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Periodontal infrabony defects: systematic review of healing by defect morphology following regenerative surgery

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ABSTRACT

Background: It is thought that infrabony defect morphology affects the outcome of periodontal regenerative surgery. However, this has not been systematically investigated.

Aims: To investigate how well defect morphology is described in papers reporting regenerative therapy of periodontal infrabony defects and to investigate its effect on clinical and radiographic outcomes

Materials and Methods: A search was conducted in 3 electronic databases for publications reporting clinical and radiographic outcomes of periodontal intrabony defects after regenerative therapy, divided by defect morphology.

Results: The initial search resulted in 4487 papers, reduced to 143 after first and second screening. Fifteen of these publications were suitable for a fixed effects meta-analysis. Initial defect depth was found to influence radiographic bone gain 12 months post-surgery, while narrower angles and increased number of walls influenced both radiographic bone gain and clinical attachment level (CAL) gain at 12 months. These associations seemed to occur irrespective of biomaterials used. Risk of bias ranged from low to high.

Conclusion: Deeper defects with narrower angles and increased number of walls exhibit improved CAL and radiographic bone gain at 12 months post-regenerative surgery. More data are needed about other aspects of defect morphology such as extension to buccal/lingual surfaces.

CLINICAL RELEVANCE

Scientific rationale for study: It is important to establish which aspects of infrabony defect morphology influence outcomes of regenerative surgery. Principal findings: Defect depth, angle and number of walls appear to influence the healing following regenerative periodontal surgery, irrespective of biomaterials used. Practical implications: It is important to consider defect morphology when planning regenerative periodontal surgery. More data about several aspects of defect morphology should be routinely collected and correlated with treatment response, in order to improve the clinician's ability to maximise healing.

INTRODUCTION

Classically, periodontal defects have been differentiated based on bone resorption patterns into 'supraosseous' ('suprabony') and 'infraosseous' ('infrabony') (Goldman & Cohen, 1958). These authors defined suprabony defects as those where the base of the pocket is located coronal to the alveolar crest. On the other hand, infrabony defects are those with apical location of the base of the pocket relative to the bone crest. Goldman and Cohen then classified infrabony defects according to the location and number of osseous walls remaining around the pocket. It has been suggested that the term 'intrabony' means 'within or inside the bone', while 'infrabony' means 'below the crest of bone' (Weinberg & Eskow, 2000). These authors suggested that only 3-wall angular defects should be termed 'intrabony', while all other vertical bony defects should be referred to as 'infrabony'. A large body of clinical and histological evidence accumulated over the last 4-5 decades shows how healing following periodontal surgery of infrabony defects can, with the use of biomaterials, be guided towards the formation of de novo cementum, functionallyoriented periodontal ligament, new alveolar bone and gingiva (Melcher, 1976; Nyman, Lindhe, Karring, Rylander, 1982; Wikesjo & Nilveus, 1990). The emphasis on '3-wall' bony defects was due to their higher chances of successful regeneration (Weinberg & Eskow, 2000). Recent developments in periodontal regenerative techniques and materials have pushed the boundaries of what is considered 'regenerable' (P. Cortellini, Stalpers, Mollo, & Tonetti, 2020). Papapanou & Tonetti differentiated osseous defects into 'suprabony' defects, 'infrabony' and 'interradicular or furcation' defects (Papapanou & Tonetti, 2000). Infrabony defects were further divided into 'intrabony' and 'craters' and the former were sub-divided into '1-, 2- or 3-wall defects' or 'combinations'. The emphasis was placed on differentiating whether or not the defect affected more or less to the same extent the adjacent root surfaces of an interdental space (similar periodontal breakdown along the root surface of two adjacent teeth, i.e. craters) or primarily affected one of the two root surfaces of an interdental space (greater periodontal breakdown on the tooth with the defect and more coronal crest of bone on the adjacent tooth in the same interdental space, i.e. intrabony defect) (Papapanou and Tonetti 2000).

Infrabony defects have been associated with risk of periodontal progression in the absence of the appropriate therapy, but not if included in regular maintenance care programs (Heins, Hartigan, Low, & Chace, 1989; Papapanou & Wennstrom, 1991)(Pontoriero, Nyman, & Lindhe, 1988).

With the currently available regeneration procedures, materials, and technologies, intrabony defects can be successfully regenerated, subject to patient-factors such as plaque control, smoking, medical history, as well as tooth mobility, restorative and endodontic condition (Nibali et al., 2019). Several publications reported on the superiority of periodontal regenerative therapy in the treatment of intrabony defects over the conventional surgeries such as periodontal access flap, known as open flap debridement surgery (OFD), in terms of probing pocket depth (PPD) and clinical attachment loss (CAL) reductions (Castro et al., 2017; Needleman, Worthington, Giedrys-Leeper, & Tucker, 2006; Nibali et al., 2019).

A few studies have also investigated the healing potential of infrabony defects following periodontal regeneration in relation to defect architecture, suggesting that narrower defects surrounded by higher numbers of bony walls have higher regenerative potential (Ellegaard & Loe, 1971; Selvig, Kersten, & Wikesjo, 1993; Tsitoura et al., 2004). However, the effect of defect morphology on treatment outcomes following periodontal surgery has not been investigated systematically, perhaps owing to the lack of a clear classification system for osseous defects. Therefore, the aim of this review is to examine the relationship between intrabony defect morphology and treatment outcomes.

MATERIALS AND METHODS

A systematic review protocol was written in the planning stages and the PRISMA statement (Moher, Liberati, Tetzlaff, Altman, & Group, 2009) was followed both in the planning and reporting of the review (checklist attached as Supplemental Material 1). The protocol was registered on 26/03/2020 with PROSPERO (available from ID: 176697).

Focused questions

The present review aimed to answer two focused questions:

- How often and how well is defect morphology described in papers reporting regenerative therapy of periodontal infrabony defects (defect depth, number of walls, extension of the defect, defect angle)?
- How does defect morphology predict the outcomes of regenerative therapy of periodontal infrabony defects?

Eligibility criteria

In brief, the **PECOS** method was the following:

- (P) Participants: Adult human patients with periodontitis who have completed a cycle of non-surgical periodontal therapy and present with residual pockets and infrabony defects.
- (E) Exposure: Defect morphology (depth, angle, number of walls) in defects undergoing mucoperiosteal surgery including regenerative surgery with Guided Tissue Regeneration (GTR), Enamel Matrix Derivative (EMD), bone fillers or substitutes, growth factors (GF) or combination.
- (C) Comparisons: Different types of intrabony defect morphology and different types of biomaterials used.
- (O) Outcomes: CAL gain, PPD reduction and radiographic bone gain
- (S) Studies: Randomized controlled trials (RCTs), cohort studies or case series

The following additional criteria were considered:

Inclusion criteria: i) Definition of periodontal infrabony defects at least 3mm deep; ii) with at least 12-months follow-up, iii) only studies published in English. Exclusion criteria: i) animal studies, ii) reviews, iii) including less than 20 patients; iv) studies including patients with diabetes or immunocompromised.

Information sources and Search

Papers were searched on MEDLINE, Cochrane Database and Scopus databases (search details are reported in supplemental material 2).

Study characteristics

This systematic review focused specifically on intrabony defect morphology and on its impact on regenerative treatment outcomes. Data extraction was performed in duplicate (authors DS and CA) including description of the infrabony defect and treatment outcomes by defect depth, defect angle and defect type (1- wall, 2-wall, 3-wall or more description if available). The exposure 'defect morphology' was assessed as:

- Description of defect depth and width/angle

- Description of number of defect walls, divided into craters, 1-wall, 2-wall, 3-wall or combination
- Description of extension of defect to buccal and/or lingual walls, for example following the definition of trench (Karn, Shockett, Moffitt, & Gray, 1984) or circumferential defects or 'moats'(Karn et al., 1984)
- Description of materials used
- Description of study outcomes (clinical, radiographic, patient-reported) divided by defect type and materials used

Risk of bias analysis

In order to assess the quality of the included studies, Risk of Bias was assessed using the Cochrane Collaborations Tool for assessing risk of bias for RCTs, the Newcastle Ottawa Tool for cohort studies and the The Modified Delphi Tool for case series. Assessment across all key domain was summarized in order to draw a conclusion of the overall risk of bias within and across trials. This judgment was made independently by two reviewers (DS, CA); any discrepancies were resolved by discussion.

Summary measures and planned method of analysis

Studies were initially divided by reporting of defect morphology (based on criteria above). Among publications reporting defect morphology (even if just one aspect of defect morphology was reported), study outcomes were investigated and compared with type of regenerative materials used (when possible) and by defect morphology reported. A meta-analysis was considered appropriate and was performed in the presence of at least two studies with the same follow-up and reporting the same data. The outcomes of interest were CAL gain, PPD reduction and radiographic bone gain. The impact of initial defect depth, defect walls, and defect angle on bone gain and CAL gain were pooled and weighted mean difference (WMD) were estimated using a computer program [Review Manager (RevMan). Version 5.0. Copenhagen; The Nordic Cochrane Centre, The Cochrane Collaboration, 2008]. In addition, the coefficient estimates and the standard errors of the investigated variables, including defect depth, defect angle, and number of walls, from each publication were also pooled to assess the odds ratio (OR) and 95% confidence interval (CI) of the primary and secondary outcomes. The contribution of each article was weighed. Forest plots were produced to graphically represent the difference in outcomes. A p value= 0.05 was used as the

level of significance. Heterogeneity was assessed with p value for chi-square test. Random effects meta-analyses of the selected studies were applied if the p value for chi-square test was > 0.05. Fixed effects meta-analyses were applied if the p value for chi-square test was ≤ 0.05 to avoid any bias being caused by methodological differences among studies. In addition, the funnel plot was used to assess the presence of the publication bias. The reporting of these meta-analyses adhered to the PRISMA (Preferred Reporting Items for Systematic Review and Meta-Analyses) statement (Liberati et al., 2009).

