

Up-regulation of Skp2 after Prostate Cancer Cell Adhesion to Basement Membranes Results in BRCA2 Degradation and Cell Proliferation*

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Aberrant interaction of carcinoma cells with basement membranes (BM) is a fundamental pathophysiological process that initiates a series of events resulting in cancer cell invasion and metastasis. In this report, we describe the results of our investigations pertaining to the events triggered by the adhesion of normal (PNT1A) and highly metastatic (PC-3) prostate cells onto BM proteins. Unlike PNT1A, PC-3 cells adhered avidly to Matrigel BM matrix as well as to isolated collagen type IV, laminin, and heparan sulfate proteoglycan perlecan, main BM components. This aberrantly increased cancer cell adhesion resulted in sustained BRCA2 protein depletion and vigorous cell proliferation, a cascade triggered by β_1 integrin-mediated phosphatidylinositol 3-kinase activation leading to BRCA2 degradation in the proteasome. The latter effect was orchestrated by phosphatidylinositol-dependent up-regulation of Skp2, a subunit of the E3 ubiquitin complex that directly assembles the ubiquitin-proteasome complex. This effect was demonstrated by coimmunoprecipitation, in vitro ubiquitination, and ultimately by proteasome inhibition. Inhibition of Skp2 by siRNA or proteasome inhibitors prevented BRCA2 degradation and cell proliferation, thus demonstrating the dependence on the role of BRCA2 in cell proliferation. These results and elucidate the molecular mechanism of BRCA2 up-regulation in cancer cells with a view to identifying a key step in the biology of metastasis. The identification of this molecular pathway may provide novel therapeutic strategies aimed at modulating cell proliferation in prostate carcinoma.

Basement membranes (BM)² are thin layers of specialized extracellular matrix (ECM) that surround and closely associate

with epithelial and endothelial cells, muscle fibers, and nerves. They consist mostly of collagen type IV (COL4) admixed with laminins (LN), nidogens, and the heparan sulfate proteoglycan perlecan (PLN) and may contain small amounts of fibronectin (FN) (1, 2). Although the BM structural role in defining tissue architecture and compartmentalization has long been recognized, its dynamic role in the regulation of cell behavior has only recently been appreciated.

Aberrant interactions with BM proteins play a crucial role in cancer cells. Cancer cells must be able to adhere to BM proteins and increase cell motility to become motile, which is essential for invading a BM directly or indirectly. Cell behavior is regulated by the passage of various signals across the BM. These signals are transduced into the cell, where they play a pivotal role in the response to environmental cues. Key molecules such as growth factors (3, 4), integrins (5, 7), MAPK/ERK 1/2 (8, 9), and PI 3-kinase (10) mediate these various interactions help in modulating the expression of genes exerting stringent control upon cell growth, survival, and cell proliferation (12).

Cellular protein degradation via the ubiquitin-proteasome pathway is a prime pathway through which cells normally regulate processes involved in cell growth and proliferation (13, 14). There is evidence that a number of growth inhibitory molecules and tumor suppressor proteins, such as p53, p21, p27, p130, the β_{1C} integrin, and FOXO1, are preferentially degraded by the ubiquitin-proteasome system in carcinoma cells (14–17). Furthermore, E3 ubiquitin ligase family members Skp2 and Mdm2 have been shown to play a role in prostate cancer development and progression (18–20). In a previous report, we provided evidence for a novel pathological mechanism whereby prostate carcinoma cell adhesion to collagen type I (COL1), a major ECM protein at osseous metastatic sites, promotes cancer cell proliferation through depletion of BRCA2 protein, the product of a tumor suppressor gene whose inactivation accounts for an increased risk in cancer development (21–23). This newly described effect resulted from β_1 integrin-dependent activation of the PI 3-kinase pathway, which promoted BRCA2 ubiquitination and degradation in the proteasome (24).

In this study, we extended our investigations to elucidate the mechanisms by which β_1 integrin signaling in prostate cancer

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² The abbreviations used are: BM, basement membranes; ECM, extracellular matrix; COL4, collagen type IV; LN, laminin; FN, fibronectin; PLN, perlecan; MAPK, mitogen-activated protein kinase; ERK, extracellular signal-regulated kinase; PI 3-kinase, phosphatidylinositol 3-kinase; Me₂SO, dimethyl sulfoxide; E3, ubiquitin-protein isopeptide ligase; siRNA, small interference RNA.

cells resulted in BRCA2 protein degradation in the proteasome. We also provide evidence demonstrating that the BRCA2-associated trophic effect is not restricted to the osseous environment but is quite active in mediating cancer cell proliferation after interaction with BM proteins.

