

pubs.acs.org/jced

Thermodynamic Behavior of Polyalcohols and Speciation Studies in the Presence of Divalent Metal Cations

Rosalia Maria Cigala,* Francesco Crea, Concetta De Stefano, Anna Irto, Demetrio Milea, and Silvio Sammartano



relevant polyalcohols (erythritol, sorbitol, maltol, and ethylmaltol) toward bivalent metal cations have been determined by potentiometric measurements with an ISE-H⁺ glass electrode in NaNO₃ aqueous solutions at $I = 0.151 \text{ mol·kg}^{-1}$ and T = 298.15 K. For the erythritol system, the investigations have been carried out in the ionic strength range $0.1 \leq I/\text{mol·kg}^{-1} \leq 1.0$, and the dependence on the ionic strength of the protonation and stability constants has been modeled by the specific ion interaction theory (SIT). The sequestering ability of the different ligands toward the considered metal cations (Ca²⁺, Zn²⁺, and Sn²⁺) has been evaluated by the pL_{0.5} parameter. For example, for M²⁺/maltol systems, at T = 298.15 K in NaNO_{3(aq)}, $I = 0.151 \text{ mol·kg}^{-1}$, and pH 7.4, the sequestering ability toward Zn²⁺ (pL_{0.5} = 4.60) is higher than that toward Sn²⁺ (pL_{0.5} = 3.94) and Ca²⁺ (pL_{0.5} = 1.27).

$\mathbf{F}_{\mathbf{r}}^{\mathbf{r}} = \begin{bmatrix} 100 & & & \\ 80 & & & \\ 80 & & & \\ 60 & & & \\ 40 & & & \\ 40 & & & \\ 0 & & & \\ 3 & 4 & 5 & 6 & 7 \\ \mathbf{pH} \end{bmatrix}$

1. INTRODUCTION

The polyalcohols or polyols are monomers or polymers with more than one hydroxyl functional group available for reactions. They are of great importance in food sciences, since they are used as sweeteners to substitute the sugar. Recently, the interest toward their consumption has increased, due to their potential beneficial effect on health; the principal qualities are noncariogenicity, a low energy value, a low glycemic and insulin index, they are digested very slowly, and they have osmotic properties (hydration of the colon, with laxative and depurative effect). As regards the last property, the polyalcohols favor the formation of saccharolytic and acidic anaerobic organisms in the colon, purifying it from endotoxic, rotting, and pathological organisms, which explains their importance and clinical relevance. In addition, polyols help to form short-chain fatty acids that are beneficial for colon health.¹ As said above, polyalcohols are employed as food additives, which are substances used in the food industry during the preparation, storage, and marketing of foodstuffs.² The additives are classified according to their function and subdivided into three large groups:

- additives helping to preserve the freshness of food, therefore, preservatives, which slow down the growth of microbes, and antioxidants, which prevent rancidity phenomena
- additives improving the sensory characteristics of foods, such as dyes, thickeners, emulsifiers, sweeteners, and flavor enhancers

• technological additives used to facilitate the processing of food but which do not have a specific function in the final product, for example, anti-foaming and anti-caking agents.

Food additives undergo a safety assessment process before being authorized for food use. In Europe, the evaluation is performed from European Food Safety Authority (EFSA) and by Joint Expert Committee on Food Additives (JECFA), an international scientific expert committee administered jointly by the Food and Agriculture Organization of the United Nations (FAO) and World Health Organization (WHO). The additives authorized at the European level are marked with a numerical code preceded by the letter E.

Erythritol (1,2,3,4-Butanetetrol, abbreviated to the acronym *Ery*, see Scheme 1) is a polyalcohol naturally present in fruit and fermented foods,³ industrially obtained from sugary substrates, (for example, starch, glucose, sucrose, etc.), by microbial fermentation by selected osmophilic yeasts (e.g., *Moniliella pollinis*).⁴ The European Commission has included erythritol in the list of polyvalent food additives, with the abbreviation E968. It is used as a sweetener, but it can be employed with different functions, for example, to give body and disguise unwanted

Received: February 3, 2020 Accepted: April 16, 2020 Published: April 29, 2020





Scheme 1. Structures of (a) Erythritol, (b) Sorbitol, (c) Maltol, and (d) Ethylmaltol



aftertaste. The metabolic profile of this polyalcohol is unique, due to its small size and therefore its low molecular weight (122.12 g mol⁻¹); over 90% of ingested erythritol is absorbed in the small intestine through passive diffusion (i.e., without energy consumption by the cell, or osmosis) and is excreted unchanged through the urine. The remaining fraction (<10%) reaches the large intestine where it is only partially metabolized. As a consequence, the total caloric value of erythritol is very low and varies from 0 to a maximum of 0.2 kcal g^{-1} .^{5,6} Furthermore, it is characterized by almost zero value of glycemic index (IG) and insulin index (II) and therefore it represents a valid substitutive ingredient of sucrose to reduce the glycemic impact of the diet, suitable for those at risk or suffering from diabetes.¹ Recent studies have also highlighted the anti-radical activity of erythritol, which has proved to be an excellent scavenger of hydroxyl radicals, with protective properties for cell membranes.' It was certified as a tooth-friendly product, and its acariogenicity is widely demonstrated: erythritol is not converted into acids by the bacteria present in the mouth; therefore, it does not favor dental caries. On the contrary, it seems to have a protective role, similar to that of xylitol, against bacterial plaque.

Sorbitol ((2S,3R,4R,5R)-Hexane-1,2,3,4,5,6-hexol abbreviated to Sorb, see Scheme 1), a naturally occurring polyol, is widely used as an additive, E420, in the food industry as a sweetener, humectant, and texturizing agent, in toothpastes, shampoo, and soap as a softening ingredient, as an excipient in pharmaceutical formulations, and as a plasticizer in film formulations.^{9,10} Sorbitol may also be used analytically as a marker for assessing liver blood flow.¹¹ Since sorbitol is an alcohol rather than a sugar, it is relatively resistant to fermentation and hampering acid formation caused by micro-organisms found in the mouth. For this reason, it is used for sweetening and bodying of noncariogenic foods and soft drinks, as said for the erythritol. It was observed that a little amount of sorbitol (0.5-3.0%) in wine is able to chelate low levels of iron and copper and smoothing out bitterness in lower-quality wine.¹² Sorbitol is an isomer of mannitol, and the difference between the two polyols consists of the planar orientation of the OH group on the second carbon atom. Each isomer is characterized by its own individual set of properties; the most important difference is the response toward moisture. Sorbitol is hygroscopic, while mannitol is resistant to moisture sorption, even at high relative humidities. Sorb forms water-soluble chelates with many bivalent and trivalent metal cations under strongly acidic and alkaline conditions.

