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1 **Bioactive and antioxidant activity from *Citrus***
2 ***Bergamia* RISSO (BERGAMOT) juice collected in**
3 **different areas of Reggio Calabria province, Italy**

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12 **Abstract**

13 The chemical composition and antioxidant activity of juice extracted from seven samples of
14 bergamot (*Citrus bergamia* Risso) collected in different areas of Reggio Calabria Province were
15 investigated. The ascorbic acid, total polyphenol, and flavonoid contents were determined. Total
16 flavonoids and polyphenols were analyzed by UV spectra, while flavanone content was analyzed
17 by HPLC. The antioxidant activity of the fractions was assessed using three representative
18 assays: ABTS^{•+}, DPPH[•] radical quenching and β -Carotene bleaching test. The main flavanones
19 were naringin, neohesperedin and neoeriocitrin, and their average content 242.4±1.8, 183.0±0.6
20 and 247.0±1.4 mg mL⁻¹ respectively. The results showed that bergamot juice possessed a good
21 quality and a valuable source of health promoting constituents. In fact it contained eriocitrin,
22 naringin, neoeriocitrin, and neohesperedin, which may contribute differentially to the antioxidant
23 capacity.

24 INTRODUCTION

25 Bergamot (*Citrus bergamia* Risso) is thought to be a hybrid of the sour orange and citron or
26 lemon. The fruit is spherical like an orange but yellow like a lemon. It has been known in the
27 Mediterranean for several centuries and was described as early as 1708. Production is mostly
28 limited to the Ionian coastal areas of the province of Reggio di Calabria (Italy). Bergamot is a
29 symbol of the entire province, which grows about 90% of world production of this citrus fruit.
30 This fruit is also cultivated in Côte d'Ivoire. However the quality of the obtained essence is not
31 comparable due to the argillite, limestone and alluvial deposits found there.^[1] There are three
32 bergamot cultivars: Fantastico, Castagnaro and Femminello. Fantastico cultivar represents the
33 most representative (90%).^[2]

34 Citrus derivatives are an important field of food technology. However, even today, the
35 technological processes often provide only partial extraction of all possible derivatives. As
36 regards, in particular, bergamot, the aim of the producer is limited almost exclusively to the
37 extraction of the essential oil; the juice and peel are used only occasionally, although they could
38 be a source of profit for growers.

39 The juice is generally considered a waste product, which represents a serious environmental and
40 economic problem for the industries. Most studies have focused on its essential oil as this
41 constitutes a raw material for the perfume and food industries.^[3-4] However, the presence of
42 neoriesperidin, naringin, and neoeriocitrin in its juice has also been reported.^[5-7]

43 Bergamot juice and albedo are rich in neoeriocitrin, neohesperidin, naringin, rutin, neodesmin
44 and rhoifolin. The profile of the flavonoids in fruits of bergamot differs from other Citrus not
45 only in terms of quality but also quantity.[8,9] Flavonoids are specially known for their
46 antioxidant activities,[10- 12] which play a significant role in cardiovascular health and in
47 prevention of cancer.[13-15] Other than fighting free radicals, they are also known for their
48 antihistamine, antimicrobial, memory enhancing and even mood-boosting properties. In recent
49 years, the beneficial properties of bergamot juice have been generating interest, and have been
50 the subject of several studies.[16-20] Some researchers have focused on the molecular
51 mechanisms of the bioactive compounds contained in the bergamot juice. The obtained results
52 suggest that the flavonoids of bergamot juice may be useful for the development of alternative
53 pharmacological strategies aimed at reducing the inflammatory process. [21-23]

54 Several studies have shown that flavonoids present in bergamot juice, lower cholesterol levels by
55 modulating hepatic HMG-CoA levels.[24-26] Recently a research group of University of
56 Catanzaro (Italy) has shown that administration of bergamot juice leads to a significant reduction
57 in serum cholesterol and triglycerides. Moreover, the flavonoids present in the juice of this citrus
58 fruit have a lipid-lowering action. Its effectiveness was tested on patients with
59 hypercholesterolemia with and without type 2 diabetes, and on rats who also had an altered lipid
60 profile. Intake of bergamot juice twenty minutes before meals helped reduce both cholesterol and
61 triglycerides in patients. Moreover, none of the participants in the trial showed intolerance or
62 side effects after taking the bergamot juice.[27] An analytical differentiation of Citrus juices,
63 based on the differing concentrations of certain minor components, including flavonoids, plays
64 an important role in determining chemotaxonomic markers to ascertain the authenticity of

65 commercial products.^[28] The aim of the present paper was to evaluate the chemical composition
66 and antioxidant activity in juice obtained from bergamot fruits collected Catona and Africo in a
67 90 km coastal stretch of the province of Reggio Calabria, Southern Italy.