EXPERIMENTAL PROCEDURES

Cell Culture—PNT1A cells (a human prostate normal cell line established by immortalization of normal adult prostate epithelial cells) and PC-3 cells (a human prostate carcinoma cell line derived from a bone metastasis) were kept in culture as described previously (16, 24).

Cell Adhesion—Cell adhesion assays to ECM proteins were carried out using 96-well tissue culture plates as described previously (24). Plates were precoated with different concentrations of FN (3 $\mu\text{g}/\text{ml}$; Sigma), LN (10 $\mu\text{g}/\text{ml}$; Invitrogen), PLN (10 $\mu\text{g}/\text{ml}$; Sigma), or COL4 (10 $\mu\text{g}/\text{ml}$; Sigma) for 16 h at 4 °C. Coating with 10 $\mu\text{g}/\text{ml}$ bovine serum albumin (Sigma) served as negative control.

Adhesion to the BM matrix Matrigel (Sigma) was tested in 96-well plates coated with 50 μl /well of a 1:3 dilution in RPMI medium (Invitrogen) before cell plating. Cells were starved in serum-free methionine/cysteine-deficient RPMI 1640 (Sigma) for 45 min at 37 °C before labeling with 100 $\mu\text{Ci}/\text{ml}$ ^{35}S protein labeling mix (Amersham Biosciences) in 1 ml of methionine/cysteine-free RPMI medium containing 5% fetal bovine serum. After 24 h, 100 μl of a 0.2×10^6 cell suspension were seeded to adhere for 1 h onto Matrigel or bovine laminin (10 $\mu\text{g}/\text{ml}$) at 37 °C and were washed three times with phosphate-buffered saline. Cells were lysed in 100 μl of 150 mM NaCl and 2 mM EDTA containing 1% NP-40. Total protein concentration was determined by a scintillation counter (Beckman LS 5000TD).

Inhibition Assays were performed by preincubating cells for 1 h on ice in the presence of the β_1 integrin blocking antibody P4C10 (1 $\mu\text{g}/\text{ml}$; Santa Cruz Biotechnology, Temecula, CA), or the monocyte chemoattractant protein-1 (MCP-1) (100 ng/ml) on vascular endothelial surface protein-1 (VSP-1) (10 $\mu\text{g}/\text{ml}$; Santa Cruz Biotechnology) used as a negative control. Cell adhesion was recorded for each experiment.

Immunoblotting Analysis and Inhibition Assays—Cells were grown either onto FN (3 $\mu\text{g}/\text{ml}$), LN (10 $\mu\text{g}/\text{ml}$), PLN (10 $\mu\text{g}/\text{ml}$), or COL4 (10 $\mu\text{g}/\text{ml}$) and lysed in RNeasy lysis buffer. Protein extracts were analyzed by immunoblotting as described previously (24). Where indicated, cells were pretreated for 1 h with either P4C10 or 1C10, or the PI 3-kinase inhibitors wortmannin (0.1 μM ; Sigma), LY294002 (10 μM ; Calbiochem), or solvent alone (Me_2SO), or added with the proteasome inhibitor MG132 (10 μM). The following antibodies were used: 1 $\mu\text{g}/\text{ml}$ anti-BRCA2 polyclonal antibody (H-300; Santa Cruz Biotechnology, Santa Cruz, CA), 10 $\mu\text{g}/\text{ml}$ monoclonal antibody to β -tubulin (Sigma), 1 $\mu\text{g}/\text{ml}$ polyclonal antibody to Skp2 (H-435; Santa Cruz Biotechnology), 2 $\mu\text{g}/\text{ml}$ monoclonal antibody to Mdm2 (D-12; Santa Cruz Biotechnology), 1 $\mu\text{g}/\text{ml}$ anti-phospho-AKT-Ser-473 polyclonal antibody (Santa Cruz Biotechnology), 1 $\mu\text{g}/\text{ml}$ anti-AKT 1/2 polyclonal antibody (H-136; Santa Cruz Biotechnology), 0.2 $\mu\text{g}/\text{ml}$ anti-phospho-ERK monoclonal antibody (E-4; Santa Cruz Biotechnology), 0.2 $\mu\text{g}/\text{ml}$ anti-ERK2

polyclonal antibody (C-14; Santa Cruz Biotechnology), or 1:1000 dilution of anti-p85 α rabbit antiserum (Sigma).

