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Effect of solar radiation and flaxseed supplementation on milk production and fatty acid profile of lactating ewes under high ambient temperature

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ABSTRACT

The objectives of this study were to evaluate the effects of protection from solar radiation and whole flaxseed supplementation on milk yield and milk fatty acid profile in lactating ewes exposed to high ambient temperature. The experiment was conducted during summer and involved 40 ewes divided into 4 groups. The ewes were either exposed (not offered shade) or protected from solar radiation (offered shade). For each solar radiation treatment, ewes were supplemented with whole flaxseed or not. Milk samples from each ewe were collected at the morning and afternoon milking every week, and analyzed for pH, total protein, casein, fat, and lactose content, somatic cell count, and renneting parameters (clotting time, rate of clot formation, and clot firmness after 30 min). At the beginning of the experiment, and then at d 23 and 44, milk samples were analyzed for milk fatty acids using gas chromatography. Flaxseed supplementation significantly increased milk yield, fat, protein, and casein yields, and somatic cell count, and increased fat and lactose contents of milk. A decrease of saturated fatty acids from C6:0 to C16:0 and an increase of C18:1 trans-11 and C18:2 *cis*-9, *trans*-11 was observed in milk from flaxseed-supplemented ewes. Flaxseed supplementation decreased saturated fatty acids content and increased total monounsaturated fatty acids content, the total content of isomers of conjugated linoleic acid, and polyunsaturated fatty acids content in milk. Flaxseed also increased the α -linolenic acid content of milk. As a result, milk from supplemented groups showed an increase in n-3 fatty acid content. Flaxseed supplementation decreased short-chain and medium-chain fatty acids, and increased long-chain fatty acid content of milk. On average, flaxseed supplementation increased the C18:2 cis-9, trans-11/C18:1 trans-11 Δ^9 -desaturase index starting from d 23 of the experiment, in correspondence with the highest C18:2 cis-9, trans-11 content

of milk from flaxseed-supplemented ewes. Flaxseed decreased atherogenic and thrombogenic indices of milk. Protection from solar radiation during summer did not improve yield and composition of ewe milk. Nevertheless, milk from ewes exposed to solar radiation showed decreased long-chain fatty acid and polyunsaturated fatty acids contents, and in particular, decreased vaccenic acid, rumenic acid, and total conjugated linoleic acid contents.

Key words: polyunsaturated fatty acid, sheep milk, high ambient temperature, flaxseed

INTRODUCTION

Sheep milk is entirely destined for dairy products, in particular cheese. Consequently, it is desirable for sheep milk to contain high contents of fat and protein to produce high-quality cheese. During summer in the Mediterranean basin, lactating ewes exposed to temperature-humidity index (**THI**) >80 exhibit a 20% reduction in milk yield (Sevi et al., 2001). The increase in ambient temperatures and the advancement of lactation contribute to a reduction in mobilization of body reserve for milk synthesis, thus inducing a decrease of milk yield and of fat and casein yields. It has been observed that exposure to direct solar radiation adversely influences the fat composition of sheep milk, with a decrease in unsaturated fatty acids and an increase in saturated fatty acids (Sevi et al., 2002).

Milk is an important source of vaccenic acid (C18:1 trans-11, VA), which is an intermediate of rumen biohydrogenation of linoleic and linolenic acids, and which is converted to rumenic acid (C18:2 cis-9,trans-11, RA) by Δ^9 -desaturase in the mammary gland. Rumenic acid is the major isomer of total conjugated linoleic acid (CLA) of milk fat, and both VA and RA are considered health-promoting fatty acids (McGuire and Mc-Guire, 2000). In ewe milk, Nudda et al. (2005) observed a reduction of VA and RA with the advancement of lactation, with the lowest values being recorded during summer.

Under high ambient temperatures, the provision of shaded areas, although contributing to the maintenance

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of ewe thermal balance, did not result in increased ewe production performance (Sevi et al., 2001). However, shaded ewes showed an increase in mammary defense ability and an improvement of milk hygienic quality.

Fat supplementation has been suggested as a strategy to increase the ration energy of the diet and reduce the adverse effects of heat stress on dairy cows; however, fat supplementation can adversely affect both milk yield and fat content (Liu et al., 2008) or only fat content of milk (Drackley et al., 2003). In addition to the improvement of physiological responses and production in heat-stressed animals, the supplementation of fat to the ruminant diet has been used to improve the fatty acid profile of milk. In sheep, several studies have found that the administration of fat supplement can change the fatty acid profile of milk to meet human dietary recommendations by a decrease of saturated and an increase of unsaturated fatty acids (Rotunno et al., 1998; Zhang et al., 2006a,b). However, lactating ewes receiving dietary fat supplementation display decreased milk yield and protein content, with adverse effects on cheese yield and quality (Rotunno et al., 1998; Toral et al., 2010a,b). To the best of our knowledge, no studies have tested the effect of fat supplementation on lactating ewes subjected to heat stress. The administration of whole flaxseed as fat supplement to cows raised under high ambient temperature sustains cow immune functions and improves the yield, composition, and nutritional properties of milk by enhancing the VA, RA, and polyunsaturated fatty acids (**PUFA**) contents of milk (Caroprese et al., 2009, 2010).

We hypothesized that the provision of shaded areas and administration of flaxseed rich in lipid content to the diet of lactating ewes subjected to a hot environment could sustain milk yield and improve the fatty acid profile of milk. Therefore, this work was conducted to evaluate the effects of protection from solar radiation and whole flaxseed supplementation on the production performance of lactating ewes and on milk fatty acid profile under high ambient temperatures.