68 MATERIALS AND METHODS

69 Plant Material and Juice Extraction

70 Citrus bergamia Risso fruits cv Fantastico were collected in February 2014 from plantations
71 located between Catona and Africo (Reggio Calabria, Italy). Plantations were chosen in seven
72 areas of the coast where bergamot is cultivated: Catona, Gallico, Arangea, Pellaro, Africo, Melito
73 Porto Salvo, Palizzi (Fig. 1). All plantations were made up of plants aged 15-20 years and
74 irrigated by sprinkler. Fruits were harvested at maturity stage and at full fruit size from a random
75 sample of 15 plants in order to obtain a set of fruits that are representative. Plants received
76 similar water and fertilizer treatments. Fruits were examined for integrity and absence of dust
77 and insect contamination. Immediately after harvesting, the juice was extracted by mechanical
78 pressure and frozen at -80°C until analyzed.

79 Reagent

80 Hesperidin, narirutin, naringin, neohesperidin, neoeriocitrin, linoleic acid and β -carotene were
81 purchased from Sigma Chemical Company (Milan, Italy). Ascorbic acid, Folin-Ciocalteu reagent
82 and 2,2-diphenyl-1-picrylhydrazyl radical (DPPH) and 2,2'-azinobis (3-ethyl-benzothiazoline-6-

83 sulphonate) (ABTS) were supplied by Carlo Erba (Milan, Italy). Solvents and reagents not
84 expressly specified had a high degree of purity and were supplied by Carlo Erba (Milan, Italy).

85 Physicochemical analysis

86 The bergamot were squeezed and the juice was centrifuged and filtered to determine the
87 following analyses: color of fresh juice was measured at 25°C using a Konica Minolta CM-
88 700/600d spectrophotometer (Konica Minolta Sensing, Japan). Data were expressed as L*
89 (lightness/darkness in a range 0-100), a* (greenness/redness in a range between -60 and + 60)
90 and b* (blueness/yellowness in a range between – 60 and + 60). Total soluble solids (TSS) were
91 determined using a digital refractometer PR-201 α (Atago, Tokyo, Japan), previously calibrated
92 at 20°C and the results expressed as degrees Brix; The pH was measured at ambient temperature
93 with a pH meter (Model Basic 20, Crison) previously calibrated with standard solutions pH 4 and
94 pH 7; Total acidity (TA) was determined using the International Federation of Fruit Juice
95 producers test^[29]: a potentiometric titration of the acidity of the juice, with a solution of 0.25 N
96 NaOH up to pH 8.1. The results were expressed as g L⁻¹ of anhydrous and hydrate citric acid.
97 Ascorbic acid was determined using the International Federation of Fruit Juice producers test:^[29]
98 a potentiometric titration of the acidity of the juice, with a solution of 2,6-dichloroindophenol.
99 Total flavanones were determined with a colorimetric method using alkaline diethylene glycol
100 for determination of the bitter rhamnoglycoside naringin and other flavanones that may be
101 present in citrus fruits.^[30] The bergamot juice was analyzed for total phenolics by the Folin-
102 Ciocalteu (FC) colorimetric method.^[31]

103 All determinations above described were made in triplicate.

104 **Flavanone analysis**

105 Flavanone glycosides, expressed as hesperidin equivalents (mg L^{-1}), were determined by liquid
106 chromatograph according to the official methodologies of the International Federation of Fruit
107 Juice producers.^[29] Fresh bergamot juice previously centrifuged at 4000 rpm for 20 minutes, was
108 filtered through a 0.45 μm membrane filter. Separation of flavanones was performed by HPLC
109 using a Phenomenex C18 column (150mm x 3 mm). The solvent system was buffer solution A:
110 ACN/H₂O/H₂PO₄ (70:26:4) and buffer solution B: KH₂PO₄ pH 3.5, with flow-rate 500 $\mu\text{L}/\text{min}$ in
111 gradient conditions. The analysis was monitored at 287 nm. The gradient program was as
112 follows: starting condition, 85% A, 15% B; 5 min, 70% A, 30% B; 20 min, 50% A, 50% B; 30
113 min, 25% A, 75% B; 35 min, 5% A, 95% B; 40 min, 85% A, 15% B. The column was operated
114 at 25°C and flow rate was 1 mL min^{-1} . Identification of compounds was performed by comparing
115 their retention time with those of standards and confirmed with characteristic spectra using the
116 photodiode array detector.

117 Flavanone glycoside evaluation was calculated according to the external standard method by
118 integration of the peak areas of individual compounds to that of standard curve prepared using
119 hesperidin standard.

120 **Antioxidant activity**

121 **DPPH assay:** This experimental procedure was described by Loizzo et al. ^[32] In an ethanol
122 solution of DPPH radical (final concentration was 1.0×10^{-4} M), samples at different
123 concentrations were added. The reaction mixtures were shaken and kept in the dark for 30 min.

