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Essential Oils from the Leaves of Six Medicinal Plants of Nigeria[#]

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The chemistry of *Cassia siamea* L., *C. occidentalis* L. (Fabaceae), *Cnestis ferruginea* Vahl ex DC (Connaraceae), *Anthocleista djalonensis* A. Chev (Loganiaceae), *Solanum torvum* Swartz and *S. erianthum* G. Don (Solanaceae) volatiles grown in Nigeria have been studied. The essential oils were obtained by hydrodistillation and analyzed by GC and GC-MS. The main compounds of *C. siamea* were (*E*)-geranyl acetone (5.8%), 1-octen-3-ol (5.8%), linalool (7.8%), *iso*-italicene (15.4%) and (*E*)- β -damascenone (11.0%). On the other hand, *C. occidentalis* consisted mainly of (*E*)-geranyl acetone (8.0%), hexahydrofarnesylacetone (24.0%) and (*E*)-phytol acetate (40.7%). The oil of *C. ferruginea* was comprised mainly of (*E*)-geranyl acetone (13.7%), (*E*)-*a*-ionone (9.5%), phytol (5.8%), pentadecanal (6.1%) and 1-octen-3-ol (5.5%). The main compounds of *A. djalonensis* were *a*-humalene (31.9%), β -caryophyllene (17.8%), humulene epoxide II (12.7%) and caryophyllene oxide (5.9%). The main volatiles of *S. torvum* were (*E*)-phytol acetate (38.7%), pentadecanal (25.3%) and (*E*)-geranyl acetone (5.0%). Apart from methyl salicylate (4.5%), tetradecanal (2.2%), 2-pentyl furan (1.8%), hexahydrofarnesylacetone (1.6%) and hexadecanal (1.1%), all other compounds were either present in trace quantity or in amounts less than 1%. On the other hand, *a*-humulene (46.6%) and β -caryophyllene (20.6%) were the compounds occurring in higher quantities in *S. erianthum*. The volatile oil contents of *Cassia siamea, Cnestis ferruginea, Anthocleista djalonensis* and *Solanum torvum* are being reported for the first time.

Keywords: Cassia siamea, Cassia occidentalis, Cnestis ferruginea, Anthocleista djalonensis, Solanum torvum, Solanum erianthum, Essential oil composition.

Cassia or Senna is a large genus of flowering plants in the family Fabaceae, subfamily Caesalpinioideae. This diverse genus is native throughout the tropics, with a small number of species reaching into temperate regions. The number of species is usually estimated to be about 260 [1], but some authors believe that there are as many as 350. The Cassias are typically shrubs or subshrubs, some becoming scandent when growing into other vegetation. Cassia species make good ornamental plants and are used for landscape gardening. Some are herbs or small trees. The fruit is a legume, indehiscent or tardily dehiscent. The present investigation reports on the volatile compounds identified in the leaf oils of Cassia siamea L. and C. occidentalis L. These plants have been studied extensively for their pharmacological activities and phytochemistry, and a large number of active compounds have been isolated and characterized [2-7]. However, there is little literature information on the volatile contents of these and several other Cassia species. The authors are aware of three reports on the oil contents of \overline{C} . alata [8-10], and one report each on C. occidentalis [9], C. hirsuta [9], C. fistula [11], and C. grandis [12]. C. alata and C. occidentalis essential oils were found to be cytotoxic, inhibiting the growth of Hs 578T human tumor breast cell lines. Moreover, C. alata, C. hirsuta and C. occidentalis oils displayed only moderate antimicrobial activities to the assayed standard strains of Bacillus cereus, Staphylococcus aureus, Pseudomonas aeroginosa, Escherichia coli, Candida albicans and Aspergillus niger [9]. The antioxidant activity of the oil of C. alata was reported to be low compared with that of butylated hydroxytoluene (BHT) [10].

The shrub, Cnestis ferruginea Vahl ex DC (Connaraceae) is native to Africa, with local names such as 'Oko-Aja' or 'Gboyín-Gboyín'. It is the commonest of the 13 species of forest shrubs and climbers of the genus *Cnestis*. It bears orange-red fruits with velveting hairs on the follicle. Extracts or compounds of C. ferruginea have been shown to possess analgesic, anti-inflammatory [13, 14], antidepressant, anxiolytic [15], and anticonvulsant activities [16]; inhibit haemoglobin glycosylation in vitro [17]; possess hypoglycemia activity [18]; hepatoprotective potential [19] and improve sexual dysfunction [20]. The light petroleum fraction of C. ferruginea fruit has been shown to contain octacosanyl stearate and 1-myristo-2-stearo-3-palmitin [21]. A novel isoflavone glycoside, afrormosin-7-O-β-D-galactoside with antimicrobial activity was isolated from the fruit testa [22]. Squalene, myricyl alcohol, β-sitosterol, cyanidin, delphinidin and apigenidin have also been isolated from the plant [23]. No report on its volatile contents could be found in the literature.

Anthocleista djalonensis A. Chev (Loganiaceae) is a small tree up to 15 m tall, with a bole up to 40 cm in diameter; twigs sometimes with 2 erect spines or small cushions above the leaf axils. It is widely used throughout its distribution area as a strong purgative and diuretic [24]. A root decoction is commonly taken to treat constipation, to regulate menstruation and as an abortifacient. It is used as a wash, bath or as a vapour bath to treat leprosy, venereal diseases, oedema and scrotal elephantiasis. A root infusion is taken to treat intestinal problems, acute inflammations, and boils on the

skin [25,26]. The plant is known to possess anti-inflammatory [27] and free radical scavenging activities [28]. The plant is a source of secondary metabolites which are cytotoxic [29, 30] and antifungal [31].

