

# Blistering lesions associated with Waldenström macroglobulinemia: New insights into pathogenesis

A 69-years old woman showed recurrent vesicles on both dorsal feet and hands (Figure 1A,B). The lesions have been developed over the years. A monoclonal IgM component (3350 mg/dl) and lambda light chain with no evidence of Bence-Jones proteinuria were detected. The diagnosis of Waldenström Macroglobulinemia (WM) was confirmed by a radiologic evidence of mediastinal lymphadenopathy and a bone marrow biopsy. A skin punch biopsy showed a dermo-epidermal split (Figure 1F). The direct immunofluorescence (DIF) on perilesional skin showed a linear deposition of IgM at the dermo-epidermal junction (DEJ) (Figure 1C). A strong intercellular IgM deposition was detected (Figure 1D). The indirect immunofluorescence showed a deposition of IgM at the DEJ, and on the dermal side of salt-split skin (SSS) (Figure 1E). To identify a possible autoantigen on the dermal side of the SSS, immunoblotting (IB) on keratinocyte extracts and supernatant, dermal extracts, affinity purified laminin 332 and a modified commercial ELISA to detect type VII collagen (coll VII) (MBL) IgM were performed. Although no band was detected by IB, a significant reactivity of circulating IgM to coll VII was detected by ELISA (Figure 1G). We have also analyzed the IgM binding with normal sera (Figure 1F) and on other skin autoantigens (BP180 and BP230) by modified ELISA and no reactivity was found (data not shown). ELISA is often more sensitive than IB and this could be due to the loss of native epitopes present in coated protein (ELISA) and absent in the denatured one (IB).<sup>1</sup> Modified ELISA is based on (1) coll VII coated on commercially available wells (MBL) and (2) a secondary antibody goat antihuman IgM mu chain HRP conjugated (ab97205, Abcam Inc, Toronto, ON, Canada) used to detect human IgM autoantibodies bond to coll VII.

WM belongs to non-Hodgkin's B-cell lymphoma.<sup>2,3</sup> It is characterized by IgM paraproteinemia and lymphoplasmacytoid infiltration in the lymph nodes, spleen, and bone marrow.<sup>2,3</sup> Cutaneous manifestations of WM can be classified in nonspecific and specific. The specific manifestations are associated with lymphocytic infiltration or skin deposition of IgM antibodies.<sup>2,4</sup> Nonspecific cutaneous lesions of WM include leucocytoclastic vasculitis and Raynaud's phenomenon, while specific cutaneous lesions include papules and blisters.<sup>5</sup> The presence of isolated IgM deposits in the skin is defined as cutaneous macroglobulinosis (CM).<sup>2,5</sup> CM usually presents with pruritic,

papules or plaques. Subepidermal blistering lesions are rare, specific manifestations of CM.<sup>5</sup>

Some immunological features detected in our patient, such as IgM deposition at the DEJ, IgM staining of dermal side of SSS, and intercellular IgM deposition, were previously described.<sup>2-4,6,7</sup> However, with the exception of West et al, who detected an IgM reactivity to a 290 kDa antigen by IB,<sup>6</sup> no other Author identified the antigen targeted by the patient's IgM<sup>3,4,6,8,9</sup>. In this context, we demonstrated a reactivity to coll VII by ELISA and not by IB, suggesting that IgM reacted with an epitope exclusively presents on the native protein. However, when patient serum was diluted to normalize IgM levels to those found in normal human serum, we obtained a negative result by coll VII ELISA (data not shown), suggesting that IgM could have low affinity for coll VII and/or this reactivity is due to a cross-reaction. These findings highlight that anti-coll VII IgM autoantibodies could represent the trigger for the blister formation, while the intercellular deposition of IgM could represent an epitope spreading phenomenon. Because of the high serum concentration of IgM, an unspecific result by ELISA cannot be excluded. However, it is possible that a huge concentration of IgM reacting or cross-reacting with an hemidesmosomal and/or desmosomal antigen leads to skin lesions.

