



Examples of Risk Tools for Pests in Peanut (*Arachis hypogaea*) Developed for Five Countries Using Microsoft Excel

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Abstract

Suppressing pest populations below economically-damaging levels is an important element of sustainable peanut (*Arachis hypogaea* L.) production. Peanut farmers and their advisors often approach pest management with similar goals regardless of where they are located. Anticipating pest outbreaks using field history and monitoring pest populations are fundamental to protecting yield and financial investment. Microsoft Excel was used to develop individual risk indices for pests, a composite assessment of risk, and costs of risk mitigation practices for peanut in Argentina, Ghana, India, Malawi, and North Carolina (NC) in the United States (US). Depending on pests and resources available to manage pests, risk tools vary considerably, especially in the context of other crops that are grown in sequence with peanut, cultivars, and chemical inputs. In Argentina, India, and the US where more tools (e.g., mechanization and pesticides) are available, risk indices for a wide array of economically important pests were developed with the assumption that reducing risk to those pests likely will impact peanut yield in a positive manner. In Ghana and Malawi where fewer management tools are available, risks to yield and aflatoxin contamination are presented without risk indices for individual pests. The Microsoft Excel platform can be updated as new and additional information on effectiveness of management practices becomes apparent. Tools can be developed using this platform that are appropriate for their geography, environment, cropping systems, and pest complexes and management inputs that are available. In this article we present examples for the risk tool for each country.

Key words: agronomy, crop rotation, cultivar resistance, decision tool, IPM-Agriculture, pesticide

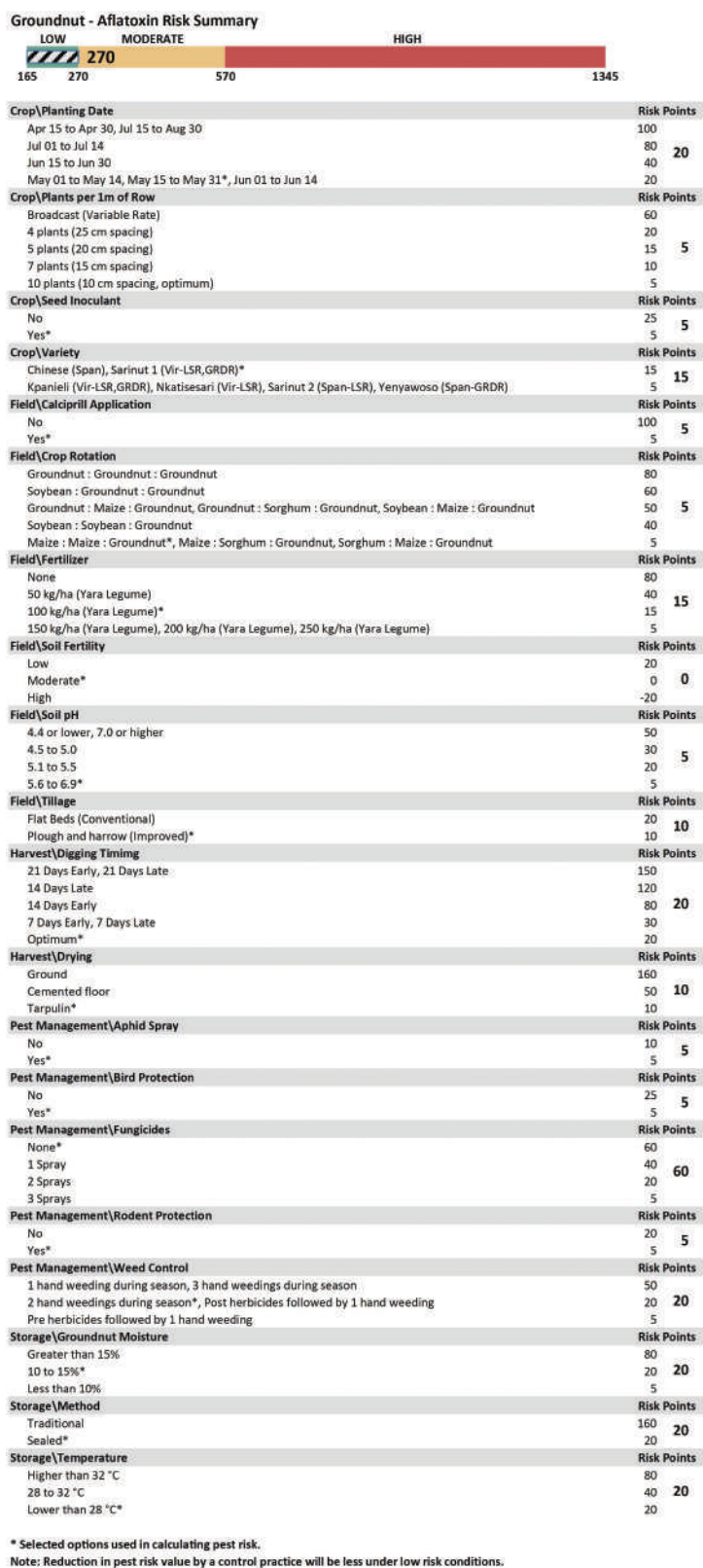


Fig. 1. Risk summary for aflatoxin contamination in the northern Ghana peanut risk tool.

Peanut (*Arachis hypogaea* L.) is an important crop in many regions of the world and contributes to food security due to the resilience it adds in cropping systems and positive contributions to the human diet (Stalker et al. 2016, Valentine 2016). However, peanut is susceptible to a wide range of biotic and abiotic stresses that can limit yield

and quality and create issues associated with food safety (Nigam et al. 2018, Jordan et al. 2018). Low yield and poor quality can affect financial sustainability of peanut-based cropping systems. Employing cost-effective practices to minimize the impact of pests can increase peanut yield and financial sustainability. Research and

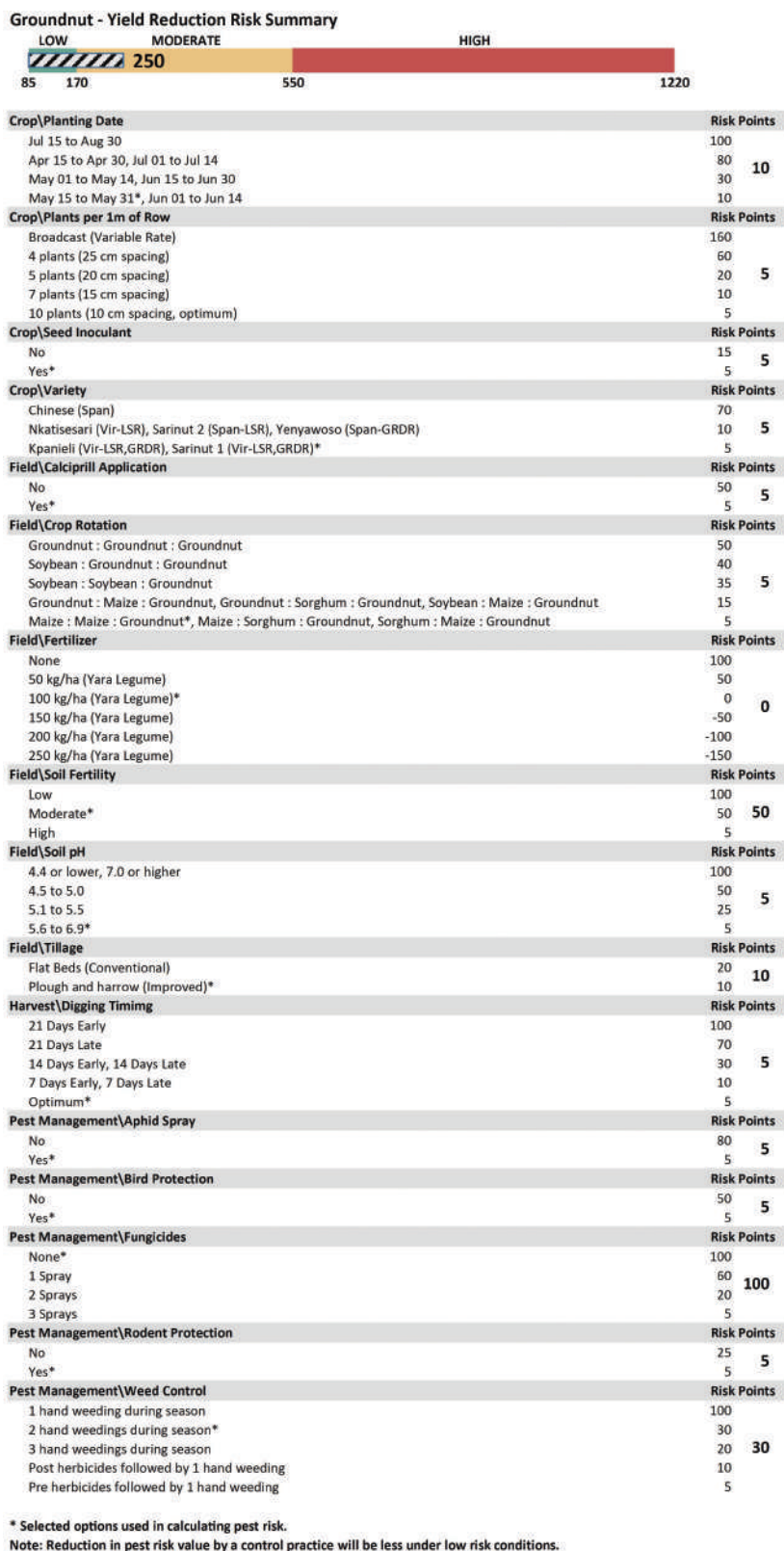


Fig. 2. Risk summary for yield in the northern Ghana peanut risk tool.

educational programs by the public institutions, the private sector, and nongovernmental organizations often provide solutions to pests that adversely affect the peanut crop. Many of these solutions are developed locally with an understanding of the financial impact of

pests and use of interventions that are available and economically practical.

