

Effects of Fungicide Treatments for the Control of Epidemic and Exotic *Calonectria* Diseases in Italy

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Abstract

Aiello, D., Cirvilleri, G., Polizzi, G., and Vitale, A. 2013. Effects of fungicide treatments for the control of epidemic and exotic *Calonectria* diseases in Italy. *Plant Dis.* 97:37-43.

The efficacy of 11 fungicides was evaluated for the control of *Calonectria* infections on bottlebrush (*Callistemon* "Masotti") and feijoa (*Acca sellowiana*), with special emphasis on *Calonectria pauciramosa* and *C. morganii*, which are the most destructive species in Italian ornamental nurseries. Three nursery experiments were performed with the selected fungicides in order to determine their ability to prevent leaf spot caused by *C. morganii* on bottlebrush and leaf spot and crown and root rot caused by *C. pauciramosa* on bottlebrush and feijoa. All fungicides were effective in reducing disease infections, except for cyproconazole, propamocarb + fosetyl-Al, and K phosphite that were the least effective in reducing *C. morganii* leaf spot. In con-

trast, K phosphite proved more effective in reducing crown and root rot caused by *C. pauciramosa*. Fungicides were also evaluated in growth-chamber experiments for their ability to reduce incidence and severity of leaf spot on bottlebrush caused by the exotic pathogens *C. pseudomexicana*, *C. tunisiana*, *C. polizzii*, and *C. mexicana*. Copper hydroxide, fosetyl-Al, prochloraz, prochloraz + cyproconazole, and tebuconazole were always effective in reducing *Calonectria* leaf spot on bottlebrush. However, some differences in levels of control might be attributable to *Calonectria* isolate. Overall, this study clearly indicates that new fungicides can be employed for chemical management of *Calonectria* infections in ornamental nurseries.

Calonectria spp. have been associated with a wide range of disease symptoms on a large number of plant hosts worldwide (9). On horticultural crops, *Calonectria* spp. have been reported mostly from the Northern Hemisphere, especially in gardens and ornamental nurseries (21). In Italy, mainly two species, *Calonectria pauciramosa* C.L. Schoch & Crous and *Calonectria morganii* Crous, Alfenas & M.J. Wingf., are widespread in nurseries and cause extensive damage on various ornamental plants. A wide range of disease symptoms has been recorded, including crown and root rot, leaf spots, stem canker, and cutting rot on several species, most commonly belonging to plants in the families Anacardiaceae, Arecaceae, Ericaceae, Fabaceae, Myrtaceae, Polygalaceae, Rhamnaceae, and Sapindaceae (30–33,37–39,43,47). Recent studies have indicated that *C. pauciramosa* included cryptic species belonging to the *C. scoparia* complex, and multigene phylogeny analysis recognized *C. polizzii* sp. nov. L. Lombard, Crous & M.J. Wingf. in Italy (22). During a more recent survey in Tunisia, additional species such as *C. pseudomexicana* sp. nov. L. Lombard, G. Polizzi & Crous; *C. tunisiana* sp. nov. L. Lombard, G. Polizzi & Crous; and *C. mexicana* C.L. Schoch & Crous, were observed to cause disease on young plants of *Callistemon* spp. and other ornamental species (23).

Chemical control of *Calonectria* diseases is necessary for reducing damage to young plants in the nursery. Various fungicides and different methods of application have been proposed to control diseases caused by *Calonectria* spp. (9,16). Only preventative measures were found effective, while no curative effects could be obtained for controlling these diseases (9). The use of fungicides (i.e., benomyl, carbendazim, chlorothalonil, copper compounds, prochloraz, and thiophanate-methyl) is a common method for control of these infections in nurseries (5–7,11–13,19,25,26). However, benzimidazole-resistant strains were reported in Italy and

Brazil (1,35,44). In different trials, regular applications of copper compounds provided good control of *Calonectria* infections (6,9,30) whereas, in other trials, repeated applications were shown to induce phytotoxicity to some seedling species (6,46). In addition, some fungicides such as prochloraz or chlorothalonil show variable control of natural infections occurring in nurseries (9,30,36,46). In Sicily (southern Italy), several fungicide applications are made in the nursery during all young growing stages (i.e., seeding, rooting, and pot transplanting) to control crown and root infections, and a fungicide treatment schedule for controlling leaf infections is necessary from August to December. Despite the application of various chemical treatments, epidemics of *Calonectria* disease frequently occur in nurseries.

Thus, the aim of this work was the evaluation of new fungicides that might be proposed to control *Calonectria* diseases in the nursery. This study evaluated fungicides using two approaches: (i) nursery experiments to assess the effectiveness of selected fungicides in controlling *C. pauciramosa* and *C. morganii* infections on bottlebrush and feijoa plants and (ii) growth chamber experiments to assess the efficacy of fungicides against *C. polizzii*, *C. pseudomexicana*, *C. tunisiana*, and *C. mexicana* on bottlebrush.