Evaluation of the strength of evidence

Evidence provided by RTCs was rated using different levels of methodological strength modified from GRADE (grading of recommendations assessments development and evaluation) (Guyatt et al., 2008). Three different strength of evidence were considered:

- High: At least 3 RCTs at low risk of bias and low heterogeneity ($I^2 < 30\%$).
- Moderate: More than 1 RCT and at least 1 RCT at low risk of bias, low heterogeneity.
- Low: Lack of RCTs or RCTs at high risk of bias or high heterogeneity.

RESULTS

Supplemental material 3 presents the flow chart from initial search to included papers. The initial search generated 4487 articles from Medline, Cochrane Library Database and Scopus combined. After screening the titles and abstracts, 404 articles were considered potentially suitable by at least one reviewer and qualified for full text screening. Following full text reading, 143 articles (from 136 original publications) met the defined inclusion criteria, while 261 were excluded. The reasons for exclusion are detailed in supplemental material 3. A total of 117 RCTs, 20 cohort studies and 6 case series were included. The publication year ranged from 1992 to 2019. The Cohen's kappa value for inter reviewer agreement was 0.57 at title and abstract screening level (94.7% agreement) and 0.87 at second screening (94.3% agreement). Every effort was made to retrieve original data from authors when needed. Data from 15 publications were available for meta-analyses. One study data (Cosyn, Cleymaet, Hanselaer, & De Bruyn, 2012) were retrieved from the study team but excluded from the meta-analysis due to high heterogeneity detected through subgroup analysis (CAL gain and bone gain data had opposite directions of association

based on defect morphology). Therefore, only 14 publications were included for quantitative analyses as detailed below for the different analyses.

Effect of defect morphology on regenerative outcomes

Defect depth: 114 papers reported average or range of defect depth measured radiographically and/or intra-surgically (often subdivided by study arm). Sixteen papers (from 15 publications) presented separate results for defects of different depth (see Table 1). Figure 1 reports forest plots for meta-analysis of defect depth data. Meta-analysis was carried out for the effect of initial defect depth > 4 mm vs. \leq 4 mm on radiographic bone gain (in mm) at 12 months, including 3 papers (125 sites). A statistically significant association was found between defect depth > 4 mm and increased bone gain (-0.75 mm, 95% CI -1.12, -0.38) with moderate heterogeneity (I²=57%) (Fig.1 A, categorical analysis with 4 mm threshold). Six publications (including 314 sites) were included in meta-analysis of the regression estimates for the effect of initial defect depth on radiographic bone gain at 12 months, showing a statistically significant association between deeper defect depth and increased bone gain (OR=1.32, 95% CI= 1.19, 1.47, I²=0) (Fig.1 B, continuous analysis). Subgroup analysis by studies using GTR or EMD (with or without adjuncts) showed a significant association between deeper defect depth and increased bone gain of similar magnitude for both (see supplemental material 4). Four studies (n=292 sites) were included in meta-analysis of the regression estimates for the effect of initial depth on CAL gain at 12 months, showing no statistically significant associations (Fig. 1 C).

Defect angle: 36 papers reported data on defect width/angle. Ten papers reported treatment outcomes by defect angle (see Table 2). However, these studies could not be meta-analysed together, owing to heterogeneity in reporting data. Figure 2 reports forest plots for meta-analysis of defect angle data. Two studies (n=91 sites) were included in meta-analysis of the regression estimates for the effect of initial defect angle on radiographic bone gain at 12 months. A statistically significant association was found between defect angle < 37° and increased bone gain (0.94 mm, 95% CI 0.48, 1.39) with moderate heterogeneity (I²= 50%) (Fig. 2 A, categorical analysis with 37° threshold). Three studies (n=201 sites) were included in meta-analysis of the regression estimates for the effect of initial defect angle on radiographic bone gain at 12 months, showing no statistically significant associations (Fig. 2 B, continuous analysis). Four studies (n=274 sites) were included in meta-analysis of the regression estimates for the effect of initial defect angle on radiographic bone gain at 12 months, showing no statistically significant associations (Fig. 2 B, continuous analysis). Four studies (n=274 sites) were included in meta-analysis of the regression estimates for the effect of initial defect angle on radiographic bone gain at 12 months, showing no statistically significant associations (Fig. 2 B, continuous analysis).

defect angle on CAL gain at 12 months, showing a statistically significant association between narrower angles and increased CAL gain (OR=0.97, 95% CI= 0.95, 0.98) with low-moderate heterogeneity (I^2 = 33%) (Fig. 2 C).

Number of walls: 122 papers reported defect morphology as described below (see supplemental material 5):

- o 1-, 2- and 3-walled defects or combinations: 78 papers
- o 1- or 2- walled defects or combinations only: 14 papers
- o 1-, 2- walled, combined 1-2 or circumferential defects: 5 papers
- o 2-walled defects only: 1 paper
- o 2- or 3-walled defects or combinations: 19 papers
- o 3-walled defects only: 1 paper
- o 1-walled defects only (or mainly 1-walled): 4 papers

Out of 122 papers reporting defect morphology 87 reported breakdowns of different types of defects included (based on defect morphology details above), while 35 did not. Out of 87 papers reporting breakdowns of different defects by number of walls, only 17 reported treatment outcomes for defects divided by baseline defect morphology. However, in 3 cases (Briguglio, Briguglio, Briguglio, Cafiero, & Isola, 2013; Crea, Dassatti, Hoffmann, Zafiropoulos, & Deli, 2008; Xu et al., 2019) only 1 type of defect was included in the study, so no comparison across different types of defects was possible. The remaining 14 papers are reported in Table 3.

Figure 3 reports forest plots for meta-analysis of number of walls data. Three publications (n= 150 sites) reported data on bone gain after regenerative treatment between 1-wall and 2-wall defects and showed significant radiographic bone gain at 12 months favoring 2-wall defects (-0.57 mm, 95% CI= -0.93, -0.21) with low heterogeneity (I²=0%) (Fig. 3 A). Two publications (n= 108 sites) reported data on radiographic bone gain 12 months after regenerative treatment between 2-wall and 3-wall defects and showed significant bone gain favoring 3-wall defects (-0.39 mm, 95% CI= -0.78, -0.01) with moderate heterogeneity (I²=54%) (Fig. 3 B). In addition, only one publication (Cortellini et al. 1993) reported data on radiographic bone gain 12 months after regenerative treatment between 1-wall and 3-wall defects and reported a significant bone gain favoring 3-wall defects (-1.18 mm, 95% CI= -1.66, -0.71).

Meta-analysis of regression estimates for radiographic bone gain at 12 months including two studies (n=101 sites) revealed a statistically significant association (OR= 3.43, 95% CI= 1.09, 10.85) with low heterogeneity (I²=0%) (Fig. 3 C). Meta-analysis of regression estimates for CAL gain at 12 months including five papers (n=431 sites) revealed a statistically significant association between more bone walls and increased CAL gain (OR=1.42, 95% CI= 1.14, 1.77) with high heterogeneity (I²=80%) (Fig. 3 D). Results of meta-analyses including Cosyn et al. 2012 are reported in supplemental material 6.

Not enough publications were available to analyze the outcome PPD.

Overall summary of results for defect depth, angle and number of walls, with associated level of evidence, is reported in table 4.

Risk of bias analysis

Supplemental materials 7, 8 and 9 report risk of bias analysis for RCT, cohort studies and case series. Risk of bias score for RCTs ranged from 0 to 10. A total of 75 papers were defined as 'good quality', 27 as 'fair' and 15 as 'poor' quality. Aspects which recorded highest risk of bias scores were allocation concealment bias and performance bias, while the area that according to our scoring showed lowest score was reporting bias. Risk of bias scores for cohort studies ranged from 5 to 8 stars, with the item 'comparability' often scored as 0. Eleven papers were identified as 'good quality' and the remaining 9 as 'poor quality'. Risk of bias scores for case series ranged from 11 to 16 out of 18. The item 'study population' was often scored low.

DISCUSSION

This systematic review investigated the effect of infrabony defect morphology on outcomes of periodontal regenerative therapy. The effect of baseline defect depth, defect angle and number of walls on radiographic bone and CAL gain was investigated. This review produced probably the largest body of systematically assessed evidence to suggest that deeper, narrower defects and defects with more walls are associated with improved clinical and radiographic outcomes 12 months post-regenerative surgery.