Even though effective strategies and tools are available to suppress pests in peanut, information about those strategies is often

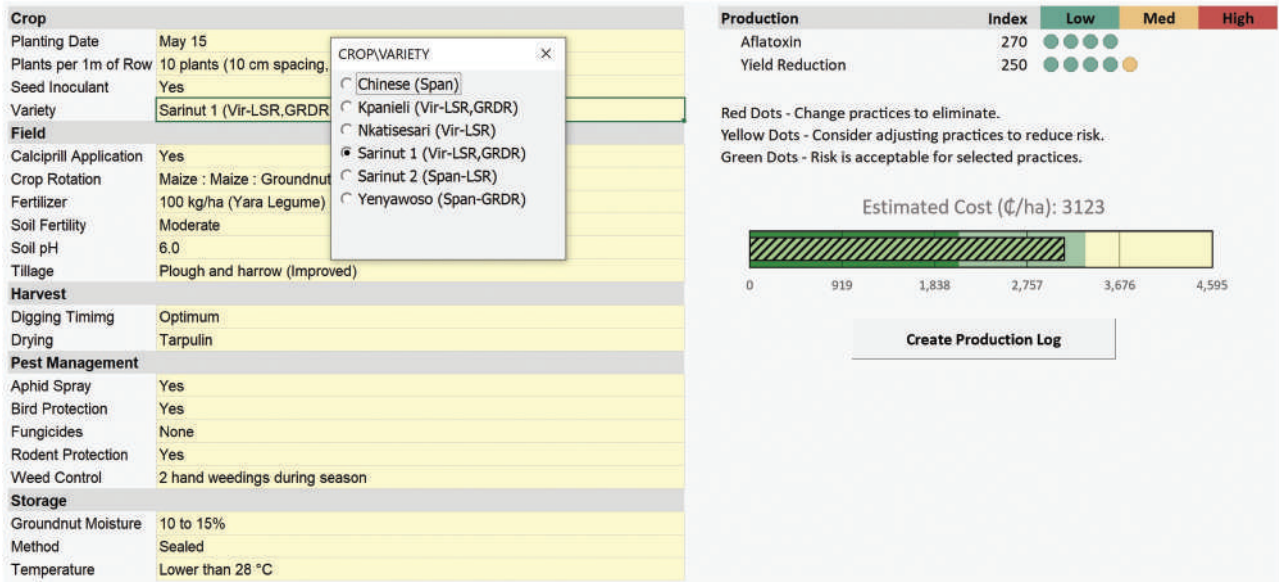


Fig. 3. Drop down menu for varieties in the northern Ghana peanut risk tool.

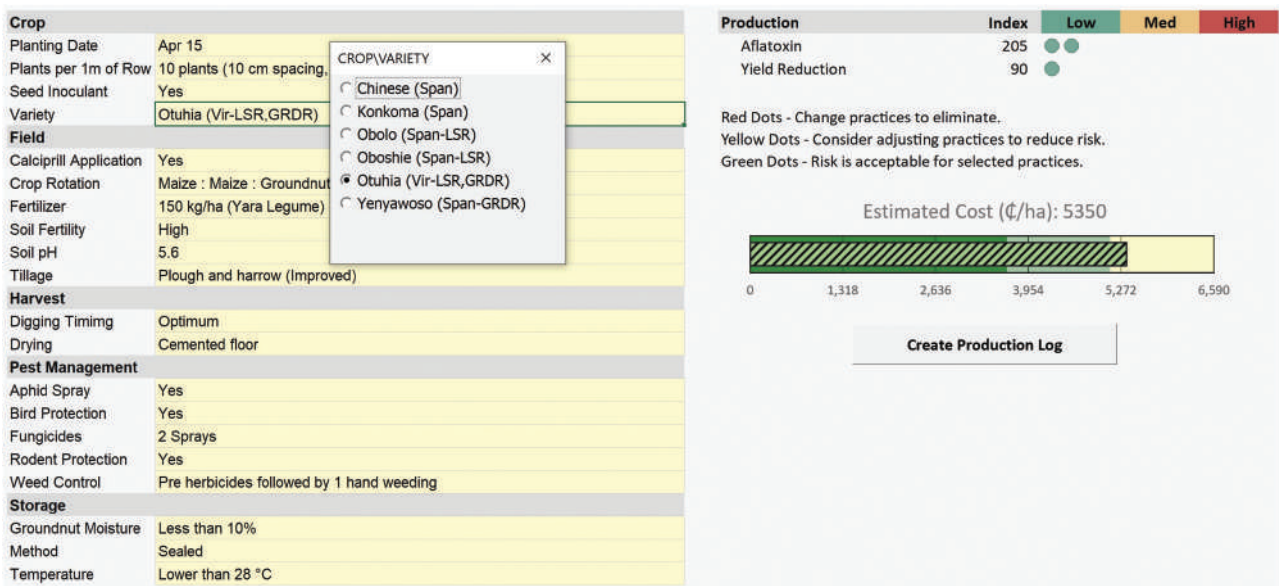


Fig. 4. Drop down menu for varieties in major season in the southern Ghana peanut risk tool.

presented for individual disciplines (e.g., entomology, plant pathology, nematology, and weed science). In some instances practitioners are required to search through resources to determine interaction across pest disciplines in order to develop a holistic approach to pest management. Several approaches have been developed to address this challenge. In the southeastern region of the US, the Peanut Rx guide allows growers and their advisors to determine the impact of production and pest management practices on tomato spotted wilt (tospovirus, Bunyaviridae) transmitted by thrips (*Frankliniella fusca* Pergande, *F. occidentalis* Hinds) and other pathogens in peanut (Anonymous 2022). In North Carolina in the United States, a Microsoft Excel platform was developed to assess overall risk from production and pest management practices for thirteen pests or groups of pests commonly found in peanut (Jordan et al. 2022). Outside of these educational resources, there

are no electronic resources in other countries for peanut that allow the research and education community and practitioners to easily assess the composite risk based on strategies that are planned for a particular field and cropping cycle across several disciplines. An electronic tool that enables farmers and their advisors to assess overall risk with different practices in a more effective manner could potentially result in greater protection of yield and increased financial sustainability.

In NC, a Microsoft Excel platform was designed to allow farmers and their advisors (e.g., private crop consultants, extension agents, agribusiness, nongovernmental organizations, and Federal and State agencies) to identify risk from a set of practices based on field history (Jordan et al. 2022). The platform computes cost of each set of practices so that farmers can observe the financial impact of changes in practices designed to reduce risk. A data log function is also a part

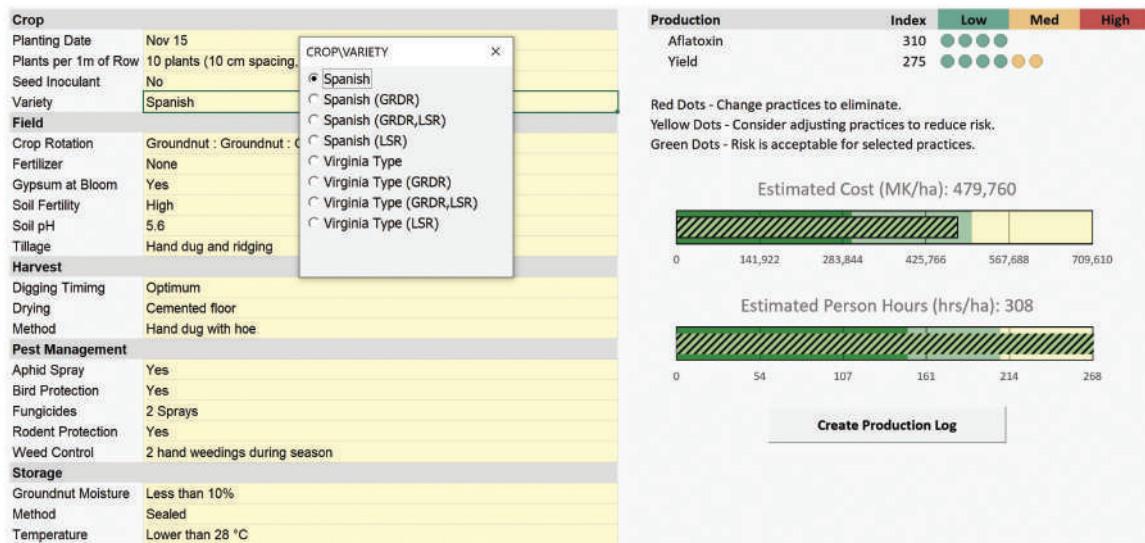


Fig. 5. Drop down menu for varieties in the Malawi peanut risk tool.