The genus Solanum (Solanaceae) is comprised of 1700 species commonly found in the temperate and tropical regions of the world [32]. The genus is represented by some 25 species in Nigeria, including S. wrightii Benth, S. torvum Swartz, S. melongena L., S. tuberosum L., S. mammorum L. and S. seaforthianum Andr. (var. disjunctum). S. erianthum D. Don is a shrub or small tree about 6 m high with dense soft stellate hairs. The leaves act as an abortifacient and are considered a potent medicine for expelling all impurities through the urine and in particular to treat leucorrhoea [32]. The plant is also used to treat stomach ache, sores in the mouth and applied externally to skin irritations and rashes. S. torvum Swartz., is a bushy, erect and spiny perennial plant used horticulturally as a rootstock for eggplant. The plant is usually 2 or 3 m in height and 2 cm in basal diameter, but may reach 5 m in height and 8 cm in basal diameter. The shrub usually has a single stem at ground level, but it may branch on the lower stem. The fruits are berries that grow in clusters of tiny green spheres (ca. 1 cm in diameter) that look like green peas. They become yellow when fully ripe. They are thinfleshed and contain numerous flat, round, brown seeds. These plants have been studied extensively for their pharmacological activities and phytochemistry, where a large number of active compounds have been isolated and characterized [33-38]. S. erianthum leaf volatile oil was reported to have potent inhibitory activity against Hs 578T and PC-3 human breast and prostate tumor cells, respectively. In addition, S. erianthum and S. macranthum essential oils exhibited significant antimicrobial activity (19.5-625 µg/mL) on pathogens employed in the assay [39].

In continuation of our studies on the volatile constituents of Nigerian medicinal plants and herbs [8, 9, 39], we report herein compounds identified from the above mentioned plant samples. The essential oil yields were: 3.4%, v/w, *C. siamea*, pale yellow; 0.15%, v/w, *C. occidentalis*, light yellow; 0.14%, v/w, *C. ferruginea*; brownish; 0.06%, v/w, *A. djalonensis*, greenish yellow; 0.51%, v/w, *S. torvum*, pale yellow; and 0.14%, v/w, *S. erianthum*, pale yellow, calculated on a dry weight basis.

The compounds identified in *C. siamea* and *C. occidentalis* can be seen in Table 1. The ubiquitous terpenoid compounds were mostly represented among the volatile contents. Oxygenated monoterpenes and sesquiterpene hydrocarbons were the main classes of compound in *C. siamea*, while the oil of *C. occidentalis* consisted largely of diterpenoids and oxygenated sesquiterpenoids.

The main compounds of C. siamea were iso-italicene (15.4%), (E)β-damascenone (11.0%), linalool (7.8%), 1-octen-3-ol (5.8%) and (E)-geranyl acetone (5.8%). However, (E)-phytol acetate (40.7%) and hexahydrofarnesylacetone (24.0%) occurred in higher amounts in C. occidentalis. The oil also features significiant amounts of (E)geranyl acetone (8.0%), β -caryophyllene (4.1%), (E)- β -ionone and farnesylacetone (ca. 3.7%). The oil of C. occidentalis previously analysed from Nigeria contained (E)-phytol (26.0%), hexadecanoic acid (17.3%) and 6,10,14-trimethyl-2-pentadecanone (9.9%) as major compounds [9]. However, hexadecanoic acid and 6,10,14trimethyl-2-pentadecanone could not be identified in the present investigation, while the phytol content was low. It may be postulated that the oil of C. occidentalis from Nigeria could exist in two chemical forms, one with an abundance of diterpenoid and oxygenated sesquiterpenoids (this study), and another whose major compounds were fatty acids and aliphatic compounds [9].

Table 1: Volatile compounds identified from C. siamea and C. occidentalis.

