## Widespread shifts in body size within populations and assemblages

Inês S. Martins ${ }^{1,2^{*}}$, Franziska Schrodt ${ }^{3}$, Shane A. Blowes ${ }^{4,5}$, Amanda E. Bates ${ }^{6}$, Anne D. Bjorkman ${ }^{7,8}$, Viviana Brambilla ${ }^{1,9}$, Juan Carvajal-Quintero ${ }^{4,12}$, Cher F. Y. Chow ${ }^{1}$, Gergana N. Daskalova ${ }^{10}$, Kyle Edwards ${ }^{11}$, Nico Eisenhauer ${ }^{4}, 12$, Richard Field ${ }^{3}$, Ada Fontrodona- Eslava ${ }^{1}$, Jonathan J. Henn ${ }^{13,14}$, Roel van Klink ${ }^{4,5}$, Joshua S. Madin ${ }^{15}$, Anne E. Magurran ${ }^{1}$, Michael McWilliam ${ }^{1}$, Faye Moyes ${ }^{1}$, Brittany Pugh ${ }^{3,16}$, Alban Sagouis ${ }^{4,5}$, Isaac TrindadeSantos ${ }^{1,17}$, Brian McGill ${ }^{18}$, Jonathan M. Chase ${ }^{4,5}$, Maria Dornelas ${ }^{1,2,9}$.

## Affiliations:

${ }^{1}$ Centre for Biological Diversity, School of Biology, University of St Andrews; St Andrews, KY16 9TH, Scotland.
${ }^{2}$ Leverhulme Centre for Anthropocene Biodiversity, Berrick Saul Second Floor, University of York; York YO10 5DD.
${ }^{3}$ University of Nottingham, School of Geography; University Park, NG7 2RD, Nottingham, UK.
${ }^{4}$ German Centre for Integrative Biodiversity Research (iDiv) Halle-Jena-Leipzig; 04103 Leipzig, Germany.
${ }^{5}$ Department of Computer Science, Martin Luther University Halle-Wittenberg; 06099 Halle (Saale), Germany.
${ }^{6}$ Department of Biology, University of Victoria, Victoria, BC, Canada.
${ }^{7}$ Department of Biological and Environmental Sciences, University of Gothenburg, Gothenburg, Sweden.
${ }^{8}$ Gothenburg Global Biodiversity Centre, Gothenburg, Sweden.
${ }^{9}$ MARE, Guia Marine Laboratory, Faculty of Sciences, University of Lisbon; 2750-374 Cascais, Portugal.
${ }^{10}$ International Institute for Applied Systems Analysis (IIASA), Laxenburg, Austria.
${ }^{11}$ Department of Oceanography, University of Hawai‘i at Mānoa, Honolulu, HI, USA.
${ }^{12}$ Institute of Biology; Leipzig University; 04103 Leipzig, Germany.
${ }^{13}$ Department of Evolution, Ecology, and Organismal Biology, University of California Riverside, Riverside, CA USA.
${ }^{14}$ Institute of Arctic and Alpine Research, University of Colorado Boulder, Boulder, CO USA.
${ }^{15}$ Hawai‘i Institute of Marine Biology, University of Hawai‘i at Manoa, Kāne‘ohe, Hawai‘i, USA.
${ }^{16}$ University College London, School of Geography; Gower Street, London, WC1E 6AE, UK.
${ }^{17}$ Macroevolution Unit, Okinawa Institute of Science and Technology Graduate University, 1919-1, Tancha, Onna-son, Kunigami-gun, 904-0495, Okinawa, Japan.
${ }^{18}$ School of Biology and Ecology and Mitchell Center for Sustainability Solutions, University of Maine, Orono, Maine, USA.
*Corresponding author: istmartins@gmail.com


#### Abstract

Biotic responses to global change include directional shifts in organismal traits. Body size, an integrative trait that determines demographic rates and ecosystem functions, is thought to be shrinking in the Anthropocene. Here, we assess the prevalence of body size change in six taxon groups across 5,025 assemblage time-series spanning 1960 to 2020. Using the Price equation to partition this change into within-species body size versus compositional changes, we detect prevailing decreases in body size through time driven primarily by fish, with more variable patterns in other taxa. Change in assemblage composition contributes more to body size changes than within-species trends, but both components show substantial variation in magnitude and direction. The biomass of assemblages remains remarkably stable as decreases in body size tradeoff with increases in abundance.


One-Sentence Summary: Variable within-species and compositional shifts combine into shrinking body size, abundance increases and stable biomass through time.

## Main Text:

The loss or gain of large organisms can have dramatic consequences for ecosystem functions in terms of total system biomass and metabolism, and thus food web energy fluxes (1). Anthropogenic changes to the biosphere are miniaturizing many communities (1-3) due to the selective removal of the largest individuals (population change e.g. (4)) and the extinction of larger species (compositional change e.g. (5)). Population shifts in body size have been attributed to anthropogenic selection forces, including preferential exploitation of larger individuals ( 6,7 ), climate change and habitat conversion (8-10). At the assemblage-level, species compositional change is a major component of biodiversity change (11, 12). Largebodied species have been particularly susceptible to extinction following temperature shifts (in either direction) and human exploitation, largely due to their life history traits and lower abundances (13). Yet shrinking trends of community body sizes are by no means universal and, when examined across many communities, some results suggest no net change in body size (14). Indeed, as species shift their ranges, some communities are gaining larger species, such as in arctic and high-elevation plant assemblages (6). Hence, the prevalence of population and compositional body size changes and their implications for assemblage abundance and biomass are unknown.

Previous assessments have investigated either compositional components of body size change (14) or within-species population changes (3), or when both components are examined, have focused on a single taxon (15). Yet, interactions of change in the two components can lead to non-intuitive outcomes in body size distributions through time. For example, the loss of competitors arising from compositional change can allow the remaining populations to increase in body size (10). Several scenarios of body size change can arise by simultaneously considering within-species population and compositional body size changes (Fig. 1). The two components can operate in the same direction (both increasing or decreasing body size), or change can involve only one component (change only in population or composition, but not both). Finally, change in one component can cancel out change in the other. Here, we explicitly set out to test to what extent each of the two components most commonly drives the changes in body size we observe across taxa and regions, and how often change in the two components occurs in opposite directions.
To examine these scenarios, we used time-series that recorded organism abundance and body size (biomass) data in the field, and quantified the contribution of both components, compositional and within-species body size changes, to change in body size distributions across taxa. Specifically, we collated 5,025 assemblages over 60 years (17), a time period of intensification of anthropogenic selection forces on the biosphere (18). These time-series range from 5-56 years of surveys and cover 4,292 species within six taxonomic groups (1971 fish sp., 1201 plants sp., 628 invertebrates sp., 66 mammals sp., 33 herpetofauna sp., and 393 marine benthic organisms) from communities distributed across multiple regions of the world (fig. S1). The time-series cover a variety of body sizes and body size change trends. We found body size shrinking in more assemblages than increasing across all datasets as well as among time-series that had stronger evidence for trends of change (lower p-values, see Fig. 2, fig. S3 and (19) for details).

We quantify the prevalence of different components of change (Fig. 1) by decomposing body size change of all assemblages into compositional and within-species changes by using an extension of the Price equation (19-21). The Price equation is a mathematical description of the relationship between statistical descriptors (mean and covariance) of selection and trait change (22). Although developed in an evolutionary context, this equation has direct application to the question of body size change through time if we equate competition and
environmental filtering as selection ((23); but see also (24)). By examining the type of covariance in these two components of change, we can determine the relative contributions of compositional and within-species change to the observed overall change (Fig. 1B).

## RESULTS AND DISCUSSION

## Partitioning patterns of body size change

Our analysis shows that over all assemblages, body size is predominantly shrinking, with substantial variation in the balance of within-species change and compositional change (Fig. 3). Two thirds of the 5,025 assemblages decreased in average body size and one-third increased. In fact, both components of body size change were present in the overwhelming majority of assemblages ( $96.4 \%$ ), with the magnitude of compositional change being greater than within-species change in $72 \%$ of assemblages. Although compositional and withinspecies change often occurred in the same direction ( $58.8 \%$ of assemblages), we found counteracting effects in $41.2 \%$ of all assemblages. For example, of the 3,415 assemblages showing within-species decreases in body size, $\sim 35 \%$ had increases in body size associated with compositional change (within-species change $<0$ and compositional change $>0$, Fig. 3). This substantial variation in magnitude and direction of the two components of body size change and their interactions suggest both components need to be considered when assessing change in body size. While duration and time period of the time-series vary, our results are robust to the length of the time-series, the start and end dates, as well as intermediate states (pairwise comparisons among years; see sensitivity analysis in fig. S5-S6).
We found that trends in body size change over time vary among taxa, realm, and latitude (Fig. 4). Our confidence in estimates of body size change is highest for the most well represented taxon in our dataset-marine fish-which show a particularly evident decrease in body size (Fig. 4A). Among other taxa, the number of available time-series is lower and body size change trends are more variable. In fact, when fish are removed from our analysis, neither increases nor decreases dominate the overall body size change trends across the remaining dataset (fig. S3C). Among non-fish assemblages (e.g., benthos-mainly marine invertebrates, plants) the role of within-species and compositional changes is also more variable, but where the patterns of within-species are stronger ( 492 of 1116 non-fish time-series) there is a tendency towards increasing body size ( $57 \%$ of assemblages), counteracting the tendency observed in fish assemblages (Fig. 4A). When we compared overlapping data (assemblages and species) with an extended dataset which uses species' average body size estimates taken from trait databases (fig. S2), we found remarkable consistency for both fish and non-fish assemblages (fig. S3E-H). Nevertheless, we maintain that considering both axes of variation (compositional and within-species) is crucial to avoid potentially misleading conclusions that arise when the two components change in opposite directions. For instance, when we estimated global trends in body size change across all available datasets by fitting a Bayesian mixed-effects model, we did not detect any clear pattern of change (with or without fish; fig. S9-11) (19). This was true regardless of whether we use the same data as in our main analysis that directly measured body size trends, or the full extended dataset which includes 20,173 assemblage time-series with body size inferred from trait databases (fig. S2-S3) (see also (14)). This extended dataset also highlights that neither increases nor decreases dominate mean body size change other than in marine fish, even when there are few more (substantially more for birds) available time-series for other taxa (fig. S12). This result emphasizes that we should take caution against extrapolating or over-interpreting trait changes across taxa, particularly when data on within-species change are not available. More data are needed to determine the prevalence of body size change through time in non-fish taxa.

Despite caveats, we consistently detect the signal of shrinking body size in marine fish for both types of body size data, and regardless of whether we used ordinary least-squares (OLS) slope estimates or the Price equation approach. Our observation of shrinking among marine fish assemblages aligns with previous evidence (25-27). For marine fishes, these changes are often linked to the selective exploitation of large-bodied individuals by humans (25), to warming (26), and/or to decreased resource availability (27). Furthermore, disturbances and selective removal of larger individuals affects the age and size structure of populations, fish or otherwise, as well as the genetic structure within populations (e.g., plants (28)). Such responses may be particularly prevalent among fish because of the widespread effects of overexploitation. Across assemblages, it is likely that combinations of these drivers result in the high variability in trends and prevalence of counteracting effects that we observed in these time-series.

Collectively, our analyses reveal that both within-species and compositional changes combine to create high variability in the observed outcomes of assemblage-level body size changes through time. These findings highlight the importance of considering the separate and interactive effects of compositional and within-species body size change. Specifically, the community context is necessary to understand within-species change. For example, removing top predators (often the larger-body size individuals in an assemblage) can trigger mesopredator release, which alters assemblage size structure and composition (29). While it is possible that some of these dynamics are missed if predators and prey belong to different sampled assemblages, the assemblage level is where regulation will play out, for example, in the context of ecological carrying capacity (30).

The selection forces acting on body size are varied and have heterogeneous distributions in space and time. By partitioning body size change into within-species and compositional change, as we have done here, we can begin to explain the wide variation in body size change patterns through time found in literature. For example, global warming is simultaneously selecting for smaller body size (for metabolic reasons), affecting species' phenology, and causing range shifts (2). Global warming and species phenology effects can best be seen in within-species changes, while range shifts induce compositional change. The net result of these processes will depend on the environmental context. For example, in the Arctic Tundra, warming promotes larger shrubs (6), because species from warmer areas are expanding their ranges and because there are longer growing seasons. In contrast, warming is associated with smaller fish in the North Sea (9), although selective harvesting/exploitation is likely also contributing to this change (31). By considering both within-species and compositional changes in individual-level body size, alongside changes in relative abundance, future research should be able to better elucidate the mechanisms involved in how body size is changing through time.

## Relationships between changes in body size, abundance, and biomass

Body size is usually tightly linked to abundance (32) through both metabolic (33) and trophic $(34,35)$ processes. This relationship can have implications for assemblage biomass mediated by a trade-off between size and abundance (36). Hence, we further investigated if the changes in body size were associated with changes in assemblage abundance, biomass, or both (Fig. 5; fig. S13). We found that abundance has, on average, slightly increased through time across assemblages, while the overall change in assemblage biomass is indistinguishable from zero (Fig. 5). Previously, no (14), or complex (37) relationships have been found between body size and abundance changes, although for invertebrates such a relationship is often negative (38). While our results confirm that the relationship between abundance change and body size
change is complex and variable, there are signs that the overall reduction in body size is being counteracted by increasing overall abundance (Fig. 5A and C). Such trade-offs between abundance and body size are often expected (32) and affect ecosystem metabolic rate and function (24,39). We detect a strong positive covariance between change in biomass and abundance (Fig. 5D), but much weaker covariance between abundance and body size, with the strongest trends among these two variables tending to negative covariance (Fig. 5C). These patterns in covariance are robust to removing fish time series from the analysis, despite the fact that overall change in body size is not detected in that case (Fig. S14). In fact, $79 \%$ of the assemblages with detectable trends in both variables have abundance increases and body size decreases. These patterns suggest that assemblage body size, abundance and biomass are linked and that change in one has implications for change in the others. There is evidence of widespread regulation of assemblage-level variables (species richness and abundance) whereby assemblages tend to return to previous levels after disturbances (40). The lack of a clear directional trend in biomass in our study suggests that it may be more tightly regulated than body size and abundance, which may cause trade-offs in change of the latter two variables.

## Conclusions

We find evidence of widespread body size shrinking through time as a result of both population and community-level changes despite substantial variation, and overall stable assemblage-level biomass. Not all taxa contributed equally to the observed changes we report. We find the most widespread declines among fish assemblages, but a greater balance of increases and declines in other taxa. Body size is an easily measured, integrative, and important morphological trait that scales with many ecological characteristics of organisms and ecosystems, such as demographic rates, metabolism and resource requirements (41, 42). We reiterate pleas for more regular monitoring of body size (43), especially for taxa other than marine fish and ideally in conjunction with abundance estimates. Future research could focus on the implications of body size changes for ecosystem functions. For instance, cascading food web effects of shrinking body size could negatively affect human nutrition and associated economics (e.g., affecting crop plants and protein sources such as fish; (44). Moreover, shrinking body size through compositional change is likely to bring changes in other traits, and therefore trigger additional impacts on ecosystem functioning (8). Our study suggests the ubiquitous turnover in biodiversity composition currently unfolding $(11,12)$ is a profound re-shuffling of not only species, but also key characteristics of living organisms.

