

The Sequences of Human and Bovine Genes of the Phosphate Carrier from Mitochondria Contain Evidence of Alternatively Spliced Forms*

(Received for publication, December 8, 1993)

Vincenza Dolce‡, Vito Iacobazzi‡§, Ferdinando Palmieri‡, and John E. Walker§¶

From the ‡Department of Pharmaco-Biology, Laboratory of Biochemistry and Molecular Biology, University of Bari, Bari 70125, Italy and the §Medical Research Council, Laboratory of Molecular Biology, Hills Road, Cambridge CB2 2QH, United Kingdom

The sequences of the human and bovine genes for the phosphate carrier from the inner membranes of mitochondria have been determined. The genes have similar structures and each is divided into nine exons. In both genes, two exons, named IIIA and IIIB, are closely related, and they appear to be alternatively spliced. The human exon IIIB sequence is found in a published human heart cDNA sequence, and bovine exon IIIA forms part of a published bovine heart cDNA sequence. By further examination of the human heart cDNA library, sequences arising from both alternatively spliced forms of the phosphate carrier have been characterized. Both forms were also found in several bovine tissues, but the ratios of expression of the two forms varied. The form containing exon IIIA was expressed most highly in bovine heart and liver, less highly in brain and kidney, and only in low amounts in lung. The opposite hierarchy was found for the form containing exon IIIB; it was most highly expressed in lung and least in heart and liver. The alternative splicing mechanism affects amino acids 4–45 of the mature phosphate carrier protein, which is believed to form one of six transmembrane segments of the phosphate carrier and to emerge into a large extramembranous loop. The alternative splicing mechanism changes 13 and 11 amino acids in the human and bovine carrier proteins, respectively. As the function of this region of the phosphate carrier is not known, the effects of the changes on carrier function are not understood at present.

The inner membranes of mitochondria contain a number of proteins that are responsible for the transport of various metabolites back and forth (Krämer and Palmieri, 1989, 1992; Walker, 1992; Walker and Runswick, 1993). An example is provided by the phosphate carrier, which catalyzes the transport of phosphate into the mitochondrial matrix, either by proton co-transport or in exchange for hydroxyl ions. The purified protein from both pig and bovine heart (Bisaccia *et al.*, 1984; Kolbe *et al.*, 1984) has been reconstituted into liposomes in an active form. The primary structure of the bovine phosphate carrier is

313 amino acids long and contains three related segments, each of about 100 amino acids arranged in tandem (Runswick *et al.*, 1987). The rat, human, *Saccharomyces cerevisiae*, and *Caenorhabditis elegans* sequences have similar features (Ferreira *et al.*, 1989; Dolce *et al.*, 1991; Phelps *et al.*, 1991; Runswick *et al.*, 1994). These repetitive elements are related to those found in the other characterized members of the mitochondrial carrier family, namely, the ADP/ATP (Saraste and Walker, 1982) oxoglutarate-malate (Runswick *et al.*, 1990) and citrate carriers (Kaplan *et al.*, 1993), and the uncoupling protein from brown fat (Aquila *et al.*, 1985). They are also found in a number of other proteins of known sequence but of unknown function, which therefore belong to the same protein superfamily (Walker, 1992; Runswick *et al.*, 1994). Among the family members, the mammalian, but not the yeast, phosphate carrier is exceptional in having a processed N-terminal sequence that helps to target the protein into mitochondria (Runswick *et al.*, 1987; Dolce *et al.*, 1991; Phelps *et al.*, 1991; Zara *et al.*, 1992; Ferreira *et al.*, 1989). By examination of the transmembrane topography of the phosphate carrier in the inner mitochondrial membrane, it has been shown that the N-terminal and the C-terminal regions of the phosphate carrier both protrude toward the cytosol. Therefore, its polypeptide chain spans the membrane an even number of times (Capobianco *et al.*, 1991; Palmieri *et al.*, 1993).

It is known that isoforms encoded by different ADP/ATP carrier genes are present in several species. Two bovine and three human genes have been detected for ADP/ATP carrier (Walker *et al.*, 1987a; Battini *et al.*, 1987; Neckelmann *et al.*, 1987; Houldsworth and Attardi, 1988; Cozens *et al.*, 1989; Powell *et al.*, 1989). Three isoforms have been characterized in *S. cerevisiae* (Gawaz *et al.*, 1990; Kolarov *et al.*, 1990). Single genes for the phosphate and oxoglutarate carriers were detected in the human and bovine genomes by Southern blotting (Runswick *et al.*, 1987, 1990), and there is a unique gene for the uncoupling protein (Ricquier *et al.*, 1991).

In this paper, the sequences of the genes of the human and bovine phosphate carriers are described. The human sequence has been determined from genomic clones obtained by screening a human genomic library with two human cDNA probes. The bovine genomic sequence has been derived from genomic fragments generated by polymerase chain reactions, using primers and probes based upon the bovine cDNA (Runswick *et al.*, 1987). The two genes are spread over 7.9 and 6.1 kb¹ of human and bovine DNA, respectively. Both contain nine exons separated by eight introns. The sequences show that there are two related exons IIIA and IIIB in each gene that appear to be alternatively spliced. The two alternatively spliced forms have

* This work was supported in part by the Consiglio Nazionale delle Ricerche Target Project "Ingegneria Genetica" and the Ministero dell'Università e della Ricerca Scientifica e Tecnologica (MURST). The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked "advertisement" in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

The nucleotide sequence(s) reported in this paper has been submitted to the GenBank™/EMBL Data Bank with accession number(s) X77337 and X77338.

¶ To whom correspondence should be addressed. Fax: 44-223-412178.

¹ The abbreviation used is: kb, kilobase(s).

