

Annotated key to the genera of Mymaridae (Hymenoptera: Chalcidoidea) in Argentina

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Abstract

A key to the genera of Mymaridae occurring in Argentina is given, and information on the 178 determined and several undetermined species in 35 genera is provided, including data on their distribution and host associations. New host records are provided for several species of *Anagrus* Haliday and *Gonatocerus* Nees. The genus *Kikiki* Huber & Beardsley is recorded for the first time from the New World, and the genera *Australomymar* Girault, *Camptopteroides* Viggiani, *Gahanopsis* Ogloblin, *Myrmecomymar* Yoshimoto, and *Ptilomymar* Annecke & Doutt are reported for the first time from Argentina. *Eucleruchus* Ogloblin, **syn. nov.** is synonymized under *Cleruchus* Enock, and its type species *E. neivai* Ogloblin is transferred to *Cleruchus* as *C. neivai* (Ogloblin), **comb. nov.**

Key words: Argentina, Mymaridae, fairyflies, egg parasitoid

Introduction

The Mymaridae (fairyflies or mymarids) is a family of minute wasps containing more than 1400 species in 103 genera (Noyes 2005). Mymarids range from 0.2 to about 4.0 mm (average 0.5–1.0 mm) in length. Most mymarids can be easily distinguished from other members of Chalcidoidea by the following combination of characters: head with trabeculae and associated sutures on vertex and on face lateral to each torulus; toruli generally closer to the eyes than to each other; forewing marginal vein with a hypochaeta; venation usually greatly reduced, terminating within the first two-fifths of wing (except in *Arescon* Walker, *Australomymar* Girault, *Borneomymar* Huber, *Eustochomorpha* Girault, *Kikiki* Huber & Beardsley, *Krokella* Huber, and *Myrmecomymar* Yoshimoto) and without postmarginal vein (except in *Borneomymar* and *Eustochomorpha*); hind wing narrow and stalked basally (except membrane extending to the base of the hind wing in *Anagroidea* Girault, *Australomymar*, and *Eubroncus* Yoshimoto, Kozlov & Trjapitzin); scutellum often subdivided into anterior and posterior parts; antenna at least as long as head plus mesosoma, usually much longer, distinctly clavate in females and filiform in males.

Mymaridae is cosmopolitan and species are found in all terrestrial habitats, from the deserts to rainforests, grasslands, ponds, and in a wide altitudinal range (Huber 1997, 2006; Noyes 2005). They are internal parasitoids of insect eggs; apparently most of them are solitary egg parasitoids but there are gregarious species. The primary hosts are Coleoptera (especially Curculionidae, Chrysomelidae, and Dytiscidae), Hemiptera (Cicadellidae, Delphacidae, Cercopidae, Miridae, Membracidae, Tingidae, etc.), Psocoptera, Odonata (Anisozygoptera and Zygoptera), and some Orthoptera. Because some of their hosts are agricultural or forest pests, mymarids are considered beneficial insects, and several species have been used successfully in classical biological control programs against insect pests (Clausen 1940). Huber (1986) reviewed the history,

systematics, biology, and hosts of Mymaridae. Recently Huber *et al.* (2006) recorded *Stethynium* Enock species from Australia as parasitoids of larvae of gall inducing Eulophidae (Hymenoptera).

The following publications among others refer to the biology and host associations of the mymarid wasps: MacGill (1934), Sahad (1982a, 1982b), Chantarasa-ard (1984), Chantarasa-ard *et al.* (1984a, 1984b), Sahad (1984), Meyerdirk & Moratorio (1987a, 1987b), Cronin & Strong (1990, 1993), Moratorio (1990), Settle & Wilson (1990), Moratorio & Chiappini (1995), Al-Wahaibi & Walker (2000a, 2000b), Triapitsyn *et al.* (2002, 2003), Irvin & Hoddle (2005a, 2005b), Chen *et al.* (2006), Pilkington & Hoddle (2006). Regarding the Neotropical species, information on the bionomics of Mymaridae is scarce, most of it published during the last decade (Vilas Boas & Andrade 1990; Pires *et al.* 1993; Gladstone *et al.* 1994; Oliveira & Spotti Lopes 2000; Virla 2001, 2004; Logarzo *et al.* 2004; Liljesthröm & Virla 2004; Virla *et al.* 2005; Triapitsyn, Vickerman *et al.* 2006; Luft Albarracin *et al.* 2006).

Mymarids could be considered uncommon insects, but if the correct collecting methods are used, they are among the most abundant groups of Hymenoptera. They may be collected by sweeping (preferably total), but using several trapping methods (suction traps, fine mesh Malaise traps, flight interception traps, pitfall traps, and yellow pan traps) usually is more effective. The samples collected should be sorted under a low power microscope to extract the specimens, especially the smallest ones. Another collecting method is employment of sentinel eggs of known hosts in natural and agricultural environments, thus being particularly helpful for revealing host associations of the parasitoids and also for obtaining data for biological control surveys.

In the Neotropics, the mymarid fauna has been little studied, with only about 280 species in 48 genera known so far. According to Noyes (2005), Argentina is the country with the highest number of recorded taxa with 166 species (most of them described during 1930s–1960s by A.A. Ogloblin), followed by Brazil (39), Ecuador (25), Chile (18), Venezuela (12), Costa Rica (12), Peru (11), and less than 10 reported species from other countries, but that of course does not represent the true diversity. Despite the numerous taxonomic studies on this family, most of them referred to a single or few species so the information is very fragmentary. The more important recent taxonomic publications on the Neotropical fauna of Mymaridae are of Yoshimoto (1990), Triapitsyn (2002), and Triapitsyn, Berezovskiy *et al.* (2007).

Most of the contributions on the mymarids of Argentina were done by six taxonomists (and their collaborators): Brèthes (1913, 1922), Ogloblin (1934, 1935a, 1935b, 1935c, 1936, 1938a, 1938b, 1938c, 1939a, 1939b, 1940, 1947, 1949, 1953, 1955a, 1955b, 1955c, 1957a, 1957b, 1959a, 1959b, 1960a, 1960b, 1962, 1963, 1964, 1967), De Santis (1967, 1979, 1981, 1989), De Santis & Fidalgo (1994), Fidalgo (1982, 1988, 1989, 1991a, 1991b), Triapitsyn (1997, 1999, 2002, 2008), Triapitsyn & Berezovskiy (2002, 2004, 2007), Triapitsyn & Virla (2004), Triapitsyn, Berezovskiy *et al.* (2006, 2007), Triapitsyn, Logarzo *et al.* (2007), Triapitsyn & Aquino (2008), and Triapitsyn *et al.* (2008).

Although the mymarid fauna of Argentina is among the better studied in South America, its current state of knowledge still can be considered insufficient and fragmentary. Therefore, the aim of this study was to summarize existing knowledge of this family in Argentina by assembling all the known information about them and adding new distributional records and/or host associations. Another objective of this study is to provide an illustrated key to the genera of Mymaridae that occur in Argentina.

Material and methods

The information was obtained mainly from publications, a partial study of the large A.A. Ogloblin collection of Mymaridae (all on slides) at the Museo de Ciencias Naturales de La Plata in La Plata, Buenos Aires, Argentina, and our own research. In the latter case, material was collected from various localities in several provinces of Argentina. Most of the specimens were collected using sentinel eggs of Cicadellidae on leaves of potted citrus or/and corn plants [for references regarding methodology see Luft Albarracin & Triapitsyn (2007)]. Other collecting methods were used as well, e.g., Malaise traps, yellow pan traps, and sweeping with a net.

Specimens were preserved in 70% ethanol, then dried using a critical point drier and point- or card-mounted, and later some of them slide-mounted in Canada balsam. Terms for morphological features are those of Gibson (1997). Voucher specimens are deposited in the collections of Fundación e Instituto Miguel Lillo, San Miguel de Tucumán, Tucumán, Argentina (IMLA), Museo de Ciencias Naturales de La Plata, La Plata, Buenos Aires, Argentina (MLPA), and Entomology Research Museum, University of California, Riverside, California, USA (UCRC).

Results

There are 178 determined and several undetermined species classified in 35 genera in Argentina (Table 1). The most speciose genera are: *Gonatocerus* Nees (45 determined species), *Acmopolynema* Ogloblin (18), *Erythmelus* Enoch (17), *Agalmopolynema* Ogloblin (16), and *Anagrus* Haliday (12). The distributional ranges of 13 species were established at the provincial level. In addition, new host associations are provided for six species of *Anagrus* and *Gonatocerus*.

Eucleruchus Ogloblin, **syn. nov.**, is synonymized here under *Cleruchus* Enoch, and its type species *E. neivai* Ogloblin [holotype female in MLPA, examined] is transferred to *Cleruchus* as *C. neivai* (Ogloblin), **comb. nov.** Presently, *Cleruchus* is too narrowly defined, and *Eucleruchus* was defined by Ogloblin (1940) based on minor species (or at most species-group) characters, which are well within the broad range of variability within *Cleruchus*.

TABLE 1. List of genera and species of Mymaridae in Argentina. Provincial acronyms: BA = Buenos Aires, CA = Catamarca, CHA = Chaco, CHU = Chubut, C = Córdoba, CO = Corrientes, ER = Entre Ríos, F = Formosa, J = Jujuy, LP = La Pampa, LR = La Rioja, M = Misiones, ME = Mendoza, N = Neuquén, S = Salta, SE = Santiago del Estero, SF = Santa Fe, SJ = San Juan, RN = Río Negro, T = Tucumán, TF = Tierra del Fuego and South Atlantic Islands. In the first column (under "Taxon"), references are provided only for the species originally described from Argentina.

Taxon	Distribution	Reference(s)	Hosts in Argentina
<i>Acmopolymena</i> Ogloblin			
<i>aberrans</i> Fidalgo 1989: 10	BA	Fidalgo (1989); De Santis & Fidalgo (1994)	-----
<i>callopterum</i> Fidalgo 1989: 12	BA, M	Fidalgo (1989); De Santis & Fidalgo (1994)	-----
<i>carinatum</i> Fidalgo 1989: 14	M	Fidalgo (1989); De Santis & Fidalgo (1994)	-----
<i>commune</i> Fidalgo 1989: 15	M, S, T	Fidalgo (1989); De Santis & Fidalgo (1994)	-----
<i>delphacivorum</i> Fidalgo 1989: 19	M, S	Fidalgo (1989); De Santis & Fidalgo (1994)	-----
<i>gracilicorne</i> Fidalgo 1989: 21	M	Fidalgo (1989); De Santis & Fidalgo (1994)	-----
<i>inaequale</i> Fidalgo 1989: 24	S	Fidalgo (1989); De Santis & Fidalgo (1994)	-----
<i>infuscatum</i> Fidalgo 1989: 25	T	Fidalgo (1989); De Santis & Fidalgo (1994)	-----
<i>kronidiphagum</i> Fidalgo 1989: 26	ER, M	Fidalgo (1989); De Santis & Fidalgo (1994)	<i>Kronides incumbens</i> (Germar) (Membracidae)

to be continued.