## References and Notes

1. B. J. Enquist, A. J. Abraham, M. B. J. Harfoot, Y. Malhi, C. E. Doughty, The megabiota are disproportionately important for biosphere functioning. Nature Communications 11, 699 (2020).
2. J. L. Gardner, A. Peters, M. R. Kearney, L. Joseph, R. Heinsohn, Declining body size: a third universal response to warming? Trends in ecology \& evolution 26, 285-291 (2011).
3. I. A. Hatton, R. F. Heneghan, Y. M. Bar-On, E. D. Galbraith, The global ocean size spectrum from bacteria to whales. Science Advances 7, eabh3732 (2021).
4. M. Heino, B. Díaz Pauli, U. Dieckmann, Fisheries-Induced Evolution. Annual Review of Ecology, Evolution, and Systematics 46, 461-480 (2015).
5. F. A. Smith, R. E. Elliott Smith, S. K. Lyons, J. L. Payne, Body size downgrading of mammals over the late Quaternary. Science 360, 310-313 (2018).
6. A. D. Bjorkman et al., Plant functional trait change across a warming tundra biome. Nature 562, 57-62 (2018).
7. W. J. Ripple et al., Are we eating the world's megafauna to extinction? Conservation Letters 12, e12627 (2019).
8. I. Bartomeus et al., Historical changes in northeastern US bee pollinators related to shared ecological traits. Proceedings of the National Academy of Sciences 110, 46564660 (2013).
9. M. Daufresne, K. Lengfellner, U. Sommer, Global warming benefits the small in aquatic ecosystems. Proceedings of the National Academy of Sciences 106, 1278812793 (2009).
10. F. He et al., The global decline of freshwater megafauna. Global Change Biology 25, 3883-3892 (2019).
11. S. A. Blowes et al., The geography of biodiversity change in marine and terrestrial assemblages. Science 366, 339-345 (2019).
12. M. Dornelas et al., Assemblage Time Series Reveal Biodiversity Change but Not Systematic Loss. Science 344, 296-299 (2014).
13. J. Dembitzer, R. Barkai, M. Ben-Dor, S. Meiri, Levantine overkill: 1.5 million years of hunting down the body size distribution. Quaternary Science Reviews 276, 107316 (2022).
14. J. C. D. Terry, J. D. O’Sullivan, A. G. Rossberg, No pervasive relationship between species size and local abundance trends. Nature Ecology \& Evolution 6, 140-144 (2022).
15. H.-F. Wang, M. Xu, Individual size variation reduces spatial variation in abundance of tree community assemblage, not of tree populations. Ecology and Evolution 7, 1081510828 (2017).
16. T. Dayan, D. Simberloff, Size patterns among competitors: ecological character displacement and character release in mammals, with special reference to island populations. Mammal Review 28, 99-124 (1998).
17. M. Dornelas et al., BioTIME: A database of biodiversity time series for the Anthropocene. Global Ecology and Biogeography 27, 760-786 (2018).
18. T. Carroll, F. Cardou, M. Dornelas, C. D. Thomas, M. Vellend, Biodiversity change under adaptive community dynamics. Global Change Biology 00, 1-14 (2023).
19. I. S. Martins et al., Supplementary Materials for "Widespread shifts in body size within populations and assemblages.". (2023).
20. S. A. Frank, Natural selection. IV. The Price equation*. Journal of Evolutionary Biology 25, 1002-1019 (2012).
21. A. Gardner, Price's equation made clear. Philosophical Transactions of the Royal Society B: Biological Sciences 375, 20190361 (2020).
22. G. R. Price, Selection and Covariance. Nature 227, 520-521 (1970).
23. M. Vellend, Conceptual synthesis in community ecology. The Quarterly review of biology 85, 183-206 (2010).
24. P. Pillai, T. C. Gouhier, Not even wrong: the spurious measurement of biodiversity's effects on ecosystem functioning. Ecology 100, e02645 (2019).
25. N. E. Bosch et al., Effects of human footprint and biophysical factors on the body-size structure of fished marine species. Conservation Biology 36, e13807 (2022).
26. J. L. Gardner et al., Temporal patterns of avian body size reflect linear size responses to broadscale environmental change over the last 50 years. Journal of Avian Biology 45, 529-535 (2014).
27. J. A. Sheridan, D. Bickford, Shrinking body size as an ecological response to climate change. Nature Climate Change 1, 401-406 (2011).
28. D. B. Lindenmayer, W. F. Laurance, J. F. Franklin, Global Decline in Large Old Trees. Science 338, 1305-1306 (2012).
29. T. M. Newsome et al., Top predators constrain mesopredator distributions. Nature Communications 8, 15469 (2017).
30. J. H. Brown, S. M. Ernest, J. M. Parody, J. P. Haskell, Regulation of diversity: maintenance of species richness in changing environments. Oecologia 126, 321-332 (2001).
31. S. Jennings et al., Long-term trends in the trophic structure of the North Sea fish community: evidence from stable-isotope analysis, size-spectra and community metrics. Marine Biology 141, 1085-1097 (2002).
32. E. P. White, S. K. M. Ernest, A. J. Kerkhoff, B. J. Enquist, Relationships between body size and abundance in ecology. Trends in Ecology \& Evolution 22, 323-330 (2007).
33. J. Damuth, Population density and body size in mammals. Nature 290, 699-700 (1981).
34. J. E. Cohen, T. Jonsson, S. R. Carpenter, Ecological community description using the food web, species abundance, and body size. Proceedings of the National Academy of Sciences 100, 1781-1786 (2003).
35. J. L. Blanchard et al., How does abundance scale with body size in coupled sizestructured food webs? Journal of Animal Ecology 78, 270-280 (2009).
36. Ethan P. White, S. K. M. Ernest, Katherine M. Thibault, Trade-offs in Community Properties through Time in a Desert Rodent Community. The American Naturalist 164, 670-676 (2004).
37. M. J. Genner et al., Body size-dependent responses of a marine fish assemblage to climate change and fishing over a century-long scale. Global Change Biology 16, 517527 (2010).
38. M. A. K. Gillespie, T. Birkemoe, A. Sverdrup-Thygeson, Interactions between body size, abundance, seasonality, and phenology in forest beetles. Ecology and Evolution 7, 1091-1100 (2017).
39. J. H. Brown, J. F. Gillooly, A. P. Allen, V. M. Savage, G. B. West, Toward a metabolic theory of ecology. Ecology 85, 1771-1789 (2004).
40. N. J. Gotelli et al., Community-level regulation of temporal trends in biodiversity. Science Advances 3, e1700315 (2017).
41. M. Flannery, Small, Medium or Large: Why Is Size so Important? The American Biology Teacher 51, 122-125 (1989).
42. M. Tseng et al., Decreases in beetle body size linked to climate change and warming temperatures. Journal of Animal Ecology 87, 647-659 (2018).
43. G. Woodward et al., Body size in ecological networks. Trends in Ecology \& Evolution 20, 402-409 (2005).
44. A. Audzijonyte, A. Kuparinen, R. Gorton, E. A. Fulton, Ecological consequences of body size decline in harvested fish species: positive feedback loops in trophic interactions amplify human impact. Biology Letters 9, 20121103 (2013).
45. I. S. Martins et al., Code and data for "Widespread shifts in body size within populations and assemblages.". Zenodo https://doi.org/10.5281/zenodo.7969814, (2023).
46. L. H. Antão et al., Temperature-related biodiversity change across temperate marine and terrestrial systems. Nature Ecology \& Evolution, (2020).
47. R. Barnes, K. Sahr, dggridR: Discrete Global Grids for R. R package version 2.0.4. "https://github.com/r-barnes/dggridR/" doi:10.5281/zenodo.1322866. (2017).
48. N. J. Gotelli, R. K. Colwell, Quantifying biodiversity: procedures and pitfalls in the measurement and comparison of species richness. Ecology Letters 4, 379-391 (2001).
49. B. F. Oliveira, V. A. São-Pedro, G. Santos-Barrera, C. Penone, G. C. Costa, AmphiBIO, a global database for amphibian ecological traits. Scientific Data 4, 170123 (2017).
50. J. Kattge et al., TRY - a global database of plant traits. Global Change Biology 17, 2905-2935 (2011).
51. H. Wilman et al., EltonTraits 1.0: Species-level foraging attributes of the world's birds and mammals. Ecology 95, 2027-2027 (2014).
52. R. Froese, D. Pauly, Editors. FishBase. World Wide Web electronic publication. www.fishbase.org, version (02/2019) (2019).
53. K. Homburg, N. Homburg, F. Schäfer, A. Schuldt, T. Assmann, Carabids.org - a dynamic online database of ground beetle species traits (Coleoptera, Carabidae). Insect Conservation and Diversity 7, 195-205 (2014).
54. S. A. Chamberlain, E. Szöcs, taxize: taxonomic search and retrieval in R. F1000Research 2, (2013).
55. P.-C. Bürkner, brms: An R Package for Bayesian Multilevel Models Using Stan. Journal of Statistical Software 80, 28 (2017).
56. RCoreTeam. (R Foundation for Statistical Computing, http://www.R-project.org/, Vienna, Austria, 2019).
57. R. Holmes, F. Sturges, Bird community dynamics and energetics in a northern hardwoods ecosystem. The Journal of Animal Ecology 1, 175-200 (1975).
58. R. T. Holmes, T. W. Sherry, Assessing population trends of New Hampshire forest birds: local vs. regional patterns. The Auk 105, 756-768 (1988).
59. R. T. Holmes, T. W. Sherry, Thirty-year bird population trends in an unfragmented temperate deciduous forest: importance of habitat change. The Auk 118, 589-609 (2001).
60. R. T. Holmes, T. W. Sherry, F. W. Sturges, Bird Community Dynamics in a Temperate Deciduous Forest: Long-Term Trends at Hubbard Brook. Ecological Monographs 56, 201-220 (1986).
61. K. J. Gaston, T. M. Blackburn, Pattern and process in macroecology., (WileyBlackwell, Oxford, England, 2000).
62. G. Beven, Changes in breeding bird populations of an oak-wood on Bookham Common, Surrey, over twenty-seven years. London Naturalist 55, 23-42 (1976).
63. D. W. Gibbons, J. B. Reid, R. A. Chapman, The new atlas of breeding birds in Britain and Ireland: 1988-1991. (T \& AD Poyser London, 1993).
64. B. Stone et al., Population estimates of birds in Britain and in the United Kingdom. British Birds 90, 1-22 (1997).
65. P. Lack, The atlas of wintering birds in Britain and Ireland. (A\&C Black, 2010).
66. P. Standley, N. Bucknell, A. Swash, I. Collins, The Birds of Berkshire. (Berkshire Atlas Group, Reading, UK, 1996).
67. M. Williamson, in Symposium of the British Ecological Society. (1987).
68. C. B. Halpern, C. Dyrness, "Plant succession and biomass dynamics following logging and burning in the Andrews Experimental Forest Watersheds 1 and 3, 1962-Present". Long-Term Ecological Research. Forest Science Data Bank, Corvallis. Available at: http://andrewsforest.oregonstate.edu/data/abstract.cfm?dbcode=TP073, accessed 2012. (2010).
69. C. B. Halpern, J. A. Lutz, Canopy closure exerts weak controls on understory dynamics: a 30-year study of overstory-understory interactions. Ecological Monographs 83, 221-237 (2013).
70. C. B. Halpern, J. A. Lutz, "Canopy closure exerts weak controls on understory dynamics: a 30 -year study of overstory-understory interactions". Available at: Dryad DigitalRepository, http://doi:10.5061/dryad.1q88j, accessed 2013 (2013).
71. A. J. Brooks, Moorea Coral Reef LTER: Coral Reef: Long-term Population and Community Dynamics: Fishes. Available at: knb-lter-mcr.6.54 http://doi:10.6073/pasta/d688610e536f54885a3c59d287f6c4c3, accessed 2016 (2016).
72. A. J. Brooks, "MCR LTER: Coral Reef: Long-term Population and Community Dynamics: Fishes". Moorea Coral Reef. Available at: http://mcr.lternet.edu/cgi-bin/showDataset.cgi?docid=knb-lter-mcr.6, accessed 2012.
73. M. Williamson, The land-bird community of Skokholm: ordination and turnover. Oikos, 378-384 (1983).
74. W. L. Vickery, T. D. Nudds, Detection of Density-Dependent Effects in Annual Duck Censuses. Ecology 65, 96-104 (1984).
75. NERC, "Fluctuations and long-term trends in the relative densities of tetraonid populations in Finland, 1964-77." NERC Centre for Population Biology, Imperial College. The Global Population Dynamics Database v2.0. . Available at: https://www.imperial.ac.uk/cpb/gpdd2/secure/register.aspx, accessed 2012.
76. H. Lindén, P. Rajala, Fluctuations and long-term trends in the relative densities of tetraonid populations in Finland, 1964-77. Finnish Game Research 39, 13-34 (1981).
77. NERC, "A transect survey of small land carnivore and red fox populations on a subarctic fell in Finnish Forest Lapland over 13 winters". NERC Centre for Population Biology, Imperial College, The Global Population Dynamics Database v2.0. Available at: http://www3.imperial.ac.uk/cpb/databases/gpdd, accessed 2012.
78. E. Pulliainen, A transect survey of small land carnivore and red fox populations on a subarctic fell in Finnish Forest Lapland over 13 winters. Annales Zoologici Fennici, 270-278 (1981).
79. P. Grant, An 11-year study of small mammal populations at Mont St. Hilaire, Quebec. Canadian Journal of Zoology 54, 2156-2173 (1976).
80. P. Grant, "An 11-year study of small mammal populations at Mont St. Hilaire, Quebec". NERC Centre for Population Biology, Imperial College. The Global Population Dynamics Database v2.0. Available at: http://www3.imperial.ac.uk/cpb/databases/gpdd, accessed 2012 (1976).
81. M. Friggens, "Sevilleta LTER Small Mammal Population Data", Albuquerque, NM: Sevilleta Long Term Ecological Research Site Database: SEV008. Available at: http://sev.lternet.edu/data/sev-8, accessed 2012 (2008).
82. R. B. Waide, Bird abundance - point counts. Long Term Ecological Research Network. Available at: http://dx.doi.org/10.6073/pasta/0d96957379936a038ebbbcc6135b2fab, accessed 2012. (2010).
83. R. B. Waide, "Bird abundance - point counts. El Verde Field Station, Puerto Rico: Luquillo Long Term Ecological Research Site Database: Data Set 23". Available at: http://luq. Iter net.edu/data/luqmetadata23, accessed 2012. (2010).
84. S. Ernest, T. J. Valone, J. H. Brown, Long-term monitoring and experimental manipulation of a Chihuahuan Desert ecosystem near Portal, Arizona, USA. Ecology 90, 1708-1708 (2009).
85. R. Condit, Tropical forest census plots: Methods and results from Barro Colorado Island, Panama and a Comparison with other plot. Springer Verlag and RG Landes Company, Berlin, (1998).
86. R. Condit et al., The importance of demographic niches to tree diversity. Science 313, 98-101 (2006).
87. R. Condit, R. A. Chisholm, S. P. Hubbell, Thirty years of forest census at Barro Colorado and the importance of immigration in maintaining diversity. PloS one 7, e49826 (2012).
88. R. Condit et al., Dataset: Barro Colorado Forest Census Plot Data (Version 2012). (2012).
89. R. Condit et al., Tree species abundance through time in tropical forest census plots, Panama. DataONE Dash, Dataset, Available at: https://doi.org/10.15146/R3MM4V, (2018).
90. S. P. Hubbell, R. Condit, R. B. Foster, "Barro Colorado Forest Census Plot Data ". Available at: https://ctfs.arnarb.harvard.edu/webatlas/datasets/bci, accessed 2012. (2005).
91. K. P. Robinson, "CRRU (Cetacean Research and Rescue Unit) Cetacean sightings in Scotland waters". Available at: http://www.emodnetbiology.eu/component/imis/?module=dataset\&dasid=2819, accessed 2012. (2010).
92. K. P. Robinson et al., The summer distribution and occurrence of cetaceans in the coastal waters of the outer southern Moray Firth in northeast Scotland (UK). Lutra 50, 19 (2007).