MATERIALS AND METHODS

Animals and Experimental Design

The experiment lasted 44 d and was conducted during the summer (July and August 2007) at the Segezia research station of the Council for Research and Experimentation in Agriculture (CRA-ZOE). Forty latelactation Sarda ewes (d 202.1 \pm 5.3 of lactation, mean \pm SD) were divided into 4 groups of 10 each, balanced for parity (2.6 \pm 0.7), milk yield (740.5 \pm 9.43 g/d), BW (39.11 \pm 0.26 kg), and BCS (1.61 \pm 0.06). During the study, animals were either exposed to (**EXP**; not

offered shade) or protected from solar radiation (**PRO**; offered shade). Groups were separately reared in external pens of 5×12 m bounded with mesh fence. Shade was provided by $3- \times 8$ -m and 3.5-m-high brickwork rooms adjacent to the open pens; the trough and the crib were located in the external areas. Both exposed and protected animals were fed twice daily with vetch and oat hay and pelleted concentrate (Pecorlat 19, Raggio di sole mangimi SpA, Potenza, Italy; **EXP-C** and **PRO-C** groups); **EXP-F** and **PRO-F** ewes were supplemented with 21% whole flaxseed (Lin Tech, Tecnozoo srl, Torreselle di Piombino Dese, Italy). Ingredients of the diets are reported in Table 1. Dry matter intake was determined for each experimental group by weighing the refusals at 0800, 1200, 1600, and 2000h. Averages of daily DMI were 2.12, 2.09, 2.13, and 2.10 kg/ewe in EXP-C, EXP-F, PRO-C, and PRO-F groups, respectively. Water was available ad libitum for all groups from automatic drinking troughs at any time of day.

During the trial, ambient temperature and relative humidity in protected and exposed area were monitored with thermo-hygrographs (LSI, I-20090 Settala Premenugo-Milano, Italy) placed 1.5 m from the floor. Average of THI was calculated using the Kelly and Bond (1971) formula.

Chemical Composition of Diets

The chemical composition of vetch and oat hay, pelleted concentrate and whole flaxseed was carried out by standard procedures (AOAC, 1990): CP, fat (by ether extract), NDF, ADF, and acid detergent lignin were analyzed. Net energy for lactation was calculated using NRC (2001). The chemical composition of the diets is reported in Table 1.

Table 1. Ingredients and chemical composition of the diets (DM basis)

Item	Control diet	Flaxseed- supplemented diet		
Ingredients, % of DM				
Vetch and oat hay	60.77	60.37		
Concentrate ¹	39.23	31.18		
Whole flaxseed ²	0	8.45		
Chemical composition				
DM, %	90.56	90.79		
Ether extract, % of DM	2.71	4.75		
CP, % of DM	12.89	13.03		
ADF, % of DM	25.51	26.06		
NDF, % of DM	50.14	49.21		
ADL, % of DM	2.99	3.01		
NE_L , $Mcal/kg$	1.38	1.36		

¹Percolat 19 (Raggio di sole mangimi SpA, San Nicola di Melfi, Italy). ²Lin Tech (Tecnozoo srl, Torrreselle di Piombino Dese, Italy).

Lipid extraction for fatty acid analysis of the diet ingredients was carried out according to a modified Folch method (Ardvisson et al., 2009). Separation and quantification of the methyl esters were carried out using a gas chromatograph (Agilent 6890N) equipped with a flame-ionization detector, autosampler, split injection port, and a fused silica capillary column (100 m, internal diameter 0.25 mm, film thickness 0.25 μ m; HP-88 capillary column, Agilent Technologies SpA, Cernusco sul Naviglio, Milan, Italy). Helium was used as the gas carrier (0.42 mL/min). Injector temperature was maintained at 220°C whereas the detector temperature was of 250°C. Fatty acid profile was determined using the following temperature gradient program: initial temperature of 150°C that was increased to 220°C at 1°C/min. Each peak was identified and quantified using pure methyl ester standards: 37-component FAME mixture and methyl trans vaccenate C18:1 trans-11 octadecenoate (Matreya Inc., Pleasant Gap, PA); CLA isomers were identified by comparison of retention times with CLA methyl ester standards: C18:2 cis-9, trans-11 octadecadienoate and methyl C18:2 trans-10, cis-12 octadecadienoate (Matreya Inc.). Results were expressed as grams per 100 grams of total fatty acids analyzed (Table 2).

Sampling and Chemical Analyses of Milk

Ewes were milked twice daily (0700 and 1400 h) in a parlor using a pipeline milking machine (Alfa Laval Agri, Tumba, Sweden). Milk samples from each ewe were collected at the morning and afternoon milkings once a week on the same day throughout the experiment. One aliquot was stored at -20° C for fatty acid analysis. Fresh samples were used for the following chemical analysis: pH (GLP 21 Crison, Barcelona, Spain), total protein, casein, fat, and lactose contents using an infrared spectrophotometer (MilkoScan FT120, Foss Electric, Hillerød, Denmark) according to the International Dairy Federation standard (IDF, 1990), and SCC using a Fossomatic Minor (Foss Electric; IDF, 1995). Evaluation of the renneting parameters (clotting time, rate of clot formation, and clot firmness after 30 min) was measured by a Foss Electric formagraph. The milk coagulating index (CoI) was calculated as the clot firmness to clotting time + rate of clot formation ratio.

Milk Fatty Acid Analysis

At the beginning of the experiment and then at d 23 and 44, each milk sample was analyzed for milk fatty acids. Milk fat was extracted according to the procedure of Luna et al. (2005) and transesterification of fatty acids according to ISO-IDF (2002) procedures, as reported in Caroprese et al. (2010). Briefly, fatty acid methyl esters were separated and measured using a gas chromatograph (Agilent 6890N) equipped with CP-Sil 88 fused-silica capillary column (100 m \times 0.25 mm i.d. with $0.25 \mu m$ film thickness). Operating conditions were a helium flow rate of 1 mL/min, a flame-ionization detector at 260°C, a split-splitless injector at 260°C, and an injection volume of 1 μ L with a split ratio 1:50. The temperature program of the column was set at $100^{\circ}C$ with a subsequent increase to $240^{\circ}C$ at $3.5^{\circ}C/$ min and held for 15 min. Fatty acid were reported as grams per 100 grams of FA. The content of short-chain fatty acids (C4:0 to C12:1, SCFA), medium-chain fatty acids (C14:0 to C16:1, MCFA) and long-chain fatty acids (>C18:0, LCFA) were calculated. Saturated fatty acids (SFA), monounsaturated fatty acids (MUFA), and PUFA were calculated. The desaturase indexes were calculated as described by Kelsev et al. (2003). Atherogenic (AI) and thrombogenic (TI) indices were calculated according to Ulbricht and Southgate (1991) formula:

AI =
$$(C12:0 + 4 \times C14:0 + C16:0)/[\Sigma MUFA + \Sigma PUFA(n-6) and (n-3)];$$

 $TI = (C14:0 + C16:0 + C18:0)/[0.5 \times \Sigma MUFA + 0.5 \times \Sigma PUFA(n-6) + 3 \times \Sigma PUFA(n-3) + (n-3)/(n-6)].$

Table 2. Fatty acid composition (g/100 g of total fatty acids) of feed ingredients

Fatty acid	Vetch and oat hay	$Concentrate^1$	$\begin{array}{c} \text{Whole} \\ \text{flaxseed}^2 \end{array}$
C16:0	20.21	16.09	5.62
C18:0	2.66	2.64	3.67
C18:1 cis-9	14.15	19.05	19.50
C18:2n-6 cis-9, cis-12	20.48	53.73	16.15
C18:3n-3	24.17	6.87	54.03
Other fatty acids	18.35	1.77	1.18

¹Percolat 19 (Raggio di sole mangimi SpA, San Nicola di Melfi, Italy).

²Lin Tech (Tecnozoo srl, Torrreselle di Piombino Dese, Italy).

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Statistical Analysis

All variables were tested for normality using the Shapiro-Wilk test (Shapiro and Wilk, 1965). Data were processed using ANOVA for repeated measures (SAS Institute, 1999), with exposure to solar radiation (non-repeated factor), diet (nonrepeated factor), the time of sampling, and their interactions as repeated factors. Individual animals were nested within treatments. Where significant effects were found (P < 0.05), Student's *t*-test was used to locate significant differences between means.

RESULTS

Meteorological Data

Meteorological data are reported in Figure 1. The averages of THI during daytime were 2.3 to 6.3 units higher in the exposed areas than in the protected areas. In contrast, during nighttime, averages of THI were 5.4 to 7.8 units higher in the protected areas than in the exposed areas.

In the exposed areas, the THI values were near 77, on average, during daytime and 63 during nighttime. In the protected areas, THI was 73, on average, during daytime and was more than 6 points higher than in the exposed areas during nighttime.

Milk Yield and Quality

Flaxseed supplementation increased milk yield and fat, protein, and casein yields (P < 0.001; Table 3). Milk and fat yields were affected by time of sampling and the interaction of diet × time of sampling (P < 0.001 and P < 0.05, respectively); flaxseed increased milk and fat yields starting from wk 2 of milk sampling and thereafter (Figure 2). Flaxseed supplementation increased fat and lactose contents of milk (P < 0.01and P < 0.001, respectively), whereas exposure to solar radiation resulted in increased contents of fat and casein (P < 0.05) and lactose (P < 0.001) in milk. Fat and lactose contents were affected by time of sampling with fluctuations connected to milk yield (P < 0.001).

Flaxseed supplementation shortened milk clotting time (17.55 vs. 15.90 min \pm 0.47), and increased curd firmness (51.50 vs. 56.14 mm \pm 1.46) and milk coagulation index (3.25 vs. 3.63 \pm 0.13; P < 0.05), with EXP-F milk displaying the best coagulation parameters (Table 4). Flaxseed supplementation decreased SCC of milk by about 57%, with the lowest cell count being found in EXP-F ewes (P < 0.001).

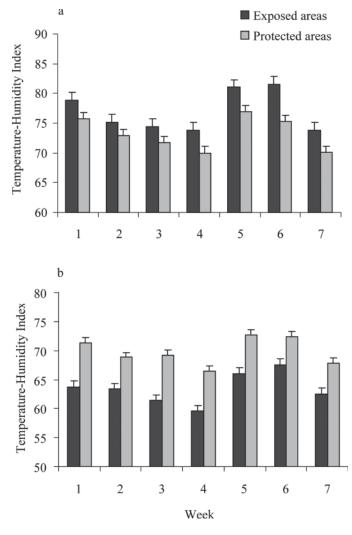


Figure 1. Means \pm SD of temperature-humidity index (THI) in exposed and protected areas measured during (a) daytime (0800 to 2000 h) and (b) nighttime (2000 to 0800 h).

Milk Fatty Acids Composition

The fatty acid composition of milk is reported in Table 5. A decrease in SFA from C6:0 to C16:0 (P < 0.001) was observed in milk from flaxseed-supplemented ewes. Flaxseed supplementation also led to a decrease of C17:0 (P < 0.01) and C17:1 (P < 0.001). An effect of time was observed because the aforementioned fatty acids began to decrease in the flaxseed-supplemented groups starting on d 23 of the experiment. Exposure to solar radiation increased C16:0 (23.31 vs. 24.26 \pm 0.25 g/100 g of total fatty acids; P < 0.01), and the highest C16:0 content was observed in EXP-C milk (P < 0.05). In contrast, C18:0 (stearic acid) was higher in milk from ewes receiving flaxseed supplementation