Materials and Methods

Fungal isolates. Six pathogenic isolates belonging to the genus *Calonectria*—*C. morganii* (CBS 120930 from *Callistemon* hybrid 'Rose Opal', Catania, Italy), *C. pauciramosa* (CBS 130333 from *Callistemon citrinus* (Curtis) Skeels, Catania, Italy), *Calonectria polizzii* (CBS 130351 from *Myrtus communis*, Tunis, Tunisia), *C. pseudomexicana* (CBS 130354 from *Callistemon* sp., Tunis, Tunisia), *Calonectria tunisiana* (CBS 130357 from *Callistemon laevis* An., Tunis, Tunisia), and *Calonectria mexicana* (CBS 130353 from *Dodonaea viscosa*, Tunis, Tunisia)—were used in this study (9,22,23,32). In previous studies, *C. pauciramosa* CBS 130333 was reported as resistant to benzimidazoles, with minimum inhibitory concentration (MIC) between 10 and 100 $\mu\text{g ml}^{-1}$ (32,35), while *C. morganii* CBS 120930 was found sensitive to benzimidazoles (37,44). In more recent laboratory assays, the employed isolates of *C. pseudomexicana*, *C. tunisiana*, and *C. mexicana* had an MIC to benzimidazoles between 1 and 10 $\mu\text{g ml}^{-1}$ whereas an MIC > 100 $\mu\text{g ml}^{-1}$ was found for *C. polizzii* (G.

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Accepted for publication 16 July 2012.

Polizzi, data not published). Fresh cultures of each isolate were obtained by transferring agar plugs from stock cultures onto synthetic low-nutrient agar plates (24). Inoculum was prepared by culturing the fungi on potato dextrose agar (PDA) dishes incubated at 25°C for 10 to 14 days. Conidial suspensions were prepared by flooding the dishes with sterile distilled water, gently rubbing the colony surface with a sterile loop, and filtering the suspension through a triple layer of cheesecloth into a flask to collect conidia. Final conidial concentrations were determined using a hemocytometer and adjusted to a final concentration of 1 to 2.5×10^5 conidia ml⁻¹.

Fungicides. Commercial formulations of 11 fungicides representing eight chemical groups (Table 1) were evaluated for their efficacy at the standard use rates in reducing *Calonectria* leaf spot and crown and root rot on bottlebrush (*Callistemon* "Masotti") and feijoa (*Acca sellowiana* Berg.).

Nursery experiments. The fungicides employed in these experiments were selected based on preliminary data obtained according to a reliable red clover method for screening disease control measures, used in a previous study (45). Eleven fungicides were tested in three nursery experiments for ability to control artificial infections caused by *C. morganii* and *C. pauciramosa* on bottlebrush cuttings and feijoa seedlings, respectively. *C. morganii* was used only for leaf inoculation on bottlebrush, while *C. pauciramosa* was also inoculated on crown and root area of feijoa seedlings. All experiments were conducted on 4-by-4-cm-diameter potted plants grown in a commercial nursery located in Catania province (Sicily, Italy).

In experiments I and II, the fungicide treatments were evaluated for the control of bottlebrush leaf spot caused by *C. morganii* and *C. pauciramosa*. In experiment III, the efficacy of the fungicides was assessed against crown and root rot on feijoa caused by *C. pauciramosa*. Each treatment was replicated three times with 210 (for *C. morganii*) and 70 (for *C. pauciramosa*) bottlebrush cuttings/replicate and 32 feijoa seedlings/replicate. The same number of untreated and inoculated control plots was also included in each experiment. According to common nursery practices, irrigation was provided every 1 to 2 days to young plants for the entire duration of the study with overhead sprinklers placed at 1.5 m in height.

Fungicide suspensions were prepared according to manufacturers' recommended concentrations, as indicated in Table 1. Fungicide treatments were performed by spraying to run-off using a hand sprayer or by applying approximately 0.5 ml/pot as a drench to the crown of feijoa seedlings a day prior to *Calonectria* inoculations. Plant inoculations were made by using $1\text{--}2.5 \times 10^5$ conidia ml⁻¹ suspensions. Bottlebrush cuttings were sprayed to leaf wetness with a hand sprayer, while a soil drench with conidial suspension (about 6.5 ml/pot) was applied onto the crown area of feijoa seedlings. No surfactants or wetting agents were added to the

fungicide and conidial suspensions. After inoculation, bottlebrush cuttings were maintained for 72 h under plastic bags. All fungicide treatments consisted of a single application on bottlebrush cuttings whereas they were repeated twice at a 15-day interval on feijoa seedlings.

