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SHORT COMMUNICATION

Rosmarinic acid inhibits some metabolic enzymes including glutathione S-transferase, lactoperoxidase, acetylcholinesterase, butyrylcholinesterase and carbonic anhydrase isoenzymes

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Abstract

Rosmarinic acid (RA) is a natural polyphenol contained in many aromatic plants with promising biological activities. Carbonic anhydrases (CAs, EC 4.2.1.1) are widespread and intensively studied metalloenzymes present in higher vertebrates. Acetylcholinesterase (AChE, E.C. 3.1.1.7) is intimately associated with the normal neurotransmission by catalysing the hydrolysis of acetylcholine to acetate and choline and acts in combination with butyrylcholinesterase (BChE) to remove acetylcholine from the synaptic cleft. Lactoperoxidase (LPO) is an enzyme involved in fighting pathogenic microorganisms, whereas glutathione *S*-transferases (GSTs) are dimeric proteins present both in prokaryotic and in eukaryotic organisms and involved in cellular detoxification mechanisms. In the present study, the inhibition effects of rosmarinic acid on tumour-associated carbonic anhydrase IX and XII isoenzymes, AChE, BChE, LPO and GST enzymes were evaluated. Rosmarinic acid inhibited these enzymes with K_is in the range between micromolar to picomolar. The best inhibitory effect of rosmarinic acid was observed against both AChE and BChE.

Keywords

Acetylcholinesterase, butyrylcholinesterase, carbonic anhydrase, glutathione S-transferase, lactoperoxidase, rosmarinic acid

History

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Introduction

For many natural phenolic compounds, such as polyphenols directly provided by human diet, their beneficial properties have been primarily related to antioxidant activities, which are associated with their redox behaviour. Polyphenolic compounds derived from natural products are well known to possess a range of biological activities¹. They incorporate one or more hydroxyl moieties bonded directly to an aromatic carbon atom with the substitution patterns on the aromatic ring creating a large chemical variety. Therefore, free-radical scavenging by natural polyphenols has been widely studied².

Rosmarinic acid (Figure 1) is an ester of caffeic and 3,4dihydroxyphenyllactic acids. It is generally found in a wide range of species and plants. It is especially present as a secondary metabolite in medicinal and food plants. Rosmarinic acid was first isolated and characterised in 1958 from rosemary (Rosmarinus officinalis)³. This polyphenol and its derivatives are abundantly found in sage or savoury herbs. Also, these plants are widely used in Mediterranean and Anatolian folk medicine as culinary herbs. It is also used as a fragrant additive in cosmetic applications and other applications. It was reported that rosmarinic acid had several biological activities, namely antioxidant, anti-inflammatory, antimutagenic, antibacterial, antiviral, antitumor, hepatoprotective and cardioprotective properties. Also, rosmarinic acid had health promoting and beneficial effects. It is supposed to act as a preformed constitutively accumulated defence compound in plants. Plant extracts that contain rosmarinic acid are a good potential source of antioxidants for food protection and pharmaceutical applications. Rosmarinic acid is a natural polyphenol carboxylic acid, an ester of caffeic and 3,4-dihydroxyphenyllactic acids and an important phenolic bioactive compound⁴.

Carbonic anhydrases (CAs, EC 4.2.1.1) are zinc-containing metalloenzymes that catalyse the reversible hydration of carbon dioxide (CO₂) in a two-step reaction, to yield bicarbonate (HCO₃⁻) and protons (H⁺)⁵. CO₂, HCO₃⁻ and H⁺ are essential molecules or

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Figure 1. Chemical structure of rosmarinic acid as glutathione *S*-transferase, lactoperoxidase, acetylcholinesterase, butyrylcholinesterase and carbonic anhydrase isoenzymes inhibitor.

ions in many important physiologic processes in all life kingdoms, throughout the tree of life and for this reason, relatively high amounts of CAs are present in different tissues or cell compartments of most investigated organisms⁶.

