

# Dopamine D1/D5 Receptors in the Retrosplenial Cortex Are Necessary to Consolidate Object Recognition Memory

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The retrosplenial cortex (RSC) has been widely related to spatial and contextual memory. However, we recently demonstrated that the anterior part of the RSC (aRSC) is required for object recognition (OR) memory consolidation. In this study, we aimed to analyze the requirement of dopaminergic inputs into the aRSC for OR memory consolidation in male rats. We observed amnesia at 24-h long-term memory when we infused SCH23390, a D1/D5 dopamine receptors antagonist, into aRSC immediately after OR training session. However, the same infusion had no effect on OR short-term memory. Then, we analyzed whether the ventral tegmental area (VTA) is necessary for OR consolidation. VTA inactivation by intra-VTA administration of muscimol, a GABA<sub>A</sub> agonist, immediately after an OR training session induced amnesia when animals were tested at 24 h. Moreover, we observed that this VTA inactivation-induced amnesia was reversed by the simultaneous intra-aRSC delivery of SKF38393, a D1/D5 receptor agonist. Altogether, our results suggest that VTA dopaminergic inputs to aRSC play an important modulatory role in OR memory consolidation.

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# INTRODUCTION

Recognition memory refers to the recall and awareness of a familiar event, individual, item, or place, allowing animals to discriminate between novel and familiar stimuli. In particular, the object recognition (OR) task has been widely used for studying the "what" component of recognition memory. The anterior retrosplenial cortex (aRSC) was recently observed to participate in OR memory consolidation (de Landeta et al., 2020), i.e., the storage of the "what" component of recognition the OR memory consolidation process.

Understanding the mechanisms involved in memory consolidation is a main topic in memory research, which is relevant to better understand some memory disorders and to analyze molecular targets related to those disorders. In particular, exposure to novel stimuli induces dopamine release from the ventral tegmental area (VTA) into the hippocampus to form long-term memory (LTM) (Lisman and Grace, 2005). Moreover, dopamine is known to regulate OR memory in the prefrontal and perirhinal cortices (Nagai et al., 2007; Balderas et al., 2013; De Bundel et al., 2013; Rossato et al., 2013). Thus, dopamine is a strong candidate for modulating OR memory consolidation in the RSC.

1

In this regard, the RSC receives dopaminergic projections from the VTA (Berger et al., 1985; Oades and Halliday, 1987), a structure that consists mainly of dopaminergic neurons (Morales and Margolis, 2017) and that was observed to be necessary for OR memory consolidation (Rossato et al., 2013). In addition, the RSC expresses D1/D5 receptors (Diop et al., 1988) and D1/D5 activity in the aRSC is necessary and sufficient to form a long-lasting aversive memory (Katche et al., 2013). In this scenario, we hypothesized that dopaminergic inputs from VTA to aRSC are essential for OR memory consolidation. Here, we combined pharmacological and behavioral approaches to assess the role of the dopaminergic tone in the aRSC during OR memory consolidation.

### **METHODS**

#### **Subjects**

We used a total of 92 2.5-month-old male Wistar rats (Instituto de Biología Celular y Neurociencia, CONICET-UBA) weighing about 220–300 g. Animals were housed in groups of three per cage and maintained under a 12 h light/dark cycle (lights on at 7:00 a.m.) at 21–23°C with water and food *ad libitum*. Experimental procedures followed the guidelines of the US National Institutes of Health (NIH) Guide for the Care and Use of Laboratory Animals and were approved by the Animal Care and Use Committee at the University of Buenos Aires (CICUAL).

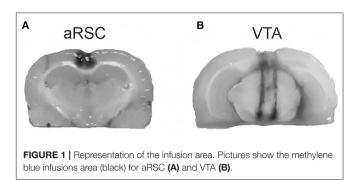
#### Surgery

Rats were implanted bilaterally under deep ketamine/xylazine anesthesia (40 and 2 mg/kg, respectively) with a 1-cm 22 G guide cannula in the aRSC at AP -3.9, L  $\pm 0.5$ , DV -1.8, and VTA at AP -5.3, L  $\pm 1.0$ , and DV -7.2, coordinates in mm from Bregma according to the atlas of Paxinos and Watson (Paxinos and Watson, 2007). Cannulas were fixed to the skull with dental acrylic. Obturators were then inserted into the cannula to prevent blockage. After 4 or 5 days of recovery from surgery, the animals were handled gently once a day for 2 days and then trained in the OR task.