124 The absorbance of the resulting solutions was measured in 1 cm cuvettes using a Perkin Elmer
125 Lambda 40 UV/VIS spectrophotometer at $\lambda = 517$ nm against blank without DPPH. A decrease of
126 DPPH solution absorbance indicates an increase of DPPH radical scavenging activity. This
127 activity is given as % DPPH radical-scavenging that is calculated in the equation:

128 % DPPH radical-scavenging = $[1 - (\text{sample absorbance with DPPH} - \text{sample absorbance without}$
129 $\text{DPPH} / \text{control absorbance})] \times 100$

130 **ABTS assay:** ABTS assay was based on the method previously described by Loizzo et al. ^[32]
131 with slight modifications. ABTS radical cation (ABTS^+) was produced by the reaction of a 7 mM
132 ABTS solution with 2.45 mM potassium persulphate. The mixture was stored in the dark at room
133 temperature for 12 h before use. The ABTS^+ solution was diluted with ethanol to an absorbance
134 of 0.70 ± 0.05 at $\lambda = 734$ nm. After addition of 25 μL of sample or Trolox standard to 2 mL of
135 diluted ABTS^+ solution, absorbance was measured at exactly 6 min after mixing. Appropriate
136 solvent blanks were run in each assay. The scavenging ability of the sample was calculated
137 according to the following equation: $\text{ABTS scavenging activity (\%)} = [(A_0 - A) / A_0] \times 100$
138 where A_0 is the absorbance of the control reaction and A is the absorbance in the presence of
139 samples.

140 **β -Carotene Bleaching Test:** Antioxidant activity was determined using β -carotene bleaching as
141 previously described by Menichini et al. ^[33] Briefly, β -carotene solution was added to linoleic
142 acid and 100% Tween 20. The emulsion was mixed with of samples at different concentrations
143 and tubes were placed at 45°C in a water bath for 60 min. Propyl gallate was used as standard.

144 The absorbance of the samples, standard and control was measured at 470 nm against a blank at t
145 = 0 and successively at 30 and 60 min.

146 **Statistical analysis**

147 All experiments were carried out in triplicate. Data were expressed as means \pm standard
148 deviation (S.D.). Analysis of variance (one-way ANOVA) was conducted using SPSS version
149 17.0 for Windows (SPSS Inc., Chicago, IL) and the Tukey's test was used to determine any
150 significant difference among all treatments at $P < 0.05$. The concentration giving 50% inhibition
151 (IC_{50}) was calculated by nonlinear regression with the use of Prism GraphPad Prism version 4.0
152 for Windows (GraphPad Software, San Diego, CA, USA). The dose-response curve was
153 obtained by plotting the percentage inhibition versus concentration. Differences within and
154 between groups were evaluated by one-way analysis of variance test (ANOVA) followed by a
155 multicomparison Dunnett's test compared with the positive controls.

156 **RESULTS AND DISCUSSION**

157 The bergamot juice extracted from fruits collected in seven different areas of the coast of the
158 province of Reggio Calabria (Italy), were examined in order to determine their antioxidant
159 activity as related to their phenolic composition. Table 1 shows the areas where the fruits were
160 collected with their latitude and longitude. There were significant differences ($P < 0.05$) in all
161 color attributes among samples studied (Table 2). With respect to lightness (L^*), the lowest value

162 corresponded to the Melito, Catona and Palizzi samples, while the highest values were those
163 samples from Gallico, Arangea and Pellaro.

164 Table 3 shows the values of the chemical-physical characteristics (pH, titratable acidity, soluble
165 solid and formol index) of the bergamot juice. The pH values of the juices analyzed are within
166 normal range (2.54-2.84) and the differences between them are significant ($P < 0.05$). The lowest
167 value was in the Africo sample and the highest (pH 2.84) in the Catona sample. Titratable acidity
168 was significantly different: values were in the range 11.38-14.83. Soluble solid content measured
169 for bergamot juice was in the range 8.11-11.61 °Brix. The formol index was used to estimate the
170 total content of amino acids in a juice, and partly to estimate its purity. In the seven juices
171 analyzed the values obtained show that there are significant differences.

172 Table 4 shows that the ascorbic acid, total flavonoid and total polyphenol content in all treated
173 samples was significantly different ($P < 0.05$). Vitamin C (ascorbic acid), flavonoid and phenolic
174 compounds are essential components for human health and have a high antioxidant activity,
175 providing protection against free radicals and consequently participating in the prevention of
176 many degenerative diseases. Ascorbic acid content was in the range 89.40-285.35 mg 100 mL⁻¹.
177 The Gallico sample was significantly lower (89.4±2.1 mg 100 mL⁻¹) than that of other samples.
178 Similarly, the total flavonoid content showed the lowest value for the Gallico and Palizzi
179 samples (51.1±2.2 mg 100 mL⁻¹ and 74.0±2.1 mg 100 mL⁻¹, respectively). The highest content of
180 total flavonoids was found in the samples from Arangea and Pellaro (148.2±3.4 mg 100 mL⁻¹
181 and 147.8±4.6 mg 100 mL⁻¹, respectively). With regard to total polyphenols significant
182 differences in the various samples analyzed were observed, their content being between

183 180.5±2.9 mg 100 mL⁻¹ and 233.4±0.5 mg 100 mL⁻¹. The Catona sample was significantly lower
184 (180.5±2.9 mg 100 mL⁻¹) compared to the other samples.

