

Aroma Profile and Essential Oil Composition of *Helichrysum* speciesRose Vanessa Bandeira Reidel^{1*}, Pier Luigi Cioni¹, Barbara Ruffoni², Claudio Cervelli² and Luisa Pistelli¹¹Dipartimento di Farmacia, Università di Pisa, Via Bonanno 6, 56126 Pisa, Italy²Consiglio per la ricerca in agricoltura e l'analisi dell'economia agraria (CREA)- Unità di ricerca per la floricoltura e le specie ornamentali, Corso degli Inglesi 508, 18038 San Remo, Italy

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Essential oils and volatile emission from leaves and flowers of three *Helichrysum* species (*H. arenarium*, *H. nudifolium*, *H. graveolens*) were analysed by GC-MS and SPME analysis. A total of 112 components were identified in the EOs representing 92.3-99.9% of the whole oil composition, while 109 constituents were identified in the spontaneous volatile emission accounted for 78.7-98.8% of the total volatile organic compounds (VOCs). Sesquiterpenes represented the main class of constituents in the EOs of the majority studied species. β -caryophyllene was the key compound of *H. arenarium* and *H. nudifolium* leaves, even though (*E*)-2-hexenal showed high percentage in *H. arenarium* leaves. *H. graveolens* EO showed a good amount of diterpenes (cembrene and beyerene), absent in the other species. SPME analyses were carried out for the first time to complete the chemotaxonomic investigation on the volatile organic constituents of these plants.

Keywords: *Helichrysum arenarium*, *H. nudifolium*, *H. graveolens*, Asteraceae, HS-SPME, GC-MS.

Genus *Helichrysum* Mill. (*Asteraceae*) consists of approximately 500 species distributed worldwide, of which 250 are indigenous to South Africa (including Namibia), while only 16 are spontaneous in Europe and in the Mediterranean areas, including Italy [1-5].

Three *Helichrysum* species (*H. arenarium*, *H. nudifolium*, *H. graveolens*) native from different geographical areas but acclimatised and grown in Italy in homogeneous conditions (CREA-Sanremo collection) were selected in this work to evaluate their aromatic profile and contribute to their ornamental or industrial applications. *Helichrysum arenarium* (L.) Moench, is a perennial herb, 50 cm height, that grows in dry sandy and semi-desert soils, dunes, steppes of a large geographic area including Scandinavia, Atlantic Europe, Balkans, European Russia, Siberia, Caucasus, Asia Minor, Central Asia, Mongolia and China. This species has greyish leaves, with stems that are covered with white trichomes and the colour of flowers varies from yellow to orange. *H. arenarium* is used to treat stomach pain, asthma, arthritis disorders, cystitis and jaundice and its inflorescences are traditionally used in Central Europe as antiseptic and spasmolytic remedies [6]. It is also used as repellent against brown house moths [7]. Many studies confirmed several biological properties of the species *H. arenarium* including antioxidant, hepatoprotective, antibacterial, antiviral, antifungal, anti-inflammatory and antiproliferative activities [8-11] together with psychotropic effects [12]. Essential oil from *H. arenarium* showed antioxidant and antimicrobial properties [13].

Helichrysum nudifolium (L.) Less is found in grasslands throughout Southern Africa, with solitary flowering stems which can be tall till 1.5 m. The inflorescence is flat-topped contracted head-like. This species has radical leaves nerved, linear or lanceolate, elliptic to ovate, thinly and thickly white-woolly [14, 15, 16]. The leaves of *H. nudifolium* are externally used for wounds and respiratory infections; moreover, the poultice of this plant is recommended for swellings and the roots are employed as antiemetic and antiseptic remedies [2b]. No papers on biological activity of *H. nudifolium* extracts are reported in the literature to confirm its folk uses.