High strength of evidence suggests that deeper defects are associated with more radiographic bone gain at 12 months (both continuous and categorical analyses). The magnitude of additional radiographic bone gain was approximately 0.7 mm for defects initially deeper than 4 mm compared with those 3-4 mm deep. It is interesting to notice that the association between defect depth and bone gain seems to occur irrespective of biomaterials used and was of the same magnitude for EMD (including studies with or without adjunctive bone replacement grafts) or GTR. On the other hand, the more clinically meaningful CAL outcome was not associated with baseline defect depth.

Low level of evidence suggests that narrower angles are associated with more radiographic bone gain (only at categorical analysis with 37° threshold) and with more CAL gain (magnitude approximately 1 mm more CAL gain for angle $< 37^{\circ}$). Furthermore, more walls are associated with more radiographic bone gain and CAL gain (magnitude approximately 0.5 mm per extra wall from 1 to 2 to 3). The increased chance of favorable treatment outcomes following periodontal surgery by number of residual walls has also been widely reported in publications not included in this review (Rosling, Nyman, Lindhe, & Jern, 1976). When categorizing infrabony defects by number of walls, we should not forget that often defects are categorized as 'combinations' and relative proportions of 1-, 2- or 3-wall components of the defects are reported (P. Cortellini et al., 2008). The prevalence of 'combination' defects in some GTR studies was as high as 31% to 56% (Christgau et al., 2002; Falk, Laurell, Ravald, Teiwik, & Persson, 1997). Furthermore, a 1-wall component may be present in the majority of sites (P. Cortellini & Tonetti, 2011). A gradient effect on percentage of defect fill has been shown from the 3-wall component of the defects (95 \pm 6.2%) to the 2-wall component ($82 \pm 18.7\%$) and the 1-wall component ($39 \pm 62.4\%$) in a study using GTR (Cortellini et al., 1993). In contrast, other researchers reported limited influence of the defect's characteristics on the clinical outcome as defect characteristics showed weak or no correlations to defect fill (Polson & Heijl, 1978; Renvert, Garrett, Nilveus, Chamberlain, & Egelberg, 1985). It is clear that defect depth, narrow angle and increased number of walls are correlated, since usually the deepest part of the defect has increased walls and it is narrower. Therefore, it might be difficult to disentangle the relative contribution of each of these morphology aspects on regenerative surgery outcomes. It is also important to highlight that these observations, based on studies inclusion, were specific to defects with at least 3 mm radiographic infrabony component. These findings are further confounded by the fact that, in combination defects, the subcomponents with a lower number of walls are the more superficial ones, which may be negatively affected by the oral environment and the wound healing process.

Another important finding of this review is that, although most publications reported some description of the study defects, only a minority of publications did report outcomes based on defect morphology. This is somehow surprising, since it has long been suspected that regeneration of intraosseous defects is thought to depend on uninterrupted maturation of the fibrin clot, favoured by stability of the wound and good soft tissue coverage of the defect (Hiatt, Stallard, Butler, & Badgett, 1968; Wikesjo & Nilveus, 1990). As such, intraosseous defect morphology is believed a crucial factor to facilitate predictable regeneration by influencing stability of the blood clot. These initial theories are supported by observations in animal models, showing that bone and cementum regeneration was positively correlated to the number of bone walls limiting the infrabony periodontal defects (Kim et al., 2004). Interestingly, the description of defect morphology in the included papers was limited to depth, angle and number of residual walls in the interproximal area and not to whether the defects extended to buccal and/or lingual walls, with the exception of a few studies including circumferential defects (Al Machot, Hoffmann, Lorenz, Khalili, & Noack, 2014; Hoffmann, Al-Machot, Meyle, Jervoe-Storm, & Jepsen, 2016; Meyle et al., 2011). Such extension is often pivotal for decision making on biomaterials to be used and on flap design (for example MIST vs. M-MIST or single-flap approach). No mention of 'craters' was found in the reviewed papers, although other papers not included in this review have described attempted regeneration of this type of defects (Falk et al., 1997). Therefore, no meaningful conclusion can be drawn on the regenerative potential of craters and on the potential effect of defect extension on outcomes of regenerative therapy.

Different types of regenerative materials can be adapted to the defect morphology: some materials are supportive and space-maintaining, like non-resorbable membranes, bone grafts, and combination of resorbable membrane and bone grafts, and others are non-supportive and non-space maintaining materials, like resorbable membranes alone, enamel matrix proteins, and growth factors (Pierpaolo Cortellini & Tonetti, 2015). Some publications have suggested that supportive biomaterials may overcome the negative effect of defect morphology and improve the outcomes of regeneration in non-space maintaining defects as they have the ability to create and maintain space

for regeneration and provide increased stability to the blood clot (Palmer, Cortellini, & Group, 2008; Reynolds, Aichelmann-Reidy, Branch-Mays, & Gunsolley, 2003; Slotte, Asklow, Sultan, & Norderyd, 2012; Tonetti et al., 2004; Tonetti et al., 1993; Tonetti, Prato, et al., 1996; Trombelli & Farina, 2008). On the other hand, for non-supportive biomaterials, such as EMD, their added benefit is thought to be greater in defects with a predominantly 3-wall anatomy compared with one-wall defect (Tonetti et al., 2002). However, these concepts are in constant evolution together with developments in the surgical procedure itself, specifically flap design and suturing technique. The most common complications of periodontal regeneration procedures were dehiscence of interdental tissues, graft exfoliation, membrane collapse and/or exposure, with the subsequent bacterial contamination which negatively affects the outcomes of regeneration, such as CAL gain and bone gain (P. Cortellini et al., 1993; P. Cortellini et al., 2001; Selvig, Kersten, Chamberlain, Wikesjo, & Nilveus, 1992). Therefore, new surgical techniques were developed with the aim of soft tissue preservation in order to achieve tension-free primary closure over the defect and the regenerative materials, and to ensure wound stability and blood clot stability during the early healing phase. It has been shown that stable flap designs, such as achieved by minimally-invasive surgical therapy (MIST) can lead to such favorable regenerative outcomes that the use of regenerative materials may not offer any additional benefits (Liu, Hu, Zhang, Li, & Song, 2016). No clear effect of biomaterials (supportive or not) on these results, although the only factor where this could be formally analyzed was defect depth.

A recent systematic review has concluded that EMD and GTR with resorbable membranes appear to be the gold standard for the surgical treatment of deep (\geq 3mm) infrabony defects which have not resolved following completion of non-surgical therapy and that among the possible replacement biomaterials, Deproteinized Bovine Bone Mineral (DBBM) improved clinical outcomes of both EMD and resorbable GTR compared with OFD and it should be considered a viable treatment option especially in non-supporting defects (Nibali et al., 2019). These authors also suggested that papillary preservation flaps may improve the clinical outcomes and should be considered a surgical pre-requisite when performing any regeneration procedure (Nibali et al., 2019). These observations are the basis for the recently published EFP S3 clinical guidelines (Sanz et al., 2020). The topic of periodontal regeneration of periodontal infraosseous defects is developing quickly with other biologically active agents such as growth factors (Smith, Martínez, Cáceres, & Martínez, 2015) and bone morphogenetic proteins (Larsson et al., 2016). Therefore, the frontier of what is 'regenerable' is quickly moving and the 'bar' is being raised.

Strengths of this review are the analysis of a large body of literature and the relatively low heterogeneity, leading to moderate to high strength of evidence for most meta-analyses. The exclusion of one paper (Cosyn et al., 2012) significantly reduced heterogeneity for the radiographic bone gain outcomes. Risk of bias revealed that only a minority of papers were defined as 'poor quality' across all study designs, with 'reporting bias' for RCT and 'comparability' for cohort studies resulting as areas requiring improvement. The main limitation of this review is that despite the inclusion of more than a hundred papers, only 15 papers could be included in meta-analyses, due to limited or heterogeneous data reporting.

From these data, it emerges clearly how infrabony defect morphology has an important influence on outcomes of regenerative periodontal surgery. Baseline defect depth seems to positively influence radiographic bone gain 12 months post-surgery, while narrower angles and increased number of walls positively influence both bone and CAL gain. A good description and definition of the infraosseous defects can help in planning the most appropriate treatment option. Such specific definition can only really be obtained intra-surgically or perhaps through CBCT scan, although combined accurate assessment of probing pocket depths and periapical radiographs has good value (Wolf, von Bethlenfalvy, Hassfeld, Staehle, & Eickholz, 2001) and should be sufficient for treatment planning of most cases. A detailed classification system for infraosseous defects, which takes into account also other aspects of defect morphology, such as extension to buccal-lingual walls, should be used widely to improve our understanding of regenerative potential and of appropriate biomaterials for different types of defects.