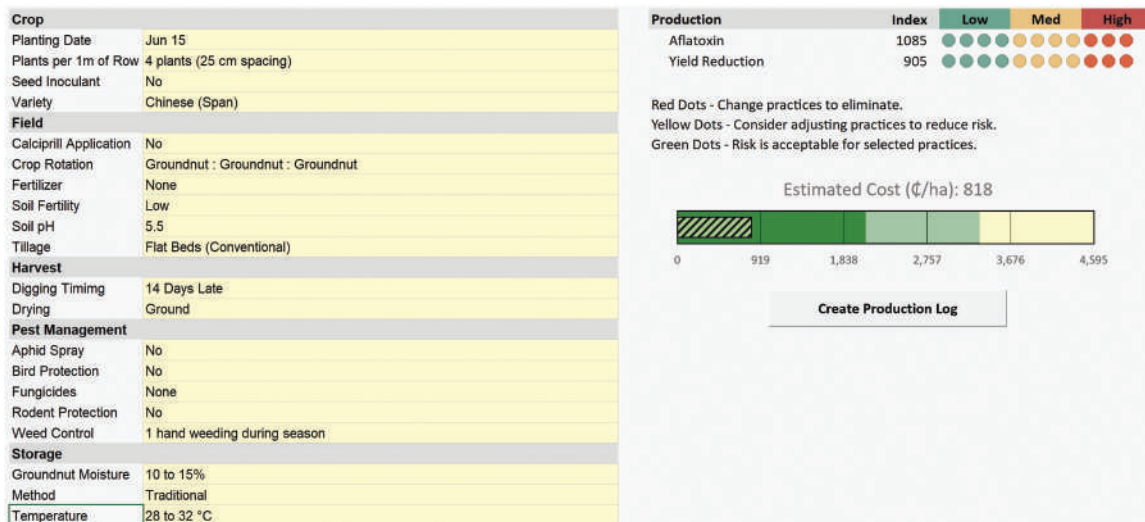


Fig. 6. Risk to aflatoxin contamination, yield, and cost of production for the limited input system in the northern Ghana peanut risk tool.

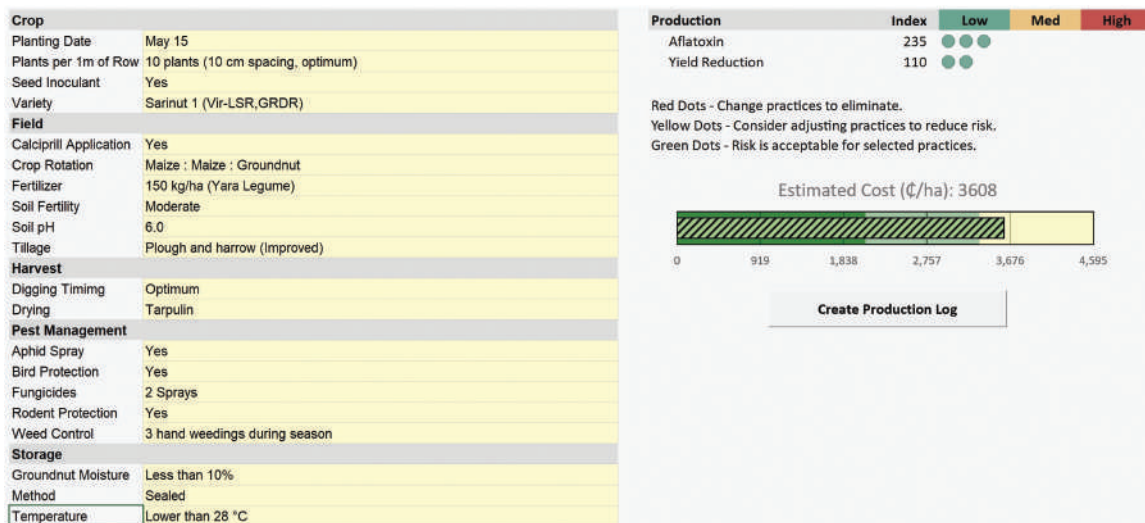


Fig. 7. Risk to aflatoxin contamination, yield, and cost of production for the high input system in the northern Ghana peanut risk tool.

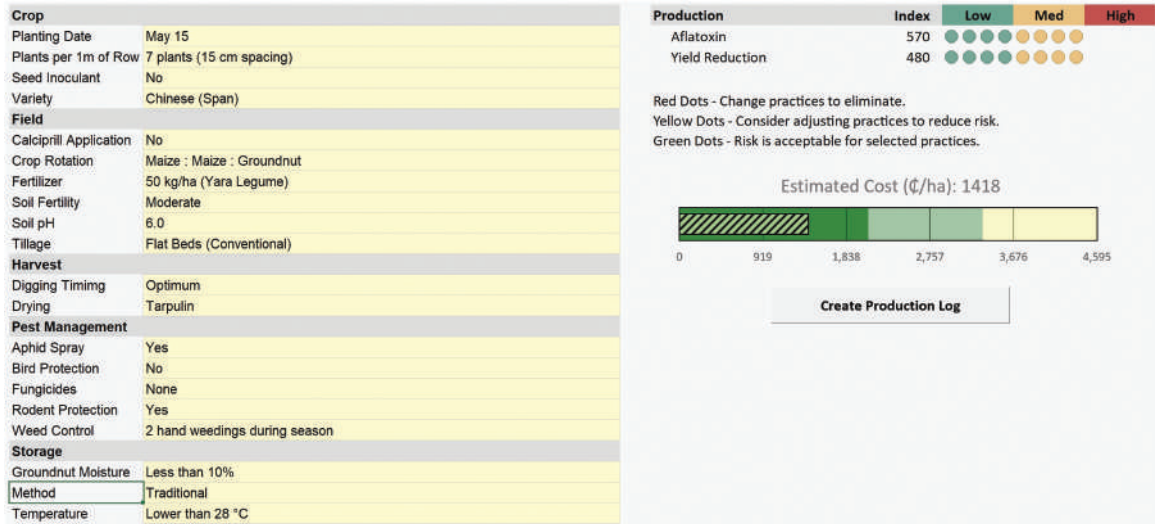


Fig. 8. Risk to aflatoxin contamination, yield, and cost of production for the medium input system in the northern Ghana peanut risk tool.

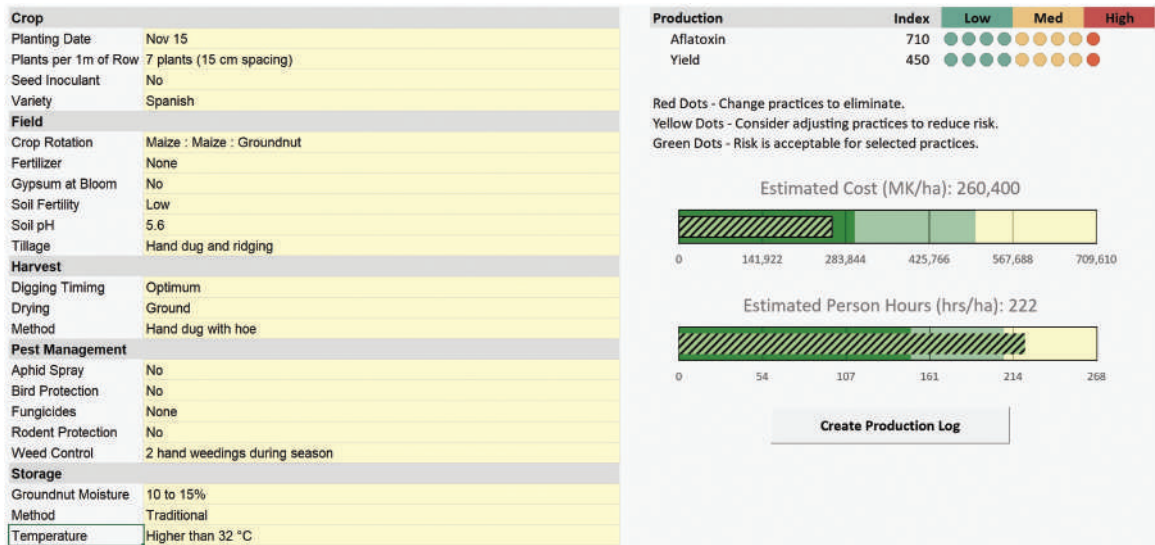


Fig. 9. Risk to aflatoxin contamination, yield, and cost of production for the limited input system in the Malawi peanut risk tool.