H. graveolens (M. Bieb.) Sweet grows in high pastures, subalpine and alpine meadows of Crimea, Caucasus, Balkan Peninsula, Asia Minor, Armenia and Kurdistan. It is a greyish-green plant with yellow flowers and linear or linear lanceolate leaves, acute, often spinulose [16]. Flowers from *H. graveolens* are used in the folk medicine of Turkey for treatment of diabetes mellitus, wound, jaundice and as diuretic [17, 18, 19]. Antidiabetic and antioxidant effects together with its use to treat urolithiasis have been reported in the literature for *H. graveolens* flowers [19, 20-21].

The chemical composition of the EOs and the spontaneous emission of volatiles (SPME) obtained from *H. arenarium* (*Ha*), *H. nudifolium* (*Hn*) and *H. graveolens* (*Hg*) flowers (*F*) and leaves (*Le*) were analysed by GC-FID and GC/MS analyses. The results are reported in Table 1 and 2, respectively. A total of 112 components were identified in the EOs representing 92.3-99.9% of the whole oil composition, while 109 compounds were identified by SPME analysis accounted for 78.7-98.8% of the total volatile organic compounds (VOCs).

From the Hierarchical analysis, HCA and the principal component analysis (PCA) three groups of EOs were obtained (Fig 1 and 2, respectively). One of these groups (I) was formed by *Ha* (flowers and leaves) and *Hn* flowers. The composition of EOs from *Ha* leaves and *Hn* flowers was dominated by sesquiterpene hydrocarbons (SH), whereas the *Ha* flower showed a similar amount of SH and non-terpene (NT) compounds. Monoterpenes were completely missing in *Hn* sample. The main constituents in group I were: β -caryophyllene, β -pinene, 1,8-cineole, pentadecanoic acid, 6,10,14-trimethyl-2-pentadecanone and bis-(2-methylpropylester)-1,2-benzendicarboxylic acid. *Hn* leaves are inserted in group II because they were characterized by sesquiterpene hydrocarbons (SH) and non-terpenes (NT) by β -caryophyllene (43.9%) for SH and (*E*)-2-hexenal (41.0%) and (*Z*)-lanceol acetate (11.6%) for NT. Group III (*Hg* leaves and flowers) showed mainly sesquiterpene hydrocarbons (α -neo-clovene, *cis*- β -guaiane, *trans*- β -guaiane, selina-1(2)4-diene) and oxygenated sesquiterpenes (OS: γ -eudesmol, 7-epi- α -eudesmol and valerianol). The diterpenes (DT)

Table 1: Chemical composition of EOs from the three selected *Helichrysum* species and organs.