Maltol (3-hydroxy-2-methyl-4H-piran-4-one, abbreviated to *Malt*, see Scheme 1) is naturally present mainly in larch bark and pine needles; in the food industry, it is used as a flavor enhancer (identified with E636) in food, beverage, tobacco, brewing, and cosmetics for its flavor and anti-oxidant properties.¹³ It exhibits interesting anti-neoplastic activities attributed to the formation of reactive oxygen species^{14,15} as well as coordination properties toward metal cations; for this reason, ligands containing maltol

have been developed and exploited as new potential metal-based anti-tumor drugs. 16,17

Ethylmaltol (2-Ethyl-3-hydroxy-4H-pyran-4-one, abbreviated to *EMalt*, see Scheme 1) differs to maltol for the presence of an ethyl group rather than a methyl one. Also, this polyol is used as a flavor enhancer and is identified with E637. The 3-hydroxy-4pyranones are employed both for the control of metal levels in the human body and for oral administration in view of diagnostic or therapeutic purposes.¹⁸ Furthermore, they have been suggested for the treatment of anemic patients for introduction of iron or its removal (also in the case of aluminum) from patients suffering of bodily overload as in thalassemia. This family of compounds also has a role in the administration of ⁶⁷Ga or ¹¹¹In for radiopharmaceuticals and of gadolinium for magnetic resonance imaging (MRI).¹⁹ The beneficial therapeutic effects of compounds of zinc have been recognized, and more recently, the isotopes ⁶³Zn and ^{69m}Zn have been employed in radiopharmaceuticals. The absorption of zinc by the human body depends on the form in which it is administered. In fact, it seems that the simple aqua-ion $Zn^{2+}{}_{(aq)}$ and its phytate complex are not well absorbed, but the complexes with citrate, histidine, prolinate, several EDTA derivatives, and 3-hydroxy-4-pyranones and 3-hydroxy-4-pyridinones are absorbed by the body. In order to increase the bioavailability of zinc, several complexes such as Zn-maltol and Zn-ethylmaltol have been synthesized.²⁰ Following this study, Zn-ethylmaltol complex could be considered as a suitable zinc species for the treatment of metal deficiency in replacement of Zn^{2+} -sulfate. The determination of stability constants of Zn-maltol derivatives may be a new way for the administration of zinc.

The metal cations selected for this study are Ca^{2+} , Zn^{2+} , and Sn^{2+} , and the first two are essential elements for humans, playing a central, if still only dimly understood, role in a number of healing processes, while the main use of tin is for the manufacture of cans and containers, typically as tinplate.^{21,22} As a consequence, the direct contact of tin with foods may result in migration from the inside wall of cans and other packaging materials to edible contents.

In this paper, the studies, by potentiometric techniques, on the acid—base properties of erythritol and complexing ability of erythritol, sorbitol, maltol, and ethylmaltol toward Ca²⁺ (only for maltol and ethylmaltol), Zn²⁺, and Sn²⁺ are reported in NaNO₃ aqueous solutions at $I = 0.151 \text{ mol·kg}^{-1}$ and at T =298.15 K. For the erythritol system, the investigations have been performed at different ionic strengths, $0.1 \leq I/\text{mol·kg}^{-1} \leq 1.0$, and the dependence on I of the protonation and stability constants has been modeled by the Specific ion Interaction Theory (SIT). The sequestering ability of the different ligands toward the investigated metal cations, Ca²⁺, Zn²⁺, and Sn²⁺, has been analyzed by the empirical pL_{0.5} parameter.

2. MATERIALS AND METHODS

2.1. Materials. The polyalcohol solutions (erythritol, sorbitol, maltol, and ethylmaltol from Sigma-Aldrich-Merck) were prepared, with analytical grade water ($R = 18 \text{ M}\Omega \text{ cm}^{-1}$) and grade A glassware, from products of high available purity. The hydrochloric acid (HCl) and sodium hydroxide (carbonate free, NaOH) solutions were prepared from dilution of concentrated ampoules (Riedel-deHäen) and standardized against sodium carbonate and potassium hydrogen phthalate, respectively. The NaOH solutions were preserved from atmospheric CO₂ by means of soda lime traps. The solutions of ZnCl₂ and SnCl₂ were prepared, without further purification,

Table 1. Calculated Deprotonation and Formation Constants of Zn^{2+} and Sn^{2+}/Ery Species in NaNO _{3(aq)} at Different Ionic
Strengths and Ionic Strength Dependence Parameters, eq 3, at $T = 298.15$ K and $p = 0.1$ MPa ⁴

						$I/{ m mol}\cdot { m kg}^{-1}$		
equilibrium	$\log \beta^0 b$	z^*	$\Delta \varepsilon^{c}$	0.15	0.25	0.50	0.75	1.00
$Ery = [Ery(H)_{-1}]^{-} + H^{+}$	-12.54 ± 0.03^{d}	-2	0.35 ± 0.01^{d}	-12.24 ^e	-12.16	-12.01	-11.89	-11.78
$Ery = [Ery(H)_{-2}]^{2-} + 2H^+$	-24.24 ± 0.04	-6	0.51 ± 0.01	-23.42	-23.24	-22.94	-22.71	-22.51
$Zn^{2+} + Ery = [ZnEry(H)_{-1})]^{+} + H^{+}$	-5.62 ± 0.03^{d}	2	1.37 ± 0.01^{d}	-5.67	-5.57	-5.29	-4.98	-4.66
			Model 1					
$\mathrm{Sn}^{2+} + \mathrm{Ery} = [\mathrm{Sn}\mathrm{Ery}]^{2+}$	2.82 ± 0.01^{d}	0	-1.44 ± 0.01^{d}	2.61	2.46	2.10	1.74	1.38
$Sn^{2+} + Ery = [SnEry(H)_{-1}]^{+} + H^{+}$	-0.65 ± 0.02	2	0.53 ± 0.01	-0.82	-0.80	-0.73	-0.63	-0.52
			Model 2					
$Sn^{2+} + 2Ery = [Sn(Ery)_2]^{2+}$	4.95 ± 0.08	0	-1.31 ± 0.12	4.76	4.62	4.30	3.97	3.64
$Sn^{2+} + Ery = [SnEry(H)_{-1}]^+ + H^+$	-1.01 ± 0.06^{d}	2	0.92 ± 0.08^{d}	-1.12	-1.07	-0.90	-0.70	-0.50
a_{Char} \mathbf{J}_{char} \mathbf{J}_{char} \mathbf{J}_{char} \mathbf{J}_{char} \mathbf{J}_{char} \mathbf{J}_{char} \mathbf{J}_{char}		-111	b _D	. 1 6		· :		¢

"Standard uncertainties: u(T) = 0.1 K, u(I) = 0.001 mol·kg⁻¹. "Deprotonation and formation constants at infinite dilution, eq 3. "Empirical parameter of eq 3. " \pm Std. dev. "Calculate formation constants according to eq 3.

by weighing the dihydrated pure salt. Their purity was checked by titrations with EDTA standard solutions,²³ and it was in all the cases \geq 99.5%.

therefore to know the formation percentage of the last one in the distribution diagrams.