Fungicides efficacy was evaluated by calculating disease incidence (DI) and symptoms severity (SS) 10 to 20 and 80 to 120 days after *Calonectria* inoculations on bottlebrush cuttings and feijoa seedlings, respectively, once symptoms were observed in untreated plots. DI was calculated as a percentage resulting from number of plants showing symptoms out of the total number of plants examined $\times 100$. The SS value, calculated based on a percentage of infected foliar surface caused by *C. morganii* or *C. pauciramosa* on bottlebrush cuttings, was collected on a five-point scale, as follows: 1 = healthy seedling and 2 = 1 to 5%, 3 = 6 to 25%, 4 = 26 to 50%, 5 = more than 50% of infected surface up to blighting of entire leaf. A single value was obtained using the mean of five leaves per plant. The SS of crown and root rot caused by *C. pauciramosa* was determined on all plants after their extirpation using a 1-to-5 scale on the basis of percent area with crown and root rot, where 1 = no infection and 2 = 1 to 16%, 3 = 17 to 33%, 4 = 34 to 50%, and 5 = more than 50% of infected crown and root area. SS values were converted to mean disease rating (MDR) that was calculated as Σ (the number of disease plants or leaves in this class \times the disease class)/number of plants or leaves scored. These experiments were conducted twice.

Growth chamber experiments. Four experiments were carried out in growth chambers to evaluate the effects of eight fungicides (Table 1) in reducing bottlebrush leaf spot caused by *C. polizzii* (occasionally reported in Italy) and *C. pseudomexicana*, *C. tunisiana*, and *C. mexicana* (exotic fungi; not reported in Italy). Each treatment was replicated three times with 70 bottlebrush cuttings/replicate. Pathogen inoculations and disease rating were done as in the *C. pauciramosa* and *C. morganii* nursery experiments. Fungicides were applied 3 h before pathogen inoculations by spraying bottlebrush cuttings to run-off. Bottlebrush cuttings were covered with plastic bags and incubated in a growth cabinet (Hitec; Mectec S.A.S.) at 25°C under near-UV light with a 16-h light-and-dark regimen. Evaluation of fungicide effects was determined 6 to 8 days after inoculations. Each experiment was performed twice.

Statistical analyses. Data from the seven experiments were analyzed separately by using the Statistica package software (version 7; Statsoft Inc.). In all repeated experiments, the arithmetic means of DI and MDR were calculated, averaging the values determined for the single replicates of each treatment. Percentage data concerning DI were transformed into the arcsine (\sin^{-1} square root x) prior to analysis of variance (ANOVA), whereas SS values were not transformed. Initial analyses of DI were conducted by calculating *F* and *P* values associated for all experiments to evaluate

Table 1. Features and rates of fungicides used for nursery and growth chamber experiments

Active ingredient	Group name (FRAC code) ^x	Rates (g or ml per 100 liters)	Trade name	Manufacturer	Formulation (%) ^y	Fungicide use ^z
Thiophanate-methyl	MBC (1)	100	Faro	Gowan Italia	38.3 SC	All
Cu hydroxide	Inorganic (11)	250	Ekoram 2000	Du Pont De Nemours	35 WG	All
Fosetyl-AI	Phosphonate (33)	300	Aliette	Bayer CropScience	80 WG	All
Cyproconazole	DMI (3)	30	Caddy	Bayer CropScience	10 WG	I and III
Prochloraz + cyproconazole	DMI (3, 3)	300	Tiptor Xcell	Syngenta Crop Protection	16.15 + 2.15 EC	All
Prochloraz	DMI (3)	100	Octave	Basf Italia	46.1 WP	All
Tebuconazole	DMI (3)	60	Horizon	Bayer CropScience	25.8 EW	I, II, IV–VII
Azoxystrobin	QoI (11)	100	Quadris	Syngenta Crop Protection	22.9 SC	All
Trifloxystrobin	QoI (11)	25	Flint	Bayer CropScience	50 WG	All
Propamocarb + fosetyl-AI	Carbamate (28), phosphonate (33)	300	PrevicurEnergy	Bayer CropScience	47.2 + 27.6 SL	I and III
K phosphite (P ₂ O ₅ + K ₂ O)	Fertilizer (NC)	250	Kaliphos	Agriphos	30 + 20 DC	I and III

^x MBC = methyl benzimidazole carbamates; DMI = demethylation inhibitor; QoI = quinone outside inhibitors. FRAC code number is reported in parentheses; NC = not classified (<http://www.frac.info/frac/publication/anhang/FRAC-Code-List2011-final.pdf>).

^y Percentage of active ingredient. WP = wettable powder, WG = water-dispersible granule, EC = emulsifiable concentrate, DC = dispersible concentrate, SC = suspension concentrate, EW = emulsion oil in water, and SL = soluble concentrate.

^z Combinations of experiments I through VII or "All". All fungicides were preliminarily tested in growth chamber assays on red clover.

whether there is significant trial-treatment interaction. In the post-hoc analyses, the corresponding mean values of DI were subsequently separated by the Newman-Keuls (NK) test ($P = 0.01$). Untransformed arithmetic means of DI are presented in the tables.