Constituents LRI LRI LRI C.s MI 2-Heptanane 850 880 - Tr Cmd 2-Heptanane 900 901 - Tr Cmd Berzaldehyde 962 952 Tr O.4 Cmd 1-Octenol 978 984 0.8 1.2 Cmd 6-Methyl-S-hepten-2-one 986 981 1.1 0.8 Cmd 6-Acetryl-2-pentenyl) Furan 1000 - 0.6 Cmd Cmd 6-Acetryl-2-pentenyl) Furan 1004 - - Cmd Cmd 6-Acetryl-ansole 1019 1051 1.3 - Cmd 1/mas-Linalool oxide (furanoid) 1084 Tr - Cmd 1/mas-Linalool oxide (furanoid) 1088 1087 0.7 - Cmd 1/mas-Linaloo oxide (furanoid) 1088 1082 - Cod Cmd 1/mas-Linaloo 1160 1.6 - Cod <td< th=""><th>Constituent</th><th>1 1 1 3</th><th>Inth</th><th>C</th><th>C</th><th>M</th></td<>	Constituent	1 1 1 3	Inth	C	C	M
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1.3-Dimetholy benzaldehyde 1169 1165 - 0.3 Cmrl 2.4-Dimethyl benzaldehyde 1180 - 0.7 - Cmrl Naphthalene 1182 1178 0.6 Tr Cmrl a-Terpineol 1190 1186 1.4 - Cmrl bethyl silcylate 1192 110 0.5 - Cmrl f-Cyclocitral 1217 1217 1.6 0.6 Cmrl ac-Terpinyl aphthalene 1348 1345 1.0 - Cmrl ar-Terpinyl aphthalene 1381 1383 11.0 0.3 Cmrl 2.7-Dimethyl naphthalene 1400 1401 0.6 Cmrl 2.6-Dimethyl naphthalene 1407 - 0.5 Cmrl 2.7-Dimethyl naphthalene 1407 - 0.5 Cmrl 2.6-Dimethyl naphthalene 1407 - 0.5 Cmrl 2.6-Dimethyl naphthalene 1407 - Cmrl Cmrl 7.2-Dimethyl naphthalene 1418 1418 2.2 4.1 Cmrl						
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(E) - β -Damascenone 1381 1383 11.0 0.3 Cmrl $2,7$ -Dimethyl naphthalene 1392 - - 0.6 Cmrl xso -Italicene 1400 1401 15.4 - Cmrl $Longifolene$ 1400 1401 0.6 - Cmrl L_0 -Dimethyl naphthalene 1407 1.401 0.6 - Cmrl J_0 -Dimethyl naphthalene 1418 1.418 2.2 4.1 Cmrl J_0 -Dimethyl naphthalene 1421 1419 0.4 - Cmrl J_0 -Caryophylene 1432 1431 - 0.6 Cmrl J_0 -Gurinone 1432 1431 - 0.5 Cmrl J_0 -Gurinone 1437 1434 - tr Cmrl $I^-Methoxy naphthalene 1446 1444 1.0 Cmrl I^-Methoxy naphthalene 1445 1453 5.8 8.0 Cmrl (E)-Geranyl acetone 1445 1447 $						-
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Irans-a-Bergamotene 1439 1432 - 0.5 Cmrl I-Methoxy naphthalene 1446 1444 1.0 Cmrl Cmrl (E)-Geranyl acetone 1454 1453 5.8 8.0 Cmrl y-Muurolene 1477 1478 - 0.3 Cmrl (E)-β-lonone 1485 1487 1.9 3.7 Cmrl cis -Eudesma-6,11-diene 1489 1489 0.4 - Cmrl β -Selinene 1490 1489 0.4 - Cmrl Cmrl (Z)-a-Bisabolene 1504 1506 2.2 Tr Cmrl β -Curcumene 1512 1514 0.6 - Cmrl δ -Cadinene 1520 1520 0.5 - Cmrl θ -Curcumene 154 1555 0.5 - Cmrl δ -Cadinene 1542 1544 1.1 - Cmrl θ -Cadinene 1542 1555 0.5 <t< td=""><td></td><td></td><td></td><td></td><td></td><td></td></t<>						
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(E)-Geranyl acetone 1454 1453 5.8 8.0 Cmrl γ -Muurolene 1477 1478 - 0.3 Cmrl (E)-β-Ionone 1485 1487 1.9 3.7 Cmrl cis -Eudesma-6,11-diene 1489 1489 3.1 - Cmrl β -Selinene 1490 1489 0.4 - Cmrl Q -Methyl eugenol 1495 1494 0.6 - Cmrl (E)-Methyl eugenol 1495 1494 0.6 - Cmrl α -Selinene 1504 1506 2.2 Tr Cmrl (Z) - α -Bisabolene 1512 1514 0.6 - Cmrl δ -Cadinene 1524 1522 1.9 0.9 Cmrl δ -Cadinene 1542 1544 1.1 - Cmrl Δ -Cadinene 1542 1544 1.1 - Cmrl G -Dalporenol 1554 1586 - 0.7 <td< td=""><td>trans-α-Bergamotene</td><td></td><td></td><td>-</td><td>0.5</td><td>Cmrl</td></td<>	trans-α-Bergamotene			-	0.5	Cmrl
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(Z) - α -Bisabolene 1504 1506 2.2 Tr Cmrl β -Curcumene 1512 1514 0.6 - Cmrl (E) -Dihydroapofarnesal 1520 1520 0.5 - Cmrl δ -Cadinene 1524 1522 1.9 0.9 Cmrl β -Thujaplicinol 1536 1529 1.4 1.0 Cmrl α -Calacorene 1542 1544 1.1 - Cmrl Elemicin 1554 1555 0.5 - Cmrl (E) -Nerolidol 1564 1561 1.0 0.5 Cmrl Caryophyllene oxide 1581 1582 - 0.6 Cmrl (G) -Oplopenone 1604 1607 - 0.1 Cmrl β -Oplopenone 1664 1667 - 0.3 Cmrl α -Cadinol 1653 1652 - 0.3 Cmrl α -Cadinol 1653 1652 - 0.6 Cmr		1498	1498	Tr	0.4	Cmrl
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(E)-Nerolidol 1564 1561 1.0 0.5 Cmrl Caryophyllene oxide 1581 1582 - 0.6 Cmrl Gleenol 1585 1586 - 0.7 Cmrl Longiborneol 1594 1599 - 0.4 Cmrl β-Oplopenone 1604 1607 - 0.1 Cmrl geliphic 1652 1656 0.5 - Cmrl o-Cadinol 1653 1652 0.3 Cmrl o-Cadalene 1674 1675 - 0.6 Cmrl Pentadecanal 1717 1717 - 0.5 Cmrl Pentadecanal 1717 1717 - 0.5 Cmrl Farnesylacetone ^c 1927 1913 1.5 3.7 Cmrl Hexadecanoic acid 1950 1942 - 0.4 Cmrl Hexadecanoic acid 1959 1959 2.8 - Cmrl Abietatrien		1554	1555	0.5	-	Cmrl
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Abietatriene 2054 2055 1.0 - Cmrl (E)-Phytol acetate 2218 2218 - 40.7 Cmrl Total 91.0 98.6 98.6 99.0 99.0 99.0 Oxygenated monoterpenes 1.5 0.9 0.9 0.9 0.9 0.9 0.9 0.9 0.9 0.9 0.9 0.9 0.9 0.9 0.9 0.9 0.0 0.1 1.0 4.1 0.1	Hexadecanoic acid	1959	1959	2.8	-	Cmrl
(E)-Phytol acetate 2218 2218 - 40.7 Cmrl Total 91.0 98.6 Monoterpene hydrocarbons 1.5 0.9 Oxygenated monoterpenes 24.8 12.3 Sesquiterpene hydrocarbons 39.2 7.8 Oxygenated sequiterpenes 6.1 31.2 Diterpenoids 1.0 41.1 Aliphatic/fatty acids 10.2 2.1 Aromatic compounds 7.4 3.2		2054	2055	1.0		Cmrl
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Oxygenated sequiterpenes 6.1 31.2 Diterpenoids 1.0 41.1 Aliphatic/fatty acids 10.2 2.1 Aromatic compounds 7.4 3.2	Sesquiterpene hydrocarbons			39.2	7.8	
Diterpenoids 1.0 41.1 Aliphatic/fatty acids 10.2 2.1 Aromatic compounds 7.4 3.2				6.1	31.2	
Aliphatic/fatty acids 10.2 2.1 Aromatic compounds 7.4 3.2						
Aromatic compounds 7.4 3.2						
			L			

