

Antibacterial, antifungal and cytotoxic properties of novel N-substituted sulfonamides from 4-hydroxycoumarin

ZAHID H. CHOCHAN^{†,1}, ALI U. SHAIKH², ABDUL RAUF³, & CLAUDIU T. SUPURAN⁴

¹Department of Chemistry, Bahauddin Zakariya University, Multan 60800, Pakistan, ²Department of Chemistry, University of Arkansas at Little Rock, Little Rock, AR, 72204, USA, ³Department of Chemistry, Islamia University, Bahawalpur, Pakistan, and ⁴Università degli Studi di Firenze, Polo Scientifico, Laboratorio di Chimica Bioinorganica, Rm, 188, Via della Lastruccia 3, 50019 Sesto Fiorentino (Florence), Italy

(Received 9 March 2006; in final form 24 April 2006)

Abstract

A new series of 4-([2, 4-dioxo-2H-chromen-3 (4H)-ylidene] methyl) amino sulfonamides have been obtained by the condensation reaction of 4-hydroxycoumarin with various sulfonamides (sulfanilamide, sulfaguanidine, p-aminomethylsulfanilamide, p-aminoethylsulfanilamide, sulfathiazole, sulfamethoxazole, sulfamethazine and 4-[(2-amino-4-pyrimidinyl) amino] benzenesulfonamide) in the presence of an excess of ethylorthoformate. These compounds were screened for their *in-vitro* antibacterial activity against four Gram-negative (*E. coli*, *S. flexneri*, *P. aeruginosa* and *S. typhi*) and two Gram-positive (*B. subtilis* and *S. aureus*) bacterial strains and for *in-vitro* antifungal activity against *T. longifusus*, *C. albicans*, *A. flavus*, *M. canis*, *F. solani* and *C. glaberata*. Results revealed that a significant antibacterial activity was observed by compounds (4) and (5), (6) and (8) against two Gram-negative, (*P. aeruginosa* and *S. typhi*) and two Gram-positive (*B. subtilis* and *S. aureus*) species, respectively. Of these (4) was found to be the most active. Similarly, for antifungal activity compounds (3) and (8) showed significant activity against *M. canis* and, (6) and (8) against *F. solani*. The brine shrimp bioassay was also carried out to study their *in-vitro* cytotoxic properties and only two compounds, (4) and (8) possessing LD₅₀ = 2.9072 × 10⁻⁴ and 3.2844 × 10⁻⁴ M, respectively, displayed potent cytotoxic activity against *Artemia salina*.

Keywords: 4-Hydroxycoumarin, sulfonamides, antibacterial, antifungal, cytotoxicity

Introduction

Previously, we prepared and investigated some Schiff bases of aromatic/heterocyclic sulfonamide derived chromones [1–7] as inhibitors of the zinc enzyme carbonic anhydrase. The 15 CA isozymes presently known in humans are involved in many physiological and pathological processes, and their inhibition may thus be exploited clinically for the treatment of glaucoma in which CA II and CA XII are targeted by sulfonamide or sulfamate inhibitors [8,9]. Coumarins, member of the class benzopyrones are also good inhibitors for carbonic anhydrase [10] and display a variety of pharmacological properties

depending on their substitution pattern. Natural [11] and synthetic coumarins are known to possess antifungal/antibacterial properties [12–14]. The diverse biological activity of coumarin derivatives as anticoagulant is also well known [15,16]. The rapid spread of the acquired immunodeficiency syndrome (AIDS) epidemic has stimulated discovery of therapeutic agents to arrest the replication of the causative virus, human immunodeficiency virus (HIV). Recently, 3-substituted-4-hydroxycoumarin, phenoprocaumen [3-(*d*-ethyl-benzyl)-4-hydroxycoumarin] and analogous compounds have been identified as active nonpeptidic HIV protease inhibitors [17–21].

Correspondence: Professor Claudiu Supuran, Università degli Studi di Firenze, Polo Scientifico, Laboratorio di Chimica Bioinorganica Rm, 188, Via della Lastruccia 3, Sesto Fiorentino (Florence) 50019, Italy. Tel.: 39 055 4573005. Fax: 39 055 4573385.
E-mail: claudiu.supuran@unifi.it

[†]Present address: Department of Chemistry, University of Arkansas at Little Rock, Little Rock, AR 72204, U.S.A.
E-mail: zchohan@mul.paknet.com.pk

The coumarin ring forms a part of many heterocyclic compounds of pharmacological interest. Based on these findings and on the recent report [22] for the new synthetic routes to this class of compounds and their different important and interesting biological and pharmacological activities initiated us to synthesize a new class of sulfonamide derived coumarins and to explore their biological activities with the aim of obtaining more potent antibacterial and antifungal compounds. These synthesized compounds (1)–(8), were tested for *in-vitro* antibacterial activity against four Gram-negative (*Escherichia coli*, *Shigella flexneri*, *Pseudomonas aeruginosa* and *Salmonella typhi*) and two Gram-positive (*Bacillus subtilis* and *Staphylococcus aureus*) bacterial strains and for *in-vitro* antifungal activity against *Trichophyton longifusus*, *Candida albicans*, *Aspergillus flavus*, *Microsporium canis*, *Fusarium solani* and *Candida glaberata*. The coumarin-derived sulfonamide reported in this paper formulate not only a new class of antibacterial and antifungal agents but may also participate as good candidates for the globally alarming drug resistance problems in the clinic.