Because ordinal scales were adopted for MDR calculation, different nonparametric approaches were used for SS data. Kendall's coefficient of concordance (W) was calculated to assess whether the rankings of the SS data are similar within each trial. In experiment I, where $W > 0.9$, the SS values were at first analyzed by using Friedman's nonparametric rank test, whereas, in the remaining experiments ($W < 0.75$), the Kruskal-Wallis nonparametric one-way test was applied for individual trials, calculating χ^2 and P value associated to both tests. Following Friedman and Kruskal Wallis tests, nonparametric analyses of SS scores in all possible pairwise comparisons were performed with the Wilcoxon signed-rank and Mann-Whitney (MW) tests at $P < 0.05$, respectively.

Results

Fungicide efficacy in controlling *C. morganii* and *C. pauciramosa* infections in the nursery. In all nursery experiments where *C. morganii* and *C. pauciramosa* were sprayed on young bottlebrush cuttings and feijoa plants, there was always a significant effect of fungicides on *Calonectria* infections ($P < 0.001$). A significant treatment-trial effect was observed for all experiments with the exception of experiment I ($P = 0.15$), which was combined. Kendall's coefficient of concordance was 0.91 for SS data in presence of *C. morganii* infections, thus indicating very high concordance between two trials (Table 2). Therefore, the two trials were combined.

All fungicides were effective in controlling *Calonectria* leaf symptoms according to NK at $P = 0.01$ and nonparametric tests at $P < 0.05$ (Tables 3 and 4). In experiment I, tebuconazole and thiophanate-methyl were the most effective treatments in controlling DI and MDR of *C. morganii* leaf spot, while cyproconazole, propamocarb + fosetyl-AI, and K phosphite (fungicides tested only for this pathogen) provided the lowest reductions of DI and MDR compared with the control. A good efficacy in controlling *C. morganii* infections was also observed for fosetyl-AI, prochloraz + cyproconazole, and Cu hydroxide, followed by strobilurins and prochloraz (Table 3). In experiment II, all fungicides significantly reduced DI and MDR of leaf infections caused by *C. pauciramosa* compared with the controls. In the second trial, DI for tebuconazole was significantly lower than azoxystrobin, thiophanate-methyl, and prochloraz and MDR significantly lower than azoxystrobin (Table 4).

In experiment III, conducted on feijoa seedlings, all fungicides significantly ($P = 0.01$) reduced crown and root rot caused by *C.*

pauciramosa compared with the controls (Table 5). K phosphite, fosetyl-AI, and prochloraz + cyproconazole, followed by thiophanate-methyl, Cu hydroxide, trifloxystrobin, cyproconazole, and azoxystrobin, provided a good control of crown and root rot. Otherwise, propamocarb + fosetyl-AI was the least effective treatment in controlling crown and root infections while prochloraz provided variable efficacy against disease in two single trials (Table 5).

Effects of fungicides against other *Calonectria* spp. in growth chamber experiments. In the four experiments conducted on *Calolistemon* "Masotti", a significant effect of treatments ($P \leq 0.001$) and treatment-trial interactions ($P \leq 0.008$) was observed for DI (Table 2). In addition, the Kendall's coefficient for SS showed low concordance ($W < 0.75$) for repeated trials within each experiment (Table 2). Therefore, individual trials are presented.

When fungicide effects were tested against *C. polizzii* in experiment IV, the DI and MDR values in the control plants ranged from 35.2 to 100% and 1.3 to 4.2, respectively. Fosetyl-AI, followed by prochloraz + cyproconazole, consistently reduced DI and MDR of *Calonectria* infections the most compared with the controls, while thiophanate-methyl was ineffective in the first trial and the least effective in the second trial. In the first trial, trifloxystrobin was also ineffective in reducing DI and MDR of *Calonectria* infections,

Table 3. Effects of fungicides on disease incidence (DI) and mean disease rating (MDR) of leaf spot caused by *Calonectria morganii* on bottlebrush under nursery conditions

Treatment	DI (%) ^x	MDR ^{x,y}
Thiophanate-methyl	2.2 a	1.0 a
Tebuconazole	4.6 a	1.0 a
Cu hydroxide	10.8 abc	1.1 bc
Fosetyl-AI	10.0 ab	1.1 c
Prochloraz + cyproconazole	10.7 abc	1.1 cd
Trifloxystrobin	20.4 bc	1.3 e
Azoxystrobin	20.5 bc	1.3 e
Prochloraz	28.0 c	1.3 de
Propamocarb + fosetyl-AI	60.4 d	2.0 f
K phosphite (P ₂ O ₅ + K ₂ O)	60.3 d	2.0 f
Cyproconazole	63.6 d	2.2 f
Control ^z	93.3 e	3.8 g

^x Pooled results of two trials. Data are the mean of three replicates of 210 young bottlebrush plants. Values followed by the same letters within a column are not significantly different according to the Newman-Keuls test ($P = 0.01$) for DI. Arcsine square root transformation was applied on percentage prior to data analysis.

^y Differences among MDR (1-to-5 scale) data for each treatment were analyzed with Friedman two-way analysis of variance by mean rank scores ($P < 0.001$) followed by all pairwise multiple comparison with Wilcoxon signed-rank test ($P \leq 0.05$).