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Item		Treatment					$Effects, P-value^1$		
	EXP-C	EXP-F	PRO-C	PRO-F	SEM	\mathbf{SR}	Diet	Time	
Yield, g/d									
Milk	599.14^{b}	$712.43^{\rm a}$	623.00^{b}	728.57^{a}	15.08	NS	***	***	
Fat	41.82^{b}	$50.72^{\rm a}$	41.66^{b}	51.39^{a}	1.02	NS	***	**	
Protein	$34.51^{\rm b}$	$40.21^{\rm a}$	$35.29^{ m b}$	41.03^{a}	0.86	NS	***	***	
Casein	27.27^{b}	$32.34^{\rm a}$	27.11^{b}	32.53^{a}	0.37	NS	***	***	
Milk composition, %									
Fat	7.09^{a}	7.18^{a}	6.74^{b}	7.15^{a}	0.20	*	**	***	
Protein	5.81	5.66	5.69	5.66	0.16	NS	NS	NS	
Casein	4.59^{a}	4.56^{a}	4.36^{b}	4.50^{ab}	0.14	*	NS	NS	
Lactose	4.64^{b}	4.86^{a}	4.33°	4.66^{b}	0.07	***	***	***	

Table 3. Milk yield and composition in ewes when exposed to solar radiation and fed control diet (EXP-C), exposed to solar radiation and fed flaxseed (EXP-F), protected from solar radiation and fed control diet (PRO-C), and protected from solar radiation and fed flaxseed (PRO-F)

^{a-c}Means within a row followed by different letters are significantly different at P < 0.05.

 ${}^{1}SR = solar radiation; Time = time of sampling.$

*P < 0.05; **P < 0.01; ***P < 0.001.

than in nonsupplemented ewes (P < 0.001), starting on d 23 of the trial (P < 0.001). As a result, the diet based on flaxseed supplementation reduced the SFA content of milk (P < 0.001), starting on d 23 of the experiment (P < 0.05, Figure 3).

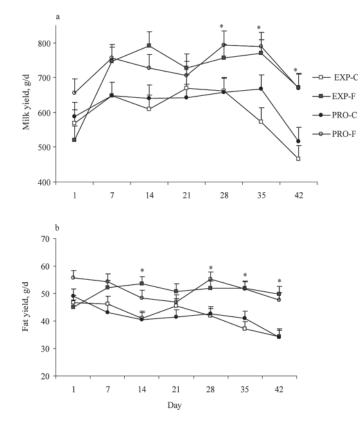


Figure 2. Temporal changes in milk and fat yield in ewes when exposed to solar radiation and fed control diet (EXP-C), exposed to solar radiation and fed flaxseed (EXP-F), protected from solar radiation and fed control diet (PRO-C), or protected from solar radiation and fed flaxseed (PRO-F). Differences between the groups at each time point are represented by * (P < 0.05).

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As expected, the diet supplemented with flaxseed influenced the levels of C18:1 trans-9, C18:1 trans-11, and C18:1 cis-9 in milk (P < 0.001). In addition, milk from EXP ewes showed lower average C18:1 trans-9 than milk from PRO ewes (0.38 vs. 0.43 \pm 0.02 g/100 g of total fatty acids; P < 0.05). The C18:1 trans-11 (VA) content in milk was lower in EXP than in PRO groups at the beginning of the experiment. Vaccenic acid content in milk from supplemented ewes increased starting on d 23 of the trial (Figure 4; P < 0.001).

The total MUFA content of milk from ewes fed flaxseed also showed a significant increase (P < 0.001, Figure 3) starting from d 23 (P < 0.001). Flaxseed supplementation resulted in increased levels of PUFA (P < 0.001) starting from d 23 in the exposed groups. Fat supplementation influenced SCFA, MCFA, and LCFA contents of milk (P < 0.001) by decreasing SCFA and MCFA and increasing LCFA content of milk. In addition, milk from EXP-C had higher content of MCFA than milk from PRO-C (Figure 3).

Flaxseed supplementation increased C18:2 trans-9, trans-12 and C18:2 cis-9, trans-11 contents of milk (P < 0.001). At the beginning of the experiment (Figure 4), the C18:2 *cis*-9,*trans*-11 content was higher in milk of ewes protected from solar radiation (P < 0.01), aside from flaxseed supplementation; subsequently, on d 23, the flaxseed-supplemented groups showed an increase of C18:2 cis-9, trans-11. On average, flaxseed supplementation led to an increase in C18:2 cis-9, trans-11 of about 80% in milk from ewes exposed to solar radiation and of 63% in milk of ewes protected from solar radiation. Consequently, an increase of the total content of CLA isomers in groups subjected to the flaxseed diet (EXP-F, PRO-F) was observed, starting on d 23 of the trial (Figure 4). Milk from EXP-F ewes showed, on average, a 57% higher content of CLA isomers than

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Table 4. pH, renneting parameters, and SCC in milk of ewes when exposed to solar radiation and fed control diet (EXP-C), exposed to solar radiation and fed flaxseed (EXP-F), protected from solar radiation and fed control diet (PRO-C), and protected from solar radiation and fed flaxseed (PRO-F)

		Treatment					Effects, P -value ¹		
Item	EXP-C	EXP-F	PRO-C	PRO-F	SEM	SR	Diet	Time	
H	6.49	6.47	6.51	6.51	0.01	NS	NS	***	
Clotting time, min	$17.44^{\rm a}$	$14.51^{\rm b}$	$17.66^{\rm a}$	$17.29^{\rm a}$	0.66	*	*	NS	
Rate of clot formation, min	1.27	1.53	1.40	1.43	0.15	NS	NS	NS	
Clot firmness, mm	51.96^{b}	59.75^{a}	51.04^{b}	52.52^{b}	2.07	*	*	NS	
CoI ²	3.29^{b}	4.05^{a}	3.21^{b}	3.22^{b}	0.44	*	*	NS	
SCC, $\times 10^3$ cells/mL	$957.49^{\rm a}$	354.46°	814.14^{a}	655.99^{b}	52.91	NS	***	**	

^{a-c}Means within a row followed by different letters are significantly different at P < 0.05.

 ${}^{1}SR = solar radiation; Time = time of sampling.$

²Milk coagulating index (CoI) = clot firmness to clotting time + rate of clot formation ratio.

*P < 0.05; **P < 0.01; ***P < 0.001.