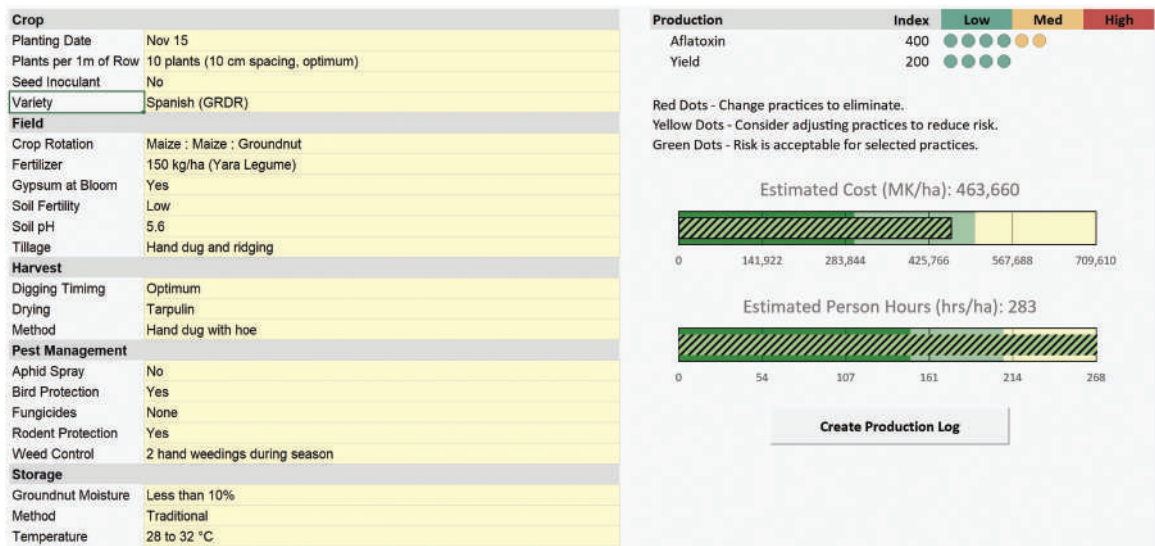


Fig. 10. Risk to aflatoxin contamination, yield, and cost of production for the high input system in the Malawi peanut risk tool.

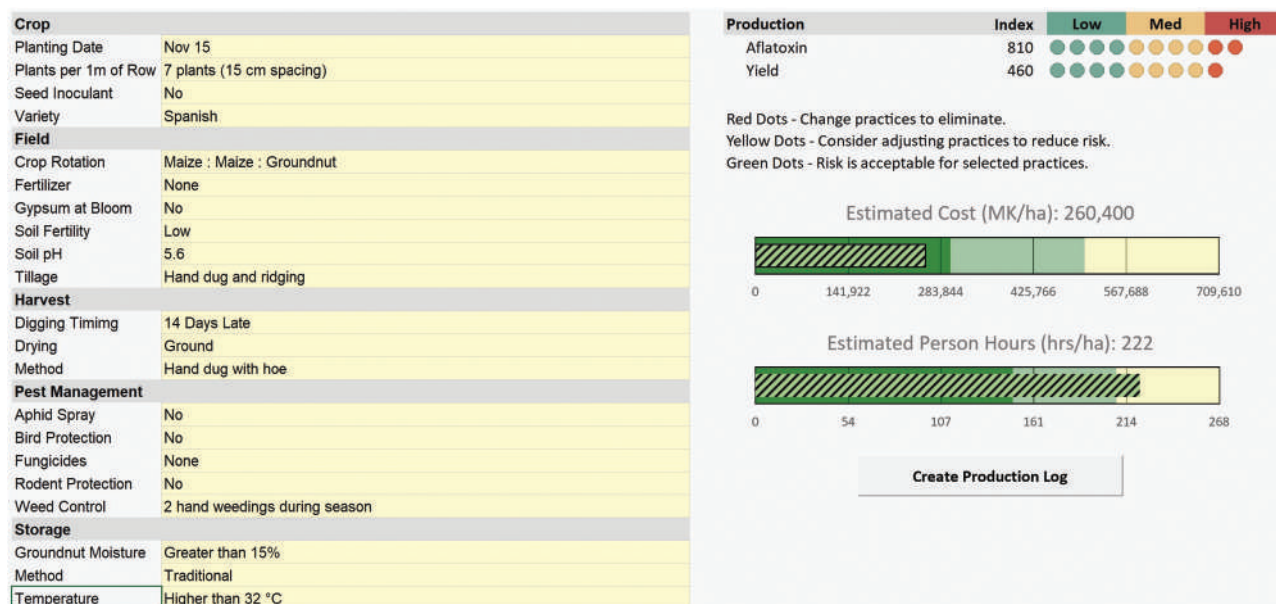


Fig. 11. Influence of timing of digging, drying method, and approaches to storage on aflatoxin contamination, yield, and cost of production in Malawi peanut risk tool with poor practices.

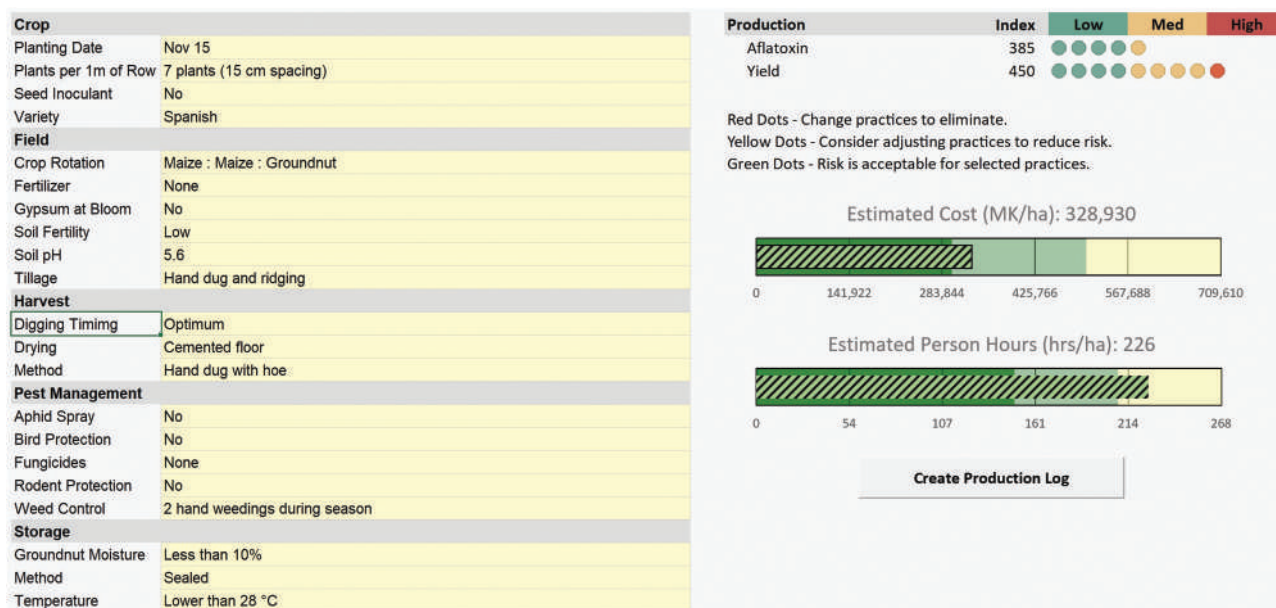


Fig. 12. Influence of timing of digging, drying method, and approaches storage on aflatoxin contamination, yield, and cost of production in Malawi peanut risk tool with improved practices.

of the platform that enables the user to electronically record production and pest management practices for the field and other important factors including yield, market grade characteristics, and rainfall. A detailed description of the NC peanut risk tool and examples of pests and pest management interactions are provided elsewhere (Jordan et al. 2022). When the NC peanut risk tool was under development, the decision to use Microsoft Excel as the platform was made so that tools for other states in the US or other countries could use the platform to create their own risk management tool. A portion of the funding for the development of the NC peanut risk tool was from the USAID Feed the Future Innovation Lab for Peanut with a specific goal of creating a tool that was transferable to partnering countries and ultimately a risk management tool that is available for the general public. In this

paper, we provide examples of Microsoft Excel based peanut risk tools developed for Argentina, Ghana, India, and Malawi using the peanut risk tool initially developed for NC. The current iteration of each of these tools, a blank template, and an instructional video for creation of a risk tool can be found at: <https://cropmanagement.cals.ncsu.edu/risk-tools/peanut.html>.

Peanut Risk Tools in Ghana and Malawi

The peanut risk tools for Ghana and Malawi were developed simultaneously with information from both countries exchanged among scientists and practitioners. Risk to yield and contamination by aflatoxin (produced by *Aspergillus flavus* and *A. parasiticus*)

Crop			
Cultivar	GG 20		
Planting Date	Jun 25		
Plants per 1m Row	6 or fewer		
Row Pattern	Single (75 - 1		
Field			
Intercrop	None		
Irrigation	Irrigated		
Soil pH	7.0		
Tillage	Conventional		
Field Crop Rotation			
1 Crop Season Ago	Groundnut		
2 Crop Seasons Ago	Groundnut		
3 Crop Seasons Ago	Groundnut		
4 Crop Seasons Ago	Groundnut		
Field Nematode Population			
Root Knot	High		
Root Lesion	High		
Stunt	High		
Field Pest History			
Aflaroot	Problem even		
Alternaria Leaf Spot	Problem even		
Collar Rot	Problem even with good fungicide/biocontrol program		
Defoliators	Problem even with good pesticide program		
Early Leaf Spot Rainfed	Problem even with good fungicide program		
Early Leaf Spot Summer	Problem even with good fungicide program		
Late Leaf Spot Rainfed	Problem even with good fungicide program		
Late Leaf Spot Summer	Problem even with good fungicide program		
PBND	Problem even with good fungicide program		
Rust	Problem even with good fungicide program		
Soil Pests	Problem even with good pesticide program		
Stem Rot	Problem even with good fungicide/biocontrol program		
Sucking Pests	Problem even with good pesticide program		
Natural Pest Enemy Population			
Defoliators	Low/Below Average		
Sucking Pests	Low/Below Average		
Pest Control			
Nematicide	None		