TABLE 1. (continued)

Taxon	Distribution	Reference(s)	Hosts in Argentina
<i>longicorne</i> Fidalgo 1989: 29	M	Fidalgo, 1989; De Santis & Fidalgo, 1994	-----
<i>mirabile</i> Fidalgo 1989: 30	M	Fidalgo (1989); De Santis & Fidalgo (1994)	-----
<i>missionicum</i> Fidalgo 1989: 31	M	Fidalgo (1989); De Santis & Fidalgo (1994)	-----
<i>obscuricorne</i> Fidalgo 1989: 32	LR	Fidalgo (1989); De Santis & Fidalgo (1994)	-----
<i>perterebrator</i> Fidalgo 1989: 34	M, T	Fidalgo (1989); De Santis & Fidalgo (1994)	-----
<i>poecilopterum</i> Fidalgo 1989: 35	BA	Fidalgo (1989); De Santis & Fidalgo (1994)	-----
<i>polyrrhiza</i> Fidalgo 1989: 37	M	Fidalgo (1989); De Santis & Fidalgo (1994)	-----
<i>scapulare</i> Fidalgo 1989: 38	M	Fidalgo (1989); De Santis & Fidalgo (1994)	-----
<i>uma</i> Schauff	BA, J, M, S, T	Fidalgo (1989); De Santis & Fidalgo (1994)	-----
<i>Agalmopolynema</i> Ogloblin			
<i>australe</i> Fidalgo 1988: 47	N	Fidalgo (1988); De Santis & Fidalgo (1994)	-----
<i>ayra</i> Fidalgo 1988: 49	N	Fidalgo (1988); De Santis & Fidalgo (1994)	-----
<i>aza</i> Fidalgo 1988: 49	N	Fidalgo (1988); De Santis & Fidalgo (1994)	-----
<i>bicoloricorne</i> Fidalgo 1988: 51	N, RN	Fidalgo (1988); De Santis & Fidalgo (1994); Loíacono <i>et al.</i> (2005)	-----
<i>calyptera</i> Fidalgo 1988: 53	N, R	Fidalgo (1988); De Santis & Fidalgo (1994)	-----
<i>caudatum</i> Fidalgo 1988: 55	N	Fidalgo (1988); De Santis & Fidalgo (1994)	-----
<i>chusqueanum</i> Fidalgo 1988: 56	N	Fidalgo (1988); De Santis & Fidalgo (1994)	-----
<i>denticulatum</i> Fidalgo 1988: 56	N	Fidalgo (1988); De Santis & Fidalgo (1994)	-----
<i>longisetosum</i> Fidalgo 1988: 57	N	Fidalgo (1988); De Santis & Fidalgo (1994)	-----
<i>mirabile</i> Fidalgo 1988: 58	N, RN	Fidalgo (1988); De Santis & Fidalgo (1994)	-----
<i>nubeculatum</i> Fidalgo 1988: 59	N	Fidalgo (1988); De Santis & Fidalgo (1994)	-----

to be continued.

TABLE 1. (continued)

Taxon	Distribution	Reference(s)	Hosts in Argentina
<i>nantuense</i> Fidalgo 1988: 59	N	Fidalgo (1988); De Santis & Fidalgo (1994)	-----
<i>ogloblini</i> Fidalgo 1988: 60	N	Fidalgo (1988); De Santis & Fidalgo (1994)	-----
<i>rufithorax</i> Fidalgo 1988: 61	N	Fidalgo (1988); De Santis & Fidalgo (1994)	-----
<i>shajovskoi</i> Fidalgo 1988: 62	N	Fidalgo (1988); De Santis & Fidalgo (1994)	-----
<i>succineum</i> (Ogloblin 1960a: 2)	N	Ogloblin (1960a); Fidalgo (1988)	-----
<i>Alaptus</i> Westwood			
<i>caecilii</i> Girault	BA, M	De Santis (1979)	-----
<i>Anagrus</i> Haliday			
<i>atomus</i> (Linnaeus)	ME	Triapitsyn (1999, 2002)	-----
<i>breviphragma</i> Soyka	BA, C, CHA ⁽⁺⁾ , J ⁽⁺⁾ , M, S, SE ⁽⁺⁾ , T	Triapitsyn (1997, 1999, 2002)	<i>Agalliana ensigera</i> Oman ⁽⁺⁾ , <i>Chlorotettix fraterculus</i> (Berg) ⁽⁺⁾ , <i>Ciminius platensis</i> (Berg) ⁽⁺⁾ , <i>Dalbulus maidis</i> (DeLong & Wolcott), <i>Dechacona missionum</i> (Berg) ⁽⁺⁾ , <i>Exitianus obscurinervis</i> (Stål) ⁽⁺⁾ , <i>Hortensia similis</i> (Walker) ⁽⁺⁾ , <i>Xerophloea viridis</i> (Fabricius) ⁽⁺⁾ (Cicadellidae); <i>Delphacodes kuscheli</i> Fennah (Delphacidae)
<i>empoascae</i> Dozier	M	Triapitsyn (1997, 1999, 2002)	-----
<i>flaveolus</i> Waterhouse	BA, C, J ⁽⁺⁾ , M, ME ⁽⁺⁾ , N ⁽⁺⁾ , S ⁽⁺⁾ , SE ⁽⁺⁾ , T	Triapitsyn (1997, 1999, 2002); Virla (2004)	<i>Amplicephalus simpliciusculus</i> Linnavuori, <i>Dalbulus maidis</i> (DeLong & Wolcott), <i>Exitianus obscurinervis</i> (Stål) (Cicadellidae); <i>Delphacodes kuscheli</i> Fennah, <i>Peregrinus maidis</i> (Ashmead), <i>Saccharosydne</i> sp. ⁽⁺⁾ , <i>Toya propinqua</i> (Fieber) (Delphacidae)
<i>lineolus</i> Triapitsyn 1999: 217	BA, J ⁽⁺⁾ , M, T ⁽⁺⁾	Triapitsyn (1999, 2002)	<i>Chlorotettix fraterculus</i> (Berg), <i>Plesiommata mollicella</i> (Fowler) (Cicadellidae) ⁽⁺⁾
<i>miriamae</i> Triapitsyn & Virla 2004: 383	BA, M, N, SE, T	Triapitsyn & Virla (2004)	<i>Delphacodes sitarea</i> Marino de Remes Lenicov & Tesón (Delphacidae)

to be continued.

TABLE 1. (continued)

Taxon	Distribution	Reference(s)	Hosts in Argentina
<i>nigriventris</i> Girault	BA, ER, J, M, S, SF, T	Triapitsyn (1997, 1999, 2002); Luft Albarracin <i>et al.</i> (2006)	<i>Dalbulus maidis</i> (DeLong & Wolcott) (Cicadellidae)
<i>ogloblini</i> Triapitsyn 1999: 216	M	Triapitsyn (1999, 2002)	-----
<i>stethynioides</i> Triapitsyn 2002: 216	M	Triapitsyn (2002)	-----
<i>subfuscus</i> Foerster	CHU	Triapitsyn (1999, 2002)	-----
<i>ustulatus</i> Haliday	ME	Triapitsyn (1999, 2002)	-----
<i>yawi</i> Fullaway	BA, M, S	Triapitsyn (1997, 1999, 2002)	<i>Tenthecoris bicolor</i> Scott (Miridae)
Anaphes Haliday			
<i>amplipennis</i> Ogloblin 1962: 49	N	Ogloblin (1962)	-----
<i>atomarius</i> (Brèthes 1913: 100)	ER	Brèthes (1913)	-----
<i>conotracheli</i> Girault	BA	De Santis (1989)	-----
<i>neuquenensis</i> Ogloblin 1962: 54	N	Ogloblin (1962)	-----
<i>nitens</i> (Girault)	BA, ER	De Santis (1967)	<i>Gonipterus gibberus</i> Boisduval, <i>G. platensis</i> Marshall (Curculionidae)
<i>nunezi</i> Ogloblin 1962: 51	TF	Ogloblin (1962)	-----
<i>pucarobius</i> Ogloblin 1962: 52	N	Ogloblin (1962)	-----
Arescon Walker			
<i>dallasi</i> (Ogloblin 1938c: 97)	M	Ogloblin (1938c)	-----
<i>elongatus</i> (Ogloblin 1957a: 39)	M	Ogloblin (1957a)	-----
<i>maculipennis</i> (Ogloblin 1957a: 37)	M	Ogloblin (1957a)	-----
<i>platensis</i> (Ogloblin 1957a: 42)	BA	Ogloblin (1957a)	-----
<i>pusillus</i> (Ogloblin 1957a: 40)	M	Ogloblin (1957a)	-----
Australomyar Girault			
<i>A. sp(p).</i>	LR ⁽⁺⁾ , N ⁽⁺⁾	⁽⁺⁾	-----
Callodicopus Ogloblin			
<i>crassula</i> Ogloblin 1955a: 378	M	Ogloblin (1955a)	-----
<i>cursor</i> Ogloblin 1955a: 384	M	Ogloblin (1955a)	-----
<i>longicornis</i> Ogloblin 1955a: 381	M	Ogloblin (1955a)	-----
<i>silvestriana</i> Ogloblin 1955a: 383	M	Ogloblin (1955a)	-----
Camptoptera Foerster			
<i>angustipennis</i> Ogloblin 1947: 504	M	Ogloblin (1947)	-----
<i>loretoensis</i> Ogloblin 1947: 495	M	Ogloblin (1947)	-----
<i>missionica</i> Ogloblin 1947: 499	M	Ogloblin (1947)	-----

to be continued.