Analysis of BRCA2 ubiquitination was performed as described previously (24). To analyze BRCA2 association with Skp2, cell extracts were precleared and incubated overnight with 2 μg of polyclonal antibody to BRCA2. Immunocomplexes were recovered with protein A-Sepharose (Sigma), washed five times with phosphate-buffered saline containing 1% Nonidet P-40, 2 mM phenylmethylsulfonyl fluoride, 10 mM sodium fluoride, 1 mM sodium orthovanadate, 5 mM sodium pyrophosphate and were subjected to 10% SDS-PAGE under reducing conditions followed by transfer to polyvinylidene difluoride membranes. Filters were immunoblotted using 1 $\mu\text{g}/\text{ml}$ monoclonal antibody to Skp2 (Zymed Laboratories Inc., San Francisco, CA) or 2 $\mu\text{g}/\text{ml}$ monoclonal antibody to BRCA2 (clone 5.23; Chemicon) following the manufacturers' instructions. Alternatively, whole cell extracts were immunoprecipitated with 2 μg of monoclonal antibody to Skp2 (Zymed Laboratories Inc.) and separated by 6% SDS-PAGE, and filters were immunoblotted using 1 $\mu\text{g}/\text{ml}$ anti-BRCA2 polyclonal antibody (H-300; Santa Cruz Biotechnology) or 1 $\mu\text{g}/\text{ml}$ polyclonal antibody to Skp2 (H-435; Santa Cruz Biotechnology).

Transfection and Luciferase Assays—Cells were transfected with BRCA2 cDNA (a kind gift from Dr. M. D. Anderson, University of Texas M. D. Anderson Cancer Center) or a negative form of the same construct (pcDNA3; Invitrogen) as described previously (24). A siRNA specific for BRCA2 (Santa Cruz Biotechnology) and used as a negative control was transfected into cells according to the manufacturer's instructions. Luciferase reporter assays were performed as described previously (24).

Statistical analysis of the data are reported as the mean \pm S.E. Statistical significance was determined by the Student's *t* test. All experiments were repeated at least twice.

RESULTS

Prostate Normal and Carcinoma Cells Adhere Differently to BM—We investigated the adhesive properties of PNT1A and PC-3 cells to LN, PLN, and COL4, major components of BM, and to FN, a widely expressed extracellular matrix protein (25) enriched at the prostatic stroma but a minor component in BM. As shown in Fig. 1A, PNT1A cells adhered efficiently to FN, to a lesser extent to LN and PLN ($\sim 60\%$ of FN levels), and showed no adhesion to COL4. Highly metastatic PC-3 cells exhibited a reversed affinity pattern, with partial loss of adhesion to FN and newly gained adhesion capabilities to BM proteins. Because cell adhesion to FN, LN, COL4, and PLN (26) is mediated by integrin receptors that share a common β_1 subunit ($\alpha_5\beta_1$ to FN, $\alpha_6\beta_1$ to LN, $\alpha_1\beta_1$, $\alpha_2\beta_1$, and $\alpha_3\beta_1$ to COL4, $\alpha\beta_1$ to PLN), we tested the effect of the β_1 integrin-blocking antibody P4C10 on cell adhesion. As shown in Fig. 1B, adhesion to FN was reduced by $42 \pm 6\%$ ($p < 0.002$) and $39 \pm 7\%$ ($p < 0.001$) in PNT1A and PC-3 cells, respectively. Adhesion to LN was inhibited by $\sim 25\%$ in PNT1A and $\sim 75\%$ in PC-3 cells, adhesion to PLN was almost completely inhibited in both cell types, and onto COL4 PC-3