CROP\CULTIVAR

BG 3

CSMG 84-1

GG 2

GG 20

GG 7

ICG FDRS 10

ICGS 44

ICGS 76

ICGV 86590

M 335

M 522

R 8808

R 9201

Somnath

TG 37 A

TPG-41

Disease (Foliar)	Index	Low	Med	High
Alternaria Leaf Spot	118	●●●	●●●	●●●
Early Leaf spot - Rainfed	118	●●●	●●●	●●●
Early Leaf Spot - Summer	118	●●●	●●●	●●●
Late Leaf Spot - Rainfed	118	●●●	●●●	●●●
Late Leaf Spot - Summer	118	●●●	●●●	●●●
PBND	136	●●●	●●●	●●●
Rust	123	●●●	●●●	●●●
Disease (Stem)	Index	Low	Med	High
Aflaroot	106	●●●	●●●	●●●
Collar Rot	106	●●●	●●●	●●●
Stem Rot	126	●●●	●●●	●●●
Insect	Index	Low	Med	High
Defoliators	136	●●●	●●●	●●●
Sucking Pests	136	●●●	●●●	●●●
Nematode	Index	Low	Med	High
Root Knot	80	●●●	●●●	●●●
Root-Lesion	80	●●●	●●●	●●●
Stunt-Kalahasty Malady	80	●●●	●●●	●●●
Soil	Index	Low	Med	High
Pests	131	●●●	●●●	●●●

Red Dots - Change practices to eliminate.
 Yellow Dots - Consider adjusting practices to reduce risk.
 Green Dots - Risk is acceptable for selected practices.

Fig. 13. Drop down menu for varieties in India peanut risk tool.

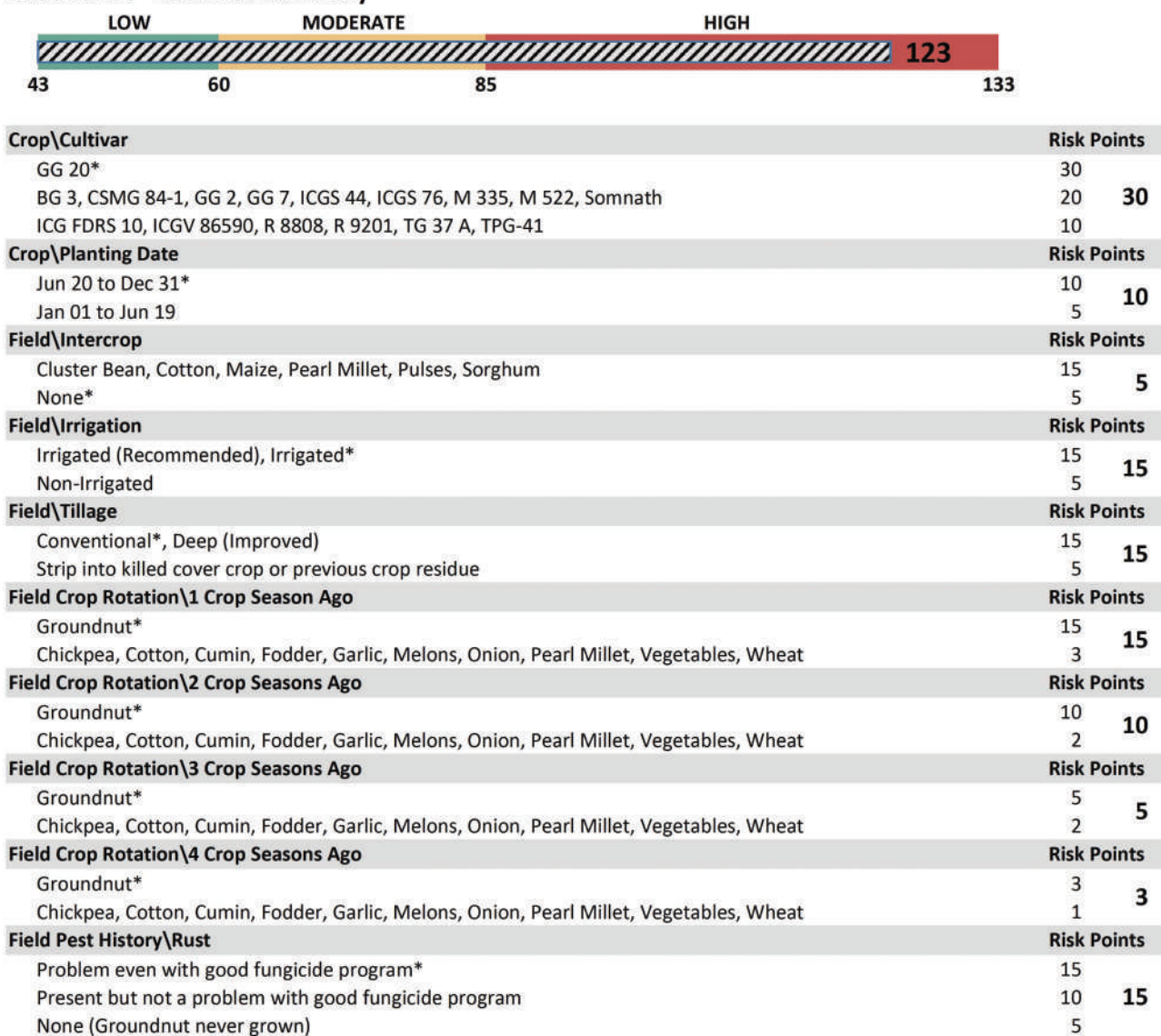
were compared using information for five categories of practices. Examples of the components of the northern Ghana peanut risk tool are presented in Figs. 1 and 2. The risk tools differed between the countries primarily in areas of cultivar selection, planting patterns and plant population, and planting dates. Also, peanut production in Ghana is impacted by a bimodal rainfall pattern in southern Ghana and a unimodal rainfall pattern in northern Ghana. Malawi has a single production season similar to northern Ghana. Risk tools in Ghana and Malawi include estimates of production and pest management costs. The Malawi risk tool also includes estimates of the time required to complete tasks (e.g., labor costs in person hours). Cultivar selection is a major driver of yield and is an important element of risk tools in Ghana and Malawi. Drop down menus for cultivar selection for both risk tools in Ghana (unimodal and bimodal rainfall seasons) and the risk tool in Malawi are presented in Figs. 3–5.

Risk to yield and aflatoxin contamination for three levels of input for northern Ghana are contrasted in Figs. 6–8. When inputs are limited, risk to both yield and aflatoxin are high as noted with three dots in the red category for both parameters (Fig. 6). This approach to peanut production in many areas of Ghana is not uncommon where availability of interventions are limited and financial constraints exist (e.g., financial credit and access to loans) (Abudulai et al. 2020, Appaw et al. 2020). Estimated cost of production for this low input system was \$131/ha (818 Ghana cedes/ha). When resources are available and interventions are included across all categories, risk to yield and contamination by aflatoxin was essentially eliminated but at a cost that is over four times the cost of the low input system (\$577/ha or 3,608 Ghana cedes/ha) (Fig. 7). Few peanut farmers in Ghana have access to all interventions

and/or financing to purchase available resources prior to the cropping cycle. A reasonable alternative to both the low and high input systems is presented in Fig. 8. Risk in this scenario remains relatively high (e.g., yellow dots for yield and aflatoxin) but with lower costs at \$226/ha (1,418 Ghana cedes/ha). Although cost is greater than the low input system, risk to yield and aflatoxin is lowered considerably compared with the low input system.

The Malawi peanut risk tool allows practitioners to observe not only changes in cost of production as risk is addressed but also gives an estimate of the labor involved as practices are modified. For example, cost of production when inputs are limited is \$322/ha (260,400 Malawian kwacha/ha) with 222 person hours required in the limited input system (Fig. 9). In contrast, risk was lowered with increased inputs (e.g., fertilizer, gypsum, fungicide, and additional hand weeding) but required an increase to 283 person h/ha and a cost of \$574/ha (463,660 Malawian kwacha/ha) (Fig. 10). The Malawi risk tool also demonstrates the value of adopting improved practices associated with digging peanut, drying, and storage to mitigate aflatoxin contamination (Figs. 11 and 12). Two red dots were present when peanut was dug 14 d after optimum pod maturity, dried on the ground to moisture exceeding 15%, and stored in a traditional setting at temperatures exceeding 32°C (Fig. 11). Risk was reduced to only one yellow dot when peanut was dug at optimum maturity, dried on cement flooring to less than 10% moisture, and stored in sealed bags at 28°C or lower (Fig. 12). Although not captured in this version of the Malawi risk tool, previous research (Appaw et al. 2020) reported that drying on tarps and storing in hermetically-sealed bags prevented increases in aflatoxin contamination during storage compared with traditional practices (e.g., drying on soil and storing in non-sealed bags) and also resulted in more higher quality

Groundnut - Rust Risk Summary



* Selected options used in calculating pest risk.

Note: Reduction in pest risk value by a control practice will be less under low risk conditions.

Fig. 14. Risk summary for rust in the India peanut risk tool.

kernels for the market. Less time and labor would be needed by the farmer because quality of peanut is at a higher level due to improved harvest, drying, and storage.