TABLE 1. (continued)

Taxon	Distribution	Reference(s)	Hosts in Argentina
<i>pulla</i> Girault	BA, S	De Santis (1967)	-----
<i>reticulata</i> Ogloblin 1947: 501	M	Ogloblin (1947)	-----
<i>semialbata</i> Ogloblin & Annecke 1961: 302	M	Ogloblin & Annecke (1961)	-----
<i>Camptopteroides</i> Viggiani			
<i>C. (Alalinda) sp.</i>	M ⁽⁺⁾	(+)	-----
<i>Cleruchus</i> Enock			
<i>brevipennis</i> Ogloblin 1940: 597	M	Ogloblin (1940)	-----
<i>longicornis</i> Ogloblin 1955b: 499	M	Ogloblin (1955b)	-----
<i>neivai</i> (Ogloblin 1940: 601)	M	Ogloblin (1940)	-----
<i>terebrator</i> (Ogloblin 1959a: 71)	BA	Ogloblin (1959a)	-----
<i>vagatus</i> (Ogloblin 1959a: 74)	M	Ogloblin (1959a)	-----
<i>Cnecomymar</i> Ogloblin			
<i>bimaculatus</i> Ogloblin 1963: 74	BA, M, S	Ogloblin (1963)	-----
<i>major</i> Ogloblin 1963: 65	M	Ogloblin (1963)	-----
<i>meridionalis</i> Ogloblin 1963: 67	BA	Ogloblin (1963)	-----
<i>parvulus</i> Ogloblin 1963: 73	M	Ogloblin (1963)	-----
<i>pauperatus</i> Ogloblin 1963: 71	BA	Ogloblin (1963)	-----
<i>terebrator</i> Ogloblin 1963: 77	M	Ogloblin (1963)	-----
<i>Dicopomorpha</i> Ogloblin			
<i>macrocephala</i> Ogloblin 1955a: 387	M	Ogloblin (1955a)	-----
<i>pulchricornis</i> (Ogloblin 1955a: 390)	M	Ogloblin (1955a)	-----
<i>stramineus</i> (Ogloblin 1955a: 394)	M	Ogloblin (1955a)	-----
<i>Dicopus</i> Enock			
<i>citri</i> Mercet	BA	De Santis (1979)	-----
<i>Erdosiella</i> Soyka			
<i>decorata</i> (Ogloblin 1967: 184)	M	Ogloblin (1967)	-----
<i>Erythmelus</i> Enock			
<i>angustatus</i> Ogloblin 1934: 256	M	Ogloblin (1934); Triapitsyn, Berezovskiy <i>et al.</i> (2007)	-----
<i>brachialis</i> Ogloblin 1934: 255	BA, M, S	Ogloblin (1934); Triapitsyn, Berezovskiy <i>et al.</i> (2007)	-----
<i>cingulatus</i> Ogloblin 1934: 254	CA, M	Ogloblin (1934); Triapitsyn, Berezovskiy <i>et al.</i> (2007)	-----
<i>clavatus</i> Ogloblin 1934: 247	BA, J, LR, M, ME, S	Ogloblin (1934); Triapitsyn, Berezovskiy <i>et al.</i> (2007)	-----

to be continued.

TABLE 1. (continued)

Taxon	Distribution	Reference(s)	Hosts in Argentina
<i>coviellai</i> Triapitsyn in Triapitsyn, Berezovskiy <i>et al.</i> 2007: 13	BA, M, ME, S	Triapitsyn, Berezovskiy <i>et al.</i> (2007)	<i>Engytatus</i> sp. (Miridae)
<i>fidalgoi</i> Triapitsyn in Berezovskiy <i>et al.</i> 2007: 46	LR	Triapitsyn, Berezovskiy <i>et al.</i> (2007)	-----
<i>gak</i> Triapitsyn	N	Triapitsyn, Berezovskiy <i>et al.</i> (2007); Triapitsyn (2008)	-----
<i>hirtipennis</i> Ogloblin, 1934: 250	M	Ogloblin (1934); Triapitsyn, Berezovskiy <i>et al.</i> (2007)	-----
<i>logarzoi</i> Triapitsyn in Triapitsyn, Berezovskiy <i>et al.</i> 2007: 16	BA	Triapitsyn, Berezovskiy <i>et al.</i> (2007)	-----
<i>miridiphagus</i> Dozier	M	Triapitsyn, Berezovskiy <i>et al.</i> (2007)	<i>Lygus</i> sp. (Miridae)
<i>pastoralis</i> Ogloblin 1934: 252	M	Ogloblin (1934); Triapitsyn, Berezovskiy <i>et al.</i> (2007)	<i>Lygus</i> sp. (Miridae)
<i>rex</i> (Girault)	BA, ME	Triapitsyn, Berezovskiy <i>et al.</i> (2007); Triapitsyn (2008)	-----
<i>rosascostai</i> Ogloblin 1934: 249	BA, M	Ogloblin (1934); Triapitsyn, Berezovskiy <i>et al.</i> (2007)	-----
<i>tingitiphagus</i> (Soares)	BA, F, M	Triapitsyn, Berezovskiy <i>et al.</i> (2007)	unidentified Tingidae
<i>toreador</i> Triapitsyn in Triapitsyn, Berezovskiy <i>et al.</i> 2007: 25	C, SF	Triapitsyn, Berezovskiy <i>et al.</i> (2007)	<i>Corythaica cucullata</i> (Berg), <i>Leptobyrsa</i> sp. (Tingidae)
<i>verticillatus</i> Ogloblin 1934: 245	M	Ogloblin (1934); Triapitsyn, Berezovskiy <i>et al.</i> (2007)	unidentified Miridae
<i>yuzhanin</i> Triapitsyn 2008: 67	C	Triapitsyn (2008)	? unidentified Miridae
Gahanopsis Ogloblin			
<i>G. spp.</i>	M ⁽⁺⁾ , S ⁽⁺⁾	(+)	-----
Gonatocerus Nees			
<i>abbreviatus</i> (Ogloblin 1953: 7)	BA, J ⁽⁺⁾ , M, S ⁽⁺⁾ , SE ⁽⁺⁾ , T ⁽⁺⁾	Ogloblin (1953)	<i>Plesiommata mollicella</i> (Fowler), <i>Scopogonia</i> <i>subolivacea</i> (Stål), <i>Tapajosa</i> <i>rubromarginata</i> (Signoret) (Cicadellidae) ⁽⁺⁾
<i>acanophorae</i> (Ogloblin, 1938a: 97)	M	Ogloblin (1938a)	<i>Calloconophora pugionata</i> (Germar) (Membracidae)
<i>aethalionis</i> (Ogloblin 1938a: 93)	M	Ogloblin (1938a)	<i>Aetalion reticulatum</i> (Linnaeus) (Aetalionidae)

to be continued.

TABLE 1. (continued)

Taxon	Distribution	Reference(S)	Hosts in Argentina
<i>annulicornis</i> (Ogloblin 1936: 41)	CA, CO, F, J, LR, M, ME, S, SJ, T	Ogloblin (1936); Virla <i>et al.</i> (2008)	<i>Oncometopia tucumana</i> Schröder, <i>Tapajosa rubromarginata</i> (Signoret) (Cicadellidae)
<i>anomocerus</i> Crawford	M	Ogloblin (1953)	unidentified Membracidae
<i>appendiculatus</i> (Ogloblin 1939a: 239)	M	Ogloblin (1939a)	-----
<i>atriclavus</i> Girault (as <i>G. triguttatus atriclavus</i> Girault)	J	Triapitsyn (2006)	unidentified Proconiini (Cicadellidae)
<i>blesticus</i> (Ogloblin 1957b: 39)	RN	Ogloblin (1957b)	-----
<i>bonaerensis</i> (Ogloblin 1939a: 246)	BA	Ogloblin (1939a)	-----
<i>bonariensis</i> (Brèthes 1922: 129)	BA	Brèthes (1922)	-----
<i>brachyurus</i> (Ogloblin 1938b: 32)	BA	Ogloblin (1938b)	-----
<i>carahuensis</i> (Ogloblin 1957b: 36)	N	Ogloblin (1957b)	-----
<i>caudatus</i> Ogloblin 1935c: 74	M	Ogloblin (1935c, 1953)	-----
<i>chusqueicolus</i> (Ogloblin 1957b: 33)	N, RN	Ogloblin (1957b)	-----
<i>concinus</i> (Ogloblin 1936: 46)	M	Ogloblin (1936)	-----
<i>deleoni</i> Triapitsyn, Logarzo & Virla in Triapitsyn <i>et al.</i> 2008: 6	ME	Triapitsyn <i>et al.</i> (2008)	<i>Tapajosa rubromarginata</i> (Signoret) (Cicadellidae)
<i>dimorphus</i> (Ogloblin 1938a: 101)	M	Ogloblin (1938a)	<i>Kronides incumbens</i> (Germar) (Membracidae)
<i>excisus</i> (Ogloblin 1936: 53)	M	Ogloblin (1936)	-----
<i>gracilicornis</i> (Ogloblin 1936: 50)	M	Ogloblin (1936)	-----
<i>grandis</i> (Ogloblin 1936: 38)	M	Ogloblin (1936)	-----
<i>granulosus</i> (Ogloblin 1959b: 189)	M	Ogloblin (1959b)	-----
<i>h-luteum</i> (Ogloblin 1938b: 29)	BA	Ogloblin (1938b)	-----
<i>inauditus</i> (Ogloblin 1936: 36)	M	Ogloblin (1936)	-----
<i>membraciphagus</i> Ogloblin 1935c: 65	M	Ogloblin (1935c, 1938a)	<i>Bolbonota pictipennis</i> Fairmaire (Membracidae)
<i>metanotalis</i> (Ogloblin 1938b: 35)	C ^(*) , ER ^(*) , J ⁽⁺⁾ , LR ^(*) , M, ME ^(*) , S ⁽⁺⁾ , T ⁽⁺⁾	Ogloblin (1938b); Virla <i>et al.</i> (2008)	<i>Oncometopia tucumana</i> Schröder, <i>Plesiommata mollicella</i> (Fowler) ⁽⁺⁾ , <i>Scopogonalia subolivacea</i> (Stål) ⁽⁺⁾ , <i>Tapajosa rubromarginata</i> (Signoret) (Cicadellidae)
<i>mexicanus</i> Perkins	S	Ogloblin (1955c)	-----

to be continued.