Material and methods

All reagents and solvents were used as obtained from the supplier or recrystallized/ redistilled as necessary. Thin-layer chromatography was performed using aluminum sheets (Merck) coated with silica gel 60 F₂₅₄. Infrared spectra (KBr discs) were recorded with a Hitachi Model 200–50 IR spectrophotometer. ¹H NMR spectra were recorded in d₆-DMSO with Bruker AM 300 and AM 400 spectrometers (Rheinstetten–Forchheim, Germany) operating at 300 and 400 MHz, respectively. Tetramethylsilane was used as an internal standard. Microanalytical data were determined using an Elemental Analyzer Flash EA 1112. Melting points were taken on a Gallenkamp apparatus and are uncorrected. *In-vitro* antibacterial and antifungal properties were studied at HEJ Research Institute of Chemistry, International Center for Chemical Sciences, University of Karachi, Pakistan.

General procedure for the preparation of compounds (1)–(8)

To a stirred solution of 4-hydroxycoumarin (1.62 g, 0.01 mole) and ethylorthoformate (2.25 g, 0.015 mole) in 2-butanol (30 mL) was added the respective sulfonamide (0.01 mole). The mixture was refluxed for 3 h. The precipitates formed during refluxing were cooled to room temperature and collected by suction

filtration. Washing with hot ethanol, afforded TLC pure products in good yield.

4-([2,4-dioxo-2H-chromen-3(4H)-ylidene]methyl)amino benzenesulfonamide (1). Yield 90%; m.p. 294–295 °C; IR (KBr, cm⁻¹): 3450 (NH₂), 1740 (lactone, C=O), 1680 (ketone, C=O), 1440 (S=O); ¹H NMR (DMSO-d₆, δ, ppm): 7.35 (d, 2H, benzene C_{3,5}-H), 7.42 (dd, 1H, chromen C₈-H), 7.54 (d, 2H, benzene C_{2,6}-H), 7.65 (ddd, 1H, chromen C₆-H), 7.70 (br s, 2H, SO₂NH₂), 7.78 (ddd, 1H, chromen C₇-H), 8.12 (dd, 1H, chromen C₅-H), 8.62 (s, 1H, =CH-N-), 9.85 (s, 1H, =C-NH-); Anal. Calcd. for C₁₆H₁₂N₂O₅S (344.34): C, 55.81; H, 3.51; N, 8.14. Found: C, 55.95; H, 3.40; N, 8.25%.

3-[4-([amino(imino)methyl]amino)sulfonyl]anilino]methylidene-2,4-dioxo-2H-chromene (2). Yield 96%; m.p. 276–277 °C; IR (KBr, cm⁻¹): 3440 (NH₂), 3315 (NH), 1745 (lactone, C=O), 1690 (ketone, C=O), 1580 (guanidine, C=N), 1445 (S=O); ¹H NMR (DMSO-d₆, δ, ppm): 7.40 (d, 2H, benzene C_{3,5}-H), 7.43 (dd, 1H, chromene C₈-H), 7.58 (d, 2H, benzene C_{2,6}-H), 7.67 (ddd, 1H, chromene C₆-H), 7.72 (s, 2H, -N=C-NH₂), 7.76 (ddd, 1H, chromene C₇-H), 7.82 (s, 1H, SO₂NH-), 8.12 (s, 1H, -C=NH), 8.14 (dd, 1H, chromene C₅-H), 8.65 (s, 1H, =CH-N-), 9.89 (s, 1H, =C-NH-); Anal. Calcd. for C₁₇H₁₄N₄O₅S (386.38): C, 52.84; H, 3.65; N, 14.50. Found: C, 52.60; H, 3.80; N, 14.65%.

4-([2,4-dioxo-2H-chromen-3(4H)-ylidene]methyl)amino)methyl]benzenesulfonamide (3). Yield 78%; m.p. 218–219 °C; IR (KBr, cm⁻¹): 3430 (NH₂), 3310 (NH), 1750 (lactone, C=O), 1700 (ketone, C=O), 1445 (S=O); ¹H NMR (DMSO-d₆, δ, ppm): 4.32 (d, 2H, -CH₂-), 7.36 (dd, 1H, chromen C₈-H), 7.44 (d, 2H, benzene C_{3,5}-H), 7.62 (ddd, 1H, chromen C₆-H), 7.65 (d, 2H, benzene C_{2,6}-H), 7.78 (s, 2H, SO₂NH₂), 7.75 (ddd, 1H, chromen C₇-H), 8.10 (dd, 1H, chromen C₅-H), 8.50 (s, 1H, =CH-N-), 8.62 (s, 1H, =C-NH-); Anal. Calcd. for C₁₇H₁₄N₂O₅S (358.37): C, 56.98; H, 3.94; N, 7.82. Found: C, 56.75; H, 3.80; N, 7.98%.