3. RESULTS AND DISCUSSION

Particular attention was employed for the preparation of tin solutions, in order to prevent the oxidation of Sn(II) to Sn(IV) and the starting of the hydrolysis process, owing to hamper this, the $SnCl_2$ solutions were acidified with HCl to reach pH < 2 and a piece of metallic tin was added to the solutions after the preparation. Furthermore, these solutions were bubbled with purified N₂ to exclude O₂.

The ionic medium aqueous solutions, NaNO₃, were prepared by weighing the pure salt (Fluka), previously dried in an oven, at T = 383.15 K, for at least 2 h.

2.2. Potentiometric Technique. The determination of the protonation and complex formation constants of the ligands were carried out by potentiometric ISE-H⁺ measurements. A 25 cm³ portion of the solutions containing the polyalcohol (at different concentrations) and the ionic medium, NaNO₃, at a pre-established ionic strength value was titrated with NaOH standard solutions, in thermostatted cells. In the case of the complex formation studies, an amount of the different metal cations considered and a known aliquot of strong acid (HCl) were added to the investigated solutions. All of the potentiometric titrations were carried out under magnetic stirring and bubbling purified presaturated N_{2(g)} through the solutions, to exclude O_{2(g)} and CO_{2(g)} inside. More details on the procedures for the potentiometric measurements have already been reported in previous papers.²⁴

2.3. Computer Programs. All of the potentiometric parameters such as standard electrode potential (E^0) , basic junction coefficient (j_b) , analytical concentration of the ligands, ionic product of water (log K_w), and protonation constants were refined using the ESAB2M²⁵ computer program, while the complex formation constants of the different metal-ligand systems were determined by using the BSTAC²⁶ program. The study of the dependence on the ionic strength of the protonation and complex formation constants and the refinement of the corresponding parameters was performed by the least-squares computer program LIANA.²⁷ The ES4ECI²⁸ and HYSS²⁹ computer programs have been used to draw the speciation diagrams and to calculate the species formation percentages. In the case of the second program, it is possible, over considering the protonation, hydrolysis, and formation constants of all of the species present in the speciation model, to take into account also the solubility product of the sparingly soluble species, and

3.1. Acid–Base Properties of Polyalcohols. The acid– base properties of erythritol (*Ery*) have been studied by experimental potentiometric measurements, while, in the case of the sorbitol (*Sorb*),^{30–32} maltol (*Malt*), and ethylmaltol (*EMalt*),³³ literature data have been considered. The hydrolytic constants of the metal cations, Ca^{2+} , Zn^{2+} , Sn^{2+} , and in the last case also the solubility product of the sparingly soluble species $Sn(OH)_{2(s)}$, have been taken into account in the elaboration of the complex formation constants.^{34,35} The conversion from the molar to the molal concentration scale of the constants was carried out by using the appropriate density values.³⁶

3.2. Acid–Base and Complexing Properties of Erythritol. The acid–base properties and the complexing ability toward Zn²⁺ and Sn²⁺ of *Ery* have been studied by the potentiometric technique in NaNO₃ aqueous solutions at different ionic strengths $(0.1 \le I/\text{mol·kg}^{-1} \le 1.0)$ and at T = 298.15 K. For the determination of the acid–base behavior of this ligand, concentrations in the range $2.0 \le c_{Ery}/\text{mmol·kg}^{-1} \le 10.0$ were used, in water and a suitable amount of NaNO₃, to obtain the desired ionic strength value. The elaboration of the potentiometric data, in the pH range 7.0-11.5, led us to determine two deprotonation constants, expressed by means of the equilibria

$$Ery = [Ery(H)_{-1}]^{-} + H^{+} \qquad \log K_{-1}^{H}$$
(1)

and

$$Ery = [Ery(H)_{-2}]^{2-} + 2H^{+} \qquad \log \beta_{2}^{H}$$
(2)

The deprotonation constants determined at different ionic strengths have been fitted according to the Specific ion Interaction Theory (SIT) approach,^{37,38} to obtain the ionic strength dependence parameters

$$\log \beta = \log \beta^{0} - z^{*} \cdot 0.51 \cdot (I^{0.5} / (1 + 1.5 \cdot I^{0.5})) + \Delta \varepsilon \cdot I$$
(3)

where $\log \beta^0$ is the equilibrium constant at infinite dilution, z^* is

$$z^* = \sum (\text{charges})_{\text{reag}}^2 - \sum (\text{charges})_{\text{prod}}^2$$

and $\Delta \varepsilon$ is the summation of the specific interaction coefficients of the species involved in the equilibrium and the ions of the background electrolyte. Model 1

pubs.acs.org/jced

Model 2



Figure 1. Distribution diagram of the Sn²⁺/erythritol system: models 1 and 2, in NaNO_{3(aq)} at $I = 0.151 \text{ mol·kg}^{-1}$ and at T = 298.15 K. Experimental conditions: $c_{\text{Sn}}^{2+} = 1.5 \text{ mmol·kg}^{-1}$ and $c_{\text{Ery}} = 5.0 \text{ mmol·kg}^{-1}$. Standard uncertainties: u(pH) = 0.01, u(T) = 0.1 K, $u(I) = 0.001 \text{ mol·kg}^{-1}$. Legend: **Model 1:** 1. SnEry; 2. SnEry(H₋₁); 3. Sn(OH)_{2(s)}; 4. Sum of the Sn²⁺ hydrolytic species; 5. Sum of the SnCl_n species; 6. Free Sn²⁺. **Model 2:** 1. SnEry₂; 2. SnEry(H₋₁); 3. Sn(OH)_{2(s)}; 4. Sum of the Sn²⁺ hydrolytic species; 5. Sum of the SnCl_n species; 6. Free Sn²⁺. Charges were omitted for simplicity. The shaded zone refers to the pH range where the sparingly soluble species, Sn(OH)_{2(s)}, is present.

This procedure allowed us to calculate the deprotonation constants at different ionic strengths and at infinite dilution, as reported in Table 1.