TABLE 1. (continued)

Taxon	Distribution	Reference(s)	Hosts in Argentina
<i>monrosi</i> (Ogloblin 1959b: 185)	BA	Ogloblin (1959b)	-----
<i>nasutus</i> (Ogloblin 1939a: 244)	M	Ogloblin (1939a)	-----
<i>necator</i> (Ogloblin 1939a: 241)	BA, LP, SJ	Ogloblin (1939a)	unidentified Cicadellidae and Delphacidae
<i>nigriceps</i> (Ogloblin 1955: 19)	J	Ogloblin (1955c)	-----
<i>nigrithorax</i> (Ogloblin 1953: 2)	J ⁽⁺⁾ , M, SE ⁽⁺⁾ , T ⁽⁺⁾	Ogloblin (1953)	<i>Tapajosa rubromarginata</i> (Signoret) (Cicadellidae) ⁽⁺⁾
<i>parcepilosus</i> (Ogloblin 1957b: 35)	N	Ogloblin (1957b)	-----
<i>perforator</i> (Ogloblin 1953: 4)	M	Ogloblin (1953)	-----
<i>piriformis</i> (Ogloblin 1955: 17)	M	Ogloblin (1955c)	-----
<i>pratensis</i> (Ogloblin 1936: 48)	M, S, T	Ogloblin (1936)	-----
<i>pusilus</i> Ogloblin 1935c: 68	M	Ogloblin (1935c, 1938a)	<i>Entylia carinata</i> (Forster) (Membracidae)
<i>quirogai</i> (Ogloblin 1936: 44)	M	Ogloblin (1936)	-----
<i>schajovskoi</i> (Ogloblin 1957b: 37)	N	Ogloblin (1957b)	-----
<i>spiracularis</i> Ogloblin 1935c: 70	M	Ogloblin (1935c)	-----
<i>stenopterus</i> (Ogloblin 1936: 33)	M	Ogloblin (1936)	-----
<i>tuberculifemur</i> (Ogloblin 1957b: 38)	N	Ogloblin (1957b); Triapitsyn <i>et al.</i> (2008)	
<i>uat</i> Triapitsyn in Triapitsyn, Vickerman <i>et al.</i> 2006: 58	J, S ⁽⁺⁾ , SE ⁽⁺⁾ , T	Triapitsyn, Vickerman <i>et al.</i> (2006)	<i>Tapajosa rubromarginata</i> (Signoret) (Cicadellidae)
<i>urocerus</i> Ogloblin 1935c: 72	BA	Ogloblin (1935c)	-----
<i>valentinae</i> (Ogloblin 1959b: 192)	M	Ogloblin (1959b)	-----
<i>virilai</i> Triapitsyn, Logarzo & León in Triapitsyn, Logarzo <i>et al.</i> 2007: 62	C, CO, ER, LR, ME, S, T	Triapitsyn, Logarzo <i>et al.</i> (2007)	<i>Ciminius platensis</i> (Berg), <i>Dechacona missionum</i> (Berg), <i>Molomea consolidata</i> Schröder, <i>Plesiommata mollicella</i> (Fowler), <i>Tapajosa rubromarginata</i> (Signoret) (Cicadellidae)
<i>Kalopolynema</i> Ogloblin			
<i>discrepans</i> Ogloblin 1960a: 6	BA	Ogloblin (1960a); Triapitsyn & Berezovskiy (2002)	-----
<i>poema</i> Triapitsyn & Berezovskiy 2002: 614	BA	Triapitsyn & Berezovskiy (2002)	<i>Megamelus scutellaris</i> Berg (Delphacidae)
<i>Kikiki</i> Huber & Beardsley			
<i>K. sp.</i>	CA ⁽⁺⁾	(+)	-----
<i>Litus</i> Haliday			

to be continued.

TABLE 1. (continued)

Taxon	Distribution	Reference(s)	Hosts in Argentina
<i>argentinus</i> (Ogloblin 1935a: 60)	M	Ogloblin (1935a); Triapitsyn & Berezovskiy (2004)	-----
<i>missionicus</i> Ogloblin 1955b: 495	M	Ogloblin (1955b)	-----
<i>neotropicus</i> Ogloblin 1955b: 492	M	Ogloblin (1955b)	-----
<i>Myrmecomymar</i> Yoshimoto			
<i>M. sp(p).</i>	BA ⁽⁺⁾ , M ⁽⁺⁾ , S ⁽⁺⁾	(+)	-----
<i>Neomymar</i> Crawford			
<i>mirabilicorne</i> (Ogloblin 1939b: 218)	F, M	Ogloblin (1939b); Triapitsyn, Berezovskiy <i>et</i> <i>al.</i> (2006)	-----
<i>soror</i> (Ogloblin 1939b: 223)	M	Ogloblin (1939b); Triapitsyn, Berezovskiy <i>et</i> <i>al.</i> (2006)	-----
<i>Neostethynium</i> Ogloblin			
<i>stenopterum</i> (Ogloblin 1964: 107)	M	Ogloblin (1964)	-----
<i>Notomymar</i> Doutt & Yoshimoto			
<i>aptenosoma</i> Doutt & Yoshimoto 1970: 293	TF (South Georgia Island)	Doutt & Yoshimoto (1970); De Santis (1981)	-----
<i>Omyomymar</i> Schauff			
<i>clavatum</i> (Ogloblin 1935b:152)	M	Ogloblin (1935b)	-----
<i>silvanum</i> (Ogloblin 1935b:149)	M	Ogloblin (1935b)	-----
<i>Parapolynema</i> Fidalgo			
<i>sagittifer</i> Fidalgo 1982: 98	BA	Fidalgo (1982, 1991b); De Santis (1989)	-----
<i>tucumanum</i> Fidalgo 1991b: 153	T	Fidalgo (1982, 1991b)	-----
<i>Platypolynema</i> Ogloblin			
<i>cautum</i> Ogloblin 1960a: 8	M	Ogloblin (1960a, 1967); Triapitsyn & Berezovskiy (2002)	-----
<i>Polynema</i> Haliday			
<i>luteolum</i> (Ogloblin 1960b: 73)	M	Ogloblin (1960b)	-----
<i>nupogodi</i> Triapitsyn & Aquino 2008: 62	BA, LR	Triapitsyn & Aquino (2008)	-----
<i>pallidiventre</i> (Ogloblin 1960b: 77)	M	Ogloblin (1960b); Triapitsyn & Aquino (2008)	-----
<i>platense</i> (Brèthes 1913: 44)	BA	Brèthes (1913)	-----

to be continued.

TABLE 1. (continued)

Taxon	Distribution	Reference(s)	Hosts in Argentina
<i>polychromum</i> (Ogloblin 1960b: 71)	J, M	Ogloblin (1960b)	-----
<i>reticulatum</i> (Ogloblin 1946: 282)	M	Ogloblin (1946); Loiácono <i>et al.</i> (2005)	-----
<i>saga</i> (Girault)	N, RN	Ogloblin (1960b)	<i>Opsius stactogalus</i> Fieber (Cicadellidae)
<i>Polynemula</i> Ogloblin			
<i>rufosignata</i> Ogloblin 1967: 190	BA	Ogloblin (1967); Triapitsyn & Aquino (2008)	-----
<i>Ptilomymar</i> Annecke & Doutt			
<i>P. sp.</i> (p).	BA ⁽⁺⁾ , M ⁽⁺⁾	⁽⁺⁾	-----
<i>Schizophragma</i> Ogloblin			
<i>basalis</i> Ogloblin 1949: 347	BA, M	Ogloblin (1949)	<i>Aconophora sp.</i> (Membracidae)
<i>nana</i> Ogloblin 1949: 356	M	Ogloblin (1949)	-----
<i>parvula</i> Ogloblin 1949: 355	BA	Ogloblin (1949)	-----
<i>saltensis</i> Ogloblin 1949: 352	S	Ogloblin (1949)	-----
<i>squamosa</i> Ogloblin 1949: 350	M	Ogloblin (1949)	-----
<i>Stephanodes</i> Enock			
<i>missionicus</i> (Ogloblin 1967: 194)	M	Ogloblin (1967); Huber & Fidalgo (1998)	-----
<i>similis</i> (Foerster)	BA, LR, M, S, SF, T	De Santis & Fidalgo (1994); Huber & Fidalgo (1998)	-----
<i>Tetrapolynema</i> Ogloblin			
<i>ogloblini</i> Fidalgo 1991a: 133	M	Fidalgo (1991a)	-----
<i>Xenopolynema</i> Ogloblin			
<i>areolatum</i> Ogloblin 1960a: 9	N, RN	Ogloblin (1960a); Triapitsyn & Berezovski (2007)	-----

⁽⁺⁾: Unpublished records by S.V. Triapitsyn and/or E. Luft Albarracin.

^(*): Personal communication by G.A. Logarzo and/or E.G. Virila.

Key to the genera of Mymaridae in Argentina (females)

1	Tarsi 3-segmented (Fig. 1).....	<i>Kikiki</i> Huber & Beardsley
-	Tarsi 4-or 5-segmented.....	2
2	Tarsi 5-segmented.....	3
-	Tarsi 4-segmented.....	13
3(2)	Metasoma broadly sessile (Figs 2, 3) or subsessile (in <i>Litus</i> Haliday with petiole wider than long and not evident in dry specimens).....	4
-	Metasoma constricted at base, with petiole of varied length, but evident in dry specimens (Fig. 31)	8
4(3)	Funicle 5-segmented (Fig. 4), posterior margin of forewing excised behind venation (Fig. 5). <i>Alaptus</i> Westwood	
-	Antennal funicle 6- or 7-segmented; posterior margin of forewing not or little excised behind venation	5
5(4)	Head and mesosoma strongly sculptured; metasoma subsessile (petiole wider than long and not evident in dry specimens) (Fig. 3); funicle 6-segmented.....	<i>Litus</i> Haliday