93. N. Derezuyk, "Phytoplankton of the Ukrainian Black Sea shelf (1985-2005)". Available at: http://www.emodnetbiology.eu/component/imis/?module=dataset\&dasid=2694, accessed 2012.
94. BSTS, "Baltic Seabirds Transect Surveys", Institute of Ecology of Vilnius University -OBIS-SEAMAP Available at: http://www.emodnetbiology.eu/component/imis/?module=dataset\&dasid=1971, accessed 2012.
95. P. A. Henderson, The long-term study of the fish and crustacean community of the Bristol Channel. Available at http://www.pisces-conservation.com/, accessed 2013.
96. P. A. Henderson, A. E. Magurran, Direct evidence that density-dependent regulation underpins the temporal stability of abundant species in a diverse animal community. Proceedings of the Royal Society B: Biological Sciences 281, (2014).
97. P. A. Henderson, A. E. Magurran. (Dryad Data Repository, 2014).
98. E. Woehler, "Seabirds of the Southern and South Indian Ocean - Australian Antarctic Data Centre". Available at: http://www.iobis.org, accessed 2012.
99. NIWA, "South Western Pacific Regional OBIS Data Asteroid Subset", NIWA (National Institute of Water and Atmospheric Research - New Zealand) MBIS (Marine Biodata Information System) accessed through South Western Pacific OBIS. Available at: http://www.iobis.org/mapper/?dataset=219, accessed 2012.
100. D. Clark, B. Branton, DFO Maritimes Research Vessel Trawl Surveys, OBIS Canada Digital Collections. Bedford Institute of Oceanography, Dartmouth, Nova Scotia, Canada, OBIS Canada, (2007).
101. CRED, "Towed-Diver Fish Biomass Surveys in the Pacific Ocean 2000-2010". Coral Reef Ecosystem Division (CRED), Pacific Island Fisheries Sciences Center, National Marine Fisheries Service. Available at: http://www.iobis.org/mapper/?dataset=1581, accessed 2012 (2011).
102. S. Sherman, "Maine Department of Marine Resources Inshore Trawl Survey, 20002009". Maine Department of Marine Resources, Maine. Available at:
http://www.usgs.gov/obis-usa/data search_and access/datasets.html, accessed 2012. (2010).
103. M. Reichert, "MARMAP Chevron Trap Survey 1990-2009". SCDNR/NOAA MARMAP Program, SCDNR MARMAP Aggregate Data Surveys, The Marine Resources Monitoring, Assessment, and Prediction (MARMAP) Program, Marine Resources Research Institute, South Carolina Department of Natural Resources U.S.A. Available at: http://www.usgs.gov/obis-usa/data search_and_access/participants.html, accessed 2012. (2009).
104. M. Reichert, "MARMAP Neuston Nets 1990-2009". SCDNR/NOAA MARMAP Program, SCDNR MARMAP Aggregate data surveys, The Marine Resources Monitoring, Assessment, and Prediction (MARMAP) Program, Marine Resources Research Institute, South Carolina Department of Natural Resources U.S.A. Available at: http://www.usgs.gov/obis-usa/data search_and_access/participants.html, accessed 2012. (2010).
105. M. Reichert, "MARMAP Florida Antillean Trap Survey 1990-2009". SCDNR/NOAA MARMAP Program, SCDNR MARMAP Aggregate Data Surveys, The Marine Resources Monitoring, Assessment, and Prediction (MARMAP) Program, Marine Resources Research Institute, South Carolina Department of Natural Resources U.S.A. Available at: http://www.usgs.gov/obis-usa/data search_and_access/participants.html, accessed 2012. (2009).
106. NPRB, "The Observer Program database", accessed through the OBIS-USA North Pacific Groundfish Observer (North Pacific Research Board). Available at: http://www.iobis.org, accessed 2012. .
107. A. Diamond, A. Gaston, R. Brown, Converting PIROP Counts of Seabirds at Sea to Absolute Densities. Progress Notes No 164. Canadian Wildlife Service, Ottawa, (1986).
108. F. Huettmann, An ecological GIS research application for the northern Atlantic-The PIROP database software, environmental data sets and the role of the internet. Riekert/Tochtermann, (1998).
109. PIROP, "PIROP Northwest Atlantic 1965-1992 - OBIS SEAMAP". Available at: http://www.iobis.org/mapper/?dataset=2245, accessed 2012.
110. R. G. Brown, Atlas of eastern Canadian seabirds. (1975).
111. P. N. Halpin et al., OBIS-SEAMAP: The world data center for marine mammal, sea bird, and sea turtle distributions. Oceanography 22, 104-115 (2009).
112. A. Read, P. Halpin, L. Crowder, B. Best, E. Fujioka, OBIS-SEAMAP: mapping marine mammals, birds and turtles. World Wide Web electronic publication. http://seamap. env. duke. edu. Accessed 15, (2009).
113. J. R. Jahncke, C., "CalCOFI and NMFS Seabird and Marine Mammal Observation Data, 1987-2006". California Cooperative Oceanic Fisheries Investigations (CalCOFI) and National Marine Fisheries Service (NMFS) cruises, 1987-2006 - OBIS SEAMAP. Available at: http://www.iobis.org, accessed 2012. (2006).
114. C. Rintoul, B. Langabeer-Schlagenhauf, K. Hyrenbach, K. Morgan, W. Sydeman, Atlas of California Current marine birds and mammals: Version 1. Unpublished Report, PRBO Conservation Science, Petaluma, CA, (2006).
115. P. Yen, W. Sydeman, S. Bograd, K. Hyrenbach, Spring-time distributions of migratory marine birds in the southern California Current: Oceanic eddy associations and coastal habitat hotspots over 17 years. Deep Sea Research Part II: Topical Studies in Oceanography 53, 399-418 (2006).
116. P. P. Yen, W. J. Sydeman, K. D. Hyrenbach, Marine bird and cetacean associations with bathymetric habitats and shallow-water topographies: implications for trophic transfer and conservation. Journal of Marine Systems 50, 79-99 (2004).
117. BMMROOS, "Bahamas Marine Mammal Research Organisation Opportunistic Sightings - OBIS SEAMAP". Available at: http://www.iobis.org, accessed 2012.
118. POPA, "cetacean, seabird, and sea turtle sightings in the Azores area 1998-2009 OBIS SEAMAP". . Available at: http://www.iobis.org/mapper/?dataset=4257, accessed 2012.
119. P. Amorim et al., Spatial variability of seabird distribution associated with environmental factors: a case study of marine Important Bird Areas in the Azores. ICES Journal of Marine Science 66, 29-40 (2008).
120. M. Machete, R. Santos, in Proceedings of the 5th International Fisheries Observer Conference. (2007), pp. 15-18.
121. T. Morato et al., Evidence of a seamount effect on aggregating visitors. Marine Ecology Progress Series 357, 23-32 (2008).
122. M. Kennedy, J. Spry, Atlantic Zone Monitoring Program Maritimes Region plankton datasets. Fisheries and Oceans Canada-BioChem archive. OBIS Canada, Bedford Institute of Oceanography, Dartmouth, Nova Scotia, Canada, (2011).
123. ECNA, "East Coast North America Strategic Assessment Project, Groundfish Atlas for the East Coast of North America". Available at: http://www.iobis.org, accessed 2012.
124. E. Wade, Snow crab research trawl survey database (Southern Gulf of St. Lawrence, Gulf region, Canada) from 1988 to 2010. OBIS Canada, Bedford Institute of Oceanography, Dartmouth, Nova Scotia, Canada, (2011).
125. USVI, "St. John, USVI Fish Assessment and Monitoring Data (2002 - Present)", Silver Spring, MD Publisher: NOAAs Ocean Service, National Centers for Coastal Ocean Science (NCCOS). National Oceanic and Atmospheric Association (NOAA)National Ocean Service (NOS)-National Centers for Coastal Ocean Science (NCCOS)-Center for Coastal Monitoring and Assessment (CCMA)-Biogeography Team. Available at: http://www.iobis.org/mapper/?dataset=1672, accessed 2012 (2007).
126. USVI, "St. Croix, USVI Fish Assessment and Monitoring Data (2002 - Present)", Silver Spring, MD Publisher: NOAAs Ocean Service, National Centers for Coastal Ocean Science (NCCOS). National Oceanic and Atmospheric Association (NOAA)National Ocean Service (NOS)-National Centers for Coastal Ocean Science (NCCOS)-Center for Coastal Monitoring and Assessment (CCMA)-Biogeography Team. Available at: http://www.iobis.org/mapper/?dataset=167,3 accessed 2012 (2007).
127. AADC, "Whale Catches in Southern Ocean". OBIS - Australian Antarctic Data Centre. Available at: http://www.iobis.org, accessed 2013.
128. USGS, Patuxent Wildlife Research Center "North American Breeding Bird Survey" ftp data set, version 2014.0. Available at:
ftp://ftpext.usgs.gov/pub/er/md/laurel/BBS/DataFiles/, accessed 2013.
129. J. J. Moore, C. M. Howson, "Survey of the rocky shores in the region of Sullom Voe, Shetland, A report to SOTEAG from Aquatic Survey \& Monitoring Ltd", Cosheston, Pembrokeshire. 29 p. Available at: http://www.soteag.org.uk, accessed 2013.
130. DATRAS, ICES Scottish West Coast Bottom Trawl Survey (SWC-IBTS) 1985-2014. Available at https://datras.ices.dk, accessed 2015. (2015).
131. DATRAS, ICES Baltic International Trawl Survey For Commercial Fish Species (1991-2013). Available at https://datras.ices.dk, accessed 2013 (2013).
132. DATRAS, "Fish trawl survey: Scottish Rockall Survey for commercial fish species. ICES Database of trawl surveys (DATRAS)." The International Council for the Exploration of the Sea, Copenhagen. Available at: http://www.emodnet-biology.eu/data-catalog?\%3Fmodule=dataset\&dasid=2767, accessed 2013. (2010).
133. DATRAS, "Fish trawl survey: Northern Irish Ground Fish Trawl Survey. ICES Database of trawl surveys (DATRAS)." The International Council for the Exploration of the Sea, Copenhagen. Available at: http://www.emodnet-biology.eu/datacatalog?\%3Fmodule=dataset\&dasid=2764, accessed 2013. (2010).
134. DATRAS, "Fish trawl survey: Irish Ground Fish Survey for commercial fish species. ICES Database of trawl surveys (DATRAS)." The International Council for the Exploration of the Sea, Copenhagen. Available at: http://www.emodnet-biology.eu/data-catalog?\%3Fmodule=dataset\&dasid=2762, accessed 2013. (2010).
135. DATRAS, "Fish trawl survey: ICES French Southern Atlantic Bottom Trawl Survey for commercial fish species. ICES Database of trawl surveys (DATRAS)." The International Council for the Exploration of the Sea, Copenhagen. Available at: http://www.emodnet-biology.eu/data-catalog?\%3Fmodule=dataset\&dasid=2759, accessed 2013. (2010).
136. DATRAS, "Fish trawl survey: ICES Beam Trawl Survey for commercial fish species. ICES Database of trawl surveys (DATRAS)." The International Council for the Exploration of the Sea, Copenhagen. Available at: http://www.emodnet-biology.eu/data-catalog?\%3Fmodule=dataset\&dasid=2761, accessed 2013. (2010).
137. DATRAS, "Fish trawl survey: ICES North Sea International Bottom Trawl Survey for commercial fish species. ICES Database of trawl surveys (DATRAS)." The International Council for the Exploration of the Sea, Copenhagen. Available at: http://www.emodnet-biology.eu/data-catalog?\%3Fmodule=dataset\&dasid=2763, accessed 2013. (2010).
138. M. Reichert, "MARMAP Fly Net 1990-2009". SCDNR/NOAA MARMAP Program, SCDNR MARMAP Aggregate Data Surveys, The Marine Resources Monitoring, Assessment, and Prediction (MARMAP) Program, Marine Resources Research Institute, South Carolina Department of Natural Resources USA. Available at: http://www.usgs.gov/obis-usa/, accessed 2013. (2010).
139. M. Reichert, "MARMAP Yankee Trawl 1990-2009". SCDNR/NOAA MARMAP Program, SCDNR MARMAP Aggregate data surveys, The Marine Resources Monitoring, Assessment, and Prediction (MARMAP) Program, Marine Resources Research Institute, South Carolina Department of Natural Resources USA. Available at: http://www.usgs.gov/obis-usa, accessed 2013. (2010).
140. NMFS, "Northeast Fisheries Science Center Bottom Trawl Survey Data (OBISUSA)." NOAA's National Marine Fisheries Service (NMFS) Northeast Fisheries Science Center. Woods Hole, Massachusetts, USA. Available at: http://www.iobis.org/mapper/?dataset=1435, accessed 2013. (2005).
141. M. F. Harmon, J., "Long-term growth, mortality and regeneration of trees in permanent vegetation plots in the Pacific Northwest, 1910 to present." Long-Term Ecological Research. Forest Science Data Bank, Corvallis. Available at:
http://andrewsforest.oregonstate.edu/data/abstract.cfm?dbcode=TV010, accessed 2012. (2012).
142. HMANA, "Hawk Migration Association of North America (HMANA)". Available at: http://www.hmana.org/, accessed 2012.
143. NatureCounts, "Ontario Breeding Bird Atlas (2001-2005): point count data." NatureCounts, a node of the Avian Knowledge Network. Bird Studies Canada. Available at: http://www.birdscanada.org/birdmon/, accessed 2012.
144. USFS, "Landbird Monitoring Program (UMT-LBMP)." US Forest Service. Available at: http://www.avianknowledge.net/, accessed 2012.
145. NatureCounts, "Maritimes Breeding Bird Atlas (2006-2010): point count data." NatureCounts, a node of the Avian Knowledge Network. Bird Studies Canada. Available at: http://www.birdscanada.org/birdmon/, accessed 2012.
146. NatureCounts, Bird Studies Canada "Marsh Monitoring Program." NatureCounts, a node of the Avian Knowledge Network. Available at: http://www.birdscanada.org/birdmon,/ accessed 2012. (2012).
147. W. O. McLarney, J. Meador, J. Chamblee, "Upper Little Tennessee River Biomonitoring Program Database." Coweeta Long Term Ecological Research Program. Available at: https://coweeta.uga.edu/dbpublic/dataset details.asp?accession=4045, accessed 2012. (2010).
148. J. Trexler, "Consumer Stocks: Fish, Vegetation, and other Non-physical Data from Everglades National Park (FCE), South Florida from February 2000 to Present." Florida Coastal Everglades LTER Program. Available at: http://fcelter.fiu.edu/data/core/metadata/EML/?datasetid=LT CD Trexler 001, accessed 2012. (2007).
149. D. Williams, "Pelagic Fish Observations 1968-1999." Australian Antarctic Data Centre. Available at: http://www.gbif.org/dataset/85b0a82a-f762-11e1-a43900145 eb45e9a, accessed 2012.
150. J. J. Battles, Johnson, C., Hamburg, S., Fahey, T., Driscoll, C. \& Likens, G., "Forest Inventory of a Northern Hardwood Forest: Watershed 6 2002." The Hubbard Brook Ecosystem Study LTER Program. Available at: http://www.hubbardbrook.org/data/dataset.php?id=35, accessed 2012. (2003).
151. J. J. Battles, Fahey, T. \& Cleavitt, N., "Forest Inventory of a Northern Hardwood Forest: Watershed 6 1965, Hubbard Brook Experimental Forest." The Hubbard Brook Ecosystem Study LTER Program. Available at: http://www.hubbardbrook.org/data/dataset.php?id=29, accessed 2016.
152. J. J. Battles, Fahey, T. \& Cleavitt, N., "Forest Inventory of a Northern Hardwood Forest: Watershed 6 1977, Hubbard Brook Experimental Forest." The Hubbard Brook Ecosystem Study LTER Program. Available at: http://www.hubbardbrook.org/data/dataset.php?id=30, accessed 2016.
153. J. J. Battles, Fahey, T. \& Cleavitt, N., "Forest Inventory of a Northern Hardwood Forest: Watershed 6 1987, Hubbard Brook Experimental Forest." The Hubbard Brook Ecosystem Study LTER Program. Available at: http://www.hubbardbrook.org/data/dataset.php?id=32, accessed 2016.
154. J. J. Battles, Fahey, T. \& Cleavitt, N., "Forest Inventory of a Northern Hardwood Forest: Watershed 6 1992, Hubbard Brook Experimental Forest." The Hubbard Brook Ecosystem Study LTER Program. Available at: http://www.hubbardbrook.org/data/dataset.php?id=33, accessed 2016.
155. J. J. Battles, Fahey, T. \& Cleavitt, N., "Forest Inventory of a Northern Hardwood Forest: Watershed 6 1997, Hubbard Brook Experimental Forest." The Hubbard Brook