$$CO_2 + H_2O \iff H_2CO_3 \iff HCO_3^- + H^+$$

CA

These enzymes are present in all organisms, from the very simple to the complex ones. This metalloenzyme superfamily includes six distinct genetic families (α -, β -, γ -, δ -, ζ - and η -CAs) known to date, which constitute an interesting example of convergent evolution at the molecular level⁷. They vary in their preference for the catalytic metal ions used within the active site, since Zn^{2+} , Cd^{2+} or Fe^{2+} can be used within their active sites⁸. The α -CA isoforms differ significantly in their localization and tissue distribution. They are present in vertebrates, protozoa, algae and cytoplasm of green plants and in some bacteria. CA I, II, III, VII and XIII are cytosolic isoforms, CA IV, IX, XII and XIV are membrane-bound, CA VA and VB are mitochondrial, whereas CA VI is secreted. CA IX and XII are known as the membrane tumour-associated CAs, being found in a limited number of normal tissues, such as the gastrointestinal mucosa and body cavity lining9. An important role of CA IX and XII as tumour pHregulating enzymes, involved in the survival or proliferation of the tumour cells within the hypoxic, acidic niche typical of many solid cancers. CA I, II, IX and XII were studied in this study. CA I and II are present at high concentrations in the cytosol of erythrocytes and the gastrointestinal tract. CA IX is found in tumour cells and absent or is present in very limited amount in normal tissues¹⁰.

Cholinesterases (ChE) are an enzyme family that catalyse the hydrolysis of acetylcholine (ACh) into choline and acetic acid, an essential process for the restoration of the cholinergic neuro-transmission. There are two cholinesterase types: acetylcholinesterase (AChE; EC 3.1.1.7) and butyrylcholinesterase (BChE; EC 3.1.1.8)¹¹.



Butyrylcholinesterase is plasma ChE and a non-specific ChE enzyme that hydrolyses many different choline-based esters. In humans, it is made in the liver, found mainly in blood plasma and encoded by the BChE gene. AChE is known to be abundant in the muscle, brain and erythrocyte membrane, whereas BChE has a higher activity in liver, intestine, heart, kidney and lung. AChE and BChE share 65% amino acid sequence homology and have similar molecular forms and active sites despite being products of different genes on the human chromosomes¹².



Both cholinesterases participate in cholinergic neurotransmission by hydrolysing ACh in the central and peripheral nervous system. The symptomatic Alzheimer's disease (AD) treatment involves the use of cholinesterase inhibitors (ChEIs) such as Rivastigimine. ChEIs are the first-line drugs in the symptomatic treatment of AD, as by inhibiting cholinesterase they lead to an increased synaptic level of the neurotransmitter¹³.

Lactoperoxidase (LPO, E.C. 1.11.1.7) is of growing interest due to its distinctive biological activity, such as biocidal and biostatic ones. The mechanism of the LPO in antimicrobial action has been studied thoroughly regarding the conversion of thiocyanate (SCN⁻) to hypothiocyanite ion (OSCN⁻) as antimicrobial product and some other highly reactive and short-lived oxidation products. These oxidations occur in the presence of hydrogen peroxide¹⁴.

$$SCN^- + H_2O_2 \xrightarrow{LPO} OSCN^- + H_2O_2$$

LPO is found in the salivary glands, in the breast secretory epithelial cells, lacrimal glands and in their secretions, such as saliva, milk and tears. Based on its antibacterial characteristics, currently LPO has extensive applications, including preservation of raw milk during collection or transportation to processing plants in dairy industry, the extending shelf-life of pasteurised milk and the supplementation of salivary peroxidase antimicrobial system in toothpastes and mouth rinses to reduce acid production by oral microorganisms¹⁵.

Glutathione *S*-transferases (GSTs, E.C. 2.5.1.18) catalyse the conjugation of the reduced form of glutathione (GSH) to xenobiotic substrates for the purpose of detoxification. They are multifunctional enzymes for the cellular defence against xenobiotics and provide protection for organism. They are essential and found in all kingdoms of life. GSTs catalyse the conjugation of GSH by a sulfhydryl group (-SH) to electrophilic centres on a wide variety of substrates in order to make the compounds more water soluble¹². For example, the detoxification of 1,2-epoxyethylbenzene can be given.