- Head and mesosoma very weakly sculptured; metasoma sessile; funicle 6- or 7-segmented 6
- 6(5) Antennal scape with 2 or 3 setaceous teeth; mandible blade-like, with a tooth and a little denticle, projected downward and not overlapping (Fig. 6); funicle 7-segmented..... *Dicopus* Enock
- Antennal scape without setaceous teeth; mandible bidentate, overlapping each other; funicle 6- or 7-segmented. 7
- 7(6) Forewing narrow and elongate, anterior and posterior margins almost parallel; occiput slightly emarginate medially (Fig. 2); funicle 7-segmented (F2 sometimes ring-like)..... *Callodicopus* Ogloblin
- Forewing elongately spatulate; occiput forming an acute V-shaped emargination; funicle 6- or 7-segmented (F2 sometimes ring-like)..... *Dicopomorpha* Ogloblin
- 8(3) Funicle 4- or 5-segmented..... 9
- Funicle at least 6-segmented 10
- 9(8) Fully winged; venation extending over 0.5 length of forewing (Fig. 7); ocelli present, stemmaticum quadrate; occipital carina absent; pronotum, mesoscutum and scutellum not transverse *Arescon* Walker
- Wingless; ocelli absent; stemmaticum absent; occipital carina present; pronotum, mesoscutum and scutellum transverse, 2.5-3.0 x as wide as long (only winged males known from Argentina; similar to males of *Arescon*, but in *Myrmecomymar* stemmaticum absent)..... *Myrmecomymar* Yoshimoto
- 10(8) Funicle 6- or 7-segmented; if 7-segmented, second segment ring-like, much shorter than first segment 11
- Funicle 8-segmented (Fig. 8)..... 12
- 11(10) Mandible with 2 teeth..... *Camptopteroides* Viggiani
- Mandible with 1 tooth *Camptoptera* Foerster
- 12(10) Dorsellum rhomboidal or strap-shaped, with its posterior margin straight *Gonatocerus* Nees s. l.
- Dorsellum mostly strap-shaped but with posterior margin angulate medially, with a little projection (Fig. 9) *Gahanopsis* Ogloblin
- 13(2) Clava 3-segmented (Fig. 10) 14
- Clava 1- or 2-segmented..... 15
- 14(13) Fully winged..... *Neostethynium* Ogloblin
- Wingless *Notomyrmar* Doutt & Yoshimoto
- 15(13) Clava 1-segmented 19
- Clava 2-segmented 16
- 16(15) Marginal vein with hypochaeta about midway between proximal and distal macrochaetae (Fig. 11); propodeum strongly sloping relative to scutellum and with a median longitudinal carina (Fig. 12)..... *Anaphes* Haliday, part
- Marginal vein with hypochaeta next to proximal macrochaeta; propodeum almost in same plane as scutellum and smooth 17
- 17(16) Scutellum longitudinally divided by a distinct median groove (Fig. 13); forewing usually strongly expanded apically (Fig. 14) *Schizophragma* Ogloblin
- Scutellum entire or at most longitudinally divided posteriorly by a faint median groove; forewing at most slightly expanded apically 18
- 18(17) Clava usually with apical finger-like projection (Fig. 15); mandibles reduced; scutellum often longitudinally divided posteriorly by a faint median groove *Omyomyrmar* Schauff
- Clava rounded, without apical finger-like projection (Fig. 16); mandibles not reduced; scutellum undivided..... *Cleruchus* Enock, part
- 19(15) Venation almost 0.5 length of forewing (Fig. 17) *Australomyrmar* Girault
- Venation less than 0.33 length of forewing..... 20
- 20(19) Funicle 8-segmented..... *Ptilomyrmar* Annecke & Doutt
- Funicle 6-segmented, rarely 5-segmented (in *Erythmelus* (*Parallelaptera*) *rex* (Girault) and sometimes in minute specimens of subgenus *Erythmelus* Enock s. str.) 21
- 21(20) Metasoma sessile or subsessile (petiole wider than long) (Figs 2, 3) 22
- Metasoma markedly petiolate (petiole of varied length, but always longer than wide) (Fig. 31). 25
- 22(21) Metasoma with apical sternite much larger than preceding sternite, forming a distinct, V-shaped hypopygium often extending slightly beyond apex of gaster (Fig. 18); mandibles reduced to minute stubs *Erythmelus* Enock
- Metasoma with apical sternite similar to preceding sternites, short and inconspicuous; mandibles normal, overlapping and with 3 teeth..... 23
- 23(22) Marginal vein with hypochaeta about midway between proximal and distal macrochaetae (Fig. 11); propodeum strongly sloping relative to scutellum and with a medial longitudinal carina (Fig. 12); body usually black or very dark brown. *Anaphes* Haliday, part
- Marginal vein with hypochaeta next to proximal macrochaeta; propodeum almost in same plane as scutellum and smooth; body usually yellow, gray, light brown or brown, sometimes dark brown..... 24
- 24(23) Pronotum entire; scutellum clearly divided into anterior and posterior parts and the posterior part longitudinally

	divided by a median groove (Fig. 19); face evenly convex in lateral view.....	<i>Anagrus</i> Haliday
-	Pronotum divided medially; scutellum entire (Fig. 20); face in lateral view usually angular, strongly receding below toruli.....	<i>Cleruchus</i> Enock, part
25(21)	Scutellum divided medially by a longitudinal groove (Fig. 21)	<i>Tetrapolynema</i> Ogloblin
-	Scutellum entire.....	26
26(25)	Toruli almost touching vertex, very close to transverse trabecula (much less than 1 torular diameter) (Fig. 22)	<i>Neomymar</i> Crawford
-	Toruli separated from transverse trabecula by at least 1 torular diameter (Fig. 23).....	27
27(26)	Forewing broadly spatulate (Fig. 24); stigmal vein with a short apical RS-process (Fig. 25).....	<i>Parapolynema</i> Fidalgo
-	Forewing shape different, not broadly spatulate; stigmal vein without an apical process.....	28
28(27)	Basal vein indicated by distinct, curved darkening of membrane (Fig. 26).....	<i>Xenopolynema</i> Ogloblin
-	Basal vein not indicated	29
29(28)	Face with distinct V-shaped subantennal grooves (Fig. 23).....	30
-	Face without subantennal grooves or with subparallel subantennal grooves (in <i>Stephanodes</i> Enock)	31
30(29)	Head about as wide as mesosoma; forewing without narrowing of the blade beyond venation, marginal vein relatively short (Figs 23, 27).....	<i>Kalopolynema</i> Ogloblin
-	Head very large and high, markedly wider than mesosoma; forewing with a narrowing of the blade beyond venation, marginal vein relatively long (Fig. 28).....	<i>Platypolynema</i> Ogloblin
31(29)	Metatibia with conspicuous, strong setae at least as long as width of metatibia (Fig. 29) .	<i>Cnecomymar</i> Ogloblin
-	Metatibia without such setae (setae shorter than width of metatibia)	32
32(31)	Antennal funicle segments 5 and 6 about as wide as clava (Fig. 30).....	<i>Erdosiella</i> Soyka
-	Antennal funicle segments 5 and 6 much narrower than clava.....	33
33(32)	Petiole attached to gastral tergum.....	34
-	Petiole attached to gastral sternum.....	36
34(33)	Propleura not abutting anteriorly, thus prosternum “open”; face without a pit next to each torulus	<i>Polynema</i> Haliday s. str.
-	Propleura abutting each other anteriorly along the midline, thus prosternum “closed”; face with a pit next to each torulus	35
35(34)	Propodeum either smooth or with a median carina, or sometimes with transverse wrinkles	<i>Polynema</i> (subgenus <i>Doriclytus</i> Foerster)
-	Propodeum with two subparallel submedian carinae connected by a transverse carina near anterior margin of propodeum.....	<i>Polynemula</i> Ogloblin
36(33)	Propodeum medially with V-shaped median carina of various size and shape (Figs 31, 32)	<i>Acmopolynema</i> Ogloblin
-	Propodeum either smooth or with a median carina.....	37
37(36)	Scape with rasp-like sculpture (Fig. 33).....	<i>Stephanodes</i> Enock
-	Scape either smooth or serrate (with cross-ridges)	<i>Agalmopolynema</i> Ogloblin

Material examined of the newly recorded genera

[Excluding *Gahanopsis*, which is currently under revision (S.V. Triapitsyn unpublished data).]

Australomymar sp(p).

Australomymar Girault 1929: 343. Type species: *Australomymar aurigerum* Girault, by monotypy.

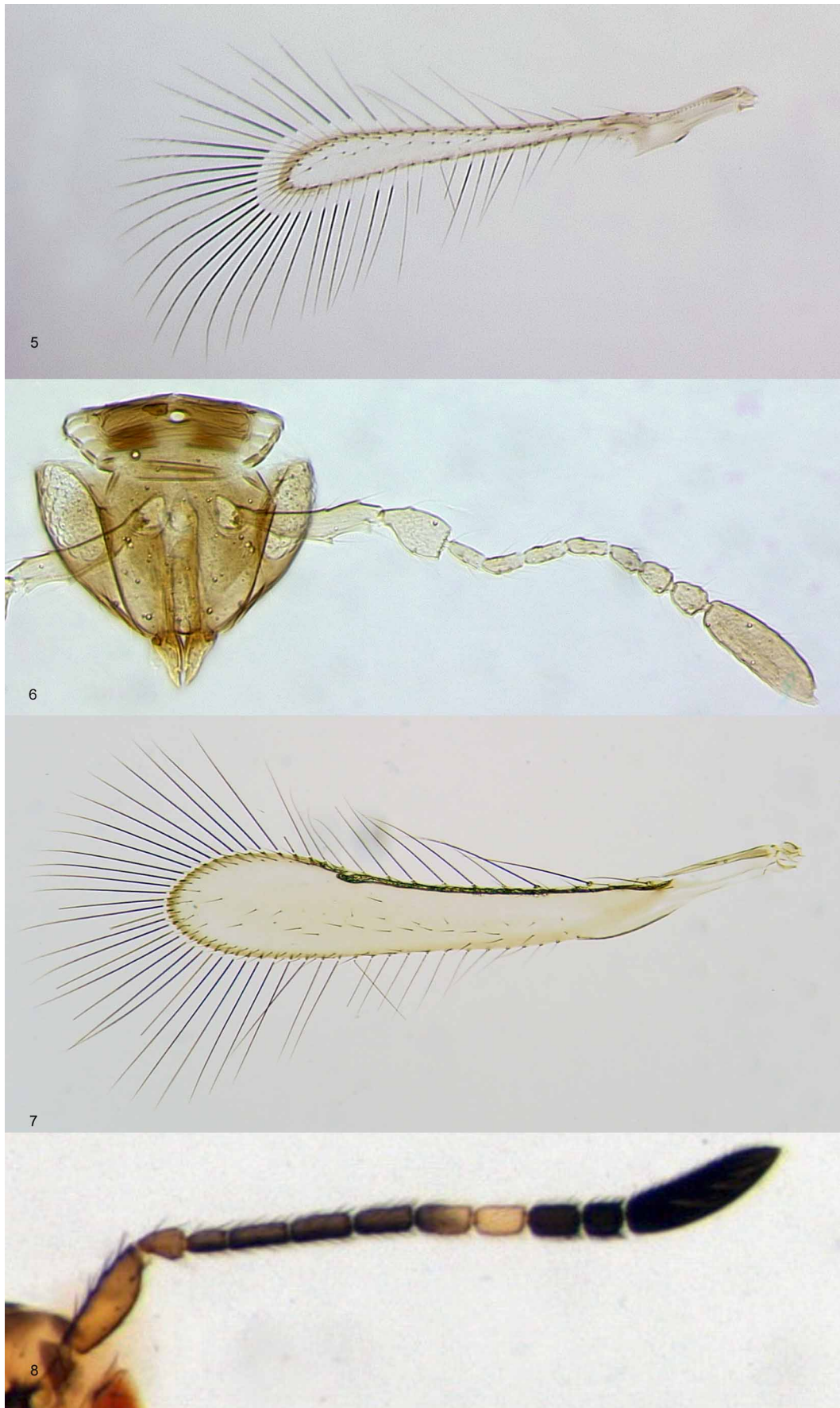
Material examined. ARGENTINA: La Rioja, Santa Vera Cruz, 28°40'42.7''S 66°57'50.4''W, 1660 m, 1–15.xii.2002, P. Fidalgo, Malaise trap [1 male, UCRC]. Neuquén, San Martín de los Andes, iii.1951, S.S. Schajovskoi [1 female, MLPA].