Table 5. Least square means of milk fatty acid composition (g/100 g of total fatty acids) of ewes when exposed to solar radiation and fed control diet (EXP-C), exposed to solar radiation and fed flaxseed (EXP-F), protected from solar radiation and fed control diet (PRO-C), and protected from solar radiation and fed flaxseed (PRO-F)

		Treat		Effects, P -value ¹				
Fatty acid	EXP-C	EXP-F	PRO-C	PRO-F	SEM	SR	Diet	Time
C4:0	4.53	4.35	4.30	4.38	0.13	NS	NS	*
C6:0	2.29^{a}	1.98^{b}	2.30^{a}	1.98^{b}	0.14	NS	***	***
C8:0	1.74^{a}	$1.44^{\rm b}$	1.81^{a}	1.43^{b}	0.13	NS	***	***
C10:0	5.07^{a}	4.04^{b}	5.36^{a}	4.02^{b}	0.37	NS	***	***
C12:0	3.14^{a}	2.52^{b}	3.38^{a}	2.68^{b}	0.15	NS	***	***
C14:0	10.69^{a}	8.86^{b}	10.72^{a}	9.48^{b}	0.34	NS	***	**
C14:1 cis-9	0.51	0.72	0.53	0.48	0.13	NS	NS	NS
C15:0	$1.04^{\rm a}$	$0.90^{ m b}$	1.04^{a}	0.92^{b}	0.03	NS	***	***
C16:0	26.07^{a}	22.44°	24.27^{b}	22.35°	0.54	**	***	NS
C16:1 cis-9	1.32^{a}	$0.95^{ m b}$	1.29^{a}	$0.95^{ m b}$	0.08	NS	***	***
C17:0	$0.50^{\rm a}$	0.46^{ab}	0.50^{a}	$0.43^{ m b}$	0.02	NS	**	***
C17:1	0.27^{a}	0.18^{b}	0.26^{a}	0.19^{b}	0.01	NS	***	***
C18:0	8.78^{b}	$11.92^{\rm a}$	$9.12^{\rm b}$	11.20^{a}	0.48	NS	***	***
C20:0	0.26^{ab}	$0.24^{\rm ab}$	$0.27^{\rm a}$	$0.23^{\rm b}$	0.01	NS	***	***
C20:1	0.19^{b}	$0.54^{\rm a}$	$0.24^{\rm b}$	$0.54^{\rm a}$	0.04	NS	***	*
C18:1 trans-9	0.31^{b}	0.45^{a}	$0.37^{ m b}$	$0.50^{\rm a}$	0.03	*	***	NS
C18:1 trans-11	$2.68^{\rm b}$	$4.00^{\rm a}$	3.18^{b}	3.77^{a}	0.00	NS	***	***
C18:1 <i>cis</i> -9	$25.42^{\rm b}$	$27.89^{\rm a}$	$25.43^{\rm b}$	$28.60^{\rm a}$	0.63	NS	***	NS
C18:2 trans-9.trans-12	$0.11^{\rm b}$	$0.20^{\rm a}$	$0.12^{\rm b}$	$0.20^{\rm a}$	0.01	NS	***	***
C18:2 cis-9, cis-12	2.98^{ab}	$2.84^{\rm bc}$	3.11^{a}	2.75°	0.01	NS	***	***
C18:2 cis-9,trans-11	$0.442^{\rm b}$	$0.792^{\rm a}$	$0.510^{\rm b}$	$0.835^{\rm a}$	0.03	NS	***	NS
C18:2 trans-10, cis-12	$0.442 \\ 0.164^{\rm ab}$	$0.169^{\rm ab}$	$0.172^{\rm a}$	$0.161^{\rm b}$	0.03 0.004	NS	NS	**
C18.2 <i>trans</i> -10, <i>cis</i> -12 C18:3n-3	$1.05^{\rm b}$	1.65^{a}	$1.20^{\rm b}$	1.55^{a}	0.004	NS	***	***
C20:3n-3	0.01	0.02	0.02	0.01	0.08	NS	NS	**
C20:5n-3, eicosapentaenoic acid	$0.01 \\ 0.005^{\rm b}$	$0.02 \\ 0.037^{a}$	$0.02 \\ 0.004^{\rm b}$	$0.01 \\ 0.030^{a}$	0.01 0.004	NS	***	*
	0.003 0.012	0.037	$0.004 \\ 0.004$	0.030 0.010	$0.004 \\ 0.003$	*	NS	*
C22:6n-3, docosapentaenoic acid C20:3n-6	0.012 0.10^{ab}	0.013 0.08^{b}	$0.004 \\ 0.10^{a}$	$0.010 \\ 0.07^{\circ}$		NS	NS ***	***
C20:3n-6 C20:4n-6	$0.10 \\ 0.19^{a}$	$0.08 \\ 0.15^{\rm b}$	$0.10 \\ 0.19^{\rm a}$	$0.07 \\ 0.12^{\rm c}$	0.01	NS	***	*
	$0.19 \\ 0.61^{\rm b}$	$0.15 \\ 0.96^{a}$	0.19 0.68^{b}	1.00^{a}	$0.01 \\ 0.04$	NS	***	NS
Total conjugated linoleic acid		14.34^{b}		$1.00 \\ 14.49^{b}$			***	*
Short-chain fatty acids	16.78 ^a	14.34° 33.91°	$17.16^{\rm a} \\ 37.92^{\rm b}$		0.52	NS	***	
Medium-chain fatty acids	39.68 ^a			34.62°	0.51	NS	***	NS
Long-chain fatty acids	$43.53^{\rm b}$	51.75 ^a	44.92^{b}	51.32^{a}	94.00	NS	***	$_*^{\rm NS}$
Saturated fatty acids	64.17 ^a	59.21 ^b	63.13^{a}	59.14 ^b	0.71	NS	***	
Monounsaturated fatty acids	$30.70^{\rm b}$	34.73 ^a	$31.31^{\rm b}$	$35.07^{\rm a}$	1.03	NS		$\underset{***}{\operatorname{NS}}$
Polyunsaturated fatty acids	5.04 ^b	$5.93^{\rm a}$	5.43 ^{ab}	5.74^{a}	0.20	NS	***	
n-3	1.07^{b}	$1.72^{\rm a}$	1.23^{b}	$1.60^{\rm a}$	0.11	NS	***	***
n-6	3.98	4.24	4.22	4.15	0.13	NS	NS	**
n-3/n-6	0.27^{b}	0.40^{a}	0.29^{b}	0.38^{a}	0.02	NS	***	***

^{a-c}Means within a row followed by different letters are significantly different at P < 0.05.