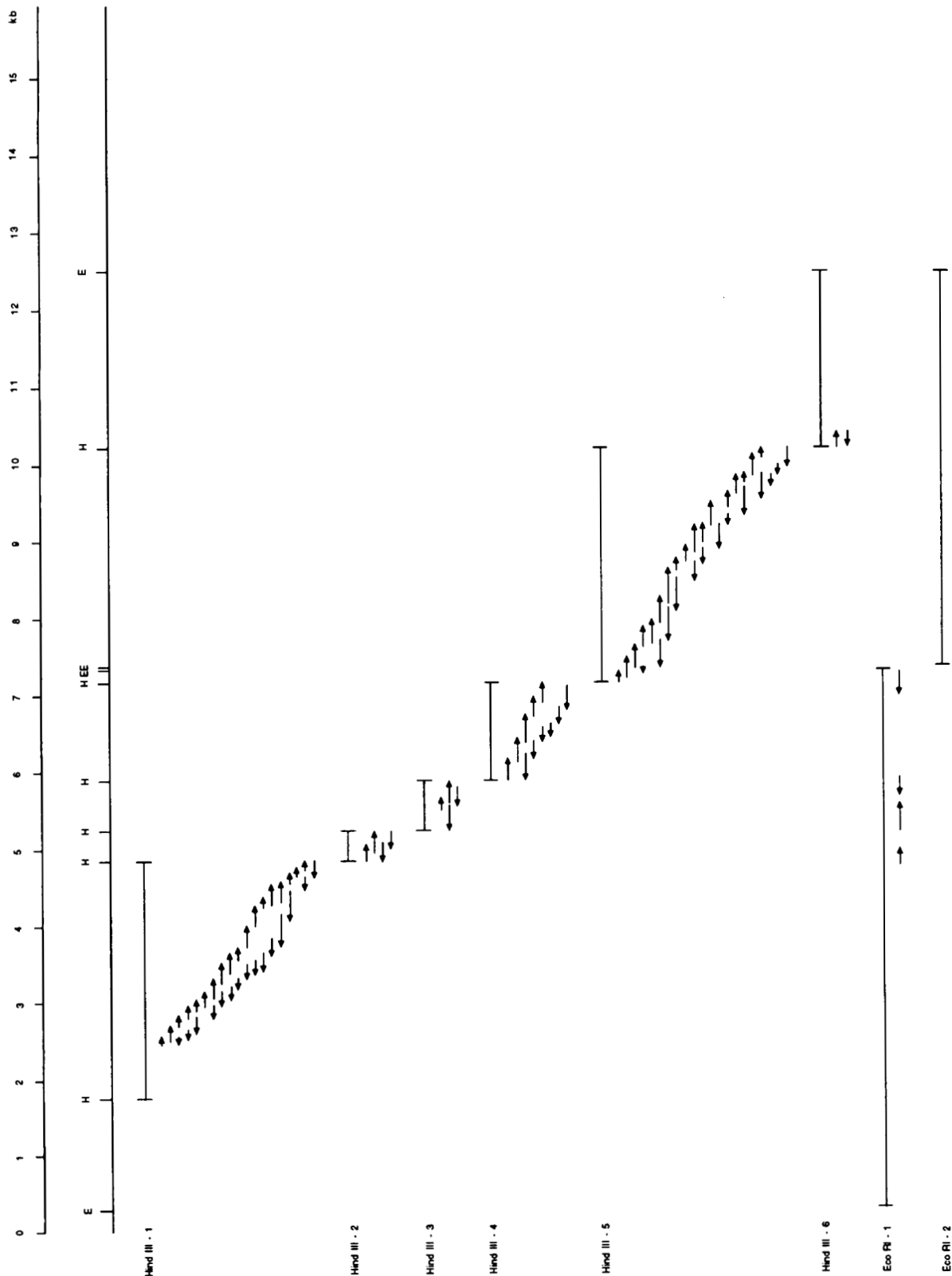


FIG. 1. **Sequence analysis of the human gene for the mitochondrial phosphate carrier.** The restriction map of the λ 12D3 clone is shown at the top, where E and H represent EcoRI and HindIII restriction sites, respectively. The sequence was determined by sequencing fragments prepared by subdigesting two EcoRI fragments (6.7 and 5.0 kb) with HindIII. The HindIII sites were overlapped by priming on the intact EcoRI fragments with appropriate oligonucleotides. The two EcoRI fragments were joined by the sequence of an overlapping HindIII fragment (nucleotides 4710-7769 in Fig. 2). The arrows denote the directions and the extents of the individual DNA sequences from which the complete sequence was compiled.

been characterized from a human cDNA library, and their expression has been investigated in various bovine tissues.

MATERIALS AND METHODS

Synthetic Oligonucleotides—Oligonucleotide primers based on the bovine phosphate carrier cDNA were synthesized in an Applied Biosystems model 320B DNA synthesizer. Some were used as primers in polymerase chain reactions, and others as hybridization probes with the products. Forward and reverse primers 25 bases in length were made with an appropriate 5'-linker (*EcoRI*, *HindIII*, *BamHI*, or *XbaI*). Oligonucleotides were also employed as primers in sequencing reactions. The purification, radiolabeling, and use as hybridization probes of synthetic oligonucleotides have been described before (Powell *et al.*, 1989; Walker *et al.*, 1989).

Isolation of Human Genomic Clones—The human genomic library consisted of partial *Sau3A* fragments cloned into λ 2001 (LeFranc *et al.*, 1986). About 7×10^5 recombinants were screened by plaque hybridization (see Dyer and Walker (1993)). Duplicate filters were screened with two different probes; one was derived from the 5' end of the human phosphate carrier cDNA (nucleotides 1–562), and the other from the 3' end of the cDNA for the human phosphate carrier (nucleotides 612–1114) (Dolce *et al.*, 1991). DNA was purified by glycerol step gradient centrifugation (Garber *et al.*, 1983) from two positively hybridizing recombinant phages, λ 12D3 and λ 2F2. Restriction fragments from these phages were cloned into M13mp18 or M13mp19.