Argentina, India, and the United States

In contrast to Ghana and Malawi, farmers in Argentina, India, and NC (USA) have greater resources and inputs at their disposal to manage pests. While discussed in detail elsewhere, the NC risk tool includes individual risk indices for 13 pests or groups of pests and a wide range of pesticides available to suppress pest populations (Jordan et al. 2022). At the current time, risk tools for Argentina and India do not have cost of inputs. Improved cultivars are widely available for adoption as they are released because of a reliable certified seed delivery system. With the exception of tomato spotted wilt, a

significant number of pesticides is available to suppress all pests that are economically important for peanut. However, cultural practices also contribute to suppression of pests.

The India peanut risk tool includes 16 pests or groups of pests under five categories (Fig. 13). A drop down menu for cultivars is presented in Fig. 13. Practices that affect rust (*Puccinia arachidis* Speg.) in peanut are presented in Fig. 14. When peanut was grown continuously and intercropped with corn (*Zea mays* L.) with limited inputs, risk was high (e.g., numerous red dots) for all pests (Fig. 15). In contrast, establishing a more effective rotation sequence with two cycles of cotton (*Gossypium hirsutum* L.), not intercropping, and planting a cultivar with resistance to this pathogen decreased risk substantially (Fig. 16). The current India peanut risk tool does not include the cost associated with production and pest management practices.

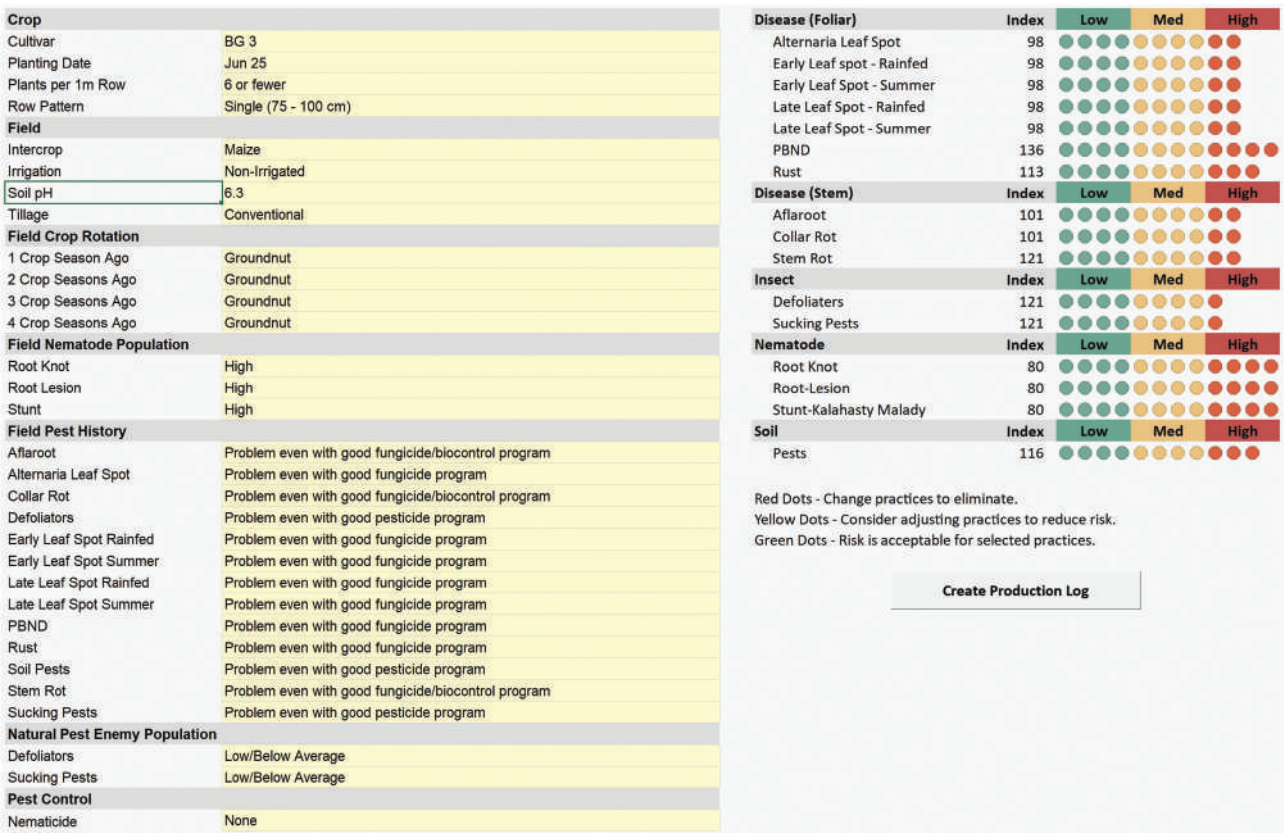


Fig. 15. Risk from pests in the India peanut risk tool with limited inputs and practices.

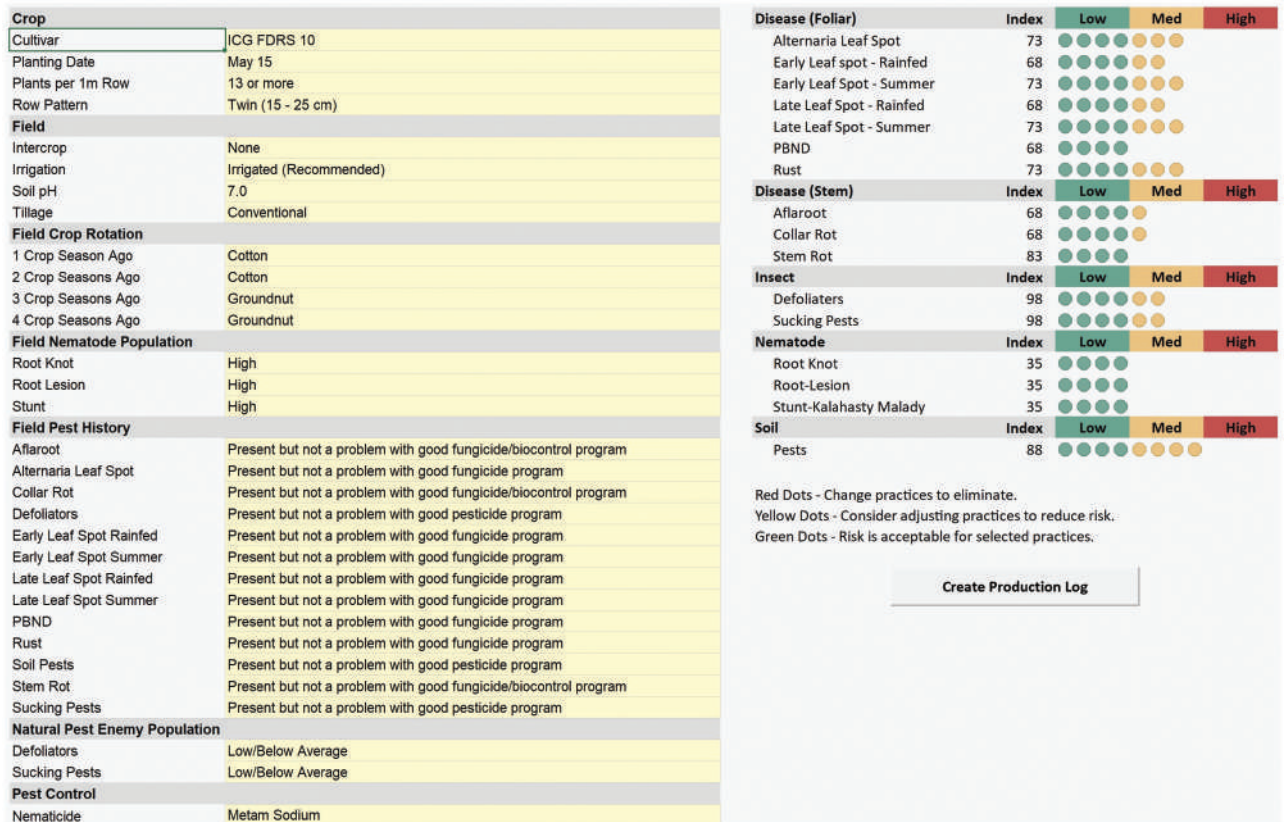


Fig. 16. Risk from pests in the India peanut risk tool with improved inputs and practices.