Ecosystem Study LTER Program. Available at: http://www.hubbardbrook.org/data/dataset.php?id=34, accessed 2016.
156. J. J. Battles, Fahey, T. , N. Cleavitt, "Forest Inventory of a Northern Hardwood Forest: Watershed 6 1982, Hubbard Brook Experimental Forest." The Hubbard Brook Ecosystem Study LTER Program. Available at: http://www.hubbardbrook.org/data/dataset.php?id=31, accessed 2016.
157. K. B. Gido, "Fish population on selected watersheds at Konza Prairie - CFP01." Konza Prairie LTER Program. Available at: http://www.konza.ksu.edu/KNZ/pages/data/Knzdsdetail.aspx?datasetCode=CFP01, accessed 2012.
158. E. Muldavin, "Pinon-Juniper (Core Site) Quadrat Data for the Net Primary Production Study at the Sevilleta National Wildlife Refuge, New Mexico (2003-Present)." Sevilleta Long Term Ecological Research Program. Available at: http://sev.lternet.edu/node/1718, accessed 2013.
159. A. Paquette et al., Lac Croche understory vegetation data set (1998-2006). Ecology 88, 3209-3209 (2007).
160. F. Day, "Long-term N-fertilized vegetation plots on Hog Island, Virginia Coastal Barrier Islands, 1992-2014." Virginia Coast Reserve Long-Term Ecological Research Project. Available at: http://www.vcrlter.virginia.edu/cgi-bin/showDataset.cgi?docid=knb-lter-vcr.106, accessed 2013. (2010).
161. F. P. Day, C. Conn, E. Crawford, M. Stevenson, Long-term effects of nitrogen fertilization on plant community structure on a coastal barrier island dune chronosequence. Journal of Coastal Research, 722-730 (2004).
162. NatureCounts, Bird Studies Canada "BC Coastal Waterbird Survey (2004)." NatureCounts, a node of the Avian Knowledge Network. Available at: http://www.birdscanada.org/birdmon/, accessed 2012. (2012).
163. H. Chen et al., Long-term monitoring dataset of fish assemblages impinged at nuclear power plants in northern Taiwan. Scientific data 2, 150071 (2015).
164. L. Rudstam, "Zooplankton survey of Oneida Lake, New York, 1964 - 2012", KNB Data Repository. Available at: https://knb.ecoinformatics.org/\#view/kgordon.17.56, accessed 2016. (2008).
165. Z. Shi et al., Evidence for long-term shift in plant community composition under decadal experimental warming. Journal of Ecology 103, 1131-1140 (2015).
166. P. F. Thomsen et al., Resource specialists lead local insect community turnover associated with temperature - analysis of an 18-year full-seasonal record of moths and beetles. Journal of Animal Ecology 85, 251-261 (2016).
167. M. Reichert, "MARMAP Blackfish Trap Survey 1990-2009". SCDNR/NOAA MARMAP Program. SCDNR MARMAP Aggregate Data Surveys. The Marine Resources Monitoring. Assessment. and Prediction (MARMAP) Program. Marine Resources Research Institute. South Carolina Department of Natural Resources USA. Available at: http://www.usgs.gov/obis-usa/, accessed 2013. (2010).
168. N. LTER, "North Temperate Lakes LTER: Phytoplankton - Madison Lakes Area 1995 - current." North Temperate Lakes Long Term Ecological Research Program, Center for Limnology, University of Wisconsin-Madison. Available at: https://Iter.limnology.wisc.edu/dataset/north-temperate-lakes-lter-phytoplankton-madison-lakes-area-1995-current, accessed 2013.
169. K. D. Woods, Multi-decade, spatially explicit population studies of canopy dynamics in Michigan old-growth forests. Ecology 90, 3587-3587 (2009).
170. D. C. Reed, "SBC LTER: Reef: Kelp forest community dynamics: Abundance and size of giant kelp (Macrocystis pyrifera), ongoing since 2000". Santa Barbara Coastal