 ${}^{1}SR = solar radiation; Time = time of sampling.$

 $^{*}P < 0.05; \, ^{**}P < 0.01; \, ^{***}P < 0.001.$

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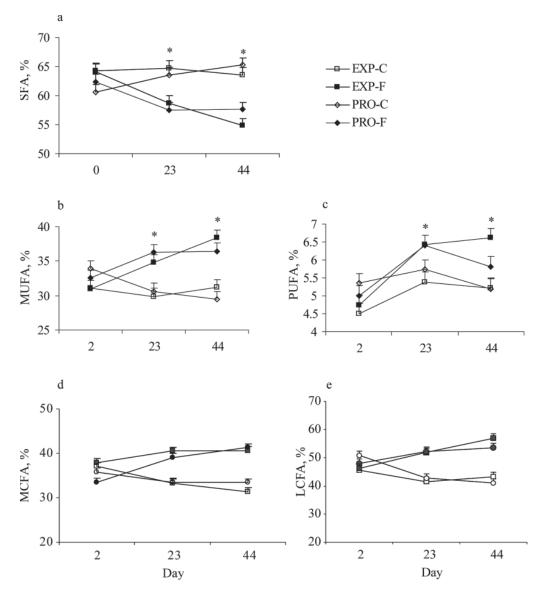


Figure 3. Temporal changes in saturated fatty acids (SFA), monounsaturated fatty acids (MUFA), polyunsaturated fatty acids (PUFA), medium-chain fatty acids (MCFA), and long-chain fatty acids (LCFA) content in ewes when exposed to solar radiation and fed control diet (EXP-C), exposed to solar radiation and fed flaxseed (EXP-F), protected from solar radiation and fed control diet (PRO-C), protected from solar radiation and fed flaxseed (PRO-F). Differences between the groups at each time point are represented by * (P < 0.05).

EXP-C milk, and PRO-F milk displayed a 47% higher content of CLA than PRO-C milk.

Flaxseed supplementation also positively influenced the α -linolenic acid content of milk (C18:3n-3; P < 0.001), which began to increase in the supplemented groups starting from d 23 (P < 0.001; Figure 4). The diet based on flaxseed also resulted in an increase of eicosapentaenoic acid (C20:5n-3; P < 0.001). As a result, milk from supplemented groups showed an increase in n-3 PUFA content (P < 0.001). In particular, increases in n-3 PUFA content of about 60% in milk from EXP-F compared with EXP-C ewes, and of 30% in milk from PRO-F compared with PRO-C ewes were observed. The increase of milk n-3 PUFA content in the flaxseed-supplemented groups also resulted in a higher n-3/n-6 ratio in EXP-F and PRO-F milk compared with EXP-C and PRO-C milk (P < 0.001).

On average, flaxseed supplementation positively influenced the C18:2 *cis*-9,*trans*-11/C18:1 *trans*-11 Δ^9 -desaturase index (P < 0.001; Table 6), starting on d 23 of the experiment (P < 0.001), in correspondence with the highest C18:2 *cis*-9,*trans*-11 content of milk from

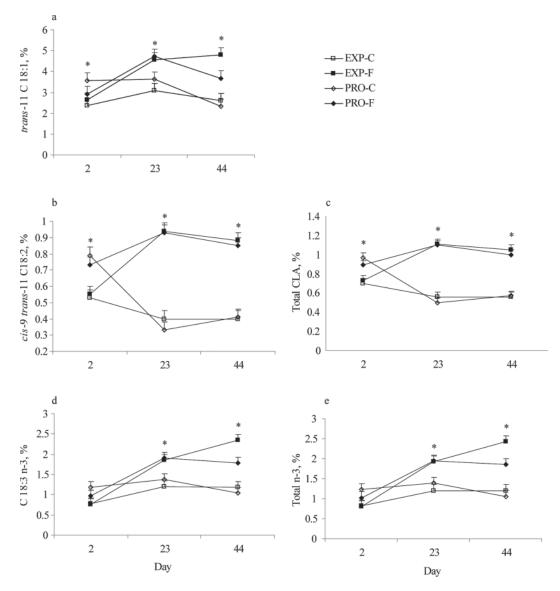


Figure 4. Temporal changes in C18:1 trans-11, C18:2 cis-9, trans-11, total conjugated linoleic acid (CLA), C18:3 n-3, and total n-3 polyunsaturated fatty acids (PUFA) in ewes when exposed to solar radiation and fed control diet (EXP-C), exposed to solar radiation and fed flaxseed (EXP-F), protected from solar radiation and fed control diet (PRO-C), protected from solar radiation and fed flaxseed (PRO-F). Differences between the groups at each time point are represented by * (P < 0.05).

EXP-F and PRO-F ewes. Flaxseed supplementation decreased the atherogenic and thrombogenic indices of milk (P < 0.001).