^z Control = untreated, inoculated seedlings.

Table 2. Effects of treatments and treatment-trial interactions on disease incidence (%) and severity (mean disease rating) of leaf spot and crown and root rot caused by *Calonectria* spp. on inoculated bottlebrush and feijoa plants

Experiment	Model effect	Disease incidence ^y			Disease severity ^z	
		df	F	P value	χ^2	W
1	Treatment	11	65.31	<0.001	>2,500	...
1	Treatment-trial	11	1.53 ns	0.15	59.74	0.91
2	Treatment	8	30.19	<0.001	>300	...
2	Treatment-trial	8	2.98	0.011	27.64	0.58
3	Treatment	10	63.08	<0.001	>100	...
3	Treatment-trial	10	4.19	<0.001	44.14	0.74
4	Treatment	8	48.15	<0.001	>700	...
4	Treatment-trial	8	3.18	0.008	28.02	0.58
5	Treatment	8	59.69	<0.001	>150	...
5	Treatment-trial	8	19.71	<0.001	30.80	0.64
6	Treatment	8	147.70	<0.001	>1,000	...
6	Treatment-trial	8	3.19	0.008	22.17	0.46
7	Treatment	8	24.66	<0.001	>300	...
7	Treatment-trial	8	4.52	<0.001	25.07	0.52

^y F test of fixed effects, df = degrees of freedom, and P value associated to F; ns = not significant data.

^z The χ^2 values for Kruskal-Wallis one-way analysis of variance test (treatment) and Friedman two-way analysis of variance (treatment-trial), respectively; W = Kendall's coefficient of concordance between repeated trials in single experiment.

while azoxystrobin and Cu hydroxide were not able to significantly reduce DI values when compared with the control. Prochloraz and tebuconazole always showed a good activity in reducing DI and MDR of leaf spot caused by *C. polizzii* (Table 6). In experiment V, the MDR values on bottlebrush plants ranged from 2.4 to 3.2 while the DI was similar in two single trials, and all fungicides significantly reduced both DI and MDR of *C. pseudomexicana* leaf spot

compared with the control (Table 7). However, few differences were observed among treatments in the first trial, where trifloxystrobin, followed by prochloraz, were the least effective treatments in reducing MDR of leaf spot caused by *C. pseudomexicana* (Table 7).

In experiment VI, DI and SS values of *C. tunisiana* on untreated bottlebrush seedlings were high, ranging from 92.9 to 100% and 2.9 to 4.0, respectively. All fungicides significantly reduced the DI

Table 4. Effects of fungicides on disease incidence (DI) and mean disease rating (MDR) of leaf spot caused by *Calonectria pauciramosa* on bottlebrush under nursery conditions^w

Treatment	First trial		Second trial	
	DI (%) ^x	MDR ^{x,y}	DI (%) ^x	MDR ^{x,y}
Fosetyl-Al	5.2 a	1.1 a	1.4 ab	1.0 ab
Trifloxystrobin	4.8 a	1.1 a	1.9 ab	1.0 ab
Prochloraz	3.8 a	1.1 a	7.6 b	1.1 ab
Cu hydroxide	4.8 a	1.1 a	1.9 ab	1.0 ab
Tebuconazole	10.5 a	1.1 a	0.0 a	1.0 a
Prochloraz + cyproconazole	9.1 a	1.2 a	1.9 ab	1.0 ab
Thiophanate-methyl	13.8 a	1.1 a	9.0 b	1.1 ab
Azoxystrobin	11.9 a	1.1 a	11.4 b	1.1 b
Control ^z	56.2 b	1.8 b	38.6 c	1.7 c

^w The χ^2 and *P* values indicate the significance of the Kruskal-Wallis test. First trial, $\chi^2 = 394.1$ and *P* < 0.0001; second trial, $\chi^2 = 325.2$ and *P* < 0.0001.

^x Data are the mean of three replicates of 70 young bottlebrush plants. Values followed by the same letters within a column are not significantly different according to the Newman-Keuls test (*P* = 0.01) for DI. Arcsine square root transformation was applied on percentage prior to data analysis.

^y Differences among MDR (1-to-5 scale) data for each treatment were analyzed with Kruskal-Wallis one-way analysis of variance by mean rank scores followed by all pairwise multiple comparison with Mann-Whitney test (*P* ≤ 0.05).

^z Control = untreated, inoculated seedlings.