REFERENCES

- Al Machot, E., Hoffmann, T., Lorenz, K., Khalili, I., & Noack, B. (2014). Clinical outcomes after treatment of periodontal intrabony defects with nanocrystalline hydroxyapatite (Ostim) or enamel matrix derivatives (Emdogain): a randomized controlled clinical trial. *Biomed Res Int, 2014*, 786353. doi:10.1155/2014/786353
- Bratthall, G., Lindberg, P., Havemose-Poulsen, A., Holmstrup, P., Bay, L., Soderholm, G., . . . Skold Bell, H. (2001). Comparison of ready-to-use EMDOGAIN-gel and EMDOGAIN in

patients with chronic adult periodontitis. *J Clin Periodontol, 28*(10), 923-929. doi:10.1034/j.1600-051x.2001.028010923.x

- Briguglio, F., Briguglio, E., Briguglio, R., Cafiero, C., & Isola, G. (2013). Treatment of infrabony periodontal defects using a resorbable biopolymer of hyaluronic acid: a randomized clinical trial. *Quintessence Int*, 44(3), 231-240. doi:10.3290/j.qi.a29054
- Castro, A. B., Meschi, N., Temmerman, A., Pinto, N., Lambrechts, P., Teughels, W., & Quirynen, M. (2017). Regenerative potential of leucocyte- and platelet-rich fibrin. Part A: intra-bony defects, furcation defects and periodontal plastic surgery. A systematic review and meta-analysis. *J Clin Periodontol, 44*(1), 67-82. doi:10.1111/jcpe.12643
- Christgau, M., Bader, N., Felden, A., Gradl, J., Wenzel, A., & Schmalz, G. (2002). Guided tissue regeneration in intrabony defects using an experimental bioresorbable polydioxanon (PDS) membrane. A 24-month split-mouth study. J Clin Periodontol, 29(8), 710-723. doi:10.1034/j.1600-051x.2002.290808.x
- Cortellini, P., Nieri, M., Prato, G. P., & Tonetti, M. S. (2008). Single minimally invasive surgical technique with an enamel matrix derivative to treat multiple adjacent intra-bony defects: clinical outcomes and patient morbidity. *J Clin Periodontol*, 35(7), 605-613. doi:10.1111/j.1600-051X.2008.01242.x
- Cortellini, P., Pini Prato, G., & Tonetti, M. S. (1993). Periodontal regeneration of human infrabony defects. II. Re-entry procedures and bone measures. *J Periodontol*, *64*(4), 261-268. doi:10.1902/jop.1993.64.4.261
- Cortellini, P., Stalpers, G., Mollo, A., & Tonetti, M. S. (2020). Periodontal regeneration versus extraction and dental implant or prosthetic replacement of teeth severely compromised by attachment loss to the apex: A randomized controlled clinical trial reporting 10-year outcomes, survival analysis and mean cumulative cost of recurrence. *J Clin Periodontol, 47*(6), 768-776. doi:10.1111/jcpe.13289
- Cortellini, P., & Tonetti, M. S. (2011). Clinical and radiographic outcomes of the modified minimally invasive surgical technique with and without regenerative materials: a randomized-controlled trial in intra-bony defects. *J Clin Periodontol, 38*(4), 365-373. doi:10.1111/j.1600-051X.2011.01705.x
- Cortellini, P., & Tonetti, M. S. (2015). Clinical concepts for regenerative therapy in intrabony defects. *Periodontology 2000, 68*(1), 282-307. doi:10.1111/prd.12048

- Cortellini, P., Tonetti, M. S., Lang, N. P., Suvan, J. E., Zucchelli, G., Vangsted, T., . . . Adriaens,
 P. (2001). The simplified papilla preservation flap in the regenerative treatment of deep intrabony defects: clinical outcomes and postoperative morbidity. *J Periodontol*, 72(12), 1702-1712. doi:10.1902/jop.2001.72.12.1702
- Cosyn, J., Cleymaet, R., Hanselaer, L., & De Bruyn, H. (2012). Regenerative periodontal therapy of infrabony defects using minimally invasive surgery and a collagen-enriched bovine-derived xenograft: a 1-year prospective study on clinical and aesthetic outcome. *J Clin Periodontol, 39*(10), 979-986. doi:10.1111/j.1600-051X.2012.01924.x
- Crea, A., Dassatti, L., Hoffmann, O., Zafiropoulos, G. G., & Deli, G. (2008). Treatment of intrabony defects using guided tissue regeneration or enamel matrix derivative: a 3-year prospective randomized clinical study. *J Periodontol*, 79(12), 2281-2289. doi:10.1902/jop.2008.080135
- Crea, A., Deli, G., Littarru, C., Lajolo, C., Orgeas, G. V., & Tatakis, D. N. (2014). Intrabony
 defects, open-flap debridement, and decortication: a randomized clinical trial. J Periodontol, 85(1), 34-42. doi:10.1902/jop.2013.120753
- Ehmke, B., Rudiger, S. G., Hommens, A., Karch, H., & Flemmig, T. F. (2003). Guided tissue regeneration using a polylactic acid barrier. *J Clin Periodontol, 30*(4), 368-374. doi:10.1034/j.1600-051x.2003.00312.x
- Eickholz, P., Horr, T., Klein, F., Hassfeld, S., & Kim, T. S. (2004). Radiographic parameters for prognosis of periodontal healing of infrabony defects: two different definitions of defect depth. *J Periodontol*, 75(3), 399-407. doi:10.1902/jop.2004.75.3.399
- Eickholz, P., Rollke, L., Schacher, B., Wohlfeil, M., Dannewitz, B., Kaltschmitt, J., ... Kim, T. S.
 (2014). Enamel matrix derivative in propylene glycol alginate for treatment of infrabony defects with or without systemic doxycycline: 12- and 24-month results. *J Periodontol*, 85(5), 669-675. doi:10.1902/jop.2013.130290
- Ellegaard, B., & Loe, H. (1971). New attachment of periodontal tissues after treatment of intrabony lesions. *J Periodontol*, 42(10), 648-652. doi:10.1902/jop.1971.42.10.648
- Falk, H., Laurell, L., Ravald, N., Teiwik, A., & Persson, R. (1997). Guided tissue regeneration therapy of 203 consecutively treated intrabony defects using a bioabsorbable matrix barrier. Clinical and radiographic findings. *J Periodontol*, 68(6), 571-581. doi:10.1902/jop.1997.68.6.571

- Francetti, L., Trombelli, L., Lombardo, G., Guida, L., Cafiero, C., Roccuzzo, M., . . . Del Fabbro,
 M. (2005). Evaluation of efficacy of enamel matrix derivative in the treatment of intrabony defects: a 24-month multicenter study. *Int J Periodontics Restorative Dent, 25*(5), 461-473.
- Goldman, H. M., & Cohen, D. W. (1958). The Infrabony Pocket: Classification and Treatment. *The Journal of Periodontology*, 29(4), 272-291. doi:10.1902/jop.1958.29.4.272
- Grusovin, M. G., & Esposito, M. (2009). The efficacy of enamel matrix derivative (Emdogain) for the treatment of deep infrabony periodontal defects: a placebo-controlled randomised clinical trial. *Eur J Oral Implantol, 2*(1), 43-54.
- Guyatt, G. H., Oxman, A. D., Vist, G. E., Kunz, R., Falck-Ytter, Y., Alonso-Coello, P., . . . Group,
 G. W. (2008). GRADE: an emerging consensus on rating quality of evidence and strength of recommendations. *BMJ*, 336(7650), 924-926. doi:10.1136/bmj.39489.470347.AD
- Heins, P., Hartigan, M., Low, S., & Chace, R. (1989). Relative stability of deep- versus shallow-side bone levels in angular proximal infrabony defects. *J Clin Periodontol*, 16(1), 59-64. doi:10.1111/j.1600-051x.1989.tb01613.x
- Hiatt, W. H., Stallard, R. E., Butler, E. D., & Badgett, B. (1968). Repair following mucoperiosteal flap surgery with full gingival retention. *J Periodontol*, *39*(1), 11-16. doi:10.1902/jop.1968.39.1.11
- Hoffmann, T., Al-Machot, E., Meyle, J., Jervoe-Storm, P. M., & Jepsen, S. (2016). Three-year results following regenerative periodontal surgery of advanced intrabony defects with enamel matrix derivative alone or combined with a synthetic bone graft. *Clin Oral Investig*, 20(2), 357-364. doi:10.1007/s00784-015-1522-4
- Ilgenli, T., Dundar, N., & Kal, B. I. (2007). Demineralized freeze-dried bone allograft and plateletrich plasma vs platelet-rich plasma alone in infrabony defects: a clinical and radiographic evaluation. *Clin Oral Investig*, *11*(1), 51-59. doi:10.1007/s00784-006-0083-y
- Karn, K. W., Shockett, H. P., Moffitt, W. C., & Gray, J. L. (1984). Topographic classification of deformities of the alveolar process. *J Periodontol*, 55(6), 336-340. doi:10.1902/jop.1984.55.6.336
- Kim, C. S., Choi, S. H., Chai, J. K., Cho, K. S., Moon, I. S., Wikesjo, U. M., & Kim, C. K. (2004).
 Periodontal repair in surgically created intrabony defects in dogs: influence of the number of bone walls on healing response. *J Periodontol*, 75(2), 229-235. doi:10.1902/jop.2004.75.2.229

