995.
- Zhang, Y., Mo, G., Li, X., Zhang, W., Zhang, J., Ye, J., Huang, X., & Yu, C. (2011). A graphene modified anode to improve the performance of microbial fuel cells. *Journal of Power Sources*, 196(13), 5402–5407.

- Zhang, Y., Zhang, M., Jiang, H., Shi, J., Li, F., Xia, Y., Zhang, G., & Li, H. (2017). Bioinspired layered chitosan/graphene oxide nanocomposite hydrogels with high strength and pH-driven shape memory effect. *Carbohydrate Polymers*, 177, 116– 125.
- Zhao, D., Yu, S., Sun, B., Gao, S., Guo, S., & Zhao, K. (2018). Biomedical applications of chitosan and its derivative nanoparticles. *Polymers*, 10(4), 462.
- Zhao, J., Li, Y., Wang, Y., Mao, J., He, Y., & Luo, J. (2017). Mild thermal reduction of graphene oxide as a lubrication additive for friction and wear reduction. *RSC Advances*, 7(3), 1766–1770.
- Zhong, Q. P., & Xia, W. S. (2008). Physicochemical properties of edible and preservative films from chitosan/cassava starch/gelatin blend plasticized with glycerol. *Food Technology and Biotechnology*, 46(3), 262–269.
- Zhou, T., Qi, X., Bai, H., & Fu, Q. (2016). The different effect of reduced graphene oxide on the performance of chitosan by using homogenous fillers. *RSC Advances*, *6*, 34153–34158.
- Zhu, Y., Murali, S., Cai, W., Li, X., Suk, J. W., Potts, J. R., & Ruoff, R. S. (2010). Graphene and graphene oxide: Synthesis, properties, and applications. Advanced Materials, 22(35), 3906–3924.
- Zivanovic, S., Chi, S., & Draughon, A. E. (2005). Antimicrobial Activity of Chitosan. *Journal of Food Science*, 70(1), 45–51.