

1 **First Report of Asiatic Brown rot (*Monilinia polystroma*) and Brown rot**
2 **(*Monilinia fructicola*) on Pears in Italy**

3 **C. Martini^a, A. Di Francesco^a, A. Lantos^b, and M. Mari^a**

4 ^aCriof, DipSa, University of Bologna, Via Gandolfi, 19, 40057, Cadriano, Bologna, Italy.

5 ^bFaculty of Horticultural Science, Department of Plant Pathology, Corvinus University of
6 Budapest, Ménesi Road 44, 1118 Budapest, Hungary.

7
8 *E mail:camilla.martini2@unibo.it
9

10 Worldwide, brown rot caused by *Monilinia* spp. is an important fruit postharvest decay causing
11 severe losses in stone and pome fruits with a significant economic impact. In Italy three *Monilinia*
12 species *M. laxa*, *M. fructicola* and *M. fructigena* are the causal agents of blossom and twig blight,
13 and brown fruit rot in stone fruit. *M. polystroma* has been observed on peaches in Italy (Martini C.
14 et al. 2014), and has been reported in Czech Republic and Hungary (Petroczy M. et al. 2012),
15 Poland (Poniatowska A. et al. 2013), Serbia (Vasic M. et al. 2013), and Switzerland (Hilber-
16 Bodmer M. et al. 2012) on pome fruits and apricots. In September 2013, during a survey for fungal
17 postharvest pathogens, stored 'Abate Fetel' pears showing brown rot symptoms were observed in
18 Emilia Romagna region. In the 20% of symptomatic pears, circular and brown to black decay spots
19 were observed; these spots were covered by a large number of yellowish or buff-colored stromata
20 while decayed tissues remained firm. Overall, these symptoms resembled those originated by *M.*
21 *polystroma*. In other 13% of stored pears the decayed tissues remained firm and. These decay
22 lesions were covered with numerous grayish pustules containing spores. Putative pathogens were
23 isolated on Potato Dextrose agar (PDA) and incubated at 25°C in darkness for 5 days. The colonies
24 grown on PDA were yellowish in color, with irregular black stromatal crusts at the edges of the
25 colonies after 10-12 days of incubation. Some colonies developed sporogenous tissue was slightly
26 elevated above the colony surface, color buff/pale luteous, and was present at the margin of
27 colonies (Poniatowska A. et al. 2013). Conidia developing from such cultures were one-celled,
28 ovoid or limoniform, smooth and hyaline, measuring 12.2-20.4 x 8.4-12.3µm when grown on V8
29 Juice agar (V8) at 22°C and matched the description of those for *M. polystroma*. Other colonies
30 developed a gray mass of spores in concentric rings with the reverse side black were
31 morphologically identified as *M. fructicola*. The colony margins were smooth edged and the conidia
32 were one-celled, limoniform, hyaline, and measuring 12.1 to 17.4 x 8.1 to 11.2 µm when grown on
33 V8 Juice agar (V8) at 22°C. The identification of the isolates was obtained using the universal
34 primers for *Monilinia* spp. designed by Petroczy et al. 2012. Pathogenicity was confirmed using
35 surface-sterilized mature 'Abate Fetel' and 'William' pears wounded with a sterile needle, and

1 inoculated with 20 μ l of a *M. polystroma* or *M. fructicola* conidial suspension (10^3 spores/ml).
2 Control pears received sterile water and each treatment contained ten fruit. After seven days of
3 incubation at 20°C in plastic containers with high humidity, typical symptoms of Asiatic brown rot
4 or brown rot developed on both the wounds of all inoculated pears, while controls remained
5 symptomless. Mean colony diameters measured after 7 days were 47.3 mm for Asiatic brown rot
6 and 44.1 mm for brown rot, and there were no significant differences in colony diameter after
7 seven days between *M. polystroma* and *M. fructicola* ($\alpha < 0.05$).

8 After 14 days yellowish exogenous stromata appeared on the surface of pears infected *M.*
9 *polystroma*, whereas numerous grayish pustules containing spores appeared on pears inoculated
10 with *M. fructicola*. Control pears still remained symptomless. The fungus isolated from inoculated
11 fruit exhibited the same morphological features of the original isolates, and PCR/sequencing
12 analysis using primers ITS1 and ITS4 to amplify ribosomal ITS1-5.8S-ITS2 region confirmed the
13 molecular results with primers by Petroczy et al. 2012 (GenBank Accession Nos. GU067539.1 and
14 HQ893748.1). Although the presence of *M. polystroma* and *M. fructicola* has been already
15 documented in Italy, this is the first time these two species are observed on Italian pears. This report
16 suggests a broader impact since *M. polystroma* and *M. fructicola* have not been previously reported
17 on pears in Europe. Because of the importance of pears in Italian fruit industry, knowledge about
18 the occurrence of new pathogens will facilitate the adoption of adequate control strategies in order
19 to reduce postharvest losses.

20 **References**

- 21 (1) Hilber-Bodmer M. et al. *Plant Dis.*91: 146, 2012. (2) Martini C. et al. *Plant Dis* 98: 1585,
22 2014. (3) Petroczy M. et al. *Trees* 26:153-164, 2012. (4) Poniatowska A. et al. *Eur J Plant*
23 *Pathol*135:855-865, 2013. Vasic M. et al. *Plant Dis.*97: 145, 2013.