- Klein, F., Kim, T. S., Hassfeld, S., Staehle, H. J., Reitmeir, P., Holle, R., & Eickholz, P. (2001).
 Radiographic defect depth and width for prognosis and description of periodontal healing of infrabony defects. *J Periodontol*, 72(12), 1639-1646. doi:10.1902/jop.2001.72.12.1639
- Larsson, L., Decker, A. M., Nibali, L., Pilipchuk, S. P., Berglundh, T., & Giannobile, W. V.
 (2016). Regenerative Medicine for Periodontal and Peri-implant Diseases. *J Dent Res*, 95(3), 255-266. doi:10.1177/0022034515618887
- Liberati, A., Altman, D. G., Tetzlaff, J., Mulrow, C., Gotzsche, P. C., Ioannidis, J. P., . . . Moher, D. (2009). The PRISMA statement for reporting systematic reviews and meta-analyses of studies that evaluate health care interventions: explanation and elaboration. *J Clin Epidemiol*, 62(10), e1-34. doi:10.1016/j.jclinepi.2009.06.006
- Linares, A., Cortellini, P., Lang, N. P., Suvan, J., Tonetti, M. S., & European Research Group on,
 P. (2006). Guided tissue regeneration/deproteinized bovine bone mineral or papilla preservation flaps alone for treatment of intrabony defects. II: radiographic predictors and outcomes. *J Clin Periodontol, 33*(5), 351-358. doi:10.1111/j.1600-051X.2006.00911.x
- Liu, S., Hu, B., Zhang, Y., Li, W., & Song, J. (2016). Minimally Invasive Surgery Combined with Regenerative Biomaterials in Treating Intra-Bony Defects: A Meta-Analysis. *PLoS One*, *11*(1), e0147001. doi:10.1371/journal.pone.0147001
- Loos, B. G., Louwerse, P. H., Van Winkelhoff, A. J., Burger, W., Gilijamse, M., Hart, A. A., & van der Velden, U. (2002). Use of barrier membranes and systemic antibiotics in the treatment of intraosseous defects. *J Clin Periodontol, 29*(10), 910-921. doi:10.1034/j.1600-051x.2002.291006.x
- Losada, M., Gonzalez, R., Garcia, A. P., Santos, A., & Nart, J. (2017). Treatment of Non-Contained Infrabony Defects With Enamel Matrix Derivative Alone or in Combination With Biphasic Calcium Phosphate Bone Graft: A 12-Month Randomized Controlled Clinical Trial. *J Periodontol*, 88(5), 426-435. doi:10.1902/jop.2016.160459

Melcher, A.H. (1976) On the repair potential of periodontal tissues. J Periodontol 47:256-260.

Meyle, J., Hoffmann, T., Topoll, H., Heinz, B., Al-Machot, E., Jervoe-Storm, P. M., . . . Jepsen, S. (2011). A multi-centre randomized controlled clinical trial on the treatment of intra-bony defects with enamel matrix derivatives/synthetic bone graft or enamel matrix derivatives alone: results after 12 months. *J Clin Periodontol*, *38*(7), 652-660. doi:10.1111/j.1600-051X.2011.01726.x

- Moher, D., Liberati, A., Tetzlaff, J., Altman, D. G., & Group, P. (2009). Preferred reporting items for systematic reviews and meta-analyses: the PRISMA statement. *J Clin Epidemiol*, 62(10), 1006-1012. doi:10.1016/j.jclinepi.2009.06.005
- Needleman, I. G., Worthington, H. V., Giedrys-Leeper, E., & Tucker, R. J. (2006). Guided tissue regeneration for periodontal infra-bony defects. *Cochrane Database Syst Rev*(2), CD001724. doi:10.1002/14651858.CD001724.pub2
- Nevins, M., Kao, R. T., McGuire, M. K., McClain, P. K., Hinrichs, J. E., McAllister, B. S., ...
 Giannobile, W. V. (2013). Platelet-derived growth factor promotes periodontal regeneration in localized osseous defects: 36-month extension results from a randomized, controlled, double-masked clinical trial. *J Periodontol, 84*(4), 456-464. doi:10.1902/jop.2012.120141
- Nibali, L., Koidou, V. P., Nieri, M., Barbato, L., Pagliaro, U., & Cairo, F. (2019). Regenerative surgery versus access flap for the treatment of intrabony periodontal defects. A systematic review and meta-analysis. *J Clin Periodontol*. doi:10.1111/jcpe.13237
- Nyman, S., Lindhe, J., Karring, T., Rylander, H. (1982) New attachment following surgical treatment of human periodontal disease. *J Clin Periodontol* 9: 290-296
- Palmer, R. M., Cortellini, P., & Group, B. o. E. W. o. P. (2008). Periodontal tissue engineering and regeneration: Consensus Report of the Sixth European Workshop on Periodontology. J Clin Periodontol, 35(8 Suppl), 83-86. doi:10.1111/j.1600-051X.2008.01262.x
- Papapanou, P. N., & Tonetti, M. S. (2000). Diagnosis and epidemiology of periodontal osseous
 lesions. *Periodontol 2000, 22*, 8-21. doi:10.1034/j.1600-0757.2000.2220102.x
- Papapanou, P. N., & Wennstrom, J. L. (1991). The angular bony defect as indicator of further alveolar bone loss. *J Clin Periodontol*, *18*(5), 317-322. doi:10.1111/j.1600-051x.1991.tb00435.x
- Polson, A. M., & Heijl, L. C. (1978). Osseous repair in infrabony periodontal defects. J Clin Periodontol, 5(1), 13-23. doi:10.1111/j.1600-051x.1978.tb01902.x
- Pontoriero, R., Nyman, S., & Lindhe, J. (1988). The angular bony defect in the maintenance of the periodontal patient. J Clin Periodontol, 15(3), 200-204. doi:10.1111/j.1600-051x.1988.tb01570.x
- Renvert, S., Garrett, S., Nilveus, R., Chamberlain, A. D., & Egelberg, J. (1985). Healing after treatment of periodontal intraosseous defects. VI. Factors influencing the healing response. *J Clin Periodontol*, *12*(9), 707-715. doi:10.1111/j.1600-051x.1985.tb01396.x

- Reynolds, M. A., Aichelmann-Reidy, M. E., Branch-Mays, G. L., & Gunsolley, J. C. (2003). The efficacy of bone replacement grafts in the treatment of periodontal osseous defects. A systematic review. *Ann Periodontol*, 8(1), 227-265. doi:10.1902/annals.2003.8.1.227
- Rosling, B., Nyman, S., Lindhe, J., & Jern, B. (1976). The healing potential of the periodontal tissues following different techniques of periodontal surgery in plaque-free dentitions. A 2-year clinical study. *Journal of clinical periodontology*, *3*(4), 233-250. doi:10.1111/j.1600-051x.1976.tb00042.x
- Sanz, M., Herrera, D., Kebschull, M., Chapple, I., Jepsen, S., Beglundh, T., . . . Lang France Lambert, N. (2020). Treatment of Stage I-III Periodontitis -The EFP S3 Level Clinical Practice Guideline. *J Clin Periodontol*. doi:10.1111/jcpe.13290
- Sanz, M., Tonetti, M. S., Zabalegui, I., Sicilia, A., Blanco, J., Rebelo, H., . . . Suvan, J. E. (2004).
 Treatment of intrabony defects with enamel matrix proteins or barrier membranes: results from a multicenter practice-based clinical trial. *J Periodontol*, 75(5), 726-733. doi:10.1902/jop.2004.75.5.726
- Selvig, K. A., Kersten, B. G., Chamberlain, A. D., Wikesjo, U. M., & Nilveus, R. E. (1992).
 Regenerative surgery of intrabony periodontal defects using ePTFE barrier membranes: scanning electron microscopic evaluation of retrieved membranes versus clinical healing. *J Periodontol*, 63(12), 974-978. doi:10.1902/jop.1992.63.12.974
- Selvig, K. A., Kersten, B. G., & Wikesjo, U. M. (1993). Surgical treatment of intrabony periodontal defects using expanded polytetrafluoroethylene barrier membranes: influence of defect configuration on healing response. *J Periodontol, 64*(8), 730-733. doi:10.1902/jop.1993.64.8.730
- Silvestri, M., Sartori, S., Rasperini, G., Ricci, G., Rota, C., & Cattaneo, V. (2003). Comparison of infrabony defects treated with enamel matrix derivative versus guided tissue regeneration with a nonresorbable membrane. *J Clin Periodontol*, *30*(5), 386-393. doi:10.1034/j.1600-051x.2003.10146.x
- Slotte, C., Asklow, B., Sultan, J., & Norderyd, O. (2012). A randomized study of open-flap surgery of 32 intrabony defects with and without adjunct bovine bone mineral treatment. J *Periodontol*, 83(8), 999-1007. doi:10.1902/jop.2011.110490
- Smith, P. C., Martínez, C., Cáceres, M., & Martínez, J. (2015). Research on growth factors in periodontology. *Periodontology 2000*, 67(1), 234-250. doi:10.1111/prd.12068