LTER. Available at: http://sbc.lternet.edu/cgi-bin/showDataset.cgi?docid=knb-ltersbc. 18 doi:10.6073/pasta/d90872297e30026b263a119d4f5bca9f, accessed 2016 (2014a).
171. R. A. Davis, T. S. Doherty, Rapid recovery of an urban remnant reptile community following summer wildfire. PloS one 10, e0127925 (2015).
172. C. B. Halpern, J. A. Lutz, "Canopy closure exerts weak controls on understory dynamics: a 30 -year study of overstory-understory interactions.". Available at: Dryad DigitalRepository http://doi:10.5061/dryad.1q88j, accessed 2013. (2013).
173. G. J. Edgar, R. D. Stuart-Smith, Systematic global assessment of reef fish communities by the Reef Life Survey program. Scientific Data 1, 140007 (2014).
174. R. H. Wiley, "Population estimates of Appalachian salamanders". Coweeta LTER. Available at: http://coweeta.uga.edu/eml/1044.xml, accessed 2016.
175. J. Merritt, Long Term Mammal Data from Powdermill Biological Station 1979-1999. Environmental Data Initiative. Available at: http://dx.doi.org/10.6073/pasta/83c888854e239a79597999895bb61cfe, accessed 2016 (1999).
176. D. W. Kaufman, Seasonal summary of numbers of small mammals on 14 LTER traplines in prairie habitats at Konza Prairie. Konza Prairie Long-Term Ecological Research. . Available at: http://lter.konza.ksu.edu/content/csm01-seasonal-summary-numbers-small-mammals-14-lter-traplines-prairie-habitats-konza, accessed 2016.
177. H. H. T. Prins, I. Douglas-Hamilton, Stability in a multi-species assemblage of large herbivores in East Africa. Oecologia 83, 392-400 (1990).
178. D. Lightfoot, "Lizard pitfall trap data (LTER-II, LTER-III)". Jornada Basin LTER. Available at: http://jornada.nmsu.edu/lter/dataset/49821/view, accessed 2016. (2013).
179. R. Twilley, V. H. Rivera-Monroy, E. Castaneda, "Mangrove Forest Growth from the Shark River Slough, Everglades National Park (FCE), South Florida from January 1995 to Present". Florida Coastal Everglades LTER. http://dx.doi.org/10.6073/pasta/bec6c029df692768f349106c69162df7. Available at: http://fcelter.fiu.edu/data/core/metadata/?datasetid=LT PP Rivera 002, accessed 2016. (2005).
180. SANParks, "Karoo National Park Census Data. 1994-2009". Available at: http://datadryad.org/handle/10255/dryad.13079?show=full, accessed 2016. (2011).
181. D. J. Wilgers, E. A. Horne, B. K. Sandercock, A. W. Volkmann, Effects of rangeland management on community dynamics of the herpetofauna of the tallgrass prairie. Herpetologica 62, 378-388 (2006).
182. D. Lightfoot, R. L. Schooley, "SMES rodent trapping data, Small Mammal Exclosure Study". Jornada LTER. Available at: http://jornada.nmsu.edu/sites/jornada.nmsu.edu/files/data_files/JornadaStudy_086_sm es rodent trapping data 0.csv, accessed 2016.
183. F. Venturoli, J. M. Felfili, C. W. Fagg, Temporal evaluation of natural regeneration in a semideciduous secondary forest in Pirenópolis, Goiás, Brazil. Revista Arvore 35, 473-483 (2011).
184. D. Kelt, P. Meserve, J. Gutiérrez, W. B. Milstead, M. Previtali, Long-term monitoring of mammals in the face of biotic and abiotic influences at a semiarid site in northcentral Chile. Ecology 94, 977-977 (2013).
185. D. Scott, B. Metts, S. Lance, "The Rainbow Bay Long-term Study". Available at: http://srelherp.uga.edu/projects/rbay.htm, accessed 2016.
186. G. D. Grossman, "Stream fish assemblage stability in a southern Appalachian stream at the Coweeta Hydrologic Laboratory from 1984 to 1995". Coweeta Long Term Ecological Research Program. Available at:
http://dx.doi.org/10.6073/pasta/f0baf5f59c89f670e04f537f5cc05290, accessed 2016. (2007).
187. G. D. Grossman, P. B. Moyle, J. O. Whitaker Jr, Stochasticity in structural and functional characteristics of an Indiana stream fish assemblage: a test of community theory. American Naturalist, 423-454 (1982).
188. S. Svensson, A. Thorner, N. Nyholm, Species trends, turnover and composition of a woodland bird community in southern Sweden during a period of fifty-seven years. Ornis Svecica 20, 31-44 (2010).
189. F. Carvalho, J. J. Zocche, R. Á. Mendonça, Morcegos (Mammalia, Chiroptera) em restinga no município de Jaguaruna, sul de Santa Catarina, Brasil. Biotemas 22, 193201 (2009).
190. D. Billett, B. Bett, W. Reid, B. Boorman, I. Priede, Long-term change in the abyssal NE Atlantic: The 'Amperima Event'revisited. Deep Sea Research Part II: Topical Studies in Oceanography 57, 1406-1417 (2010).
191. D. Billett et al., Long-term change in the megabenthos of the Porcupine Abyssal Plain (NE Atlantic). Progress in Oceanography 50, 325-348 (2001).
192. L. A. Kuhnz, H. A. Ruhl, C. L. Huffard, K. L. Smith, Rapid changes and long-term cycles in the benthic megafaunal community observed over 24years in the abyssal northeast Pacific. Progress in Oceanography 124, 1-11 (2014).
193. H. A. Ruhl, K. L. Smith, Shifts in deep-sea community structure linked to climate and food supply. Science 305, 513-515 (2004).
194. M. G. Bradford, H. T. Murphy, A. J. Ford, D. L. Hogan, D. J. Metcalfe, Long-term stem inventory data from tropical rain forest plots in Australia. Ecology 95, 2362-2362 (2014).
195. P. Stapp, SGS-LTER Long-Term Monitoring Project: Small Mammals on Trapping Webs on the Central Plains Experimental Range, Nunn, Colorado, USA 1994-2006, ARS Study Number 118. Environmental Data Initiative. Available at: http://dx.doi.org/10.6073/pasta/2e311b4e40fea38e573890f473807ba9, accessed 2017 (2013).
196. J. G. Dickson, R. N. Conner, J. H. Williamson, Neotropical migratory bird communities in a developing pine plantation. Procedings on the Annual Conference. SEAFWA 47, 439-446 (1993).
197. D. C. Reed, "SBC LTER: Reef: Kelp Forest Community Dynamics: Fish abundance". Santa Barbara Coastal LTER. Available at: http://doi:10.6073/pasta/e37ed29111b2fddffc08355252b8b8c7, accessed 2016 (2014).
198. G. A. Hall, A long-term bird population study in an Appalachian spruce forest. The Wilson Bulletin, 228-240 (1984).
199. A. Enemar, B. Sjöstrand, G. Andersson, T. von Proschwitz, The 37-year dynamics of a subalpine passerine bird community, with special emphasis on the influence of environmental temperature and Epirrita autumnata cycles. Ornis Svecica 14, 63-106 (2004).
200. T. J. Willis, "Hahei marine dataset (1997-2002), New Zealand fish". Institute of Marine Sciences, University of Portsmouth. Accessed 2016.
201. D. Lightfoot, "Small Mammal Exclosure Study (SMES)". Sevilleta Long Term Ecological Research Program. Available at: http://sev.lternet.edu/content/small-mammal-exclosure-study-smes-0, accessed 2016.
202. Monitoring Site 1000 Project, Biodiversity Center, M. o. E. o. Japan, "Monitoring site 1000 Village survey - Bird survey data (2005-2012)". (SAT02.zip, downloaded from http://www.biodic.go.jp/moni1000/findings/data/index.html), accessed 2016. (2014).
203. Monitoring Site 1000 Project, Biodiversity Center, M. o. E. o. Japan, "Monitoring site 1000 Village survey - Medium and large mammal survey data (2006-2012)".
(SAT03zip, downloaded from
http://www.biodic.go.jp/moni1000/findings/data/index.html), accessed 2016. (2014).
204. Monitoring Site 1000 Project, Biodiversity Center, M. o. E. o. Japan, "Monitoring site 1000 Shorebird Survey" (ShorebirdsDatapackage2012.zip, downloaded from http://www.biodic.go.jp/moni1000/ findings/data/index.html), accessed 2016. (2013).
205. Monitoring Site 1000 Project, Biodiversity Center, M. o. E. o. Japan, "Monitoring site 1000 Forest and grassland research - Surface wandering beetles survey data". (GBDataPackage2014ver1.zip, downloaded from http://www.biodic.go.jp/moni1000/findings/data/index.html), accessed 2016. (2014).
206. R. A. How, Long-term sampling of a herpetofaunal assemblage on an isolated urban bushland remnant, Bold Park, Perth. Journal of the Royal Society of Western Australia 81, 143-148 (1998).
207. L. W. Krefting, C. E. Ahlgren, Small Mammals and Vegetation Changes After Fire in a Mixed Conifer-Hardwood Forest. Ecology 55, 1391-1398 (1974).
208. NERC, "The Global Population Dynamics Database Version 2". Centre for Population Biology, Imperial College. Available at: http://www.sw.ic.ac.uk/cpb/cpb/gpdd.html, accessed 2016. (2010).
209. Y. R. Suarez, "Brazil Dataset 6", Mato Gosso do Sul State University. Accessed 2016.
210. D. C. Rossa-Feres, Community ecology of anura amphibia at Northwest region of Sao Paulo state, Brazil: microhabitat, seasonality, diet and multidimensional niche. State University of São Paulo, PhD thesis. Accessed 2016. (1997).
211. C.-H. Hsieh, "Icthyoplankton data collected from Yenliao Bay in 6 stations northeast of Taiwan (1995-2000)". Ecoinformatics Lab, Institute of Oceanography National Taiwan University. Accessed 2016.
212. S. Svensson, Species composition and population fluctuations of alpine bird communities during 38 years in the Scandinavian mountain range. Ornis Svecica 16, 183-210 (2006).
213. F. Pomati, B. Matthews, J. Jokela, A. Schildknecht, B. W. Ibelings, Effects of reoligotrophication and climate warming on plankton richness and community stability in a deep mesotrophic lake. Oikos 121, 1317-1327 (2012).
214. F. Pomati et al., Challenges and prospects for interpreting long-term phytoplankton diversity changes in Lake Zurich (Switzerland). Freshwater Biology 60, 1052-1059 (2015).
215. E. M. Olsen, S. M. Carlson, J. Gjøsæter, N. C. Stenseth, Nine decades of decreasing phenotypic variability in Atlantic cod. Ecology Letters 12, 622-631 (2009).
216. L. A. Rogers et al., Climate and population density drive changes in cod body size throughout a century on the Norwegian coast. Proceedings of the National Academy of Sciences 108, 1961-1966 (2011).
217. N. C. Stenseth et al., Dynamics of coastal cod populations: intra- and intercohort density dependence and stochastic processes. Proceedings of the Royal Society of London. Series B: Biological Sciences 266, 1645-1654 (1999).
218. C. Barceló, L. Ciannelli, E. M. Olsen, T. Johannessen, H. Knutsen, Eight decades of sampling reveal a contemporary novel fish assemblage in coastal nursery habitats. Global Change Biology 22, 1155-1167 (2016).
219. NIWA, "The New Zealand Freshwater Fish Database". Available at: https://www.niwa.co.nz/our-services/online-services/freshwater-fish-database, accessed 2016.
220. D. Steinberg, Zooplankton collected with a $2-\mathrm{m}, 700$-um net towed from surface to 120 m , aboard Palmer Station Antarctica LTER annual cruises off the western antarctic peninsula, 2009-2016. Environmental Data Initiative. Available at: http://dx.doi.org/10.6073/pasta/fb658789188724be5f27c81a634647d5, accessed 2017
221. A. Hoey, "Karimunjawa WCS fish data". Accessed 2016.
222. A. Hoey, "Aceh WCS fish surveys". Accessed 2016.
223. V. D. Zakharov, Biodiversity of bird population of terrestrial habitats in Southern Ural. Miass: IGZ. Ural Branch of Russian Academy of Sciences, 158 p (1998).
224. N. N. Berezovikov, The birds of settlements in Markakol Depression (Southern Altai). Russian Ornithological Journal 249, 3-15 (2004).
225. Y. I. Melnikov, Melnikova, N. \& Pronkevich, V. V., Migration of birds of prey in the mouth of the river Irkut. Russian Ornithological Journal 108, 3-17. (2000).
226. A. B. Jalilov, Andreychev, A. V. \& Kuznetsov, V. A., Monitoring and conservation of medium and large mammals in Chamzinsky District of the Republic of Mordovia.Vestnik of Lobachevsky University of Nizhni. Novgorod 4 (1), 222-227. (2014).
227. V. Y. Nedosekin, Long-term dynamics of the population and the quantity of small mammals under conditions of the reserve "Galichya Gora". Proceedings of National Nature Reserve Prisursky 30, 87-90 (2015).
228. B. I. Sheftel et al., Population dynamics of small mammals at Western Khentey during ten years. Proceedings of the international conference Ecological consequences of biosphere processes in the ecotone zone of southern Siberia vol. I, Oral reports, 230233. (2010).
229. S. Thorn et al., Changes in the dominant assembly mechanism drive species loss caused by declining resources. Ecology letters 19, 163-170 (2016).
230. S. Thorn et al., New insights into the consequences of post-windthrow salvage logging revealed by functional structure of saproxylic beetles assemblages. PloS one 9 , e101757 (2014).
231. S. Thorn et al., Response of bird assemblages to windstorm and salvage loggingInsights from analyses of functional guild and indicator species. Ecological Indicators 65, 142-148 (2016).
232. F. Neat, N. Campbell, Demersal fish diversity of the isolated Rockall plateau compared with the adjacent west coast shelf of Scotland. Biological Journal of the Linnean Society 104, 138-147 (2011).
233. D. J. Kushner, A. Rassweiler, J. P. McLaughlin, K. D. Lafferty, A multi-decade time series of kelp forest community structure at the California Channel Islands. Ecology 94, 2655-2655 (2013).
234. E. Muldavin, S. L. Collins, Prescribed Burn Effect on Chihuahuan Desert Grasses and Shrubs at the Sevilleta National Wildlife Refuge, New Mexico: Species Composition Study 2004 to present. Sevilleta LTER. Available at: http://sev.lternet.edu/data/sev166, accessed 2016. (2003).
235. O. Hogstad, in Annales Zoologici Fennici. (JSTOR, 1993), pp. 43-54.
236. J. G. Douglass, K. E. France, J. P. Richardson, J. E. Duffy, Seasonal and interannual change in a Chesapeake Bay eelgrass community: Insights into biotic and abiotic control of community structure. Limnology and Oceanography 55, 1499-1520 (2010).
237. J. S. Lefcheck, The use of functional traits to elucidate the causes and consequences of biological diversity. The College of William \& Mary, PhD thesis. Available at: http://gradworks.umi.com/36/62/3662989.html, accessed 2016. (2015).
238. B. Institute of Agricultural and Fisheries research (ILVO), Epibenthos and demersal fish monitoring at long-term monitoring stations in the Belgian part of the North Sea. Available at: http://dx.doi.org/10.14284/54, accessed 2016. (2015).
239. K. D. Woods, Multi-decade biomass dynamics in an old-growth hemlock-northern hardwood forest, Michigan, USA. PeerJ 2, e598 (2014).
240. J. Belmaker, Y. Ziv, N. Shashar, The influence of connectivity on richness and temporal variation of reef fishes. Landscape ecology 26, 587-597 (2011).
241. D. Edelist, G. Rilov, D. Golani, J. T. Carlton, E. Spanier, Restructuring the Sea: profound shifts in the world's most invaded marine ecosystem. Diversity and Distributions 19, 69-77 (2013).
242. G. B. G. Souza, M. Vianna, "Demersal fish hauls from Guanabara Bay, Brazil 20052015". Accessed 2017.
243. E. M. Sampaio, E. K. Kalko, E. Bernard, B. Rodríguez-Herrera, C. O. Handley, A biodiversity assessment of bats (Chiroptera) in a tropical lowland rainforest of Central Amazonia, including methodological and conservation considerations. Studies on Neotropical fauna and environment 38, 17-31 (2003).
244. F. Z. Farneda et al., Functional recovery of Amazonian bat assemblages following secondary forest succession. Biological Conservation 218, 192-199 (2018).
245. R. Rocha, University of Lisbon, Lisbon, Portugal., (2017).
246. R. Rocha et al., Consequences of a large-scale fragmentation experiment for Neotropical bats: disentangling the relative importance of local and landscape-scale effects. Landscape Ecology 32, 31-45 (2017).
247. R. Rocha et al., Secondary forest regeneration benefits old-growth specialist bats in a fragmented tropical landscape. Scientific Reports 8, 3819 (2018).
248. M. Valeix, H. Fritz, S. Chamaillé-Jammes, M. Bourgarel, F. Murindagomo, Fluctuations in abundance of large herbivore populations: insights into the influence of dry season rainfall and elephant numbers from long-term data. Animal Conservation 11, 391-400 (2008).
249. J. Mundava et al., Factors influencing long-term and seasonal waterbird abundance and composition at two adjacent lakes in Zimbabwe. Ostrich 83, 69-77 (2012).
250. E. Haplet, SANParks, "Monthly bird lists and bird arrival dates, Birmingham Timbivati". Available at: http://dataknp.sanparks.org/sanparks/, accessed 2018. (2009).
251. SANParks, "Northern Plains Ecological Aerial Census data 1993-1998". Available at: http://dataknp.sanparks.org/sanparks/, accessed 2018. (2009).
252. J. W. F. Chu, V. Tunnicliffe, Oxygen limitations on marine animal distributions and the collapse of epibenthic community structure during shoaling hypoxia. Global Change Biology 21, 2989-3004 (2015).
253. C. Gjerdrum, CWS-EC Eastern Canada Seabirds at Sea (ECSAS). OBIS Canada. Available at: https://obis.org/dataset/51391fb1-ae4d-44d8-9178-a06f95545604, accessed 2019.
254. A. E. Magurran et al., Divergent biodiversity change within ecosystems. Proceedings of the National Academy of Sciences USA, (2018).
255. A. E. Magurran et al. (University of St Andrews, 2018).
256. S. Matsuzaki, Fish monitoring data in Lake Kasumigaura. Version 13.11. National Institute of Genetics, ROIS. Occurrence dataset https://doi.org/10.15468/nhsh6m, accessed via GBIF.org on 2021-07-06., (2021).
257. Y. Umatani et al., Long-term observation of fish community in streams inside and outside road construction of Teshio River system, northern Hokkaido, Japan. Ecological Research 35, 986-993 (2020).
258. R. van Klink et al., InsectChange: a global database of temporal changes in insect and arachnid assemblages. Ecology 102, e03354 (2021).
259. R. van Klink et al., Meta-analysis reveals declines in terrestrial but increases in freshwater insect abundances. Science 368, 417-420 (2020).
260. C. H. Davies et al., A database of marine phytoplankton abundance, biomass and species composition in Australian waters. Scientific data 3, (2016).

## Acknowledgments:

## Funding:

Marie Sklodowska-Curie Actions Individual Fellowship (MSCA-IF), European Union's Horizon 2020, grant agreement no. 894644. (I.S.M.)

German Research Foundation grant to the German Centre for Integrative Biodiversity Research (iDiv) Halle-Jena-Leipzig (DFG FZT-118, 202548816) sTeTra working group through sDiv, the Synthesis Centre of iDiv (F.S. and M.D.)
German Research Foundation grant to the German Centre for Integrative Biodiversity Research (iDiv) Halle-Jena-Leipzig (DFG FZT-118, 20254881) (S.A.B., J.M.C., N.E., R.vK. and A.S)

German Research Foundation grant to the Gottfried Wilhelm Leibniz Prize (Ei 862/29-1) (N.E.)
CAPES (Coordenação de Aperfeiçoamento de Pessoal de Nível Superior Coordination for the Improvement of Higher Education Personnel), process number: \#88881.129579/2016-01 (Finance Code 001) (I.T.S.)
Leverhulme Trust (RPG-2019-402) (A.E.M., M.D.)
Leverhulme Trust (ECF-2021-512) (M.M.)
Leverhulme Trust through the Leverhulme Centre for Anthropocene Biodiversity (RC-2018-021) (I.S.M., M.D.)
European Union (ERC coralINT, 101044975) (M.D.)
Schmidt Science Fellowship (G.N.D.)
Fisheries Society of the British Isles PhD Studentship (A.F-.E)
Knut och Alice Wallenberg Foundation (A.D.B.)
USDA Hatch grant MAFES (1011538) (B.M.)
NSF EPSCOR Track II grant (2019470) (B.M.)

## Author contributions:

Conceptualization: all coauthors
Data curation: I.S.M., V.B., C.F.Y.C., R.vK, A.B., A.F-E. and F.M.
Methodology: all coauthors
Formal analysis: I.S.M.
Supervision/Project administration: I.S.M., M.D., F.S., and J.M.C.
Visualization: I.S.M., M.D., F.S., C.F.Y.C., B.P., A.F-E., A.E.B., M.M., and S.A.B.

Writing - original draft: I.S.M., M.D., F.S., R.F. and J.M.C
Writing - review and editing: all coauthors
Funding acquisition: I.S.M, F.S. and M.D.
Competing interests: The authors declare that they have no competing interests.

Data and materials availability: The published BioTIME data (17) can be accessed on Zenodo (https://doi.org/10.5281/zenodo.2602708) or through the BioTIME website (http://biotime.standrews.ac.uk/); Links to the individual datasets are also provided in table S1. The selected data used in this article and the R scripts used to generate the main results of the study are archived online on Zenodo (45).

## Supplementary Materials

Materials and Methods
Figs. S1 to S14
Table S1
References (46-260)


Fig. 1. Components of temporal changes in mean body size. (A) Shifts in mean assemblage-level body size can occur due to within-species changes (vertical axis), compositional changes (horizontal axis), or a combination of both components, displayed as change between two time points: from time 1 to time 2 . Boxes represent assemblages made up of individual organisms (icons), with different colors and shapes representing different species. Icon size represents the body size of an individual within each species. The body size distribution at time 1 is shown in the middle (i.e., the example of no change from time 1 to time 2), with different change outcomes for time 2 shown in the other cartoons. Note that the vertical placement (axis) represents the within-species (population or intraspecific) changes through time in mean body size. This could be a mix of increases due to smaller individuals growing larger or being replaced by larger individuals (shown in the right-hand boxes) and decreases in the average size of individuals (shown in the left-hand boxes). The horizontal placement (axis) indicates change in mean body size resulting from the gain or loss of species (compositional turnover), or a change in the relative abundance of the species present in an assemblage (even without local extinction or immigration of species). (B) The two components of body size change can reinforce or counteract each other. Assemblage body size change is most pronounced when both components operate in the same direction (towards either shrinking or increasing body size), such that the covariance between compositional and within-species changes is positive. When only one component is involved (i.e., change in one axis but not the other), body size change tends to be lower, and with negative covariance it is possible to have change in one component cancel out change in the other (middle panel). If they counteract each other, the overall direction of change will depend on which component shows the higher absolute effect (contribution).


Fig. 2. Changes in mean body size across the $\mathbf{5 , 0 2 5}$ assemblages. (A) Average individual size across the full set of assemblage time-series. Each point is coloured by density break with colder colors indicating lower densities. (B) Density plots of the distribution of slopes of change in average individual body size. The full set of 5,025 assemblage time-series is shown in light gray. Yellow, blue and orange represent respectively the subset of assemblages for which strong evidence ( $\mathrm{P}<0.01$ ), moderate evidence $(\mathrm{P}<0.05)$ and weak evidence $(\mathrm{P}<0.1)$ of change was detected when testing slopes against 0 . Dotted lines show slope of 0 , while blue dashed lines show the mean slope across the blue data (traditional significance value) and the respective $90 \%$ credible interval.