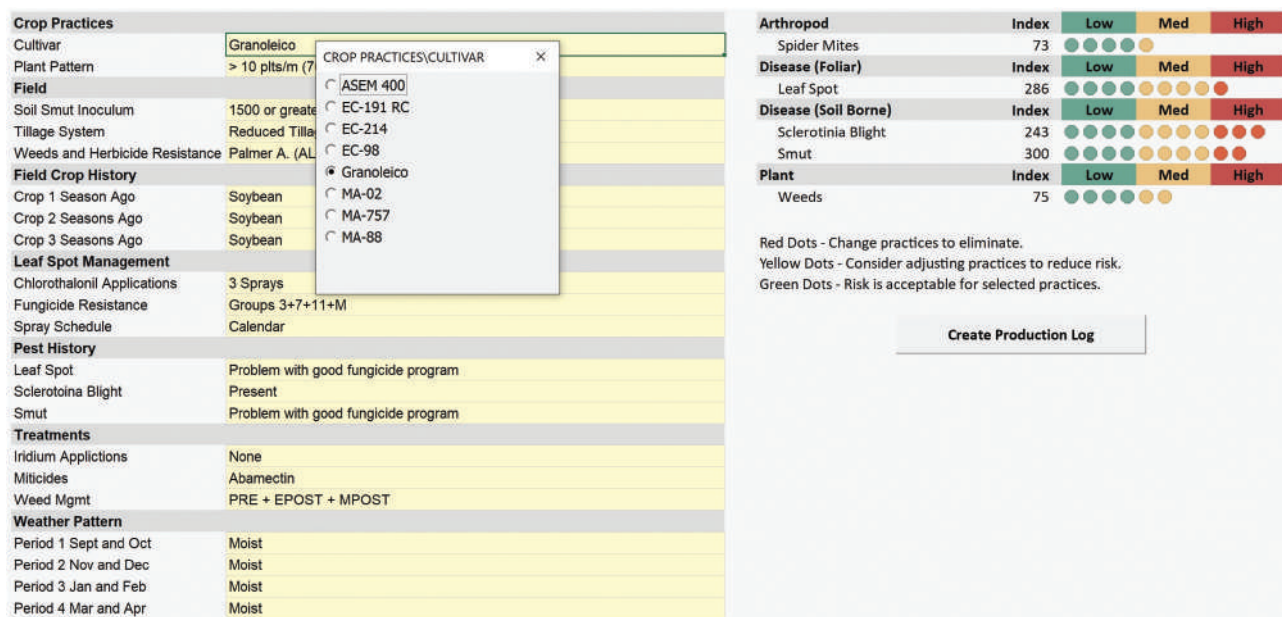


Fig. 17. Drop down menu for varieties in the Argentina peanut risk tool.



Fig. 18. High risk of smut disease in the Argentina peanut risk tool with short rotations and planting a cultivar without resistance to this disease.

The Argentina peanut risk tool includes indices for two-spotted spider mites (*Tetranychus urticae* Koch), peanut smut disease (caused by *Thecaphora frezii* Carranza and Lindquist), early leaf spot disease [caused by *Mycosphaerella arachidicola* W.A. Jenkins (*syn. Passalora arachidicola* W.A. Jenkins)], late leaf spot disease [caused by *Nothopassalora personata* (Berk. & M.A. Curtis) U. Braun, C. Nakash., Videira & Crous], Sclerotinia blight (*Sclerotinia minor* Jagger), and weeds (Fig. 17). Eight cultivars are listed in the drop down menu for Argentina (Fig. 17). Similar to the India peanut risk tool, the current tool for Argentina does not include a cost comparison for management inputs. Risk to smut disease was high when the cultivar Granoleico was planted and the rotation prior to peanut was corn, peanut, and soybean [*Glycine max* (L.) Merr.] (Fig. 18). Adding one more year of corn prior to peanut and planting the cultivar EC-191 RC eliminated risk of smut disease (Fig. 19).

Similar to the NC peanut risk tool (Jordan et al. 2022), the Argentina peanut risk tool includes a drop down menu for resistance to fungicides with respect to leaf spot disease and herbicides (Figs. 20 and 21). Three scenarios associated with risk to two-spotted spider mites are presented in Figs. 22–24. Applying chlorothalonil (a broad spectrum and non-systemic fungicide) three times during the season created greater risk for two-spotted spider mites compared with only one application of this fungicide (Fig. 22). Chlorothalonil and other fungicides can decrease presence of beneficial fungal pathogen *Entomophthora fresenii* Nowakowski, that adversely affects two-spotted spider mites in peanut, especially when moisture is limited (Carner and Canerday 1968, Campbell 1978). Chlorothalonil can also increase risk of Sclerotinia blight (Figs. 22 and 23) but is an effective fungicide for resistance management because it is a multi-site fungicide (Culbreath et al. 2002). Abamectin moderated risk to two-spotted mites (Fig. 24).



Fig. 19. Risk of smut disease in the Argentina peanut risk tool when the number of years between peanut plantings is increased, a smut tolerant variety is planted, and iridium is applied.

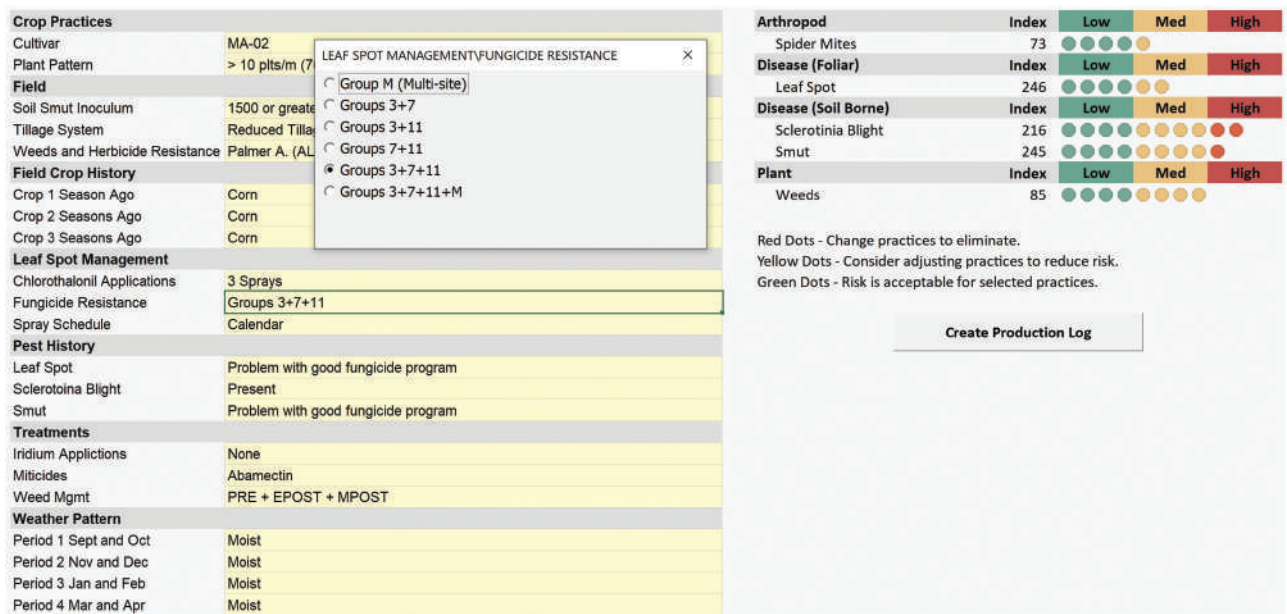


Fig. 20. Drop down menu for fungicide resistance in the Argentina peanut risk tool.

Future Goals for Peanut Risk Tools

The risk tools described for peanut in this article serve as a starting point and are designed for modification as well as expansion to other peanut production areas. In the process of developing these tools several limitations have been identified due to dynamic nature of risk components. First, it is possible that modifications to create tools or portions of tools do not reflect the current knowledge of peanut production systems. Of course, the current versions are not complete in the sense that empirical data sets are a foundation for all of the point designations within and across pest disciplines and individual pests. A considerable amount of the information used in these tools

reflects information provided by practitioners that are not verified by experimental data. However, it is important that risk tools created represent the current knowledge base for peanut production and pest management. When tools are modified there also needs to be a reference file that is considered ‘official’ so that the risk tool is consistent in format and content. With that said, modifications that represent other production areas are a recommended and are a key reason why the initial risk tool was created in Microsoft Excel, especially given the ubiquitous nature of this platform.

A second limitation to the current platform is that it is designed primarily as a planning tool with limited options once the cropping cycle begins. Integrating the tool with other outreach platforms or

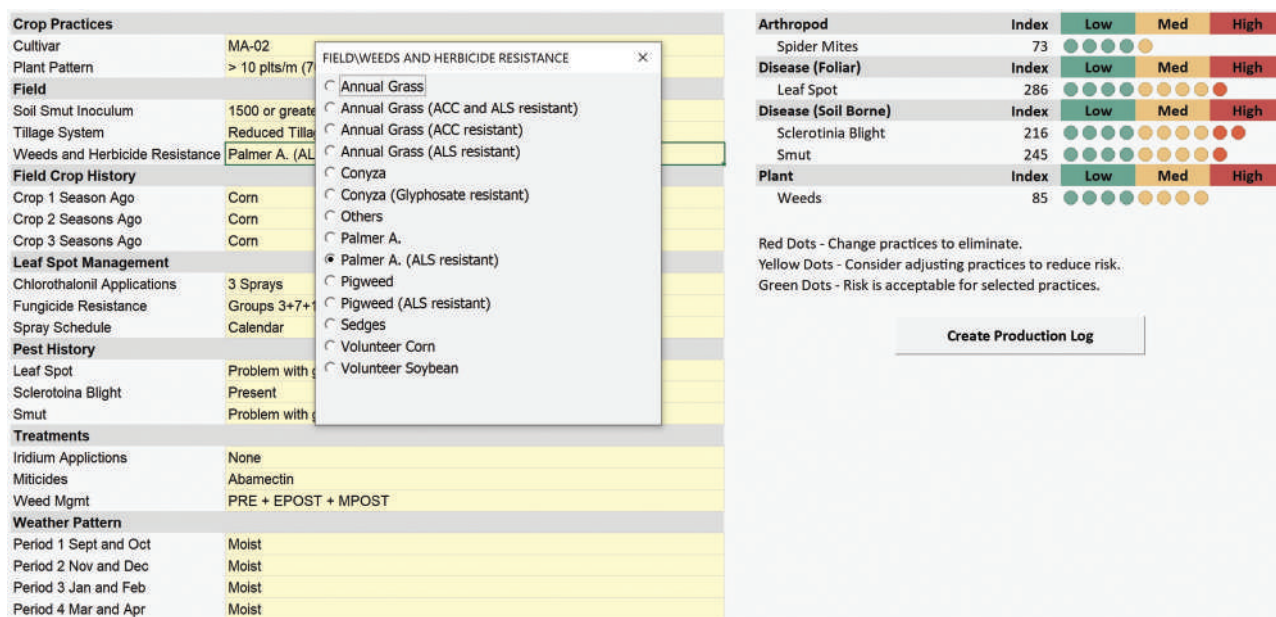


Fig. 21. Drop down menu for weeds based on herbicides resistance in the Argentina peanut risk tool.