Amplification of Bovine Genomic DNA Sequences—Overlapping segments of bovine genomic DNA from liver (Walker *et al.*, 1987b) were amplified in four polymerase chain reactions (Walker *et al.*, 1992; Iacobazzi *et al.*, 1992), using synthetic oligonucleotide primers (with appropriate linkers) and probes based on the bovine cDNA sequence (Runswick *et al.*, 1987). These oligonucleotides are listed in Table I. The reactions were carried out for 30 cycles and, after addition of fresh portions of enzyme and primers, for an additional 30 cycles. The reaction products were analyzed on 1.4% high melting agarose gels. The DNA fragments were transferred to Hybond-N membranes (Amersham International, Amersham, United Kingdom), cross-linked to membranes by irradiation with uv light, and hybridized with radioactively labeled synthetic oligonucleotides at 5 °C below the minimum dissociation temperature. Fragments that hybridized with the probes were recovered from the gel by the Gene Clean procedure (Bio 101, La Jolla, CA), and the products were cloned into M13mp18 and M13mp19 vectors.

Characterization of Human cDNAs—A human cDNA library derived from cardiac muscle cloned in λ gt11 was kindly donated by Dr. E. R. M. McCabe. This library was screened as described previously (Dolce *et al.*, 1991) with sequences derived from the bovine cDNA for the phosphate carrier (Runswick *et al.*, 1987). These sequences were an *EcoRI-HindIII* fragment and a *PstI* fragment corresponding to nucleotides 628–1130 and 104–339 of the bovine sequence, respectively.

Polymerase Chain Reactions with Bovine mRNAs—Poly(A⁺) RNA was prepared from total RNA from bovine heart, liver, brain, kidney and lung (Chirgwin *et al.*, 1979; Viñas *et al.*, 1990). Samples of poly(A⁺) RNA (2 μ g) from each tissue were employed as templates in polymerase chain reactions carried out as described above, except that the annealing temperature was 50 °C. The primers from exon IIIA (IIIA-F and IIIA-R) and from exon IIIB (IIIB-F and IIIB-R) were 19 bases long (see Fig. 4 for their sequences). Two sets of the amplified products from each tissue were analyzed on an agarose gel, transferred to Hybond membranes, hybridized with the 16-mer oligonucleotide probes (IIIA-P and IIIB-P), and autoradiographed at room temperature for 1 h. The products from heart and liver were recovered from the gel, cloned, and sequenced.

DNA Sequence Analysis—DNA sequences were determined by the modified dideoxy chain termination method (Sanger *et al.*, 1977; Biggin *et al.*, 1983) with T7 DNA polymerase (Sequenase, U. S. Biochemical Corp.). Sequencing reactions were primed with either the 17-mer universal primers or the primers used in the polymerase chain reactions. Other 17-mer synthetic primers were used to extend existing sequences. Both DNA strands were sequenced completely. In order to avoid sequence errors being introduced by the polymerase chain reaction, at least three independent M13-isolated clones were sequenced from each reaction. When an ambiguity was observed, further clones were sequenced. Compressions were resolved by the use of dITP instead of dGTP in sequencing reactions. Data bases were compiled and analyzed with the computer program DBUTIL (Staden, 1982).

RESULTS AND DISCUSSION

Cloning and Sequencing the Human and Bovine Genes for the Phosphate Carrier—Two independent and overlapping

TABLE I

Synthetic oligonucleotide primers and probes used to generate partial overlapping clones comprising the bovine gene for the mitochondrial phosphate carrier protein

Their sequences were based on the bovine cDNA sequence. F, R, and P denote forward and reverse primers and probes, respectively, employed in polymerase chain reactions PCR-1 to PCR-4.

Oligonucleotide	Position in bovine cDNA
1F	1–25
1R	285–310
1P	194–211
2F	250–275
2R	445–470
2P	340–357
3F	420–445
3R	847–862
3P	735–752
4F	822–847
4R	1300–1325
4P	973–990

genomic clones, λ 12D3 and λ 2F2, were isolated from a human genomic library. Together they cover about 26 kb of genomic sequence. It was shown by Southern blot analysis that λ 12D3 contained the entire phosphate carrier gene, and by sequencing appropriate restriction fragments from this phage (see Fig. 1), a sequence of about 7,969 bases encompassing the human phosphate carrier gene was established (see Fig. 2).

In the case of the bovine gene, a different strategy was adopted. Four overlapping sequences about 1.0–1.8 kb in length (PCR-1 to PCR-4 in Fig. 3) were amplified by the polymerase chain reaction using bovine genomic DNA as template, and with synthetic oligonucleotide primers based on the bovine cDNA sequence (see Table I and legend to Fig. 3). The sequences of PCR-1 to PCR-4 were then determined with the use of synthetic primers. In this way, a bovine genomic sequence of 6,161 nucleotides containing the phosphate carrier gene was determined (see Fig. 4).

Gene Structures—The exons of the human and bovine genes for the phosphate carrier were identified by comparison of the genomic sequences with the human and bovine cDNA sequences (Dolce *et al.*, 1991; Runswick *et al.*, 1987). Consensus rules for splice sites that predict conservation of the dinucleotides GT and AG, respectively, next to the 5' and 3' boundaries of the introns, were also taken into consideration (Breathnach and Chambon, 1981). In this way, it was found that the human gene contains nine exons separated by eight introns, and that the bovine gene has the same organization. The introns in the bovine gene are mostly smaller than those in the human gene (Fig. 5). The 3' extremities of both genes were established by the cDNA sequences, but the transcriptional start sites of the human and bovine phosphate carrier genes have not been mapped. Therefore, the 5' extremity of neither of exons I is known, and it is unlikely that the rather short 5'-noncoding sequences present in the cDNAs represent the full extent of these exons.

The eight introns in both genes interrupt the coding sequences at exactly the same positions. In other members of the carrier family, it has been noted that the introns tend to interrupt the coding sequences in or near to the extramembranous loops (Cozens *et al.*, 1989; Krämer and Palmieri, 1992). A similar tendency is found in the phosphate carrier genes with introns B, C, D, E, F, and H. However, intron G appears to be in the middle of an α -helical segment.