4-[2-([2,4-dioxo-2H-chromen-3(4H)-ylidene]methyl)amino)ethyl] benzenesulfonamide (4). Yield 88%; m.p. 256–257 °C; IR (KBr, cm⁻¹): 3435 (NH₂), 3320 (NH), 1745 (lactone, C=O), 1710 (ketone, C=O), 1450 (S=O); ¹H NMR (DMSO-d₆, δ, ppm): 3.10 (t, 2H, -CH₂-aromatic ring-), 3.32 (t, 2H, -CH₂-N-C=C-),

7.36 (dd, 1H, chromen C₈-H), 7.39 (d, 2H, benzene C_{3,5}-H), 7.61 (ddd, 1H, chromen C₆-H), 7.52 (d, 2H, benzene C_{2,6}-H), 7.70 (s, 2H, SO₂NH₂), 7.74 (ddd, 1H, chromen C₇-H), 8.10 (dd, 1H, chromen C₅-H), 8.47 (s, 1H, =CH-N-), 8.56 (s, 1H, =C-NH-); Anal. Calcd. for C₁₈H₁₆N₂O₅S (372.39): C, 58.05; H, 4.33; N, 7.52. Found: C, 57.80; H, 4.20; N, 7.70%.

4-({[2,4-dioxo-2H-chromen-3(4H)-ylidene]methyl}-amino)-N-(1,3-thiazol-2-yl) benzenesulfonamide (5). Yield 95%; m.p. 254 °C (decomp.); IR (KBr, cm⁻¹): 3315 (NH), 1740 (lactone, C=O), 1680 (ketone, C=O), 1615 (thiazole, C=N), 1440 (S=O); ¹H NMR (DMSO-d₆, δ, ppm): 6.35 (d, 1H, thiazole C₅-H), 6.67 (d, 1H, thiazole C₄-H), 7.44 (dd, 1H, chromen C₈-H), 7.48 (d, 2H, benzene C_{3,5}-H), 7.62 (d, 2H, benzene C_{2,6}-H), 7.68 (ddd, 1H, chromen C₆-H), 7.77 (ddd, 1H, chromen C₇-H), 7.88 (s, 1H, SO₂NH-), 8.16 (dd, 1H, chromen C₅-H), 8.67 (s, 1H, =CH-N-), 9.92 (s, 1H, =C-NH-); Anal. Calcd. for C₁₉H₁₃N₃O₅S₂ (427.45): C, 53.39; H, 3.07; N, 9.83. Found: C, 53.56; H, 2.70; N, 9.68%.

4-({[2,4-dioxo-2H-chromen-3(4H)-ylidene]methyl}-amino)-N-(5-methyl-3-isoxazolyl) benzenesulfonamide (6). Yield 80%; m.p. 273–274 °C; IR (KBr, cm⁻¹): 3310 (NH), 1735 (lactone, C=O), 1675 (ketone, C=O), 1605 (isoxazolyl, C=N), 1435 (S=O); ¹H NMR (DMSO-d₆, δ, ppm): 2.32 (s, 1H, -CH₃), 5.78 (s, 1H, isoxazolyl C₄-H), 7.44 (dd, 1H, chromen C₈-H), 7.50 (d, 2H, benzene C_{3,5}-H), 7.64 (d, 2H, benzene C_{2,6}-H), 7.69 (ddd, 1H, chromen C₆-H), 7.78 (ddd, 1H, chromen C₇-H), 7.90 (s, 1H, SO₂NH-), 8.16 (dd, 1H, chromen C₅-H), 8.68 (s, 1H, =CH-N-), 9.94 (s, 1H, =C-NH-); Anal. Calcd. for C₂₀H₁₅N₃O₆S (425.41): C, 56.47; H, 3.55; N, 9.88. Found: C, 56.67; H, 3.62; N, 9.68%.

N-(4,6-dimethyl-2-pyrimidinyl)-4-({[2,4-dioxo-2H-chromen-3(4H)-ylidene]methyl}amino) benzenesulfonamide (7). Yield 72%; m.p. 259–260 °C; IR (KBr, cm⁻¹): 3305 (NH), 1725 (lactone, C=O), 1670 (ketone, C=O), 1590 (pyrimidinyl, C=N), 1425 (S=O); ¹H NMR (DMSO-d₆, δ, ppm): 2.48 (s, 6H, -CH₃), 7.48 (dd, 1H, chromen C₈-H), 7.52 (d, 2H, benzene C_{3,5}-H), 7.74 (d, 2H, benzene C_{2,6}-H), 7.77 (ddd, 1H, chromen C₆-H), 7.84 (ddd, 1H, chromen C₇-H), 7.88 (s, 1H, pyrimidinyl C₅-H), 8.20 (dd, 1H, chromen C₅-H), 8.28 (s, 1H, SO₂NH-), 8.78 (s, 1H, =CH-N-), 10.21 (s, 1H, =C-NH-); Anal. Calcd. for C₂₂H₁₈N₄O₅S (450.46):

C, 58.66; H, 4.03; N, 12.44. Found: C, 58.48; H, 4.16; N, 12.55%.

4-({[2-({[2,4-dioxo-2H-chromen-3(4H)-ylidene]methyl}amino)-4-pyrimidinyl]amino} benzenesulfonamide (8). Yield 75%; m.p. 306 °C (decomp.); IR (KBr, cm⁻¹): 3300 (NH), 1720 (lactone, C=O), 1670 (ketone, C=O), 1575 (pyrimidinyl, C=N), 1420 (S=O); ¹H NMR (DMSO-d₆, δ, ppm): 7.31 (d, 1H, pyrimidinyl C₆-H), 7.55 (dd, 1H, chromen C₈-H), 7.62 (d, 2H, benzene C_{3,5}-H), 7.72 (ddd, 1H, chromen C₇-H), 7.77 (d, 2H, benzene C_{2,6}-H), 7.80 (ddd, 1H, chromen C₆-H), 7.84 (s, 1H, SO₂NH-), 7.92 (d, 1H, pyrimidinyl C₅-H), 8.22 (dd, 1H, chromen C₅-H), 8.88 (s, 1H, =CH-N-), 9.22 (s, 1H, pyrimidinyl-NH-benzene), 10.62 (s, 1H, =C-NH-); Anal. Calcd. for C₂₀H₁₅N₅O₅S (437.43): C, 54.91; H, 3.46; N, 16.01. Found: C, 54.72; H, 3.55; N, 15.84%.