### **Drug Infusion**

To study the dopaminergic input we infused into the aRSC, the D1/D5 dopamine receptor antagonist SCH23390 hydrochloride (Sigma Aldrich, Germany) and the agonist SKF38393 hydrochloride (Sigma Aldrich, Germany) at a dose of 0.75  $\mu$ g per side and 12.5  $\mu$ g per side, respectively. We infused the GABA<sub>A</sub> receptor agonist muscimol (Sigma Aldrich, Germany) at a dose of 0.1  $\mu$ g per side into the VTA immediately after the training session to study memory consolidation.

All drugs except SKF38393 were dissolved in sterile saline; SKF38393 was dissolved in 10% DMSO and sterile saline. Solutions used for dissolving the drugs were infused in the control group of the experiments (Vehicle, Veh). For all drugs infused, the entire infusion procedure took around 4 min, and the infusion rate was 1  $\mu$ l/min. Infusions into the aRSC were 1  $\mu$ l/side, while those in the VTA were of 0.5  $\mu$ l/side. Injector needles were 0.1 and 0.15 cm longer than the cannula for aRSC and VTA, respectively. Injectors were left in place for an



additional minute following infusion before they were removed carefully to avoid backflow.

### **Cannula Placement**

Cannula placement was verified after the end of the behavioral procedures by infusions of 1  $\mu$ l into the aRSC (**Figure 1A**) or 0.5  $\mu$ l into the VTA (**Figure 1B**) of 4% methylene blue in saline. A histological examination of cannula placements was performed. Only the behavioral data from animals with the cannula located in the intended site were included in the final analysis (20 animals were excluded from the analysis).

## Y-Shape Object Recognition

We performed the OR task as previously described (de Landeta et al., 2020, 2021). In brief, we habituated the animals to the empty Y-maze for 10 min, and the following day we trained the animals with two identical objects for 5 min. We then test memory 3 or 24 h after training; during the test session, we let the animals explore one object from the training session (familiar object) and one novel object for 3 min. The novel object or its position were selected by chance and were counterbalanced between animals. Objects were made of glass, metal, or plastic. The objects and apparatus were cleaned with a solution of soap, alcohol, and water before being presented to each animal.

In both training and test sessions, we used manual timers to score the time, the rodent spent exploring the objects (sniffing or touching while sniffing or facing the object). We calculated the novel object discrimination index as the exploration time of the novel object minus the exploration time of the familiar object divided by the total exploration time. Indexes significantly greater than zero were indicators of memory. We analyzed data from animals that had a minimum exploration time of 15 s/per object during the training session showing no preference for any of the sampled objects (<65% of preference for one object during training session) and that explored more than 15 s during the test (nine animals were excluded from the analysis). Total exploration times for each experiment and manipulation are shown in **Table 1**.

# **Data Analysis**

As we used a between-subjects design for our experiments, behavioral data were analyzed using the unpaired Studen's t-test between groups or the theoretical value 0 and the two-way

TABLE 1   Total training and test sessions' exploration times for each
manipulation.

Figure	Group	Training		Test		dF
		Expl time (s)	p-value	Expl time (s)	p-value	
2A			0.77		0.89	12
	Veh	$63.1 \pm 17.2$		$42.7\pm17.2$		
	SCH	$65.6\pm16.0$		$41.4\pm12.2$		
2B			0.35		0.05	13
	Veh	$73.5\pm16.6$		$44.2\pm15.2$		
	SCH	$84.7\pm28.0$		$27.7\pm14.6$		
ЗA			0.12		0.16	19
	Veh	$71.6\pm12.9$		$41.3\pm15.8$		
	Mus	$83.0\pm19.5$		$32.2\pm11.0$		
3B			0.27		0.42	37
	Veh-Veh	$100.4\pm19.5$		$40.4\pm16.5$		
	Veh-Musc	$95.7\pm12.4$		$35.3\pm16.8$		
	SKF-Veh	$98.3\pm23.9$		$47.5\pm18.6$		
	SKF-Musc	$83.9\pm24.3$		$39.0\pm13.6$		