Fig. 3. Compositional versus within-species body size change through time in $\mathbf{5 , 0 2 5}$ assemblages representing $\mathbf{4 , 2 9 2}$ species of fish, plants, invertebrates, mammals, herpetofauna, and marine benthic organisms. (A) Relationship between population-level (i.e., within-species) changes and assemblage-level (i.e., compositional) changes. Both axes show \% changes standardised by the number of years between the first and last year sample of the assemblage (duration); assemblages (points) are coloured by density break (colder colours indicating lower densities). Dashed lines show $x=0, y=0, x=y$ and $y=-x$. (B) Frequency distributions (in percentage) of the number of assemblages ( $\mathrm{n}=5025$ ) in the different scenarios depicted in Fig. 1B. Assemblages with $\%$ change year ${ }^{-1}$ higher than $10 \%$ ( $n=517$; see fig. S2 \& S7) are not shown in panel (A), but are included in (B).


Fig. 4. Patterns of temporal body size change vary across (A) taxa, (B) realms, (C) climates, and the (D) globe. Plots show the frequency distributions (in percentage) of the number of assemblages across different groups for each scenario depicted in Fig. 1B. Dashed lines mark the 50\% threshold.


Fig. 5. Changes in assemblage abundance, biomass and body size. Density plots of the distribution of slopes of (A) change in total abundance of individuals (of all species), and (B) change in total biomass in an assemblage, as a function of time, for the same assemblages as shown in Fig. 3. The full set of 5,025 assemblage time-series is shown in light gray. Yellow, blue and orange represent respectively the subset of assemblages for which strong evidence ( $\mathrm{P}<0.01$ ), moderate evidence ( $\mathrm{P}<0.05$ ) and weak evidence ( $\mathrm{P}<0.1$ ) of change was detected when testing slopes against 0 . Dotted lines show slope of 0 , while blue dashed lines show the mean slope across the blue data (traditional significance value) and the respective $90 \%$ credible interval; (C and D) The bottom panels show the different relationships between variables. Only assemblages for which strong or moderate evidence $(\mathrm{P}<0.05)$ were detected for both variables plotted are shown in blue, while purple highlights the assemblages for which significant changes through time were detected in all 3 variables ( $\mathrm{n}=50$ ), all remaining assemblages are shown in light grey: (C) change in average body size as a function of abundance changes (note that $79 \%$ of the blue dots are in the quadrant where abundance increases and body size decreases), and (D) change in biomass as a function of abundance changes.

# Science <br> MIAAAS 

## Supplementary Materials for

## Widespread shifts in body size within populations and assemblages.

Inês S. Martins*, Franziska Schrodt, Shane A. Blowes, Amanda E. Bates, Anne D. Bjorkman, Viviana Brambilla, Juan Carvajal-Quintero, Cher F. Y. Chow, Gergana N. Daskalova, Kyle Edwards, Nico Eisenhauer, Richard Field, Ada Fontrodona-Eslava, Jonathan J Henn, Roel van Klink, Joshua S. Madin, Anne E. Magurran, Michael McWilliam, Faye Moyes, Brittany Pugh, Alban Sagouis, Isaac Trindade-Santos, Brian McGill, Jonathan M. Chase, Maria Dornelas.

*correspondence to: istmartins@gmail.com

This PDF file includes:
Materials and Methods
Figs. S1 to S14
Table S1
References (46-260)

## Materials and Methods

Our analysis is based on bringing together trait data and ecological assemblage time-series. We used two sources of body size trait data: direct measurements of species' body size (biomass) taken over time in the field (hereafter called type 1, fig. S1) and species' average body size estimates from major published trait databases (hereafter called type 2 data, fig. S2). Both datasets of time-series cover a variety of body sizes and body size change trends (fig. S3). Below, we first describe the a priori quality criteria, standardisations and subsequent calculations and statistical analyses that we used to provide an analysis of the processes behind current assemblage- and population-level body size changes using type 1 data. The results of these analyses are presented in the main text. Then, we present the methodology we followed to explore the global patterns of body size change when considering compositional changes alone and using different types of trait data (type 1 and type 2; see Supplementary Analysis section).

## Assemblage time-series data (BioTIME Database)

BioTIME is the largest global, open-access database of assemblage composition time-series (17). This database includes data on multiple multicellular taxa (e.g., plants, fish, birds, mammals, invertebrates), with over 12.5 million species-level records representing $\sim 46,000$ species. Each BioTIME study contains distinct samples measured (with a consistent methodology) over time, which could be fixed plots (i.e., 'single-site' studies where measures are taken from a set of specific georeferenced sites at any given time) or wide-ranging surveys, transects, tows, and so on (i.e., 'multi-site' studies where measures are taken from multiple sites that may or may not align from year to year). Because the spatial extent varies across studies, we followed previous approaches (11,40) to identify and standardise 'multi-site' studies using a global grid of $96 \mathrm{~km}^{2}$ hexagonal cells using dggridR (47). Studies that were contained within a single cell were not partitioned. Following this step, each sample was assigned a different combination of study ID and grid cell (based on its latitude and longitude) resulting in a unique identifier for each assemblage time-series within grid cells, thus allowing for the integrity of each study and each sample to be maintained. Then sample-based rarefaction was applied to standardise the number of samples per year within each time-series (11, 46, 48). Finally, we only retained observations sampled after the year 1960 ( $99.4 \%$ of all the data was recorded after this period) and restricted our analysis to time-series with at least five different sampled years. For the analyses presented in the main text, we further subsetted the database, and considered only records that contain both abundance information (i.e., counts of the number of individuals) and biomass estimates directly measured in the field. In total, we considered 5,025 time-series from 45 studies across the globe (fig. S1; Table S1).

## Trait data (BioTIME Database)

We extracted body size trait data directly from BioTIME using both abundance and biomass estimates measured at the same time and place. From each record $i$, we estimate average body size of individuals (BS) by considering that:

$$
B S_{r, s, t, i}=\frac{B_{r, s, t, i}}{N_{r, s, t, i}},
$$

where $B$ is biomass and $N$ is abundance recorded in year $t$, for species $s$ within the assemblage time-series $r$. Note that biomass is only measured at the individual scale (abundance=1) for $\sim 22 \%$ of the data, thus we refer to this measure as average individual body size. Here, we include all taxonomic groups for which the appropriate data were available including groups poorly represented in most trait compilations (e.g., invertebrates). In total, we considered the following taxon groups: fish, benthos, plants, mammals, invertebrates, and herpetofauna (reptiles
and amphibians). We did not distinguish taxa across realms, but studies that included multiple taxa (i.e., studies that sampled multiple taxon groups simultaneously) were re-classified based on the dominant taxa represented. On average, we estimated multiple body size measurements for 4,292 species within the 5,025 assemblage time-series (average $\sim 39$ species per assemblage time-series; fig. S1).

## Partitioning body size change - Price equation

To partition temporal changes in average individual body size, we used an extension of the Price equation (20,22), that allows an exact partition of trait change, $\Delta z$, in an observed dataset:

$$
\Delta z=\Sigma\left(\Delta q_{s}\right) z_{s}+\Sigma q_{s}^{\prime}\left(\Delta z_{s}\right)
$$

The first term on the left-hand side of the equation for $\Delta z$ accounts for total body size change caused by changes in frequency due to species selection, i.e., changes in the relative abundance or presence of species with a certain property value (body size; e.g., local extirpations or colonisations). This term reflects the effect of species turnover. The second term describes the part of total change caused by changes in mean property values, reflecting the effect of withinspecies variation (e.g., larger individuals within a species being replaced by smaller individuals of the same species). Together, the two terms sum up to the actual change in communityweighted mean (CWM) body size in an assemblage, $\Delta z$. Given this, we quantified changes in frequency as $\Delta q_{s}=q_{s}^{\prime} q_{s}$, where $q_{s}$ and $q_{s}^{\prime}$ are, respectively, the before and after relative abundance of species $s$; and changes in mean property value as $\Delta z_{s}=z_{s^{-}}^{\prime} z_{s}$, where $z_{s}$ and $z^{\prime}{ }_{s}$ represent, respectively, the before and after mean individual body size of species $s$. In assemblage time-series where multiple individual $B S_{r, s, t, i}$ estimates were available for the same year and species (e.g. when the assemblage was monitored more than once a year), an abundance-weighted mean was used instead. Finally, when $z_{s}$ was not available (i.e., colonisations did not occur) we considered $z_{s}=z_{s}^{\prime}$ for that species, and thus $\Delta z_{s}$ is equal to 0 and no change occurs due to changes in mean property values (within-species changes).

For each assemblage time-series, we used the Price equation to partition body size changes that occurred between two years, the last year $\left(t_{2}\right)$ and the first year $\left(t_{1}\right)$, where $t=t_{2}-t_{1}+1$ is the full length of the assemblage time-series. In order to ensure comparability among timeseries of different durations, both the within-species and the composition component of body size change were converted to proportional changes relative to the starting size of the assemblage. This was done by dividing each component of change by the initial assemblage CWM and standardising it by duration (i.e., dividing by t ). These quantities were expressed in units of $\%$ change.year ${ }^{-1}$. Patterns across all assemblages are represented in Fig. 3 and fig. S4, and patterns across the different taxa, realms and climates are shown in Fig. 4.

## Sensitivity analyses

Many of the assemblage time-series varied in length ( $27.3 \pm 12 \mathrm{yr}$, mean $\pm \mathrm{SD}$ ), with varying start and end points. To examine whether our results were sensitive to such effects, we repeated the analysis using alternative start $\left(t_{1}\right)$ and end times $\left(t_{2}\right)$ within the same assemblage. This analysis included a scenario where the first year was fixed, a scenario where the last year was fixed and a scenario where both years varied randomly. For each of the three scenarios, we repeated the analysis 100 times, where for each iteration we used the Price equation to partition body size changes that occurred between the selected two years in a given assemblage (as done in our main analysis); and reported the median effect of each component and their dispersion (interquartile range) across all iterations (fig. S5). Additionally, we also used the Price equation to partition body size changes that occurred between all pairs of consecutive years in a given assemblage, to investigate the bias found in intermediate states (fig. S6). Despite slight
differences across scenarios, the results were largely concordant and yielded the same directional trends.

Given that some of the estimates were extreme with very large changes to assemblage level body size ( $\sim 0.7 \%$ of assemblages show increases in compositional change in body size.year ${ }^{-1}$ $>100 \%$; fig. S4), there could be some concerns about errors in the measurement or measurement reporting of the abundance and biomass estimates (and consequently body size estimates) in the original datasets. We performed in-depth checks of the raw data within affected individual datasets (see fig. S7 for an example) and found that such effects seem to be a true representation of changes occurring in the assemblages. Nevertheless, these few assemblages have the potential to over-influence the overall effects found, so we chose to report robust statistics (median and interquartile range) that de-emphasise such extreme cases without removing them, when appropriate (e.g., fig. S5 and S8).

For some assemblage time-series the sample-based rarefaction process could lead to a different species composition. To ensure our results were robust to the random samples selected by the sample-based rarefaction process, we performed a bootstrap analysis re-running the analysis described in the main text (using first and last year only) 100 times, each time using a different dataset after the sample-based rarefaction process was applied. Only the results of one iteration are presented in the main text, but plots of the distribution of results across the 100 rarefaction iterations can be seen in fig. S8.

## Supplementary Analyses

At large scales and often using species-level trait values, previous studies generally conclude that body size has, on average, been decreasing ( $2,3,27$ ). This is due both to compositional changes, whereby bigger species are disproportionately replaced by smaller species (3) and within-species changes associated with the removal of larger individuals (42). However, data limitations on individual-level body size make it difficult to assess the importance (and signal) of the latter, thus, constraining global assessments to quantify body size change using species-level trait values alone. Here, we follow a similar approach but use BioTIME data to test if the assemblage body size shrinkage patterns presented in the main text are observed at the global scale when using different types of trait data. For this analysis, we used two distinct subsets of BioTIME data: the 45 studies with directly measured estimates of body size (as described earlier in the Materials and Methods; type 1 data), and a larger subset of studies for which estimates of body size could be retrieved from major published databases (species trait averages; type 2 data). Note that when matching with trait databases (i.e., the type 2 data), we did not work with only the subset of 45 studies featured in the main text but considered instead all BioTIME data that reported counts of the number of individuals and met our duration criteria. The different steps of data preparation and analysis for the former (type 1) are summarized earlier in the Materials and Methods of this document (19), any additional data, statistical analyses and supplementary results are described in detail below.

## Additional Trait data (Trait databases)

Five open-access global databases were identified as having partially overlapping observations with species listed in the BioTime dataset: AmphiBIO (49); https://doi.org/10.6084/m9.figshare.4644424.v5), TRY database (50); https://www.try-db.org/), EltonTraits 1.0 (51); https://doi.org/10.6084/m9.figshare.c.3306933.v1), FishBase (52); www.fishbase.org) and Carabids.org (53); http://www.carabids.org). While a number of traits related to body size are available in these datasets, here, and given the broad-scale goal of the paper, we choose to select the body size trait that had the higher cover for each taxon. For birds and mammals, it was body mass ('body_size_mm' field). However, for ectotherms, length was used instead, as it is a more commonly available measure and considered more reliable than
body mass. For fish and beetles (hereafter: Invertebrates) we used maximum body length ('MaxLengthTL' and 'maxSize' fields, respectively), for amphibians we used snout-vent length ('body_size_mm' field), and for plants we used plant height ('veg_height' field). Only specieslevel trait values were considered and when there were multiple values for a particular species (i.e., TRY database), we took the median.

## Synthesis and harmonisation of data

To merge and harmonise the assemblage time-series and trait databases data and optimise species matching, we first followed a series of steps to deal with species names and incompatibilities between the two sources of data. As a preliminary step, we reviewed species names listed in BioTIME with the taxize R package (54). An additional field was created with potential synonyms, or alternative scientific names (identified misspelling errors). Finally, common names were also flagged and, when possible, converted to scientific synonyms using additional data sources (ITIS and NCBI), helped by manual inspection based on the description provided. This process allowed for 913 species ( $11 \%$ ) extra matches between the two sources of data. Because the sample-based rarefaction could result in different species composition for some assemblage time-series, we decided to work with all 100 BioTIME resamples (see 'Sensitivity analysis' section in Material and Methods (19)). The matching was then done for each BioTIME resample dataset separately, and filtered to keep only records from where trait values were available (i.e., common species across datasets). On average, we retained records for $\sim 5,000$ species across $\sim 20,200$ assemblage time-series (fig. S2), with an average of $\sim 75 \%$ completeness (i.e., proportion of species in the assemblage time-series from which trait data was retrieved).

## Calculating body size

The way we calculated average body size of individuals in a given year $\left(\widehat{B S}_{r, t}\right)$ depended on the source of the trait data. For assemblages time-series matched with trait databases data (type 2 data), $\widehat{B S}_{r, t}$ was calculated, by averaging all species' individual body sizes, weighted by their abundances within the assemblage. For assemblage time-series with directly measured estimates of body size (type 1 data), we first calculated for each year assemblage-level total abundance and total biomass by tallying the number of individuals and biomass (regardless of species) sampled within that year, respectively. The average individual size in a given year $(\widehat{B S} r, t)$ was then retrieved by dividing the sum of the biomass by the total abundance reported in that year for that assemblage time-series. To make all body size estimates comparable, all values were standardised using classic z -scores, where individual observations in a group are scaled relative to the mean and standard deviation of all observations of that group. This was done for each assemblage time-series within each dataset separately.