The knowledge of the acid-base properties of erythritol is important for solutions with pH > 10.5, because the displacement of the proton of the -OH groups (eqs 1 and 2) occurs at high pH values. As expected, the acidity of the two alcoholic groups is very similar and their values are in agreement with protonation (or deprotonation) data reported in the literature for alcoholic groups (e.g., sorbitol and glycerol). The complexation of Zn²⁺ and Sn²⁺ was investigated preparing solutions at the following experimental conditions: $1.0 \le c_{Zn}^{2+}$ /mmol·kg⁻¹ \le 3.0, $2.0 \le c_{Ery}$ /mmol·kg⁻¹ \le 9.0, and $0.5 \le c_{Sn}^{2+}$ /mmol·kg⁻¹ \le 1.5, $2.0 \le c_{Ery}$ /mmol·kg⁻¹ \le 8.2, with HCl in different amounts and NaNO₃ to adjust the ionic strength values, $0.1 \le I/\text{mol} \cdot \text{kg}^{-1}$ \leq 1.0, at *T* = 298.15 K. From the potentiometric investigations of the Zn^{2+}/Ery system, the $ZnEry(H)_{-1}$ species has been determined. During the measurements, the formation of a sparingly soluble species was observed below pH \sim 7.5, probably due to the formation of a neutral species between Zn^{2+} and erythritol (e.g., $ZnEry_2(H)_{-2}$), but its formation was not obtained from the calculations. The formation pH of the sparingly soluble species depends on the m_{Zn}^{2+} : m_{Ery} molal ratio. The dependence on the ionic strength of the formation constant has been studied by eq 3, and the relative parameters are reported in Table 1, together with the calculated formation constants at different ionic strengths. From the elaboration of the experimental data of the Sn²⁺/erythritol system, two speciation models have been proposed: model 1 consists of SnEry and SnEry(H)₋₁ species, while model 2 consists of $Sn(Ery)_2$ and $SnEry(H)_{-1}$ species. The formation constant values of the species and their corresponding equilibria are reported in Table 1 together with the $\Delta \varepsilon$ parameter of eq 3. Considering the criteria generally used to select the best speciation model, namely, the simplicity and the likelihood of the proposed species, the formation percentages and the pH range where they form, the consistency of various models under the different conditions, and the values of the variance ratio, both models can be considered reliable, even if the consistency of model 1 under the different conditions is preferable. The formation of the Sn²⁺/Ery species occurs at acidic pH values and

is poorly stable, leading to the formation of the sparingly soluble hydroxo species at pH ~ 2.5, although the concentration of Sn^{2+} was maintained low, to avoid the precipitation of the $\text{Sn}(\text{OH})_{2(s)}$ species. Figure 1 reports the distribution diagrams of the species at $I = 0.151 \text{ mol·kg}^{-1}$ and T = 298.15 K, for the two speciation models. As it can be seen, the formation percentage of the Sn*Ery*H₋₁ species is low at ~8 and ~2% for models 1 and 2, respectively, while the Sn*Ery* and Sn(*Ery*)₂ species reach about 62 and 40%, respectively. In both cases, the solubility product of the Sn(OH)_{2(s)} species has been taken into account, and as evidenced in Figure 1, its formation occurs at pH ~ 2.7, for model 1, and at pH ~ 2.6, for model 2.

3.3. Acid–Base and Complexing Properties of Sorbitol. The deprotonation constants of the sorbitol are already known and reported in the literature, ^{31,32} where generally a single deprotonation step has been reported. On the contrary, Gaidamauskas et al.³⁰ report two deprotonation steps with constant values very close: $\log K^{\rm H}_1 = -13.6$ and $\log \beta^{\rm H}_2 = -27.0$ ($\log K^{\rm H}_2 = -13.4$, the equilibria are reported in Table 2). The complexing ability of sorbitol toward Sn²⁺ has been investigated by the potentiometric technique, at I = 0.151 mol·kg⁻¹ in NaNO_{3(aq)} and T = 298.15 K. The analytical concentration ranges used for the measurements are $2.0 \le c_{Sorb}/$ mmol·kg⁻¹ ≤ 4.5 and $0.5 \le c_{\text{sn}}^{2+}/$ mmol·kg⁻¹ ≤ 1.5 , with HCl in

Table 2. Deprotonation and Formation Constants of Sn²⁺/ Sorb Species in NaNO_{3(aq)} at $I = 0.151 \text{ mol·kg}^{-1}$, T = 298.15K, and $p = 0.1 \text{ MPa}^{a}$

equilibrium	$\log \beta$			
$Sorb = [Sorb(H_{-1})]^{-} + H^{+}$	-13.6^{b}			
$Sorb = [Sorb(H)_{-2}]^{2-} + 2H^+$	-27.0^{b}			
Model 1				
$Sn^{2+} + Sorb = [SnSorb]^{2+}$	2.63 ± 0.02^{c}			
$Sn^{2+} + Sorb = [SnSorb(H)_{-1}]^{+} + H^{+}$	-0.84 ± 0.04			
Model 2				
$Sn^{2+} + 2Sorb = [Sn(Sorb)_2]^{2+}$	5.13 ± 0.02			
$Sn^{2+} + Sorb = [SnSorb(H)_{-1}]^{+} + H^{+}$	-0.90 ± 0.05			

^{*a*}Standard uncertainties: u(T) = 0.1 K, u(I) = 0.001 mol·kg⁻¹. ^{*b*}Calculated from literature values.³⁰⁻³² ^{*c*}±Std. dev.

pubs.acs.org/jced

Article

Table 3. Deprotonation and Stability	Constants of the M^{2+}/M	<i>1alt</i> and M ²⁺ / <i>EMalt</i> Sys	stems in NaNO _{3(aq)} :	at <i>I</i> = 0.151 mol·kg ⁻¹	, T =
298.15 K, and <i>p</i> = 0.1 MPa ^{<i>a</i>}					

equilibrium		log	β			
$Malt = [Malt(H_{-1})]^- + H^+$		-8.41	17 ^b			
$EMalt = [EMalt(H_{-1})]^{-} + H_{-1}$	I ⁺	-8.50)4 ^b			
	Ca ²⁺	Zn ²⁺	Sn ²⁺			
Maltol						
$\mathbf{M}^{2+} + Malt = [\mathbf{M}Malt]^{2+}$	2.33 ± 0.02^{c}	5.598 ± 0.008^c	10.66 ± 0.10^{c}			
$M^{2+} + 2Malt = [M(Malt)_2]^{2+}$		10.246 ± 0.012	18.26 ± 0.05			
$M^{2+} + 3Malt^{-} = [M(Malt)_3]^{2+}$		11.62 ± 0.15				
Ethylmaltol						
$\mathbf{M}^{2+} + EMalt = [\mathbf{M}EMalt]^{2+}$	$2.302 \pm 0.015^{\circ}$	5.639 ± 0.010^{c}	10.83 ± 0.10^{c}			
$M^{2+} + 2EMalt = [M(EMalt)_2]^{2+}$		10.33 ± 0.02	18.61 ± 0.03			
$\mathbf{M}^{2+} + 3EMalt = [\mathbf{M}(EMalt)_3]^{2+}$		12.10 ± 0.10				
^a Standard uncertainties: $u(T) = 0.1$ K, $u(I) = 0.001$ mol·kg ⁻¹ , ^b Constant values from ref 33, ^c +Std, dev.						