Antibacterial bioassay (in-vitro)

The synthesized compounds (1)–(8) were screened *in vitro* for their antibacterial activity against four Gram-negative (*E. coli*, *S. flexneri*, *P. aeruginosa* and *S. typhi*) and two Gram-positive (*B. subtilis* and *S. aureus*) bacterial strains by the agar-well diffusion method [23]. The wells (6 mm in diameter) were dug in the media with the help of a sterile metallic borer with centers at least 24 mm apart. Two to eight hours old bacterial inocula containing approximately 10⁴–10⁶ colony-forming units (CFU/ml) were spread on the surface of the nutrient agar with the help of a sterile cotton swab. The recommended concentration of the test sample (1 mg/ml in DMSO) was introduced in the respective wells. Other wells supplemented with DMSO and reference antibacterial drug, imipenem, served as negative and positive controls, respectively. The plates were incubated immediately at 37 °C for 24 h. Activity was determined by measuring the diameter of zones showing complete inhibition (mm). In order to clarify any participating role of DMSO in the biological screening, separate studies were carried out with the solutions alone of DMSO and they showed no activity against any bacterial strains.

Antifungal activity (in-vitro)

Antifungal activities of all compounds were studied against six fungal cultures, *T. longifusus*, *C. albicans*, *A. flavus*, *M. canis*, *F. solani* and *C. glabrata*. Sabouraud dextrose agar (Oxoid, Hampshire, England) was seeded with 10⁵ (cfu) ml⁻¹ fungal spore suspensions and transferred to petri plates. Discs soaked in 20 ml (200 µg/mL in DMSO) of all

compounds were placed at different positions on the agar surface. The plates were incubated at 32 °C for seven days. The results were recorded [24] as zone of inhibition and compared with standard drugs mic-nazole and amphotericin B.

Cytotoxic activity

Brine shrimp (*Artemia salina leach*) eggs were hatched in a shallow rectangular plastic dish (22 × 32 cm), filled with artificial seawater, which was prepared with commercial salt mixture and double distilled water. An unequal partition was made in the plastic dish with the help of a perforated device. Approximately 50 mg of eggs were sprinkled into the large compartment, which was darkened while the other compartment was opened to ordinary light. After two days nauplii were collected by a pipette from the lighted side. A sample of the test compound was prepared by dissolving 20 mg of each compound in 2 mL of DMF. From this stock solutions 500, 50 and 5 µg/mL were transferred to 9 vials (three for each dilutions were used for each test sample and LD₅₀ is the mean of three values) and one vial was kept as control having 2 mL of DMF only. The solvent was allowed to evaporate overnight. After two days, when shrimp larvae were ready, 1 mL of sea water and 10 shrimps were added to each vial (30 shrimps/dilution) and the volume was adjusted with sea water to 5 mL per vial. After 24 h the numbers of survivors were counted [25]. Data were analyzed by a Finney computer program to determine the LD₅₀ values [26].