Alternative Splicing of Exons IIIA and IIIB—The most striking feature, uncovered by the comparison of the human and bovine genomic sequences with the corresponding cDNAs, was that both genes contain evidence of alternative splicing. Exon

AGTCTTGTCTGCTGCCAGGCTGGAGTGTAGTGGTCCATGGTCTCACTGCAACCTCCACCTCCCTCAGGCTCAAGCGATTGTTCTGTCTCACCTCACCTCCCGAGTAGCTGGAT 3840

TACAGGTGCACGCCACATGCCCGTCCAATTTTTTATATTTTAGTAGAGATGGGTTTCACTGTGTGCCAGGCTGGTCTGAACTCCTGAGCTCAGGTAATCCACCTGCCCTCAGCCTCC 3960

CAAAGCGATTGGGATTACAGATGTGAGCCACCGTGACCTGGCAGGAATTACTGTAGTTACCTTTTGTGTAGTTGCTGAAATTTATATGGAACCCAAACAAAATAATCATGAGATTAAT 4080

50 Exon IV 60

TTTATGTAGCTGTTGGTGCATTAAATTTTTTTTCTGTGTTTAAATAAAGTGGACCCCAAAAGTACAAGGCATATTTAACGGATTCTCAGTTACACTTAAAGAGGATGGTCT 4200

70 80 90 100

CGTGGTTGGCTAAAGAGTAGGGCTCCGACTTCTCTGGCTACTCCATGCAGGACTCTGCAAGTTTGGCTTTTATGAGTCTTTAAAGTCTTGTATAGCAATATGCTTGGAGAGTATGT 4320

AATTAACCTTAAATGAATGTTCCGAGTGTAAAGACTTCCGAGTGTCTTAGATTTTTGTCTGTCTGCCTATTTAGCACTGAAGAAAATGGTCTACAAGTGGCAACTGTTTT 4440

AATCGTATGCCTGTGTGAGCAGAGACAGGTTGATAAATGTATTCAATTAATACAATTCCTTTTTAATATACATCTGTTTTCTAAGTAAGTTGGATTTTGTAGTTTTTATGAGAAGGG 4560

CTCTGGGACTCCTTGGTAGTCCGGGGCATGGCATCTAGATATTTTTGAATATTCATGTTGTTTACATAGTTACGTGTTTTGTTAATGAGTTATGAGACCACATATATATCCATA 4680

TCATCGTGTACTTTATAAAATGTCTGAAGCTTAGTTGTTTTCTAGAACTGTCACCTTCAATGAAAACCAACAACAACAATTCACATCCCTTCTGTGTTTTGGATTTTAGGAGA 4800

110 Exon V 120 130 140

N T Y L W R T S L Y L A A S A S A E F F A D I A L A P M E A A K V R I Q T Q P G 4920

ATACTTATCTCTGGGCACATCACTATATTTGGCTGCCCTGCTGCAAGTCTTGTGACATTTGCCCTGCTCCTATGGAAGCTGCTAAGGTTCAAGTTCGAATCAAACCCAGCAGGTT

150 160

Y A N T L R D A A P K M Y K E E G L K A 5040

ATGCCAACCTTTGAGGATGCAGCTCCCAAAATGTATAAGGAAGAAGCCATAAAGCTAAGTAAACACTTAAAAATTTATACTATGAAAGTACTTATTTAAGTGAACCTCATTTTTT

TTTGTTTTTTTTTTGTGTTTTTTTGTGACGAAAAGTCTCGCTCTGTCCACAGGCTGGAGTGCAGTGGCTGATCTCGGCTCACTGCAAGCTCCGCTCCCGGTTACGCCCATTTCTCC 5160

TGCCCTCAATCCCAAGAGCTAGGACTACAGGCGCTGCACTGCCAGCTAATTTTTGTATTTTTTTCAGTATAGACGGAGTTTCAACGCTCCTCACTCCTGACCTCATGATCCGCCACCT 5280

TGGCCTCCCAAAATGCTGGGATTACAGGTGTGAGCCACCAGCACCCGGCTCTTTTTTTTTTTTCTTTGAGACGGAGTTTGTCTTTGTTGCCAGGCTGGATGCAATGGCATATCTCA 5400

GCTCACTGCAACCTCCGCTCCAGGTTCAAGTGTGTCTGCTCAGCTGCTGAATAGCTGGGATTACAGGCATCACACCACGCTGGCTAATTTTGTATTTTTAGTAGAGACTCCAT 5520

GTTGGTCAGGCTGGTCTCGAATCCTGACCTCAGTGTCCACCGCTCGCCTCCCAAGTGTGGGATTACAGGCTGAGACACCGCTCTGGCTGTGAACCTCATTTTTTAGTATTAAG 5640

CCTACCAGACTTTTTAAAAAGACTATATAACATTTAATGCTATTTACCTGTATGAGAACAATAACTTGAGAGGTAAGATAAGTTAATACATGGTTTGACCTGGACTGAGTTTTT 5760

TTTTTTCCCTAGAGTAAAGACCATTGTGCGTCTAGAAATGACTTGTATGTTGGCTAAACATTTACTATTAACCTTTTAGGGAGGCTTGTAGTATCCTTGTGATCTATTACAGTTA 5880

AGAATTTCCATGACCACAACATGCTTCTTAGATCCACCTTTGTGGATGAATCTTGAACCTGAGTTCACCTTGTAAACTTCTGTTTCTTGTGGTTCCAGTAGTCAAAGAAACATCCAGCA 6000

ACTTTTTGTTGTATAGTCAAAGGCTGTTGAGTCAATGGCATGTAAGAGAAATATACTGCATGTTAGTCTAACGTATCTGATAGAAATGACATGCATTTACTGGCCATTTGTTACTAT 6120