185 Table 5 shows the data collected for the flavonoid content of bergamot juice. The most abundant
186 components in bergamot juices are naringin, neoeriocitrin and neohesperedin, as reported by
187 Gionfriddo et al.;[5] Kawaii et al.; [6] Calabrò et al.;[34] Dugo et al.;[8] Gattuso et al.;[35]
188 Nogata. [9]

189 The lowest values of flavonoids identified by HPLC on the basis of retention times and
190 quantified from the peak areas compared to standards, are those collected from the areas of
191 Gallico and Palizzi.

192 Reactive oxygen species (ROS) are closely related to many pathological conditions such as
193 inflammation, tumours, cardiovascular disease, cerebral ischemia and diabetes. DPPH· and
194 ABTS· are stable free radicals, which have been widely accepted as a tool for estimating free
195 radical scavenging activities of antioxidants.^[36] All samples exhibited a radical scavenging
196 activity against both radicals in a concentration-dependent manner (Table 6). The area of
197 collection influenced the DPPH radical scavenging activity with a range of IC₅₀ values from
198 19.6±2.0 to 31.4±1.1 µg mL⁻¹ (Melito and Palizzi samples, respectively). The sample was also
199 Melito the most effective also in the ABTS test with IC₅₀ value of 17.4±1.6 µg mL⁻¹ followed by
200 the Africo sample (IC₅₀ value of 18.9±2.4 µg mL⁻¹). Correlation analysis revealed that the DPPH
201 assay is positively correlated with total phenol and flavonoid content. However total phenols
202 positively correlated also with ABTS data. The potential of bergamot juice to inhibit lipid
203 peroxidation was evaluated using the β-carotene/linoleic acid bleaching test, which measures the

204 capacity for inhibiting conjugated diene hydroperoxide formation during linoleic acid oxidation.
205 Even in this case the Melito was sample the most active (IC_{50} values of 25.7 ± 1.2 and 24.9 ± 3.0
206 $\mu\text{g mL}^{-1}$ at 30 and 60 minutes of incubation). The correlation analysis revealed that data obtained
207 from this assay are positively correlated with vitamin C and total flavonoids, which are mainly
208 responsible for antioxidant activity. A significant positive correlation was also observed with
209 eriocitrin, naringin, neohesperidin and neohesperidin content. Previously Gardner et al.^[37]
210 reported that vitamin C was found to account for 65-100% of the antioxidant potential of *Citrus*
211 juice.

212 The antioxidant potential of *C. bergamia* juice has been previously investigated by Trovato et
213 al.^[38] that found a noticeable effect on scavenging DPPH radicals with IC_{50} value of $25.01 \mu\text{L}$.
214 Xu et al.^[39] studied the total phenolic content and antioxidant activities of fifteen *Citrus* variety
215 juices. The total phenolic content ranged from 751.82 to 1555.49 mg L^{-1} for *C. limon* and hybrid
216 439 (*C. reticulata* x *C. sinensis*), respectively. It was interesting that Hybrid 439 achieved the
217 highest DPPH inhibitory activity with a percentage of inhibition of 61.62% followed by *C.*
218 *sinensis* Osbeck cv Hamlin and cv Liubencheng that exhibited a DPPH scavenging ability of
219 60.24 and 60.13%. Interestingly in *Citrus* investigated by Xu et al.^[39] neohesperidin was found
220 only in two varieties.

221 CONCLUSIONS

222 For all the studied characteristics there were statistically significant differences ($P < 0.05$)
223 between juices analyzed. The purpose of this thesis was to see how the microclimate of a given

224 area of cultivation may influence the quality and, therefore, the chemical composition of the
225 juice itself. Results obtained in this study are a contribution to the characterization of bergamot
226 fruits (*Citrus bergamia* Risso) cultivated in different areas of the province of Reggio Calabria
227 (Italy).

228 REFERENCES

229 Mandalari, G.; Bennett, R.B.; Bisignano, G.; Saija, A.; Dugo, G.; Lo Curto, R.B.; Faulds, C.B.;
230 Waldron, K.W. Characterization of Bergamot (*Citrus bergamia* Risso) Pulp, a Major Co-product
231 of Essential Oil Extraction. *Journal of Agricultural and Food Chemistry* **2006**, 54, 197-203.

232 Dugo, G.; Lamonica, G.; Cotroneo, A.; Trozzi, A.; Crispo, F.; Licandro, G.; Giofrè, D. La
233 composizione della frazione volatile dell'essenza di bergamotto. *Stazione sperimentale per*
234 *l'industria delle essenze e dei derivati agrumari Reggio Calabria* (1987). Collana di monografie
235 sugli oli essenziali e sui derivati agrumari.