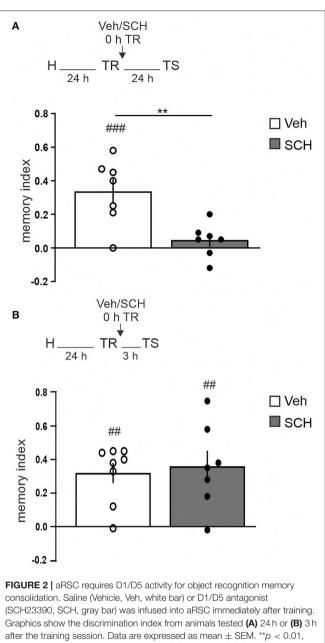
Mean  $\pm$  SD exploration time for each experiment during training and test sessions. Results of two-tailed Student's t-test or ANOVA for the exploration time in each experiment.

ANOVA. We checked the normality of data using the Shapiro–Wilk test. We used Graph Pad Prism 8 (Graphpad, USA) for statistical analysis. For all analyses, the  $\alpha$  level was set at 0.05 and the statistical power at 90% (G\*Power, Universität Düsseldorf). All data are presented as mean  $\pm$  SEM.

### RESULTS

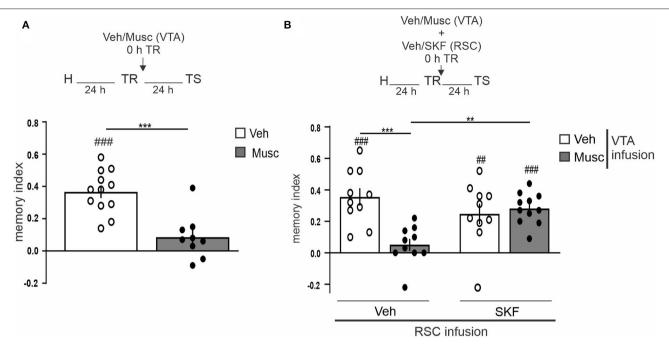
To analyze the requirement of the dopaminergic input for OR memory consolidation in the aRSC, we infused SCH23390 (0.75 µg/side, D1/D5 receptors antagonist) into the aRSC immediately after the training session and tested 3h after for short-term memory (STM) or 24h for LTM. We observed a clear-cut amnesia at 24 h in animals infused with SCH23390, while the control group had intact memory (Figure 2A, Studen's t-test. SCH vs. Vehicle: p = 0.0042, t = 3.522, df = 12. SCH vs. 0: p =0.2842, t = 1.176, df = 6. Veh vs. 0: p = 0.0039, t = 4.549, df = 6.  $n_{\rm SCH} = 7$ ,  $n_{\rm Veh} = 7$ ). However, we did not observe differences in the exploration pattern between control and SCH-infused animals when testing STM, showing both groups preference for the novel object (Figure 2B, Studen's *t*-test. Veh vs. SCH: p =0.7419, t = 0.3365, df = 13. SCH vs. 0: p = 0.0097, t = 3.734, df = 6. Veh vs. 0: p = 0.0012, t = 5.261, df = 7.  $n_{SCH} =$ 7,  $n_{\text{Veh}} = 8$ ). These results show that blocking dopaminergic signaling in the aRSC prevents OR memory consolidation but not initial formation.