Result and discussion

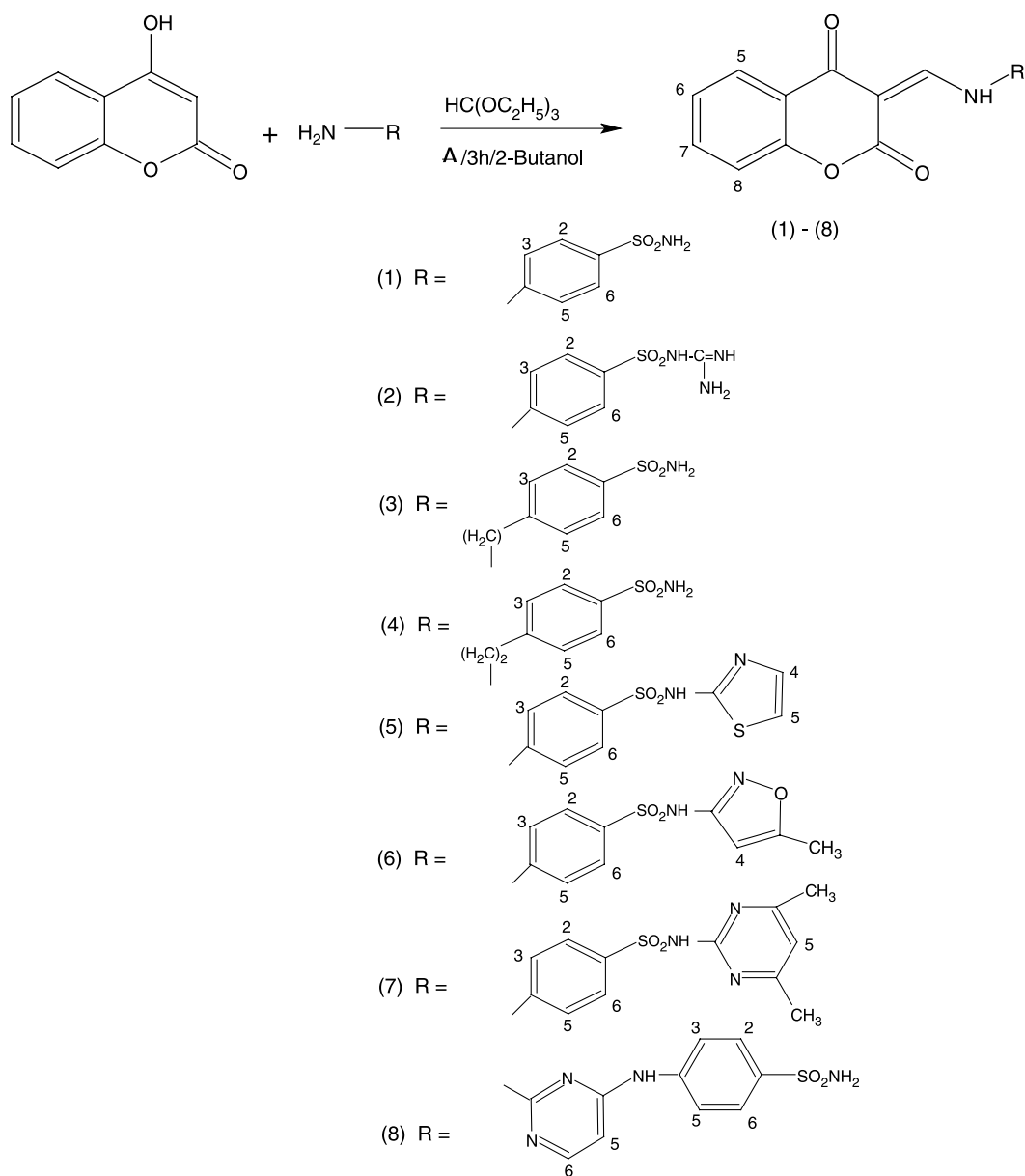
Chemistry

All synthesized compounds were prepared by refluxing an equimolar ratio of 4-hydroxycoumarin and the respective sulfonamide such as sulfanilamide, sulfaguanidine, p-aminomethylsulfanilamide, p-aminoethylsulfanilamide, sulfathiazole, sulfamethoxazole, sulfamethazine and 4-[(2-amino-4-pyrimidinyl) amino] benzenesulfonamide in 2-butanol. The reaction was carried out in the presence of ethyl orthoformate (Scheme 1) which successfully led to a new series of corresponding 4-({[2,4-dioxo-2H-chromen-3 (4H)-ylidene] methyl} amino) sulfonamides in good yield (72–96%). The reaction was rapid, and no observation supported any kind of competition from the intermolecular condensation of 4-hydroxycoumarin [27]. All the products were obtained as solids and their purities were checked by thin layer chromatography (eluent = ethanol/chloroform/ethyl-acetate, 1/2/1 v/v). All the synthesized compounds

were characterized by spectroscopic techniques (IR & ¹H-NMR) and their elemental analyses.

IR Spectra. The IR spectra of (1)–(8) showed bands resulting from the S=O, ketone (C=O) and lactone (C=O) stretchings in the region at 1420–1455, 1670–1710 and 1720–1750 cm⁻¹, respectively, in all the cases. In addition, the spectra of (2), (5)–(8) showed bands resulting from the guanidine (C=NH), thiazole (C=N), isoxazolyl (C=N), -SO₂-NH-pyrimidinyl (C=N) and -NH-pyrimidinyl (C=N) stretchings at 1580, 1615, 1605, 1590 and 1575 cm⁻¹, respectively. The IR spectra of (1)–(4), exhibited the -NH₂ stretchings in the region at 3430–3450 cm⁻¹. Also the IR spectra of (2)–(8) showed a band resulting from the NH stretching in the 3305–3320 cm⁻¹ region. All this evidence was supportive [28,29] of the formation of compounds (1)–(8).

¹H-NMR Spectra. The ¹H-NMR spectra of compounds (1)–(8) displayed the coumarin C₅-H and C₈-H protons at δ 8.10–8.22 and δ 7.36–7.55, respectively, as double doublets. The C₇-H and C₆-H protons of all the compounds appeared [30] as a separate doublet of double doublet at δ 7.72–7.84 and δ 7.61–7.80, respectively. The ¹H-NMR spectra displayed singlets between δ 9.56 and δ 10.62 and between δ 8.47 and δ 8.88 attributed to the NH and ethenyl protons. However, benzene C_{2,6}-H and C_{3,5}-H protons appeared as separate doublets at δ 7.52–7.77 and δ 7.35–7.62, respectively. The SO₂NH₂ or SO₂NH- protons in all cases appeared as a singlet at (7.70–δ 8.28. The ¹H-NMR spectrum of compound (2) also displayed -C=NH and -N=C-NH₂ protons as a singlet at δ 8.12 and δ 7.72, respectively. In the case of compound (3), the ¹H-NMR spectrum exhibited [31] methylenic (-CH₂-) protons as a doublet at δ 4.32. Similarly, in the spectrum of compound (4), the methylenic, (=C-NH-CH₂) and (-N=C-CH₂-) protons appeared as a triplet at δ 3.32 and δ 3.10, respectively. The ¹H-NMR spectrum of compound (5) displayed thiazole C₄-H and C₅-H protons as doublets at δ 3.32 and δ 3.10. In the case of compounds (6) & (7), the isoxazolyl C₄-H and pyrimidinyl C₅-H protons appeared as a singlet at δ 5.78 and δ 7.88, respectively. The spectra also displayed the methyl protons as a singlet at δ 2.32 and δ 2.48, respectively. The ¹H-NMR spectrum of compound (8) showed signals between δ 7.92 and δ 7.31 as a doublet, due to pyrimidinyl C₅-H and C₆-H protons, respectively. The spectrum also displayed the pyrimidinyl-NH-benzene proton as a singlet at δ 9.22. The elemental analysis data of these



Scheme 1. Synthesis of compounds (1)–(8)

compounds was also found to be in good agreement with the proposed structures of the synthesized compounds.