236 Ricci H.; Rovesti, P. Impieghi cosmetologici dell'olio essenziale di bergamotto. *EPPOS* **1979**,
237 61:18.

238 Poiana, M.; Reverchon, E.; Sicari, V.; Mincione, B.; Crispo, F. Supercritical carbon dioxide
239 extraction of bergamot oil. Bergapten content in the extracts. *Italian Journal of Food Science*
240 **1994**, 4, 59–66.

241 Gionfriddo, F.; Postorino, E.; Bovalo, F. I flavanoni glucosidici nel succo di bergamotto. *Essenza*
242 *e Derivati Agrumari* **1996**, 4, 404–416.

243 Kawaii, S.; Tomono, Y.; Katase, E.; Ogawa, K.; Yano, M. Quantitation of flavonoid
244 constituents in Citrus fruits. *Journal of Agricultural and Food Chemistry* **1999**, 47, 3565–3571.

245 Pernice, R.; Borriello, G.; Ferracane, R.; Borrelli, RC.; Cennamo, F.; Ritieni, A. Bergamot: A
246 source of natural antioxidants for functionalized fruit juices. *Food Chemistry* **2009**, 112, 545–
247 550.

248 Dugo, P.; Lo Presti, M.; Ohman, M.; Fazio, A.; Dugo, G.; Mondello, L. Determination of
249 flavonoids in citrus juices by micro-HPLC-ESI-MS. *Journal of Separation Science* **2005**, 28,
250 1149-1156.

251 Nogata, Y.; Sakamoto, K.; Schiratsuchi, H.; Ishii, T.; Yano, M.; Ohta, H. Flavonoid composition
252 of fruit tissues of citrus species. *Bioscience Biotechnology and Biochemistry* **2006**, 70, 178-192.

253 Kumar, S.; Pandey, A.K. Chemistry and biological activities of flavonoids: An Overview.
254 *Scientific World Journal*. **2013**

255 Wong, J.Y.; Matanjun, P.; Ooi Y.B.H.; Chia, K.F. Evaluation of antioxidant activities in relation
256 to total phenolics and flavonoids content of selected Malaysian wild edible plants by multivariate
257 analysis. *International Journal of Food Properties* **2014**, 17, 1763-1778.

258 Li, L.; Wang, S.; Chen, J.; Xie, J.; Wu, H.; Zhan, R.; Li, W. Major antioxidants and in vitro
259 antioxidant capacity of eleven mango (*Mangifera Indica* L.) cultivars. *International Journal of*
260 *Food Properties* **2014**, 17, 1872-1887.

261 Hollman, P.C.; Katan, M.B. Dietary flavonoids: intake, health effects and bioavailability. *Food*
262 *Chemistry Toxicology* **1999**, 37, 937-942.

263 Navarra, M.R.; Ursino, N.; Ferlazzo, M.; Schumacher, U.; Valentiner, U. Effect of *Citrus*
264 *bergamia* juice on human neuroblastoma cells *in vitro* and in metastatic xenograft models.
265 *Fitoterapia* **2014**, 95, 83 – 92.

266 Delle Monache, S.; Sanità, P.; Trapasso, E.; Ursino, M.R.; Dugo, P.; Russo, M.; Ferlazzo, N.;
267 Calapai, G.; Angelucci, G.; Navarra, M. “Mechanisms underlying the anti-tumoral effects of
268 *bergamia* juice,” *PLoS ONE* **2013**, vol. 8. no. 4.

269 Barbera, N.; Pendino, F. Bergamot symposium (Reggio Calabria, November 30–December 2)
270 1998.

271 Matsumoto, H.; Ikoma, Y.; Sugiura, M.; Yano, M.; Hasegawa, Y. Identification and
272 quantification of the conjugated metabolites derived from orally administered hesperidin in rat
273 plasma. *Journal of Agricultural and Food Chemistry* **2004**, 52:6653-6659.

274 Picerno, P.; Sansone, F.; Mencherini, T.; Prota, L.; Aquino, R.P.; Rastrelli, L.; Lauro, M.R.
275 *Citrus bergamia* juice: phytochemical and technological studies. *Natural Product*
276 *Communications* **2011**, 6, 951-955.

277 Ferlazzo, N.; Visalli, G.; Smeriglio, A.; Cirmi, S.; Lombardo, G.E.; Campiglia, P.; Di Pietro, A.;
278 Navarra, M. Flavonoid fraction of orange and bergamot juices protect human lung epithelial cells
279 from hydrogen peroxide-induced oxidative stress. *Evid Based Complement Alternat Med.* **2015**,
280 Published online 2015 Jun 21. doi: 10.1155/2015/957031

281 Ursino, M.; Mondello, M.R.; Sdrafkakis, V.; Campolo, L., Taviano, M.F.; Miceli, N. Attività
282 citoprotettiva del succo di citrus *bergamia* Risso & Poiteau sull'apparato gastrointestinale.
283 *Fitomed* 2006 (Taormina, July 6–8).