170

CAGGACTCGACTCGTGTGGGACATTTCTGTTAATAATGACAGTCTCTGATAGATGAGTGTATGCAACTGTGTAACAAGTCTCTGATTTCCTAGATTCTACAAGGGGTGCTCTCC 6240

F Y K G V A P

Exon VI 180 190 200 210

L W M R Q I P Y T M M K F A C F E R T V E A L Y K F V V P K P R S E C S K P E Q 6360

TCTGGATGAGACAGATACCATAACCATGATGAAGTTCGCTGCTTGAACCTACTGTTGAAGCACTGTACAAGTTTGTGGTTCTAAGCCCGCAGTGAATGTTCAAAGCCAGAGCAGC

220

L V V T F V A G Y I 6480

TGGTGTAAACATTTGTAGCAGGTACATAGTTACGAATTACTTAGAACACACTTGTCTGAAATATGAAACAATAATCATTCCATTAITGGCGTTTTTGTAGAGGGAAAAATACTCCAA

AGGTTATAAAGCAGTGTTTTTGTTTTTGGTTTCTGAGGCATAGTCTCGCTCTGTGCCAGGCTGGAGTGCAGTGGCATATAATAGCTCACTGCAACCTCCTCCTCGGTTCAA 6600

GTGATTTCTTCCCGCAGCCTCCCAAGTAGCTGGGACAGGCATGCACCGCCATGCCGCGTATTTTTGTTTTTGTTTTTTTTTTGTAGATGGAGTTTTGCCCTTGTGCCAGG 6720

CTGGAGTGCAGTGGTGAATCTTGGCTACTGCAGCCTCCGCTCCTGGTTCAGCGATITCTCTGCTCAGCCTCCTGAGTAGCTGGGATTGCAGGCTCTGCCTACGCCCCGCTAA 6840

TTTTAGTATTTTTAGTAGAGAGCGTTTCACTATGTTGCCAGGCTGTTCAAACTCCTGGGCAAGTGTATGCTGCTCCGCTCCCAAGTGTGGATTAACAGCGTGCAGCCAC 6960

CGCCGACCTGGCCTGATTTTTGTAAAGATGGGTTTCAACATGTTGATGAGTCTGTTCTCAAACTCCTGACCTCAGGTGATGCTGCTGCTTAAAGCAGTTTTATCTTCCATAACAT 7080

TTTATATGCTTTTTAGGATTCATAAATACTTGTAACTAGTCTCTAAAAGGCTGTTTCACTGGGATGTCATTAITAGGCTTGGGAATGTATTGATCTTACTGTTGCCACCATTTTTTGT 7200

GTTGTTTTAAGAATTTATACAGAAAGTGTGAATAAAATCTGAAAACCTTTAAAAGATGTTCCGTTTTACATAGATAGACAGCTTGATAACTGTACCCTGTACCCTGAGTCTGA 7320

Fig. 2—continued

TTTTTATTTTGAAGCTAGAAAAATATTATTTTAAAAATCATAAAAAATGATTATTGGCTCTGTGAAGTTTGTGTTTAACTGGCAGCTGATGGGATCCTTTATCTTTTTTCAGCTGGAGTCT 7440
 A G V
 230 Exon VII 240 250 260
 F C A I V S H P A D S V V S V L N K E K G S S A S L V L K R L G F K
 TTTGTGCAATTGTTTCTCACCCCTGCTGATTCTGTGGTATCTGTGTGAATAAAGAAAAGGTAGCAGTGTCTTCTGGTCCTCAAGAGACTTGGATTAAAGGTAGGATGATGTTTTTTT 7560
 Exon VIII 270
 G V W K G L F A R I I M I G T L T A
 CTTGAAAGAAGAACAACAGTTTGGATATGTTGCATTTTTTTCATTGCTTTTCCTGTTGAACCAAGGTGTATGGAAGGGACTGTTGCCCGTATCATGATTGGTACCCTGACTGC 7680
 280 290 300 310
 L Q W F I Y D S V K V Y F R L P R P P P P E M P E S L K K K L G L T Q *
 ACTACAGTGGTTTATCTATGACTCCGTGAAGTCTACTTCAGACTTCCTCGCCCTCCTCCACCCGAGATGCCAGAGTCTCTGAAGAAGAAGCTTGGGTTAACTCAGTAGTTAGATCAAAG 7800
 CAAATGTGGACTGAATCTGCTTGTGATCAGTGTGTTGAAGAAAGTCAAAGGAAGCTTTTATATATTGACAGTGTAGGAAATGTCTATTCTGATATAATTACTGTAGTACTCTTGCT 7920
 TAAGGCAAGAGTTTCAGATTTACTGTGAAATAAACCCCAACTGTTTCATG 7969