DISCUSSION

Sheep are thought to be among the most resistant ruminants to climatic extremes (being less affected by extremes), especially to high ambient temperatures. Experiments conducted on Comisana ewes by Sevi et al. (2001, 2002) demonstrated a reduction in milk yield after exposure to average daily temperatures of 35°C, even for short periods, or after prolonged ewe exposure to mean ambient temperatures of 30° C. Sarda ewes are highly sensitive to high temperatures. Peana et al. (2007) found that in Sarda ewes, milk yield could be reduced by 15% if the maximum ambient temperatures are higher than 21 to 24°C, and by 20% if the minimum temperature changed from 9 to 12°C to 18 to 21°C. In light of previous findings, we can hypothesize that the effect of higher maximum temperatures on the milk yield of ewes exposed to solar radiation during daytime was counterbalanced by the effect of higher minimum temperatures acting on protected ewes during nighttime. This resulted in failure to find differences in milk yield between shaded and nonshaded ewes. In addition,

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Table 6. Least square means of desaturase and nutritional indexes of milk in ewes when exposed to solar radiation and fed control diet (EXP-C), exposed to solar radiation and fed flaxseed (EXP-F), protected from solar radiation and fed control diet (PRO-C), and protected from solar radiation and fed flaxseed (PRO-F)

		Treatment					Effects, P -value ¹		
Item	EXP-C	EXP-F	PRO-C	PRO-F	SEM	SR	Diet	Time	
Desaturase index ² Conjugated linoleic acid (CLA) Nutritional index ³	0.15^{b}	$0.18^{\rm a}$	0.14^{b}	$0.19^{\rm a}$	0.01	NS	***	***	
Atherogenic index Thrombogenic index	2.04^{a} 2.32^{a}	$1.531^{ m b}\ 1.93^{ m b}$	${1.96^{ m a}}\over{2.18^{ m a}}$	$1.60^{ m b}$ $1.93^{ m b}$	$\begin{array}{c} 0.06 \\ 0.06 \end{array}$	NS NS	*** ***	NS ***	

^{a,b}Means within a row followed by different letters are significantly different at P < 0.05.

 ${}^{1}SR = solar radiation; Time = time of sampling.$

²CLA desaturase index was calculated as [cis-9, trans-11 CLA]/[cis-9, trans-11 CLA + trans-11 18:1]; Kelsey et al. (2003).

³Atherogenic and thrombogenic indexes were calculated using Ulbricht and Southgate (1991) formulas.

***P < 0.001.

a physiological decrease in milk yield in ewes occurs during summer with late lactation, which could completely or partly hide the negative effect of high temperatures on milk production.

Flaxseed supplementation increased milk yields by about 18% in both exposed and protected ewes. In particular, in the exposed group, milk yield was higher at the end of the experiment than at the beginning. Our results are in line with those obtained by Zhang et al. (2006a,b), whereas flaxseed supplementation had no effects on milk yield in cows (Caroprese et al., 2010). The increase in milk yield observed in supplemented groups is not easy to explain, given that control and flaxseedsupplemented diets had very similar NE_L, whereas in the Zhang et al. (2006a,b) experiments, the increase in ewe milk yield reflected the higher energy content of the flaxseed-supplemented diet. The possibility that increased lactose resulted in high levels of production in ewes receiving flaxseed cannot be excluded.

The increase in milk yield in flaxseed-supplemented ewes led to increases in fat, protein, and casein yields. Flaxseed in the diet increased fat content of milk; the effects of lipid supplementation on milk fat are controversial depending on the type of supplemented fat, the level of supplementation, and the species. In goat and sheep milk, no effects of extruded linseed were found on fat content (Nudda et al., 2006; Gómez-Cortés et al., 2009). In previous experiments, the supplementation of 8% rumen-protected fat to sheep or whole flaxseed to cows contributed to increase the fat content of milk (Rotunno et al., 1998; Caroprese et al., 2010). The increase in fat and casein contents of milk observed in EXP-C ewes can be attributed to the reduction of milk yield registered in that group. Dietary fat usually decreases milk protein synthesis because of a reduction in amino acid availability to the mammary gland (Rotunno et al., 1998; Pulina et al., 2006). Only a few studies have reported the effects of supplemental fat to sheep diet on milk protein composition (Goulas et al., 2003; Zhang et al., 2006a,b). In our study, flaxseed supplementation had a positive effect on casein content of milk, which was associated with better rheological properties of milk from ewes fed flaxseed. It is well known that milk coagulation is strictly connected to milk composition and SCC (Albenzio et al., 2004). Milk from ewes fed flaxseed was characterized by improved milk composition and lower SCC, resulting in better coagulation properties. Few studies focused on the effects of direct solar radiation or fat supplementation on milk SCC (Sevi et al., 2001; Liu et al., 2008; Caroprese et al., 2010) and did not show a direct effect of both factors on SCC.

Different dietary strategies to improve the fatty acid profile of ewe milk have been assessed, with an improvement of the saturated to unsaturated fatty acid ratio (Rotunno et al., 1998; Zhang et al., 2006a,b). The administration of an appropriate dietary fat supplement can be effective in increasing the content of unsaturated fatty acids of ewe milk especially in late lactation, when a reduction of mobilization of body reserves for milk synthesis occurs (Rotunno et al., 1998). Accordingly, in the present study, flaxseed supplementation increased both the MUFA and PUFA contents of sheep milk, together with a reduction of SFA, SCFA, and MCFA. The increase in PUFA in the mammary gland could explain the reduction of both SCFA and MCFA in milk from flaxseed-supplemented ewes; previous findings confirmed that PUFA compete for esterification with SCFA and MCFA synthesized in the mammary gland, causing a feedback inhibition of lipogenic enzymes (Gómez-Cortés et al., 2008, 2009). The decrease of C16:0 observed in the milk from supplemented ewes can be ascribed both to the inhibition of its synthesis and to the reduced C16:0 content in flaxseed compared with the C16:0 content of vetch and oat hay and concentrate. In addition, in a previous experiment, an increase of the proportions of C12:0, C14:0, and C16:0 fatty acids in milk from ewes exposed to direct solar radiation during summer was observed (Sevi et al., 2002).