Table 5. Effects of fungicides on disease incidence (DI) and mean disease rating (MDR) of crown and root rot caused by *Calonectria pauciramosa* on feijoa under nursery conditions^w

Treatment	First trial		Second trial	
	DI (%) ^x	MDR ^{x,y}	DI (%) ^x	MDR ^{x,y}
K phosphite (P ₂ O ₅ + K ₂ O)	9.4 a	1.1 a	10.4 a	1.1 a
Fosetyl-Al	12.5 a	1.2 a	8.4 a	1.1 a
Prochloraz + cyproconazole	14.6 a	1.3 a	9.4 a	1.2 a
Thiophanate-methyl	15.6 a	1.3 a	11.5 a	1.2 ab
Cu hydroxide	16.7 a	1.3 a	14.6 a	1.3 ab
Trifloxystrobin	17.7 a	1.3 a	13.5 a	1.3 ab
Cyproconazole	16.6 a	1.3 a	13.5 a	1.3 ab
Azoxystrobin	17.7 a	1.4 ab	16.6 a	1.3 ab
Prochloraz	37.5 b	1.8 c	16.7 a	1.3 ab
Propamocarb + fosetyl Al	34.4 b	1.6 bc	27.1 b	1.6 b
Control ^z	77.1 c	2.5 d	51.0 c	2.1 c

^w The χ^2 and *P* values indicate the significance of the Kruskal-Wallis test. First trial, $\chi^2 = 196.9$ and *P* < 0.0001; second trial, $\chi^2 = 101.2$ and *P* < 0.0001.

^x Data are the mean of three replicates of 32 young feijoa plants. Values followed by the same letters within a column are not significantly different according to the Newman-Keuls test (*P* = 0.01) for DI. Arcsine square root transformation was applied on percentage prior to data analysis.

^y Differences among MDR (1-to-5 scale) data for each treatment were analyzed with Kruskal-Wallis one-way analysis of variance by mean rank scores followed by all pairwise multiple comparison with Mann-Whitney test (*P* ≤ 0.05).

^z Control = untreated, inoculated seedlings.

Table 6. Effects of fungicides on disease incidence (DI) and mean disease rating (MDR) of leaf spot caused by *Calonectria polizzii* on bottlebrush under controlled environment conditions^w

Treatment	First trial		Second trial	
	DI (%) ^x	MDR ^{x,y}	DI (%) ^x	MDR ^{x,y}
Fosetyl-Al	4.8 a	1.1 a	1.0 a	1.0 a
Prochloraz + cyproconazole	6.7 a	1.1 a	2.4 ab	1.0 a
Prochloraz	4.3 a	1.1 a	5.7 bc	1.1 ab
Tebuconazole	7.1 a	1.2 a	15.2 c	1.3 b
Cu hydroxide	9.5 ab	1.1 a	9.5 bc	1.1 ab
Azoxystrobin	12.8 ab	1.3 ab	5.7 bc	1.1 ab
Trifloxystrobin	22.4 ab	1.3 bc	7.6 bc	1.1 ab
Thiophanate-methyl	24.8 ab	1.3 c	57.6 d	2.6 c
Control ^z	35.2 b	1.4 c	100 e	4.2 d

^w The χ^2 and *P* values indicate the significance of the Kruskal-Wallis test. First trial, $\chi^2 = 162.0$ and *P* < 0.0001; second trial, $\chi^2 = 1,089.0$ and *P* < 0.0001.

^x Data are the mean of three replicates of 70 young bottlebrush plants. Values followed by the same letters within a column are not significantly different according to the Newman-Keuls test (*P* = 0.01) for DI. Arcsine square root transformation was applied on percentage prior to data analysis.

^y Differences among MDR (1-to-5 scale) data for each treatment were analyzed with Kruskal-Wallis one-way analysis of variance by mean rank scores followed by all pairwise multiple comparison with Mann-Whitney test (*P* ≤ 0.05).

^z Control = untreated, inoculated seedlings.

and MDR values of leaf spot compared with the controls, and thiophanate-methyl and prochloraz + cyproconazole were more effective than Cu hydroxide in reducing both DI and MDR of leaf spot caused by this pathogen in the first trial (Table 8).

In experiment VII, a moderate infection level caused by *C. mexicana* was observed in both trials (DI ranged from 46.2% to 57.1% and SS from 1.5 to 2.0 on control bottlebrush plants). However, all fungicides were effective in reducing DI and MDR of *Calonectria*

leaf spot except for DI of azoxystrobin and prochloraz in the first trial. In the second trial, prochloraz + cyproconazole and fosetyl-AI were more effective than thiophanate-methyl in reducing both DI and MDR of leaf spot on bottlebrush caused by *C. mexicana* (Table 9).

Discussion

This research adds valuable information contributing to a valid strategy for using new and already known fungicides for managing

Table 7. Effects of fungicides on disease incidence (DI) and mean disease rating (MDR) of leaf spot caused by *Calonectria pseudomexicana* on bottlebrush under controlled environment conditions^w

Treatment	First trial		Second trial	
	DI (%) ^x	MDR ^{x,y}	DI (%) ^x	MDR ^{x,y}
Fosetyl-AI	1.4 a	1.0 a	1.9 a	1.0 a
Prochloraz + cyproconazole	3.3 a	1.0 a	1.0 a	1.0 a
Thiophanate-methyl	3.3 a	1.1 a	1.0 a	1.0 a
Cu hydroxide	4.3 a	1.0 a	3.4 a	1.0 a
Tebuconazole	4.3 a	1.0 a	2.4 a	1.0 a
Azoxystrobin	8.6 a	1.2 ab	4.8 a	1.0 a
Prochloraz	20.0 a	1.2 bc	1.0 a	1.0 a
Trifloxystrobin	21.4 a	1.3 c	3.4 a	1.0 a
Control ^z	80.4 b	2.4 d	77.2 b	3.2 b

^wThe χ^2 and *P* values indicate the significance of the Kruskal-Wallis test. First trial, $\chi^2 = 792.4$ and *P* < 0.0001; second trial, $\chi^2 = 1,102.6$ and *P* < 0.0001.