BM Proteins Reduce BRCA2 Expression in Cancer Cells

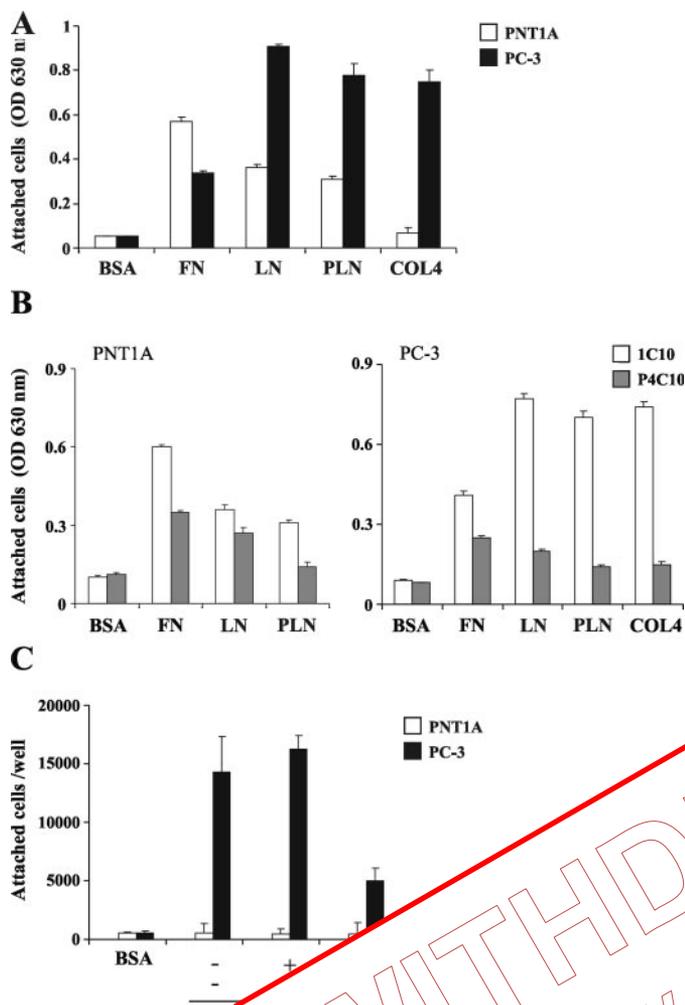


FIGURE 1. Prostate normal adhesion and BRCA2 protein depletion. **A**, 1.5×10^5 PNT1A or PC-3 cells were grown onto plates coated with BSA (10 μ g/ml), FN (10 μ g/ml), LN (10 μ g/ml), PLN (10 μ g/ml), or COL4 (10 μ g/ml) at 37 °C for 1 h. Cell adhesion was quantified by measuring the optical density at 630 nm. **B**, PNT1A or PC-3 cells were grown onto plates coated with BSA, FN, LN, PLN, or COL4 as described above. PNT1A or PC-3 cells were incubated for 1 h on ice in the presence or absence of the β_1 integrin-blocking antibody P4C10 or 1C10 as control and then grown onto plates coated with BSA, FN, LN, PLN, or COL4 as described above. **C**, PNT1A or PC-3 cells were incubated for 1 h on ice in the presence or absence of P4C10 or 1C10 before being allowed to adhere to BSA (10 μ g/ml) or Matrigel (0.2 mg of protein/well) for 1 h at 37 °C. Following three sequential saline washes, cells were lysed and the radioactivity was measured. Data are expressed as mean \pm S.E. of triplicate wells. A representative experiment of three is shown.

cell adhesion was almost completely inhibited ($90 \pm 7\%$, $p < 0.0001$).

To investigate whether these findings could be reproduced onto naturally occurring BM layers, we tested the adhesive properties of normal and cancer prostate cells to Matrigel, a reconstituted BM preparation (27). As shown in Fig. 1C, whereas PC-3 cells adhered efficiently to Matrigel in a manner that was significantly inhibited by the anti- β_1 integrin-blocking antibody ($\sim 70\%$ inhibition, $p < 0.004$), normal prostate cell adhesion was nil in the time period considered.

Adhesion of PC-3 Cells to BM Proteins Decreases BRCA2 Protein Levels in a β_1 Integrin-dependent Manner—We asked whether PC-3 cell adhesion to BM proteins exercised a modu-

latory effect upon BRCA2 expression. To this effect, PNT1A and PC-3 cells were grown onto plates coated with FN, LN, PLN, COL4, or Matrigel and BRCA2 levels were assessed. As depicted in Fig. 2A, whereas PNT1A cells transiently increased BRCA2 protein after adhesion to BM proteins (2.2 ± 0.3 -fold onto FN after 2.5 h, $p < 0.002$), PC-3 cells exhibited almost complete (PLN) or complete (LN, COL4, Matrigel) loss of detectable BRCA2 after adhesion for 6 h. The strongest down-regulatory effect was observed onto COL4, which caused BRCA2 protein levels to decrease after only 2.5 h by $68 \pm 4\%$ ($p < 0.01$). BRCA2 protein levels did not recover after 12 h of cell adhesion (data not shown). The β_1 integrin-blocking antibody P4C10 partly rescued BRCA2 protein to $45 \pm 6\%$ ($p < 0.001$) after 6 h onto COL4 (Fig. 2B) as well as onto other BM proteins (data not shown). As COL4 is the most abundant component of the BM, most of the subsequent experiments were performed onto this BM protein.