Next, we decided to study the possible involvement of the VTA in the dopaminergic modulation of OR LTM in aRSC. We transiently inactivated the VTA by infusing muscimol (0.1  $\mu$ g/side, GABA<sub>A</sub> agonist) immediately after the training session and tested 24 h LTM. The VTA-inactivated group did not show memory, while the control group showed preference



Veh vs. SCH, two-tailed Student's *t*-test; ##p < 0.01, ###p < 0.001, Group vs. 0, two-tailed Student's *t*-test. (A) n = 7, (B) n = 7-8.

for the novel object (**Figure 3A**, Studen's *t*-test. p = 0.0001, t = 4.722, df = 19; Musc vs. Veh: p = 0.0996, t = 1.862, df = 8; Musc vs. 0: p < 0.0001, t = 9.558, df = 11; Veh vs. 0:  $n_{\text{Musc}} = 9$ ,  $n_{\text{Veh}} = 12$ ). This result indicates that VTA is required for OR memory consolidation, and it is consistent with previous results using another OR task (Rossato et al., 2013). Thus, we then studied whether this amnesia could be prevented by mimicking the dopamine input in the aRSC. We observed that the co-infusion of SKF38393 (12.5 µg/side, D1/D5 receptors agonist) into the aRSC immediately after the training session reversed the amnesic effect of muscimol-induced VTA



**FIGURE 3** VTA dopaminergic input to aRSC is required for object recognition memory consolidation. (A) Saline (Vehicle, Veh, white bar) or GABA<sub>A</sub> agonist (Muscimol, Musc, gray bar) infusions were made into VTA immediately after the training session; animals were tested 24 h after the training. (B) simultaneously, saline (Vehicle, Veh, white bar) or GABA<sub>A</sub> agonist (Muscimol, Musc, gray bar) was infused into VTA and 10% DMSO (Vehicle, Veh) or D1/D5 agonist (SKF38393, SKF) was infused into aRSC immediately after the training. Test session was performed 24 h after training. Data are expressed as mean discrimination index  $\pm$  SEM. (A) \*\*\*p < 0.001, Veh vs. Musc, two-tailed Student's *t*-test. (B) \*\*\*p < 0.001, \*\*p < 0.01, Tukey's multiple comparison test after two-way ANOVA (only biologically relevant comparisons are shown). (A,B) ###p < 0.001, ##p < 0.01, Group vs. 0, two-tailed Student's *t*-test. (A) n = 9-12, (B) n = 10-11.

inactivation (**Figure 3B**, p<sub>interaction</sub> = 0.0012,  $F_{\text{interaction}} = 12.23$ , Tukey's multiple comparisons test after two-way ANOVA<sub>(1,37)</sub> factors: infusion into VTA and infusion into aRSC. Studen's *t*-test: p = 0.0001, t = 6.474, df = 9; Veh–Veh vs. 0: p = 0.0044, t = 3.768, df = 9; Veh–SKF vs. 0: p = 0.2177, t = 1.325, df = 9; Musc–Veh vs. 0: p < 0.0001, t = 9.453, df = 10; Musc–SKF vs. 0:  $n_{Veh-Veh} = 10$ ,  $n_{Veh-SKF} = 10$ ,  $n_{Musc-Veh} = 10$ ,  $n_{Musc-SKF} = 11$ ). This result suggests that dopamine from VTA is not only necessary but also sufficient for OR memory consolidation in aRSC.

# DISCUSSION

Our results suggest that the dopaminergic input from the VTA to the aRSC is necessary for modulating long-term OR memory consolidation. The results shown here are in line with others that showed the modulation of the dopaminergic system in OR memory by observing the enhancement of LTM when using systemic injections of dopamine D1/D5 receptor agonist SKF38393 (de Lima et al., 2011) or inhibiting the catechol-O-methyltransferase (Detrait et al., 2016). Moreover, infusion of the D1/D5 antagonist SCH23390 into the perirhinal cortex (Balderas et al., 2013), hippocampus (De Bundel et al., 2013; Furini et al., 2014; Neves et al., 2020, but see Rossato et al., 2013), amygdala (Rossato et al., 2013), or prefrontal cortex (Nagai et al., 2007; De Bundel et al., 2013; Rossato et al., 2013) produced 24 h OR amnesia, like our result when infusing SCH23390 into

the aRSC. In addition, blocking dopamine reuptake in the insular cortex of an Alzheimer's disease mice model reversed the STM and LTM object amnesia in those mice (Guzmán-Ramos et al., 2012). Moreover, hippocampal dopaminergic tone is essential for object memory persistence (Neves et al., 2020; Vargas et al., 2020; Lima et al., 2022) and reconsolidation (Rossato et al., 2015; Gonzalez et al., 2021).