Entry	Class	Components	LRI	Content (%)					
				Ha Fl	Ha Le	Hn Fl	Hn Le	Hg Fl	Hg Le
1	NT	(E)-2-Hexenal	854		1.8		41.0		0.6
2	MH	α -Pinene	939		3.5			tr	0.9
3	MH	Tricyclene	926	2.0					
4	MH	Camphene	953	tr	0.3				tr
5	NT	2-(Z)-Heptenal	956					0.1	
6	MH	β -Pinene	980	7.4	9.5				0.3
7	MH	Myrcene	991		0.6				
8	NT	6-Methyl-5-hepten-2-one	985						0.1
9	NT	2-Pentyl-furan	995	0.3				0.4	0.2
10	NT	<i>trans</i> -2-(2-Pentenyl) furan	1004						0.1
11	NT	<i>n</i> -Octanal	1001	0.2	1.0			0.4	
12	MH	α -Terpinene	1018	0.2	0.4				0.1
13	MH	<i>p</i> -Cymene	1026		0.3				
14	MH	Limonene	1031	2.0	5.2			tr	
15	OM	1,8-Cineole	1033	2.4	8.3			tr	1.6
16	MH	γ -Terpinene	1062	0.7	2.7			0.2	0.2
17	MH	Terpinolene	1088	0.5	1.3			tr	tr
18	NT	<i>n</i> -Nonanal	1102	0.5	1.8	1.6		1.7	0.1
19	OM	Exo-fenchol	1121	tr	0.3				tr
20	OM	Pinocampheol	1173					0.3	
21	OM	Borneol	1165	0.3	tr				
22	OM	Terpinen-4-ol	1177	tr	0.4				tr
23	OM	α -Terpineol	1189		1.6			tr	
24	OM	γ -Terpineol	1198	1.3					
25	AC	Safranal	1199						
26	NT	<i>n</i> -Decanal	1204	0.8	0.3	5.3		1.2	
27	NT	Butanoic acid, 3-methyl-3-hexen-1-yl-ester	1199		0.9				
28	NT	Hexyl-3-methylbutanoate	1243		0.9				
29	OM	Geraniol	1255						0.5
30	NT	(E)-2-Decenal	1261					0.2	
31	OM	Perilla aldehyde	1271		0.5				tr
32	NT	1-Dihydroedulan	1295						0.2
33	NT	Undecanal	1306			3.4		0.2	
34	NT	(2E, 4Z)-Decadienal	1314					0.3	
35	SH	α -Ylangene	1372	tr	0.2			0.8	
36	SH	α -Copaene	1376	0.9	1.6			1.4	0.8
37	SH	α -Duprezianene	1388					1.2	
38	NT	Tetradecane	1399	0.2				0.4	
39	NT	Dodecanal	1407			0.9		tr	
40	SH	β -Caryophyllene	1418	27.5	46.0	27.0	43.9	1.9	1.9
41	SH	<i>trans</i> - α -Bergamotene	1439			8.6			
42	SH	β -Gurjunene	1432					tr	0.2
43	SH	α -Guaiene	1439					0.5	
44	SH	α -Neo-clovene	1445					3.2	3.5
45	SH	<i>cis</i> -Muurolo-3,5-diene	1446						1.8
46	SH	(E)- β -Farnesene	1458		3.6				1.2
47	SH	β -Cadinene	1474						2.5
48	SH	<i>trans</i> -Cadin-1(6),4-diene	1470					1.8	
49	SH	α -humulene	1454	1.4	1.3	1.1			
50	SH	9-Epi-(E)-caryophyllene	1467	2.2					
51	SH	<i>cis</i> -Muurolo-4(14), 5-diene	1460					1.6	0.4
52	SH	γ -Muurolole	1477	1.2	0.9				9.2
53	SH	Viridiflorene	1493		0.3				
54	SH	<i>cis</i> - β -Guaiene	1490					6.4	7.3
55	SH	Valencene	1491	0.3				2.1	
56	SH	δ -Amorphene	1505	0.9				2.0	1.4
57	SH	Cadin-1(10),4-diene	1527						2.1
58	OS	10-Epi-cubebol	1534					1.2	
59	SH	Bicyclgermacrene	1494			8.0	tr		
60	SH	<i>trans</i> - β -Guaiene	1500					5.6	1.8
61	NT	Tridecanal	1509			5.3			
62	SH	<i>trans</i> - γ -Cadinene	1513	1.0	0.7				1.8
63	SH	δ -Cadinene	1524	3.2	1.7			2.3	3.8
64	SH	α -Cadinene	1538	0.2					
65	SH	<i>trans</i> -Cadin-1(2),4-diene	1533						0.6
66	SH	Selina-3,7-(11) diene	1542	tr				4.7	0.4
67	SH	α -Calacorene	1542	tr				0.6	
68	SH	β -Calacorene	1563					1.2	
69	OS	Caryophyllene alcohol	1568	0.2					
70	OS	Caryophyllene oxide	1581	1.7	1.3	7.8	tr		
71	OS	Viridiflorol	1590					0.6	0.7
72	OS	Guaial	1595					0.6	1.2
73	NT	Hexadecane	1600	0.1		1.5			
74	OS	5-Epi-7-epi- α -eudesmol	1606		0.2				
75	OS	1,10-Di-epi-cubenol	1614	0.7				0.5	
76	OS	Cadin-4-en-7-ol	1637						1.6
78	OS	α -Acorenol	1630			1.1			
79	OS	10-Epi- γ -eudesmol	1619					0.4	
80	OS	γ -Eudesmol	1630					6.1	10.4
81	OS	1-Epi-cubenol	1627	0.5	0.3				
82	OS	Hinesol	1638						4.7
83	OS	Epi- α -cadinol	1640		tr			2.6	
84	OS	α -Muurolole	1645	1.1					
85	OS	Valerianol	1655					4.0	2.7
86	OS	7-Epi- α -eudesmol	1605	0.4				2.1	7.3
87	OS	α -Cadinol	1652	0.6					
88	OS	Elemol acetate	1680					0.2	
89	NT	1-Heptadecene	1695	0.6					
90	OS	β -Acorenone	1685						0.1
91	NT	Davanol acetate	1610						0.1
92	OS	Juniper camphor	1691						0.2
93	NT	δ -Dodecalactone	1671			7.5			
94	NT	Pentadecanal	1718	1.2				0.8	
95	NT	Tetradecanoic acid	1766	1.7				2.3	