Fig. 2—continued

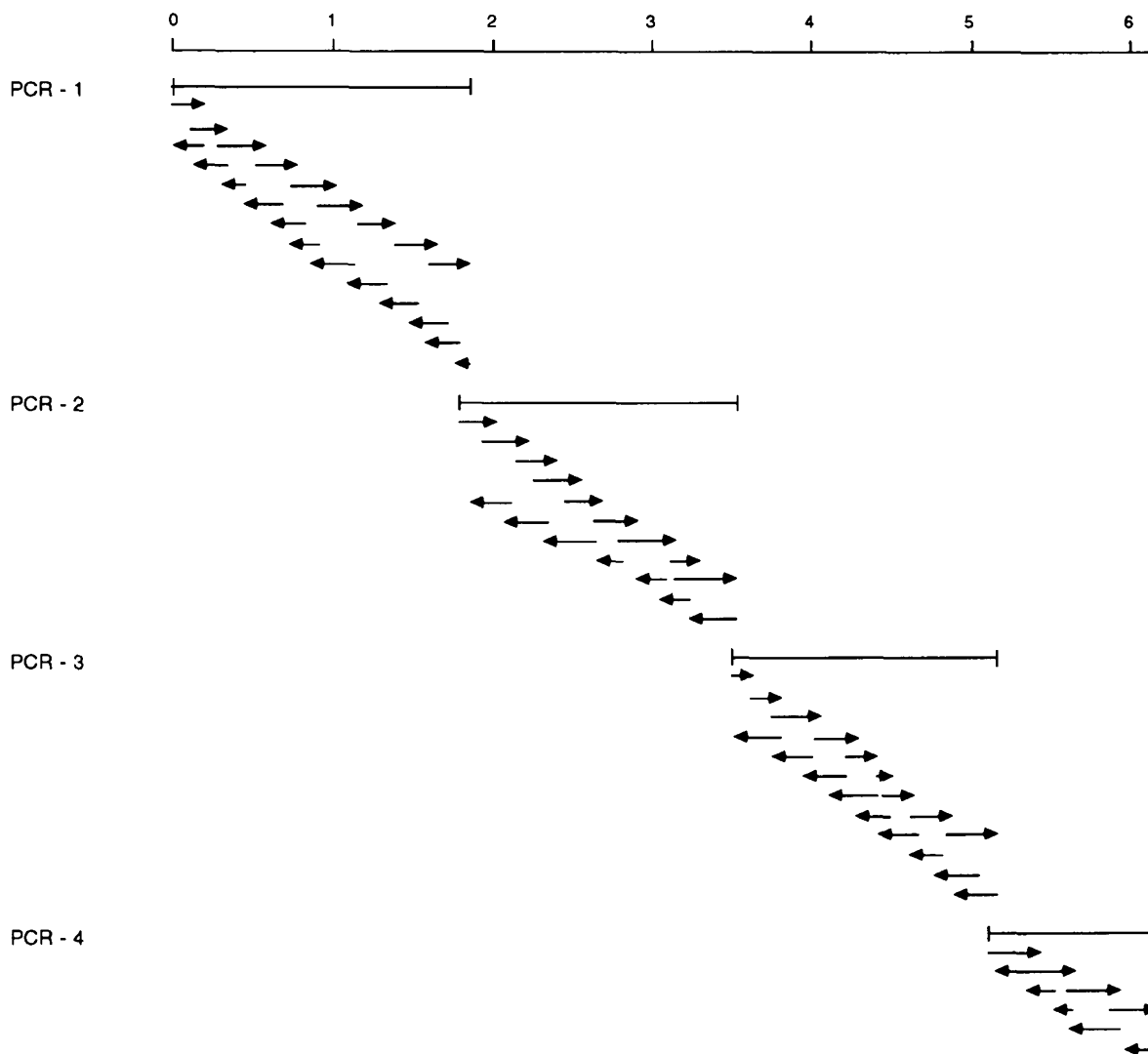


FIG. 3. Generation by polymerase chain reactions and sequence analysis of genomic clones of the bovine phosphate carrier gene. The heavy horizontal lines are proportional to the lengths of these DNA segments, and the arrows represent the directions and the extents of the determined DNA sequences. The primers are listed in Table I.

IIIA in the bovine gene encodes amino acids 4–45 of the known sequence of the bovine phosphate carrier (Runswick *et al.*, 1987), whereas human exon IIIB encodes amino acids 4–45 of the published human phosphate carrier (Dolce *et al.*, 1991). The nucleotide sequence of bovine exon IIIA is identical to nucleotides 220–344 in the published bovine cDNA sequence, and

human exon IIIB is identical to nucleotides 206–327 of the cDNA sequence. The sequences of amino acids 4–44 of the rat phosphate carrier encoded in a cDNA derived from liver (Ferreira *et al.*, 1989) and of the *C. elegans* phosphate carrier are both more closely related to the protein sequence encoded in exon IIIB than to the sequence encoded in exon IIIA. The N-