Camptopteroides (Alalinda) sp.

Camptopteroides Viggiani 1974: 23. Type species: *Camptopteroides armata* Viggiani, by original designation.



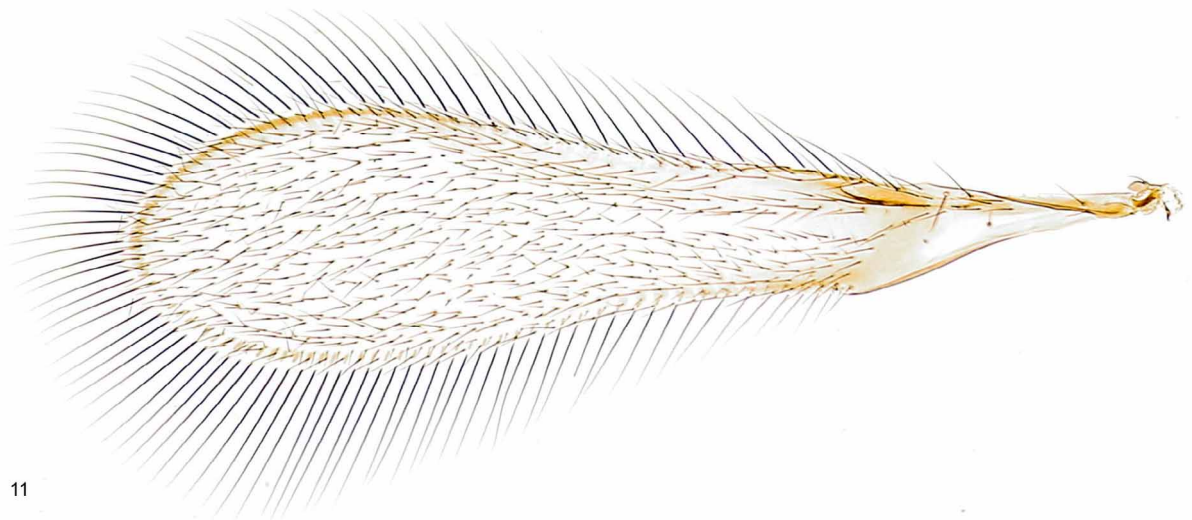
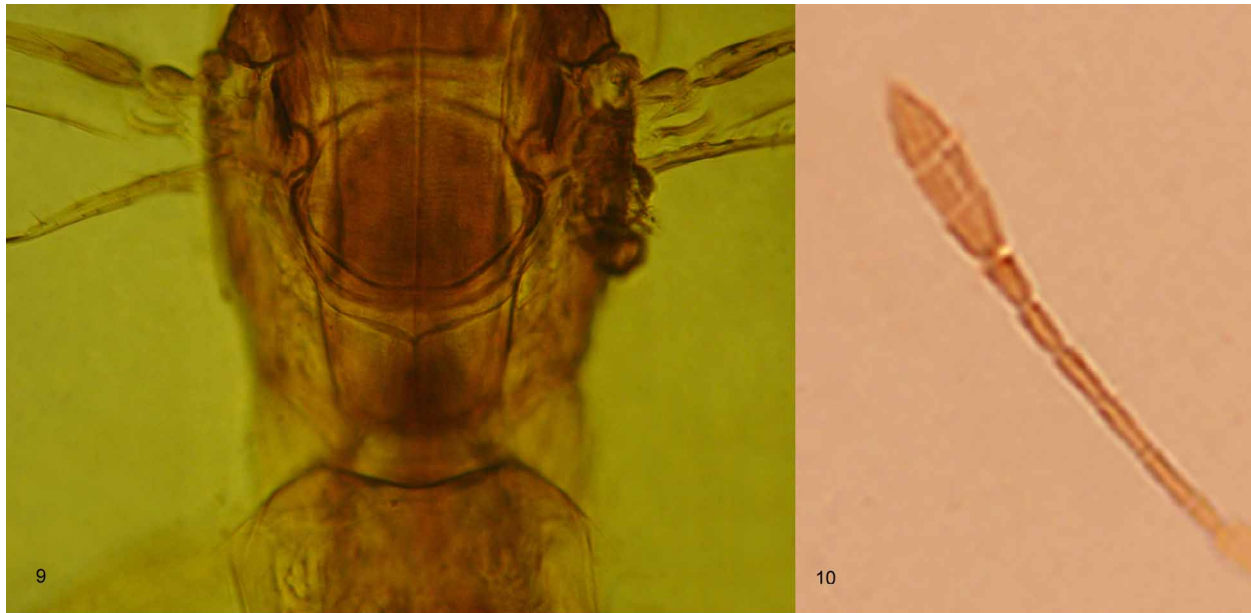
FIGURES 1–4. 1. *Kikiki*, tarsi. 2. *Callodicopus*, female body (dorsal view). 3. *Litus*, mesoscutum and scutellum (dorsal view). 4. *Alaptus*, female antenna.



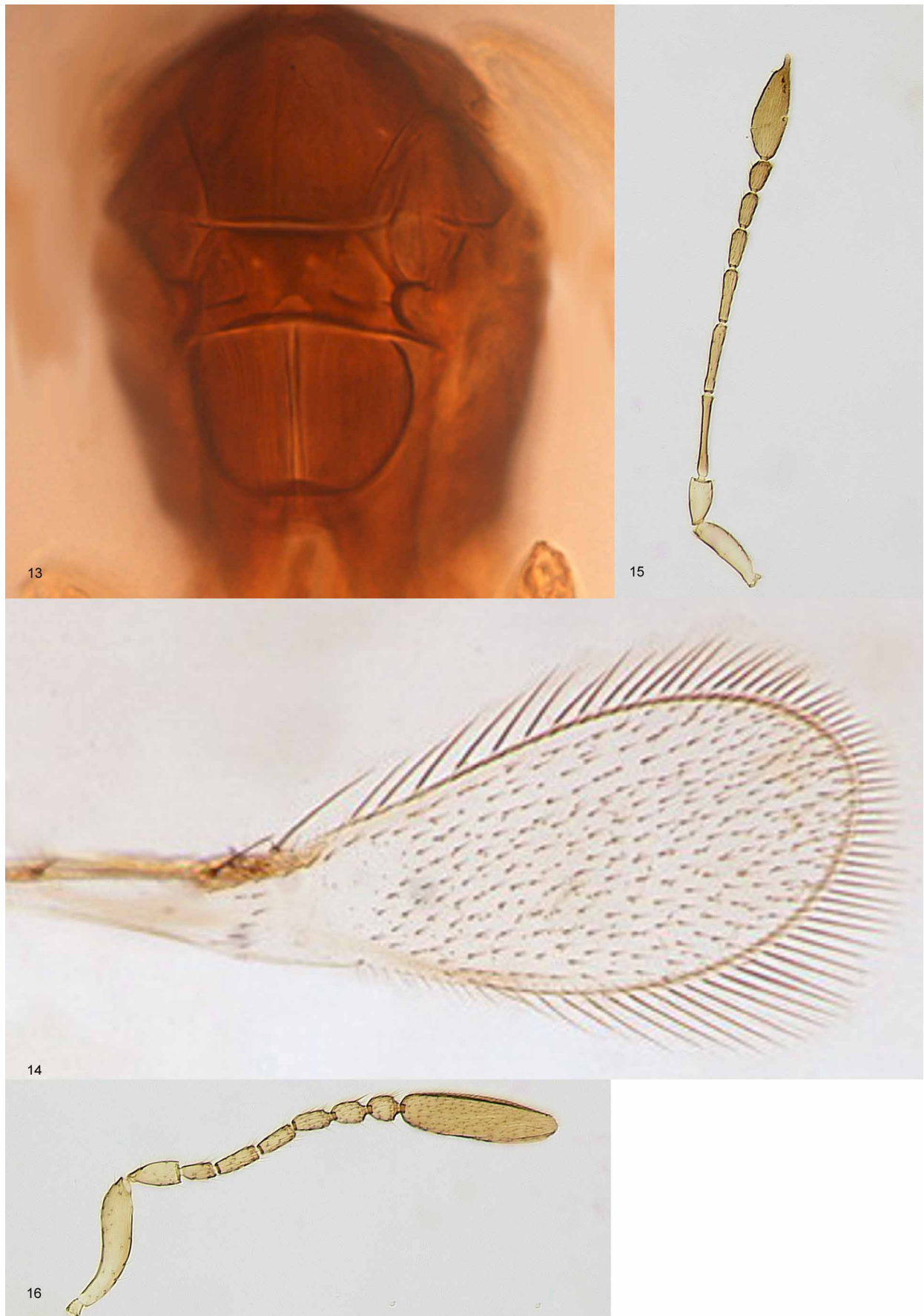
FIGURES 5–8. 5. *Alaptus*, forewing. 6. *Dicopus*, female head (anterior view) and antenna. 7. *Arescon*, forewing. 8. *Gonatocerus*, female antenna.