## Statistical analyses - Models of body size change

We explored the global patterns of body size change using mixed-effects models. Year (mean-centered) was included as a fixed effect, and was also included as a random slope varying across studies and assemblages, with assemblages nested into the original studies from which they originated in order to account for the non-independence of the time-series. All statistical models were fitted in a Bayesian framework using the package 'brms' (55) in R (v3.6.3; (50)). We modelled average individual body size change assuming a skew-normal distribution and an identity link function. The overall model structure implemented using the bms syntax was:

$$
y_{j, i, t} \sim \operatorname{SkewNormal}\left(\mu_{j, i, t}, \sigma, a\right)
$$

$$
\mu_{j, i, t}=\beta_{0}+\beta_{0 j}+\beta_{0 j, i}+\left(\beta_{1}+\beta_{1 j}+\beta_{1 j, i}\right) \text { year }_{j, i, t},
$$

where $y_{j, i, t}$ is the average individual body size change in year $t$ of the $i$ th assemblage in the $j$ th study. year $r_{j, i, t}$ is the time in years, $\beta_{0}$ and $\beta_{1}$ are the global intercept and slope (fixed effects), $\beta_{0 j}$ and $\beta_{1 j}$ are the study-level departures from $\beta_{0}$ and $\beta_{1}$ (respectively; study-level random intercept and slope), and $\beta_{0 j, i}$ and $\beta_{1 j, i}$ are the (nested) assemblage-level departures from $\beta_{0 j, i}$ and $\beta_{1 j, i}$ (respectively; assemblage-level random intercept and slope). We used weakly regularizing priors for the global intercept and slope, residual variation $(\sigma)$, and skew parameter (a):

$$
\begin{gathered}
\beta_{0} \sim N(0,2), \\
\beta_{1} \sim N(0,1), \\
\sigma \sim \text { student } t(3,0,2.5), \\
a \sim N(0,4) .
\end{gathered}
$$

Group level parameters were drawn from the student-t distribution:

$$
\sigma_{0 j}=\sigma_{1 j}=\sigma_{0 j i}=\sigma_{1 j i} \sim \operatorname{student} t(3,0,2.5) .
$$

Correlations between levels of the grouping-factors were estimated using the Cholesky decomposition (L) of the correlation matrix, with a Lewandowski-Dorota- Joe (LKJ) prior:

$$
L \sim L K J(2)
$$

The model was fit to 100 resamples of each dataset (i.e., type 1 and type 2 ) to adjust for any variation in species composition arising when sample effort was standardised using samplebased rarefaction. For each model fit, we extracted the 100 draws from the posterior distribution, which were combined for making inferences.

## Body size, abundance, and biomass change

As mean body size emerges from the ratio of biomass and abundance (see section "Trait data (BioTIME Database)"), change in either biomass or abundance can be responsible for any observed body size changes. To explore these effects, we quantified trends in biomass and abundance across individual assemblage time-series with directly measured estimates (type 1 data). This was achieved by fitting ordinary least-squares (OLS) regression models for each assemblage separately, with either average individual body size, total abundance, or total biomass (centered and scaled) as a function of time (year, mean-centered). All sampled years were considered. The set of slopes $(\beta)$ of these linear models is shown in Fig. 2 (change in body size) and Fig.5A and B (change in abundance and biomass, respectively). Additionally, we evaluated the associations between the temporal trends in total abundance, total biomass, and mean body size, by comparing the slopes of change of assemblages for which statistically significant trends were found across two or more variables (Fig. 5C and D; fig. S13-14). Lastly, the same approach was used to explore the variability of body size change trends present in type 1 and type 2 data (fig. S3). All calculations and statistical analyses were performed in R-3.6.3 (50).


Fig.S1.
Distribution of study data characteristics (BioTIME data, type 1). (A) Location of the studies with direct measures of body size (based on central coordinates; $n=45$ ), (B) number of assemblage time-series in each study, (C) species richness observed across assemblages, (D) total number of body size observations across assemblages, (E) duration of sampling, and taxonomic distribution of: (F) species represented, (G) body size observations and (H) assemblage time-series.


Fig. S2.
Distribution of study data characteristics (BioTIME data, type 2). (A) location of the studies with indirect measures of body size (based on central coordinates; $\mathrm{n}=151$ ), (B) number of assemblage time-series in each study, (C) species richness observed across assemblages, (D) total number of body size observations across assemblages, (E) duration of sampling, and taxonomic distribution of: (F) species represented, (G) body size observations and (H) biodiversity time-series.


Fig. S3.
Changes in body size across type 1 and type 2 assemblages. The left-hand graphs show the patterns of body size change using direct measures of body size (type 1 data), while the right- hand graphs show patterns using species' average body size estimates from trait databases (type 2 data). A-B shows patterns across the full sets of assemblages, and $\mathbf{C - D}$ when fish assemblages are excluded. E-F show changes in body size across common type 1 and type 2 assemblages considering only species where both types of data were available, using either (E) direct measures of body size or (D) species' average body size estimates; and G-H when fish assemblages are excluded.


Fig. S4.
Patterns of body size change through time in $\mathbf{5 , 0 2 5}$ assemblages. Relationship between population-level (i.e., within-species) changes and assemblage-level (i.e., compositional) changes. Both axes show \% changes standardised by the number of years between the first and last year sample on the assemblage (duration); assemblages (points) are coloured by density break (colder colours indicating lower densities); Dashed lines show $x=0, y=0, x=y$ and $y=-x$. Please see fig. S7 for more details.


Fig. 55.
Patterns of body size shrinkage through time are not influenced by start and/or end points.
Results using alternative start and/or end points. Analysis in the main text only compared the first and the last year of the time-series. The sensitivity analyses used instead an (A) fixed start year and random last year, a (B) random first year but fixed end year, or a (C) random start and end year. Points (assemblages) and grey lines indicate the assemblage median body size change and its IQR across 100 iterations (for each iteration changes were calculated using years chosen randomly according to the scenario assumptions). Points are coloured by density break (colder colours indicating lower densities). Inset histograms show $\%$ of assemblage where the median and IQR interval falls below (shrinkage) or above (growth) the $y=-x$ line. Assemblages where the variation (IQR) crosses the $y=-x$ line (and hence neither shrinking nor growing) are not shown.


Fig. S6.
Body size change through time between two consecutive years. (A) Partitions for consecutive years across the 5,025 assemblages ( $n=43,804$ ), where each point represents one pair of consecutive years in a given assemblages (coloured by density break). (B) Frequency distributions (in percentage) of the number of pairs in the different scenarios depicted in Fig. 1B. Points with \% change year ${ }^{-1}$ higher than $50 \%$ are not shown in panel (A), but are included in (B).


Fig. S7.
Body size change through time: a look into the extremes. The size of the assemblage through time will be determined by changes in species composition and within-species body size changes of each species. More extreme changes can occur in locations where assemblages are composed of species with very different body sizes to begin with, or/and when turnover occurs. For example, in the assemblage highlighted in (A) a wide spectrum of benthic organisms was sampled together. (B) In the first year (1979; orange), only small benthic organisms were recorded, although in high abundances $\left(\mathrm{CWM}_{\text {before }}=6.8 \times 10^{-3}\right)$, however, by the last year (2006; green) a complete turnover in the assemblage had occurred. Despite decreases in both species' richness and abundance, the size of the assemblage increased $\left(\mathrm{CWM}_{\text {after }}=3.193\right)$, with the addition of several large-bodied species, including several individuals of the genus Dasyatis. This led to an assemblage $\sim 467.5$ times bigger (in weight) than the original, an increase of $\sim 1666 \%$ per year due to compositional changes alone. Note that the illustrations are not to scale, and thus the size of the icons does not represent the real difference between the average body size of the species in the wild.


Fig. S8.
Sensitivity analysis using different rarefaction subsets. (A) 20.9\% of assemblages (black points; $\mathrm{n}=1052$ ) are affected by the sample-based rarefaction process. Grey lines indicate the variation (IQR) found in this subset of assemblages across 100 resamples. Assemblages unaffected by this process are shown in grey on the background. (B) Assemblages where the variation (IQR) crosses the $\mathrm{y}=-\mathrm{x}$ line (and hence neither shrinking nor growing; $\mathrm{n}=365$ ). (C) Histograms show \% of assemblage where the median and IQR interval falls below (shrinkage) or above (growth) the $y=-x$ line after excluding assemblages highlighted in (B). For clarity, assemblages with $\%$ change.year ${ }^{-1}$ higher than $20 \%$ are not shown in panels (A) and (B), but are included in (C).


Study-level slope estimates
Fig. S9.
Global temporal trend in body size change across assemblages. Overall trend in average individual body size (centred and scaled) change as a function of year when considering (A) species' average body size estimates or (B) direct measurements of size. Lines depict the global median and $90 \%$ credible interval across all assemblages. Black lines show study-level variation. (C) Density ridges of posterior distributions of the study-level slope coefficients for a given taxon. (D) Estimates of change for each taxon, each point represents a single study, with the bar showing the $90 \%$ credible interval; studies are arranged by their median value (point). In all plots, colour represents the type of body size data: orange $=$ average body size from trait databases ( 20,173 assemblages across 149 studies), dark green $=$ body size from direct measurements as reported in the BioTIME database ( 5,025 assemblages across 45 studies).


Fig. S10.
Patterns of average individual body size change across the different realms. Estimates of change for each realm when considering (A) species' average body size estimates ( 149 studies) or (B) direct measurements of size ( 45 studies). Each data point represents a single study, with the bar showing the $90 \%$ credible interval; studies are arranged by their median change value (point).

# Submitted Manuscript: Confidential <br> Template revised November 2022 



Fig. S11- Same as fig. S9, A and B, but when fish are excluded from both datasets.


Fig. S12.
Patterns of temporal body size change associated with compositional change alone (type 2 data). Patterns across (A) taxa, (B) realms, (C) climates, and the (D) globe as shown in Fig.4. Dashed lines mark the $50 \%$ threshold.


Fig. S13.
Body size change through time as a function of biomass change. Only assemblages for which strong or moderate evidence ( $\mathrm{P}<0.05$ ) was detected for both variables plotted are shown in blue. Purple highlights the assemblages for which significant changes through time were detected in all three variables ( $\mathrm{n}=50$ ), all remaining assemblages are shown in light grey.


Fig. S14.
Same as Fig. 5, but when fish are excluded from the dataset (n=1116). Symbols represent different non-fish taxa.

Table S1. Details on the datasets used in this study.

| $\begin{gathered} \text { Study } \\ \text { ID } \\ \hline \end{gathered}$ | Start year | End year | $\begin{aligned} & \hline \mathrm{Nr} \\ & \mathrm{yrs} \\ & \hline \end{aligned}$ | Taxon | Realm | Climate | type1 | type2 | References |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 39 | 1970 | 2015 | 45 | Birds | Terrestrial | Temperate |  | x | (57-60) |
| 42 | 1960 | 1979 | 20 | Birds | Terrestrial | Temperate |  | x | (61-67)* |
| 44 | 1962 | 1977 | 16 | Plants | Terrestrial | Temperate | x | x | (68-70)* |
| 45 | 2005 | 2010 | 5 | Fish | Marine | Tropical | x | x | $(71,72)$ |
| 46 | 1960 | 1979 | 20 | Birds | Terrestrial | Temperate |  | x | (73) |
| 47 | 1960 | 1977 | 18 | Birds | Terrestrial | Temperate |  | x | (74) |
| 51 | 1964 | 1977 | 14 | Birds | Terrestrial | Temperate |  | x | $(75,76)$ |
| 52 | 1968 | 1980 | 13 | Mammals | Terrestrial | Polar |  | x | $(77,78)$ |
| 53 | 1966 | 1976 | 10 | Mammals | Terrestrial | Temperate |  | x | $(79,80)$ |
| 56 | 1989 | 2019 | 31 | Mammals | Terrestrial | Temperate | x | x | (81) |
| 58 | 1991 | 2008 | 18 | Birds | Terrestrial | Tropical |  | x | $(82,83)$ |
| 59 | 1977 | 2002 | 26 | Mammals | Terrestrial | Temperate |  | x | (84) |
| 60 | 1982 | 2005 | 6 | Plants | Terrestrial | Tropical |  | x | (85-90) |
| 81 | 1997 | 2010 | 14 | Mammals | Marine | Temperate |  | x | $(91,92)$ |
| 86 | 1985 | 2007 | 16 | Plants | Marine | Temperate | x |  | (93) |
| 91 | 1992 | 1999 | 8 | Birds | Marine | Temperate |  | x | (94) |
| 100 | 1981 | 2011 | 31 | Fish | Marine | Temperate |  | x | (95-97)* |
| 108 | 1981 | 2006 | 25 | Birds | Marine | Global |  | x | (98) |
| 112 | 1973 | 2005 | 22 | Fish | Marine | Temperate/Tropical |  | x | (99) |
| 119 | 1970 | 2010 | 41 | Fish | Marine | Temperate | x | x | (100) |
| 121 | 2000 | 2010 | 8 | Fish | Marine | Temperate/Tropical |  | x | (101) |
| 123 | 2000 | 2009 | 10 | Fish | Marine | Temperate | x | x | (102) |
| 125 | 1988 | 2000 | 12 | Fish | Marine | Temperate | x | x | (103) |
| 126 | 1974 | 1980 | 7 | Fish | Marine | Temperate |  | x | (104) |
| 127 | 1980 | 1989 | 10 | Fish | Marine | Temperate | x | x | (105) |
| $\dagger \dagger 163$ | 1993 | 2004 | 12 | Benthos | Marine | Temperate | x |  |  |
| $\dagger+163$ | 1993 | 2004 | 12 | Birds | Marine | Temperate |  | x | (106) |
| $\dagger$ †163 | 1993 | 2004 | 12 | Fish | Marine | Temperate |  | x |  |
| †166 | 1966 | 1990 | 23 | Birds | Marine | Global |  | x |  |
| †166 | 1971 | 1988 | 13 | Mammals | Marine | Global |  | x | (107-112) |
| $\dagger 169$ | 1987 | 2006 | 20 | Birds | Marine | Temperate |  | x |  |
| $\dagger 169$ | 1987 | 2003 | 7 | Fish | Marine | Temperate |  | x | (113-116) |
| $\dagger 169$ | 1987 | 2006 | 20 | Mammals | Marine | Temperate |  | x |  |
| 171 | 1992 | 2008 | 17 | Mammals | Marine | Temperate/Tropical |  | x | (117) |
| $\dagger 172$ | 2000 | 2009 | 10 | Birds | Marine | Temperate |  | x |  |
| $\dagger 172$ | 1998 | 2009 | 12 | Mammals | Marine | Temperate |  | x | (118-121) |
| 176 | 1999 | 2010 | 12 | Invertebrates | Marine | Temperate | x |  | (122) |
| 180 | 1970 | 1995 | 26 | Fish | Marine | Polar/Temperate |  | x | (123) |
| 182 | 1988 | 2009 | 22 | Fish | Marine | Temperate |  | x | (124) |
| 189 | 2001 | 2010 | 10 | Fish | Marine | Tropical |  | x | (125) |
| 190 | 2001 | 2010 | 10 | Fish | Marine | Tropical |  | x | (126) |
| 192 | 1960 | 1980 | 19 | Mammals | Marine | Polar/Temperate |  | x | (127) |
| 195 | 1978 | 2007 | 30 | Birds | Terrestrial | Temperate |  | x | (128) |
| 196 | 1987 | 2012 | 12 | Fish | Marine | Temperate |  | x | (129) |
| 197 | 1985 | 2013 | 28 | Fish | Marine | Temperate |  | x | (130)* |
| 198 | 1991 | 2013 | 23 | Fish | Marine | Temperate |  | x | (131)* |
| 205 | 2001 | 2009 | 8 | Fish | Marine | Temperate |  | x | (132)* |
| 206 | 1993 | 2008 | 16 | Fish | Marine | Temperate |  | x | (133)* |
| 207 | 2003 | 2008 | 6 | Fish | Marine | Temperate |  | x | (134)* |
| 208 | 1997 | 2007 | 11 | Fish | Marine | Temperate |  | x | (135)* |