284 Impellizzeri, D.; Bruschetta, G.; Di Paola R.; Ahmad, A.; Campolo, M.; Cuzzocrea, S.; Esposito,
285 E.; Navarra, M. “The anti-inflammatory and antioxidant effects of bergamot juice extract (BJe)
286 in an experimental model of inflammatory bowel disease,” *Clinical Nutrition* **2014**, n. 27.

287 Sommella, E.; Pepe, G.; Pagano, F.; Tenore, G.; Marzocco, S.; Manfra, M.; Calabrese, G.;
288 Aquino, R.P.; Campiglia, P. UHPLC profiling and effects on LPS-stimulated J774A.1
289 macrophages of flavonoids from bergamot (*Citrus bergamia*) juice, an underestimated waste
290 product with high anti-inflammatory potential. *Journal of Functional Foods* 2014, 7, 641–649.

291 Risitano, R.; Currò, M.; Cirimi, S.; Ferlazzo, N.; Campiglia, P.; Caccamo, D.; Ientile,
292 R.; Navarra, M. “Flavonoid fraction of bergamot juice reduces LPS-induced inflammatory
293 response through SIRT1-mediated NF-kappaB inhibition in THP-1 monocytes,” *PLoS ONE*,
294 2014, vol. 9, no. 9.

295 Terpstra, A.H.; Lapre, J.A.; De Vries, H.T.; Beynen, A.C. Dietary pectin with high viscosity
296 lowers plasma and liver cholesterol concentration and plasma cholesteryl ester transfer protein
297 activity in hamsters. *Journal of Nutrition* **1998**, 128, 1944–9.

298 Kim, H.K.; Jeong, T.S.; Lee, M.K.; Park, Y.B.; Choi, M.S. Lipid-lowering efficacy of hesperetin
299 metabolites in high-cholesterol fed rats. *Clin. Chim. Acta* **2000**, 3327, 129-137.

300 Marounek, M.; Volek, Z.; Synytsya, A.; Copikova, J. Effect of pectin and amidated pectin on
301 cholesterol homeostasis and cecal metabolism in rats fed a high-cholesterol diet. *Physiological*
302 *Research* **2007**, 56, 433–42.

- 303 Mollace, V.; Sacco, I.; Janda, E.; Malara, C., Ventrice, D.; Colica, C.; Visalli, V.; Muscoli, S.;
- 304 Ragusa, S.; Muscoli, C.; Rotiroti, D.; Romeo, F. Hypolipemic and hypoglycaemic activity of
- 305 bergamot polyphenols: from animal models to human studies. *Fitoterapia* **2011**, 82, 309-16.
- 306 Hammond, D.A. Authenticity of fruit juices, jams and preserves. P.R. Ashurst, M.J. Dennis
- 307 (Eds.), *Food authentication*, Chapman & Hall, London, UK (1996), pp. 32–36.
- 308 IFU (3-17-58) International Federation of Fruit Juice Producers. *Methods of Analysis and*
- 309 *Microbiological Methods*. <http://www.ifu-fruitjuice.com/ifu-methods>.
- 310 Davis, W.B. Determination of flavanones in citrus fruits. *J. Ind. Eng. Chem.* **1947**, 19, 476.
- 311 Singleton, V.L.; Orthofer, R.; Lamanuela-Raventòs, R.M. Analysis of total phenols and other
- 312 oxidation substrates and antioxidants by means of folin-ciocalteu reagent. *Methods in*
- 313 *enzymology* **01/1999**; 299C:152-178.
- 314 Loizzo, M.R.; Tundis, R.; Bonesi, M.; Menichini, F.; Mastellone, V.; Avallone, L.; Menichini, F.
- 315 Radical scavenging, antioxidant and metal chelating activities of *Annona cherimola* Mill.
- 316 (cherimoya) peel and pulp in relation to their total phenolic and total flavonoid contents. *Journal*
- 317 *of Food Composition and Analysis* **2012**, 25, 179-184.
- 318 Menichini, F.; Tundis, R.; Bonesi, M.; Loizzo, M.R.; Conforti, F.; Statti, G.; De Cindio, B.;
- 319 Houghton, P.J.; Menichini, F. The influence of fruit ripening on the phytochemical content and
- 320 biological activity of *Capsicum chinense* Jacq. cultivar Habanero. *Food Chemistry* **2009**, 114,
- 321 553-560.
- 322 Calabro, M.L., Galtieri, V.; Cutroneo, P.; Tommasini, S.; Ficarra, P.; Ficarra, R. Study of the
- 323 extraction procedure by experimental design and validation of a LC method for determination of

324 flavonoids in *Citrus bergamia* juice. Journal of Pharmacological and Biomedical Analysis **2004**,
325 35, 349-363.