different amounts and NaNO3 to obtain the desired ionic strength value. The pH range investigated is 2.0-3.5 because, at higher values, the formation of the sparingly soluble species $Sn(OH)_{2(s)}$ occurs. From the elaboration of the experimental data, as for the Sn^{2+}/Ery system, two speciation models have been obtained: model 1, SnSorb and SnSorb(H)-1 species; model 2, $Sn(Sorb)_2$ and $SnSorb(H)_{-1}$ species. The formation constant values and the corresponding formation equilibria are reported in Table 2. Applying the criteria of selection already cited, we can consider model 1 as the most reliable with respect to another one. Taking into account the most common databases,^{39,40} where the results of some speciation studies concerning the interaction of glycerol and sorbitol with different metal cations (Ln³⁺, B³⁺, As³⁺, Al³⁺, Fe³⁺, Cu²⁺, etc.) are reported, we can observe that, independent of the temperature, ionic strength, and ionic medium, the formation of ternary hydrolytic metal-ligand species seems to be favored. However, these studies were carried out at a single ionic strength value, generally at I = 0.1 or 1.0 mol·kg⁻¹, and no systematic investigation for the dependence of the stability constants on ionic strength is reported.

3.4. Complexing Properties of Maltol and Ethylmaltol. The acid-base behavior of maltol and ethylmaltol was already studied and reported in a previous paper.³³ The complexing ability of these two ligands has been studied toward Ca^{2+} , Zn^{2+} , and Sn²⁺ by the potentiometric technique, in NaNO₃ aqueous solutions at $I = 0.151 \text{ mol·kg}^{-1}$ and T = 298.15 K. The experimental conditions used are ligand $1.3 \le c_{Malt}$ or $c_{EMalt}/$ mmol·kg⁻¹ ≤ 22.3 , metal cation $1.0 \le c_{Ca}^{2+}/$ mmol·kg⁻¹ ≤ 4.1 or $1.3 \le c_{Zn}^{2+}/$ mmol·kg⁻¹ ≤ 4.3 or $0.5 \le c_{Sn}^{2+}/$ mmol·kg⁻¹ ≤ 2.0 , an amount of strong acid (HCl) and NaNO3. The potentiometric data were collected in the pH ranges 6.0-10.5, 3.0-9.0, and 2.0-9.5 for Ca²⁺, Zn²⁺, and Sn²⁺ systems, respectively. The different pH range investigated, for the different systems, is dependent on the different hydrolytic behavior of the considered metal cations and in particular on the formation of the sparingly soluble species during the alkalimetric titrations. The experimental results highlighted that, for a given metal cation (Ca²⁺, Zn^{2+} , or Sn^{2+}), the same speciation models were obtained independent of the ligand (maltol or ethylmaltol) considered. For Ca^{2+} , only the formation of the CaL species (L = maltol or ethylmaltol) was observed. Other species were checked, but they were systematically rejected by the BSTAC computer program. In the case of the $Zn^{2+}/Malt$ and $Zn^{2+}/EMalt$, the formation of three complex species was obtained, namely, ZnL, ZnL₂, and ZnL₃, while, for the Sn²⁺/Malt and Sn²⁺/EMalt, the speciation

scheme is represented by two complexes: SnL and SnL₂. The stability constants of the $M^{2+}/Malt$ and $M^{2+}/EMalt$ species are reported in Table 3. In the case of the Sn^{2+} systems, the experimental measurements were carried out with different $m_{\rm Sn}^{2+}:m_{\rm L}$ molal ratios but always with a large excess of the ligands. Owing to the high ligand concentrations and $m_{Sn}^{2+}:m_L$ molal ratios used, it has been possible to investigate a wide pH range, 2-9.5, without the formation of the sparingly soluble species $Sn(OH)_{2(s)}$. Concerning maltol, from a comparison of the stability constant values of the ML species, it is possible to note that the stability of the SnL complex is higher than that obtained for ZnL and CaL, i.e., 10.66 ± 0.10, 5.598 ± 0.008, and 2.33 ± 0.02 , respectively. There is not a significant difference between the formation constant values of maltol and ethylmaltol species, which indicates the absence of an influence of the methyl and ethyl groups of the lateral chain on the stability of the complexes.

In Figure 2, the distribution diagram of the $Zn^{2+}/maltol$ species shows that the Zn*Malt* species exists in the pH range 3–7.5 and it achieves 60% of formation even if the main species is the Zn(*Malt*)₂ that at pH ~ 7.5 reaches 96%. The Zn(*Malt*)₃ species forms at pH ~ 7 and achieves 15% of formation. The



Figure 2. Distribution diagram of the Zn²⁺/maltol system, in NaNO_{3(aq)} at $I = 0.151 \text{ mol·kg}^{-1}$ and at T = 298.15 K. Experimental conditions: $c_{Zn}^{2+} = 2.6 \text{ mmol·kg}^{-1}$ and $c_{Malt} = 14.9 \text{ mmol·kg}^{-1}$. Standard uncertainties: u(pH) = 0.01, u(T) = 0.1 K, $u(I) = 0.001 \text{ mol·kg}^{-1}$. Charges were omitted for simplicity.

Journal of Chemical & Engineering Data

literature reports some data on the complexation of maltol with different metal cations; for example, for the Zn²⁺/Malt system, Jakusch et al.⁴¹ report, in the molar concentration scale, at T = 298.15 K in KCl aqueous solution and I = 0.2 mol·dm⁻³, the formation of three species: ZnL, ZnL₂, and ZnL₃ with log $\beta = 5.57 \pm 0.02$, 10.29 ± 0.02 , and 12.71 ± 0.08 , respectively. These data are in good agreement with the results reported in this paper, in the same order, log $\beta = 5.60$, 10.25, and 11.62 (see Table 3).

3.5. Sequestering Ability. The sequestering ability of the polyalcohols toward the metal cations has been evaluated by the empirical parameter $pL_{0.5}$.⁴² It represents the total concentration of ligand required to sequester 50% of a metal cation when present in trace amount $(10^{-12} \text{ mol} \cdot \text{kg}^{-1})$ in solution. The $pL_{0.5}$ value depends on the experimental conditions such as pH, ionic strength, ionic medium, and temperature. From a mathematical point of view, this can be expressed by a sigmoidal Boltzmann type equation

$$\chi_{\rm M} = \frac{1}{1 + 10^{(\rm pL-pL_{0.5})}} \tag{4}$$

where $\chi_{\rm M}$ is the mole fraction of metal cation complexed by the ligand, pL = $-\log c_{\rm L}$, and pL_{0.5} = $-\log c_{\rm L}$, at $\chi_{\rm M}$ = 0.5.