96	OS	Hinesol acetate	1783						0.2
97	OS	Nootkatone	1999					0.7	
98	NT	Octadecane	1800			1.3			
99	NT	Pentadecanoic acid, methyl ester	1820	31.0					
100	NT	6,10,14-Trimethyl-2 pentadecanone	1844			10.5			
101	NT	Bis (2-methylpropylester)-1,2 benzenedicarboxylic acid	1873			6.4			
102	OS	Cedren-13-ol-acetate	1788						2.6
103	NT	<i>n</i> -Nonadecane	1900			0.3			
104	DT	Totarene	1922						1.6
105	NT	(<i>Z</i>)-Lanceol acetate	1860				11.6	0.7	0.3
106	DT	Beyerene	1924					0.7	12.7
107	DT	Cembrene	1929					25.7	0.3
108	OS	6-Acetate- α -chenopodiol	1960					0.6	0.4
109	SH	Bifloratriene	1977						1.0
110	NT	<i>n</i> -Hexadecanoic acid	1978						0.2
111	DT	Neocembrene	1959					1.9	
112	NT	1-Eicosene	2000			1.2	1.1	0.4	
Total identified			96.8	99.7	98.8	99.8	92.3	96.0	
Non-terpene derivatives			36.6	6.7	45.2	53.7	7.1	1.6	
Monoterpene hydrocarbons			12.8	23.8			0.2	1.5	
Oxygenated monoterpenes			4.0	11.1			0.3	2.1	
Sesquiterpene hydrocarbons			38.8	56.3	44.7	43.9	36.1	43.1	
Oxygenated sesquiterpenes			5.2	1.8	8.9		20.3	33.6	
Diterpenes							28.3	14.6	

^{a)} Compounds present at less than 0.1% were excluded from the table. ^{b)} LRI: Linear retention index. Relative contents of the EO constituents expressed as percentages. ^{c)} Species and organs analyzed: Ha Fl: *H. arenarium* flowers; Ha Le: *H. arenarium* leaves; Hn Fl: *H. nudifolium* flowers; Hn Le: *H. nudifolium* leaves; Hg Fl: *H. graveolens* flowers; Hg Le: *H. graveolens* leaves.

Table 2: Spontaneous volatile emission of the three selected *Helichrysum* species (SPME analysis)