- Stavropoulos, A., Karring, E. S., Kostopoulos, L., & Karring, T. (2003). Deproteinized bovine bone and gentamicin as an adjunct to GTR in the treatment of intrabony defects: a randomized controlled clinical study. *J Clin Periodontol, 30*(6), 486-495. doi:10.1034/j.1600-051x.2003.00258.x
- Tonetti, M. S., Cortellini, P., Lang, N. P., Suvan, J. E., Adriaens, P., Dubravec, D., . . . Zybutz, M. (2004). Clinical outcomes following treatment of human intrabony defects with GTR/bone replacement material or access flap alone. A multicenter randomized controlled clinical trial. *J Clin Periodontol*, *31*(9), 770-776. doi:10.1111/j.1600-051X.2004.00562.x
- Tonetti, M. S., Lang, N. P., Cortellini, P., Suvan, J. E., Adriaens, P., Dubravec, D., . . . Wallkamm,
 B. (2002). Enamel matrix proteins in the regenerative therapy of deep intrabony defects. *J Clin Periodontol, 29*(4), 317-325. doi:10.1034/j.1600-051x.2002.290407.x
- Tonetti, M. S., Pini Prato, G., Stalpers, G., & Cortellini, P. (1996). Guided tissue regeneration of deep intrabony defects in strategically important prosthetic abutments. *Int J Periodontics Restorative Dent, 16*(4), 378-387.
- Tonetti, M. S., Pini-Prato, G., & Cortellini, P. (1993). Periodontal regeneration of human intrabony defects. IV. Determinants of healing response. *J Periodontol*, *64*(10), 934-940. doi:10.1902/jop.1993.64.10.934
- Tonetti, M. S., Prato, G. P., & Cortellini, P. (1996). Factors affecting the healing response of intrabony defects following guided tissue regeneration and access flap surgery. *J Clin Periodontol*, 23(6), 548-556. doi:10.1111/j.1600-051x.1996.tb01823.x
- Trombelli, L., & Farina, R. (2008). Clinical outcomes with bioactive agents alone or in combination with grafting or guided tissue regeneration. *J Clin Periodontol*, 35(8 Suppl), 117-135. doi:10.1111/j.1600-051X.2008.01265.x
- Tsitoura, E., Tucker, R., Suvan, J., Laurell, L., Cortellini, P., & Tonetti, M. (2004). Baseline radiographic defect angle of the intrabony defect as a prognostic indicator in regenerative periodontal surgery with enamel matrix derivative. *J Clin Periodontol, 31*(8), 643-647. doi:10.1111/j.1600-051X.2004.00555.x
- Weinberg, M. A., & Eskow, R. N. (2000). Osseous defects: proper terminology revisited. J Periodontol, 71(12), 1928. doi:10.1902/jop.2000.71.12.1928
- Wikesjo, U. M., & Nilveus, R. (1990). Periodontal repair in dogs: effect of wound stabilization on healing. *J Periodontol*, 61(12), 719-724. doi:10.1902/jop.1990.61.12.719

- Wolf, B., von Bethlenfalvy, E., Hassfeld, S., Staehle, H. J., & Eickholz, P. (2001). Reliability of assessing interproximal bone loss by digital radiography: intrabony defects. *J Clin Periodontol*, 28(9), 869-878. doi:10.1034/j.1600-051x.2001.028009869.x
- Xu, Y., Qiu, J., Sun, Q., Yan, S., Wang, W., Yang, P., & Song, A. (2019). One-Year Results Evaluating the Effects of Concentrated Growth Factors on the Healing of Intrabony Defects Treated with or without Bone Substitute in Chronic Periodontitis. *Med Sci Monit*, 25, 4384-4389. doi:10.12659/MSM.917025
- Yukna, R. A. (1994). Clinical evaluation of coralline calcium carbonate as a bone replacement graft material in human periodontal osseous defects. *J Periodontol*, 65(2), 177-185. doi:10.1902/jop.1994.65.2.177
- Zucchelli, G., Amore, C., Montebugnoli, L., & De Sanctis, M. (2003). Enamel matrix proteins and bovine porous bone mineral in the treatment of intrabony defects: a comparative controlled clinical trial. *J Periodontol*, 74(12), 1725-1735. doi:10.1902/jop.2003.74.12.1725
- Zucchelli, G., Bernardi, F., Montebugnoli, L., & De Sanctis, M. (2002). Enamel Matrix Proteins and Guided Tissue Regeneration With Titanium-Reinforced Expanded PolytetrafluoroethyleneMembranes in the Treatment of Infrabony Defects: A Comparative Controlled Clinical Trial. *J Periodontol*, 73(1), 3-12. doi:10.1902/jop.2002.73.1.3

TABLES	

Author	Study characteristics	Results by intrabony defect depth
Sanz et al. 2004	EMD vs. GTR (Guidor)	Intrabony defect depth did not influence significantly CAL gain.
		Estimate= -0.4 ± 0.2 , P-value= 0.07
Meyle et al. 2011	EMD/synthetic bone graft	Deeper intrabony defect depth was associated with more defect fill
	vs. EMD	(estimate= 3.068, P-value= 0.003)
Loos et al. 2002	GTR (Guidor)/Antibiotic vs.	Intrabony defect depth did not significantly influence bone gain (P-
	GTR alone vs.	value= 0.38)
	OFD/Antibiotic vs. OFD	
	alone	
Grusovin &	EMD vs. Placebo	Initial intrabony defect depth didn't significantly influence CAL gain
Esposito 2009		and radiographic bone gain at 1 year. P-value= 0.41 and 0.81
		respectively
Ehmke et al 2003	GTR (Guidor)	Intrabony defect depth significantly influenced alveolar bone gain (b-
		weight \pm SD= 0.32 \pm 0.15, P-value= 0.045)
Tonetti et al 1996	GTR Titanium ePTFE vs.	Borderline significance for initial intabony defect depth on CAL gain
	GTR ePTFE vs. OFD	at 1 year (P-value= 0.055)
Klein et al. 2001	GTR (ePTFE/bioabsorbable)	Statistically significant positive influence of baseline intrabony depth
		on bone gain (P-value= <0.0001). More bone fill for initially deep
		intrabony defects (≥3 mm) but no association with CAL gain
Eickholz et al.	GTR (ePTFE/bioabsorbable)	Deep (≥4 mm) infrabony defects exhibited statistically significantly
2004		more favorable bony fill than defects ≤ 4 mm (bone fill 2.50 ± 1.99 ar
		-0.57 \pm 2.16 respectively). Intrabony defect depth had statistically
		significant positive influence on bone fill (estimate= 0.314, P-value=
		0.033)
Eickholz et al.	EMD/Doxycycline vs.	Baseline intrabony defect depth influenced bone gain positively (P-
2014	EMD/Placebo	value= 0.04)
Francetti et al.	EMD vs. OFD	Statistically significant difference in bone gain between the EMD and
2005		OFD groups only at 12 months in the \leq 6-mm subgroup (P-value= 0.0
		in favor to EMD)
Bratthall et al.	ready-to-use EMD-gel vs.	Higher CAL gain in sites with deeper baseline defects. Defects gaining
2001	marketed EMD	>4 mm had deeper bony defects at baseline compared to the other
2001		

1.33 respectively)

Т	onetti et al 2004	GTR (Bio-Guide)/Bio-oss vs. OFD	Depth of the intrabony component did not have a significant impact on CAL gain (estimate= 0.01 ± 0.09 , P-value= 0.8751)
Z	ucchelli et al.	EMP/bovine porous bone	Intrabony defect depth significantly influenced CAL gain (more CAL
20	003	mineral vs. EMP	gain in cases with deeper intrabony component, F-ratio= 19.62, P-value= 0.00001)
Z	ucchelli et al	EMD vs. GTR (ePTFE) vs.	Intrabony defect depth did not influence significantly CAL gain. F-
20	002	OFD	ratio= 2.01, P-value= 0.1603
Li	inares et al. 2006	GTR (collagen	Initial radiographic intrabony defect depth was a significant covariate
(s	ame clinical study	membrane)/Deproteinized	(p-value= 0.0001) to predict bone gain after 1 year
as	Tonetti et al.	Bovine Bone Mineral vs.	
20	004)	OFD	
Ilş	genli et al. 2007	Demineralized freeze-dried bone allograft/PRP vs. PRP	Initial defect depth was positively correlated to the bony fill (p-value= 0.047

Table 1. Details of papers reporting treatment outcomes divided by baseline defect depth.