^aRetention indices on HP-5MS capillary column; ^bLiterature retention indices ([42] and lower version); M.I = Modes of identification: cmrl, Co-injection, mass fragmentation pattern, Retention indices from column and Literature Retention indices; cmr, Coinjection, Mass fragmentation pattern, Retention indices from column, literature other than [42] and its lower versions; ^c correct isomer not identified; - not identified and not present in literature; Tr < 0.1%; *C.s = C. siamea*; 2, *C.o = C. occidentalis*.

Table 2: Compounds identified	d from C. ferruginea.
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Constituents	LRI ^a	LRI ^b	Percent	M.I
(E)-2-Hexenal	855	846	1.2	Cmrl
1-Hexanol	868	863	0.3	Cmrl
2-Heptanone	890	889	Tr	Cmrl
Heptanal	900	901	1.1	Cmrl
Benzaldehyde	962	952	0.9	Cmrl
1-Heptanol	970	959	0.4	Cmrl
1-Octen-3-ol	978	974	5.5	Cmrl
6-Methyl-5-hepten-2-one	986	981	3.8	Cmrl
2-Pentyl furan	986	984	2.3	Cmrl
δ-2-Carene	1001	1001	0.7	Cmrl
trans-2(2-pentenyl) Furan	1004	-	1.4	Cmr
Octanal	1005	998	Tr	Cmrl
Dihydro-tagetone	1050	1046	0.4	Cmrl
m-Tolualdehyde	1068	1064	0.7	Cmrl
1-Octanol	1071	1063	Tr	Cmrl
Terpinolene	1088	1086	0.6	Cmrl
Linalool	1099	1095	1.9	Cmrl
Nonanal	1103	1100	3.5	Cmrl
Isophorone	1120	1118	0.6	Cmrl
4-keto-Isophorone	1143	1140	0.6	Cmrl
iso-Isopulegol	1155	1145	Tr	Cmrl
(E)-2-Nonenal	1162	1157	Tr	Cmrl
2,4-Dimethyl Benzaldehyde	1171	-	1.6	Cmr
Menthol	1174	1167	1.0	Cmrl
4-Terpineol	1178	1174	0.5	Cmrl
p-Methyl acetophenone	1179	1179	0.7	Cmrl
Naphthalene	1180	1178	0.7	Cmrl
α-Terpineol	1190	1186	Tr	Cmrl
Methyl salicylate	1192	1190	3.5	Cmrl
Safranal	1200	1196	2.0	Cmrl
Decanal	1204	1201	0.8	Cmrl
β-Cyclocitral	1217	1217	3.7	Cmrl
Pulegone	1237	1233	1.4	Cmrl
(E)-β-Damascenone	1381	1383	1.3	Cmrl
β-Elemene	1392	1389	0.7	Cmrl
(E)-α-Ionone	1426 1439	1428 1432	9.5 0.4	Cmrl Cmrl
<i>trans</i> -α-Bergamotene	1439	1432	13.7	Cmrl
(E)-Geranyl acetone (E)-β-Ionone	1454	1455	2.5	Cmrl
(E)-p-totione (E)-Nerolidol	1483	1487	0.8	Cmrl
(E)-Nerolidol Caryophyllene oxide	1564	1561	0.8	Cmrl
Tetradecanal	1611	1582	0.6	Cmrl
7-epi- α-Eudesmol	1665	1662	0.0	Cmrl
Elemol acetate	1680	1680	0.9	Cmrl
Pentadecanal	1717	1717	6.1	Cmrl
(E,E) - α -farnesyl acetate	1843	1843	1.0	Cmrl
Hexahydrofarnesylacetone	1845	1843	2.4	Cmrl
Farnesylacetone °	1927	1913	4.3	Cmrl
Phytol	1949	1945	5.8	Cmrl
	'otal		93.1%	
Monoterpene hydrocarbons			1.3	l
Oxygenated monoterpenes			41.3	
	Sesquiterpene hydrocarbons			l
	Oxygenated sesquiterpenes			
Diterpenoids			10.7 5.8	l
Aliphatic compounds			16.6	
Aromatic compounds			8.3	
Fatty acids			6.7	

^a Retention indices on HP-5MS capillary column; ^b Literature retention indices (Adams, [42] and its lower versions); Tr, trace amount < 0.1%; - not present in literature; ^c Correct isomer not identified; M.I. = Modes of identification which are: cmrl, Co-injection, Mass fragmentation pattern, Retention indices from column and Literature Retention indices; cmr, Co-injection, Mass fragmentation pattern, Retention indices from column, literature other than Adams [42] and its lower version.