Entry	Class	Components	LRI	Content (%) ^{f)}					
				Ha Fl	Ha Le	Hn Fl	Hn Le	Hg Fl	Hg Le
1	NT	Hexanal	810			tr		0.1	
2	NT	2-Octene	815					0.1	
3	NT	(<i>E</i>)-3-Hexen-1-ol	858			0.9			
4	NT	1-Nonene	860	2.5					
5	NT	Hexanol	870	0.4		0.5		0.5	
6	NT	(<i>Z</i>)-3-Hexenol	858		tr		15.0		
7	NT	Isopentyl acetate	876			0.2			
8	NT	3-Nonene	899	0.2				tr	
9	MH	Tricyclene	934	0.2				tr	
10	MH	α -Pinene	940	15.8	4.0	0.2	0.2	0.3	2.2
11	MH	Camphene	953	0.5		tr			
12	NT	<i>n</i> -Heptanol	973	0.5			0.1	tr	
13	MH	β -Pinene	981	55.2	5.8			tr	0.2
14	NT	1-Octen-3-ol	978			0.3			
15	NT	3-Octanone	987				6.6		
16	MH	Myrcene	993	0.5				tr	0.1
17	NT	3-Octanol	994			0.1			
18	MH	<i>p</i> -Mentha-1(7),8-diene	1004	0.4					
19	NT	Octanal	1002			0.1			
20	NT	(<i>E</i>)-3-Hexenol acetate	1007	0.2		0.8		tr	
21	NT	(<i>Z</i>)-3-Hexenol acetate	1008		49.6		70.5		0.1
22	NT	<i>n</i> -Hexyl acetate	1010	tr		0.1			
23	MH	α -Terpinene	1018	0.4				0.2	
24	MH	Limonene	1032	5.2				0.2	tr
25	OM	1,8-Cineole	1036	5.1	7.4	tr		0.9	
26	MH	(<i>Z</i>)- β -Ocimene	1041					0.3	
27	MH	(<i>E</i>)- β -Ocimene	1053	0.7	0.3			0.5	
28	MH	Terpinolene	1089	0.9				0.2	
29	MH	γ -Terpinene	1062	1.4	0.3	0.2			
30	NT	Isopentyl isovalerate	1106				0.3		
31	OM	Linalool	1102		0.6				
32	NT	<i>n</i> -Nonanal	1103	Tr		0.4	0.8	tr	tr
33	OM	Exo-fenchol	1117	0.1					
34	OM	<i>cis-p</i> -Menth-2-en-1-ol	1122	0.2					
35	OM	(<i>E</i>)-Myroxide	1143	0.1				tr	
36	OM	Camphor	1148		1.2	tr			
37	OM	Camphene hydrate	1150	0.1					
38	NT	1,4-Dimethoxy benzene	1163			0.8			
39	OM	4-Terpineol	1180	0.5				tr	
40	OM	Borneol	1169	0.3	0.6				
41	NT	(<i>Z</i>)-3-Hexenyl butyrate	1191		0.4				
42	OM	α -Terpineol	1190	1.6				tr	
43	NT	Decanal	1206	0.1		0.2		tr	
44	OM	Dihydro carveol	1196		0.5				
45	NT	<i>n</i> -Decanal	1206		0.3		0.3		0.1
46	NT	Butanoic acid, (<i>Z</i>)-3-methyl-3-hexenyl ester	1231		11.1				
47	NT	Hexyl-3-methyl butanoate	1242	tr	1.6				
48	OM	Perilla aldehyde	1280	tr	0.2			tr	
49	OM	Isobomyl acetate	1287		3.0	0.1		tr	
50	SH	Presilphiperfol-7-ene	1328					0.1	
51	SH	δ -Elemene	1340			1.8			0.2
52	SH	7-Epi-silphiperfol-5-ene	1345	tr	tr			0.3	
53	PP	Eugenol	1361		0.2				
54	SH	α -Ylangene	1372	tr	0.2			4.4	3.7
55	SH	α -Copaene	1376	0.5	0.8	tr		4.8	
56	SH	Isoledene	1375						1.9
57	SH	β -Patchoulene	1380					1.1	
58	NT	(<i>Z</i>)-3-Hexenyl ester hexanoic acid	1386		0.8				
59	SH	Isolongifolene	1385					0.3	0.2
60	SH	β -Cubebene	1389					0.3	
61	NT	Hexyl <i>n</i> -hexanoate	1388		0.2				