BRCA2 Protein Depletion by Cancer Cell Adhesion to BM Increases DNA Synthesis—To investigate whether the signaling cascade initiated by cell adhesion to BM proteins had any effect on cell proliferation, we measured the incorporation of [3 H]thymidine in PNT1A and PC-3 cells on various substrates (Fig. 3A). We observed no variations irrelevant to cell adhesion. Adhesion to COL4 enhanced [3 H]thymidine incorporation by $291 \pm 24\%$ ($p < 0.0002$). This trophic response was almost completely inhibited by PNT1A adhesion to PLN ($\sim 175\%$) (data not shown). The response to COL4 was partially inhibited by P4C10 to 55% (data not shown). This response was also inhibited and confirmed by MG132 (Fig. 3B), adhesion to Matrigel increased [3 H]thymidine incorporation by $509 \pm 27\%$ and P4C10 inhibited this response by 55% ($p < 0.0002$). The increase in [3 H]thymidine incorporation upon adhesion to BM proteins could be reversed by transfecting PC-3 cells with wild-type BRCA2 cDNA (Fig. 3C). In these experiments, PC-3 cells were transiently transfected with BRCA2 cDNA or empty vector for 36 h, after which we measured BRCA2 protein levels (upper panel) and [3 H]thymidine incorporation upon 12 h of cell adhesion to COL4 in the presence or absence of the proteasome inhibitor MG132 (lower panel). After transfection, BRCA2 protein levels increased 2.3-fold compared with mock-transfected cells. Upon COL4 adhesion, BRCA2 decreased by $44 \pm 8\%$ ($p < 0.03$) in transfected cells, but this reduction had no effect in [3 H]thymidine incorporation, which remained at basal levels throughout the experiment. On the contrary, mock-transfected cells exhibited complete disappearance of BRCA2 protein upon adhesion to COL4, which resulted in a burst in [3 H]thymidine incorporation. This trophic effect could be reversed with the proteasome inhibitor MG132, which decreased [3 H]thymidine incorporation by 94% ($p < 0.003$). Treatment of BRCA2-transfected cells with MG132 resulted in BRCA2 accumulation to $245 \pm 29\%$ ($p < 0.001$).

Skp2 Promotes BRCA2 Protein Depletion in PC-3 Cells Adherent to BM—Because it is known that F-box proteins Skp2 and Mdm2 mediate the ubiquitin-dependent degradation of several negative regulators of cell proliferation in cancer (17, 28,