Author	Study characteristics	Results by intrabony defect width/angle
Losada et al. 2017	EMD/Biphasic calcium	Probability of gaining ≥3mm of attachment diminished as the
	phosphate vs. EMD	angulation score increased (OR=2.57 higher if the treatment
		was performed in an angle <24.75 than in wider angles, but
		not statistically significant)
Cortellini et al.	EMD	CAL gain significantly associated with the baseline
2008		radiographic defect angle (estimate= -0.05, SE= 0.02, P-
		value= 0.0038)
Loos et al 2002	GTR	Radiographic defect angle did not influence significantly bo
	(Guidor)/Antibiotic vs.	gain (P-value= 0.20)
	GTR alone vs.	
	OFD/Antibiotic vs.	
	OFD alone	
Ilgenli et al. 2007	Demineralized freeze-	No significant differences between narrow and wide defects
	dried bone allograft/PRP	the PRP-alone therapy group (P-value= 0.89 for CAL gain &
		0.90 for defect bone fill). More CAL gain and defect bone fi

d		vs. PRP	in favor of the narrow defects (P-value was 0.03 for both) in the DFDBA + PRP group
	Eickholz et al. 2004	GTR (ePTFE/bioabsorbable)	Initially narrow (angle $<37^{\circ}$) defects exhibited statistically significantly more favorable bony fill than did wide defects (bone fill 2.30 ± 1.88 mm and -0.72 ± 2.49 mm respectively). Baseline defect angle had statistically significant positive influence on bone fill. Estimate= -0.064, P-value= 0.003
1	Zucchelli et al. 2003	EMP/bovine porous bone mineral vs. EMP	Defect angle did not influence significantly CAL gain (F- ratio= 2.20, P-value= 0.1439)
	Tonetti et al. 1993	GTR (GoreTex)	Defect angle significantly affected CAL gain and bone gain (estimate= -0.05/P-value= 0.0026, and estimate= -0.05/P- value= 0.0031 respectively)
	Tsitoura et al. 2004	EMD vs. OFD	Radiographic defect angle statistically significantly associated with CAL gain (p-value= 0.0477). The probability of obtaining CAL gain >3mm was 2.464 times higher (with a 95% confidence interval: 1.017– 5.970) when the radiographic defect angle was $\leq 22^{\circ}$, than when the radio- graphic defect angle was $\geq 36^{\circ}$
	Klein et al. 2001	GTR (ePTFE/bioabsorbable)	Initially narrow (angle $< 26^{\circ}$) infrabony defects exhibited more favorable CAL gain than wide defects (not statistically significant) and statistically significantly more favorable bony gain ($P < 0.05$)
	Linares et al.	GTR (collagen	Radiographic defect angle did not have a statistically
	2006	membrane)/Deproteiniz	significant effect on CAL gain (p-value= 0.8138).
		ed Bovine Bone Mineral	Radiographic defect angle did not have statistically significant
		vs. OFD	effect on bone gain either (p-value= 0.6179)
	Table 2. Details	s of papers reporting tre	eatment outcomes divided by baseline defect width/angle

Acce

AUTHORS	DEFECTS	REGENERATIVE	COMPARISON RESULT BY DEFECT TYPE
	INCLUDED	TREATMENT PROVIDED	
Cortellini et al.	1-, 2-, 3-wall, and	MIST+EMD	No statistically significant association for CAL
2008	combined		change at multilevel regression analysis for 3-wall vs
			other defects at 12 months (p=0.135)
Cortellini et al.	1-, 2-, 3-wall, and	Gore-Tex	Bone gain at 12 months: 3-wall=2.7 ±2.2, 2-
1993	combined		wall=1.6±1.6, 1-wall=0.4±1.6 (Bone fill: 95 ±6.2%,
			$82\pm\!18.7\%$ and $39\pm\!62.4\%$ for 3- , 2- and 1-wall
			components respectively)
Cosyn et al.	1-, 2-, 3-wall, and	M-MIST vs. MIST+Bio-oss	Association between 1-wall defects and both failure
2012	combined		(CAL gain < 2mm at 12 months, OR=10.4 for 1-wall
			vs. 2-wall defects) and increased buccal recession
			(OR=58.8 for 1- vs. 2-wall defects)
Crea et al. 2014	2-, 3-wall, or	OFD+ Intramarrow	Radiographic defect depth change: 2-wall OFD
	combined	Penetration IMP vs. OFD	(n=4):1.00 ± 1.82, OFD+IMP (n=13): 3.14 ± 1.36, 3-
			wall OFD (n=9): 2.00 ± 1.12, OFD+IMP (n=15):
			$3.00 \pm 1.76.$
			CAL change: 2-wall OFD (n=4):1.75 \pm 3.33,
			OFD+IMP (n=13): 3.00 ± 1.62, 3-wall OFD (n=9):
			1.78 ± 2.63, OFD+IMP (n=15): 3.14 ± 1.85
Loos et al.	1-, 2- or 3-wall	Bioresorbable membrane or	No statistically significant association between
2002		not (with or without systemic	number of walls and CAL gain
		antibiotics)	
Losada et al.	1-2 wall, or	EMD+BC vs. EMD	Probability of gaining \geq 3 mm CAL: 2.57 (0.36-
2017	combined		18.33) times higher for narrow defect angles
			(<24.75°) and 0.55 (0.16-1.92) for 1-wall vs. 2-wall
Meyle et al.	1-, 2-, combined	EMD + synthetic bone graft	Tukey's plot showing bone fill in each defect type
2011	1- and 2-wall,	vs. EMD	
	circumferential		
Silvestri et al.	2-, 3-wall	EMD vs. GTR	CAL gain: 3-wall= 5.02 ± 1.86 mm, 2-wall= $4.25\pm$
2003			2.34 mm
Stavropoulos et	1-, 2-, combined	GTR (resorbable membrane)	Estimated differences for 1-wall vs. 2-wall: PPD gain
al. 2003	1- and 2-wall	vs. resorbable membrane +	(0.40 mm, CI= -0.32;1.12, p=0.26), CAL gain (0.44
		Bio-Oss	mm, CI= 1.68; 0.74, p=0.44), bone gain (1.33 mm,
		impregnated with saline vs.	CI= 0.14; 2.53, p=0.03)
		resorbable membrane + with	
		Bio-	

		Oss impregnated with	
		gentamicin vs. flap surgery	
Tonetti et al.	1-, 2- and 3-wall	Papilla preservation flap with	OR of CAL gain ≥3mm: 3-wall vs. 1-wall: 2.69
2002		EMD vs. no regenerative	(CI=1.1-7.5)
		material	
Tonetti et al.	1-, 2- and 3-wall	Papilla preservation flap with	Estimated difference in CAL gain in 1- vs. 3-wall= -
2004		GTR vs. no regenerative	0.5 ± 0.04
		material	
Yukna et al.	1-, 2-, 3-wall, and	Calcium carbonate graft vs.	Relative Defect Fill number ($\geq 90\%$, $\geq 50\%$, $<50\%$,
1994	combined	OFD	<10% of defect) presented for each defect type
Tonetti et al.	1-, 2-, 3-wall, and	GTR Titanium ePTFE vs.	General linear model showing lack of significance of
1996	combined	GTR ePTFE vs. OFD	depth of 1-, 2- or 3-wall on 1-year CAL gain (p-
			value= 0.664, 0.24, 0.19 respectively)
Nevins et al.	1,2 wall,	β -TCP + sodium acetate (SA)	Graph showing the influence of defect type on CAL
2013	combined or	vs. β -TCP + SA + 0.3 mg/mL	gain with time
	circumferential	rhPDGF-BB vs. β -TCP + SA	
		+ 1.0 mg/mL rhPDGF-BB	

Table 3. Details of papers reporting treatment outcomes divided by baseline defect morphology.

Morphol	ogy parameter	Rad	liographic bone ga	in		CAL gain		Strength of
		Effect size if statistically	Heterogeneity (I ²)	Number of studies	Effect size if statistically	Heteroge neity (I ²)	Number of studies	evidence
		significant		with low RoB	significant		with low RoB	
Defect	Threshold	-0.75 mm	52%	3	No data	-	-	LOW
depth	(<4mm)							
	Regression	OR 1.32	0%	6	NS	4%	4	HIGH
	analysis							
Defect	Threshold	0.94 mm	50%	2	-	-	-	LOW
angle	(< 37°)							
	Regression	NS	80%	2	OR 0.97	33%	2	LOW-
	analysis							MODERATE
Number	1 vs. 2	-0.57 mm	0%	2	No data	-	-	MODERATE

of walls 2	vs. 3	-0.39 mm	54%	1	No data	-	-	LOW
1	vs. 3	-1.18 mm	-	0	No data	-	-	-
Re	egression	OR 3.43	0%	2	OR 1.42	80%	3	MODERATE-
an	nalysis							LOW

Table 4. Summary of all meta-analyses findings. OR= Odds Ratio, NS= not statistically significant, RoB= Risk of Bias. Regression analyses were carried out having outcomes (bone gain, CAL gain) as continuous variables.

FIGURE LEGENDS

Fig. 1. Forest plots of meta-analysis of effect of defect depth on healing following regenerative surgery: categorical analysis for the effect of defect depth > 4mm on radiographic bone gain at 12 months (1A), regression estimates for the effect of initial defect depth on radiographic bone gain at 12 months (1B), regression estimates for the effect of initial depth on CAL gain at 12 months (1C).

Fig. 2. Forest plots of meta-analysis of effect of defect angle on healing following regenerative surgery: categorical analysis for the effect of initial defect angle $< 37^{\circ}$ on radiographic bone gain at 12 months (2A), regression estimates for the effect of initial defect angle on radiographic bone gain at 12 months (2B), regression estimates for the effect of defect angle on CAL gain at 12 months (2C).

Fig. 3. Forest plots of meta-analysis of effect of number of walls on healing following regenerative surgery: categorical analysis for the effect of 1-wall vs. 2-wall defects on radiographic bone gain at 12 months (3A), categorical analysis for the effect of 2-wall vs. 3-wall on radiographic bone gain at 12 months (3B), regression estimates for the effect of number of walls on radiographic bone gain at 12 months (3C), regression estimates for the effect of number of walls on CAL gain at 12 months (3D).