62	SH	Sativene	1395					0.1	
63	SH	β -Elemene	1392	tr	0.2	tr			
64	SH	Cyperene	1398						0.4
65	SH	Isoitalicene	1397					0.3	
66	SH	Isocaryophyllene	1405	tr		1.2			
67	PP	Methyl eugenol	1407		0.2				
68	NT	Dodecanal	1409				0.1		
69	SH	α -Gurjunene	1409	tr				6.1	4.0
70	SH	β -Caryophyllene	1418	1.0	1.0	79.4	2.5	6.0	
71	SH	β -Gurjunene	1432					1.4	9.6
72	SH	β -Copaene	1429	0.1	0.2				
73	SH	γ -Elemene	1434			1.3		tr	1.0
74	SH	Aromadendrene	1439		0.4	0.2			
75	SH	α -Guaiene	1440	tr	0.1	tr		1.2	
76	SH	α -Neoclovene	1454					10.6	11.6
77	SH	α -Himachalene	1450		tr				
78	SH	α -Humulene	1456			2.5			
79	AC	Geranyl acetone	1455	tr	0.4		0.4		
80	SH	(<i>E</i>)- β -Farnesene	1460	2.5	2.3				10.3
81	SH	<i>cis</i> -Muurolo-4(14), 5-diene	1463					4.5	1.7
82	SH	Epizonarene	1464					0.8	
83	SH	Alloaromadendrene	1462			0.1		1.2	
84	SH	γ -Himachalene	1474					tr	0.3
85	SH	<i>trans</i> -Cadin-1(6),4-diene	1470	tr	0.1				
86	SH	γ -Muurolole	1477	0.3	0.5	tr		3.0	12.9
87	SH	α -Muurolole	1480						3.2
88	SH	Germacrene D	1481		1.4				
89	SH	β -Selinene	1485	tr				14.6	5.7
90	SH	<i>cis</i> - β -Guaiene	1489					11.0	5.5
91	SH	δ -Selinene	1492						10.5
92	SH	Valencene	1493	0.1				1.7	1.6
93	SH	γ -Amorphene	1494		0.3				
94	SH	Bibyclogermacrene	1495			5.5	0.3		
95	SH	<i>trans</i> - β -Guaiene	1500	0.1	0.5	tr			1.2
96	SH	δ -Amorphene	1505	tr	0.1				5.5
97	SH	γ -Cadinene	1513	0.2	0.1				0.1
98	SH	δ -Cadinene	1523		0.6				1.1
99	SH	<i>trans</i> -Cadin-1(2),4-diene	1533	tr				0.1	
100	SH	Selina-3,7(11)-diene	1542					0.5	1.2
101	SH	α -Cadinene	1537	tr				0.7	
102	OS	Spathulenol	1568			0.2			
103	OS	Prenopsan-8-ol	1571						0.1
104	OS	Caryophyllene oxide	1581			1.4		tr	
105	NT	Hexadecane	1599		0.1				
106	OS	10-Epi- γ -eudesmol	1627					0.1	0.2
107	OS	Hinesol	1638						0.2
108	OS	Caryophylla-4(14)8(15)-dien-5-ol	1636			0.3			
109	OS	(<i>Z</i>)-Lanceol acetate	1856						0.2
Total identified				97.8	97.6	98.8	97.0	78.7	96.6
Non-terpene derivatives				3.9	64.1	4.4	93.7	0.7	0.2
Monoterpene hydrocarbons				81.2	10.4	0.4	0.2	1.7	2.5
Oxygenated Monoterpenes				8.0	13.5	0.1		0.9	
Sesquiterpene hydrocarbons				4.8	8.8	92.0	2.8	75.1	93.4
Oxygenated Sesquiterpenes						1.9		0.3	0.5
Phenylpropanoids					0.4				
Apocarotenoids					0.4		0.4		

^a) Compounds present at less than 0.1% were excluded from the table. ^b) *LRI*: Linear retention index. Relative contents of the volatile constituents expressed as percentages. ^c) Species and organs analysed- *Ha* Fl: *H. arenarium* flowers; *Ha* Le: *H. arenarium* leaves; *Hn* Fl: *H. nudifolium* flowers; *Hn* Le: *H. nudifolium* leaves; *Hg* Fl: *H. graveolens* flowers; *Hg* Le: *H. graveolens* leaves.