TERMS OF USE

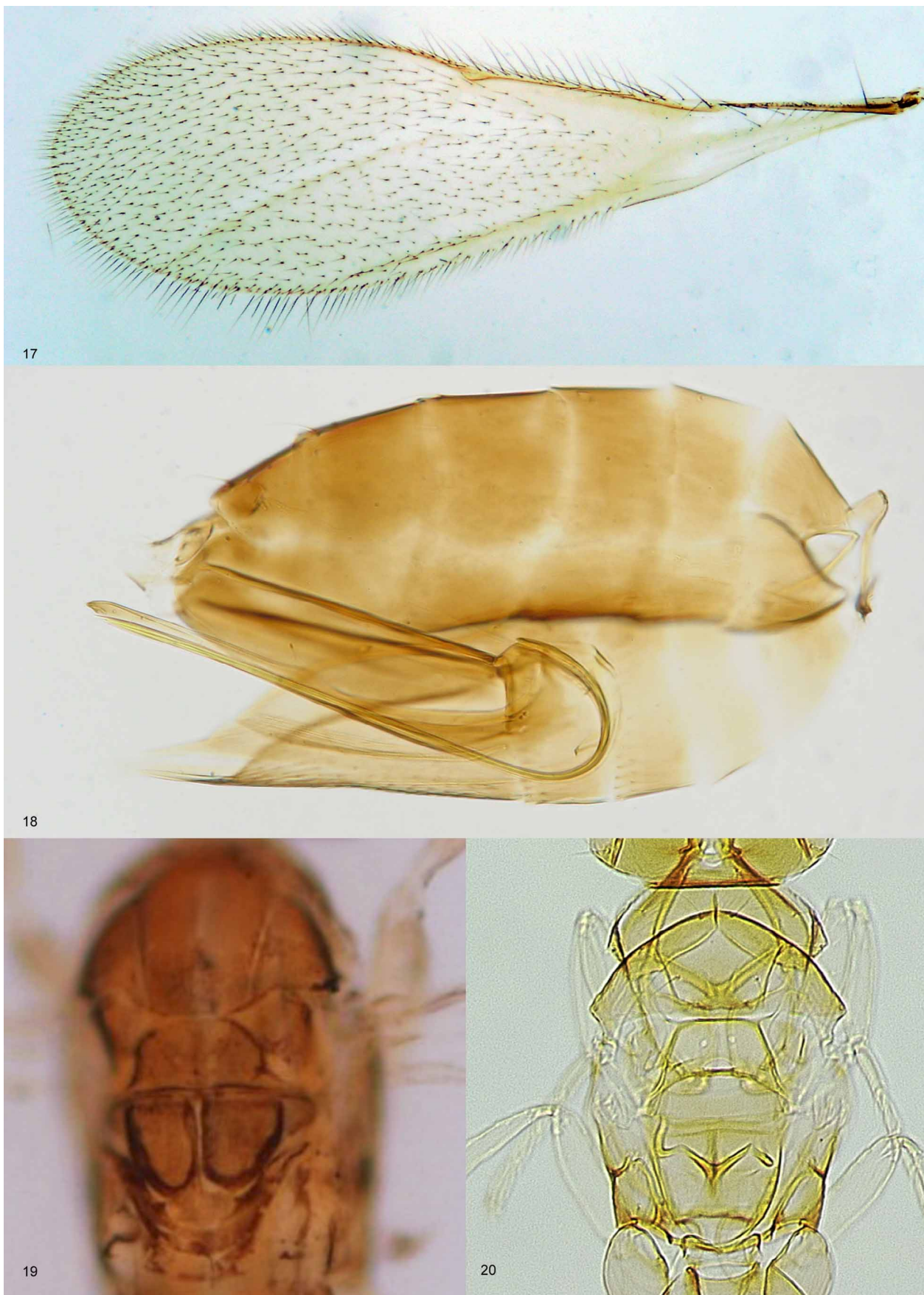
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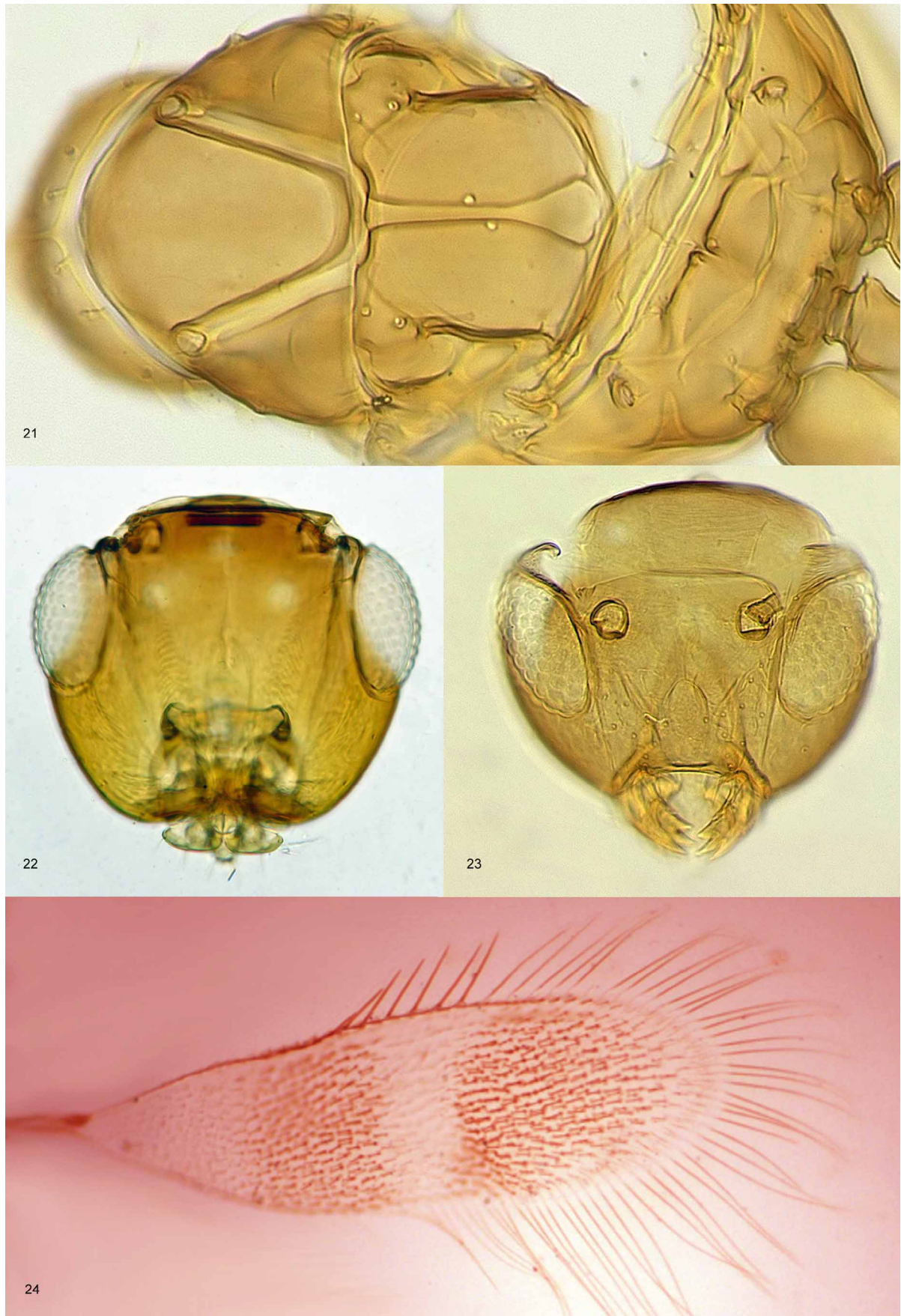
FIGURES 9–12. 9. *Gahanopsis*, mesosoma (dorsal view). 10. *Neostethynium*, female flagellum. 11. *Anaphes*, forewing. 12. *Anaphes*, propodeum (dorsal view).