^xData are the mean of three replicates of 70 young bottlebrush plants. Values followed by the same letters within a column are not significantly different according to the Newman-Keuls test (*P* = 0.01) for DI. Arcsine square root transformation was applied on percentage prior to data analysis.

^yDifferences among MDR (1-to-5 scale) data for each treatment were analyzed with Kruskal-Wallis one-way analysis of variance by mean rank scores followed by all pairwise multiple comparison with Mann-Whitney test (*P* ≤ 0.05).

^zControl = untreated, inoculated seedlings.

Table 8. Effects of fungicides on disease incidence (DI) and mean disease rating (MDR) of leaf spot caused by *Calonectria tunisiana* on bottlebrush under controlled environment conditions^w

Treatment	First trial		Second trial	
	DI (%) ^x	MDR ^{x,y}	DI (%) ^x	MDR ^{x,y}
Thiophanate-methyl	0.0 a	1.0 a	5.7 a	1.1 a
Prochloraz + cyproconazole	0.0 a	1.0 a	6.7 a	1.1 a
Trifloxystrobin	4.3 ab	1.1 ab	3.8 a	1.0 a
Prochloraz	3.3 ab	1.0 ab	5.7 a	1.1 a
Tebuconazole	3.3 ab	1.0 ab	6.7 a	1.1 a
Fosetyl-AI	3.8 ab	1.0 ab	5.7 a	1.1 a
Cu hydroxide	1.4 ab	1.0 ab	11.9 a	1.1 a
Azoxystrobin	11.9 b	1.1 b	6.7 a	1.1 a
Control ^z	92.9 c	2.9 c	100 b	4.0 b

^wThe χ^2 and *P* values indicate the significance of the Kruskal-Wallis test. First trial, $\chi^2 = 1,299.0$; *P* < 0.0001; second trial, $\chi^2 = 1,160.5$ and *P* < 0.0001.

^xData are the mean of three replicates of 70 young bottlebrush plants. Values followed by the same letters within a column are not significantly different according to the Newman-Keuls test (*P* = 0.01) for DI. Arcsine square root transformation was applied on percentage prior to data analysis.

^yDifferences among MDR (1-to-5 scale) data for each treatment were analyzed with Kruskal-Wallis one-way analysis of variance by mean rank scores followed by all pairwise multiple comparison with Mann-Whitney test (*P* ≤ 0.05).

^zControl = untreated, inoculated seedlings.

Table 9. Effects of fungicides on disease incidence (DI) and mean disease rating (MDR) of leaf spot caused by *Calonectria mexicana* on bottlebrush under controlled environment conditions^w

Treatment	First trial		Second trial	
	DI (%) ^x	MDR ^{x,y}	DI (%) ^x	MDR ^{x,y}
Prochloraz + cyproconazole	1.4 a	1.0 a	0.0 a	1.0 a
Fosetyl-AI	4.3 a	1.0 a	0.0 a	1.0 a
Trifloxystrobin	7.6 a	1.1 a	2.9 ab	1.0 ab
Cu hydroxide	6.7 a	1.1 a	4.8 ab	1.0 ab
Tebuconazole	3.3 a	1.0 a	5.7 ab	1.1 ab
Thiophanate-methyl	1.0 a	1.0 a	13.3 b	1.3 b
Azoxystrobin	19.5 ab	1.2 b	4.8 ab	1.0 ab
Prochloraz	22.9 ab	1.2 b	3.8 ab	1.0 ab
Control ^z	46.2 b	1.5 c	57.1 c	2.0 c

^wThe χ^2 and *P* values indicate the significance of the Kruskal-Wallis test. First trial, $\chi^2 = 336.4$; *P* < 0.0001; second trial, $\chi^2 = 591.7$; *P* < 0.0001.

^xData are the mean of three replicates of 70 young bottlebrush plants. Values followed by the same letters within a column are not significantly different according to the Newman-Keuls test (*P* = 0.01) for DI. Arcsine square root transformation was applied on percentage prior to data analysis.

^yDifferences among MDR (1-to-5 scale) data for each treatment were analyzed with Kruskal-Wallis one-way analysis of variance by mean rank scores followed by all pairwise multiple comparison with Mann-Whitney test (*P* ≤ 0.05).