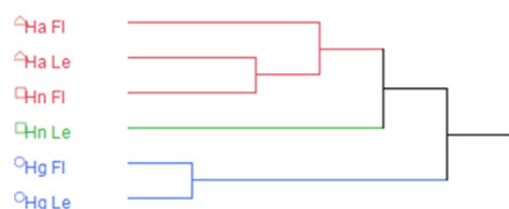


Figure 1: Dendrogram of the Hierarchical Cluster Analysis (HCA) of the EO composition of the three *Helichrysum* spp.

totarene, beyerene, cembrene, neocembrene, which were absent in *Ha* and *Hn*, contributed to the characterization of this group. β -caryophyllene represented a marker in the aroma of *H. arenarium* and *H. nudifolium*. High amount of sesquiterpenes characterized the EO composition of the majority examined species except for *Hn* leaves.

These results agree with Maggio *et al.* [22] that reported the predominance of β -caryophyllene in the EOs from *Helichrysum stoechas*, *H. rupestre* and *H. italicum* cultivated in Italy. This

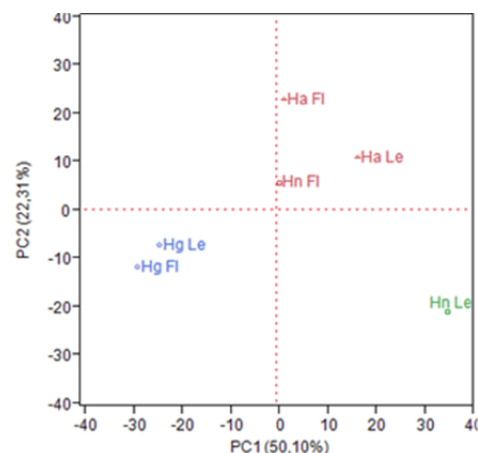


Figure 2: Principal Component Analysis (PCA) plot of the EO composition from the three examined *Helichrysum* spp.

compound, together with δ -cadinene, heneicosane and octadecane, was the main constituent of *H. arenarium* EO collected in Lithuania [7]. However aliphatic acids (34.6%) and their esters (28.5%)



Figure 3: Dendrogram of the Hierarchical Cluster Analysis (HCA) of volatiles emitted by the three *Helichrysum* spp..

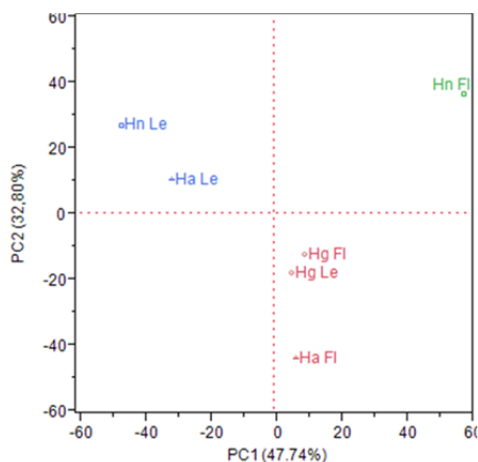


Figure 4: Principal Component Analysis (PCA) plot of volatile compounds spontaneously emitted by the three examined *Helichrysum* species.

together with linalool (1.7%), anethole (3.2%), carvacrol (3.6%) and α -muurolol (1.3%) were the major compounds in *H. arenarium* EO from Caucasus [8]. In a previous study from the literature aerial parts of *H. graveolens* grown in Turkey showed α -cubebene, β -caryophyllene, caryophyllene oxide and azulene-octahydro as main compounds [23] confirming the predominance of SH constituents, per other works [24, 25]. The EO of *H. arenarium* collected in Italy showed a similar composition of the same plant from Lithuania. Studies present in the literature showed the anti-inflammatory effects of β -caryophyllene and the strong antimicrobial properties of (*E*)-2-hexenal against pathogens confirming the use of these plants in the traditional medicine [26]. A different trend was found in the literature for *Helichrysum armenium* EO, another species growing in Iran, where limonene (21.2-29.2%) was the main constituent in all the aerial parts studied while in the present study was found in very low amount (0-5.2%) [27].