The sequestering ability of a ligand can be graphically represented by a dose—response curve, with asymptotes equal to 1 for $pL \rightarrow -\infty$ and 0 for $pL \rightarrow +\infty$, obtained by plotting the mole fraction of metal complexed vs the pL. A high sequestering ability of a ligand toward a metal cation is represented by high $pL_{0.5}$ values. In the calculation of this empirical parameter, all of the interactions in the system, such as the protonation of the ligand, the hydrolysis of the metal cation, and the interactions with other components, are taken into account in the speciation model.

From the speciation study, it was seen that the acid–base properties and complexing ability of maltol and ethylmaltol appear fairly similar, but analyzing the obtained $pL_{0.5}$ values, we observe a significantly different behavior between Ca²⁺, Zn²⁺, and Sn²⁺. As examples in Figures 3 and 4, the sequestering



Figure 3. Calcium sequestration diagram by maltol and ethylmaltol. Molar fraction of Ca²⁺ complexed vs the total ligand concentration (pL) at T = 298.15 K and I = 0.151 mol·kg⁻¹ in NaNO_{3(aq)} at pH 7.4. (\bigcirc) Ca²⁺/ethylmaltol system, pL_{0.5} = 1.16; (\square) Ca²⁺/maltol system, pL_{0.5} = 1.27. Standard uncertainties: u(pH) = 0.01, u(T) = 0.1 K, u(I) = 0.001 mol·kg⁻¹.



Figure 4. Sequestration diagram of maltol toward Ca²⁺, Sn²⁺, and Zn²⁺ cations. Molar fraction of M²⁺ complexed vs the total ligand concentration (pL) at T = 298.15 K and I = 0.151 mol·kg⁻¹ in NaNO_{3(aq)} at pH 7.4. (\Box) Ca²⁺/maltol system, pL_{0.5} = 1.27; (\bigcirc) Sn²⁺/maltol, pL_{0.5} = 3.94; (\triangle) Zn²⁺/maltol system, pL_{0.5} = 4.60. Standard uncertainties: u(pH) = 0.01, u(T) = 0.1 K, u(I) = 0.001 mol·kg⁻¹.

abilities of *Malt* and *EMalt* toward Ca²⁺ and *Malt* toward Ca²⁺, Sn²⁺, and Zn²⁺ cations, respectively, are reported. In Figure 3, the sequestration diagram is drawn at pH 7.4, in NaNO_{3(aq)} at *I* = 0.151 mol·kg⁻¹ and *T* = 298.15 K. As can be observed, the two ligands show the same sequestering ability toward Ca²⁺; in fact, for the Ca²⁺/ethylmaltol system, the pL_{0.5} value is 1.16, while, for the Ca²⁺/maltol, it is 1.27. In Figure 4, the diagram evidences that the sequestering ability of *Malt* is very different toward the various metal cations; the pL_{0.5} values are 1.27, 3.94, and 4.60 for Ca²⁺, Sn²⁺, and Zn²⁺, respectively. Therefore, the sequestering ability of *Malt* and *EMalt*, under these experimental conditions, follows the trend Zn²⁺ > Sn²⁺ > Ca²⁺.

It is important to note that, in spite of the higher stability of the Sn*Malt* and Sn*EMalt* species than that of the Zn*Malt* and Zn*EMalt* ones, an opposite trend in terms of sequestering ability is obtained. A comparison of the sequestering ability of maltol, ethylmaltol, and erythritol toward Zn²⁺ is shown in Figure 5. From the calculation of the pL_{0.5} parameter, it comes out that maltol and ethylmaltol sequestrate the Zn²⁺ in a more effective way than erythritol, with pL_{0.5} values of 4.60, 4.56, and 1.72 for the Zn²⁺/maltol, Zn²⁺/ethylmaltol, and Zn²⁺/erythritol systems, respectively. In Figure 6, comparison in terms of the sequestering ability of erythritol and sorbitol toward Sn²⁺ is reported. These two ligands have the same behavior with respect to this metal cation, in NaNO_{3(aq)} at $I = 0.151 \text{ mol·kg}^{-1}$, T = 298.15 K, and pH 2.0. The pL_{0.5} values are 2.61 for Sn²⁺/erythritol and 2.62 for Sn²⁺/sorbitol.

CONCLUSIONS

In this paper, the speciation study of four polyalcohols, namely, erythritol, sorbitol, maltol, and ethylmaltol, has been reported in NaNO_{3(aq)}, at different ionic strengths and T = 298.15 K. The results obtained can be reassumed in some points:

 The acid-base properties of the ligands seem to be influenced by their structure; in fact, erythritol and sorbitol, which have the same structure but two -(CH₂)-OH groups more in the latter, have the same behavior. Maltol and ethylmaltol, that differ only for an extra methylene group, also have an equal protonation



Figure 5. Zinc sequestration diagram by maltol, ethylmaltol, and erythritol. Molar fraction of Zn²⁺ complexed vs the total ligand concentration (pL) at T = 298.15 K and I = 0.151 mol·kg⁻¹ in NaNO_{3(aq)} at pH 7.4. (\Box) Zn²⁺/maltol system, pL_{0.5} = 4.60; (\bigcirc) Zn²⁺/ ethylmaltol, pL_{0.5} = 4.56; (\triangle) Zn²⁺/erythritol system, pL_{0.5} = 1.72. Standard uncertainties: u(pH) = 0.01, u(T) = 0.1 K, u(I) = 0.001 mol·kg⁻¹.



Figure 6. Tin(II) sequestration diagram by erythritol and sorbitol, with respect to model 1. Molar fraction of Sn^{2+} complexed vs the total ligand concentration (pL) at T = 298.15 K and I = 0.151 mol·kg⁻¹ in NaNO_{3(aq)} at pH 2.0. (\Box) Sn²⁺/erythritol system, pL_{0.5} = 2.61; (\bigcirc) Sn²⁺/sorbitol, pL_{0.5} = 2.62. Standard uncertainties: u(pH) = 0.01, u(T) = 0.1 K, u(I) = 0.001 mol·kg⁻¹.

constant, but the acidity is higher than erythritol and sorbitol, probably because of a greater delocalization of the charge due to aromatoid heterocycle.

• The stability of the complexes of the different systems can be explained considering the ML species; for example, in the case of SnL species, the stability constant for erythritol and sorbitol is the same, $\log K_{SnEry} = 2.61$ and $\log K_{SnSorb} =$ 2.63, while, for maltol and ethylmaltol, it is higher than the first two ligands; in fact, $\log K_{SnMalt} = 10.66$ and $\log K_{SnEMalt} = 10.83$. For all of the ligands, the interaction with the metal cations is through the oxo group, but in the case of the pyranones, the major stability of the complexes probably is due to the electronic interaction with the ring system. The stability of the ML species that maltol forms with Ca^{2+} , Zn^{2+} , and Sn^{2+} follows the trend SnMalt (10.66) > ZnMalt (5.598) > CaMalt (2.33).