Antibacterial bioassay (In-vitro)

All compounds were tested against four Gram-negative (*E. coli*, *S. flexneri*, *P. aeruginosa* and *S. typhi*) and two Gram-positive (*B. subtilis* and *S. aureus*) bacterial strains according to the literature protocol [23] The results were compared with those of the standard drug imipenem. All the synthesized compounds exhibited varying degree of inhibitory effects on the growth of different tested strains (Table I). A significant activity

was observed by compounds (4) and (5), (6) and (8) against two Gram-negative (*P. aeruginosa* and *S. typhi*) and two gram positive (*B. subtilis* and *S. aureus*) species, respectively. Of these (4) was found to be the most active. However, compound (1) and (2) showed moderate activity against *P. aeruginosa*, *S. typhi*, *B. subtilis* and *S. aureus*; (3), against *P. aeruginosa*, *B. subtilis* and *S. aureus*; (4), (5), (6) and (8) against *E. coli* and *S. flexneri*; (7) against *S. typhi*, *B. subtilis* and *S. aureus*. Compounds (1), (2) and (3) were found to be inactive against *E. coli* and *S. flexneri*; in addition (3) was also found to be inactive against *S. typhi*. Compound (7) was found to be inactive against *E. coli*, *P. aeruginosa* and *S. flexneri*.

Table I. Antibacterial bioassay of compounds (1)–(8) (1 mg/mL in DMSO).

Bacteria	Compound (zones of inhibition in mm)								SD*
	1	2	3	4	5	6	7	8	
Gram-negative									
<i>E. coli</i>	00	00	00	10	07	08	00	07	30
<i>S. flexneri</i>	00	00	00	09	08	08	00	07	27
<i>P. aeruginosa</i>	10	12	15	20	19	17	00	18	24
<i>S. typhi</i>	08	11	00	20	19	18	12	17	25
Gram-positive									
<i>S. aureus</i>	07	10	08	27	27	26	16	25	33
<i>B. subtilis</i>	08	10	08	27	25	24	14	25	33

10 < : weak; 10-16: moderate; > 16: Significant.

* Standard Drug (Imipenem)

Antifungal bioassay (In vitro)

The antifungal screening of all compounds was carried out against *T. longifusus*, *C. albicans*, *A. flavus*, *M. canis*, *F. solani* and *C. glaberata* fungal strains according to the literature protocol [24]. The results were compared with those from the standard drugs miconazole and amphotericin B. These results (Table II) indicate that most of the compounds were found to be inactive against all fungal species except for compounds (3) and (8) which showed significant activity against *M. canis* and (6) and (8) against *F. solani*. However, compounds (1), (3), (4), (6), (7) and (8) exhibited low to moderate activity against at least one of the fungal strains, *F. solani*, *A. flavus*, *C. albicans*, *C. glaberata* and *T. longifusus*.

Minimum inhibitory concentration (MIC)

Preliminary screening showed that compounds (4), (5), (6) and (8) were the most active against both gram-negative and Gram-positive organisms. These four compounds were selected for minimum inhibitory concentration (MIC) studies (Table III). The MIC of all the four active compounds varied from 2.2806×10^{-8} – 1.1430×10^{-7} M. Compound

(4) again proved to be the most active. It inhibited the growth of *P. aeruginosa*, *S. typhi* and *S. aureus* at 2.6853×10^{-8} M. The minimum inhibitory concentration was determined using the disc diffusion technique [23] by preparing discs containing 10, 25, 50 and 100 $\mu\text{g/mL}$ of the compounds and applying the literature protocol [32].

Cytotoxic bioassay

All the synthesized compounds were screened for their cytotoxicity (brine shrimp bioassay) using the protocol of Meyer *et al* [25]. From the data recorded in Table IV, it is evident that only two compounds, (4) and (8) displayed potent cytotoxic activity against *Artemia salina*, while the other compounds were almost inactive in this assay. Compound (4) showed maximum activity ($\text{LD}_{50} = 3.2844 \times 10^{-4}$ M) in the present series of compounds, whereas the other active compound (8) of the series demonstrated slightly less activity ($\text{LD}_{50} = 2.9072 \times 10^{-4}$ M) than compound (4). The relationship between cytotoxicity and activity however, reveals that cytotoxicity is approximately 100-fold greater than concentration for the activity of the most active compound against the selected bacterial strains.

Table II. Antifungal bioassay of compounds (1)–(8) (200 $\mu\text{g/mL}$ in DMSO).