In this study, we investigated the inhibition effect of rosmarinic acid against human carbonic anhydrase IX and XII isoenzymes, AChE, BChE, LPO and GST enzymes.

Experimental section

Biochemical assays

An applied photophysics stopped-flow instrument was used to assay the catalytic/inhibition of four CA isozymes, as reported by Khalifah¹⁶. Briefly, phenol Red (20 mM) was used as an indicator, with an absorbance maximum of 557 nm, with HEPES (10 mM, pH 7.4) as a buffer and 0.1 M Na₂SO₄ or NaClO₄ for maintaining constant the ionic strength; these anions are not inhibitory at the used concentration. The CA-catalysed CO₂ hydration was followed for a period of 10–100 s.

For the determination of kinetic parameters and inhibition constants, the saturated CO_2 concentrations ranged from 1.7 to 17 mM. For rosmarinic acid, at least six traces of the initial 5–10% of the reaction were used for determining the initial velocity. The uncatalysed rates were determined in the same manner and subtracted from the total observed rates. Stock solutions of inhibitor (10 mM) were prepared in distilled-deionised water, and dilutions up to 0.01 μ M were performed with distilled-deionised water. Rosmarinic acid and enzyme solutions were preincubated together for 15 min at room

temperature prior to the assay to allow for the formation of the EI complex. The inhibition constant of rosmarinic acid was obtained by non-linear least-squares methods using PRISM 3, as reported earlier, and represents the mean from at least three different determinations. Both CA isozymes (CA IX and XII) were prepared in recombinant form as reported earlier¹⁷.

The inhibitory effect of rosmarinic acid on AChE/BChE activities were measured according to spectrophotometric method of Ellman et al.¹⁸ as described previously¹². Acetylthiocholine iodide (AChI) or butyrylthiocholine iodide (BChI) was used as substrates of the reaction. 5,5'-Dithio-bis(2-nitro-benzoic)acid (DTNB) was used for the measurement of the AChE/BChE activities. The hydrolysis of both substrates was monitored spectrophotometrically by formation of the yellow 5-thio-2-nitrobenzoate anion as the result of the reaction of DTNB with thiocholine, released by enzymatic hydrolysis of AChI/BChI, with absorption maximum at a wavelength of 412 nm¹⁹.

CNBr-activated-sepharose 4B was used for the purification of LPO from bovine milk²⁰. LPO purity was checked by sodium dodecyl sulphate polyacrylamide gel electrophoresis (SDS-PAGE)²¹. Protein concentration was determined according to the Bradford method as reported previously²². The stacking and running gels contained 3% (w/v) and 10% (w/v) acrylamide, respectively, and SDS (0.1%, w/v), according to a previously published procedures²³.

LPO activities were determined according to the procedure of Shindler and Bardsley²⁴. One unit of enzyme is defined as the amount of enzyme catalysing the 100 oxidation of 1 μ mol of ABTS min⁻¹ at 298 K (Molar absorption coefficient, 32.400 M⁻¹ cm⁻¹). This activity method is based on the oxidation of ABTS as a chromogenic substrate by H₂O₂, resulting in a product that absorbs at 412 nm.

GST activity was measured as according to the previous study¹². The kinetic constants of GST-catalysed reaction were determined using GSH (20 mM) or CDNB (25 mM), in phosphate buffer (pH 7.2) at room temperature. The enzyme solution replaced by phosphate buffer was used as the control.

In order to determine the effect of rosmarinic acid on enzymes, different rosmarinic acid concentrations were added into the reaction medium. enzymes activities was measured, and an experiment in the absence of rosmarinic acid was used as control²⁵. The IC₅₀ values were obtained from activity (%) versus rosmarinic acid concentration plots. To determine the K_i constant in the media with rosmarinic acid as inhibitor, different substrate concentrations were used. Rosmarinic acid solution was added into the reaction medium, resulting in three different fixed rosmarinic acid concentrations as inhibitor. Lineweaver–Burk graphs²⁶ were used to determine V_{max} and other kinetic parameters. The K_i was calculated from these graphs²⁷.