• The sequestering ability of the different ligands has been evaluated by $pL_{0.5}$ parameters: for maltol systems, at T = 298.15 K in NaNO_{3(aq)}, I = 0.151 mol·kg⁻¹ and pH 7.4, the sequestration trend toward Ca²⁺, Sn²⁺, and Zn²⁺ cations is 1.27 < 3.94 < 4.60, respectively. Under these experimental conditions, comparing the pL_{0.5} values of the Zn²⁺/maltol, Zn²⁺/ethylmaltol, and Zn²⁺/erythritol systems, the zinc sequestration by maltol and ethylmaltol is the same, with pL_{0.5} = 4.60 and 4.56, respectively, while for erythritol this ability is low, pL_{0.5} = 1.72.

AUTHOR INFORMATION

Corresponding Author

Rosalia Maria Cigala – Dipartimento di Scienze Chimiche, Biologiche, Farmaceutiche e Ambientali, Università di Messina, 98166 Messina, Italy; © orcid.org/0000-0003-2054-9191; Email: rmcigala@unime.it

Authors

- Francesco Crea Dipartimento di Scienze Chimiche, Biologiche, Farmaceutiche e Ambientali, Università di Messina, 98166 Messina, Italy; o orcid.org/0000-0002-9143-9582
- **Concetta De Stefano** Dipartimento di Scienze Chimiche, Biologiche, Farmaceutiche e Ambientali, Università di Messina, 98166 Messina, Italy
- Anna Irto Dipartimento di Scienze Chimiche, Biologiche, Farmaceutiche e Ambientali, Università di Messina, 98166 Messina, Italy
- **Demetrio Milea** Dipartimento di Scienze Chimiche, Biologiche, Farmaceutiche e Ambientali, Università di Messina, 98166 Messina, Italy; o orcid.org/0000-0003-1188-8837
- Silvio Sammartano Dipartimento di Scienze Chimiche, Biologiche, Farmaceutiche e Ambientali, Università di Messina, 98166 Messina, Italy

Complete contact information is available at: https://pubs.acs.org/10.1021/acs.jced.0c00120

Notes

The authors declare no competing financial interest.

ACKNOWLEDGMENTS

The authors thank Procter & Gamble Ltd. for financial support.

REFERENCES

 Livesey, G. Health potential of polyols as sugar replacers, with emphasis on low glycaemic properties. *Nutr. Res. Rev.* 2003, *16* (2), 163.
 Food additives. https://www.efsa.europa.eu.

(3) Shindou, T.; Sasaki, Y.; Miki, H.; Eguchi, T.; Hagiwara, K.; Ichikawa, T. Determination of erythritol in fermented foods by high performance liquid chromatography. *Shokuhin Eiseigaku Zasshi* **1988**, 29 (6), 419.

(4) Moon, H. J.; Jeya, M.; Kim, I. W.; Lee, J. K. Biotechnological production of erythritol and its applications. *Appl. Microbiol. Biotechnol.* **2010**, *86*, 1017.

(5) De Cock, P.; Bechert, C. L. Erythritol. Functionality in noncaloric functional beverages. *Pure Appl. Chem.* **2002**, *74*, 1281.

(6) Munro, I. C.; Berndt, W. O.; Borzelleca, J. F.; Flamm, G.; Lynch, B. S.; Kennepohl, E.; Bar, E. A.; Modderman, J. Erythritol: An interpretive summary of biochemical, metabolic, toxicological and clinical data. *Food Chem. Toxicol.* **1998**, *36* (12), 1139.

Journal of Chemical & Engineering Data

pubs.acs.org/jced

(7) Den Hartog, G. J. M.; Boots, A. W.; Adam-Perrot, A.; Brouns, F.; Verkooijen, I. W. C. M.; Weseler, A. R.; Haenen, G. R. M. M.; Bast, A. Erytritol is a sweet antioxidant. *Nutrition* **2010**, *26*, 449.

(8) Kawanabe, J.; Hirasawa, M.; Takeuchi, T.; Oda, T.; Ideda, T. Noncariogenicity of erythritol as a substrate. *Caries Res.* **1992**, *26*, 358.

(9) Cervera, M. F.; Heinämäki, J.; Krogars, K.; Jörgensen, A. C.; Karjalainen, M.; Iraizoz Colarte, A.; Yliruusi, J. Solid-state and mechanical properties of aqueous chitosan-amylose starch films plasticized with polyols. *AAPS PharmSciTech* **2004**, 5 (1), 109.

(10) Krogars, K.; Heinämäki, J.; Karjalainen, M.; Rantanen, J.; Luukkonen, P.; Yliruusi, J. Development and characterization of aqueous amylose-rich maize starch dispersion for film formation. *Eur. J. Pharm. Biopharm.* **2003**, *56*, 215.

(11) Burggraaf, J.; Schoemaker, R. C.; Lentjes, E. G. W. M.; Cohen, A. F. Sorbitol as a marker for drug-induced decreases of variable duration in liver blood flow in healthy volunteers. *Eur. J. Pharm. Sci.* **2000**, *12*, 133.

(12) Featherstone, S. A Complete Course in Canning and Related Processes (Fourteenth Edition). *Microbiology, Packaging, HACCP and Ingredients* **2015**, *2*, 64.

(13) Gralla, E. J.; Stebbins, R. B.; Coleman, G. L.; Delahunt, C. S. Toxicol. Appl. Pharmacol. 1969, 15 (3), 604.

(14) Hironishi, M.; Kordek, R.; Yanagihara, R.; Garruto, R. M. Maltol (3-hydroxy-2-methyl-4-pyrone) Toxicity in Neuroblastoma Cell Lines and Primary Murine Fetal Hippocampal Neuronal Cultures. *Neurodegeneration* **1996**, 5 (4), 325.

(15) Yasumoto, E.; Nakano, K.; Nakayachi, T.; Morshed, S. R.; Hashimoto, K.; Kikuchi, H.; Nishikawa, H.; Kawase, M.; Sakagami, H. Cytotoxic Activity of Deferiprone, Maltol and Related Hydroxyketones against Human Tumor Cell Lines. *Anticancer Res.* **2004**, *24*, 7.

(16) Amatori, S.; Ambrosi, G.; Fanelli, M.; Formica, M.; Fusi, V.; Giorgi, L.; Macedi, E.; Micheloni, M.; Paoli, P.; Pontellini, R.; Rossi, P. Synthesis, Basicity, Structural Characterization, and Biochemical Properties of Two [(3-Hydroxy-4-pyron-2-yl)methyl]amine Derivatives Showing Antineoplastic Features. J. Org. Chem. 2012, 77 (5), 2207.