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1A	Defect dept	th 4 mm o	r less	Defect depth > 4 mm			Defect depth > 4 mm Std. Mean Difference						Std. I	Mean Diffe	erence	
Study or Subgroup	Mean	SD	Total	Mean	SD	Total	Weight	IV, Fixed, 95% C	I	IV,	Fixed, 95	% CI				
Ehmke et al. 2003	1.3	1	12	1.9	1.5	17	24.5%	-0.44 [-1.19, 0.31]								
Eickholz et al. 2004	-0.92	1.92	21	1.87	2.25	29	35.6%	-1.30 [-1.92, -0.68]			-					
Losada et al. 2017	2.227	1.27	22	3.167	2.548	24	39.9%	-0.45 [-1.04, 0.13]			-					
Total (95% CI)			55			70	100.0%	-0.75 [-1.12, -0.38]			•					
Heterogeneity: Chi ² = Test for overall effect:			= 57%						⊢ -10 Fa	-5 avors > 4	0 mm Fav	5 ors 4 mm	10 or less			

1B			Odds Ratio	Odds Ratio
Study or Subgroup	log[Odds Ratio] S	E Weight	IV, Fixed, 95% C	I IV, Fixed, 95% CI
Ehmke et al. 2003	0.32 0.1	5 12.6%	1.38 [1.03, 1.85]	
Eickholz et al. 2004	0.314 0.08	5 39.3%	1.37 [1.16, 1.62]	
Eickholz et al. 2014	0.339 0.134	4 15.8%	1.40 [1.08, 1.83]	
Losada et al. 2017	0.534 1.40	2 0.1%	1.71 [0.11, 26.63]	
Meyle et al. 2011	0.202 0.09	5 31.4%	1.22 [1.02, 1.47]	
Stavropoulos et al. 2003	-0.4 0.62	2 0.7%	0.67 [0.20, 2.27]	
Total (95% CI)		100.0%	1.32 [1.19, 1.47]	•
Heterogeneity: Chi ² = 2.33	, df = 5 (P = 0.80); l ² = 0%	/ 0		
Test for overall effect: Z =	5.23 (P < 0.00001)			0.1 0.2 0.5 1 2 5 10 Favors shallow defect Favors deep defect

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1C				Odds Ratio			Od	ds Ra	ıtio		
Study or Subgroup	log[Odds Ratio]	SE	Weight	IV, Fixed, 95% CI			IV, Fix	(ed, 9	5% CI		
Losada et al. 2017	0.244	1.608	0.2%	1.28 [0.05, 29.83]	←						
Sanz et al. 2004	0.1	0.1	43.8%	1.11 [0.91, 1.34]							
Stavropoulos et al. 2003	-0.74	0.48	1.9%	0.48 [0.19, 1.22]				+			
Tonetti et al. 2004	0.01	0.09	54.1%	1.01 [0.85, 1.20]				-			
Total (95% CI)			100.0%	1.04 [0.91, 1.18]				•			
Heterogeneity: Chi ² = 3.12,	df = 3 (P = 0.37); l	² = 4%				0.2	0.5		- <u>+</u>	5	
Tost for overall effect: $\mathbf{Z} = 0$	54 (P - 0.50)				0.1	0.2	0.5	1	2	5	

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Favors shallow defect Favors deep defect

Test for overall effect: Z = 0.54 (P = 0.59)

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2A	defect an	gle < 37 de	egree	defect ang	gle >= 37 d	egree	St	d. Mean Difference		Std. M	/lean Differe	nce	
Study or Subgroup	Mean	SD	Total	Mean	SD	Total	Weight	IV, Fixed, 95% C	I	IV,	Fixed, 95%	CI	
Eickholz et al. 2004	1.72	2.19	18	-1.13	2	20	40.9%	1.33 [0.62, 2.05]					
Losada et al. 2017	3	2.204	36	1.7	1.059	17	59.1%	0.67 [0.08, 1.26]					
Total (95% CI)			54			37	100.0%	0.94 [0.48, 1.39]			•	•	
Heterogeneity: Chi ² = Test for overall effect:			l² = 50%						-4 Favors ang	-2 le >= 37 deg	0 gree Favors	2 s angle < 3	4 4 37 degree

2B			Odds Ratio	Odds Ratio
Study or Subgroup	log[Odds Ratio] S	E Weight	IV, Fixed, 95% C	I IV, Fixed, 95% CI
Eickholz et al. 2004	-0.064 0.02	2 45.1%	0.94 [0.90, 0.98]	— — —
Linares et al. 2006	0.01 0.0	2 54.6%	1.01 [0.97, 1.05]	
Losada et al. 2017	-0.502 0.25	2 0.3%	0.61 [0.37, 0.99]	·
Total (95% CI)		100.0%	0.98 [0.95, 1.00]	
Heterogeneity: Chi ² = 9	0.79, df = 2 (P = 0.007); I		-+ $+$ $+$ $+$ $+$ $+$ $+$ $+$ $+$ $+$	
Test for overall effect: 2	Z = 1.70 (P = 0.09)			Favors sharp angle Favors flat angle

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2C	leg[Odde Detic] SE V	Noight	Odds Ratio	Odds		
Study or Subgroup	log[Odds Ratio] SE V	Veight	IV, Fixed, 95% C		l, 95% Cl	
Cortellini et al. 2008	-0.05 0.02	14.3%	0.95 [0.91, 0.99]			
Linares et al. 2006	0.005 0.02	14.3%	1.01 [0.97, 1.05]			
Losada et al. 2017	-0.035 0.01	57.1%	0.97 [0.95, 0.98]			
Tsitoura et al. 2004	-0.04 0.02	14.3%	0.96 [0.92, 1.00]			
Total (95% CI)	1	00.0%	0.97 [0.95, 0.98]	•		
Heterogeneity: Chi ² = 4	4.48, df = 3 (P = 0.21); l ² = 3			 		
Test for overall effect: 2	,		0.85 0.9 1	1.1	1.2	
				Favors sharp angle	Favors flat ano	gle

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3A	1-wa	II defec	cts	2-wa	II defea	cts	:	Std. Mean Difference	Std. Mean Difference
Study or Subgroup	Mean	SD	Total	Mean	SD	Total	Weight	IV, Fixed, 95% CI	IV, Fixed, 95% CI
Cortellini et al. 1993	0.4	1.6	40	1.6	1.6	40	64.2%	-0.74 [-1.20, -0.29]	
Losada et al. 2017	2.3	1.703	10	2.833	2.171	36	26.8%	-0.25 [-0.95, 0.45]	
Meyle et al. 2011	2	1	3	2.45	1.619	21	9.0%	-0.28 [-1.49, 0.94]	
Total (95% CI)			53			97	100.0%	-0.57 [-0.93, -0.21]	•
Heterogeneity: Chi ² = Test for overall effect:		•		l² = 0%					-4 -2 0 2 4 Favors 2-wall defects Favors 1-wall defects

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3B	2-wa	II defe	cts	3-wa	ll defe	cts	ts Std. Mean Difference		Std. Mean Difference
Study or Subgroup	Mean	SD	Total	Mean	SD	Total	Weight	IV, Fixed, 95% CI	I IV, Fixed, 95% CI
Cortellini et al. 1993	1.6	1.6	40	2.7	2.2	40	73.4%	-0.57 [-1.01, -0.12]	
Crea et al. 2014	3.14	1.36	13	3	1.76	15	26.6%	0.09 [-0.66, 0.83]	-+-
Total (95% CI)			53			55	100.0%	-0.39 [-0.78, -0.01]	•
Heterogeneity: $Chi^2 = 2.17$, df = 1 (P = 0.14); $l^2 = 54\%$ Test for overall effect: Z = 2.01 (P = 0.04)								-+	
rescior overall effect.	2 - 2.01	(- 0	.04)						Favors 3-wall defects Favors 2-wall defects

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3C Study or Subgroup	log[Odds Ratio] SE	Weight	Odds Ratio IV, Fixed, 95% Cl		s Ratio d, 95% Cl
Losada et al. 2017	0.107 2.084	7.9%	1.11 [0.02, 66.13]	+	• •
Stavropoulos et al. 2003	1.33 0.612	92.1%	3.78 [1.14, 12.55]		
Total (95% CI)		100.0%	3.43 [1.09, 10.85]		
Heterogeneity: $Chi^2 = 0.32$ Test for overall effect: Z = 2				0.05 0.2 Favors less walls	1 5 20 Favors more walls

3D			Odds Ratio	Odds Ratio
Study or Subgroup	log[Odds Ratio] SE	Weight	IV, Random, 95% C	I IV, Random, 95% CI
Cortellini et al. 2008	0.13 0.08	27.4%	1.14 [0.97, 1.33]	
Losada et al. 2017	0.55 0.199	15.9%	1.73 [1.17, 2.56]	
Stavropoulos et al. 2003	-0.47 0.617	3.0%	0.63 [0.19, 2.09]	
Tonetti et al. 2002	0.372 0.122	23.1%	1.45 [1.14, 1.84]	
Tonetti et al. 2004	0.5 0.04	30.6%	1.65 [1.52, 1.78]	
Total (95% CI)		100.0%	1.42 [1.14, 1.77]	◆
Heterogeneity: Tau ² = 0.04				
Test for overall effect: Z =	0.1 0.2 0.5 1 2 5 10 Favors less walls Favors more walls			

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