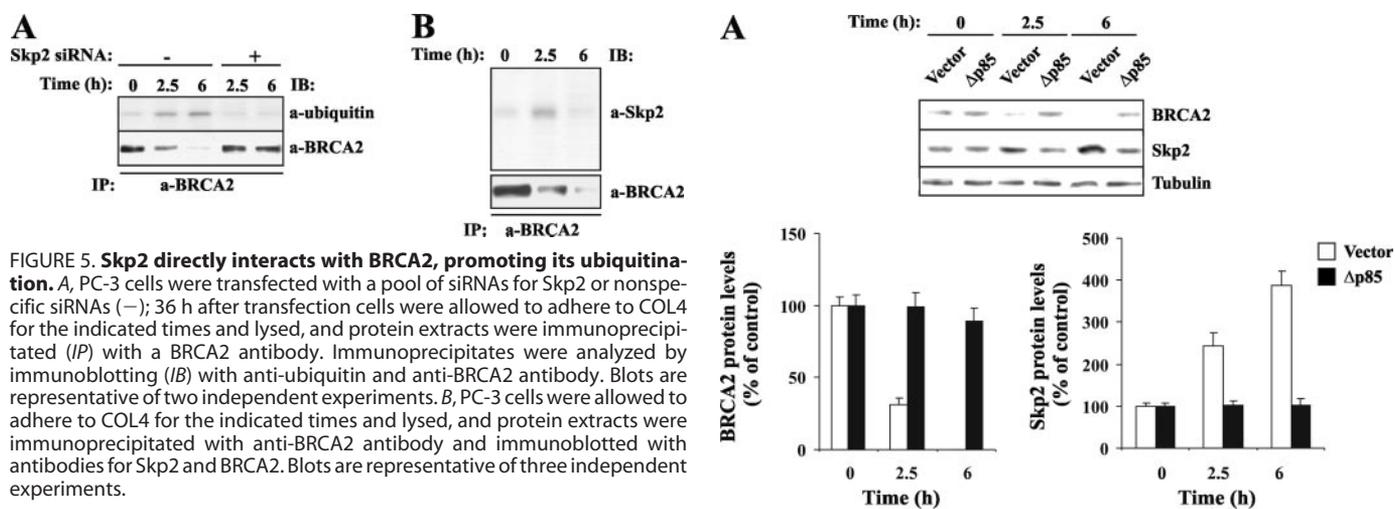


FIGURE 5. Skp2 directly interacts with BRCA2, promoting its ubiquitination. *A*, PC-3 cells were transfected with a pool of siRNAs for Skp2 or nonspecific siRNAs (–); 36 h after transfection cells were allowed to adhere to COL4 for the indicated times and lysed, and protein extracts were immunoprecipitated (IP) with a BRCA2 antibody. Immunoprecipitates were analyzed by immunoblotting (IB) with anti-ubiquitin and anti-BRCA2 antibody. Blots are representative of two independent experiments. *B*, PC-3 cells were allowed to adhere to COL4 for the indicated times and lysed, and protein extracts were immunoprecipitated with anti-BRCA2 antibody and immunoblotted with antibodies for Skp2 and BRCA2. Blots are representative of three independent experiments.

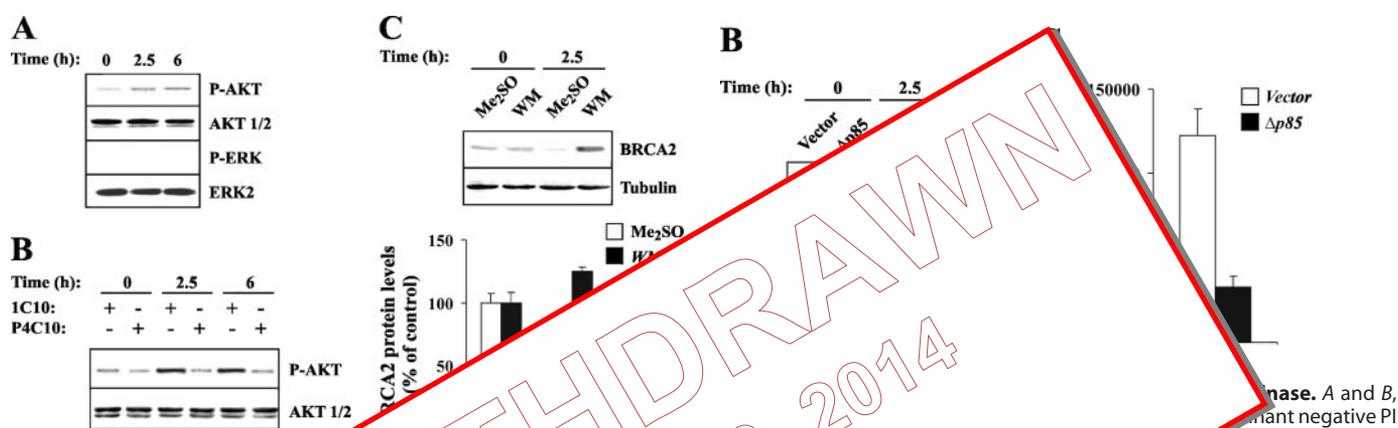
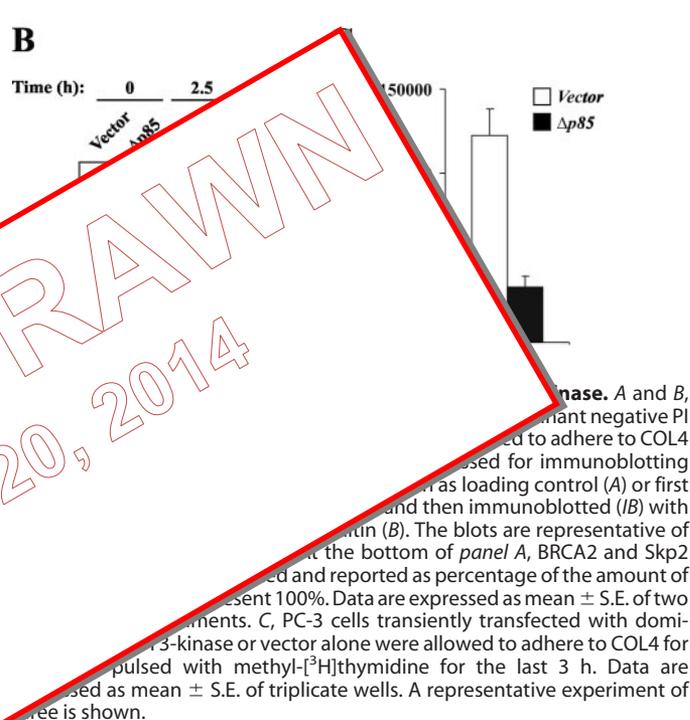