Previous results for *Cassia* species revealed that the major compounds of *C. alata* were *ar*-turmerone (13.5%), β -caryophyllene (7.3%), (*E*)-phytol (7.0%) and 6,10,14-trimethyl-2-pentadecanone (6.8%) [9]; 1,8-cineole (39.8%), β -caryophyllene (19.1%) and caryophyllene oxide (12.7%) [8], as well as linalool (23.0%), borneol (8.6%) and pentadecanal (9.3%) [9]. Also, (*E*)-phytol (30.8%) and pentadecanal (21.7%) were identified as the main components of *C. hirsuta* [9]. On the other hand, linalool (31.5%) was the main constituent of *C. grandis* from Cuba [11], while *C. fistula* from Egypt [12] had (*E*)-nerolidol (38.0%), 2-hexadecanone (17.0%) and heptacosane (12.8%) in the flower oil, while the leaf oil was characterized by an abundance of phytol (16.1%), tetradecane (10.5%) and hexadecane (8.7%).

Therefore, from various reports, the following delineation of the chemical forms of the oils of *Cassia* species analyzed from all

regions is being proposed: (i) oils whose major compounds consist of diterpenoid and fatty acids, e.g. leaves of *C. occidentalis* [9], *C. fistula* [11] and *C. hirsuta* [9]; (ii) oils whose major compounds are composed of oxygenated sesquiterpenoids and fatty acids, as seen in *C. fistula* flower [11]; (iii) oils whose major compounds were dominated by oxygenated monoterpenoid and fatty acids, e.g. *C. grandis* flower [12] and *C. alata* leaf [10]; (iv) oils with significant proportions of mono- and sesquiterpenes, as seen in leaves of *C. alata* [8,9], and *C. siamea* (this study); (v) oils in which oxygenated sesquiterpenoid and diterpenoid compounds predominate, as seen in *C. occidentalis* (this study).

Table 2 shows the compounds identified from *C. ferruginea*. Oxygenated monoterpenoids (41.3%), aliphatic compounds (16.6%) and oxygenated sesquiterpenoids (10.7%) represent the major classes of compounds identified in the oil. The main oil constituents were (*E*)-geranyl acetone (13.7%), (*E*)- α -ionone (9.5%), phytol (5.8%), pentadecanal (6.1%) and 1-octen-3-oil (5.5%). Farnesylacetone (4.3%), 6-ethyl-5-hepten-2-one (3.8%), β-cyclocitral (3.7%), nonanal (3.5%) and methyl salicylate (3.5%) also occurred above 2%. The authors are unaware of any literature information on the volatile contents of *C. ferruginea* either from Nigeria or elsewhere, and as such the present report may represent the first of its kind.

Sesquiterpene compounds (82.5%) are the dominant class of compounds in *A. djalonensis* (Table 3). The main ones were α -humulene (31.9%) β -caryophyllene (17.8%), humulene epoxide II (12.7%) and caryophyllene oxide (5.9%). Apart from phytol acetate (2.2%), (*Z*)-caryophyllene (1.7%), δ -cadinene (1.7%), hexahydrofarnesyl acetone (1.3%), α -copaene 1.0%) and cubebol (1.0%), all other compounds were identified in insignificant amount. The authors are unaware of any literature information on the volatile contents of *A. djalonensi* or any other member of the family, either from Nigeria or elsewhere, and as such the present report may represent the first of its kind.

In *S. torvum*, diterpenoids (38.7%), fatty acids (30.5%), and oxygenated monoterpenoids were the most abundant classes of compounds, while sesquiterpenoids (77.6% vs. 15.2%; hydrocarbon vs. oxygen derivatives) predominated in *S. erianthum*. The main volatiles of *S. torvum* were (*E*)-phytol acetate (38.7%), pentadecanal (25.3%) and (*E*)-geranyl acetone (5.0%). Apart from methyl salicylate (4.5%), tetradecanal (2.2%), 2-pentyl furan (1.8%), hexahydrofarnesylacetone (1.6%) and hexadecanal (1.1%), all other compounds were either present in trace quantity or amounts less than 1%. On the other hand, α -humulene (46.6%) and β -caryophyllene (20.6%) were the compounds in significant quantities were germacrene D (4.8%), humulene epoxide II (4.4%) and caryophyllene oxide (4.0%). Monoterpenoids (1.4%) were rare among the identified compounds (Table 4)

There are literature reports on the oil constituents of some *Solanum* species grown in Nigeria (Table 5), but not for *S. torvum*. The volatile oil of *S. erianthum* was characterized by the abundance of α -terpinolene (17.8%), α -phellandrene (17.5%), *p*-cymeme (15.7%) and β -pinene (11.7%) in the leaves; and α -humulene (23.1%), humulene epoxide II (20. 0%), caryophyllene oxide (16.5%), methyl salicylate (11.8%) and β -caryophyllene (10.9%) in the fruits [39]. The leaf oil of *S. macranthum* consisted of (*E*)-phytol (29.0%), pentadecanal (28.1%), pentadecane (7.7%) and ethyl palmitate (5.7%), while the fruit oil had α -humulene (36.5%), (*E*)- caryophyllene (17.8%), ethyl palmitate (9.4%), and methyl salicylate (8.2%) as major compounds [39]. Germacrene D (14.8%),

Table 3: Leaf oil constituents of A. djalonensis.