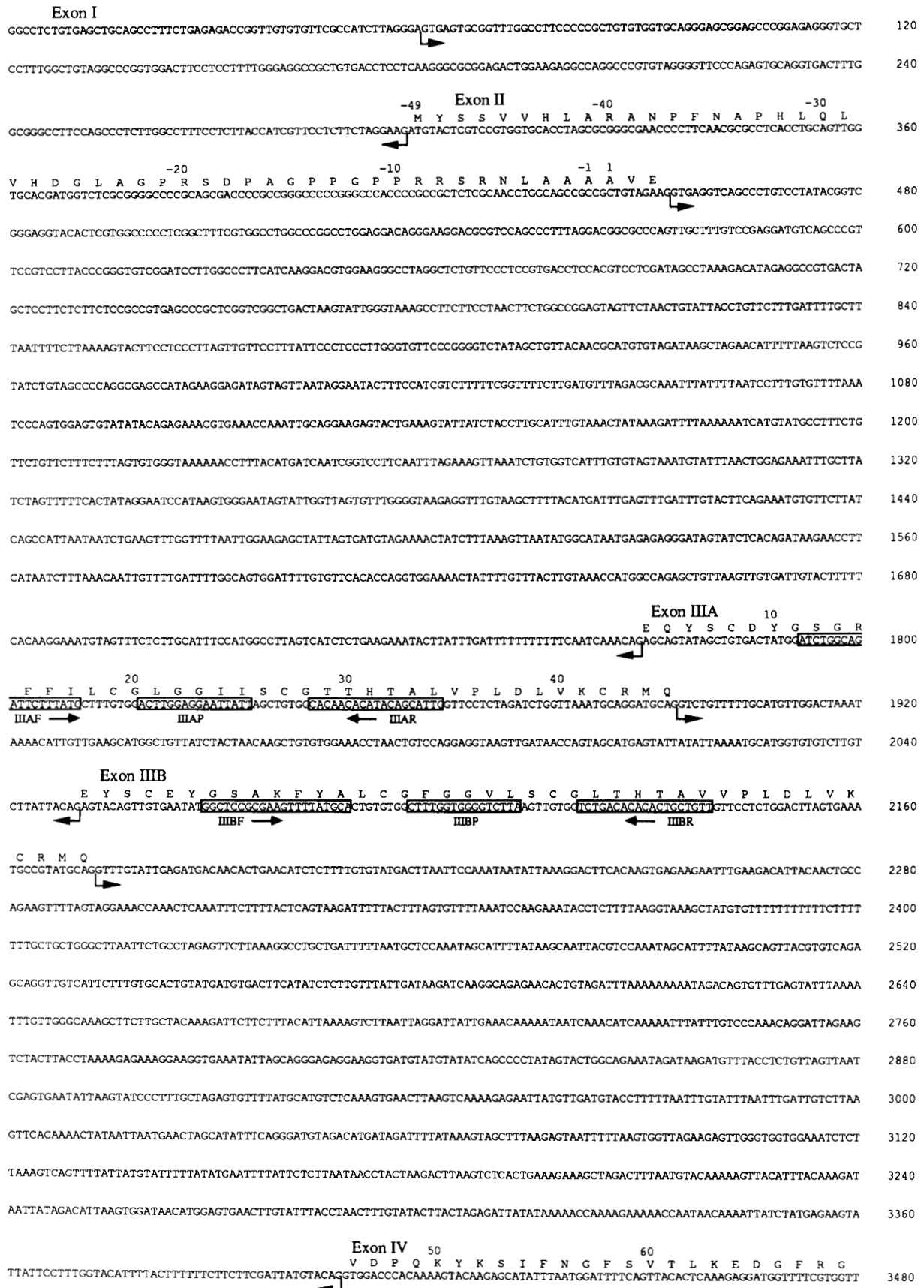


Fig. 4. DNA sequence of the bovine phosphate carrier gene. In exons IIIA and IIIB are shown the positions of primers (IIIA-F, IIIA-R, IIIB-F, and IIIB-R) and probes (IIIA-P and IIIB-P) employed in the investigation of alternatively spliced forms in various bovine tissues. For the meaning of the other symbols and further information, see legend to Fig. 2.

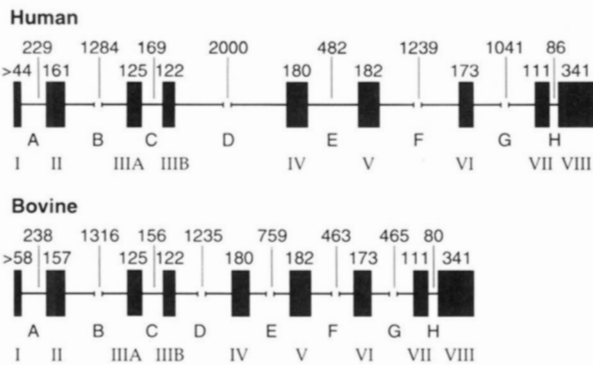


FIG. 5. Structures of the human and bovine genes encoding the mitochondrial phosphate carrier. The exons and introns are shown as filled boxes and continuous lines, respectively, and their sizes are given in base pairs.

positively hybridizing clones were identified. Three of them, λ 11D, λ 9M and λ 4D, were analyzed, but only λ 11D contained the full-length cDNA. This is the human cDNA sequence of the phosphate carrier reported previously (Dolce *et al.*, 1991). The λ 9M and λ 4D clones are partial and extend from the *Xba*I site at nucleotide 1804 in the genomic sequence. Sequences (960 and 826 bases, respectively) were determined from the 3' ends of λ 9M and λ 4D. It is clear that they contain the sequence found in human exon IIIA, whereas the reported sequence of λ 11D (Dolce *et al.*, 1991) contains exon IIIB.

Alternative Splicing of Exons IIIA and IIIB in Bovine Tissues—The distribution of the two different transcripts arising from alternative splicing of exons IIIA and IIIB were studied by polymerase chain reactions conducted on mRNAs from bovine heart, liver, brain, kidney, and lung. The primers (shown in Fig. 4) were chosen from the regions of greatest nucleotide sequence divergence between bovine exons IIIA and IIIB. The amplified products from these polymerase chain reactions were hybridized with specific probes from exons IIIA and IIIB, respectively. A hybridizing band was detected in all of the tissues that were examined (Fig. 6), and therefore both of the possible alternative transcripts were present in all instances. In order to confirm that the specific product was being amplified by each pair of primers, the products from the heart and liver polymerase chain reactions were cloned and sequenced. The sequences of the products obtained with the primers from exon IIIA corresponded to exon IIIA, and those obtained with the primers from exon IIIB corresponded to exon IIIB.

Different intensities of hybridization with the two probes were observed with the various tissues. Since the same amount of mRNA was used in each experiment, and the conditions of reaction were the same, the amounts of transcripts containing exon IIIA and exon IIIB seem to differ from tissue to tissue. The highest expression of exon IIIA was found in heart and liver, much less was found in brain and kidney, and relatively little was present in lung. In contrast, the expression of exon IIIB was greater in lung than in brain and kidney, and it was weakest in heart and liver.

The biological reasons underlying these observations are not understood at present. Exons IIIA and IIIB encode the N-terminal region of the phosphate carrier protein, which encompasses its first transmembrane segment and part of the first hydrophilic loop. The functions of these regions have not been established. The effects of the alternative splicing on the activity of the phosphate carrier require further study, probably involving their overexpression and reconstitution, as described recently for the related oxoglutarate-malate carrier (Fiermonte *et al.*, 1993). Alternative splicing has been shown to operate in one other protein associated with the inner membranes of

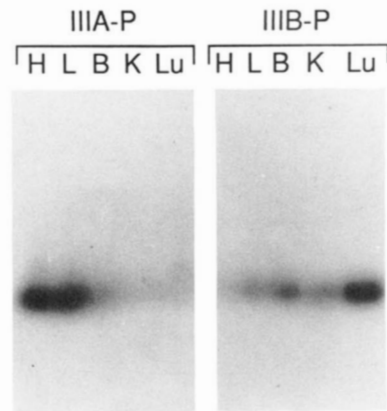


FIG. 6. Hybridization of the products of polymerase chain reactions on mRNAs from various bovine tissues using primers from bovine exons IIIA and IIIB. The products from heart (H), liver (L), brain (B), kidney (K), and lung (Lu) were separated by electrophoresis and were detected by hybridization with radiolabeled oligonucleotide probes, IIIA-P (left-hand side) and IIIB-P (right-hand side; see Fig. 4 for details of primers and probes).

mammalian mitochondria, namely the γ -subunit of the ATP synthase complex (Matsuda *et al.*, 1993), but again the biological significance is not understood.

Acknowledgments—We are indebted to Drs. T. H. Rabbitts and E. R. McCabe for providing the human genomic and cDNA libraries, respectively. We thank M. J. Runswick for help.