Monoterpenes were missing in *H. nudifolium* flowers and leaves but were present in low percentage in *H. graveolens*. Despite of several papers reported high number of monoterpenes in the EOs of several *Helichrysum* species (*H. gymnocephalum*, *H. dasyanthum*, *H. excisum*, *H. petiolare*, *H. italicum*), this class of constituents was absent also in other species like *H. felinum* [2a, 28-30].

In the SPME analysis of the *Helichrysum* spp. studied herein three groups can be observed by HCA and PCA statistical analysis (Fig 3 and 4): Group I with *Hn* and *Ha* leaves; Group II with only *Hn* flowers and Group III where *Hg* flowers and leaves had similar composition in comparison with *Ha* flowers. The leaves of *H.*

nudifolium and *H. arenarium* showed a similar aroma profile with non-terpenes (NT) as predominant compounds. (*Z*)-3-hexenol acetate (49.6% to 70.5, respectively) were the main constituent in the volatiles of *H. arenarium* and *H. nudifolium* leaves. Monoterpene hydrocarbons as β -pinene (55.2%) and α -pinene (15.8%) were the main constituents in the volatiles from *H. arenarium* flowers. *H. nudifolium* flowers represented a single group where high amount of β -caryophyllene (79.4%) was evidenced. This compound was present in low percentage in the leaves of the same species as well as in the other species studied. *H. graveolens* flowers and leaves showed sesquiterpene hydrocarbons as major class of constituents. Within this class (*E*)- β -farnesene, γ -muurolene, δ -selinene exhibited high percentage in leaves, while β -selinene, *cis*- β -guaiene in the flowers. α -neoclovene was identified in both the plant organs. Diterpenes (DT), which were predominant in the EOs from *Hg* flowers and leaves, were absent in the spontaneous volatile emission of this plant. Since no studies are reported in the literature, this work represents the first contribution to the volatile emission of *H. graveolens*.

Since essential oils from several *Helichrysum* species have pharmaceutical and cosmetic uses, the knowledge of the EO composition and the aroma profile of the selected *Helichrysum* spp. can help to identify species and chemotypes useful for industrial or ornamental applications.

Experimental

Plant material: The studied specimens (*H. arenarium*- *Ha*, *H. nudifolium*- *Hn*, *H. graveolens*- *Hg*) were supplied by CREA- Unità di ricerca per la floricoltura e le specie ornamentali (Sanremo, Italy). The correct identifications were done by one of us (Claudio Cervelli) and the voucher specimens were deposited in the Herbarium of Giardini Botanici Hanbury, La Mortola, Ventimiglia (Imperia, Italy) (*Ha*: HMGB. e/ 9006.2015. 004; *Hn*: HMGBH. e/ 9006. 2015. 005; *Hg*: HMGBH. e/ 9006. 2015. 001).

Essential Oil extraction: The aerial parts (leaves and flowers, separately) from the three *Helichrysum* species were subjected to hydrodistillation by Clevenger-type apparatus (3h). The essential oils (EOs) had very low yield ($\leq 0.01\%$), so they were collected with *n*-hexane (HPLC grade) and stored at -4°C in the dark until use.

Solid Phase Micro Extraction analyses (SPME): The analyses of the volatiles were performed using Supelco SPME device coated with polydimethylsiloxane (PDMS, 100 μm) to establish the volatile organic compound (VOC) emission of flowers and leaves of each *Helichrysum* species in according with methodology described in our previous work [31].

GC-FID and GC-MS analysis: The GC-MS analyses were performed as described in our previous studies [32-33].

Statistical analysis: JMP software package SAS (Institute, Cary, NC, USA) was used to statistical analysis. The hierarchical cluster analysis HCA was performed using Ward's method, with squared Euclidian distances as a measure of similarity. Compounds with contents below 1.0 % were excluded from statistical analysis.

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