Results and discussion

Phenolic compounds are secondary plants metabolites that play an important role in the pigmentation, growth and reproduction of plants as well as plant resistance to pathogens. Also, they are biologically active substances and possess biological activities including antioxidant, antimutagenic, anticancer, anticarcinogenic, antiviral, anti-inflammatory and antibacterial activities²⁸.

Phenolic compounds are slightly acidic and have weak tendencies to lose the proton (H^+) ion from the hydroxyl group (–OH), resulting in the highly water-soluble phenolate anion. Phenols effectively inhibit CA isoenzymes. The inhibition profile of various isozymes with this class of agent is variable, with inhibition constants ranging from the millimolar to the sub-micromolar range for many simple phenols²⁹. Also, they inhibit

the CA isozymes because of the presence of different functional groups in their scaffold, mainly the phenolic –OH and acetate (–COOH) groups, which may bind to the Zn^{2+} ion or the water coordinated to the zinc ion from the CA-active site. CA isoenzymes are interesting targets for the design of pharmacological agents that are useful in the treatment or prevention of a variety of disorders, such as glaucoma, epilepsy, as diuretics or antitumor agents and diagnostic tools. The design of CAIs as therapeutic agents is related to the large number of isoforms in humans, their rather diffuse localisation in many tissues or organs. CAIs have lately emerged that CAIs could have potential as anticancer, anti-obesity and anti-infective drugs²⁹.

Phenolic and polyphenolic compounds have a lot of biological activities in fact been evaluated as CAIs by our group earlier, being shown that many such derivatives are potent inhibitors of several physiologically relevant isoforms, such as CA IX and XII^{4,6,12,17,29}. They may constitute interesting lead molecules for identifying novel CAIs. Here, we report the inhibition effect of rosmarinic acid on two catalytically active isoforms, CA IX and XII, as well as against LPO, GST, AChE and BChE. An applied photophysics stopped-flow instrument has been used for assaying the CA catalysed CO₂ hydration activity¹⁶. Rosmarinic acid has two diphenolic moieties in its scaffolds. We discovered nanomolar inhibition against some of these metabolic enzymes. The inhibition data of Rosmarinic acid are summarized in Table 1, and the following comments can be drawn from these data:

- (1) Until now, sixteen isoforms of CA have been discovered in α -CA family; among them the dimeric transmembrane glycoproteins CA IX and XII are also human-associated CA isoforms having extracellular-active site and are found in a broad spectrum of hypoxic tumour types. CA IX and XII are overexpressed in many such tumours in response to the hypoxia inducible factor pathway and research on the involvement of these isozymes in cancer has progressed significantly in recent years³⁰. CA IX showed moderate inhibition activity with rosmarinic acid, with an inhibition constant of 4.63 μ M.
- (2) It has been known that CA IX and XII are associated with tumour progression, metastasis, and response to therapy³¹. An important role of CA IX and XII as major tumour prosurvival pH-regulating enzymes was suggested in genetic study. CA XII was also poorly inhibited by rosmarinic acid, with a K_i value of 4.71 µM. Whereas, AZA, a positive standard for CA inhibition, showed an effective inhibitory activity with a K_i value 6.0 mM¹⁷. Additionally, because the CA XII is more spread in some healthy tissues than CA IX, it is less interesting target than CA IX in the cancer therapy³¹.
- (3) ACh behaves as an excitatory neurotransmitter for voluntary muscles in the somatic nervous system and as a preganglionic and a postganglionic transmitter in the parasympathetic nervous system of vertebrates and invertebrates. On the other hand, AChE is intimately associated with the normal neurotransmission by catalysing the hydrolysis of ACh to acetate and choline and acts in combination with BChE to

Table 1. Inhibition constants (K_i) of rosmarinic acid against CA IX and XII isoenzymes, AChE, BChE, LPO and GST enzymes.