326 Gattuso, G.; Caristi, C.; Gargiulli, C.; Bellocco, E.; Toscano, G.; Leuzzi, U. Flavonoid
327 Glycosides in Bergamot Juice (*Citrus bergamia* Risso). Journal of Agricultural and Food
328 Chemistry **2006**, 54, 3929–3935.

329 Krishnaiah, D.; Sarbatly, R.; Nithyanandam, R. A review of the antioxidant potential of
330 medicinal plant species. Food and Bioproducts Processing **2010**, 89, 217-233.

331 Gardner, P.T.; White, T.A.C.; McPhail, D.B.; Duthie, G.G. The relative contributions of vitamin
332 C, carotenoids and phenolics to the antioxidant potential of fruit juices. Food Chemistry **2000**,
333 68, 471-474.

334 Trovato, A.; Taviano, M.F.; Pergolizzi, S.; Campolo, L.; De Pasquale, R.; Miceli, N. Citrus
335 bergamia Risso & Poiteau juice protects against renal injury of diet-induced
336 hypercholesterolemia in rats. Phytotherapy Research **2010**, 24, 514-519.

337 Xu, G.; Liu, D.; Chen, J.; Ye, X.; Ma, Y.; Shi, J. Juice components and antioxidant capacity of
338 citrus varieties cultivated in China. Food Chemistry **2008**, 106, 545-551.

339

340 **Table 1** Sites of collection of *Citrus bergamia* in province of Reggio Calabria (Italy)

Area of collection	Latitude	Longitude
<i>Catona</i>	38.21705	15.63691
<i>Gallico</i>	38.16559	15.65367
<i>Arangea</i>	38.10929	15.64393
<i>Pellaro</i>	38.02050	15.64856
<i>Melito Porto S.</i>	37.92072	15.78568
<i>Palizzi</i>	37.96705	15.98719
<i>Africo</i>	38.05172	16.13407

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343 **Table 2** Average colorimetric values of bergamot juice

344

<i>Area of L collection</i>	<i>L</i>	<i>a*</i>	<i>b*</i>
<i>Catona</i>	1,83±0.02	0,05±0.02	0,60±0.04
<i>Gallico</i>	7,60±0.25	0,22±0.01	4,82±0.28
<i>Arangea</i>	6,69±1.35	0,10±0.11	4,47±1.46
<i>Pellaro</i>	4,78±0.15	0,29±0.11	2,51±0.17
<i>Melito</i>	1,77±0.08	0,06±0.03	0,64±0.06
<i>Palizzi</i>	1,95±0.11	0,21±0.07	1,09±0.11
<i>Africo</i>	2,21±0.03	0,21±0.06	1,32±0.06

345

346 **Table 3** Physicochemical data of bergamot juice

Area of collection	pH	Acidity g L ⁻¹	RRS	Formol index	
<i>Catona</i>	2,84±0.00 ^a	11,38±0.31 ^e	8,11±0.11 ^d	2,96±0.01 ^a	***
<i>Gallico</i>	2,72±0.00 ^d	12,44±0.04 ^d	11,50±0.02 ^a	2,53±0.02 ^e	***
<i>Arangea</i>	2,78±0.00 ^c	14,34±0.02 ^b	11,61±0.48 ^a	2,63±0.02 ^d	***
<i>Pellaro</i>	2,81±0.00 ^b	14,61±0.03 ^{ab}	10,36±0.06 ^b	2,75±0.0 ^{2c}	***
<i>Melito</i>	2,82±0.00 ^{ab}	14,73±0.03 ^a	9,22±0.08 ^c	3,63±0.01 ^d	***
<i>Palizzi</i>	2,72±0.00 ^d	13,11±0.10 ^c	9,25±0.06 ^c	2,91±0.02 ^a	***
<i>Africo</i>	2,54±0.00 ^d	14,83±0.06 ^a	9,57±0.03 ^c	2,86±0.02 ^b	***

347 Values are the mean of three independent determinations ± standard deviation.

348 Means followed by different letter are significantly different ($P < 0.05$).

349

350 **Table 4** Values of ascorbic acid, total flavonoid and total phenolic of Citrus Bergamia (juice)

Area of collection	Ascorbic acid mg 100mL ⁻¹	Flavonoid mg 100mL ⁻¹	Polyphenol mg 100mL ⁻¹	
<i>Catona</i>	285,35±10.05 ^a	104,06±2.17 ^a	180,53±2.86 ^d	***
<i>Gallico</i>	89,40±2.10 ^d	51,11±2.18 ^b	209,72±1.29 ^b	***
<i>Arangea</i>	152,09±4.50 ^c	148,15±3.37 ^a	228,15±2.70 ^a	***
<i>Pellaro</i>	171,91±1.85 ^b	147,81±4.58 ^c	202,37±0.45 ^c	***
<i>Melito</i>	183,38±7.34 ^b	106,40±1.82 ^a	233,36±0.48 ^a	***
<i>Palizzi</i>	184,10±2.71 ^b	74,00±2.11 ^b	211,84±0.65 ^b	***
<i>Africo</i>	173,06±3.21 ^b	126,94±3.61 ^c	203,22±0.50 ^c	***