Fig. 22. Risk of two-spotted spider mite infestation when three applications of chlorothalonil are made to peanut in the Argentina peanut risk tool.

applications on smartphones would create a time sensitive approach that would be an important advance. If used appropriately, the current risk tool platform decreases the likelihood that practitioners will begin the cropping cycle with elevated risk. The risk tool also serves as a historical record of a field or group of fields by using the production log feature. In this sense the risk tool is future looking. However, greater flexibility in the risk tool for decision-making during the cropping cycle is needed.

A third limitation to these risk tools is the economic component. While this element serves the user by allowing observations of changes in risk linked to production and pest management costs, moving this component of the risk tool toward a true financial comparison using empirical and observation data based on net returns

rather than a simple cost of pest management would be an improvement. Efforts are currently underway in both Ghana and NC to address this limitation by collecting survey data from farmers using categories listed in the risk tool along with weather data, reported yield for that particular cropping cycle, and yield estimates over a longer period of time.

As with all models and tools, validation is needed with these risk tools. As these risk tools are put into practice, adjustments in distribution of points within categories in context of points in other categories need refinement. None-the-less, these risk tools provide a source of greater information exchange on the complicated nature of pest management in peanut for five countries across four continents.



Fig. 23. Risk of two-spotted spider mite infestation when one application of chlorothalonil is made to peanut in the Argentina peanut risk tool.

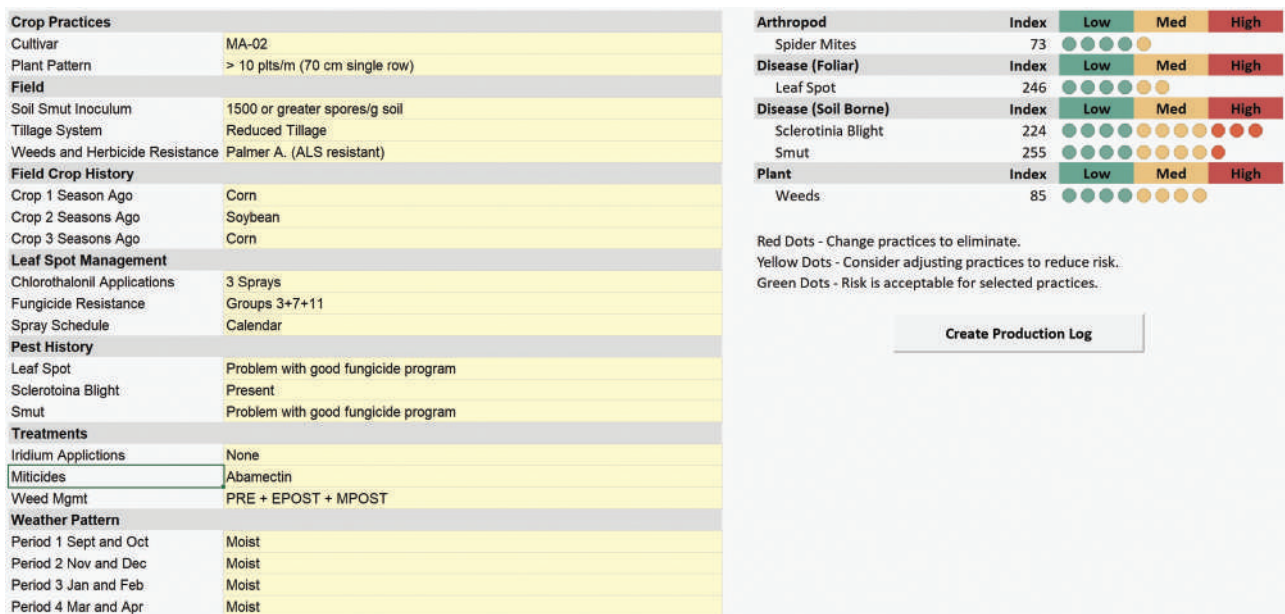


Fig. 24. Risk of two-spotted spider mite infestation when three applications of chlorothalonil are made and abamectin is applied in the Argentina peanut risk tool.

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Conceptualization, D.L.J. and G.S.B.; methodology, D.L.J. and G.S.B.; investigation, D.L.J., G.S.B., B.B.S., R.B., J.N., M.A., R.O., M.B.M., S.A., R.A., W.M.,

J.C., S.M., J.A.B., J.H.M., K.S.J., P.J., T.P.P., H.G., P.H., N.M., and G.M.; resources, D.L.J.; data curation, D.L.J. and G.S.B.; original draft preparation, D.L.J.; review and editing, D.L.J., G.S.B., R.B., J.N., M.A., R.O., M.B.M., S.A., R.A., W.M., J.C., S.M., J.A.P., J.H.M., K.S.J., P.J., T.P.P., H.G., P.H., N.M., G.M., D.H., and J.R.; project administration, D.L.J.; funding acquisition, D.L.J. All authors have read and agreed to the published version of the manuscript.

References Cited

- Appaw, W., W. O. Ellis, R. Akromah, M. B. Mochiah, A. Dankyi, M. Abudulai, D. L. Jordan, R. L. Brandenburg, J. Jelliffe, B. E. Bravo-Ureta, *et al.* 2020. Minimizing aflatoxin contamination in the field, during drying, and in storage in Ghana. *Peanut Sci.* 47: 72–80.
- Abudulai, M., G. Mahama, I. Dzomeku, A. Seidu, I. Sugri, J. A. Nboyine, N. Opoku, M. H. Alhassan, W. Appaw, W. O. Ellis, *et al.* 2020. Evaluation of agricultural practices to increase yield and financial return and

- minimize aflatoxin contamination in peanut in northern Ghana. *Peanut Sci.* 47: 156–162.
- Anonymous. 2022. Peanut Rx. University of Georgia Cooperative Extension Service. Available at: <https://peanuts.caes.uga.edu/extension/peanut-rx.html> (Accessed 5 March 2022).
- Campbell, W. V. 1978. Effect of pesticide interactions on the twospotted spider mite on peanuts. *Peanut Sci.* 5: 83–86.
- Carner, G. R., and T. D. Canerday. 1968. Field and laboratory investigations with *Entomophthora fresenii*, a pathogen of *Tetranuchus* Spp. *J. Econ Entomol.* 61: 956–961.
- Culbreath, A. K., K. L. Stevenson, and T. B. Brenneman. 2002. Management of late leaf spot of peanut with benomyl and chlorothalonil: a study in preserving fungicide utility. *Plant Dis.* 86: 349–355.
- Jordan, D., R. Brandenburg, G. Payne, D. Hoisington, N. Magnan, J. Rhoads, M. Abudulai, K. Adhikari, J. Chen, R. Akromah, *et al.* 2018. Preventing mycotoxin contamination in groundnut cultivation, pp. 181–214. *In* S. Sivasankar, D. Berguinson, P. Gaur, S. Kumar, S. Beebe, and M. Tamó (eds.), *Achieving sustainable cultivation of grain legumes; improving cultivation of particular grain legumes*, vol. 2. Burleigh Dodds Series in Agricultural Science, Burleigh Dodds Science Publishing, Cambridge, UK.
- Jordan, D. L., G. S. Buol, R. L. Brandenburg, B. B. Shew, G. G. Wilkerson, B. R. Lassiter, J. Dunne, A. Gorny, D. Washburn, D. Hoisington, *et al.* 2022. A risk tool and production log created using Microsoft Excel to manage pests in peanut (*Arachis hypogaea*). *J. Int. Pest Manag.* 13: 9:1–16.
- Nigam, S. N., D. L. Jordan, and P. Janila. 2018. Improving cultivation of groundnuts, pp. 155–180. *In* S. Sivasankar, D. Berguinson, P. Gaur, S. Kumar, S. Beebe, and M. Tamó (eds.), *Achieving sustainable cultivation of grain legumes; improving cultivation of particular grain legumes*, vol. 2. Burleigh Dodds Series in Agricultural Science, Burleigh Dodds Science Publishing, Cambridge, UK.
- Stalker, H. T., S. P. Tallury, G. R. Seijo, and S. C. Leal-Bertioli. 2016. Biology, speciation, and utilization of peanut species, pp. 27–66. *In* H. T. Stalker and R. F. Wilson (eds.), *Peanuts: genetics, processing, and utilization*, p. 478. AOCs Press Monograph on Oil Seeds, Elsevier, Amsterdam.
- Valentine, H. 2016. The role of peanuts in global food security, pp. 447–461. *In* H. T. Stalker and R. F. Wilson (eds.), *Peanuts: genetics, processing, and utilization*, p. 478. AOCs Press Monograph on Oil Seeds, Elsevier, Amsterdam.