FIGURE 6. PC-3 cell adhesion to BM proteins is dependent on β_1 integrin-dependent mechanisms. *A*, PC-3 cells were allowed to adhere to COL4 at 37 °C for 0–6 h, washed, and lysed, and protein extracts were electrophoresed onto 4–20% gradient gels. Membranes were probed with anti-phospho-AKT (P-AKT) or anti-phospho-ERK (P-ERK). Substrates were also immunoblotted with anti-AKT 1/2 (AKT 1/2). Blots are representative of three independent experiments. *B*, PC-3 cells were incubated with the β_1 integrin-blocker 1C10 or P4C10 before plating onto COL4 and being probed for AKT. Blots are representative of two independent experiments. *C*, PC-3 cells were pretreated for 1 h at 37 °C with 0.1 μ M wortmannin (W), or the solvent alone (*Me*₂SO). Cells were then allowed to adhere to COL4 at 37 °C for 0 or 2.5 h and were washed, lysed, and protein extracts were immunoblotted. Membranes were probed with anti-BRCA2 and β -tubulin as loading control. The blots are representative of two independent experiments. *Bottom*, BRCA2 protein levels were quantitated and reported as percentage of the amount of protein at 0 h, set to represent 100%. Data are expressed as mean \pm S.E. of two independent experiments.

PC-3 cells when compared with normal PNT1A cells. This latter observation could be partly correlated with the lower expression of the FN receptor β_3 integrin in PC-3³ when compared with PNT1A cells.

Another crucial finding is that after adhesion to isolated BM proteins, particularly to COL4, highly invasive carcinoma cells exhibit a burst in [³H]thymidine incorporation, a sensitive measurement of new DNA synthesis and cell proliferation. This trophic response seems to be elicited by β_1 integrin adhesion, as

³ L. Moro, unpublished data.



a similar phenomenon was previously demonstrated onto COL1, a ligand for β_1 integrin receptors (24). The proliferative response after adhesion to LN and PLN was slightly weaker than with COL4, and no effect was noticeable onto FN. When allowed to adhere onto the BM matrix Matrigel, prostate cancer cells exhibited a proliferation surge that was stronger than with isolated components, suggesting cooperation rather than competition among various proteins in activating proliferative signals.

The abnormal proliferative response following PC-3 cell adhesion onto BM was linked to a sustained BRCA2 protein depletion and could be completely averted by wild-type BRCA2 cDNA transfection. This pathway seems to switch cell proliferation on and off depending on a critical amount of BRCA2 protein, as suggested by the unresponsiveness to mild reductions in BRCA2 protein after PC-3 cell adhesion to FN. Conversely, normal cell adhesion to FN resulted in an increase in BRCA2 expression with no discernible effect upon DNA syn-

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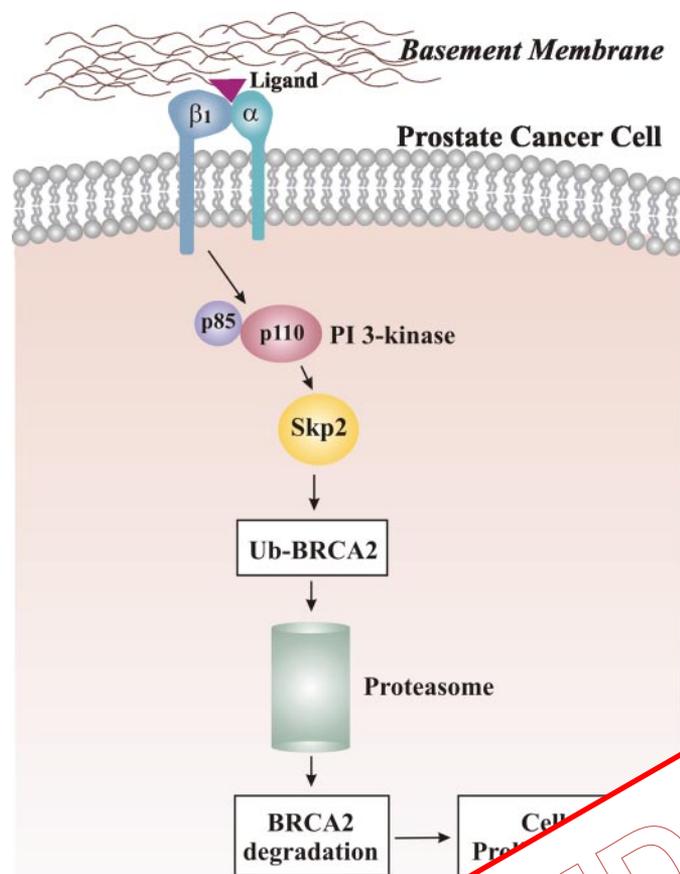