(17) Amatori, S.; Bagaloni, I.; Macedi, E.; Formica, M.; Giorgi, L.; Fusi, V.; Fanelli, M. Malten, a new synthetic molecule showing in vitro antiproliferative activity against tumour cells and induction of complex DNA structural alterations. *Br. J. Cancer* **2010**, *103* (2), 239.

(18) Hider, R. C.; Hall, A. D. Clinically useful chelators of tripositive elements. *Prog. Med. Chem.* **1991**, *28*, 41.

(19) Burgess, J. Literature Highlights-35. *Transition Met. Chem.* **1993**, 18, 9.

(20) Saghaie, L.; Badii, S.; Badii, A. Evaluation of the Intestinal Absorption of Maltol-Zn and Ethyl maltol-Zn Complexes in Rat. *J. Rep. Pharm. Sci.* **2015**, *4*.

(21) Shimbo, S.; Matsuda-Inoguchi, N.; Watanabe, T.; Sakurai, K.; Date, C.; Nishimura, A.; Nakatsuka, H.; Saito, H.; Arisawa, K.; Ikeda, M. Dietary intake of tin in Japan, and the effects on intake of canned food and beverage consumption. *Food Addit. Contam.* **2007**, *24* (5), 535–545.

(22) Blunden, S.; Wallace, T. Tin in canned food: a review and understanding of occurrence and effect. *Food Chem. Toxicol.* 2003, 41 (12), 1651–1662.

(23) Flaschka, H. A. *EDTA Titration*; Pergamon Press: London, 1959. (24) Cigala, R. M.; Crea, F.; De Stefano, C.; Irto, A.; Sammartano, S. Use of Gantrez Copolymers as Potential Chelating Agent for the Selective Sequestration of Metal Ions. Studies of the Interactions in Aqueous Solution at Different Ionic Strengths and Temperatures. *J. Chem. Eng. Data* **2018**, 63 (11), 4193–4204.

(25) De Stefano, C.; Princi, P.; Rigano, C.; Sammartano, S. Computer Analysis of Equilibrium Data in Solution. ESAB2M: An Improved Version of the ESAB Program. *Ann. Chim. (Rome)* **1987**, *77*, 643–675.

(26) De Stefano, C.; Foti, C.; Giuffrè, O.; Mineo, P.; Rigano, C.; Sammartano, S. Binding of Tripolyphosphate by Aliphatic Amines: Formation, Stability and Calculation Problems. *Ann. Chim. (Rome)* **1996**, *86*, 257–280. (27) De Stefano, C.; Sammartano, S.; Mineo, P.; Rigano, C. Computer Tools for the Speciation of Natural Fluids. In *Marine Chemistry - An Environmental Analytical Chemistry Approach*; Gianguzza, A., Pelizzetti, E., Sammartano, S., Eds.; Kluwer Academic Publishers: Amsterdam, The Netherlands, 1997; pp 71–83.

(28) De Stefano, C.; Mineo, P.; Rigano, C.; Sammartano, S. Ionic Strength Dependence of Formation Constants. XVII. The Calculation of Equilibrium Concentrations and Formation Constants. *Ann. Chim.* (*Rome*) **1993**, 83, 243–277.

(29) Alderighi, L.; Gans, P.; Ienco, A.; Peters, D.; Sabatini, A.; Vacca, A. Hyperquad simulation and speciation (HySS): a utility program for the investigation of equilibria involving soluble and partially soluble species. *Coord. Chem. Rev.* **1999**, *184*, 311–318.

(30) Gaidamauskas, E.; Norkus, E.; Vaiciuniene, J.; Crans, D. C.; Vuorinen, T.; Jaciauskiene, J.; Baltrunas, G. Evidence of two-step deprotonation of D-mannitol in aqueous solution. *Carbohydr. Res.* **2005**, 340 (8), 1553–1556.

(31) Vicedomini, M. Acid dissociation of some polyhydroxy compounds in strongly alkaline solution. *Ann. Chim. (Rome)* **1981**, 213–222.

(32) Thamsen, J. The Acidic Dissociation Constants of Glucose, Mannitol and Sorbitol, as Measured by Means of the Hydrogen Electrode and the Glass Electrode at 0° and 18 degrees°C. *Acta Chem. Scand.* **1952**, *6*, 270–284.

(33) Bretti, C.; Cigala, R. M.; De Stefano, C.; Lando, G.; Sammartano, S. Potentiometric determination of some solution thermodynamic parameters of three hydroxypyrone derivates. Effect of the ionic medium, ionic strength and temperature on the acid-base properties. *Int. J. Electrochem. Sci.* **2013**, *8*, 10621–10649.

(34) Cardiano, P.; Cigala, R. M.; Crea, F.; De Stefano, C.; Giuffrè, O.; Sammartano, S.; Vianelli, G. Potentiometric, UV and 1H NMR study on the interaction of penicillin derivatives with Zn(II) in aqueous solution. *Biophys. Chem.* **2017**, *223*, 1–10.

(35) Cigala, R. M.; Crea, F.; De Stefano, C.; Lando, G.; Milea, D.; Sammartano, S. The inorganic speciation of tin(II) in aqueous solution. *Geochim. Cosmochim. Acta* **2012**, *87*, 1–20.

(36) De Stefano, C.; Foti, C.; Sammartano, S.; Gianguzza, A.; Rigano, C. Equilibrium Studies in Natural Fluids. Use of Synthetic Seawater and Other Media as Background Salts. *Ann. Chim. (Rome)* **1994**, *84*, 159–175.

(37) Guggenheim, E. A.; Turgeon, J. C. Specific interaction of ions. *Trans. Faraday Soc.* **1955**, *51* (0), 747–761.

(38) Scatchard, G. Concentrated Solutions of Strong Electrolytes. *Chem. Rev.* **1936**, *19* (3), 309–327.

(39) Martell, A. E.; Smith, R. M.; Motekaitis, R. J. *Critically Selected Stability Constants of Metal Complexes*; National Institute of Standard and Technology, NIST, PC-based Database: Gaithersburg, MD, 2004.

(40) Pettit, L. D.; Powell, K. J. IUPAC Stability Constants Database. Academic Software, IUPAC: 2001.

(41) Jakusch, T.; Gajda-Schrantz, K.; Adachi, Y.; Sakurai, H.; Kiss, T.; Horváth, L. Solution equilibrium characterization of insulin-mimetic Zn(II) complexes. *J. Inorg. Biochem.* **2006**, *100*, 1521–1526.

(42) Crea, F.; De Stefano, C.; Foti, C.; Milea, D.; Sammartano, S. Chelating agents for the sequestration of mercury(II) and monomethyl mercury(II). *Curr. Med. Chem.* **2014**, *21* (33), 3819–3836.