## ACKNOWLEDGMENT

This article was partially supported by the "Progetto Ricerca Corrente-2019" of the Italian Ministry of Health, Rome, Italy.

## CONFLICT OF INTEREST

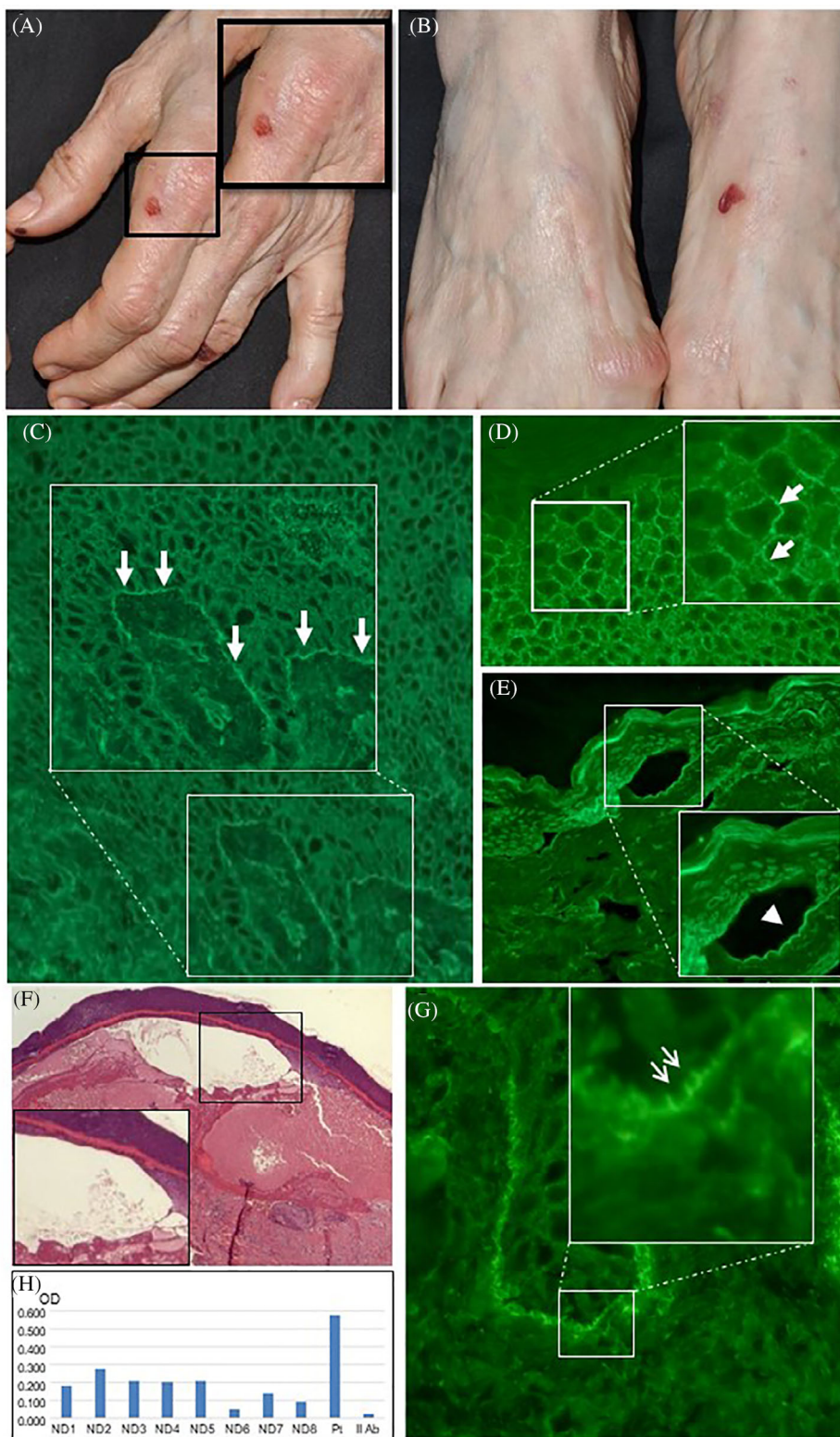
The authors declare no conflict of interest.