Organism	Compound (% Inhibition)								SD*
	1	2	3	4	5	6	7	8	
<i>T. longifusus</i>	15	00	10	00	00	11	10	09	A
<i>C. albicans</i>	00	00	00	00	00	00	10	00	B
<i>A. flavus</i>	00	00	00	06	00	00	00	00	C
<i>M. canis</i>	00	09	74	00	00	00	00	86	D
<i>F. solani</i>	00	00	00	00	00	69	00	82	E
<i>C. glaberata</i>	00	00	10	00	00	00	18	00	F

* Standard Drugs MIC $\mu\text{g/mL}$. A = Miconazole (70 $\mu\text{g/mL}$: 1.6822×10^{-7} M), B = Miconazole (110.8 $\mu\text{g/mL}$: 2.6626×10^{-7} M), C = Amphotericin B (20 $\mu\text{g/mL}$: 2.1642×10^{-8} M), D = Miconazole (98.4 $\mu\text{g/mL}$: 2.3647×10^{-7} M), E = Miconazole (73.25 $\mu\text{g/mL}$: 1.7603×10^{-7} M), F = Miconazole (110.8 $\mu\text{g/mL}$: 2.66266×10^{-7} M)

Table III. Minimum inhibitory concentration (M) for compounds (3), (4), (5), (6), (7) and (8) against selected bacteria.

No.	(4)	(5)	(6)	(8)
<i>P. aeruginosa</i>	2.6853×10^{-8}	2.3394×10^{-8}	2.3506×10^{-8}	5.7152×10^{-8}
<i>S. typhi</i>	2.6853×10^{-8}	2.3394×10^{-8}	2.3506×10^{-7}	2.2806×10^{-8}
<i>S. aureus</i>	2.6853×10^{-8}	5.8486×10^{-8}	2.3506×10^{-7}	1.1430×10^{-7}
<i>B. subtilis</i>	6.7133×10^{-8}	2.3394×10^{-7}	5.8766×10^{-8}	2.2860×10^{-7}

Table IV. Brine shrimp lethality bioassay data for compounds (1)–(8).

Compound	LD ₅₀ (M)
1	2.9041×10^{-3}
2	2.5881×10^{-3}
3	2.7904×10^{-3}
4	3.2844×10^{-4}
5	2.3394×10^{-3}
6	2.3506×10^{-3}
7	2.2199×10^{-3}
8	2.9072×10^{-4}

Acknowledgements

One of the authors (ZHC) wishes to thank the United States Department of State (U.S.A) and the Council for International Exchange of Scholars (CIES) for a William Fulbright award that made this research possible.

References

- [1] Supuran CT, Nicolae A, Popescu A. Carbonic anhydrase inhibitors. Part 35. Synthesis of Schiff bases derived from sulfanilamide and aromatic aldehydes: The first inhibitors with equally high affinity towards cytosolic and membrane-bound isoenzymes. *Eur J Med Chem* 1996;31:43–48.
- [2] Supuran CT, Popescu A, Ilisiu M, Costandache A, Banciu MD. Carbonic anhydrase inhibitors. Part 36. Inhibition of isoenzymes I and II with Schiff bases derived from chalcones and aromatic/heterocyclic sulfonamides. *Eur J Med Chem* 1996;31:439–447.
- [3] Supuran CT, Scozzafava A, Popescu A, Bobes-Tureac R, Banciu A, Creanga A, Bobes-Tureac G, Banciu MD. Carbonic anhydrase inhibitors. Part 43. Schiff base derived from aromatic sulfonamides: Towards more specific inhibitors for membrane-bound versus cytosolic isoenzymes. *Eur J Med Chem* 1997;32:445–452.
- [4] Popescu A, Simion A, Scozzafava A, Briganti F, Supuran CT. Carbonic anhydrase inhibitors. part 67 Schiff bases of some aromatic sulfonamide and their metal complexes: Towards more selective inhibitors of carbonic anhydrase isozyme IV. *J Enz Inhib* 1999;14:407–423.
- [5] Scozzafava A, Banciu MD, Popescu A, Supuran CT. Carbonic anhydrase inhibitors: Part 90. Synthesis of Schiff bases of hydroxybenzaldehyde with aromatic sulfonamides and their reactions with sulfonyl isocyanates. *J Enz Inhib* 2000; 15:533–540.
- [6] Hassan M, Scozzafava A, Chohan ZH, Supuran CT. Carbonic anhydrase inhibitors: Metal complexes of a sulfonamide derived Schiff base and their interaction with isozymes I, II and IV. *J Enz Inhib* 2001;16:499–505.
- [7] Hassan M, Chohan ZH, Scozzafava A, Supuran CT. Carbonic anhydrase inhibitors: Schiff's bases of aromatic and heterocyclic sulfonamides and their metal complexes. *J Enz Inhib Med Chem* 2004;19:263–267.
- [8] Winum JY, Casini A, Mincione F, Starnotti M, Montero JL, Scozzafava A, Supuran CT. Carbonic anhydrase inhibitors: N-(p-sulfamoylphenyl)- α -D-glycopyranosylamines as topically acting antiglaucoma agents in hypertensive rabbits. *Bioorg Med Chem Lett* 2004;14:225–229.
- [9] Vullo D, Innocenti A, Nishimori I, Pastorek J, Scozzafava A, Pastorekova S, Supuran CT. Carbonic anhydrase inhibitors. Inhibition of the transmembrane isozyme XII with sulfonamides—a new target for the design of antitumor and anti-glaucoma drugs. *Bioorg Med Chem Lett* 2005;15:963–969.
- [10] Hoult JRS, Payd M. Pharmacological and biochemical actions of simple coumarins: Natural products with therapeutic potential. *Gen Pharmacol* 1996;27:713–722.
- [11] Hossain CF, Okoyama E, Yamazaki M. A new series of coumarin derivatives having monoamine oxidase inhibitory activity. *Chem Pharm Bull* 1996;44:1535–1539.
- [12] Oliva A, Meepagala KM, Wedge DE, Harries D, Hale AI, Aliotta G, Duke SO. Natural fungicides from *Ruta graveolens* L. leaves, including a new quinolon alkaloid. *J Agric Fd Chem* 2003;51:890–896.
- [13] Manolov L, Danchev ND. Synthesis, toxicological and pharmacological assessment of some 4-hydroxycoumarin derivatives. *Eur J Med Chem* 1995;30:531–535.
- [14] Romines KR, Morris JK, Howe WJ, Tomich RK, Hong M-M, Chong K-T, Hinshaw RR, Anderson DJ, Strohbach JW, Turner SR, Mizsak SA. Cycloalkylpyranones and Cycloalkyldihydropyrones as HIV Protease Inhibitors: Exploring the Impact of Ring Size on Structure-Activity Relationships. *J Med Chem* 1996;39:4125–4130.
- [15] Zhao H, Neamah N, Hong H, Majumber A, Wang S, Sunder S, Milne GWA, Pommier Y, Bruke TR. Coumarin-Based Inhibitors of HIV Integrase. *J Med Chem* 1997;40: 242–249.
- [16] Thaisrivongs S, Romero DL, Tommasi RA, Janakiraman MN, Strohbach JW, Turner SR, Biles C, Morge RR, Johnson PD, Aristoff PA, Tomich PK, Lynn JC, Horng MM, Chong KT, Hinshaw RR, Howe WJ, Finzel BC, Watennpaugh KD. Structure-Based design of HIV protease inhibitors: 5,6-Dihydro-4-hydroxy-2-pyrones as effective, nonpeptidic inhibitors. *J Med Chem* 1996;39:4630–4642.
- [17] Chohan ZH, Hassan MU, Khan KM, Supuran CT. *In-vitro* antibacterial, antifungal and cytotoxic properties of sulfonamide-derived schiff's bases and their metal complexes. *J Enz Inhib Med Chem* 2005;20:183.
- [18] Ferroud D, Collard J, Klich M, Dupuis-Hamelin C, Mauvais P, Lassaing P, Bonnefoy A, Musicki B. Synthesis and biological evaluation of coumarincarboxylic acids as inhibitors of gyrase B, L-Rhamnose as an effective substitute for L-Noviose. *Bioorg Med Chem Lett* 1999;9:2881–2886.