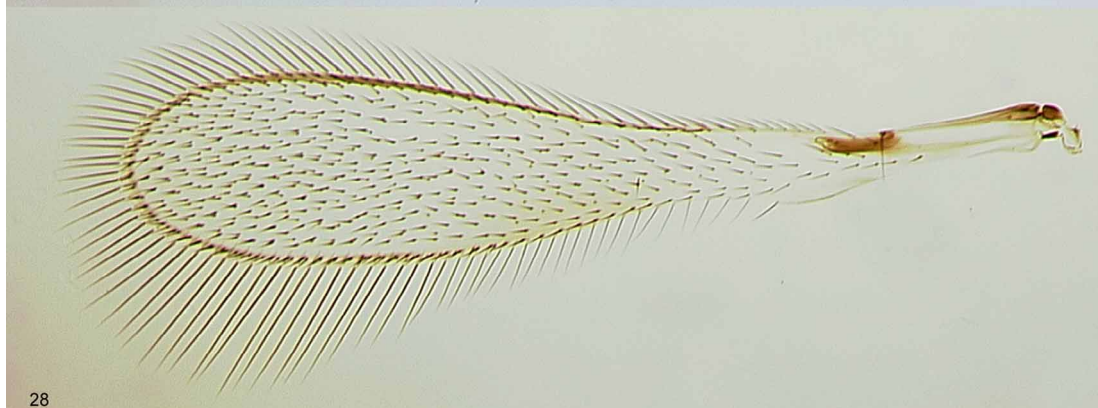
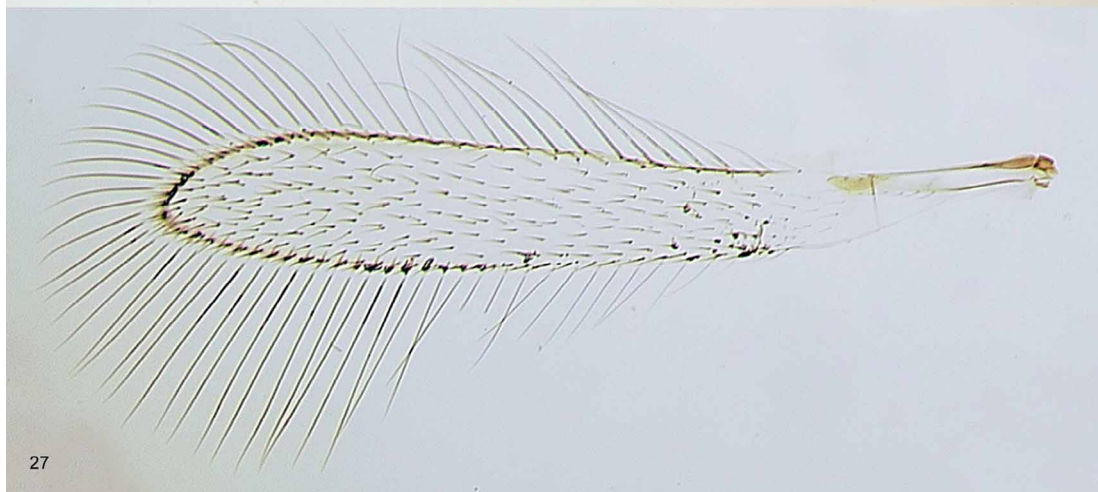
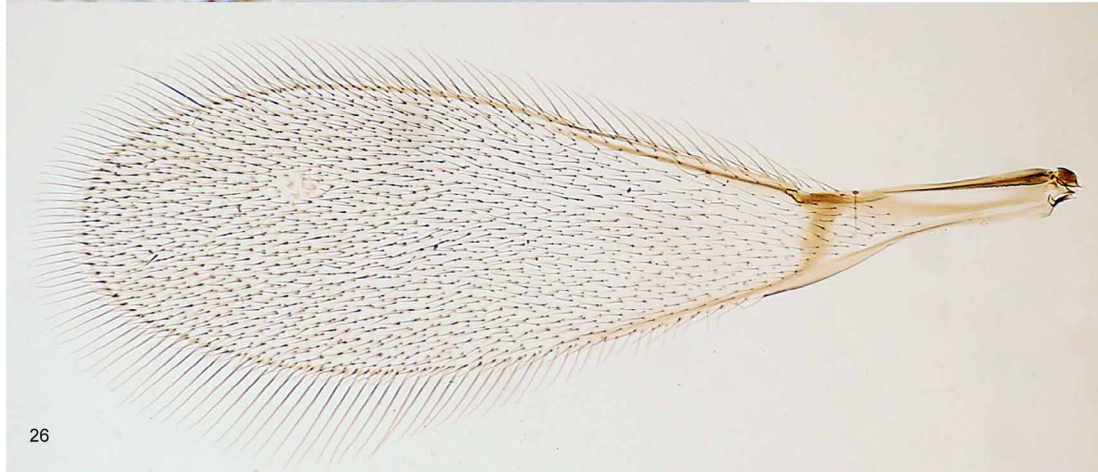
FIGURES 13–16. 13. *Schizophragma*, mesosoma (dorsal view). 14. *Schizophragma*, forewing. 15. *Omyomymar*, female antenna. 16. *Cleruchus*, female antenna.



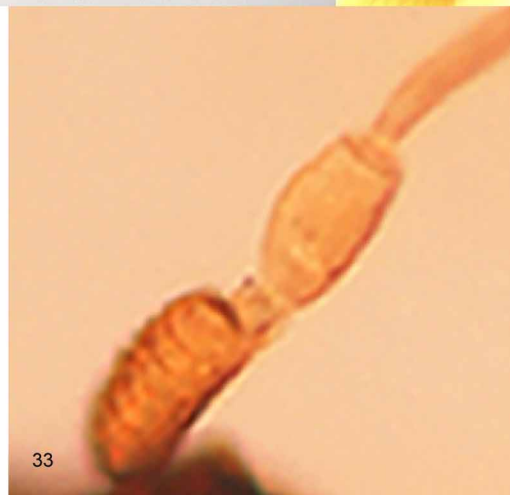
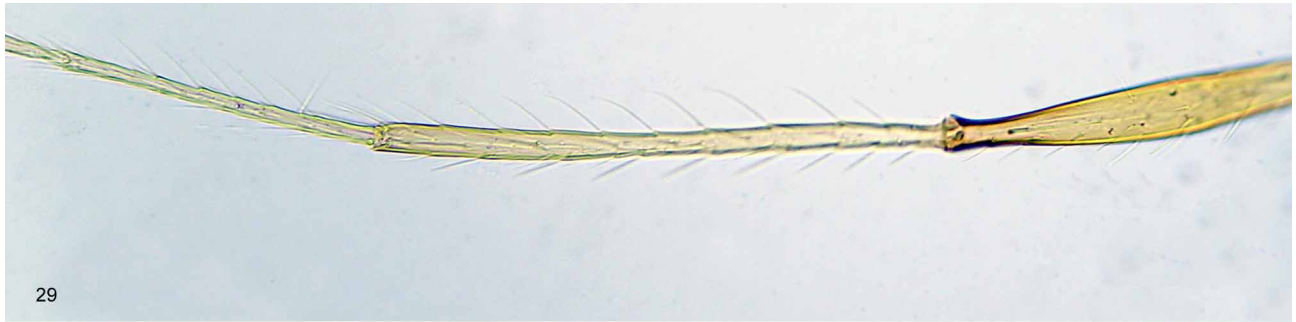
FIGURES 17–20. 17. *Australomymar*, forewing. 18. *Erythmelus*, female metasoma (lateral view). 19. *Anagrus*, mesosoma (dorsal view). 20. *Cleruchus*, mesosoma (dorsal view).



FIGURES 21–24. 21. *Tetrapolynema*, mesosoma (dorsal view). 22. *Neomymar*, head (anterior view). 23. *Kalopolynema*, head (anterior view). 24. *Parapolynema*, forewing.



FIGURES 25–28. 25. *Parapolytnema*, forewing base. 26. *Xenopolytnema areolatum*, forewing. 27. *Kalopolytnema*, forewing. 28. *Platypolytnema*, forewing.



FIGURES 29–33. 29. *Cnecomymar*, metatibia. 30. *Erdosiella*, female antenna. 31. *Acmapolynema*, habitus (male, dorsal view). 32. *Acmapolynema*, propodeum (dorsal view). 33. *Stephanodes*, scape.

Material examined. ARGENTINA: Misiones, Loreto: 10.xii.1947, A.A. Ogloblin [1 female, MLPA]; 12.xii.1947, A.A. Ogloblin (selva) [1 female, MLPA]; 17.ii.1949, A.A. Ogloblin [1 female, MLPA]; 15.ix.1949, A.A. Ogloblin (selva) [1 female, MLPA].

Kikiki sp.

Kikiki Huber & Beardsley 2000: 66. Type species: *Kikiki huna* Huber, by original designation.

Material examined. ARGENTINA: Catamarca, 28°28'54''S 65°39'10''W, 615 m, 19.i.2003, S.V. Triapitsyn, G.A. Logarzo, sweeping [1 female, UCRC].

The genus *Kikiki* Huber & Beardsley is reported for the first time for the New World. A female of an undescribed species of this genus was captured at a roadside in the jungle in lowland mountains. *Kikiki* was previously known only from specimens from the Hawaiian Islands (Huber & Beardsley 2000) and Australia (Lin *et al.* 2007), where one male specimen was found in Western Australia (S.V. Triapitsyn unpublished data).

Myrmecomymar sp(p).

Myrmecomymar Yoshimoto 1990: 28. Type species: *Myrmecomymar masneri* Yoshimoto, by original designation.

Material examined. ARGENTINA: Buenos Aires, Tigre, 34°23'50''S 58°34'32''W, 5 m: 27.xii.2005–3.i.2006, G.A. Logarzo, Malaise trap [1 male, UCRC]; 3–10.i.2006, G.A. Logarzo [1 male, UCRC]. Misiones, Loreto: 16–30.i.2001, P. Fidalgo [1 male, UCRC]; 28.ii.2001, P. Fidalgo [1 male, UCRC]; Ruinas Jesuíticas, 4.ix.2001, S.O. Martínez, P. Fidalgo, Malaise trap [1 male, UCRC]. Parque Nacional Iguazú, 25.68°S 54.45°W, 200 m, 2–7.xii.2003, B. Brown, G. Kung, Malaise trap [2 males, UCRC]. Reserva de Vida Silvestre Urugua-í, 25°58.471'S 54°06.986'W, 400 m, 10–12.xii.2003, B.V. Brown, G. Kung, Malaise trap [1 male, UCRC]. Salta, 23°14'07''S 63°23'22''W, 260 m, 24.iii.2003, J.M. Heraty [1 male, UCRC].

Ptilomymar sp(p).

Ptilomymar Annecke & Doutt 1961: 24. Type species: *Ptilomymar rete* Annecke & Doutt, by original designation.

Material examined. ARGENTINA: Buenos Aires, Tigre, 34°23'50''S 58°34'32''W, 5 m, 1–14.iv.2006, G.A. Logarzo, Malaise trap [1 female, UCRC]. Misiones, Reserva de Vida Silvestre Urugua-í, 25°58.471'S 54°06.986'W, 400 m, 7–9.xii.2003, B.V. Brown, G. Kung, Malaise trap [1 female, UCRC].

Conclusions

Although the mymarid fauna of Argentina is known better than in adjacent countries within the Neotropical region, it is still poorly known, so further studies are necessary. Host associations of Mymaridae are known only for 18.3% of the species (Table 1). Hosts are known only for 8 of the 35 genera recorded in the country, and until now, only parasitoids of Curculionidae (Coleoptera) and Hemiptera (mostly of Auchenorrhyncha, and also of some Miridae and Tingidae) were reported.

Although several genera (such as *Polynemula* Ogloblin, *Parapolynema* Ogloblin, and *Platypolynema* Ogloblin) thus far were recorded in the literature only from Argentina, it is very likely that they also occur in

similar habitats in the adjacent countries: *Polynemula* and *Parapolynema* in Uruguay and *Platypolynema* in Brazil and/or Paraguay; *Parapolynema* was recently found in Chile (S.V. Triapitsyn unpublished data). At present, doing a biogeographical analysis would be difficult because the geographic distribution of many species is poorly known and a great number of the taxa are recorded from only one locality. Not surprisingly, most species records are from the provinces where taxonomists collected most frequently, like Misiones, Buenos Aires, Neuquén, Tucumán, and Salta, but there are no data about the Mymaridae occurring in San Luis and Santa Cruz provinces of Argentina.

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