Constituents	LRI ^a	LRI ^b	Percent	M.I	
(E)-3-Hexen-1-ol	856	-	Tr	Cmr	
2-Heptanone	890	889	Tr	Cmrl	
Heptanal	900 962	901 952	Tr	Cmrl	
Benzaldehyde	962	952 981	Tr	Cmrl	
6-Methyl-5-hepten-2-one 2-Pentyl furan	986	981	Tr 0.1	Cmrl Cmrl	
Limonene	1031	1024	Tr	Cmrl	
2-Nonanone	1092	1024	Tr	Cmrl	
Nonanal	1102	1100	0.2	Cmrl	
(E,Z)-2,6-Nonadienal	1156	1150	0.2	Cmrl	
(E)-2-Nonenal	1162	1157	0.1	Cmrl	
1-Nonanol	1173	1165	Tr	Cmrl	
Naphthalene	1180	1178	Tr	Cmrl	
Decanal	1205	1201	Tr	Cmrl	
2-Undecanone	1292	1293	Tr	Cmrl	
Undecanal ^c	1306	1300	0.1	Cmrl	
α-Cubebene	1351	1345	Tr	Cmrl	
α-Copaene	1376	1374	1.0	Cmrl	
β-Bourbonene	1384	1387	0.4	Cmrl	
β-Elemene n-Tetradecane	1391 1399	1389 1400	0.4 Tr	Cmrl Cmrl	
(Z)-Caryophyllene	1399	1400	1.7	Cmrl	
β-Caryophyllene	1404	1408	1.7	Cmrl	
β-Gurjunene	1418	1418	0.2	Cmrl	
α-Guaiene	1432	1437	0.2	Cmrl	
α-Humulene	1454	1454	31.9	Cmrl	
cis-Muurola-4(14),5-diene	1461	1465	Tr	Cmrl	
γ-Muurolene	1477	1478	0.3	Cmrl	
α-Amorphene	1480	1483	Tr	Cmrl	
(E)-β-Ionone	1485	1487	1.6	Cmrl	
epi-Cubebol	1494	1493	0.3	Cmrl	
trans- β-Guaiene	1500	1502	0.4	Cmrl	
n-Pentadecane	1500	1500	0.4	Cmrl	
Germacrene A	1505	1508	0.8	Cmrl	
Cubebol	1515	1514	1.0	Cmrl	
δ-Cadinene	1524	1522	1.7	Cmrl	
α-Calacorene	1542 1549	1544 1548	Tr 0.2	Cmrl Cmrl	
Elemol (E)-Nerolidol	1565	1548	0.2	Cmrl	
Germacrene D-4-ol	1505	1574	0.2	Cmrl	
Caryophyllene oxide	1581	1582	5.9	Cmrl	
<i>cis</i> -β-Elemenone	1590	1589	0.2	Cmrl	
Cedrol	1596	1600	0.1	Cmrl	
Humulene epoxide II	1606	1608	12.5	Cmrl	
Caryophylla-4(14),8(15)-dien-5-ol	1636	-	0.2	Cmr	
Epoxy-allo-aromadendrene	1639	1639	0.9	Cmrl	
τ-Cadinol	1641	-	0.5	Cmr	
α-Cadinol	1654	1652	0.8	Cmrl	
Acorenone	1696	1692	0.3	Cmrl	
n-Heptadecane	1700	1700	0.1	Cmrl	
Pentadecanal Mint sulfide	1717 1741	1717 1740	0.5	Cmrl Cmrl	
Hexahydrofarnesylacetone	1/41 1845	1740	1.3	Cmrl	
Methyl hexadecanoate	1927	1843	0.2	Cmrl	
Phytol	1949	1942	0.2	Cmrl	
<i>n</i> -Heneicosane	2100	2100	Tr	Cmrl	
(E)-Phytol acetate	2218	2218	2.2	Cmrl	
Tot			88.2%	1	
Monoterpene hydrocarbons -					
Oxygenated monoterpenes			1.6		
Sesquiterpene hydr	57.3				
Oxygenated sesqui	25.2				
Diterpenoid	2.7				
Aliphatic compo	0.6	<u> </u>			
Aromatic compo			0.1		
Fatty acids		^b Literature 1	0.7	1	

^a Retention indices on HP-5MS capillary column; ^b Literature retention indices ([42] and its lower version); Tr, trace amount < 0.1%; - not identified and not present in literature; ^c Correct isomer not identified; M.I = Mode of identification: cmrl, Co-injection, Mass fragmentation pattern, Retention indices from column and Literature Retention indices; cmr, Co-injection, Mass fragmentation pattern, Retention indices from column, Literature other than [42] and its lower version.

Table 4: Constituents of S. torvum and S. erianthum.