On the contrary, SCH23390 failed to disrupt STM formation in the medial prefrontal cortex, perirhinal cortex, and hippocampus (Savalli et al., 2015). Despite this, another study showed that the inhibition of dopaminergic activity by SCH23390 systemic administration or its infusion into the prelimbic cortex impaired OR STM (Clausen et al., 2011). Inconsistency between the results shown in these studies could be related to methodological differences, such as drug concentration and the strain of rats used. In particular, we did not find an effect of SCH23390 infusion into aRSC when testing STM. We suggest that the discrepancy between Clausen's study and ours might be due to differences in the infusion time points and in the neocortical area analyzed. In our study, we prefer to infuse SCH23390 immediately after training rather than before training; in this way, we could check whether the effect of SCH23390 on LTM was due to dopamine requirements for memory consolidation (i.e., memory stabilization) rather than deficits in acquisition or initial formation.

Although our study and others showed OR LTM impairment by SCH23390, we cannot exclude that part of this effect might be due to SCH23390 agonist activity on serotonin  $5\text{-HT}_{2C}$ 

receptors (Millan et al., 2001). Nevertheless, it was observed that i.p. administration of a 5-HT<sub>2C</sub> agonist, compound (+)-22a, improved OR LTM in a schizophrenia model (NR1-KD mice) (Cheng et al., 2016). Also, i.p. administration of the 5-HT<sub>2C</sub> antagonist, RO 60-0491, enables OR LTM formation in animals that do not show LTM (Pitsikas and Sakellaridis, 2005), and the non-specific 5-HT<sub>2C</sub> antagonist, agomelatine, improved the OR memory of stressed mice (Gumuslu et al., 2014), though blocking of 5-HT<sub>2C</sub> receptors reinforces frontocortical dopaminergic transmission (Millan et al., 2003). Thus, 5-HT<sub>2C</sub> receptor activity could be related to OR memory formation improvement. This strengthens that our results are related to SCH23390 activity over D1/D5 receptors.

The requirement of VTA for OR memory consolidation observed in this study is similar to that previously shown in another OR task (Rossato et al., 2013). In addition, our results showed that mimicking dopamine input by the simultaneous infusion of a D1/D5 agonist into the aRSC prevented the amnesia produced by VTA inactivation. Likewise, D1/D5 activity in the medial prefrontal cortex and amygdala together, but not each structure alone, could prevent the effect of VTA inactivation (Rossato et al., 2013). The main difference between Rossato's work and ours is that we observed that local SKF38393 only in the aRSC prevents the VTA inactivation effect, suggesting that the aRSC is a prime structure for OR processing. Considering its functional connectivity with many brain regions of the OR network (de Landeta et al., 2021), we suggest that aRSC could be relevant for receiving and sending information about different features of the objects, orchestrating object memory consolidation. However, when the aRSC is not properly functioning during memory acquisition, this role might be taken over by other brain structures (de Landeta et al., 2020).

Our results showed for the first time that dopamine is required in the aRSC for OR memory consolidation; we demonstrated that dopamine is both necessary and sufficient to consolidate OR memory in the aRSC. These results also suggest the involvement

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of VTA inputs to the OR memory network for the proper memory consolidation. Considering VTA cellular diversity and the existence of neurons that co-release dopamine and either GABA or glutamate (Morales and Margolis, 2017), we cannot conclude about the nature of VTA inputs into the aRSC and their effect on OR memory. To consolidate the link between the effect of VTA transient inactivation and D1/D5 signaling in the aRSC, further experiments are needed to selectively manipulate the VTA dopaminergic neurons projecting to the aRSC.

# DATA AVAILABILITY STATEMENT

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation.

## **ETHICS STATEMENT**

The animal study was reviewed and approved by Animal Care and Use Committee of the University Buenos Aires (CICUAL). School of Medicine, University of Buenos Aires.

# **AUTHOR CONTRIBUTIONS**

ABL, JHM, and CK designed the experiments and wrote the manuscript. CK led the research. ABL performed the experiments and analyzed the data. All authors contributed to the article and approved the submitted version.

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