Diet is responsible for C18 fatty acid content in milk because the mammary gland enzymatic systems cannot extend the carbon chain from C16 to C18. Therefore, a major concentration of C18 fatty acids, especially in milk of ewes fed a flaxseed supplement, is due to the high content of C18:3n-3 in such seed (Zhang et al., 2006a,b). The high C18:0 concentration observed in milk of ewes subjected to flaxseed supplementation is the result of the biohydrogenation of unsaturated C18 fatty acids in the rumen (Lock and Garnsworthy, 2003). Generally, high values in milk C18:1 fatty acids arise from ruminal biohydrogenation of C18:2 and C18:3 fatty acids, and from desaturation of C18:0 in the mammary gland by the action of the Δ^9 -desaturase enzyme (Dhiman et al., 2000). The C18:1 cis-9 (oleic acid) in milk is a product arising from the isomerization of blood C18:0 (stearic acid) by Δ^9 -desaturase activity in the mammary gland (Gómez-Cortés et al., 2009), whereas VA is produced in the rumen and then absorbed by the gut to be transported in the mammary gland, where it is used for endogenous synthesis of RA, through stearoyl-CoA desaturase activity (Luna et al., 2008b). Flaxseed supplementation increased the content of milk VA by about 50% in the ewes exposed to solar radiation and by 18% in the ewes protected from solar radiation. The importance of increasing VA in milk arises from the observation that VA is the substrate for the production of RA, not only in the mammary gland of ruminant but also in muscle and adipose tissues in humans (Bauman et al., 2006). The accumulation of VA resulted in increased concentrations of RA in milk from ewes supplemented with flaxseed.

Sevi et al. (2002) found a reduction of both MUFA and PUFA and an increase in C12:0 to C16:0 fatty acids in milk from ewes exposed to direct solar radiation compared with shaded ewes during summer. Accordingly, at the beginning of this study, the content of MCFA was higher, and the contents of LCFA, some PUFA, such as C18:1 trans-11 and C18:2 cis-9, trans-11, and total CLA were lower in milk from ewes exposed to solar radiation. High ambient temperature can reduce the rate of digesta passage in the rumen (Silanikove, 1992), thus allowing an extensive hydrogenation of unsaturated fatty acids, and a consequent reduction of MUFA and PUFA in milk. Subsequently, the administration of flaxseed changed the fatty acid composition during the experiment, with higher content of VA, RA, and of total CLA in milk from supplemented ewes, apart from the direct exposure to solar radiation. The

administration of whole flaxseed to heat-stressed cows in a previous experiment did not result in a subsequent increase in milk C18:3 (Caroprese et al., 2010), whereas in the present experiment an increase of C18:3 in milk from ewes fed whole flaxseed was observed, according to Zhang et al. (2006a,b). As a consequence, different utilization of whole flaxseed in the rumen and then in the mammary gland has to be hypothesized in cows and sheep when they are subjected to heat stress. The fatty acid profile of sheep milk is mainly influenced by diet, even if other factors may account for variations of milk fatty acid profile, such as stage of lactation, parity, and season (De La Fuente et al., 2009). In particular, the reduction of C18:3 availability in the diet during summer resulted in a seasonal decrease of C18:3, VA, and CLA content in milk of Sarda sheep (Nudda et al., 2005). Flaxseed supplementation was effective in reducing the adverse effects of both heat stress and the advancement of lactation on milk fatty acids composition, resulting in evident increases in milk contents of VA, RA, total CLA, and C18:3n-3, starting from d 23 of the experiment until the end of the experiment. In milk from ewes fed flaxseed, contents of VA, RA, and total CLA increased during the experiment, whereas those of C18:3n-3 and total PUFA n-3 fatty acids were higher at the end than at the beginning of the experiment in milk from both EXP-F and PRO-F. The persistency of the positive effects of flaxseed supplementation on milk VA, RA, and total CLA in this study confirmed previous findings by Gómez-Cortés et al. (2009), whereas the evolution of C18:3n-3 suggested further investigation on the use of whole flaxseed to increase the content of PUFA n-3 in milk from lactating ewes in advanced stages of lactation. The increases of VA, RA, and total CLA were more marked in milk from ewes exposed to solar radiation than in milk from shaded ewes, suggesting the advantage of using whole flaxseed supplement to improve the fatty acid profile in milk of ewes subjected to high ambient temperatures.

The decrease of C12:0, C14:0, and C16:0 fatty acids caused a consequent reduction of atherogenic and thrombogenic indices in milk of ewes subjected to flaxseed supplementation. Milk fat with high atherogenic index values has detrimental effects on human health. It is known that in the human diet, SFA contributes to coronary diseases, whereas the unsaturated fatty acids, including CLA (rumenic acid), MUFA (in particular, oleic acid) and PUFA, have a protective effect against cardiovascular diseases (Williams, 2000). As it has been demonstrated that fatty acid composition of sheep cheese is dependent on the fatty acid composition of raw milk (Nudda et al., 2006), the improvement of fatty acid profile in milk can result in naturally enriched dairy products.

CONCLUSIONS

Protection from solar radiation under high ambient temperatures did not increase the yield and improve the composition of ewe milk. Nevertheless, milk from ewes exposed to solar radiation showed a worsening of fatty acid profile in terms of decreased LCFA and PUFA content, and in particular, decreased VA, RA, and total CLA contents. Flaxseed supplementation, instead, had several positive effects on the yield and quality of ewe milk. Indeed, it increased the yield and composition of milk, leading to a reduction of SCC, an increase of fat content, and better coagulation properties of milk. Sheep farmers can benefit from enhanced ewe milk yield in summer because at that time dairy factories typically stop collecting milk from sheep farms as a result of decreased milk production and deteriorating coagulation behavior. Moreover, flaxseed supplementation resulted in an enhanced fatty acid profile of milk, as demonstrated by the increase in PUFA, VA, RA, total CLA, and n-3 PUFA, and the decrease in SFA, SCFA, and MCFA. This finding is of great interest both because consumer demand for dairy products naturally enriched in healthy components has grown and because the fatty acid profile of ewe milk undergoes a marked worsening during summer.

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