Constituents	LRI ^a	LRI ^b	S. t	S. e	M.I
(E)-2-Hexenal	854	846	0.4	Tr	Cmrl
2-Heptanone	890	889	-	Tr	Cmrl
Heptanal	900	901	Tr	Tr	Cmrl
α-Pinene	939	932	-	0.3	Cmrl
Benzaldehyde	962	952	-	Tr	Cmrl
Sabinene	977	969	-	Tr	Cmrl
6-Methyl-5-hepten-2-one	986	981	0.2	Tr	Cmrl
2-Pentyl furan	986	984	1.8	0.3	Cmrl
(Z)-3-Hexenyl acetate	1007	1004	Tr	-	Cmrl
p-Cymene	1026	1020	-	Tr	Cmrl
Limonene	1031	1024	1.9	0.2	Cmrl
(E)-β-Ocimene	1051	1044	Tr	Tr	Cmrl

(E) 2 Optional	1062	1060		Te	Const
(E)-2-Octenol 1-Octanol	1063 1071	1060 1063	-	Tr Tr	Cmrl Cmrl
Terpinolene	1071	1086	_	Tr	Cmrl
Linalool	1000	1095	0.6	-	Cmrl
Nonanal	1103	1100	0.5	0.3	Cmrl
Geijerene	1143	1138	-	Tr	Cmrl
(E)-2-Nonenal	1152	1157	0.5	Tr	Cmrl
(E,Z)-2,6-Nonadienal	1156	1150	0.3	Tr	Cmrl
Naphthalene	1180 1186	1178 1184	0.4	-	Cmrl Cmrl
(Z)-3-Hexenyl butanoate Methyl salicyalte	1191	1184	4.5	-	Cmrl
<i>n</i> -Dodecane	1199	1200	0.2	-	Cmrl
Decanal	1204	1201	0.3	Tr	Cmrl
β-Cyclocitral	1217	1217	0.2	Tr	Cmrl
Pregeijerene	1288	1285	1.0	Tr	Cmrl
n-Tridecane	1299	1300	0.2	Tr	Cmrl
Undecanal ^c (E, E)-2,4-Decadienal	1306	1300 1315	- T-	Tr	Cmrl Cmrl
(E, E)-2,4-Decadienal Hexenyl tiglate	1314 1332	1315	Tr -	Tr	Cmrl
α-Cubebene	1352	1319	-	Tr	Cmrl
α-Copaene	1376	1374	Tr	0.9	Cmrl
β-Bourbonene	1384	1387	-	0.5	Cmrl
β-Elemene	1391	1389	-	0.6	Cmrl
n-Tetradecane	1400	1400	-	Tr	Cmrl
(Z)-Caryophyllene	1405	1408	-	0.3	Cmrl
β-Caryophyllene	1418	1418	0.2	20.6	Cmrl
β-Gurjunene α-Guaiene	1432 1439	1431 1437	-	0.2 Tr	Cmrl Cmrl
α-Gualene α-Humulene	1439	1457	-	46.6	Cmrl
(E)-Geranyl acetone	1456	1454	5.0	40.0	Cmrl
cis-Muurola-4(14),5-diene	1450	1465	-	Tr	Cmrl
γ-Muurolene	1477	1478	-	Tr	Cmrl
Germacrene D	1480	1484	0.3	4.8	Cmrl
(E)-β-Ionone	1485	1487	0.9	0.9	Cmrl
Bicyclogermacrene	1494	1500	-	0.2	Cmrl
epi-Cubebol	1496	1493	-	0.2	Cmrl
α-Muurolene	1499 1500	1500 1500	-	Tr	Cmrl
n-Pentadecane Germacrene A	1500	1500	0.4 Tr	- 1.5	Cmrl Cmrl
Tridecanal	1509	1509	0.3	Tr	Cmrl
Cubebol	1515	1514	-	0.7	Cmrl
(E)-Dihydroapofernesal	1522	1520	Tr	-	Cmrl
δ-Cadinene	1524	1522	-	1.4	Cmrl
(E)-Nerolidol	1565	1561	0.3	-	Cmrl
(Z)-3-Hexenyl benzoate	1570	1565	0.3	-	Cmrl
Caryophyllene oxide n-Hexadecane	1581 1600	1582 1600	- 0.6	4.0	Cmrl Cmrl
Humulene epoxide II	1606	1608	-	4.4	Cmrl
1-epi-Cubenol	1628	1627	-	0.3	Cmrl
Caryophylla-4(14),8(15)-dien-5-ol	1636	-	-	0.6	Cmr
epi-a-Cadinol	1640	1638	-	0.7	Cmrl
α-Muurolol	1645	1644	-	0.2	Cmrl
α-Cadinol	1653	1652	-	1.3	Cmrl
1-Tetradecanol Eudesma-4(15),7-dien-1-β-ol	1674	1607	0.3	-	Cmrl
n-Heptadecane	1688 1700	1687 1700	-	0.2	Cmrl Cmrl
Pentadecanal	1700	1700	25.3	0.2	Cmrl
n-Octadecane	1800	1800	0.5	-	Cmrl
Hexadecanal	1830	-	1.1	-	Cmr
Hexahydrofarnesylacetone	1845	1843	1.6	0.8	Cmrl
Benzyl salicylate	1866	1864	0.2	-	Cmrl
<i>n</i> -Nonadecane	1900	1900	0.6	-	Cmrl
Farnesylacetone ^c Hexadecanoic acid	1927 1959	1913 1958	-	1.0 0.8	Cmrl
n-Eicosane	2000	2000	0.5	- 0.0	Cmrl Cmrl
<i>n</i> -Heneicosane	2100	2100	0.5	-	Cmrl
(E)-Phytol acetate	2218	2218	38.7	-	Cmrl
Total			90.8%	95.8%	
Monoterpene hydroc			2.9	0.5	
Oxygenated monote			11.2	0.9	
Sesquiterpene hydro			0.5 1.9	77.6	
Oxygenated sesquite Diterpenoids			38.7		
Aliphatic compounds			2.7	0.3	
Aromatic compounds			2.4	0.3	
Fatty acids			30.5	1.0	
^a Potentian indians on UP 5MS	.11	1 h	r •	rotontion in	diago $([42]$

^a Retention indices on HP-5MS capillary column; ^b Literature retention indices ([42] and its lower version); Tr, trace amount < 0.1%; - not identified and not present in literature; ^c Correct isomer not identified; M.I = Mode of identification: cmrl, Co-injection, Mass fragmentation pattern, Retention indices from column and Literature Retention indices; cmr, Co-injection, Mass fragmentation pattern, Retention indices from column, Literature other than [42] and its lower version. *S. t* = *Solanum torvum; S. e* = *Solanum erianthum*

pentadecanal (11.4%), β -elemene (10.1%), α -bulnesene (7.9%), δ -cadinene (6.0%), β -caryophyllene (6.5%) and α -copaene (5.5%) were the major components of the oil of *S. nigrum* var. *virginicum* [40].