- [19] Han S, Zhou V, Pan S, Liu Y, Hornsby M, McMullan D, Klock HE, Haugen J, Lesley SA, Nathanael G, Calwell J, Gu X-J. Identification of Coumarin Derivatives as a Novel Class of Allosteric MEKI Inhibitors. *Bioorg Med Chem Lett* 2005;15:5467–5473.
- [20] Chen S, Cho M, Karlsberg K, Zhou D, Yuan Y-C. Biochemical and biological characterization of a novel anti-aromatase coumarin derivative. *J Biol Chem* 2004;12:48071–48078.
- [21] Bandyopadhyay C, Sur KR, Patra R, Sen A. Synthesis of coumarin derivatives from 4-oxo-4H-1-benzopyran-3-carboxaldehyde via 3-(arylaminoethylene)chroman-2,4-dione. *Tetrahedron* 2000;56:3583–3587.
- [22] Takechi H, Oda Y, Nishizono N, Oda K, Machida M. Screening search for organic fluorophores: Synthesis and fluorescence properties of 3-azoyl-7-diethylaminocoumarin derivatives. *Chem Pharm Bull* 2000;48:1702–1710.
- [23] Atta-ur-Rahman, Choudhary MI, Thomsen WJ. *Bioassay Techniques for Drug Development*. The Netherlands: Harwood Academic Publishers; 2001. p 16.
- [24] McLaughlin JL, Chang C-J, Smith DL. In: Atta-ur-Rahman, editor. *Studies in Natural Products Chemistry*, “Bentch-Top” Bioassays for the Discovery of Bioactive Natural Products: an update, Structure and Chemistry (part-B). vol 9. The Netherlands: Elsevier Science Publishers B.V; 1991. p 383.
- [25] Meyer BN, Ferrigni NR, Putnam JE, Jacobsen LB, Nichols DE, McLaughlin JL. *Planta Medica* 1982;45:31.
- [26] Finney DJ. *Probit Analysis*. 3rd ed., Cambridge: Cambridge University Press; 1971.
- [27] Hamdi M, Sakellariou R, Speziale V. *J Heterocyclic Chem* 1992;29:1817.
- [28] Nakamoto K. *Infrared Spectra of Inorganic and Coordination Compounds*. 2nd ed., New York: Wiley Interscience; 1970.
- [29] Bellamy LJ. *The Infrared Spectra of Complex Molecules*. New York: John Wiley; 1971.
- [30] Simmons WW. *The Sadtler Handbook of Proton NMR Spectra*. Inc: Sadtler Research Laboratories; 1978.
- [31] Pasto DJ. *Organic Structure Determination*. London: Prentice Hall International; 1969.
- [32] Bauer AW, Kirby WM, Sherris JC, Turck M, *Am J Clin Pathol* 1966;45:493.