Metabolic enzymes	$K_{\rm i}$ values
Lactoperoxidase (LPO)	0.51 µM
Human carbonic anhydrase (hCA IX)	4.63 µM
Human carbonic anhydrase (hCA XII)	4.71 µM
Acetylcholinesterase (AChE)	42.52 pM
Glutathione S-transferase (GST)	48.74 nM
Butyrylcholinesterase (BChE)	121.60 pM

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remove ACh from the synaptic cleft^{6,32}. Selective inhibition of AChE may cause accumulation of ACh in the synaptic cleft, leading to overstimulation and the disruption of nerve impulses and ultimately causing symptoms such as ataxia, central respiratory paralysis, seizures, coma and death. Inhibition of AChE activity was determined on commercially available purified AChE from the electric organ of E. electricus L. based on the method of Ellman method¹⁸. This assay has been used most widely for the detection of AChE and BChE inhibitors and for the quantification of a ChE inhibitory activity. AChE inhibition has generally been used to determine the neurotoxic properties of chemicals or drugs capable of interfering with normal neurotransmission of the sympathetic and parasympathetic nervous system of living organisms. Most AChE inhibitors (ChEIs) interact with the catalytic site of the AChE located at the bottom of the gorge, where the hydrolysis of ACh takes place¹⁹. Currently, the most prescribed ChEIs are donepezil, galantamine and rivastigmine. These drugs are used to treat patients with mild-to-moderate AD. BChE has a specific role in cholinergic neurotransmission and it has been associated with AD²¹. Individual ChEIs differ from each other with respect to their pharmacologic properties. Primary target of donepezil and galantamine is AChE; however, rivastigmine shows equal affinity for both AChE and BChE enzymes. It was also shown that the main AChE inhibitory effect was primarily associated with aromatic compounds and, to a lesser degree, with aliphatic compounds. AChE was very effectively inhibited by rosmarinic acid, with K_i value of 42.52 pM (Table 1). On the other hand, rosmarinic acid inhibited BChE with K_i value of 121.60 pM. The K_i values of rosmarinic acid for AChE and BChE were calculated from Lineweaver-Burk plots²⁶. On the other hand, donepezil hydrochloride, which is used for the treatment of mild-to-moderate AD and various other memory impairments, had been shown to lower AChE inhibition activity $(IC_{50}: 55.0 \text{ nM})^{12}$.

- (4) Milk contains a variety of constituents that protect the neonate and the milk itself from a host of deleterious microorganisms. One such constituent is LPO, which plays an important role in protecting the lactating mammary gland and the intestinal tract of new-borns against pathogenic microorganisms. Our investigation showed that rosmarinic acid moderately inhibited LPO with a K_i value of 0.51 μ M.
- (5) Recently, many studies demonstrated that GST plays important functions in cellular defence against chemical toxicity. It was reported a link between the lack of GST enzyme activity and the susceptibility to develop different types of cancer including oral, gastric and bladder cancers¹². GST was potently inhibited by rosmarinic acid, with a K_i value of 48.74 nM (Table 1). It was reported that the inhibitor of GST bearing suitable linkers could concomitantly bind to two active sites of GST and usually possess K_i values at nanomolar levels and excellent enzyme selectivity¹².

Conclusion

Rosmarinic acid is a caffeic acid ester of salvianic acid A (3,4dihydroxyphenyllactic acid). To effect of rosmarinic acid against CA IX and XII isoenzymes, AChE, BChE, LPO and GST was evaluated in this study. Rosmarinic acid showed low micromolar inhibition against both CA isoenzymes (CA IX and XII) and LPO, nanomolar inhibition against GST and picomolar inhibition against ACE and BChE. The results showed that rosmarinic acid moderately inhibited both CA isoenzymes, but effectively inhibited by GST, ACE and BChE. The catalytic inhibition mechanisms of CA isoenzymes for the physiologic reaction is well-understood processes: most types of classical inhibitors bind to the metal centre, but the discovery of new classes of CAIs possessing different inhibition mechanisms compared to the classical inhibitors. Also, the other metabolic enzymes (AChE, BChE, LPO and GST) have diverse inhibition profiles. These data may explain the beneficial health effects of Rosmarinic acid and may lead to enzyme researchers and drug design campaigns.

Declaration of interest

The authors have declared no conflict of interest.

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