^zControl = untreated, inoculated seedlings.

several *Calonectria* diseases, especially for those caused by *C. morganii* and *C. pauciramosa* that represent the majority of disease records reported from ornamental nurseries around the world (2,8,9,15,17,18,20,24,27,28,32,39). Single experiments have previously shown the potential of certain fungicides to reduce incidence and severity of infections caused by *C. morganii* and *C. pauciramosa*, well established in Italian nurseries (30,32,39,47), by other more sporadically occurring pathogens, such as *C. polizzii* (22), and by those exotic in Italian nurseries (*C. pseudomexicana*, *C. tunisiana*, and *C. Mexicana*) recently reported in Tunisia (23).

From these results, preventive applications of tebuconazole, fosetyl-Al, Cu hydroxide, thiophanate-methyl, prochloraz + cyproconazole, trifloxystrobin, azoxystrobin, and prochloraz could be suggested to control leaf spots caused by both *C. morganii* and *C. pauciramosa* in nurseries, whereas use of cyproconazole, propamocarb + fosetyl Al, and K phosphite is discouraged. Because they provided a significant reduction of crown and root rot caused by *C. pauciramosa* on feijoa plants, K phosphite, fosetyl-Al, prochloraz + cyproconazole, cyproconazole, Cu hydroxide, thiophanate-methyl, trifloxystrobin, and azoxystrobin should be included in fungicide application schedules. Overall, K phosphite did not show good efficacy against *Calonectria* leaf spot, although it performed well against crown and root rot caused by *C. pauciramosa*.

Among the tested fungicides, fosetyl-Al, prochloraz + cyproconazole, and tebuconazole showed the best activity also in controlling leaf spot on bottlebrush caused by *C. polizzii*, *C. pseudomexicana*, *C. tunisiana*, and *C. mexicana*.

Although thiophanate-methyl showed good efficacy in controlling *Calonectria* infections, it belongs to the methyl benzimidazole carbamates (MBCs) that are considered to be at high risk for the development of resistance in target populations (14). Indeed, the observed partial failure of thiophanate-methyl in reducing leaf spot caused by *C. polizzii* is due to the high resistance level (MIC > 100 µg ml⁻¹) to MBCs of the tested isolate. Because a high prevalence of MBC-resistant *Calonectria* isolates has been reported (1,35,44), the use of these fungicides should be seriously questioned and the exclusive use of MBCs for *Calonectria* spp. management avoided.

Tebuconazole reduced leaf spot caused by either *C. morganii* or *C. pauciramosa*. Unfortunately, the repeated use of tebuconazole in a short period is known to cause stunting and reduction of length and shoot number or root hypertrophy on bottlebrush and milkwort (*Polygala myrtifolia* L.; 29,46). The present study showed that copper hydroxide application could be recommended for reducing *Calonectria* crown and root rot as well as leaf spot, being careful to avoid repeated treatments that can cause phytotoxicity (46; G. Polizzi, unpublished data) and Cu accumulation in the soil. Indeed, copper compounds should be included as alternative fungicides in anti-resistance strategies to manage *Calonectria* spp. in the production of many susceptible ornamental species. Because of the variable results detected for prochloraz in this study, further investigations on the efficacy of this compound are needed.

The findings in our study suggest that effective fungicides having a different site of action may be integrated in appropriate fungicide application schedules for controlling *Calonectria* spp. For example, mixtures or alternations among tested inorganic (copper compounds), phosphonate (fosetyl-Al), quinone outside inhibitors (strobilurins), demethylation inhibitors, and MBCs fungicides could represent a sustainable strategy for achieving good control of *Calonectria* infections in nurseries. These rotation or tank mix strategies will also be able to minimize the risk of fungicide resistance, phytotoxicity, and environmental damage due to accumulation of heavy metals in the soil.

In addition, a fungicide-based management program against crown and root rot by *C. pauciramosa* could be supplemented with K phosphite fertilization, which provided very good results in the present study. Many authors have reported on the efficacy of K phosphite against fungal diseases on various crops (3,4,40,41,48). However, the efficacy of K phosphite against *Calonectria* infections was not reported until the present study. Although its mode of action against *Calonectria* spp. was not studied, the plant re-

sponses observed in this study are likely to be attributable to the direct effect of K phosphite against *Calonectria* spp., as was reported for other fungal plant pathogens (10,42). This is supported by the fact that K phosphite has shown good activity against *C. morganii* and *C. pauciramosa* on red clover seedlings in a very short period and reduced mycelial growth of these fungi in vitro on PDA amended with K phosphite (data not shown).

Overall, management of *Calonectria* diseases in the nursery cannot rely on a single control measure and the steps to developing an integrated program for efficient management of these diseases is complex. Because only preventative measures were found effective for *Calonectria* disease control (9), chemical control would always be adopted in association with good nursery practices, including reduction of primary inoculum, removal of infected plants, and utilization of uncontaminated potting medium. Moreover, the use of other sustainable strategies as well, such as soil solarization (34) or biological control agents (45), could improve disease control.

Acknowledgments

This research was supported by MIUR (project number PON01_01611), and PRA, University of Catania. We thank V. Guarnaccia, P. T. Formica, and A. Cinquerrui for technical assistance.

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