351 Values are the mean of three independent determinations ± standard deviation.

352 Means followed by different letter are significantly different ($P < 0.05$).

353

354 **Table 5** Flavanone content (mg mL⁻¹) in bergamot juice

Area of collection	Eriocitrin mg mL ⁻¹	Naringin mg mL ⁻¹	Neeriocitrin mg mL ⁻¹	Neohesperidin mg mL ⁻¹	
<i>Catona</i>	12,26±0.42 ^{bc}	261,81±0.88 ^c	282,47±1.70 ^b	222,24±0.85 ^d	***
<i>Gallico</i>	6,36±0.32 ^e	142,69±2.98 ^e	172,98±1.60 ^e	110,79±0.25 ^f	***
<i>Arangea</i>	14,09±0.17 ^a	281,52±2.43 ^b	296,77±1.51 ^a	230,87±0.40 ^c	***
<i>Pellaro</i>	14,45±0.37 ^a	295,73±1.11 ^a	279,56±1.52 ^b	244,82±0.88 ^a	***
<i>Melito</i>	11,76±0.25 ^c	265,18±1.33 ^c	264,63±0.89 ^d	235,66±0.81 ^b	***
<i>Palizzi</i>	8,07±0.22 ^d	163,18±2.80 ^d	158,59±1.05 ^f	116,63±0.39 ^e	***
<i>Africo</i>	12,87±0.18 ^b	286,74±0.78 ^b	272,73±1.26 ^c	116,87±0.48 ^e	***

355 Values are the mean of three independent determinations ± standard deviation.

356 Means followed by different letter are significantly different ($P < 0.05$).

357

358 **Table 6** Antioxidant activity of *Citrus bergamia* Risso

	DPPH				
IC ₅₀ (μg mL ⁻¹) ¹⁾	ABTS				
IC ₅₀ (μg mL ⁻¹) ¹⁾	β-carotene bleaching				
IC ₅₀ (μg mL ⁻¹) ¹⁾					
<i>Area of collection</i>			30 min	60 min	
<i>Catona</i>	20.5 ± 1.3 ^e	21.8 ± 2.0 ^d	32.6 ± 2.4 ^d	36.7 ± 2.9 ^a	***
<i>Gallico</i>	21.1 ± 1.0 ^e	25.9 ± 1.9 ^c	34.8 ± 2.7 ^b	30.8 ± 2.0 ^e	***
<i>Arangea</i>	27.3 ± 1.9 ^b	32.6 ± 2.2 ^b	36.8 ± 2.6 ^a	35.7 ± 2.5 ^b	***

<i>Pellaro</i>	23.9 ± 1.4 ^d	25.7 ± 2.2 ^d	33.5 ± 2.9 ^c	34.8 ± 2.4 ^c	***
<i>Melito</i>	19.6 ± 2.0 ^f	17.4 ± 1.6 ^f	25.7 ± 2.9 ^f	24.9 ± 3.0 ^f	***
<i>Palizzi</i>	31.4 ± 1.1 ^a	35.6 ± 1.5 ^a	33.7 ± 2.8 ^c	31.8 ± 2.6 ^d	***
<i>Africo</i>	25.7 ± 1.2 ^c	18.9 ± 2.4 ^e	30.0 ± 1.5 ^e	35.3 ± 1.9 ^b	***
Positive control					
BHT					
Ascorbic acid	5.0 ± 0.8	1.7 ± 0.3			
Propyl gallate			1.0 ± 0.04	1.0 ± 0.05	

359 Data are expressed as means ± S.D. (n= 3); DPPH Radical Scavenging Activity Assay; One-way
 360 ANOVA *** $p < 0.0001$ followed by a multicomparison Dunnett's test: *** $p < 0.01$ compared with
 361 ascorbic acid. Antioxidant Capacity Determined by Radical Cation (ABTS⁺); One-way ANOVA
 362 *** $p < 0.0001$ followed by a multicomparison Dunnett's test: *** $p < 0.01$ compared with ascorbic
 363 acid. β -carotene bleaching test 30 min One-way ANOVA *** $p < 0.0001$ followed by a
 364 multicomparison Dunnett's test: *** $p < 0.01$ compared with propyl gallate. β -carotene bleaching

365 test 60 min One-way ANOVA *** $p < 0.0001$ followed by a multicomparison Dunnett's test: *** p
366 < 0.01 compared with propyl gallate. Means followed by different letter are significantly different

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368 **Figure 1** Map (Calabrian, Southern Italy) showing the area where Bergamot is cultivated



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