REFERENCES

- Aquila, H., Link, T. A., and Klingenberg, M. (1985) *EMBO J.* **4**, 2369–2376
 Battini, R., Ferrari, S., Kaczmarek, L., Calabretta, B., Chen, S., and Baserga, R. (1987) *J. Biol. Chem.* **262**, 4355–4359
 Biggin, M. D., Gibson, T. J., and Hong, G. F. (1983) *Proc. Natl. Acad. Sci. U. S. A.* **80**, 3963–3965
 Bisaccia, F., and Palmieri, F. (1984) *Biochim. Biophys. Acta* **766**, 386–394
 Breathnach, R., and Chambon, P. (1981) *Annu. Rev. Biochem.* **50**, 349–383
 Capobianco, L., Brandolin, G., and Palmieri, F. (1991) *Biochemistry* **30**, 4963–4969
 Chirgwin, J. M., Przybyla, A. E., MacDonald, R. J., and Rutter, W. J. (1979) *Biochemistry* **18**, 5294–5299
 Cozens, A. L., Runswick, M. J., and Walker, J. E. (1989) *J. Mol. Biol.* **206**, 261–280
 Dolce, V., Fiermonte, G., Messina, A., and Palmieri, F. (1991) *DNA Sequence* **2**, 133–135
 Dyer, M. R., and Walker, J. E. (1993) *Biochem. J.* **293**, 51–64
 Ferreira, G. C., Pratt, R. D., and Pedersen, P. L. (1989) *J. Biol. Chem.* **264**, 15628–15633
 Fiermonte, G., Walker, J. E., and Palmieri, F. (1993) *Biochem. J.* **294**, 293–299
 Garber, R. L., Kuroiwa, A., and Gehring, W. (1983) *EMBO J.* **2**, 2027–2036
 Gawaz, M., Douglas, M. G., and Klingenberg, M. (1990) *J. Biol. Chem.* **265**, 14202–14208
 Houldsworth, J., and Attardi, G. (1988) *Proc. Natl. Acad. Sci. U. S. A.* **85**, 377–381
 Iacobazzi, V., Palmieri, F., Runswick, M. J., and Walker, J. E. (1992) *DNA Sequence* **3**, 79–88
 Kaplan, R. S., Mayor, J. A., and Wood, D. O. (1993) *J. Biol. Chem.* **268**, 13682–13690
 Kolarov, J., Kolarova, N., and Nelson, N. (1990) *J. Biol. Chem.* **265**, 12711–12716
 Kolbe, H. V. J., Costello, D., Wong, A., Lu, R., and Wohlrab, H. (1984) *J. Biol. Chem.* **259**, 9115–9120
 Krämer, R., and Palmieri, F. (1989) *Biochim. Biophys. Acta* **974**, 1–23
 Krämer, R., and Palmieri, F. (1992) in *Molecular Mechanisms in Bioenergetics* (Ernster, L., ed) pp. 359–384, Elsevier Science Publishers, Amsterdam
 LeFranc, M. P., Forster, A., Baer, R., Stinson, M. A., and Rabbitts, T. H. (1986) *Cell* **45**, 237–246
 Matsuda, C., Endo, H., Hirata, H., Morosawa, H., Nakanishi, M., and Kagawa, Y. (1993) *FEBS Lett.* **325**, 281–284
 Neckelmann, N., Li, K., Wade, R. P., Shuster, R., and Wallace, D. C. (1987) *Proc. Natl. Acad. Sci. U. S. A.* **84**, 7580–7584
 Palmieri, F., Bisaccia, F., Capobianco, L., Dolce, V., Fiermonte, G., Iacobazzi, V., and Zera, V. (1993) *J. Bioenerg. Biomembr.* **25**, 493–501
 Phelps, A., Schobert, C. T., and Wohlrab, H. (1991) *Biochemistry* **30**, 248–252
 Powell, S. J., Medd, S. M., Runswick, M. J., and Walker, J. E. (1989) *Biochemistry* **28**, 866–873
 Ricquier, D., Casteilla, L., and Bouillard, F. (1991) *FASEB J.* **5**, 2237–2242
 Runswick, M. J., Powell, S. J., Nyren, P., and Walker, J. E. (1987) *EMBO J.* **6**, 1367–1373
 Runswick, M. J., Walker, J. E., Bisaccia, F., Iacobazzi, V., and Palmieri, F. (1990) *Biochemistry* **29**, 11033–11040
 Runswick, M. J., Philippides, A., Lauria, G., and Walker, J. E. (1994) *DNA Se-*

- quence, in press
- Sanger, F., Nicklen, S., and Coulson, A. R. (1977) *Proc. Natl. Acad. Sci. U. S. A.* **74**, 5463-5467
- Saraste, M., and Walker, J. E. (1982) *FEBS Lett.* **144**, 250-254
- Staden, R. (1982) *Nucleic Acids Res.* **10**, 4731-4751
- Viñas, O., Powell, S. J., Runswick, M. J., Iacobazzi, V., and Walker, J. E. (1990) *Biochem. J.* **265**, 321-326
- Walker, J. E. (1992) *Curr. Opin. Struct. Biol.* **2**, 519-526
- Walker, J. E., Cozens, A. L., Dyer, M. R., Fearnley, I. M., Powell, S. J., and Runswick, M. J. (1987a) *Chem. Scr.* **27B**, 97-105
- Walker, J. E., Gay, N. J., Powell, S. J., Kostina, M., and Dyer, M. R. (1987b) *Biochemistry* **26**, 8613-8619
- Walker, J. E., Powell, S. J., Vinas, O., and Runswick, M. J. (1989) *Biochemistry* **28**, 4702-4708
- Walker, J. E., Arizmendi, J. M., Dupuis, A., Fearnley, I. M., Finel, M., Medd, S. M., Pilkington, S. J., Runswick, M. J., and Skehel, J. M. (1992) *J. Mol. Biol.* **226**, 1051-1072
- Walker, J. E., and Runswick, M. J. (1993) *J. Bioenerg. Biomembr.* **25**, 435-446
- Zara, V., Palmieri, F., Mahlke, K., and Pfanner, N. (1992) *J. Biol. Chem.* **267**, 12077-12081