FIGURE 8. Hypothesized role of BRCA2 in prostate cancer cell proliferation. BM-dependent PI 3-kinase, increasing Skp2 protein levels, promoting its ubiquitination (Ub-BRCA2) and subsequent degradation. Ultimately, BRCA2 degradation at the BM interfaces.

thesis. Our findings add to the existing knowledge suggesting that BRCA2 also functions in cell proliferation and tumor growth. In a study using recombinant BRCA2 overexpression in pancreatic adenocarcinoma cells, a significant decrease in tumorigenicity was observed in mice (31). More recently, Miyamoto et al. (32) reported that Capan-1 cells exhibited an increased proliferation rate after adhesion to COL4. The mechanisms involved are not yet well understood. Normally, part of the cellular response to DNA damage involves the activation of an ATM/p53/Mdm2 feedback loop that regulates cell cycle progression and/or apoptosis in response to relative amounts of a DNA repair complex containing BRCA2 (33–35). However, involvement of this mechanism seems unlikely because our experiments demonstrate no relationship between β_1 integrin signaling and Mdm2 levels and suggest an alternative pathway involving Skp2 (see below). Furthermore, PC-3 (and Capan-1) cells exhibit inactivating mutations in the p53 gene (36, 37).

Depletion of BRCA2 protein upon cancer cell adhesion to BM proteins and osseous COL1 (24) occurs through protein degradation in the proteasome. However, the precise mechanisms involved had not been elucidated before. In the present study, we have provided evidence that this event is mediated by

Skp2, an F-box protein that associates with Skp1, Cul1, and Roc1/Rbx1 to form the SCF(Skp2) ubiquitin ligase complex (28). Indeed, down-regulation of Skp2 by siRNA was sufficient to rescue BRCA2 levels inhibiting the burst in cell proliferation upon cancer cell adhesion to BM. Evidence that this mechanism may be relevant in prostate cancer has also been provided in immunohistochemical studies performed on 622 radical prostatectomy specimens that demonstrate that Skp2 levels and cell-labeling frequency increase dramatically in both premalignant prostatic intraepithelial lesions and prostate carcinoma (19). Changes in Skp2 levels are dependent upon β_1 integrin-mediated signaling for Skp2 up-regulation, and subsequent BRCA2 degradation could be prevented by a β_1 integrin-blocking antibody. Evidence demonstrating that cell adhesion to the ECM results in Skp2 mRNA and protein up-regulation had been provided previously (38). We confirm those results and demonstrate for the first time that β_1 integrin-mediated signaling is necessary for ECM-dependent changes in Skp2 expression.

The up-regulation of Skp2 following β_1 integrin adhesion is signaled through PI 3-kinase and could be abrogated by transfecting cells with a dominant negative form of PI 3-kinase. These observations support the hypothesis implicating PI 3-kinase/AKT in BRCA2 protein degradation.

Our findings also support the hypothesis implicating PI 3-kinase/AKT in BRCA2 protein degradation. The up-regulation of this pathway has been shown to be necessary for cell proliferation and tumor growth. In addition, immunohistochemical evidence supports the hypothesis implicating PI 3-kinase/AKT in BRCA2 protein degradation in prostate intraepithelial lesions and prostate carcinoma. Our findings also support the hypothesis implicating PI 3-kinase/AKT in BRCA2 protein degradation. We demonstrated that Skp2 directly promotes the degradation of BRCA2. As expected, this process requires PI 3-kinase/AKT activation. β_1 integrin-mediated signaling following cancer cell adhesion with BM proteins, a crucial physiopathological phenomenon at the beginning of the metastatic cascade, may be a key event in the progression of prostate carcinoma to distant organs. Furthering the understanding of this molecular pathway may prove valuable in designing new therapeutic strategies aimed at modifying the natural history of prostate carcinoma.

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