Submitted Manuscript: Confidential
Template revised November 2022

| 209 | 1987 | 2010 | 24 | Fish | Marine | Temperate |  | x | (136)* |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 210 | 1965 | 2011 | 47 | Fish | Marine | Temperate |  | x | (137)* |
| 211 | 1980 | 1987 | 8 | Fish | Marine | Temperate | x | x | (138) |
| 212 | 1973 | 1980 | 8 | Fish | Marine | Temperate | x | x | (139) |
| 213 | 1963 | 2008 | 46 | Fish | Marine | Temperate | x | x | (140) |
| 214 | 1960 | 2010 | 46 | Plants | Terrestrial | Temperate |  | x | (141) |
| 215 | 1960 | 2008 | 49 | Birds | Terrestrial | Temperate/Tropical |  | x | (142)* |
| 216 | 2001 | 2005 | 5 | Birds | Terrestrial | Temperate |  | x | (143) |
| 217 | 1992 | 2006 | 14 | Birds | Terrestrial | Temperate |  | x | (144) |
| 218 | 2006 | 2010 | 5 | Birds | Terrestrial | Temperate |  | x | (145) |
| 219 | 1995 | 2011 | 17 | Herpetofauna | Terrestrial | Temperate |  | x | (146) |
| 220 | 1995 | 2011 | 17 | Birds | Terrestrial | Temperate |  | x | (146) |
| 229 | 1988 | 2013 | 26 | Fish | Freshwater | Temperate |  | x | (147) |
| $\dagger 231$ | 2000 | 2005 | 6 | Fish | Marine | Temperate |  | x |  |
| $\dagger 231$ | 2000 | 2005 | 6 | Herpetofauna | Marine | Temperate |  | x | (148) |
| $\dagger 231$ | 2000 | 2005 | 6 | Plants | Marine | Temperate |  | x |  |
| 232 | 1975 | 1999 | 16 | Fish | Marine | Polar/Temperate | x |  | (149) |
| 234 | 1965 | 2002 | 7 | Plants | Terrestrial | Temperate | x | x | (150-150) |
| 236 | 1995 | 2006 | 12 | Fish | Freshwater | Temperate |  | x | (157) |
| 240 | 2003 | 2015 | 13 | Plants | Terrestrial | Temperate | x | x | (158) |
| 242 | 1998 | 2006 | 7 | Plants | Terrestrial | Temperate |  | x | (159) |
| 243 | 1992 | 2014 | 22 | Plants | Terrestrial | Temperate | x | x | $(160,161)$ |
| 244 | 1999 | 2012 | 14 | Birds | Marine | Temperate |  | x | (162) |
| 246 | 2000 | 2014 | 15 | Fish | Marine | Temperate |  | x | (163) |
| 247 | 1975 | 2020 | 46 | Invertebrates | Freshwater | Temperate | x |  | (164) |
| 248 | 2000 | 2012 | 13 | Plants | Terrestrial | Temperate |  | x | (165) |
| 249 | 1992 | 2015 | 24 | Invertebrates | Terrestrial | Temperate |  | x | (160) |
| 252 | 1978 | 1989 | 12 | Fish | Marine | Temperate | x | x | (167) |
| 254 | 1995 | 2018 | 24 | Plants | Freshwater | Temperate | x |  | (168) |
| 255 | 1989 | 2007 | 10 | Plants | Terrestrial | Temperate |  | x | (169) |
| 256 | 1987 | 2010 | 24 | Fish | Marine | Temperate |  | x | (136)* |
| 271 | 2000 | 2014 | 15 | Fish | Marine | Temperate |  | x | (170) |
| 275 | 2009 | 2014 | 6 | Herpetofauna | Terrestrial | Temperate | x |  | (171). |
| 277 | 1979 | 2008 | 10 | Plants | Terrestrial | Temperate |  | x | (68-70)* |
| 279 | 1979 | 2008 | 10 | Plants | Terrestrial | Temperate | x | x | (68, 69, 172)* |
| 288 | 1970 | 2005 | 29 | Fish | Marine | Temperate |  | x | (100) |
| 295 | 2008 | 2016 | 9 | Fish | Marine | Temperate | x | x | (173) |
| 296 | 2008 | 2016 | 9 | Fish | Marine | Temperate |  | x | (173) |
| 305 | 1976 | 1990 | 15 | Herpetofauna | Terrestrial | Temperate |  | x | (174) |
| 308 | 1980 | 1998 | 19 | Mammals | Terrestrial | Temperate | x | x | (175) |
| 311 | 1981 | 2013 | 33 | Mammals | Terrestrial | Temperate |  | x | (176) |
| 312 | 1961 | 1984 | 8 | Mammals | Terrestrial | Tropical |  | x | (177) |
| 316 | 1989 | 2006 | 18 | Herpetofauna | Terrestrial | Temperate | x |  | (178) |
| 317 | 1995 | 2005 | 9 | Plants | Terrestrial | Temperate |  | x | (179) |
| $\dagger$ †18 | 1994 | 2009 | 13 | Birds | Terrestrial | Temperate |  | x |  |
| $\dagger 318$ | 1994 | 2009 | 13 | Mammals | Terrestrial | Temperate |  | x | (180)* |
| 319 | 1990 | 2003 | 13 | Herpetofauna | Terrestrial | Temperate |  | x | (181) |
| 321 | 1995 | 2007 | 13 | Mammals | Terrestrial | Temperate | x | x | (182) |
| 324 | 2003 | 2007 | 5 | Plants | Terrestrial | Tropical |  | x | (183) |
| 325 | 2003 | 2007 | 5 | Plants | Terrestrial | Tropical |  | x | (183) |
| 327 | 1989 | 2005 | 17 | Mammals | Terrestrial | Temperate | x | x | (184) |
| 328 | 1979 | 2008 | 30 | Herpetofauna | Freshwater | Temperate |  | x | (185) |
| 329 | 1990 | 2010 | 6 | Plants | Terrestrial | Tropical |  | X | (185) |

Submitted Manuscript: Confidential
Template revised November 2022

| 332 | 1984 | 1995 | 12 | Fish | Freshwater | Temperate |  | x | (180) |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 335 | 1962 | 1974 | 12 | Fish | Freshwater | Temperate |  | x | (187) |
| 336 | 1989 | 2002 | 14 | Plants | Terrestrial | Temperate |  | x | (84) |
| 339 | 1960 | 2009 | 50 | Birds | Terrestrial | Temperate |  | x | (188) |
| 348 | 2006 | 2016 | 10 | Mammals | Terrestrial | Temperate/Tropical | x | x | (189) |
| 350 | 1989 | 2011 | 12 | Benthos | Marine | Temperate | x |  | $(190,191)$ |
| 351 | 1989 | 2004 | 13 | Benthos | Marine | Temperate | x |  | $(192,193)$ |
| 356 | 1971 | 2013 | 34 | Plants | Terrestrial | Tropical |  | x | (194) |
| 357 | 1994 | 2006 | 13 | Mammals | Terrestrial | Temperate |  | x | (195) |
| 358 | 1977 | 1992 | 16 | Birds | Terrestrial | Temperate |  | x | (196) |
| 359 | 2000 | 2012 | 13 | Fish | Marine | Temperate |  | x | (197) |
| 361 | 1962 | 1983 | 22 | Birds | Terrestrial | Temperate |  | x | (198) |
| 363 | 1963 | 1999 | 37 | Birds | Terrestrial | Temperate |  | x | (199) |
| 365 | 1997 | 2002 | 6 | Fish | Marine | Temperate |  | x | (200) |
| 366 | 1989 | 2013 | 25 | Mammals | Terrestrial | Temperate |  | x | (201) |
| 372 | 2005 | 2013 | 9 | Birds | Terrestrial | Temperate |  | x | (202). |
| 373 | 2005 | 2012 | 8 | Mammals | Terrestrial | Temperate |  | x | (203) |
| 374 | 2004 | 2014 | 11 | Birds | Marine | Temperate |  | x | (204) |
| 375 | 2004 | 2014 | 11 | Invertebrates | Terrestrial | Temperate | x | x | (205) |
| 377 | 2009 | 2013 | 5 | Birds | Terrestrial | Temperate |  | x | (205) |
| 381 | 1986 | 1992 | 7 | Herpetofauna | Terrestrial | Temperate |  | x | (206) |
| 382 | 1960 | 1967 | 8 | Mammals | Terrestrial | Temperate |  | x | $(207,208)$ |
| 402 | 2010 | 2015 | 6 | Fish | Freshwater | Tropical |  | x | (209) |
| 403 | 1989 | 1995 | 7 | Herpetofauna | Freshwater | Tropical |  | x | (210). |
| 412 | 1995 | 2000 | 6 | Fish | Marine | Temperate |  | x | (211) |
| 420 | 1964 | 2001 | 38 | Birds | Terrestrial | Polar/Temperate |  | x | (212) |
| 427 | 1977 | 2008 | 32 | Invertebrates | Freshwater | Temperate | x |  | $(213,214)$ |
| $\dagger 428$ | 1964 | 2015 | 43 | Birds | Marine | Temperate |  | x | (215-218) |
| $\dagger 428$ | 1960 | 2015 | 56 | Fish | Marine | Temperate |  | x |  |
| 430 | 1985 | 2016 | 32 | Fish | Freshwater | Temperate |  | x | (219) |
| 431 | 1984 | 2015 | 30 | Fish | Freshwater | Temperate |  | x | (219) |
| 432 | 1998 | 2016 | 19 | Fish | Freshwater | Temperate |  | x | (219) |
| 435 | 2012 | 2019 | 7 | Fish | Marine | Polar/Temperate | x |  | (220) |
| 435 | 2009 | 2019 | 11 | Invertebrates | Marine | Polar/Temperate | x |  |  |
| 436 | 2005 | 2012 | 5 | Fish | Marine | Tropical | x | x | (221) |
| 438 | 2006 | 2014 | 7 | Fish | Marine | Tropical | x | x | (222) |
| 439 | 1985 | 1997 | 13 | Birds | Terrestrial | Temperate |  | x | (223) |
| 440 | 1985 | 1997 | 13 | Birds | Terrestrial | Temperate |  | x | (223) |
| 441 | 1985 | 1997 | 13 | Birds | Terrestrial | Temperate |  | x | (223) |
| 442 | 1980 | 1985 | 6 | Birds | Terrestrial | Temperate |  | x | (224) |
| 444 | 1983 | 1987 | 5 | Birds | Terrestrial | Temperate |  | x | (225) |
| 446 | 2007 | 2011 | 5 | Mammals | Terrestrial | Temperate |  | x | (226) |
| 447 | 2006 | 2014 | 9 | Mammals | Terrestrial | Temperate |  | x | (227) |
| 449 | 2000 | 2009 | 10 | Mammals | Terrestrial | Temperate |  | x | (228) |
| 464 | 2007 | 2011 | 5 | Plants | Terrestrial | Temperate |  | x | (229-231) |
| 465 | 2007 | 2012 | 6 | Plants | Terrestrial | Temperate |  | x | (229-231) |
| 466 | 1986 | 2008 | 23 | Fish | Marine | Temperate |  | x | (232) |
| 469 | 1985 | 2011 | 27 | Fish | Marine | Temperate |  | x | (233) |
| 471 | 2004 | 2013 | 10 | Plants | Terrestrial | Temperate |  | x | (234) |
| 475 | 1960 | 1972 | 12 | Birds | Terrestrial | Temperate |  | x | (235) |
| 477 | 1998 | 2012 | 15 | Invertebrates | Marine | Temperate | x |  | $(236,237)$ |
| 501 | 2004 | 2012 | 9 | Fish | Marine | Temperate |  | x | (238) |
| 502 | 1962 | 2009 | 7 | Plants | Terrestrial | Temperate | x | x | (239) |


| 504 | 2003 | 2007 | 5 | Fish | Marine | Temperate |  | x | (240) |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 505 | 1990 | 2012 | 11 | Fish | Marine | Temperate |  | x | (241) |
| 511 | 2005 | 2015 | 6 | Fish | Marine | Tropical | x | x | (242) |
| 516 | 1997 | 2013 | 5 | Mammals | Terrestrial | Tropical |  | x | (243-247) |
| 521 | 1972 | 2017 | 37 | Mammals | Terrestrial | Temperate |  | x | (248)* |
| 522 | 1987 | 2013 | 20 | Birds | Terrestrial | Temperate |  | x | (249)* |
| 523 | 1992 | 2007 | 16 | Birds | Terrestrial | Temperate |  | x | (250)* |
| $\dagger 524$ | 1993 | 1998 | 6 | Birds | Terrestrial | Tropical |  | x | (251)* |
| †524 | 1993 | 1998 | 6 | Mammals | Terrestrial | Tropical |  | x |  |
| 526 | 2006 | 2013 | 8 | Fish | Marine | Temperate |  | x | (252)* |
| 527 | 2006 | 2019 | 14 | Birds | Marine | Polar/Temperate |  | x | (253)* |
| 547 | 2007 | 2011 | 5 | Plants | Terrestrial | Temperate |  | x | (229-231) |
| 548 | 2007 | 2012 | 6 | Plants | Terrestrial | Temperate |  | x | (229-231) |
| 549 | 2011 | 2015 | 5 | Fish | Freshwater | Tropical | x | x | (254, 255)* |
| 550 | 2005 | 2021 | 17 | Fish | Freshwater | Tropical | x | x | (250)* |
| 551 | 1999 | 2018 | 19 | Fish | Freshwater | Tropical |  | x | (257)* |
| 551 | 1999 | 2018 | 20 | Fish | Freshwater | Tropical | x |  |  |
| 552 | 1963 | 2010 | 7 | Invertebrates | Terrestrial | Temperate | x |  | $(258,259)^{*}$ |
| 602 | 1965 | 2002 | 7 | Plants | Terrestrial | Temperate | x | x | (150-156)* |
| 3541064 | 2003 | 2015 | 13 | Plants | Marine | Temperate/Tropical | x |  | (260) |
| 3541065 | 2008 | 2014 | 7 | Plants | Marine | Temperate/Tropical | x |  | (260) |

Note: $\dagger$ These studies were classified as 'multiple taxa' in the original data sources; $\dagger \dagger$ These studies were classified as 'benthos' in the original data sources. Approximately $83 \%$ of these studies are publicly available in the published BioTIME Database (17), those remaining are publicly available elsewhere and are indicated by an asterisk.