Table 5: Major components of Solanum oils from Nigeria.

Species/Plant Parts	Major constituents	Ref
S. erianthum (leaves)	α-terpinolene, α-phellandrene, p-cymene, β-pinene	[39]
S. erianthum (fruits)	α-humulene, humelene epoxide, caryophyllene oxide, methyl salicylate, β-caryophyllene	[39]
S. macranthum (leaves)	(E)-phytol, pentadecanal, pentadecane, ethyl palmitate	[39]
S. macranthum (fruits)	α-humelene, (E)-caryophyllene, ethyl palmitate, methyl salicylate	[39]
S. nigrum	Germacrene D, pentadecanal, β-elemene	[40]
var. nigrum (leaves)	α-bulnesene, δ-cadinene, β-caryophyllene, α-copaene	
S. torvum (leaves)	(E)-phytol acetate, pentadecanal (E)-geranyl acetone	This study
S. erianthum (leaves)	α-humulene, β-caryophyllene	This study

It is evident that the leaf oils of *S. erianthum* analysed from Nigeria exist in two chemical forms, namely one with an abundance of monoterpene hydrocarbons [39] and one whose major compounds are sesquiterpene hydrocarbons, as seen in the present study (Table 5).

Moreover, the oils of *Solanum* species so far analysed from Nigeria could be thought to exist in three chemical forms (Table 5) namely; (i) oil dominated by sesquiterpenoid compounds, such as *S. nigrum* var. *nigrum, S. erianthum* (leaf and fruit) and *S. macranthum* (fruits); (ii) oil consisting of monoterpene hydrocarbons, as seen in *S. erianthum* (leaf); and (iii) oil with abundant diterpenoids and fatty acids, typified by *S. torvum* and *S. macranthum* (leaves). Further studies are on-going to ascertain the biological activities of these essential oils and the compounds responsible for such activities.

Experimental

Plants collection: Mature leaves of *C. siamea, C. occidentalis* and *A. djalonensis* were collected at Abule-Eera, a suburb along the Badagry Expressway Lagos, in May 2011. Leaves of *C. ferruginea* were harvested at Igbesa Waterside, Agbara, also along Badagry Expressway, Lagos, in June 2011. Botanical identifications were performed at the Herbarium, Forestry Research Institute of Nigeria (FRIN), Ibadan, Nigeria, where voucher specimens FHI 109435, FHI 109436, FHI 109434 and FHI 109438, respectively have been deposited for future reference. The leaves of *S. torvum* and *S. erianthum* were obtained respectively from Ijede Town and Egan Town, Lagos. Identification was accomplished at the Herbarium, Department of Botany, University of Lagos, Nigeria, where voucher specimens LUH 5227 and LUH 5226, respectively were deposited. Leaves were air-dried for 2 weeks under laboratory shade prior to extraction.

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Extraction of the oils: Aliquots of air-dried and pulverized leaves (30 g each) were subjected to separate hydrodistillation in an all-glass Clevenger type apparatus for 3 h in accordance with the British Pharmacopoeia method [41]. The oils obtained were collected and stored under refrigeration until analyses were performed.

Gas chromatography (GC) and gas chromatography-mass spectrometry (GC-MS): GC analysis was accomplished with a HP-5890 series II instrument equipped with HP-wax and HP-5 capillary columns (both 30 m x 0.25 mm, 0.25 μ m film thickness) with the following temperature programme; 60°C for 10 min, rising from 5°C/min to 220°C. Both injector and detector temperatures were maintained at 250°C; carrier gas, nitrogen (2 mL/ min); detector, FID; ratio, 1:30. The volume injected was 0.5 μ L. The relative proportions of the oil constituents were the percentages obtained (% area) by FID peak-area normalisation without the use of response factor.

Gas chromatography-electron ionization mass spectroscopy (GC-EIMS) analysis was performed with a Varian CP-3800 gas chromatography equipped with a HP-5 capillary column (30 m x 0.25 mm; film thickness 0.25 μ m) and a Varian Saturn 2000 ion trap mass detector. Analytical conditions: injector and transfer line temperatures were 220°C and 240°C respectively. Oven temperature programmed from 60°C to 244°C at 3°C/min; carrier gas was helium at a flow rate of 1 mL/min; injection of 0.2 μ L (10% *n*-hexane solution); split ratio 1:30. Mass spectra were recorded at 70 eV. The acquisition mass range was 30-300 m/z at a scan rate of 1 scan/s.

Identification of constituents: Identification of the constituents was based on comparison of the retention times with those of authentic samples, comparing their linear indices relative to a series of *n*-hydrocarbons, and by computer matching against commercial spectra. Further identification was also made possible by the use of a self constructed spectral library built up from pure substances and components of known oils and MS literature data [42, 43]. Moreover, the molecular weights of all the identified substances were confirmed by gas chromatography-chemical ionisation mass spectrometry, using methanol as CI ionizing gas.

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