## AUTHOR CONTRIBUTIONS

**Simone Garcovich** and **Dario Didona**: Wrote the paper. **Clara De Simone** and **Valerio De Stefano**: Corrected the proof. **Feliciana Mariotti**: Performed the photos. **Giovanni Di Zenzo**: Designed the paper and corrected the proof.

## DATA AVAILABILITY STATEMENT

The data that support the findings of this study are available from the corresponding author upon reasonable request.



**FIGURE 1** Clinical and pathological manifestations of the patient. (A) Vesicular lesions on the dorsum of the hand; (B) Vesicular lesions on the dorsal aspect of the foot; (C) Direct immunofluorescence of perilesional skin showing deposits of IgM at the dermo-epidermal junction (DEJ) (arrows underline the linear staining at DEJ); (D) Direct immunofluorescence of perilesional skin showing intercellular deposition of IgM; (E) Indirect immunofluorescence showing linear staining of IgM on the dermal side (arrow head underlines the dermal staining) of salt-split skin; (F) Histopathological examination shows a dermo-epidermal detachment; (G) Modified ELISA for IgM shows a significant reactivity of patient serum (Pt) on recombinant collagen VII. Eight normal donors sera (ND1-ND8) and the antihuman IgM secondary antibody (II Ab) used show lower optical density (ODs) than Pt. We obtain the consent of the patient for the publication of identifiable details, which can include photograph(s), case history and/or details within the text

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**REFERENCES**

1. Schmidt T, Hoch M, Lotfi Jad SS, et al. Serological diagnostics in the detection of IgG autoantibodies against human collagen VII in epidermolysis bullosa acquisita: a multicentre analysis. *Br J Dermatol.* 2017;177:1683-1692.
2. Whittaker SJ, Bhogal BS, Black MM. Acquired immunobullous disease: a cutaneous manifestation of IgM macroglobulinaemia. *Br J Dermatol.* 1996;135:283-286.
3. Morita E, Horiuchi K, Yamamoto S, Hashimoto T. A case of acquired autoimmune bullous disease associated with IgM macroglobulinaemia. *J Dermatol.* 1999;26:671-676.
4. Chattopadhyay M, Rytina E, Dada M, Bhogal BS, Groves R, Handfield-Jones S. Immunobullous dermatosis associated with Waldenström macroglobulinaemia treated with rituximab. *Clin Exp Dermatol.* 2013;38:866-869.
5. Camp BJ, Magro CM. Cutaneous macroglobulinosis: a case series. *J Cutan Pathol.* 2012;39:962-970.
6. West NY, Fitzpatrick JE, David-Bajar KM, Bennion SD. Waldenström macroglobulinemia-induced bullous dermatosis. *Arch Dermatol.* 1998;134:1127-1131.
7. Kieny A, Hashimoto T, Ishii N, Antal MC, Boehm N, Lipsker D. Granular pemphigus-like IgM deposition around keratinocytes in a patient with Waldenström's macroglobulinaemia: a so far unreported finding. *J Eur Acad Dermatol Venereol.* 2017;31:e47-e49.
8. Cobb MW, Domloge-Hultsch N, Frame JN, Yancey KB. Waldenström macroglobulinemia with an IgM-kappa antiepidermal basement membrane zone antibody. *Arch Dermatol.* 1992;128:372-376.
9. Tedeschi A, Conticello C, Rizzi R, et al. Diagnostic framing of IgM monoclonal gammopathy: focus on Waldenström macroglobulinemia. *Hematol Oncol.* 2019;37:117-128.